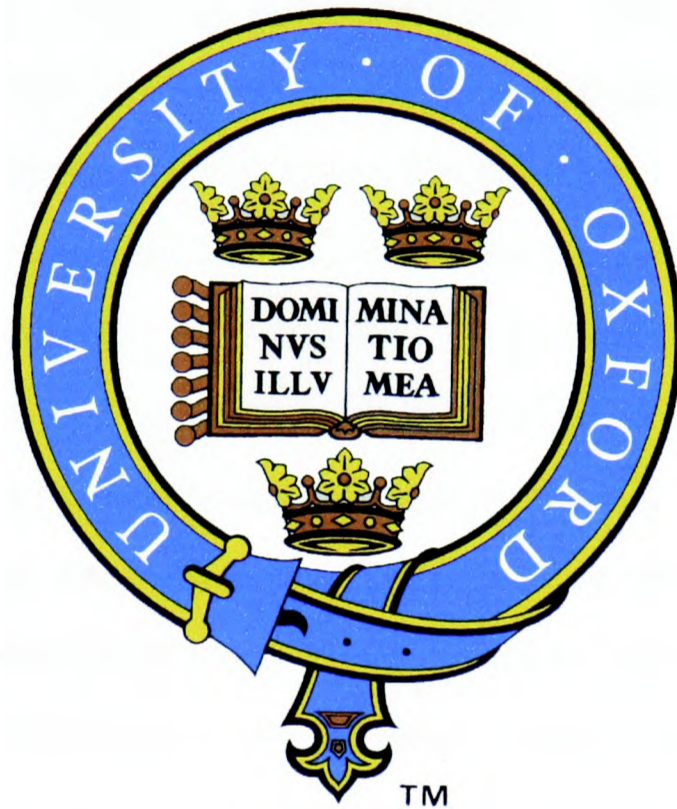


Factors Affecting Breeding in Captive Carnivora.



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ABSTRACT

Captive carnivores pose a challenge for conservationists and institutions alike, presenting many problems that range from diseases to poor welfare and unsuccessful breeding. Available databases of captive populations are rich sources of information that can help determine which factors can affect breeding success and the real potential of these populations in conservation programmes. Some species, such as tigers *Panthera tigris*, seem to preserve in captivity the same reproductive parameters seen in wild animals, making captive individuals extremely useful in the research of reproductive biology, that can be applied in evolutionary and physiological studies of the order Carnivora. Specific reproductive characteristics, mainly connected with the altriciality of the young, can make some species more prone to lose young in captivity than others, and these factors must be taken into consideration when developing *ex situ* conservation programmes. Infant mortality in captivity seems to be primarily caused by inadequate maternal behaviour, which can be connected to biological factors as well as to individual characteristics such as origin and rearing methods. Maternal infanticide, either passive or active, is also affected by biological and ecological characteristics of the species, and there may be an effect of the origin of the females, i.e. if they were wild-caught or captive-born. Housing conditions and individual history affect infant mortality, with females that suffered transfer between institutions exhibiting lower breeding success. Also, institutions with thriving research programmes presented higher infant mortality overall, independently of their latitude or management system, which can indicate an effect of human interference. Further research, both in the wild and in captivity, is needed to fully understand the factors affecting breeding success of captive carnivores.

To my dam, Judith.

In loving memory of
My father, Reinaldo, who sired me
My uncle, Elson, who fostered me
And our patriarch, Alfredo, alpha-male
You all left the group while I was away.

Goodbye.

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Chapter 1

Conservation and management aspects of carnivores in captivity.

Abstract

This chapter is a review of the conservation status of terrestrial carnivores and the challenges faced by conservation programmes. In average, a proportion of 58.6% of all families of terrestrial Carnivora are listed by IUCN in some level of threat. Conservation programmes are running to cover many species, both in the wild and captivity. The role of zoological institutions in conservation programmes is crucial, especially for those species with rapidly declining wild populations. Researchers face different challenges in the wild and captivity. In the wild, these are mostly related to conflicts between human populations and governments on the use of land assigned for conservation purposes. In captivity, zoologists face the lack of appropriate housing conditions and biological information on many species, as well as behavioural and physiological disorders caused by confinement. Conservation efforts must be interdisciplinary, with a stronger involvement from the human populations surrounding conservation areas and a direct effort from researchers and zoological institutions to exchange knowledge and set strict priorities prior to any practical investment.

1.1. Introduction.

Although the Order Carnivora¹ is not the most endangered mammalian order, it does contain some of the most charismatic species in the Animal Kingdom. This appeal has resulted in specimens being kept in hundreds of zoological institutions all around the globe, and species listed as threatened or endangered are frequently the subject of conservation and re-introduction programmes.

Managing predators or long-range foragers in captivity, however, is a very complex task. Not only are many wild species susceptible to diseases of

¹Although it appears that there is no evolutionary reason to exclude the families Phocidae and Otariidae from the Order Carnivora (e.g. Bininda-Emonds, Gittleman & Purvis 1999), in this work only terrestrial carnivores were included, due to fundamental differences in the housing, husbandry and management of aquatic and terrestrial mammals.

their domestic relatives, but also some have very specific environmental demands, which are, in many cases, close to impossible to recreate in captivity. The failure to achieve these standards may lead to welfare problems, such as stereotypies, that can lead to higher juvenile mortality (Clubb & Mason 2000), and jeopardise conservation programmes.

The role of zoos in conservation programmes is controversial (Schaller 1996). Nevertheless, many institutions claim that their main responsibility, when keeping endangered species, is to take part in captive breeding schemes, usually for subsequent re-introduction in the wild, in what is known as *ex situ* conservation (De Boer 1992). Only recently the World Conservation Union (IUCN) released the guideline policy for *ex situ* conservation, in which captive breeding is recommended under strict circumstances (IUCN 2002a). However, the results of many captive conservation programmes held by zoological institutions open to visitation were never scientifically assessed.

In the search for a better understanding of the factors that may affect breeding in captive mammals, carnivores can be a very adequate group to be looked at. The species differ largely in their life histories, including physical dimensions (Gittleman 1986a), diet (Van Valkenburg 1989), habitat use (Taylor 1989), geographical distribution (Wilson & Reeder 1993), and activity pattern (Gittleman 1986a). These factors have such an impact in husbandry techniques that species-focused guidelines for zoological institutions were urged since 1990 (Roberts 1990). Reproductive parameters are also very diverse within the order, from the age of independence of the young and the age in which the young open their eyes (Gittleman 1986a) to the general energy output in reproduction (Oftedal & Gittleman 1989), but most species are very altricial, which can prove to be a problem in captive conditions. The variety of social systems (Gittleman 1989), parental care (Baker 1994; Baker, Baker & Thompson 1996) and patterns of social communication (Peters & Wozencraft 1989; Gorman & Trowbridge 1989)

have to be considered when housing groups, and mistakes can be punished by the death of a potential founder (e.g. Brocklehurst 1997; von Schmalz Peixoto 1998).

Another advantage of using carnivores as a model for this type of research is the relative abundance of institutional records. Apart from certain institutions dedicated to a single species, most of the zoos and aquaria of the world keep at least one species of the order (ISIS 2003). The overall lack of success of captive breeding programmes for carnivores (De Boer 1992) is an additional incentive to the use of these species for this research.

1.2. Conservation status of the Order Carnivora.

There are around 240 terrestrial species in 8 families of the Order Carnivora, and 109 species, or 45.42% of those, were classified as threatened in the last edition of the Red List from IUCN, making it the fourth most threatened mammalian order with more than 100 species (IUCN 2002b). Presenting this proportion as a pooled value, though, can be confounding: within families, the proportion of species reported as in some level of risk of extinction in the IUCN Red List ranges from 23.7% for the family Herpestidae to 75% in the small families of Ursidae and Hyaenidae, with an average of 58.6%. According to the IUCN Red List Categories and Criteria version 3.1 (2001), these species are, or will be in the near future, at risk of extinction if no action is taken place. Species are considered extinct (EX) when there is no reasonable doubt that the last specimen has died, confirmed by surveys in the historic range of the species, but when it still survives in captivity or artificial propagation, it is considered extinct in the wild (EW). The threatened species are classified as critically endangered (CR), endangered (EN) or vulnerable (VU) when at high risk of extinction, at different levels of threat, through loss of habitat or individuals. If the populational or geographical loss does not meet the statistical criteria for high risk of extinction, the species is evaluated as near threatened (NT). Data deficient species (DD) do not have enough support of adequate information on distribution and abundance to be evaluated safely, even when the species is well studied and many biological aspects are known. IUCN recognises the need for more accurate information on DD species and acknowledges that a threatened status may be appropriate. Widespread, abundant species are classified as least concern (LC), and some taxa were not evaluated (NE) for the Red List, once their abundance is evident. In the order Carnivora, the proportion of species in each category varies greatly between families, but those containing species with large body size seem to be more affected than

families with medium to small body size (Figure 1.1). Body mass was one of the factors found to predict extinction in declining species (Purvis *et al* 2000). In a recent work, Cardillo *et al.* (2004) proposed a model which combines biological factors of carnivore species and human density in the areas in which they occur, explaining how a biologically sensitive species suffers higher impact of human population expansion.

Several carnivore species are subjects of *ex situ* conservation programmes, and many were reintroduced into their historical range. It appears, however, that many declining species, although in urgent need of direct conservation actions, have been overlooked. For example, in 165 reintroduction programmes of carnivores, only 28, or 16.9%, involved threatened species (Gittleman & Gompper 2001). This may reflect either the existence of multiple programmes for few threatened species (Mace *et al.* 2000) or the decision of programme managers to prioritise charismatic species over threatened ones that could benefit from these efforts (Balmford, Mace & Leader-Williams 1996). The importance of captive breeding programmes will be discussed in section 2.2.1.

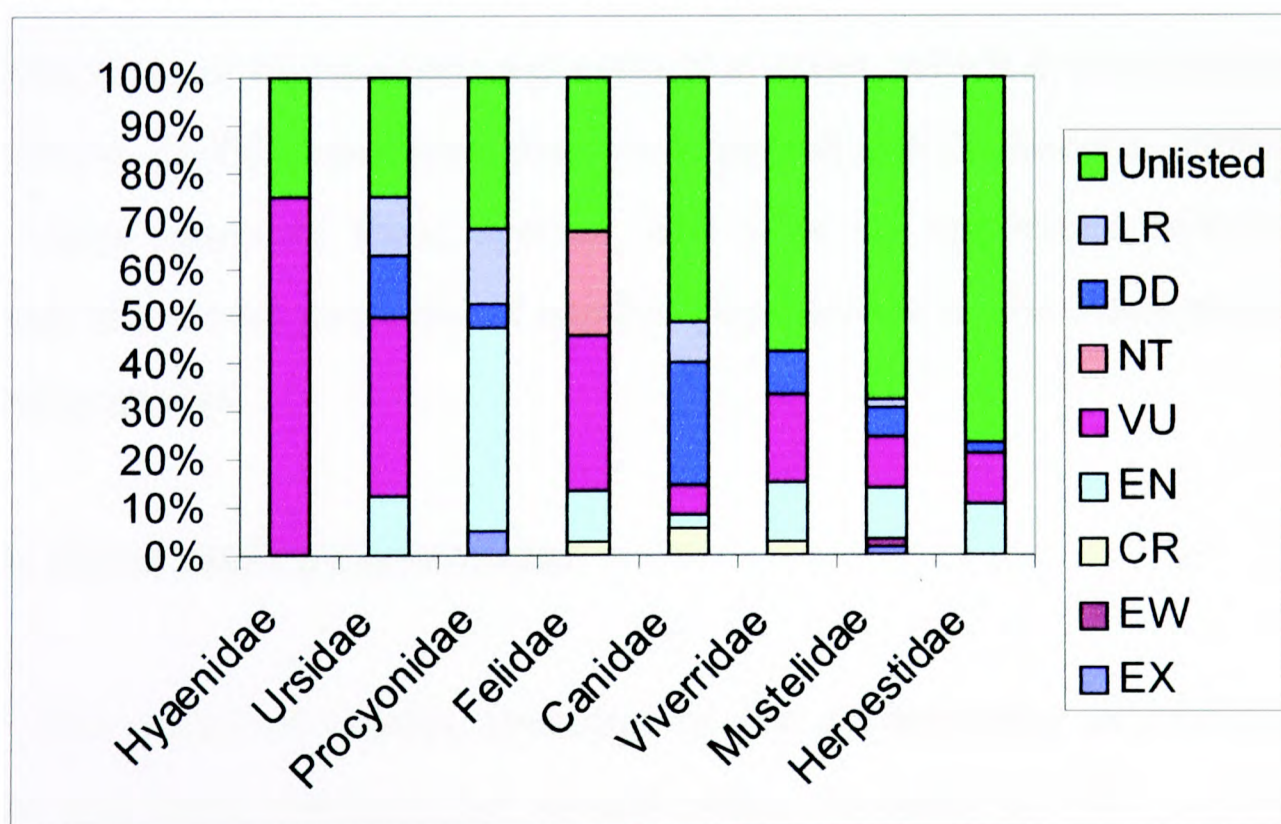


Figure 1.1: Proportion of species in each family of the order Carnivora that are unlisted or listed in diverse categories on the 2002 IUCN Red List. LR = low risk; DD = data deficient; NT = near threatened; VU = vulnerable; EN = endangered; CR = critically endangered; EW = extinct in the wild; EX = extinct.

The same situation occurs in zoological institutions around the world. While, on average, 75.3% of non-threatened carnivore species have bred more than 5 litters in captivity, worldwide, from 1986 to 1996, only 45.2% of the threatened species did so (see Chapter 2). Once again, these values vary among families (Table 1.1).

Table 1.1: Proportion of endangered species by family of carnivores, proportion of endangered species that bred at least 5 litters in captivity from 1986 to 1996 and proportion of non-endangered species that bred at least 5 litters in captivity from 1986 to 1996.

Family	Endangered species (%)	Captive-bred endangered species (%)	Captive-bred non-endangered species (%)
Canidae	50	50	61.1
Felidae	67.5	64	91.6
Herpestidae	23.7	0	20.7
Hyaenidae	75	66	100
Mustelidae	32.3	38.1	43.2
Procyonidae	68	15.4	100
Ursidae	75	83.3	100
Viverridae	42.4	0	31.5
Average	58.6	45.2	75.3

The present human-induced extinction crisis, which is developing at a rate 1000 to 10,000 times faster than the expected rate (Pimm *et al.* 1995), will soon claim many of these species, and it is an opportune moment to consider the actual potential of captive populations in the management of declining species.

1.2.1. Zoos and conservation.

The value of captive breeding in the conservation of endangered species has been discussed for several years. As early as 1979, Leyhausen argued that captive breeding could never be the solution for endangered species, since conservation efforts should consider the long term process of evolution, which would be lost in small captive populations. The most

recent guidelines for *ex situ* management published by IUCN (2002a), however, points out that the goal of conservation is to maintain “biological interactions, ecological processes and function”, through the management of wild populations and habitats, and the development of self-sustainable captive populations, aiming the production of individuals for reintroduction, when needed. Gittleman & Gompper (2001) affirm that the re-colonisation of habitats by declining species is a major goal of conservation efforts. Jalme (2002) pointed out the various aspects of captive breeding for conservation of avian species, which can be applied to many mammalian species: captive populations can replace wild populations in research and education, may provide genetic reservoirs to reinforce or found wild populations and can be a last resort for species whose wild populations cannot be maintained by other means. It is generally agreed, however, that *ex situ* conservation depends on high budgets, and must be used only as a support for ongoing *in situ* conservation programmes.

Comparing the budget of *ex situ* and *in situ* conservation programmes shows that the costs per capita for a efficient plan *in situ* are much lower than in captive breeding programmes, such as those for large mammals with scarce wild populations (Balmford, Leader-Williams & Green 1995). For instance, the costs of captive breeding programmes of larger felids are extremely high because of the species’ requirements for large territories, the management of a large founder population (due to the higher level of homozygosity of predators) and the expenses with assisted reproduction techniques (ARTs), factors present also in other families of the order Carnivora (Conway 1986). Examining the costs of keeping several taxa, from invertebrates to vertebrates, it was found that body mass increases the costs, and the result was the same when examining only mammals; the need for much larger captive populations for smaller species, due to faster breeding and quicker loss of genetic variability, seems to eventually level costs between large and small species (Balmford, Mace & Leader-Williams 1996).

The IUCN warns of the danger of leaving the development of *ex situ* techniques until a species is on the verge of extinction, since it may be too late for the formation of a viable founder population (IUCN 2002a).

The genetic management of captive populations is of great concern in captive breeding programmes. Although loss of genetic variation through generations is inevitable, there are ways to minimise it: to select founders from different wild demes; reduce the time in captivity before re-introduction; and expand the period between generations if the population has to be kept in captivity. Also, different captive populations will lose different sets of alleles, and selecting individuals from different breeding programmes to start a reintroduced population allows a larger variation that would otherwise be possible (Mace 1986). Computer databases, such as ISIS and electronic studbooks, have been used to track genetic information from captive populations, and can be used to simulate loss of variation through successive generations, giving background to a safer and more effective conservation plans (Flesness & Mace 1988). One example of how effective the use of these new techniques can be is the very successful *ex situ* programme on Rodrigues fruit bats (*Pteropus rodricensis*): the introduced populations are stable, with a safe genetic variation and continue to subdivide, occupying new areas (Carroll & Mace 1988).

Some programmes have been less successful, and can reflect the need for a very careful planning. One example is the black-footed ferret (*Mustela nigripes*) that had almost disappeared due the eradication of the prairie dogs (*Cynomys* sp.), its main prey, from North American prairies. The last wild population was discovered in 1981, and a captive breeding programme was started, as a way to increase wild populations through re-introduction. Unfortunately, an outbreak of canine distemper, spread probably by the researchers, wiped out the wild colony. Now, captive propagation is the only method that can preserve this species. Some valuable lessons have been learned, such as keeping viable genetic diversity, and increasing and

subdividing the captive population. Re-introduction efforts started in 1991 (Thorne & Oakleaf 1991), and through extensive research, the programme is starting to obtain positive results (Wolf *et al.* 2000a).

One particular programme is not achieving good results, even with the help of a large budget and several Assisted Reproduction Techniques: the conservation of the giant panda (*Ailuropoda melanoleuca*). Despite all efforts, more than half of the cubs born in 10 countries from 1963 to 1998 (226 individuals) died before 30 days after birth, and only 30% of the surviving cubs lived for 3 years in captivity. The slow breeding biology of the species and the inadequate behaviour displayed by hand-raised individuals are thought to be among the causes (Peng, Jiang & Hu 2001).

Conservation programmes should also consider problems of re-introduction. When re-introducing predators into areas where they have disappeared, prey populations can be easily affected once they are not prepared to defend themselves. Naïve prey populations were eliminated in the Pleistocene when faced by the first human hunters (Gittleman & Gompper 2001). In a study of the impact of re-colonising brown bear (*Ursus arctos*) and wolf (*Canis lupus*) populations on moose (*Alces alces*) groups in Europe and North America showed that naïve moose, which were unable to recognise predator cues, suffered higher predation-related mortality than experienced ones. Offspring predation, however, seems to elicit maternal hypersensitivity towards predator cues, contributing to subsequent increased survival rates. This shows that this species is swift in learning and adapting to changing predator populations, which may not have been true for the Pleistocene species eradicated by humans (Berger, Swenson & Persson 2001).

Naïveté can be also a problem for re-introduced predators. In the case of red wolves (*Canis rufus*), that were becoming extinct in the 1970s, the US Fish and Wildlife Service and Point Defiance Zoo started the Red Wolf Captive Breeding Program in Tacoma, Washington, in 1973. There are about

125 red wolves in captivity. They released captive-born animals in protected or rarely used areas, all individuals carrying a radio-collar. Meetings with neighbour human populations were held to enhance public recognition of the problem. Thirty-nine wolves were released and 12 died, three hit by cars and the others of “natural causes”. To avoid further deaths, the population was tracked by radio, supplementary diet was provided when necessary and reproduction was, in some cases, assisted by recapturing wolves to pair and breed within acclimation pens (Moore & Smith 1990). A recent review of 116 re-introduction programmes showed that only 26% were classified as successful over time, although these values vary between years and countries. A series of measures to improve success in these programmes was suggested, such as identifying and removing the primary cause of decline and releasing large groups of animals ($n > 100$) containing some wild-caught individuals (Fischer & Lindenmayer 2000). The release of wild-caught individuals together with a re-introduced population seems also to enhance the chances of success in African wild dogs, *Lycaon pictus* (Woodroffe & Ginsberg 1997), although some researchers put the blame of failure in re-introduction of captive-born African wild dogs on the lack of collaboration between captive breeding centres and nature conservation institutions (Frantzen, Ferguson & de Villiers 2001).

Conservation strategies have been used by The Jersey Wildlife Preservation Trust, Wildlife Preservation Trust International and WPT Canada, together with governmental organisations and zoos to research captive breeding and try to restore or create habitats to endangered species. The whole project involves also public education, personnel training and field research, with funding to captive and zoo-based breeding research and land purchase (Mallinson 1991).

Since *ex situ* techniques depend on captive founders, captive stocks are a fundamental part of any conservation programme. The role of zoological institutions in conservation programmes, however, goes beyond

captive breeding. Physiological and behavioural research, fundamental for the understanding of the biology of species, and public education cannot be provided by any other source (Smith *et al.* 2002). Today, the focus is shifting for a better understanding of the impact of captivity on the welfare of the individuals, and the scientific community is beginning to recognise the importance of research with captive populations.

1.3. Captive wild carnivores: conservation and husbandry.

Keeping wild carnivores in captivity poses several challenges, from the security of big predators to welfare and conservation problems. Research is fundamental to identifying and overcoming these difficulties. Although most of physiological problems are now well known and can be avoided with simple measures, some behavioural dysfunctions are still unclear. This section is an overview of the most common problems of wild carnivores in captivity, and which solutions have been proposed to increase the welfare of these species.

1.3.1. General health problems.

Carnivores are susceptible to a variety of common feline and canine diseases, but some can be prevented through the use of commercial vaccines. Nevertheless, a few species can develop vaccine-induced diseases, and live strains must be avoided (Table 1.2).

Panleucopaenia is a common cause of death of zoo carnivores, and the presence of feral cats inside zoo facilities can be a permanent danger. One young male clouded leopard (*Neofelis nebulosa*) was found dead when 6 months old, victimised by panleucopaenia spread by feral cats during an epidemic of the disease (Geidel & Gensch 1976). Feline infectious peritonitis

(caused by calicivirus) and panleucopaenia are responsible respectively for 6 and 4% of captive cheetahs (*Acinonyx jubatus*) deaths (Marker-Kraus 1997).

Pneumonia affects carnivores from bush dogs (*Speothos venaticus*) (Kitchener 1968) to fishing cats (*Felis viverrina*) (Jayewardene 1975), but can be treated if detected in early stages. Sand cats (*Felis margarita*) are highly sensitive to lung diseases, especially in cages with high humidity levels, but the most common infection is rhinotracheitis, spread commonly by feral cats. They are susceptible also to degenerative liver diseases and myocarditis (Sausman 1997). In felids, kidney diseases cause death of many individuals. They are the most common death cause in zoo-housed cheetahs over 6 months of age (Marker-Kraus 1997) and adult black-footed cats (*Felis nigripes*) (Olbricht & Sliwa 1997).

Table 1.2: Vaccines commonly used in captive wild carnivores and their limitations (from Roberts 1975; Scheffel & Hemmer 1975; Hulley 1976; Hinshaw, Amand and Tinkelman 1996; Deem *et al.* 2000).

Family	Vaccines used	Problems
Canidae	Canine distemper, canine infectious hepatitis, leptospirosis, parvovirus, rabies.	Cape hunting dogs (<i>Lycaon pictus</i>) can develop vaccine-induced distemper (must be used only chick-embryo-origin vaccine).
Felidae	Feline panleucopaenia (feline distemper), rhinotracheitis, calicivirus, rabies.	Should be used only killed vaccine to distemper. There is no vaccine to feline coronavirus (infectious peritonitis). Jaguarondi (<i>Herpailurus yagouarundi</i>) can develop vaccine-induced panleucopaenia, and Geoffroy's cats (<i>Felis geoffroyi</i>) can develop feline distemper.
Ursidae	Rabies	
Procyonidae	Canine and feline distemper annually. Rabies.	Kinkajous (<i>Potos flavus</i>) and red pandas (<i>Ailurus fulgens</i>) can develop vaccine-induced canine distemper from live vaccine. Red pandas are sometimes vaccinated against hepatitis and leptospirosis.
Viverridae	Canine and feline distemper annually.	
Mustelidae	Canine and feline distemper annually.	Ferrets (<i>Mustela putorius</i>) can develop vaccine-induced canine distemper. Susceptible to parvovirus.
Hyaenidae	Rabies and canine distemper annually.	

Parasitic infections are not, in general, life threatening, and can be easily controlled through antihelmintic oral solutions or vaccines. In nature, however, some species are susceptible to very specialised parasites that can lead to death. A female wild-caught maned wolf (*Chrysocyon brachyurus*), kept on Frankfurt Zoo, died of acute sero-glomerulo-nephritis. *Post-mortem* exams revealed that she had one kidney missing. This was caused by nematodes *Dioctophyma renale*, common parasites of maned wolves, that when untreated tends to destroy the kidneys (Faust & Scherpner 1975).

Malnutrition affects a large number of carnivores, due to the common mistake that feeding them solely with clean meat is sufficient to ensure good health. Felids feed largely on prey, but they consume not only the flesh, but also skin, cartilage, small bones and organs, such as liver and spleen. Giving whole carcasses instead of clean meat can prevent common nutritional problems like greenstick fractures and bone deformities (Ashton & Jones 1979). Nutritional deficiencies may be the cause of several bone deformities, but some species have a tendency to develop them. Bush dogs, like small domestic dogs, can suffer from bilateral patellar luxation, a painful condition that impeaches normal movement (Kitchener 1968).

Hand-reared animals can develop milk-induced diarrhoea that may lead to death, especially if the composition of the natural milk is not known, as in bush dogs (Kitchener 1968).

Other health problems range from hyperparathyroidism and encephalitis (Hulley 1976) to deaths by abdominal gas distension said to be caused by a sensitivity reaction, in the lack of a more accurate explanation (Murphy 1976).

Cancer occurs with some frequency in caged carnivores, and the causes of it are several. In the last years, however, veterinarians had discovered that vaccination can cause sarcoma in domestic cats (Esplin *et al.* 1993; Macy & Bergman 1995), and may be related to a local dermal inflammation resultant from the postvaccinal reaction (Macy & Hendrick 1996).

General health problems are responsible for a large number of deaths, but with appropriate veterinarian advice, most of these problems can be avoided or treated. It is important to say, however, that many of the zoological institutions included in this research do not have a veterinary doctor in their permanent staff, having an advisor that makes regular visits. Acute cases are usually treated by any veterinary surgeon available, who is sometimes not experienced in dealing with wild species. This could be the cause of the high numbers of deaths caused by these well-known conditions in some institutions.

1.3.2. Reproductive physiological problems.

Of all causes of poor breeding, low fertility is probably the easiest to identify. If a sexually mature couple, after obvious and repeated mating behaviour, fail to produce a pregnancy, infertility is immediately thought to be the cause. It is more complex, though, to define what is causing infertility in the first place.

Many factors can influence fertility in animals, apart from some obvious conditions, such as immaturity, old age, diseases and congenital abnormalities. In the wild, the three major suppressers of fertility are environmental factors (especially the variation on climate, day length and rainfall among seasons), lactation (which inhibits gonadotrophin, and then ovulation) and social dominance. In common marmosets (*Callithrix jacchus*), subordinate females had decreased gonadotrophin levels, and in naked mole-rats (*Heterocephalus glaber*), ovulation is suppressed in all females of a colony, except the "breeding queen". In both cases, removal of the infertile female from the presence of other females restores her fertility (Abbott 1988). External factors can also play a role in infertility. Environmental and industrial chemicals can influence reproductive fitness in animals, which is

well illustrated by the effects of polychlorinated biphenols in a population of Baltic ring seals (*Halichoerus grypus*) (Holloway & Moore 1988).

Mellen (1991) found negative correlations between number of litters produced by small felids, and number of medical treatments, latitude distribution of the species, and group size, and a very positive correlation with husbandry style. In general, small cats kept in groups of more than a pair (one male and one female) were unlikely to reproduce, but the ones cared for by caretakers that spent more time talking and interacting with them produced more litters. Reproductive failure of apparently healthy individuals may lead to the conclusion that the individuals do not produce gametes. In research on sperm efficiency in black-footed ferrets (*Mustela nigripes*), species in which around 55% of adult males fail to sire offspring (even when 90% of the females are receptive), it was revealed that the sperm quality of fertile and “infertile” males was the same; failure was occurring due to a combination of behavioural (improper mating position, excessive aggressiveness) and physiological (underdeveloped testes, lack of ejaculation) factors (Wolf *et al.* 2000b).

Contrasting with the low fertility problem, some species are excellent breeders, and contraception becomes the problem. There are many solutions for excessive breeding. Some zoos used to castrate males or keep female brown bears (*Ursus arctos*) alone (Van Keulen-Kromhout 1976), instead of using other available methods. Lions (*Panthera leo*) breed so well in captivity that contraceptive procedures are a common place. Hormonal implants, injections or even tablets are given to females, but the development of mammary cancer is a serious drawback to this technique. Vasectomy of males is a relatively simple surgery, and its risks are much lower (Ashton & Jones 1979). A recent trend is to find ways of interrupting early pregnancy, as with the use of antiprogestin in zoo-housed bears (Jewgenow *et al.* 2001).

It may seem that excessive breeding would be a positive step towards the conservation of a species. However, it is not only detrimental to the

health of the females, but the burden of many young may trigger brood-reducing behaviour in mothers, as it happens in rodents (Labov *et al.* 1985), or lead to infanticidal episodes in males. The major problem, though, is the issue of re-housing the young adults properly. Rare species are always in high demand, but more common species, usually the ones affected by excessive breeding, may give rise to serious housing problems. The use of reversible contraceptive methods is crucial for conservation purposes, and avoids the need in the future of using drastic “culling” methods, such as euthanasia (Graham 1996).

One of the most important aspects of reproduction that must be taken into consideration in a conservation programme is concerned with genetic variability and inbreeding. Inbreeding increases homozygosity and can allow expression of deleterious genes. Mortality during first six months after birth is higher in inbred youngsters of red pandas and leopards (*Panthera pardus*) (Carlstead 1996). In leopards, inbreeding seems to increase the frequency of cub killing (Carlstead 1996). Bred in captivity for over a century and a half, zoo populations of cheetahs are highly inbred. This genetic homogeneity seems to be responsible for the high rate of infant mortality and the low life expectancy of captive-born individuals (Marker-Kraus 1997), although this could also be an effect of differences in management practices, for values vary significantly between zoological institutions (Wielebnowski 1996).

For many species there are studbooks, where each individual of the captive population is listed, along with all its genetic relatedness to other individuals. This type of detailed record allows institutions to match couples from different genetic backgrounds, minimising the effects of inbreeding. Many rare species, however, have very small captive populations, and some, like the Sri Lankan rusty-spotted cat (*Prionailurus rubiginosa phillipsi*), have their whole captive population originated from one pair of founders from Frankfurt Zoo (Dmoch 1997). In these cases, it is

difficult to increase genetic variation, for it would imply in capturing new founders in the wild to start a conservation programme, that could be, in the end, unsuccessful. It seems, though, that inbreeding depression does not affect some species as seriously as it was thought some years ago, as in the cases of the Mexican wolf (*Canis lupus baileyi*) and the red wolf (*Canis rufus*), and this could be true to many other species (Kalinowski, Hedricks & Miller 1999).

In a captive breeding programme, having a pregnant female is a sign that only the first problem has been solved. Many gestations do not come to term, because of the high incidence of miscarriages, premature births and stillbirths among captive carnivores. For example, in cheetahs (*Acinonyx jubatus*), 20% of the cubs are stillborn, mainly because of premature births (Marker-Kraus 1997). In Okavandja Park, South-West Africa, brown hyaena (*Parahyaena brunnea*) pups at were born prematurely and died due the underdevelopment of lungs (Schulz 1966). In some species, to cross the threshold of 30 days after birth does not mean that the litter will survive. In the National Zoo, Washington, DC, USA, all the red panda cubs died before 130 days after birth, the mean age of death being 78.5 days, although the cause of these deaths were not reported (Roberts 1975).

The breeding biology of many species is not yet completely understood, and much research is still needed. Without this knowledge, the physiological causes of many of the problems listed above are mere conjecture. Furthermore, individuals demand particular treatments, so each case has to be minutely analysed. New techniques, such as non-invasive endocrinological methods, have been yielding valuable reproductive information, and can be used to survey the breeding status of captive females (Goodrowe *et al.* 2000). Maybe only the dynamic interchange of information between zoologists and veterinary surgeons can lead to a higher level of successful breeding.

1.3.3. Reproductive behavioural problems.

Captive breeding can be difficult due to inability of individuals to perform regular mating behaviour. Alternative patterns of sexual behaviour, such as masturbation and homosexuality, were recorded in the wild and cannot be described as abnormal, but their non-reproductive character may be a drawback for conservation programmes. Homosexuality is largely found among carnivores (Table 1.3), and masturbation has been recorded in several species. These behaviours promote sexual satisfaction without reproduction, but the individuals that perform them are also usually able to have, as long as a suitable partner is available, also regular reproductive activity (Bagemihl 1999). In the other hand, inadequate sexual behaviour may be originated from more complex mechanisms, like imprinting, and have long-lasting effects on the individuals, that can be unable to reproduce.

Behavioural inadequacies affecting reproductive performance may be attributed to deficient early rearing environments, the social milieu in which animals are kept in a long-term basis, or in the way potential mating pairs are put together (Lindburg & Fitch-Snyder 1994). In the case of the black-footed ferret described by Wolf *et al.* (2000b), 25% of unsuccessful breeding attempts were caused by the failure of males in adopting a proper mating position with the female. Inadequate mating behaviour is also among the causes of poor breeding success in captive giant pandas (Peng, Jiang & Hu 2001).

Table 1.3: Occurrence of homosexuality in carnivores (Bagemihl 1999)

Species	Gender	Type of behaviour	Level of occurrence	Observed in
African lion	M/F	Courtship, sexual, pair-bonding and parenting	Moderate	Wild, semiwild* and captivity
Cheetah	M/F	Courtship, sexual and pair-bonding	Moderate	Wild, semiwild and captivity
Red fox	M/F	Sexual and parenting	Incidental	Wild and captivity
Wolf	M/F	Courtship and sexual	Incidental	Captivity
Bush dog	M/F	Sexual and parenting	Incidental	Captivity
Brown bear	M/F	Sexual, pair-bonding and parenting	Moderate	Wild
American black bear	M/F	Sexual, pair-bonding and parenting	Moderate	Wild
Spotted hyena	F	Sexual	Moderate	Wild and captivity

* Such as large breeding centres or fenced nature reserves and protection areas.

1.3.4. *Infanticide.*

Infanticide has evolved as an ultimate mechanism of energy economy, or a way to minimise the others' fitness, improving the killer's own chances. Examples of nonparental infanticide in the wild include the nomadic lions in Serengeti, which expel and sometimes kill other males and eat their cubs (Schaller 1972). Killing other individuals' offspring may increase fitness by facilitating access to resources or enhancing breeding opportunities, and some species may have developed counterstrategies to minimise the occurrence of these events, such as termination of pregnancy, defence mechanisms against intruders and territoriality (Ebensperger 1998). Parental infanticide has different causes and benefits. In extreme situations, such as lack of food or threat of predation, mammals can respond by aborting or reabsorbing litters, or deserting or eating them if after birth. The benefits of these strategies may depend on the ability of the parents to produce another litter in the same season, on the cost of having another litter to the parent's reproductive value and on the probability of these youngsters, raised under adverse conditions, to breed when adults (Clutton-Brock 1991). Infanticide

also allows the alteration of reproductive patterns at the expense of others, and parental investment can be suppressed in any stage between conception and weaning (Hayssen 1984). Different types of infanticide have different aetiology and adaptive values (Hrdy 1979), but its occurrence in captivity can be detrimental to the success of breeding programmes. The impact of infanticide on the breeding success of captive carnivores is not yet well researched, and will be discussed in detail in Chapter 6.

1.3.5. Behavioural problems.

Individual behaviour can be affected by captivity (Carlstead 1996), leading to poor welfare (Mench & Kreger 1996). Even daily husbandry procedures can elevate levels of stress hormones and lead to inappropriate behaviour (Mendl *et al.* 2001). For social species, captivity puts the stability of a group under pressure, as an individual cannot abandon the group if social interactions become too agonistic, and sometimes has to change its own behaviour in a way to survive. For example, at Nairobi Zoo, two deformed pups of a litter of African wild dogs (*Lycaon pictus*) were killed when they were four months old. The only survivor had to assume a permanent submissive posture towards others (Cade 1975). Disputes over hierarchy are to blame for many fighting episodes and some casualties. In a group of dholes (*Cuon alpinus*), disputes over hierarchy led to severe fights among animals of the same sex (Sosnovskii 1975) that have lived before in apparent harmony. In a more serious event, the same disputes between a tamed male ocelot (*Leopardus pardalis*) and another male led to the death of the latter (Leyhausen 1966). This could happen because of the change in status an individual may go through when entering sexual maturity.

Respecting the social structure adopted by a species in the wild can avoid some problems with zoo-housed groups, and can even help with captive breeding efforts. For example, cheetahs frequently have poor

breeding in captivity. Although their behaviour in the wild is not completely unravelled, two factors are quite clear: 1) during much of lifetime, females are isolated from other adult conspecifics; 2) males move in hunting groups. Female isolation period goes from early pregnancy until the cubs are 16 months old, then again females are approached by males. In zoos, however, male and females are usually housed together, or at least in olfactory or visual contact, and this husbandry practice could be affecting the perception of the individuals of their housing condition (Benzon & Smith 1974). Other species are naturally solitary, and the attempt to house them in pairs or groups leads to severe injury of the animals, as it occurs with least weasels (*Mustela nivalis*) (Rosenthal 1971).

Whatever is the cause for excessive aggressiveness, there are reports of it in several species, especially felids. Table 1.4 displays some examples.

The development of stereotypies, a display of repetitive behavioural patterns with no apparent function (Mason 1991), in captive carnivores, seems to be related to the median home range size of the species (Clubb & Mason 2000). In carnivores, stereotypies are usually elicited by feeding, and pacing is the most common display. Changes in diet and feeding schedule can reduce pacing in cats (Mellen, Hayes & Shepherdson in press), and stereotypic pacing in leopard cats (*Prionailurus viverrinus*) was reduced 50% when the enclosure was provided with places to hide (Carlstead, Brown & Seidensticker 1993).

Table 1.4: Events of excessive aggressiveness in some Carnivora species in captivity.

Family	Species	No. of animals attacked	Event	Was the victim eaten?	Thought cause	Reference
Felidae	<i>Catopuma temmincki</i>	2	A male killed one female during introduction and, later, the female that was living with him.	N	Not available	Brocklehurst 1997.
Felidae	<i>Oncifelis geoffroyi</i>	1	A male killed a female.	Y	The animals were being fed live prey on the occasion.	Scheffel & Hemmer 1975.
Felidae	<i>Panthera onca</i>	1	A male preyed on a conspecific.	Y	Not available	Oliveira 1994.
Felidae	<i>Panthera tigris</i>	1	An introduced young male was killed by an adult male.	N	Competition among males. There were 4 females in the area.	Yost 1976.
Procyonidae	<i>Nasua narica</i>	1	One male was permanently under attack of females.	N	In the wild, females expel adult males from the groups.	Smith 1980.
Procyonidae	<i>Nasua nasua</i>	1	One introduced female was killed by 2 males and 7 females of a established group.	N	The group was stable, and the new female was introduced abruptly.	Von Schmalz Peixoto 1998.

Stereotypic displays can be also elicited by loud noises and cleaning procedures, as in fennec foxes (*Vulpes zerda*) (Carlstead 1991). An institution suffering frequent breeding failures has a good start towards improvement if it revises its current husbandry techniques, and checks more closely the relationship between keepers and animals.

1.4. Thesis structure and hypotheses to be tested.

This research aims to identify the factors affecting breeding success of captive carnivores, making use of the large amount of data already available from several sources. In this chapter, I review the conservation status of terrestrial carnivores, consider the importance of captive breeding, and present the most common challenges facing the management of captive carnivores. The subject and relevant questions of the next chapters are described below.

1.4.1. Chapter 2: Using databases in the research of infant mortality in captive carnivores.

In this chapter, I describe the available databases for zoo research, and the datasets on captive carnivores originating from them. I also present in detail a method of collecting and analysing this type of data, identifying the possible confounds and ways to overcome some problems. I suggest the use of relative indices as dependent variables and present the equations used in this research. I also test the variables used in multi-species analyses for possible effects of phylogeny and question the need to correct this issue.

1.4.2. Chapter 3: Factors affecting breeding in captive tigers (Panthera tigris): a studbook research.

In Chapter 3, I use the tiger studbook as a model to address the possibility of answering biological questions with record research. I point out that the tiger is a species with a large latitudinal range, thus excluding photoperiodic effects in breeding. Considering that there are abundant food and water in captivity, and that serious institutions have to provide adequate housing to their animals, it is not expected that biological

parameters, such as litter size, differ from wild populations. There are genetic, phenotypic and geographical differences between the two subspecies in the studbook, so it is expected that the dataset will reflect this and express the need of subspecies-orientated husbandry protocols. I hypothesise also that there will be little or no effect of inbreeding levels in this species for, like several carnivores, it presents high levels of homozygosity in wild populations.

1.4.3. Chapter 4: Biological factors affecting infant survival in captive carnivores.

Here I use multiple datasets to predict, through statistical models, the possible effects of the natural history of the species on infant mortality in captivity.

If conditions in captivity are providing all the demands of a species, as it would happen with a wild population during phases of abundant resources, then the specific proportion of infant mortality in captivity should be similar or slightly lower to those values for a wild population, because of the absence of natural phenomena such as fires, flash floods and predation. Otherwise, it would imply that other factors, rather than resource availability, are affecting these species' breeding success in captivity. The variance in the proportion of infant mortality between species may indicate that biological parameters have an influence on infant survival. In that case, it is possible that species with slower development were more prone to lose infants.

1.4.4. Chapter 5: Causes of mortality in young captive carnivores.

The causes of death of young carnivores in captivity are described in this chapter, and I consider the factors that can make certain species more prone to particular ailments. Also, I investigate the occurrence of infant deaths caused by inadequate maternal behaviour. I hypothesise, based on published reports, that a significant proportion of infant mortality in some species of carnivores in captivity is caused by the female. If so, most deaths shall occur in the first few weeks after birth, especially in species in which the young develop at a faster pace, due to the costs of maternal care.

1.4.5. Chapter 6: Maternal infanticide in captive carnivores.

The impact of inadequate maternal behaviour on infant mortality of captive carnivores is discussed in Chapter 6, where I also describe the mechanisms involved in maternal infanticide.

I hypothesise that species with altricial young will present higher proportion of active infanticide, and that some of the variance of infant mortality among institutions may be accounted for the history of the females (e.g., if they were wild-caught or captive-born) and institutional protocols (such as translocations and male presence during rearing).

1.4.6. Chapter 7: Increasing juvenile survival in captive carnivores: Practical considerations.

Here I review the techniques of assisted reproduction and the solutions proposed by institutions to minimise infant mortality, with special focus on hand-rearing survival rates.

A high variance in the proportion of infant mortality between institutions may suggest the influence of local characteristics such as the origin of the females and the number of translocations they were subjected to. If inadequate maternal behaviour was responsible for a significant proportion of infant deaths, and high infant mortality is related to institutional and individual characteristics, then infant mortality in captive carnivores can be greatly reduced if certain measures were taken, such as avoiding female translocations, providing more adequate housing conditions and respecting photoperiodic needs of breeding individuals.

1.5. Conclusion.

Carnivores are rapidly disappearing from many areas in the wild, and some species are declining at alarming rates. Captive stocks can help research and conservation efforts, but a change of priorities is needed, enhancing the focus on declining species rather than more abundant, but charismatic, ones. Conservation efforts need to be multidisciplinary, integrating several areas of science with local populations and governments, and a realistic approach of objectives and practical issues has to be addressed prior to any investment on the area.

The maintenance of carnivore species in zoological and research centres is yet cause of concern, and many techniques are still in their first attempts. A better communication between researchers around the world may lead to higher success, and the shifting role of zoos, from entertainment spaces to educational and research units, lifts the expectations for more successful integrative conservation programmes.

Chapter 2

Using databases in the research of infant mortality in carnivores.

Abstract

For many decades, zoological institutions have been keeping records of their animal stocks, many times with extreme detail. Main sources of information are the *International Zoo Yearbooks* published by the Zoological Society of London; the *International Species Information System*, created in 1978 by the American Association of Zoo and Aquaria; and Species Studbooks, kept initially as regional records of institutions and now available for many endangered species worldwide. Institutions keep private records of their collections and compile, many times, detailed information on individuals, including medical records and transfers. These data were very seldom used as bases for researching particular aspects of captive animals, due to the risk of bias and other statistical challenges. These problems can be overcome with the use of appropriate data collection and statistical analyses.

2.1. Introduction.

Population records for captive wild species have been kept since 1932, with the appearance of the first studbook for the European bison (*Bison bonanus*) (Glatston 1986), and gained a more standardised approach in 1960, when the *International Zoo Yearbook* (IZY) was first published by the Zoological Society of London (Olney 2003). In the 1970s, researchers at Minnesota Zoological Parks developed a computer-based questionnaire in which institutions could load data from their own collections². This system gave rise to the International Species Information System (ISIS), and grew beyond American borders to be used by a large percentage of the Western zoos (Flesness 2003).

² A collection is the total of individuals of the same species kept by a zoological institution; in some cases, institutions may keep separate collections for different subspecies, especially when the subspecies are subject of conservation programmes.

Meanwhile, curators and keepers with particular interest in one species or other, started compiling information from their own institutions and others that kept the same species in studbooks, making the number of species covered by these records grow exponentially through the decades. The basic format of the studbooks still follows the guidelines of the first attempts, but nowadays there is a more accurate account of details such as inbreeding levels, and electronic communication between institutions allowed for a better coverage of the captive population (Glatston 1986).

Zoo records have been used as a tool in the research on species conservation, animal breeding, behaviour and welfare since their standardisation in the 1980s (Flesness & Mace 1988). New methods of collection and analysis of data have been used in recent years to overcome some of the confounding factors found in the first years of data collection, and the reliability of data is getting higher due to standardised methods and the use of computer programmes.

In this chapter, I review the sources of information on zoo animals and their problems, and make use of analytical methods aimed to overcome these challenges. I describe the collection methods of the datasets used in the next chapters for statistical modelling, as well as the relative indices calculated by formulae to overcome statistical confounds. The aim of this chapter is to demonstrate that the records of captive populations, although not flawless or complete, can be safely used for statistical modelling as means of testing biological hypotheses when collected and analysed carefully.

2.2. Captive population databases.

The population of carnivores kept in zoological institutions and research or breeding centres is restricted, both in species and individuals. The number of individuals and collections reported to databases, however,

has been growing since the beginning of record keeping. One example is the number of collections and individuals published annually by the *International Zoo Yearbooks*: from 1986 to 1996, the number of collections has grown steadily, although the number of individuals has not followed the same slow rise throughout the years (figure 2.1). The *IZY* published the census of individuals and list of multiple births for the last time in 1996, when many studbooks and the ISIS were already collecting and organising most of institutional records. In posterior editions, the *IZY* printed just the list of studbooks and studbook keepers. The slight decline on the number of collections and individuals reported when the last full list was published may reflect the tendency of institutions to send reports directly to international data systems, such as studbooks or to the ISIS, instead of the Zoological Society of London, or may indicate the existence of copyright issues, held by commercial data systems such as ISIS, preventing the full publication of these lists for free access.

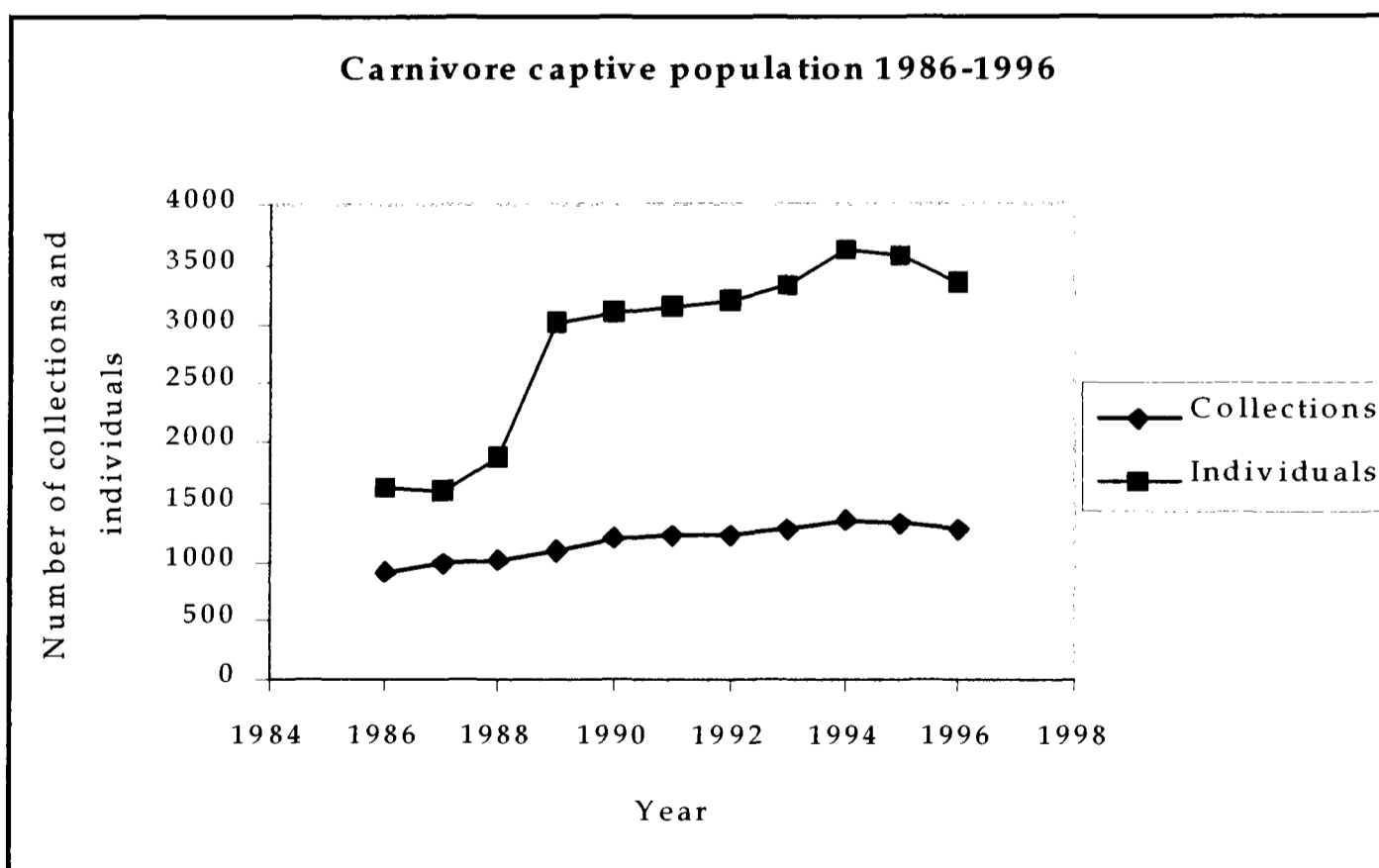


Figure 2.1: Number of collections and individuals of 35 declining Carnivora species reported to the *International Zoo Yearbooks* from 1986 to 1996. Some institutions keep subspecies in separate collections.

Nowadays, most individuals are numbered and coded, and the information from different sources is, in general, either repetitive or complementary (Glatston 1986). For example, a female in Zoo A, identified by the number 1234, may have her breeding records in the *IZY* from 1980 to 1996, and in the international studbook of the species, which also contains more detailed information of her life history, from 1984 to 2002. The decision of which repetitive data to be discarded (in this case, from 1984 to 1996) is at the discretion of the researcher. As the identity of the female is certified, complementary data, such as enclosure size and number of nesting dens, may be available on published papers and reports, or even in the institution's archives. Repetitive data can be used to crosscheck different data sources. Researchers face some challenges when collecting information, and those not attached to an institution may find their requests for data denied or subjected to copyright.

Each one of the available sources (the *IZY*, the International Tiger Studbook, zoological records collected through questionnaires and data published in papers and reports) was collected in slightly different ways, although most requested the same basic information. Institutional responses, however, may differ greatly and result in incomplete or scattered data. Overall, information on individual origins, births, deaths, enclosure characteristics and collection size will be provided by all databases. More detailed information, such as cause of death, medical treatments, transport-related issues (such as quarantine time and relocations) and aspects of husbandry (diet, number of keepers, use of contraceptives and manipulation of group size, to mention some), are rarely reported in standardised databases, but these data can be found in published reports or be requested directly to the institutions through questionnaires.

Recent efforts to keep an integrated and unique source have been proven fruitless, especially with the implementation of commercially

exploited databases, such as the ISIS. The ISIS has been trying to standardise data collection and treatment, but records are not often retroactive and older datasets are still incomplete or may be unreliable. Data organised by the ISIS have restricted access (see Section 2.2.3).

For this research, I made use of three sources or institutional records: the studbook of tigers (*Panthera tigris*); the published records on the *International Zoo Yearbooks*; and individual reports from 19 collections. Due to copyright guidelines of the International Species Information System, it was not possible to access more detailed data, apart from the annual census published in the World Wide Web (ISIS 2003).

2.2.1. Captive populations of carnivores and their implications for long term conservation.

In Chapter 1, the role of captive populations in species conservation was discussed, and some examples of successful programmes that used *ex situ* techniques were described. But captive populations are restricted, and populational researchers frequently question if these small populations are actually self-sustainable on a long-term basis. One of the possible problems is the loss of genetic variability that can affect the viability of captive populations, and therefore compromise breeding programmes.

The minimum viable size of an animal population has been the subject of great controversy in the last years. Conservation researchers disagree on the adequate values, especially when comparing research on wild and captive populations. When considering only captive populations, for example, researchers may project species persistence, which involves genetic variability, for the immediate future (e.g. Earhardt *et al.* 2001), while researchers in the wild may project species persistence for 40 or 50 generations up to into perpetuity (Reed *et al.* 2003). Surveys on numbers of rare species in captivity show that these populations are far from reaching

adequate numbers calculated for the wild (Table 2.1). In addition, a study on 13 species of carnivores suggested that the loss of genetic variability in captivity ranges from 3.1% (on the cheetah *Acinonyx jubatus*) to 22% (on the African hunting dog *Lycaon pictus*) over one generation (Earhardt *et al.* 2001). Considering that the average generation for these 13 species is 7.27 years, in only 100 years 13 generations will be produced and genetic variability may be severely reduced if not closely monitored and controlled.

Table 2.1: Minimum viable adult population size in the wild (MPVa), minimum viable adult population size in the wild corrected for 40 generations (MPVc), generation time in years (T) and population size in captivity (PC) in 2003 for five species of carnivores (Reed *et al.* 2003; Earnhardt *et al.* 2001; ISIS 2003).

Species	MVPa	MVPc	T	PC
<i>Acinonyx jubatus</i>	831	4036	5.9	1732
<i>Canis lupus</i>	1403	6332	7.05	896
<i>Lycaon pictus</i>	500	2229	-	476
<i>Panthera leo</i>	1023	5792	6.23	886
<i>Panthera tigris</i>	326	2377	7.5	253

In some species captive populations may recede, once they are not able to successfully breed in captivity and founders die of old age. For instance, the studbook keeper for the brown hyaena (*Parahyaena brunnea*) published a warning, in the 1970s, that the captive population of this declining species would disappear in the next decades (Shoemaker 1978; Shoemaker 1983). The numbers of captive individuals for the species published by the *International Zoo Yearbooks* from 1986 to 1996 seem to confirm the decline, while other large declining carnivores are apparently thriving (figure 2.2).

Captive populations can play a crucial role in conservation efforts, if genetically and demographically well managed. Institutions, on the other hand, should provide the most accurate information possible, since studbook keepers need this information to recommend breeding pairs with minimal genetic loss.

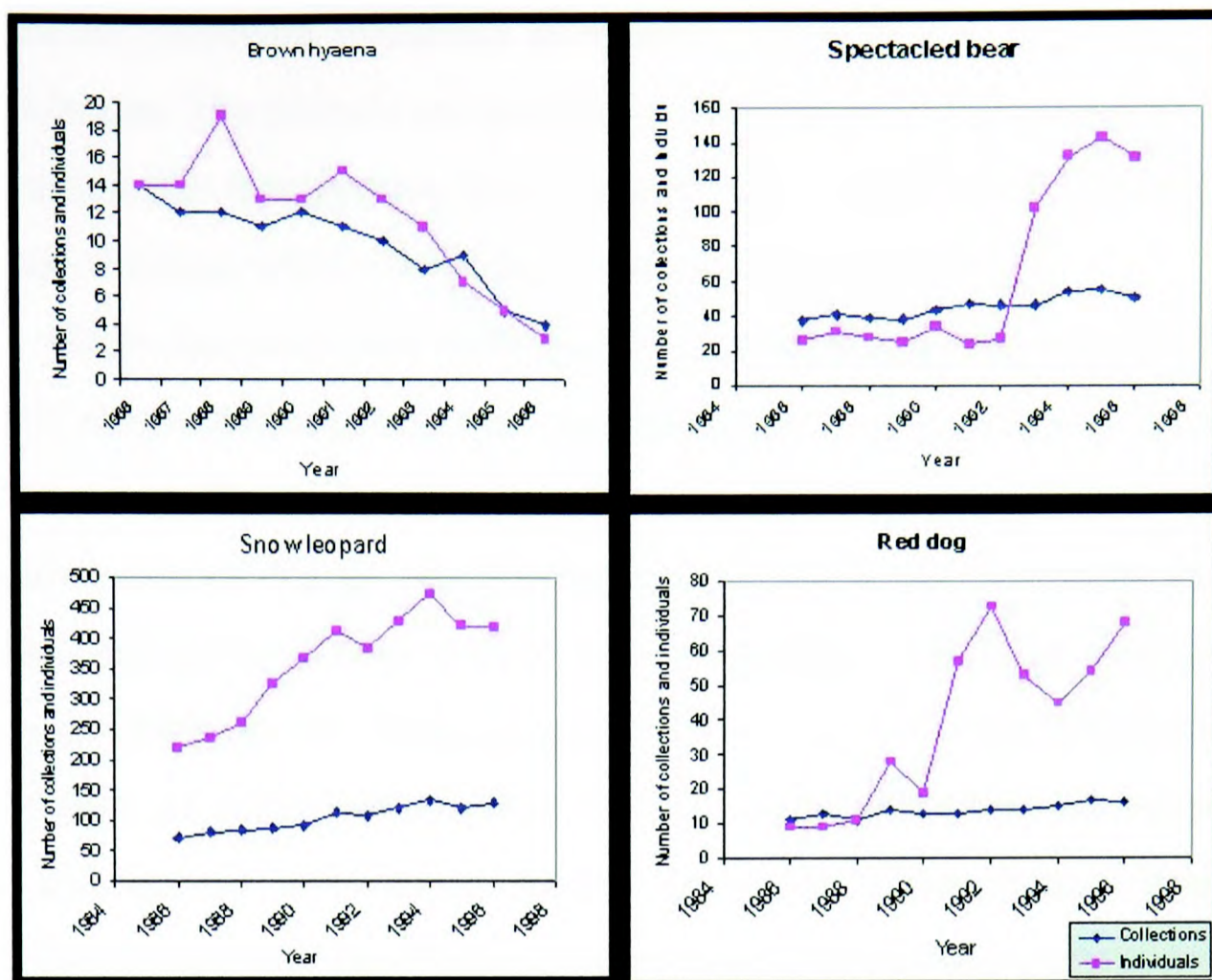


Figure 2.2: Number of collections keeping brown hyaenas (*Parahyaena brunnea*), spectacled bears (*Tremarctos ornatus*), snow leopards (*Uncia uncia*) and red dogs (*Cuon alpinus*), and total number of adult individuals reported to the *International Zoo Yearbooks* from 1986 to 1996.

2.2.2. The *International Zoo Yearbooks*.

The *International Zoo Yearbook (IZY)* was created by the Council of the Zoological Society of London in 1959, when an amount of £ 2000 was allocated to sponsor a publication aiming at zoological institutions worldwide (Olney 2003). For its first edition, Sir Zolly Zuckerman, then the Honorary Secretary of the Zoological Society, contacted personally hundreds of institutions around the world asking for papers and listings of animals and births, managing to receive information from more than 200 institutions (Morris & Jarvis 1960). From this date to 1996, the Zoological Society of London had been publishing, every year, the censuses of animals in captivity and the records of multiple births in captivity. This database today contains information from more than 600 zoos and aquaria

worldwide, covering hundreds of species from all taxa, including some invertebrates. The records are pooled by institution by year, and the census are restricted to rare species, but it contains the number of young born and number of young which died before completing 30 days of age.

When the studbooks started covering most of the species presented in the *IZY*, the publishing of listings was abolished, and now the *IZY* publishes the list of studbook keepers, for those willing to contact them. Being absolutely free of charge, any institution wishing to add its information to the database was welcome to do it, and as a result a very large collection of data is available for the years in question. The *IZY* also published detailed information on collaborating institutions, including total number of species held and annual attendance. According to the Conservation Breeding Specialist Group from the World Conservation Union, there were over 5,000 zoological institutions registered in several associations of zoos and aquaria in 1996 (www.cbsg.org), and just over 1,200 reported to the *IZY* in that year. Non-English speaking countries report in even lower proportion: from the 120 institutions registered in Brazil in 1996 (www.ibama.gov.br), only nine contacted the *IZY*.

The main advantage of the *IZY* records is the amount of data, covering most of the captive population of vertebrates and the lack of bias towards wealthier institutions; its disadvantages include the impossibility of calculating litter sizes (for the data are pooled by institution and the number of breeding females is not reported), the possible failure to report stillbirths and observational mistakes (such as the failure of noticing young that were eaten by the dam before the litter was observed). Nevertheless, the *IZY* information is still the most reliable source for the calculation of the proportion of infant mortality in zoological institutions.

2.2.2.1. *The International Zoo Yearbooks datasets.*

There were two datasets collected for this research from the *International Zoo Yearbooks*: the first contains all births and infant deaths before 30 days of age, from 1986 to 1996, for 98 species of carnivores (see Appendix 1); the second contains the characteristics of all the 535 institutions (62% of all zoos and aquaria registered in the Zoological Society of London) that provided information used to calculate the values for Appendix 1 (see Appendix 2).

The dataset on births contained information on how many young were born and died before 30 days of age every year in each collection, adding to a total of more than 9000 reported events. A proportion of mortality was calculated for each species in each zoo, and from this the median proportion of mortality was calculated for each species, as seen in Appendix 1. There is also information on the origin of the collection, if wild-caught or captive-bred, but as the information is pooled, if a collection has only one wild-caught individual, it will be listed as captive-bred. The number of breeding females in each collection was not reported.

As the database from the *IZY* is not available in electronic format, and the input of data had to be done manually, only information from the last 10 years' published listings was collected. Also, by the end of the 1980s, most members of international organisations of zoo and aquaria followed standard husbandry practices, providing a similar diet, preventive medicine and maintenance routine, and so minimising the effect of these factors in the difference of results between institutions.

The main problem of this dataset is the impossibility of calculating the number of young per female, since all births in a given collection are pooled together, independent of the number of breeding females. Also, infant deaths are only counted when happened before 30 days after birth. From this dataset, it was possible to calculate the median proportion of

infant mortality for each species that was used, together with life history information on carnivores collected by Gittleman (1983, 1986a, 1986b), to investigate biological factors that may affect breeding success in captive carnivores. Using the information presented in Appendix 2, it was possible to investigate the breeding performance of particular institutions and test the hypotheses that species from temperate areas may be affected by the latitude of the institution they are kept (cf. Chapter 4).

2.2.3. The International Species Information System.

The International Species Inventory System (ISIS) was created by Ulysses Seal and Dale Makey in 1973 as a means of standardising data collection for their research on comparative endocrinology of wild animals (Seal, Makey & Murtfeldt 1976). Its user-friendly interface and relative simplicity of data input quickly made the system interesting to other institutions, and in 1989, with its name changed to International Species Information System, it became the first worldwide collection system specifically designed for zoo records, holding the status of an independent organisation, with a Board of Trustees (Flesness 2003).

Presently, ISIS has more than 200 institutional members worldwide, but up to 90% of its members come from North America or Western Europe (figure 2.3). Its contents, however, are only fully available to its members, who are charged high prices to join. In 2003 these values range from US\$ 955 to US\$ 6355 for institutions (depending on the annual attendance) and US\$ 955 to researchers. Independent conservation researchers (such as members of Taxon Advisory Groups or Conservation Specialist Groups) and studbook keepers are granted free access for data concerning only one species. ISIS and most studbooks are interconnected, and access to unpublished information may be subjected to the same copyright laws.

Poorer institutions can opt for an alternative form of payment, called General Operating Budget. To use this option, the institution has to calculate the general operating budget, including donated services from the government; the fee will be 0.1% of the budget of the institution to a minimum of US\$ 440.

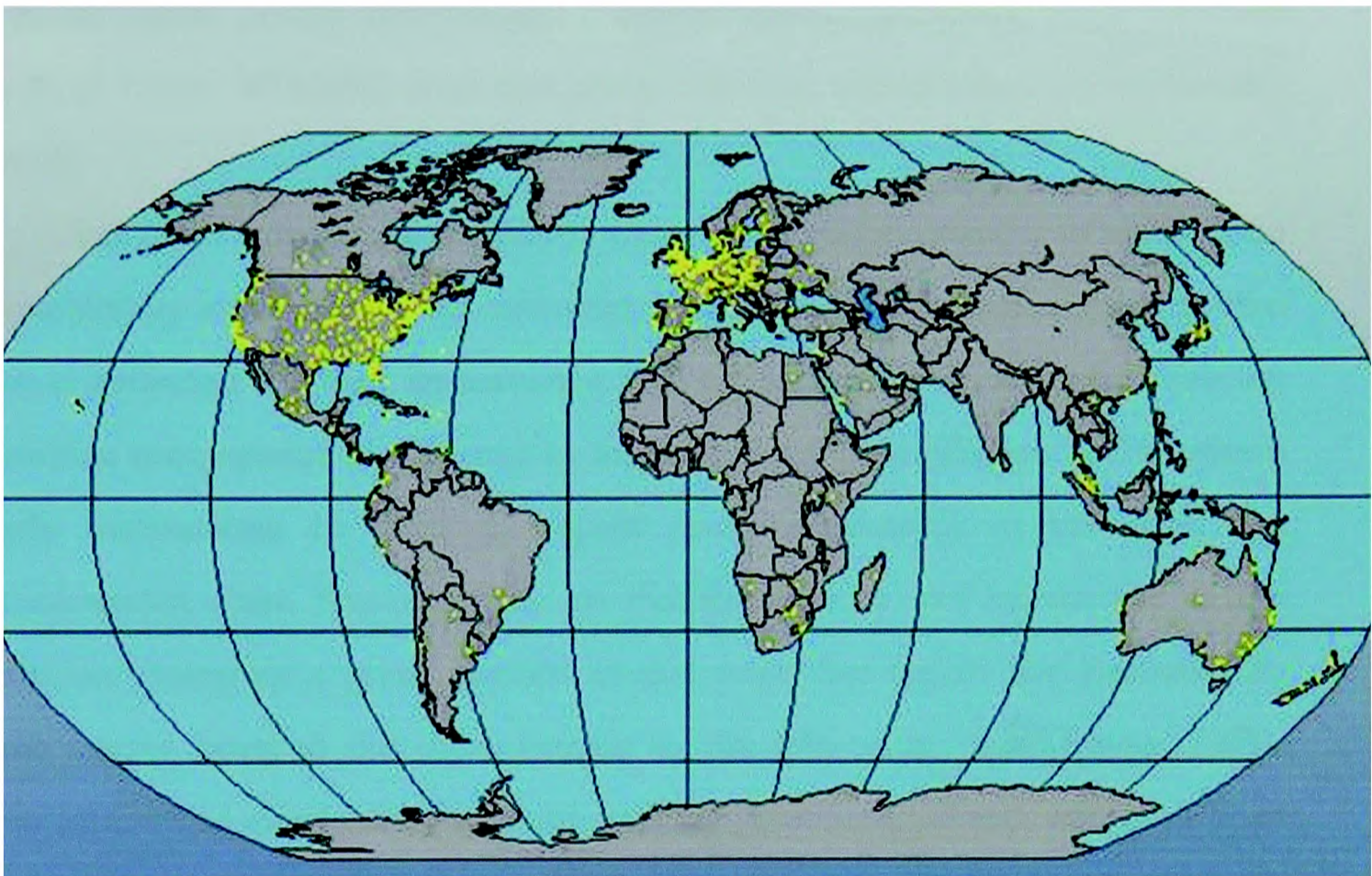


Figure 2.3: Distribution of institutional members of the International Species Information System (ISIS) in 2004, from www.isis.org.

Membership to ISIS includes a package of computer programmes that allow institutions to keep records and upload them online to the ISIS website (www.isis.org). These programmes were developed for the specific needs of record keeping by institutions. ARKS (Animal Records Keeping System) is the simplest of programmes and is suitable for maintaining simple specimen records; MedARKS (Medical Animal Records Keeping System) offers several options to keep veterinary records of collections; and SPARKS (Single Population Animal Records Keeping System) was

developed specific for veterinary records, studbooks and Species Survival Plans, and have special features that allow genetic and demographic analyses (Flesness 2003). SPARKS is provided free for studbook keepers, and is the only ISIS computer programme that can be purchased by non-members. SPARKS provides a simple touch-button form which has preset options for some items, while allowing the input of original data for aspects such as name of the individual, parental identification or more detailed medical notes. SPARKS does not cover housing information or husbandry details.

The great advantage of ISIS is allowing a large amount of data to be immediately available for its members, and the homogeneity of information that is collected through its software packaging. Its disadvantage, however, involves the general bias found in zoological records (Rieger 1979), which leads institutions to tend to report more successful events than the unsuccessful ones. For example, an institution may not report the single stillborn young of a given female in one year, but report the successfully risen young born to the same female in the following year (Rieger 1979). This procedure can lead to mistakenly high breeding success levels to both the female and the institutions. Also, because of the large amount of options offered by its softwares, such as SPARKS, zoo staff tends to leave some of these options in their default positions. For example, SPARKS has an option to describe the cause of death of individuals, which is held in the default answer "by unknown means". Analysis of the tiger information from the studbook kept by Sarah Christie at London Zoo revealed that 88.16% of the events were described as this category.

The expensive membership fees of ISIS automatically bias its information as coming from wealthy institutions, hence the majority of members being located in North America and Western Europe. Traditional institutions, such as the Zoo Negara in Kuala Lumpur, Malaysia, which have been sending records for free to the Zoological Society of London,

cannot spare resources for the fees and is out of the database, even when they keep and breed successfully some declining species, such as the Malayan sun bear (*Helarctos malayanus*) and the Persian lion (*Panthera leo persica*), among others.

For this research, ISIS was approached, through London Zoo, to give permission for the use of its records for tigers (*Panthera tigris*). Unfortunately, the access was denied because there was no connection between this research and any official conservation group at the time. However, the tiger studbook keeper, Ms. Sarah Christie (in 2000), provided her information for this research. The specific problems of this dataset will be discussed on section 2.2.4.1.

2.2.4. International studbooks.

The studbook system was created in the end of the 18th century, to keep breeding records of domestic stock, and started to be applied for wild species in 1932 (Glatston 1986). For some species of the order Carnivora, however, studbooks were not created until much later. For example, the first studbooks for the maned wolf (*Chrysocyon brachyurus*), bush dog (*Speothos venaticus*) and spectacled bear (*Tremarctos ornatus*) were published simultaneously in 1975, after their first appearance in the *IUCN Red Data Book* of 1972 (Roeben 1975).

Studbooks are primarily used, for captive populations of wild species, as the source of data for genetic and demographic management, while other sources, such as ISIS and surveys, are used for regional collection planning. ISIS data were once considered unsuitable for genetic management due to significant discrepancies with more detailed studbooks (Earhardt *et al.* 1995). The reply stated that studbooks also contained errors, especially on parental identity, but agreed that the raw data from ISIS needed thorough examination before analysis (Flesness *et al.* 1995).

The obvious suggestion for integrating both studbooks and ISIS in a single source was first made in 1986 (Glatston 1986) and was recently reiterated by an appeal to avoid duplication of data collection. The author points out, however, that before this can happen, ISIS has the challenge of becoming a more “open” system, allowing wider access to researchers and facilitating the subscription by the 500 institutions worldwide that are still not members (Flesness 2003).

In any case, studbooks are very useful tools in animal management, and the quality of data is improved by the personal involvement of the studbook keeper with the information. Once one institution sends a record to a studbook, the keeper will annually remind, sometimes three or four times, for the institution to keep sending records (Glatston 1986).

2.2.4.1. *The Tiger Studbook dataset.*

The dataset extracted from the electronic tiger studbook kept by Sarah Christie at the Zoological Society of London, was collected using the software SPARKS, from the International Species Information System (ISIS). It contains all births and infant deaths in 116 institutions mainly in the Northern hemisphere, during the period from 1986 to 1996. The records refer to two subspecies: the Sumatran (*P. tigris sumatrae*) and the amur or Siberian tiger (*P. tigris altaica*).

The dataset contains 249 litters born from 126 females (28 Sumatran and 98 Siberian), and there is additional detailed information for 43 females. There are data on the number of young born, date of birth, offspring that died up to 6 months old, date of death, litter size, litter rearing method (if reared by parents or by hand), inbreeding level and parental identity. There was no information on enclosure area or housing conditions (see Appendix 3). The cause of death of young is reported in less than 12% of the events. Studbook information is, as long as possible, checked by the studbook

keeper. Mrs. Sarah Christie pointed out that older data might present some problems such as unreported litters, especially stillbirths. The identity of the female was reassured whenever possible. In any case, the data from the tiger studbook represent a large sample of the captive population, and the values calculated from it, such as proportion of infant mortality, litter sizes and peak of births, were used to investigate reproductive parameters of tigers in captivity, check subspecific breeding differences and test the hypotheses that inbreeding levels do not affect breeding success in captive tigers.

2.2.5. *Published records.*

Since the first publication of the *International Zoo Yearbooks*, curators of collections have been publishing small records and notes on the breeding and rearing of wild animals in captivity. For decades, these reports were purely descriptive of the events, and usually contain detailed information on several aspects of husbandry. Many of these reports give details not only on parental identity, date of birth and cause of death of young, but also on the housing conditions, diet, cleaning routines and isolation protocols. All the basic information found in studbooks is usually described, and when the young were removed for hand-rearing, also the techniques used and results obtained with the procedure.

Published records were used successfully before to produce a dataset for the research on stereotypic behaviour in captive carnivores, yielding significant results (Clubb 2001). The level of detail of the reproductive data presented in published papers and notes is higher than in the other, more formal, databases. Nevertheless, it is not possible to use this type of dataset to produce a general view of the proportion of infant mortality, since there is a strong bias towards successful reproduction on these records.

2.2.5.1. *The bibliographical dataset.*

Following the methodology described by Clubb (2001), several bibliographical searches were performed using electronic resources available at the University of Oxford, such as the Web of Science and the Oxford Electronic Reference Library (ERL) databases. The results were scrutinised for descriptions of the births of individual litters. In addition, all volumes of the *International Zoo Yearbooks* and the periodical *Zoo Biology*, where these reports are common, were examined. Papers that present pooled values for several litters or that did not clarify parental identity and date of births were not included in the dataset. This dataset differs from that compiled by Clubb (2001), which contains papers describing stereotypical behaviour in captive carnivores rather than breeding events. The dataset contains information from 141 papers and notes on captive breeding of carnivores, and presents data on: litter size; date of birth and death of young; parental age, origin and rearing method; area of enclosure; number of dens; area of dens (when available); cause of death of young; and isolation protocol for pregnant females (see Appendix 4). It also contains data on removal for hand-rearing and the cause for it. The dataset comprises 69 species, and has information on 447 litters born from 212 females between 1961 and 2000. There was no information on female body weight. Table 2.2 summarises the bibliographical data presented in Appendix 4.

Table 2.2: Species, number of litters, number of zoological institutions keeping the species and number of females summarised from the data compiled from bibliographical sources.

Family	Species	Number of litters	Number of zoos	Number of females
Canidae	<i>Alopex lagopus</i>	2	1	2
Canidae	<i>Canis familiaris</i>	1	1	1
Canidae	<i>Canis latrans</i>	2	1	1
Canidae	<i>Canis lupus</i>	9	3	3
Canidae	<i>Cerdocyon thous</i>	6	2	2
Canidae	<i>Chrysocyon brachyurus</i>	20	9	13
Canidae	<i>Cuon alpinus</i>	4	2	4
Canidae	<i>Dusicyon vetulus</i>	1	1	1
Canidae	<i>Lycan pictus</i>	4	2	2
Canidae	<i>Otocyon megalotis</i>	1	1	1
Canidae	<i>Speothos venaticus</i>	5	3	3
Canidae	<i>Vulpes corsac</i>	5	1	2
Canidae	<i>Vulpes zerda</i>	7	3	4
Felidae	<i>Acinonyx jubatus</i>	22	13	16
Felidae	<i>Caracal caracal</i>	5	3	4
Felidae	<i>Catopuma temmincki</i>	22	2	5
Felidae	<i>Felis bengalensis</i>	8	2	2
Felidae	<i>Felis margarita</i>	10	2	2
Felidae	<i>Felis nigripes</i>	8	2	3
Felidae	<i>Felis pardalis</i>	1	1	1
Felidae	<i>Felis silvestris</i>	29	4	8
Felidae	<i>Herpailurus yaguarondi</i>	4	2	2
Felidae	<i>Leopardus geoffroyi</i>	15	4	6
Felidae	<i>Leopardus tigrinus</i>	7	2	3
Felidae	<i>Leopardus wiedii</i>	3	2	2
Felidae	<i>Leptailurus serval</i>	2	1	1
Felidae	<i>Lynx lynx</i>	10	3	7
Felidae	<i>Neofelis nebulosa</i>	15	4	5
Felidae	<i>Panthera onca</i>	4	2	3
Felidae	<i>Panthera pardus</i>	1	1	1
Felidae	<i>Panthera tigris</i>	9	3	3
Felidae	<i>Prionailurus viverrinus</i>	4	2	2
Felidae	<i>Uncia uncia</i>	11	4	5
Herpestidae	<i>Atilax paludinosus</i>	2	1	1
Herpestidae	<i>Cryptoprocta ferox</i>	1	1	1
Herpestidae	<i>Galidia elegans</i>	3	1	1
Herpestidae	<i>Helogale parvula</i>	14	2	3
Hyaenidae	<i>Crocuta crocuta</i>	4	3	3
Hyaenidae	<i>Parahyaena brunnea</i>	3	1	1
Mustelidae	<i>Amblonyx cinerea</i>	11	3	3
Mustelidae	<i>Arctonyx collaris</i>	1	1	1
Mustelidae	<i>Eira barbara</i>	3	2	2
Mustelidae	<i>Enhydra lutris</i>	9	4	5
Mustelidae	<i>Gulo gulo</i>	1	1	1
Mustelidae	<i>Lutra canadensis</i>	3	2	3
Mustelidae	<i>Lutra lutra</i>	7	2	4
Mustelidae	<i>Lutra perspicillata</i>	8	3	7
Mustelidae	<i>Meles meles</i>	2	1	1
Mustelidae	<i>Mellivora capensis</i>	2	1	2

Family	Species	Number of litters	Number of zoos	Number of females
Mustelidae	<i>Mustela nigripes</i>	2	1	1
Mustelidae	<i>Pteronura brasiliensis</i>	12	1	2
Procyonidae	<i>Ailurus fulgens</i>	9	3	4
Procyonidae	<i>Nasua narica</i>	4	1	4
Procyonidae	<i>Nasua nasua</i>	2	1	2
Procyonidae	<i>Potos flavus</i>	6	3	3
Ursidae	<i>Airulopoda melanoleuca</i>	11	6	7
Ursidae	<i>Helarctos malayanus</i>	12	3	5
Ursidae	<i>Tremarctos ornatus</i>	11	4	4
Ursidae	<i>Ursus arctos</i>	2	2	2
Ursidae	<i>Ursus maritimus</i>	11	4	4
Viverridae	<i>Arctictis binturong</i>	13	5	5
Viverridae	<i>Arctogalidia trivirgata</i>	1	1	1
Viverridae	<i>Fossa fossa</i>	1	1	1
Viverridae	<i>Genetta genetta</i>	1	1	1
Viverridae	<i>Hemigalus derbyanus</i>	1	1	1
Viverridae	<i>Paradoxurus hermaphroditus</i>	4	1	1
Viverridae	<i>Prionodon linsang</i>	1	1	1
Viverridae	<i>Viverra civetta</i>	9	1	3
Viverridae	<i>Viverra zibetha</i>	2	1	1

The main problem with this dataset is the lack of standards in data collection and presentation. As most of the reports follow the author's discretion, some papers will contain more detailed information than others, and the dataset will not present all variables for all species. Nevertheless, many of these papers contain data that is not available in any other source, such as detailed maternal behaviour, cause of death of the young and some husbandry protocols. The bibliographical dataset was used to investigate the causes of death of young captive carnivores and test the hypotheses that some biological factors, such as altriciality, may influence the occurrence of maternal infanticide.

2.2.6. Zoological institution records.

Zoological institutions keep records for all their specimens, including breeding notes that are fundamental for the management of collections. Recent records are usually readily available to the registrars of the collections, once the notes on many individuals that have been in exhibition for a long time may be needed for medical purposes. Most institutions,

nowadays, are transferring the data from record books to computer programmes such as SPARKS, sometimes to the detriment of some detail of information.

Traditionally, institutions keep records in a diary format, recording every single event on the life of the specimen, from vaccinations and ailments to notes on unusual behaviours (such as stereotypical pacing, excessive aggressiveness towards the keeper or changes in appetite). Female oestrous and mating are recorded when observed, as are the births and deaths of young. The animal keepers have the responsibility of keeping these records, and standardisation of the data varies greatly among institutions. Some data are fundamental and can be found, in different formats, in all institutional records, and provides a more detailed view of the conditions in which the animals are kept.

2.2.6.1. Dataset collected through questionnaires.

For this present research, 135 zoological institutions worldwide, corresponding to over 15% of zoos and aquaria registered at the Zoological Society of London, were contacted and invited to complete an electronic questionnaire. The questionnaire, a Microsoft Excel Workbook, asked for information in studbook format, with parental identification, date of birth, litter size, number of young dead, date and cause of death, and method of rearing, i.e. if the young were mother- or hand-reared (see sample in Appendix 5). Additional information on enclosure size, number of dens per enclosure, number of keepers and age, and origin of the breeding individuals was requested. As required by the institutions prior to sending the information, the identity of the females and the institutions had to be preserved and could not be printed in the dataset, being replaced by codes (Appendix 6).

To make this dataset congruent with the others in this research, and to facilitate the collection of data by the collaborating institutions, only the data from 1986 to 1996 were requested.

This dataset is restricted to nine species: red panda *Ailurus fulgens*, meerkat *Suricata suricatta*, tiger *Panthera tigris*, snow leopard *Uncia uncia*, wolf *Canis lupus*, oriental small-clawed otter *Amblonyx cinereus*, brown-nosed coati *Nasua nasua*, polar bear *Thalarctos maritimus* and maned wolf *Chrysocyon brachyurus*. The choice of species tried to reflect the diversity of habits and habitats of the Order Carnivora, and was also based in the possibility of acquiring information on the largest possible number of females, which is more probable in species with larger captive populations.

Unfortunately, only 16 institutions, comprising 31 collections, answered the request for filling the questionnaires, a response rate of less than 12%. Another 6 institutions agreed to participate but, to this date, did not return the questionnaires despite occasional reminders. Only two institutions declined participation due to lack of staff or management transition.

The dataset from the 31 collections contains 148 litters from 63 females of nine species. The cause of death of the young was reported in 61.5% of the cases.

The main drawback in this dataset was the very low level of response from the institutions, which made this dataset the smallest collected. Also, the complexity of the data discouraged the appropriate filling of the questionnaire, due to the need of reading protocol books frequently stored away in the institutions archives and the high time demand. A larger dataset, apparently, would need to be collected personally in each institution. Nevertheless, reliable information on the number of keepers, cage area, number of dens and distance travelled by the female is available in this dataset, allowing the hypotheses that husbandry protocols have an effect on the breeding success of captive carnivores to be tested.

2.2.7. Summary of data.

To avoid data duplication, the variables extracted from different sources were not the same, in a way to complement the information required for analysis. Also, complementary information was taken from other sources, such as range of latitude (Wilson & Reeder 1993), conservation level (IUCN 2002b) and biological information (Gittleman 1983, 1986a, 1986b). A summary of the variables extracted from each source and the level of duplication can be seen on Table 2.3.

For the analysis in Chapters 4, 5 and 6, some variables regarding the biology of the species were used. From Gittleman (1986a, 1986b), the variables were: gestation length; weaning age; age when the young open their eyes; female body weight; species body length; type of zonation (e.g. terrestrial, arboreal, aquatic); type of vegetation (e.g. open grassland, forest, desert); and type of diet (e.g. carnivorous, omnivorous, vegetarian). The data on delayed implantation comes from Mead (1989), and the information on juvenile mortality in the wild was presented by Gittleman (1983) in his unpublished doctoral thesis.

Table 2.3: Summary of the data used in this research, the variables extracted from each one, the sources of data, the chapters in which they were used, and the level of duplication found between datasets.

Dataset)	Variables (sources)	Duplication	Chapter
Appendix 1 (98 species)	Proportion of infant mortality, number of zoos holding the species (<i>IZY</i> vols. 26-35); conservation level (IUCN 2002b); range of latitude distribution (Wilson & Reeder 1993)	Basic dataset; encompasses the largest amount of data.	4 & 7
Appendix 2 (535 zoological institutions)	Number and type of species held, number of veterinarians and staff; presence of research staff; size; annual attendance; type of management (<i>IZY</i> vols. 30-35); latitude (calculated with software).	Complementary to the data on Appendix 1. The index of zoo performance (see section 2.3.3) was calculated using the proportion of infant mortality from Appendix 1.	4 & 7
Appendix 3 (1 species; 249 litters; 126 females)	Zoo; date of birth of litters; sire and dam; place of birth, rearing, number of transfers and age of dam; number and sex of young born and dead; age and cause of death of young; type of litter rearing; inbreeding level of the young (Mrs. Sarah Christie, International Tiger Studbook Keeper)	Over 90% of the births reported in the studbook are also reported on the <i>IZY</i> , although the studbook reports litters separately. From this dataset was possible to calculate female breeding success instead of infant mortality.	3
Appendix 4 (69 species; 447 litters; 212 females)	Zoo; origin, rearing and age of dam; litter size; number of dead young; cause and age of death of young; few present cage area, number of individuals per cage and number of dens; presence of male (141 papers published in several scientific journals from 1961 to 2000)	Some of the written reports are accounted for in the <i>IZY</i> and studbook datasets. The analysis was done exclusively with variables from this dataset, and the proportion of infant mortality was calculated for each female.	5 and 6
Appendix 6 (9 species; 148 litters; 63 females)	Age of sire; age, origin and rearing of dam; distance travelled by the dam; litter size; number and age of dead young; few present cause of death of young; number of keepers; few present cage area and number of dens (electronic questionnaires sent to institutions)	All the litters were accounted for in the <i>IZY</i> dataset, but it was possible to calculate female breeding success. Bonferroni corrections were applied when necessary.	7

2.3. Methods of collection and analysis of zoological records.

As seen in the previous sections, animal records can present many discrepancies between different sources. This is understandable since different databases have information from different groups of collections (for example, there are 535 institutions in the *IZY* dataset, but only over 200 in the *ISIS* census for 2003), although subgroups of data may overlap. They can provide, nevertheless, large samples of captive populations and can be crosschecked, in a way of verifying data reliability. For example, the overlapping information between *IZY* and the tiger studbook can be checked for the total of animals born every year or number of animals held. During the compilation of the datasets used in this research, discrepant data, comprising less than 1% of the total, were discarded. Also, pooled reports in the annual *IZY Records of Multiple Births* (represented by (b), to signify that a litter of unknown size died immediately after birth and was consumed by the dam) and incomplete data (e.g. when the number of dead animals was not printed by mistake) did not enter in the final datasets. This method of “cleaning” the data helps to minimise errors and is often used when preparing demographical datasets for human populations (Chatfield 1991). The input of data into electronic datasets must be meticulously made, and simple sums of columns and rows suffice to avoid typing errors.

There is not a standard way to treat animal records. Frequently these records are used on the research of inbreeding effects (e.g. Laikre *et al.* 1996 on brown bears *Ursus arctos*, and Wielebnowski 1996 on cheetahs *Acinonyx jubatus*), but were also used to calculate the effect of housing (Carlstead *et al.* 1999), individual behaviour (Carlstead, Mellen & Kleiman 1999) and rearing experience (Ryan *et al.* 2002) on breeding success, with very significant results. Care must be taken, however, on the multiple analysis of the datasets to assure that they represent a significant range of variance on the populations before any conclusions are made.

2.3.1. *The bias of data: adjusting the numbers.*

Zoological records tend to be biased for institutions that perform research and have breeding pairs, and perhaps even for those who managed to successfully produce young, once unsuccessful reproductions are, sometimes, not reported to international record keepers (Rieger 1979). Also, the cryptic nature of many species, together with the design of some enclosures, does not allow keepers to be aware of all births. Many females in captivity are overweight, and pregnancy can pass unnoticed, as it has happened with the oriental small-clawed otter *Amblonyx cinerea* (Leslie 1970).

In this way, these samples from animal databases are likely to represent observed births in wealthy zoos that are successful in breeding carnivores. The low participation of zoological institutions of poorer countries, as seen in the case of Brazilian zoos, is an example of this bias. Still, these institutions have very different characteristics that can be tested in the search for the factors that affect infant survival in captive carnivores.

2.3.2. *Record research and sex ratio.*

Zoological databases for captive populations could be excellent sources for the research on sex ratio of many species. In this research, however, it was found that this type of data was not reliable in the datasets analysed.

Many birth reports contain young of undetermined sex, which were observed but could not be handled up to the time of publication. Researchers discard litters that contain unsexed individuals (Faust & Thompson 2000), but it has been pointed out that there are occasions where the mother consumes some young of a large litter right after birth. This could easily pass unnoticed by keepers, and the reported litter would be included in the analysis (Rieger 1979; Christie, pers. comm.). In the datasets

used in this research, the proportion of young of undetermined gender varies greatly (Table 2.4). Within the dataset collected from the *IZY*, the variation is also high between families, being the proportion of unsexed young higher in the families Procyonidae, Mustelidae and Herpestidae, which contain small species frequently housed in groups. In species with communal rearing of the young, common in these families, manipulation of the young is discouraged and it is difficult to identify the gender before 30 days of age.

Table 2.4: Total number of young born, young of undetermined sex and proportion of young of undetermined sex in the several datasets used in this research.

Dataset	Taxa	Number of young born	Young of undetermined sex	Proportion of young of undetermined sex
Bibliographical dataset	69 species	1144	513	44,84%
Tiger studbook	<i>Panthera tigris</i>	670	70	10,44%
Census of multiple births from the <i>IZY</i> vols. 26-35	Felidae	11595	2282	19,68%
	Ursidae	2083	568	27,26%
	Viverridae	788	246	31,22%
	Canidae	7891	2474	31,35%
	Hyaenidae	406	138	33,99%
	Procyonidae	3605	1278	35,45%
	Mustelidae	4067	1735	42,66%
	Herpestidae	3372	2088	61,92%

Although the proportion of unsexed young is relatively low in the tiger studbook, the studbook keeper, Mrs. Sarah Christie, pointed out that this data is not reliable and she was still in the process of confirming the gender of the surviving individuals. The questionnaires sent to the zoological institutions asked for this information, but only one of the institutions provided this data.

2.3.3. *Relative indices as dependent variables.*

In a zoological institution there are many factors that may have potential effects on the behavioural and physiological condition of the animals. Because of this, it is appropriate to calculate relative indices that consider the possible husbandry differences between institutions, allowing the effect of other factors to be analysed. The following equations were developed for this research and are tailored to overcome specific problems of the datasets collected and analysed here.

The basic measurement on the research of infant survival is the proportion of young that died before 30 days of age, as published in the database from IZY. For the species, infant mortality will be the median of the proportion of mortality of each institution or female (equation 2.1), once there could be an effect of husbandry techniques or individual differences.

$$I = \text{median}[(\delta / \beta)_{\alpha\eta} \dots (\delta / \beta)_{\alpha\infty}]$$

Equation 2.1: Infant mortality in a species (I), where δ = total number of young dead ; β = number of young born in the in the same period of time; and $\alpha\eta \dots \alpha\infty$ = each one of the collections or females in the database.

Breeding success can be measured in many different ways. Some authors calculate breeding success including unsuccessful mating attempts, in order to assess male energy expenditure (Ilukha, Harri & Rekila 1997), and others consider infant survival a good measurement of successful breeding (Durant 1999).

For this research, females of the same collections were not considered independent, due to husbandry practices. To overcome this problem, an index of infant survival, or breeding performance, was calculated (equation 2.2). The result balances the possible confounding effect of husbandry when testing the effect of biological factors.

$$B = \frac{\beta - \delta}{\psi * \phi}$$

Equation 2.2: Female breeding performance (B), where β = total number of young born in a given period of time in an institution; δ = total number of young dead in the same period; ψ = number of years of the given period of time and ϕ = number of breeding females in that institution for the same period of time.

To understand if institutional characteristics are affecting infant survival, and taking into consideration that species have different I values, the institutional performance (equation 2.3) will give an overview of how the institutions are with all species of carnivores, of local range or imported, independent of the environmental factors, such as photoperiod, that could affect breeding in foreign species.

$$Z = \sigma / \zeta$$

Equation 2.3: Institutional performance (Z), where σ = Number of species in an institution with higher infant mortality than the value of I for the species; and ζ = total number of Carnivora species held in that institution.

The use of relative indexes to overcome statistical pitfalls such as pseudoreplication (Hurlbert 1984) can help focus analytical efforts in measuring the effect of known factors.

2.3.4 Testing for phylogenetic effects.

One of the major problems when working with several species is the possibility of phylogenetic effects in the results. Until the end of the 1970s, researchers usually took a straightforward approach when dealing with traits of several species, regardless of taxonomic levels, which led to erroneous generalisations (Harvey & Pagel 1991). A classic example is presented by

Harvey & Clutton-Brock (1985): a study presented results for mammals when 82% of the species used in the analysis were rodents, which made the results valid only for this taxon rather than all mammalian species. Ignoring phylogenetic effects in comparative studies can lead to type I and II errors, and if the data reflects a structure phylogeny, with little independence, the results can be misleading (Gittleman & Kot 1990).

To avoid these problems it is paramount to establish the independence of data and examine which percentage of the variance in the data is accounted for at different taxonomic levels (Gittleman & Luh 1992). There are several methods for checking this relation, including nested analysis of variance (Harvey & Clutton-Brock 1985) and autocorrelation statistics such as Moran's *I* (Gittleman & Luh 1992).

In this research, some comparative analysis was used in the search of whether biological aspects affected reproductive success in captive carnivores. A problem that rises in the data collected from zoological institutions is that it is not possible to cover the majority of major taxa, once not all species are kept in captivity. There are, however, data on other families of Carnivora, although not as abundant as Felidae. However, analyses of certain aspects of breeding in close taxa do not imply that these results are unimportant or invalid; care must be taken not to generalise results in a varied group such as the carnivores, but these results may point out patterns within families or, in special cases, within a species.

Table 2.5 presents a nested analysis of variance including the biological variables used in statistical models in this research. Most of these variables show independence from the genus level or below, and as phylogenetic methods reflect a more extreme form of analysis (Gittleman & Luh 1992), it was decided that a straightforward method would be adequate in this case.

Table 2.5: The percentage of variance in the data accounted for at successive taxonomic levels by each variable used for testing hypotheses. The results are based on a two-level nested ANOVA with unequal sample sizes. Apart from the age of young opening their eyes, most variance resides below the level of genus, and the response Proportion of Infant Mortality in Captivity is mostly specific.

Taxonomic level:	Family	Genus	Species	Genus and below
Variable:				
Gestation length	0	79.67	20.33	100
Proportion of infant mortality in captivity	0.58	71.97	27.45	99.42
Weaning age	30.21	0	69.79	69.79
Female weight	45.35	0	54.65	54.65
Opening eyes	67.03	27.44	5.52	32.96

It is important to point out that the results in this research shall not be generalised for all Carnivora species. For each test, the taxa more likely to behave as the results point out will be specified.

2.4. Conclusion

Captive population records have been helping to understand specific demands for better management practices, that can affect breeding success, genetic variability and, ultimately, the long term survival of captive populations. Conservation efforts should, before spending too many resources in *ex situ* programmes, consider the actual possibilities of captive populations with the help of animals databases. Research would benefit, however, from the creation of a single system of data management, incorporating records from studbooks, ISIS and detailed information from the collections, but also from a better, wider access to the databases already available.

At present, the reliability of the data is still low, but there are precautions that can be taken to minimise the error and allow relevant analysis to be performed.

This research is based in a very small sample of the total information available, for thousands of species, in the databases described in this chapter. With the technological advancements rapidly being adopted by institutions all over the world, data quality and quantity will rise. Even today, these resources must be better explored, because captive populations can decline within a decade. Conservation efforts involving captive populations are very recent, and unless the real status of these populations is known now, some species may not have the genetic backup to start new populations in the near future.

Chapter 3

Factors affecting breeding in captive tigers (*Panthera tigris*): A studbook research

Abstract

Data from the International Tiger Studbook was analysed, comprising 249 litters born from 126 females of Siberian (*Panthera tigris altaica*) and Sumatran tigers (*P. tigris sumatrae*) from 116 institutions, mostly of the Northern hemisphere. Births peaked in the month of May; the average litter size was 2.72 cubs per litter, and the average age of breeding females was 6.2 years. Female age did not have a statistically significant effect over litter sizes or proportion of infant mortality. In average, young died before the end of the third week. The median proportion of infant mortality in this dataset was 68%. There was no significant difference between Siberian and Sumatran subspecies in the number of litters produced by each female, in the interbirth interval or in the female's average number of litters per reproductive year. Also, there was not a subspecific difference in the proportion of infant mortality and heterozygosity. Sumatran tigers produced smaller litters and less young in 10 years. Sumatran cubs died significantly earlier than Siberian cubs. In both subspecies, litter size had an effect on infant mortality, and infant mortality was higher in litters born in the autumn. Institutions with a history of poor breeding of other Carnivora species also performed poorly with tiger breeding, but there was no effect of ongoing research on infant mortality. There were no detectable effects of inbreeding or female origin in infant mortality, although a larger sample size would be required.

3.1. Introduction.

Tigers (*Panthera tigris*) comprise the largest captive population of all declining species of the order Carnivora. Today, with over 1,000 individuals, including subspecific hybrids, the species is held in more than 400 collections (ISIS 2003). All five persistent subspecies (Siberian or Amur tiger *P. t. altaica*, Amoy tiger *P. t. amoyensis*, Indo-Chinese tiger *P. t. corbetti*, Sumatran tiger *P. t. sumatrae* and Bengal tiger *P. t. tigris*) are kept in separated collections and supervised by studbooks (Olney & Fischen 2003).

In this chapter, studbook data from two subspecies of tiger, *P. t. altaica* and *P. t. sumatrae*, were compiled and analysed, so as to compare

reproduction parameters in the captive population with wild populations. Also, the effect of certain aspects of the biology of the species and husbandry conditions in the reproductive success of females was tested through statistical models (see section 3.4). The results reaffirm the importance of record research in the management of captive populations and may help the development of more effective husbandry protocols.

3.2. Tiger status and conservation in the wild.

According to the World Conservation Union (IUCN), the species as a whole is endangered, with an observed continuous decline through loss of habitat and poaching, and do not possess any subpopulation with more than 250 mature individuals. Three subspecies (Siberian, Amoy and Sumatran tigers) are singled out as critically endangered, meaning that they are at extreme risk of extinction. In the case of the Siberian tigers, the population of mature individuals was estimated at less than 250, and at less than 50 individuals for the Amoy tiger (IUCN 2002). During the 20th century, three subspecies became extinct: the Bali tiger *Panthera tigris balica* in the 1940s, the Caspian tiger *P. t. virgata* in the 1970s, and the Javan tiger *P. t. sondaica*, as recently as the 1980s (Jackson 1998).

International conservation efforts started in 1994, when the Global Tiger Forum, a conference from 11 countries where the tiger occurs, took place in India. However, since the 1970s, smaller regional programmes started warning about the rapid decline of the species and campaigns were set up against the use of tiger parts, to try to stop commercial poaching (Weber & Rabinowitz 1996).

The maintenance of a commercially exploited species in poor countries faces many problems. For example, the profits of poaching tigers are very high, especially for the people living in poor conditions and without an economically viable alternative to support themselves (Saberwal 1996). Poaching activities on tiger populations peaked in the beginning of the 1990s, when the demand for tiger parts grew due to an increase in the use of traditional Chinese medicine, and only stopped growing after intense governmental intervention (Karanth & Madhusudan 1997). In 1995, researchers predicted, through mathematical models, total extinction of the species in the wild in just over one decade, if poaching was not drastically reduced (Kenney *et al.* 1995).

Human populations usually surround natural reserves, and cattle are frequently raised within the range of the tiger population. Cattle predation by tigers is an economical problem in rural areas in China (Zhang *et al.* 2002), Bhutan (Dorji & Santiapillai 1989) and India (Veeramani, Jayson & Easa 1996). Human-tiger conflicts over space due to the growing human population and the development of lands have been reported in Russia (Tkachenko 1997) and India, where there are reports of human casualties by tigers (Sukumar 1994; Veeramani, Jayson & Easa 1996, Saberwal 1996).

It has been suggested that extinction of small populations occurs by loss of genetic variability and fluctuations in demographic factors, and that population sizes are the main predictor of population extinction over time (Lande 1988). However, detailed statistical analysis of published and unpublished population reports for 10 species of large carnivores suggested that conflict over space between humans and these species is the main cause of mortality in wild populations; farmers and settlers at the edges of nature reserves were responsible, accidentally or not, for the majority of the mortality in large carnivores. Conservation efforts should thus focus largely in this aspect, especially in the case of small reserves with wide-ranging species (Woodroffe & Ginsberg 1998).

The use of land surrounding tiger territories by people can also affect tiger population over time. For example, cub survival is higher in areas with few or no roads than in areas with primary and secondary roads. Also, human disturbance reduces prey consumption and time spent on prey site, leading the animals to wander even further in their ranges (Kerley *et al.* 2002).

Humans can also compete with tigers directly for prey. In Nepal, one tiger lost ten kills to humans in an eight-month period; the tiger was driven away from the kills by humans, which then removed the carcasses from the reserve area (Sunquist & Sunquist 1989).

The effect of human-tiger conflict in tiger conservation is very strong and should be addressed at the beginning of any conservation programme. The political implications of this issue were the focus of discussion among conservation scientists in the end of last decade. In India, local human populations are generally unsupportive of conservation actions. Tigers are seen as dangerous animals that kill cattle and villagers, and nature reserves are often full of resources which the people lack (Saberwal 1996). The Indian government and conservation groups, working as part of an international effort, support the creation of a few completely inviolate areas for tiger conservation, which means relocating people and heavily securing the protected area to inhibit poaching, before the total disappearance of the species in the wild (Karanth & Madhusudan 1997). In any case, human-tiger conflict seems to be leading these carnivores to either total extinction in the wild within the next two decades, or to their confinement to small isolated populations that will not persist in the long term.

In the Royal Chitwan National Park, in Nepal, conservation efforts have been in practice for decades, and have focused frequently in the human populations surrounding the Park. As a result, tiger density in the park is the highest of the world, and the carnivore's presence seems to have also improved the population of other species in the park (Gittleman *et al.* 2001).

3.3. Tiger conservation in captivity.

While wild tigers are quickly and inescapably disappearing, the attention of researchers has been turning to the captive population and its potential to preserve the species. For two subspecies, the Siberian and the Amoy tigers, the reported captive population exceeds estimated numbers in the wild (Table 3.1).

Captive breeding programmes for all tiger subspecies are run in more than 200 institutions around the world (Olney & Fisker 2003), since the

viability of the captive population, for some, seems higher than the small, scattered remnants of wild populations. For instance, the Amoy tiger is the most critically endangered of these subspecies, with a wild population estimated as less than 50 individuals scattered in isolated pockets of habitat. It has been suggested that the subspecies is very close to extinction, and the captive population, although coming from only 6 wild-caught founders, may be the only alternative for Amoy tiger survival (Tilson, Traylor-Holzer & Jiang 1997).

Table 3.1: Estimated numbers in the wild and reported captive populations of tigers *Panthera tigris*; captive population numbers do not include individuals of unidentified gender (Jackson 1998; Olney & Fiske 2003; ISIS 2003)

Subspecies	Estimated wild population in 1998	Captive population in 2000 (studbooks)	Captive population in 2003 (ISIS)
<i>P. t. altaica</i>	360 – 406	466	365
<i>P. t. amoyensis</i>	20 – 30	51	Not available
<i>P. t. corbetti</i>	1227 – 1785	26	88
<i>P. t. sumatrae</i>	400 – 500	162	143
<i>P. t. tigris</i>	3176 – 4556	206	244
Totals	5183 – 7227	911	840

In the case of Sumatran tigers, official efforts started only in 1994, with the publication of the Indonesian Sumatran Tiger Conservation Strategy by the Ministry of Forestry of Indonesia. This document gave rise to a great number of multinational programmes, although researchers have pointed out how *in situ* and *ex situ* programmes must collaborate closely, for an international effort of this magnitude to be effective (Tilson *et al* 1997).

One specific problem faced by *ex situ* programmes is the limitation of resources, especially related to housing. In a survey of 1990, an estimated 1000 spaces existed in institutions worldwide, for tigers of all subspecies. Dividing them equally between subspecies would benefit the preservation of individual subspecies, while allocating more spaces to the subspecies

with higher genetic variability would help the conservation of the species as a whole (Maguire & Lacy 1990). The development of assisted reproduction techniques, such as cryogenic preservation of gametes and embryos, can help solve the problem by allowing the production of embryos from the present population; preserved, these could be implanted, when needed, in surrogate mothers (Donohue *et al.* 1990). Tigers have been intensively researched in these aspects and the first technical protocols are achieving positive results (e.g. Byers *et al.* 1990; Donohue *et al.* 1992; Donohue *et al.* 1996; Crichton *et al.* 2003).

Recently, researchers have discussed new priorities on tiger conservation. Earlier approaches focused on keeping a viable population for each of the five tiger subspecies in the wild, but genetic analysis showed that there is very little difference between the four continental subspecies of tiger, and only the Sumatran tiger, an island subspecies, has significant genetic differences from the continental subspecies (Ginsberg 2001).

Conservation *ex situ* has been the subject of controversy among researchers. Some view zoological institutions as Noah's Ark (the "Ark Paradigm") and believe that all declining species can be preserved through re-introduction of zoo-bred animals (Gippoliti & Carpaneto 1996); others sensibly point out the high costs of this type of programme, adding that they should be used in extremely rare situations and the indiscriminate re-introduction of captive-bred animals can be responsible for disease outbreaks in wild populations (Snyder *et al.* 1996). For a species, such as the tiger, whose ecological demands of large areas and abundant prey lead to conflict with human populations, re-introduction programmes have to be very carefully planned and managed, or the new populations will face the same pressures of the original ones, and also disappear. As pointed out recently by researchers, captive breeding and re-introduction programmes for large carnivores cannot bring about population recovery if the species has declined because of habitat destruction (Woodroffe 2001).

A species as high in the trophic chain as the tiger may soon enough be displaced by its human competitors, and may become exclusive to zoological institutions and breeding centres. This captive population, if well managed, can persist for long periods of time with minimum loss of genetic variability. It is important, though, to understand the biological patterns of this population for optimal management, especially related to husbandry techniques.

Studbooks are fundamental for conservation programmes and have been used as sources of reliable data for the research into several aspects of captive breeding in carnivores (c.f. Chapter 2), and also can provide data to the construction of mathematical models and simulations that can be applied for wild populations (Wildt, Howard and Brown 2001).

The importance of studbooks to the conservation of large carnivores is exemplified by the management plan for the African lion subspecies in North American zoos. Of the two subspecies identified in the captive stock, *Panthera leo krugeri* and *P. l. nubicus*, only *P. l. krugeri* had breeding potential to take part in a future *ex situ* conservation programme. Furthermore, with the widespread occurrence of feline immunodeficiency virus (FIV) and canine distemper in North America, the species survival plan recommended only tested individuals to be included in breeding loans and programmes, in an attempt to contain the spread of these diseases (Shoemaker & Pfaff 1997).

Biological information on captive populations may help to understand the biology of the species in the wild, at least for some species. For example, one of the most endangered of Carnivora species, the giant panda *Ailuropoda melanoleuca* has more than two decades of records in studbooks. In one study, researchers monitored the breeding biology of six wild female giant pandas. The results were compared with data from studbooks, in aspects such as litter size, interbirth interval and reproductive life span and found no difference between captive and wild counterparts (Zhu *et al.* 2001).

3.4. Hypotheses to be tested

I use the tiger studbook as a model to address the possibility of answering biological questions with record research. I point out that the tiger is a species with a large latitudinal range (Wilson & Reeder 1993) and does not seem to be strictly seasonal (Byers *et al.* 1990; Smith & MacDougal 1991), thus excluding photoperiodic effects in breeding. Considering that there are abundant food and water in captivity, and that serious institutions have to provide adequate housing to their animals, it is not expected that biological parameters, such as litter size, differ from wild populations. There are genetic, phenotypic and geographical differences between the two subspecies in the studbook (Ginsberg 2001), so it is expected that the dataset will reflect these differences on some biological parameters and express the need of subspecies-orientated husbandry protocols. I hypothesise also that there will be little or no effect of inbreeding levels in this species for, like several carnivores, it presents high levels of homozygosity in wild populations (Shivali, Jayaprakash & Patil 1998).

3.5. Methods.

In this research, data from the Tiger International Studbook were compiled and statistically analysed using the computer programmes MINITAB v. 13 and SPSS v.11. The dataset extracted from the electronic tiger studbook kept by Sarah Christie at the Zoological Society of London, was collected using the software SPARKS, from the International Species Information System (ISIS). It contains all births and infant deaths in 116 institutions mostly in the Northern hemisphere, during the period from 1986 to 1996. The records refer to two subspecies: the Sumatran (*P. tigris sumatrae*) and the amur or Siberian tiger (*P. tigris altaica*).

The dataset contains 249 litters born from 126 females (28 Sumatran and 98 Siberian) over 10 years (1986-1996), and there is additional detailed information for 43 females, such as place and date of birth, parentage and number of transfers. There are data on the number of young born, date of birth, offspring that died up to 6 months old, date of death, litter size, litter rearing method (if reared by parents or by hand), inbreeding level and parental identity. There was no information on enclosure area or housing conditions (see Appendix 3). The cause of death of young is reported in less than 12% of the events. There was no data on female body weight.

Studbook information is, as long as possible, checked by the studbook keeper. Mrs. Sarah Christie pointed out that older data might present some problems such as unreported litters, especially stillbirths, and that the data on the gender of the young may be unreliable, misleading sex ratio calculations. The identity of the female was reassured whenever possible. Although the interbirth interval could be calculated for 64 females, many institutions house animals of different genders separately, only occasionally gathering them for breeding, while others do not allow females to breed more than once a year. Housing protocols can therefore influence this result.

The dependent variables for regressions were the proportion of infant mortality for each female (I , calculated by equation 2.1, cf. Chapter 2), transformed by the arcsine of the square root; female breeding performance (B , see equation 2.2); the overall breeding success of the institution (Z , see equation 2.3) and the absolute litter size.

For the statistical tests, litters with uncertain numbers of individuals were not considered. The discarded data comprised 0.76% of the information available on the studbook. Young of undetermined gender comprised 10.44% of the dataset; however, as pointed out by Mrs. Sarah Christie, this information should not be relied on.

As the tests done in this chapter refer to the same population, it is necessary to correct for multiple tests. In this work, the method used was described by Legendre & Legendre (1998), which adjust values for multiple analyses but still allows lighter effects to be detected. All p-values presented in this chapter are adjusted. Data used in the analysis of average litter sizes were calculated separately for each female. Data on breeding age of dam, age of young at death and breeding season used pooled females. Institutional characteristics were collected from the *International Zoo Yearbooks* and can be found in Appendix 2.

3.6. Results.

3.6.1. Description of reproductive parameters in the sample.

3.6.1.1. Births.

In the tiger studbook, recorded births occurred mostly during Spring and Summer, peaking in the month of May (figure 3.1).

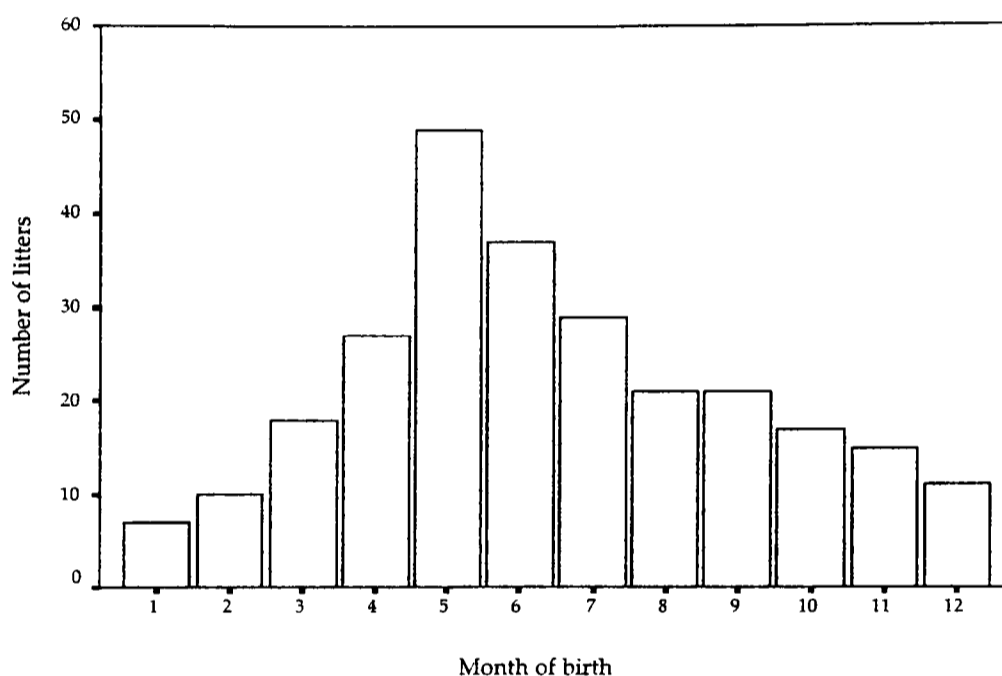


Figure 3.1: Absolute frequency of births of tiger litters in 116 Northern hemisphere institutions over ten years, by month (N=262).

In a study in Nepal, there was no evidence of breeding seasonality in wild tigers. The number of new litters appears to peak around the beginning of Summer and once again, less remarkably, in the beginning of Winter, but the difference is not statistically significant (Smith & MacDougal 1991). In the studbook data, the apparent peak during Summer months may be caused by active management of the institution: as it is known that gestation length in tigers averages 104.1 days, or approximately three and a half months (Gittleman 1986b), males and females are kept separated and

introduced to each other in the end of Winter, enabling litters to be born during Summer, when the number of visitors is higher.

From the 116 institutions present in this dataset, 107 have known latitude. Most of them, however, are located between 35 and 55 degrees of latitude (Figure 3.2). Tigers occur between the latitudes of 62 ° N and 10 ° S (Wilson & Reeder 1993), so it was decided not to include latitude as a factor in this analysis, once it is unlikely that results would be significant.

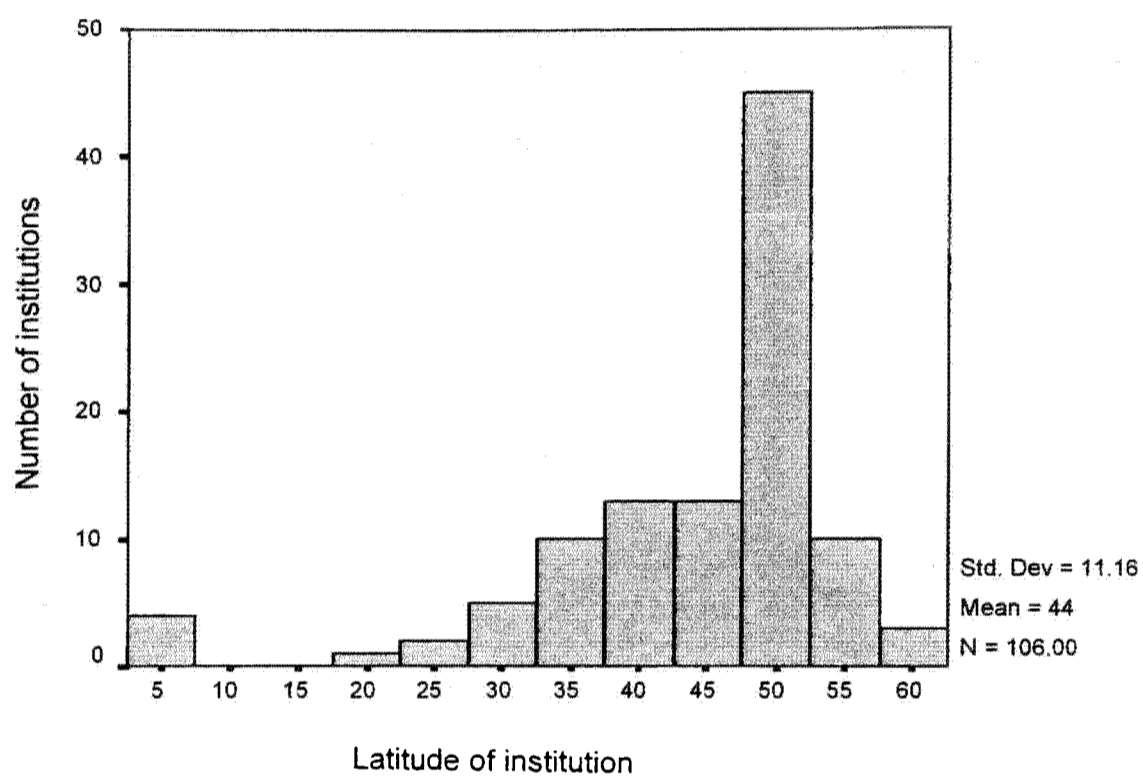


Figure 3.2: Absolute frequency of zoological institutions present in the International Tiger Studbook according to the latitude of their location (N=106).

3.6.1.2. Litter size.

In this dataset, litter sizes ranged from one to seven cubs (average of 2.72 ± 1.17 cubs per litter, $N=126$). Figure 3.3 shows the absolute frequency of litter size; most of the litters have less than four cubs.

In a wild population in Nepal, litter sizes also ranged from one to seven cubs, but seldom more than three (Sunquist 1981). In another study in the same population, litter sizes ranged from two to five cubs, with an average of 2.98 (Smith & MacDougal 1991). The average litter size in the wild for the species, calculated from published material, is from 2.5 (Gittleman 1986b) to 3 (Gittleman 1989).

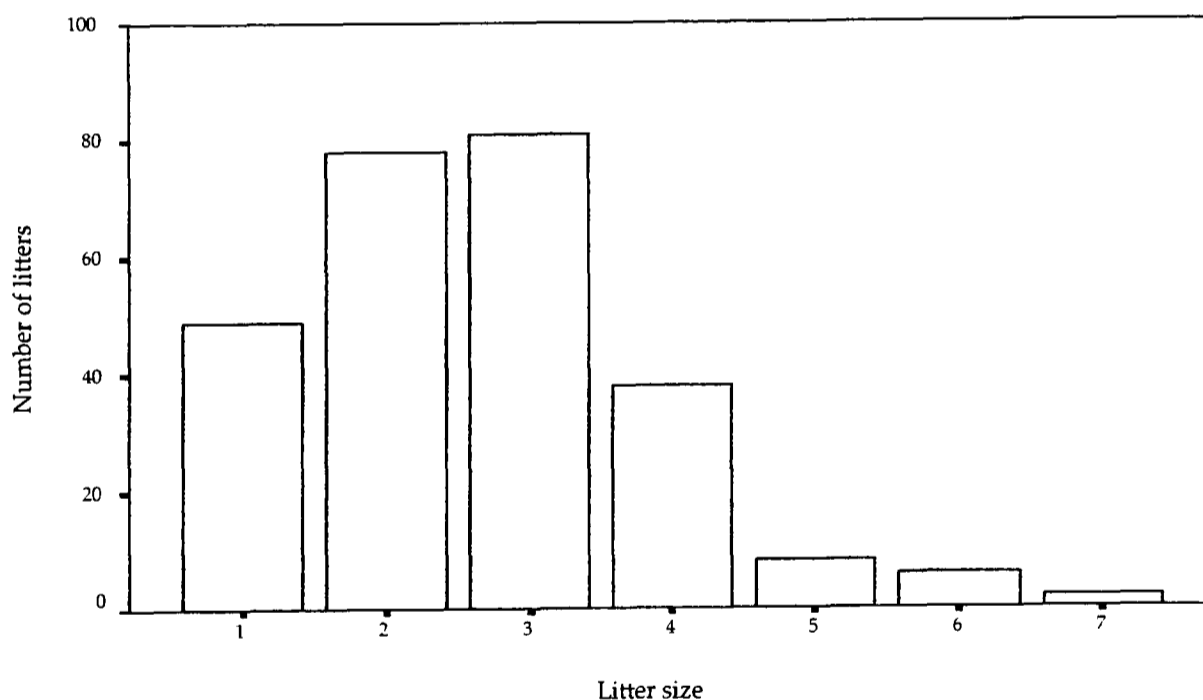


Figure 3.3: Absolute frequencies of litter size on captive tigers from 116 Northern hemisphere institutions ($N=262$).

Table 3.2 summarises this data from the wild providing sample sizes when available. In studies made previously in zoos, the average litter size reached 2.8 (Sankhala 1967).

Table 3.2: Summary of litter sizes found in wild tiger populations. Sample sizes are provided when available.

Source	Place	Litter size (N)	Sample size
Kerley <i>et al.</i> 2003	Russia	2.4±0.6	16 litters
Sunquist 1981	Nepal	2.52	22 litters
Smith & MacDougal 1991	Nepal	2.98	49 litters
Gittleman 1989	Several	3	Not available
Gittleman 1986a	Several	2.5	Not available

3.6.1.3. *Breeding age of dam.*

On average, the captive female tigers produced more litters when 6.2 years (± 2.96 , N=241), but females bred from 2 to 16 years of age (figure 3.4). There is very little information on these aspects in wild tiger populations, but oestrus was once observed in a 30 months old female in Nepal (Sunquist 1981). Other study in the same area revealed a mean age of first reproduction as 3.4 years (Smith & MacDougal 1991). In zoos, first mating was observed in females between three and six years old (Sankhala 1967). In this study, there are few reports of 2 years old females producing young, but this information is usually based in estimated age of dam and cannot be relied on. In one event, however, a female with known date of birth bred before 24 months of age, but this is probably a very rare phenomenon. Management decisions may affect the distribution of births, because studbooks keepers tend to take older animals out of the breeding stock (Christie 2000).

There was no significant effect of the age of dam over litter sizes (One-way ANOVA: $F_{15, 227} = 0.976$, $p = 0.48$) or the proportion of infant mortality (One-way ANOVA: $F_{15, 226} = 1.323$, $p = 0.18$).

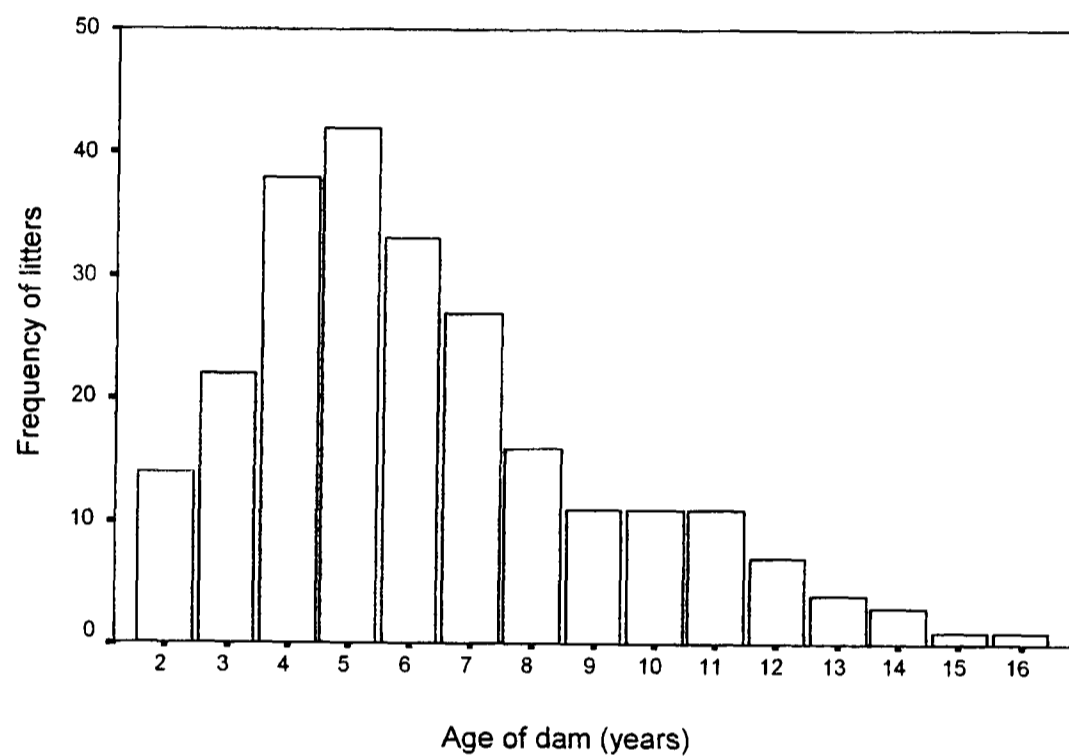


Figure 3.4: Absolute frequencies of breeding age of captive female tigers, in number of litters (N=262). Females are pooled in the analysis.

3.6.1.4. Age of death of young.

Most infant deaths in this database occurred in the first week of life (Figure 3.5), and on average the young died before the end of the third week (average week of death = 2.1 ± 2.1).



Figure 3.5: Absolute frequencies of age of death of tiger litters in captivity (N=262).

3.6.1.5. Proportion of infant mortality.

In this dataset, the median proportion of infant mortality is higher than the median proportion calculated from the database from the *International Zoo Yearbooks* (section 2.2.2.1, in Chapter 2), which contains information from 230 institutions. The median infant mortality calculated from the studbook, for the first 30 days, was 68%, against 37% of the *International Zoo Yearbooks* dataset. In the *IZY* there is no information on the number of litters and females, and the proportion of mortality was calculated by collection (i.e. all young born by year independently of

number of females or litters). In this dataset, the proportion of infant mortality for each female was calculated, and the median for the sample was taken.

In Nepal, cub mortality in the wild was recorded as 34% for the first year (Smith & MacDougal 1991), and 31 - 43% for the first two years (Sunquist 1981). In India, first year mortality in tiger cubs was calculated around 38% for the first year, but because of the difficulty of knowing actual litter sizes in the wild, it could be as high as 50% (Sunquist 1981).

3.6.1.6. Differences in reproductive parameters between subspecies.

There was no evidence of significant statistical difference between Sumatran and Siberian tigers in the number of litters produced by each breeding female ($t_{124} = -0.748$, $p = 0.46$). Also, no significant differences between these two subspecies were found in the interval, in years, in which each female had litters recorded in the studbook between 1986 and 1996 ($t_{112} = 0.208$, $p = 0.84$) or the female's average number of litters per reproductive year ($t_{112} = -0.238$, $p = 0.81$). Both the proportions of infant mortality ($t_{124} = 1.471$, $p = 0.14$) and heterozygosity - or inbreeding level - ($t_{32} = 0.721$, $p = 0.48$) did not differ significantly. However, Sumatran tigers appear to produce smaller litters ($t_{124} = -1.705$, $p = 0.09$, $\eta^2 = 0.05$) and to have produced fewer cubs in total during these 10 years ($t_{81.4} = -2.778$, $p = 0.007$, $\eta^2 = 0.035$) than the continental subspecies. Sumatran cubs died significantly earlier, in average, than Siberian cubs ($t_{123.9} = -2.770$, $p = 0.006$, $\eta^2 = 0.023$).

3.6.2. Factors affecting infant mortality.

3.6.2.1. Litter size.

Litter size had an effect on the median proportion of mortality of young (transformed by arcsine of the square root), especially when corrected for females (regression: $F_{1,124} = 77.129$, $p = 0.000$). Litter size explains 37.8% of the variance in the proportion of infant mortality. The fitted line plot can be seen on Figure 3.6. The relation of litter size and infant mortality in other species will be discussed in section 3.6.3.

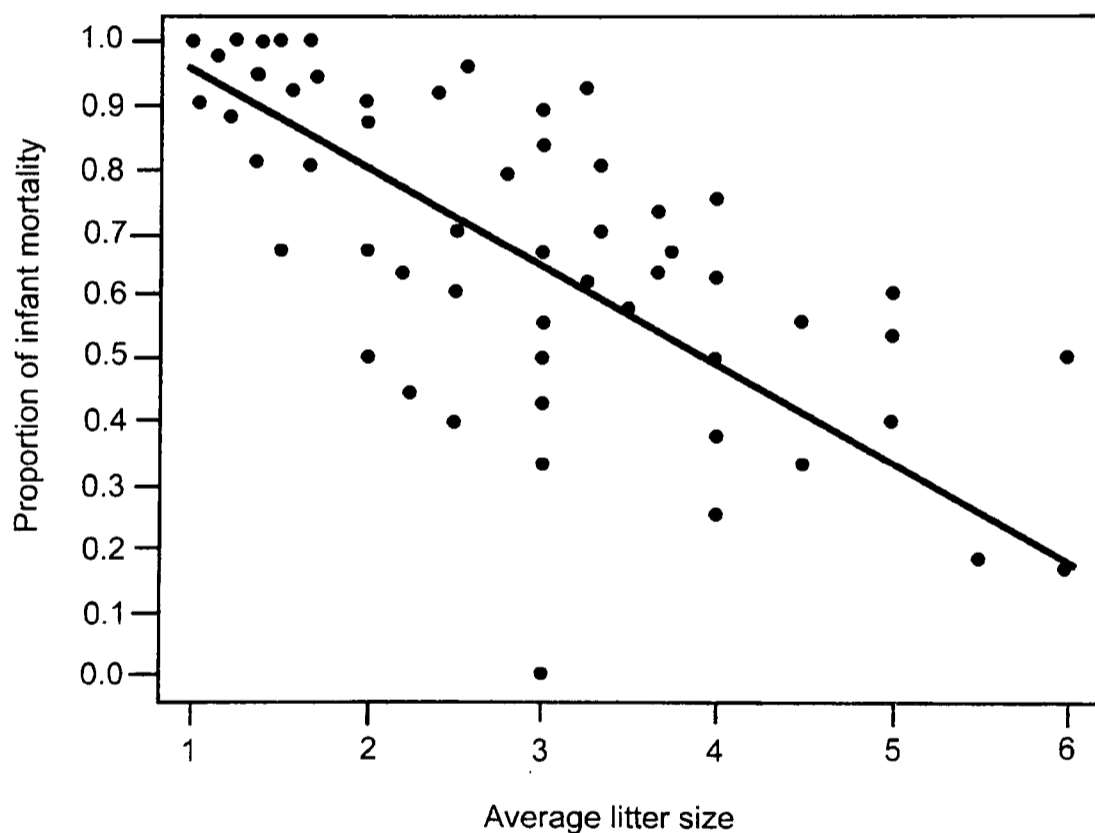


Figure 3.6: Regression plot for the effect of average litter size for female tiger, on the median proportion of infant mortality. Each dot represents one female tiger. The data plotted was not transformed.

3.6.2.2. Season of reproduction.

The median proportion of infant mortality is higher in litters that are born in autumn than in other seasons, for autumn litters are smaller than litters born in any other season, when corrected for the latitude of the institutions (One-way ANOVA: $F_{3,258} = 3.95$, $p = 0.036$; see Figure 3.7).

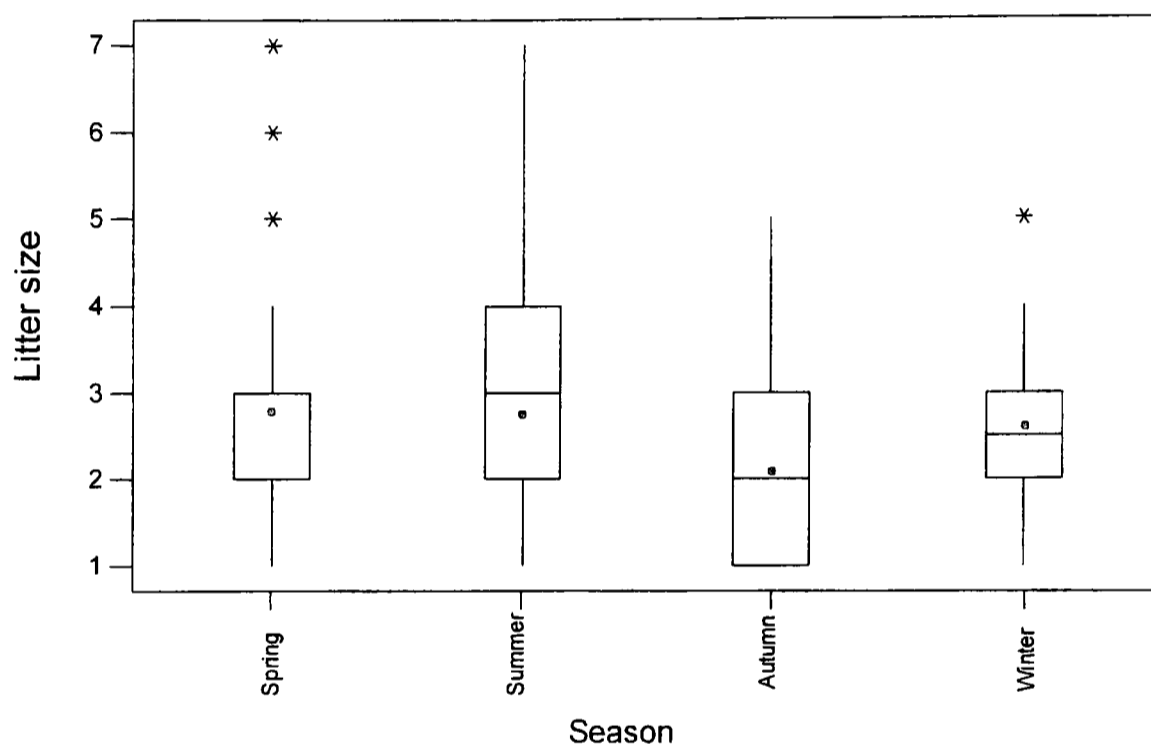


Figure 3.7: Litter size in captive tigers in each season of the year. Solid circles represent means, lines represent medians and stars represent outliers. The data plotted has pooled females.

3.6.3. Factors affecting institutional female breeding success.

3.6.3.1. Institutional characteristics.

Institutions that presented low breeding performance with species of the Order Carnivora in general (see Chapter 4) also scored poorly for breeding success in tigers (Regression: $F_{1,81} = 9.28$, $p = 0.015$), but there was no statistically significant difference between breeding success in

institutions that did and did not perform research with their animals, when the p-value was corrected, as seen overall for the order in Chapter 7 (One-way ANOVA: $F_{1,81} = 3.74$, $p = 0.17$, see figure 3.8). The higher proportion of infant mortality in this dataset, when compared to the tiger dataset from the *International Zoo Yearbooks*, may reflect the prevalence of institutions with overall poorer carnivore breeding success in the studbook, and may be related to institutional husbandry protocols. The studbook dataset also contained a higher percentage of institutions performing research (45.8%) when in comparison with the institutions listed in the dataset of the *International Zoo Yearbooks* (28.5%). The studbook excludes hybrids individuals and those of unknown subspecies, and institutions with researchers are more likely to be able to identify with certainty the subspecies of their individuals.

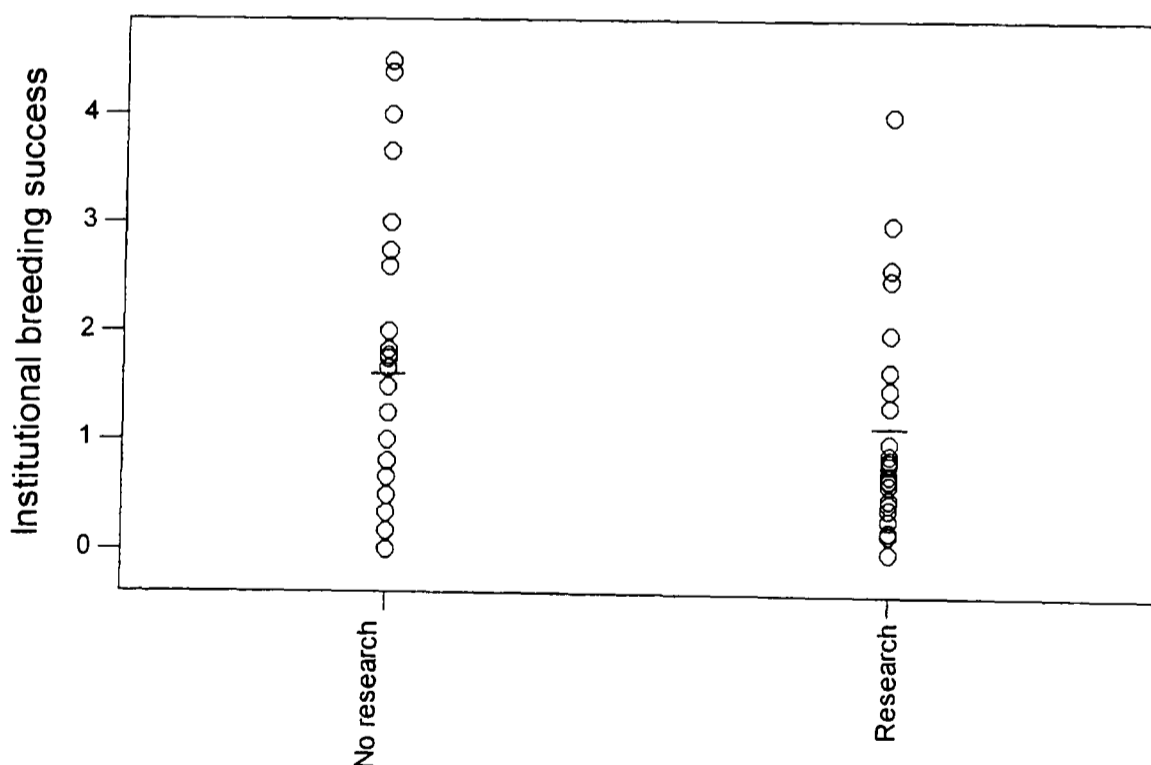


Figure 3.8: Breeding success of female tigers kept in institutions with (N=38) and without (N=45) research groups.

Unfortunately, it was not possible to collect information regarding the institutional experience on tiger reproduction, because breeding efforts are frequently informal at the beginning and are usually not reported to studbook keepers until some positive results are obtained (Christie, pers. comm.).

3.6.3.2. Female characteristics.

There is no detectable effect of inbreeding level in individual females' breeding success, although a larger sample would be needed to increase test power (Regression: $F_{1,40} = 1.24$, $p = 0.4$, $R\text{-Sq} = 3\%$). There was no evidence that wild-caught females performed worse than captive-born ones, but it must be considered that there are only nine wild-caught females in the sample, against 35 captive-born, and the power of the test is low (One-way ANOVA: $F_{1,42} = 0.32$, $p = 0.5$, $\text{power} = 0.07$). The dataset did not contain information on female body size, and there was no sample size large enough to investigate the effect of the age of dam on breeding success. For the reasons discussed before, it was not recommendable to perform tests on sex ratio.

3.7. Discussion.

3.7.1. Reproductive parameters in captivity and in the wild.

The reproductive parameters of gestation length and litter size found in the tiger studbook data were, in general, congruent with the data collected in the wild, suggesting that this species preserves much of its reproductive patterns in captivity.

The peak of births during summer found in this research may reflect more the husbandry techniques applied by institutions than the actual existence of a breeding season, and a previous research using data from the

Siberian tiger studbook for North America, seasonal analysis also showed a peak of births between April and June (Seal *et al.* 1985). Although seasonality is uncertain in tigers as a species (Byers *et al.* 1990; Smith & MacDougal 1991), a study on seven captive females indicated that oestrus and follicular cycles started in late January and ceased in early June, suggesting that Siberian tigers, at least, may be induced ovulators and seasonal breeders (Seal *et al.* 1985). Season did not seem to affect semen viability in five captive Siberian tigers monitored throughout the year, but serum concentrations of thyroxin and triiodothyronine were lowest during summer, and testosterone was higher in autumn and early winter (Byers *et al.* 1990). Unfortunately, there are no studies on seasonality on Sumatran tigers, and further research is needed for the species as a whole, if possible using larger sample sizes.

3.7.2. Subspecific differences and implications to captive breeding.

As it was said before, there are few genetic differences among the four continental subspecies of tigers; continental subspecies, however, differ greatly from the Sumatran subspecies, and this fact is leading to a change in conservation priorities for tiger subspecies (Ginsberg 2001). Subspecific hybrids happen frequently in the captive population (Olney and Fisker 2003) and also in some wild populations. For example, conservation efforts on a wild tiger population in India led to the introduction of a hybrid Siberian-Bengal tiger in an attempt to increase tiger numbers; two decades later, genetic analysis of this population showed a prevalence of Siberian tiger markers in the reserve population (i.e. subspecific hybrids), but as the hybrids were fulfilling the ecological role in the area and were unlikely to migrate to other areas, and the population would not support the removal of hybrids, the population was left in this way (Wayne & Brown 2001).

A recent review of the factors affecting the persistence of carnivore species pointed out aspects that make some species more prone to extinction: small populations; island endemics; higher trophic levels; slow life histories; complex mating displays and social structure; large home ranges; and large body sizes, which correlate with many of the previous aspects (Purvis, Mace & Gittleman 2001). Although both subspecies present the same basic characteristics, Sumatran tigers may be more vulnerable to extinction pressures because of their natural distribution (an island), smaller populations, and reduced litter sizes and female reproductive output. This may indicate that a tailor-made conservation approach is needed for the subspecies, instead of applying to Sumatran tigers the same protocols used for the Siberian subspecies.

3.7.3. Factors affecting breeding success.

Infant mortality in the dataset from the *International Zoo Yearbooks* was similar to that found in wild populations, and even smaller than the 50% rate suggested by Sunquist (1981). In the studbook dataset, however, infant mortality during the first month was extremely high (68%), but may reflect bias in the data, as discussed in Chapter 2. Further investigation, using a broader dataset, would be useful to determine the actual infant mortality rates for the species in captivity.

Of the factors that could be related to infant mortality in captive tigers, litter size seems to have a very strong effect. The fact that litter size was negatively correlated with infant mortality in captive tigers was not expected, because litter sizes are usually positively related with infant mortality in several mammalian species, such as the common marmosets *Callithrix jacchus* (Rothe, Darms & Koenig 1992) and other Callitrichidae (Jaquish, Gage & Tardif 1991), and snowshoe hares *Lepus americanus*

(O'Donogue 1994). Infant mortality can be affected also by birth weight, which is negatively correlated to litter size in mammals such as the snowshoe hare (O'Donogue 1994) and the Zambia giant mole rat *Cryptomys mechowii* (Scharff *et al.* 1999).

Litter sizes can be related to the age of death of young. For example, in captive common marmosets, perinatal infant mortality was prevalent, and stillbirths and abortions were related to litter size: most abortions occurred in singleton pregnancies, while most of stillbirths occurred in quadruplets (Rothe, Darms & Koenig 1992). Also, in a study of several species of Callitrichidae, litter size also influenced infant survivorship; survival to maturity was higher in singleton and twins than in triplets, while perinatal mortality was higher in singletons and triplets than in twins, suggesting some influence of sibling competition and maternal care (Jaquish, Gage & Tardif 1991).

An indication of what could be influencing the relation of litter size and infant mortality in captive tigers was found in other species of mammals. Litter sizes were positively correlated with maternal body fat (or nutritional status) in Virginia opossums *Didelphis virginiana* (Hossler, MacAninch & Harder 1994) and raccoon dogs *Nyctereutes procyonoides* (Kauhala & Helle 1995). In raccoon dogs, litter sizes were correlated with the abundance of prey and population density, which affected directly the fat reserves of females, but mortality before maturity, albeit high, was not correlated with litter size (Kauhala 1996). In cheetahs *Acinonyx jubatus*, maternal food intake and maternal fat reserves were positively correlated with cub growth rate up to a physiological limit (Laurenson 1995). Cub birth mass and maternal mass also affected positively infant survival in polar bears *Thalarctos maritimus*, and maternal condition affected infant survival during the first year; lack of food, and its direct effect on maternal fat stores, can be crucial during lactation and may be the main cause of mortality in polar bear cubs (Derocher & Stirling 1996). This may suggest that female

tigers that had larger litters were in a better nutritional and health status, and were able to raise the cubs successfully. One confounding factor in captive tigers, though, is the husbandry protocol that some institutions adopt: in many zoos, cubs are removed immediately after birth for hand-rearing, especially after a maternal infanticide event (e.g. Hughes 1977). This way, larger litters could have at least some individuals “rescued”, while small litters, especially those of singletons or twins, could be killed almost instantly, before the intervention of zoo staff. A more detailed database would be needed to rule out the influence of human intervention on cub survival in captive tigers.

Researching infant mortality in 18 species of canids in captivity, Ginsberg (1994) found that there is a strong correlation between extensive institutional breeding experience and pup survivorship, explaining up to 77% of the variance. Species bred in captivity for a long time will have higher infant survival.

In this research, there was an effect of the season of the year in litter size, with smaller litters occurring during autumn months. If litter sizes are related to female body fat and nutritional conditions in tigers, it would be informative examining female weight fluctuations in captive conditions, and a decrease in body fat would be expected by the end of the summer and beginning of autumn.

In any case, it is possible that there is a real relation between litter size and cub survival in tigers in captivity. It was suggested that the relation between infant survival and litter size can be affected by population dynamics, and change overtime. In a long-term study with Columbian ground squirrels *Spermophilus columbianus*, litter sizes were large during the phase in which the population was growing, and infant survival increased with litter size; when the population was decreasing, the average litter size fell from 4 to 3, and larger litters suffered higher mortality, which can be related to changes in resource availability (Festa-Bianchet & King 1991). As

resources are theoretically abundant and do not change over time in captivity, the captive population could be behaving as a population in growth phase.

The relation found between institutions that had overall poor carnivore breeding and those who were not breeding tigers successfully suggests the influence of husbandry techniques in infant mortality. Some of the aspects that may be involved are hand-rearing techniques, housing conditions and the provision of a well-balanced diet, but this data was not available.

There was no detectable effect of inbreeding levels in infant mortality of captive tigers. The effect of inbreeding depression in tigers is not yet fully researched, and opinions on the subject differ. Inbreeding depression can be reflected in many aspects of reproduction, such as litter size, infant survival and spermatozoa quality. A study in the Royal Chitwan National Park in Nepal monitored 22 breeding females and 14 breeding males for 16 years, and found that the inbreeding rate of this population, one of the largest in the Indian subcontinent, is 2% per generation. This suggested that, if low levels of heterozygosity affect the species, many tiger populations would be vulnerable to inbreeding depression (Smith & McDougal 1991).

In recent years, the idea that captive tiger populations may have low genetic variability due to inbreeding has been researched with the aid of technological innovations, and the results point in a different direction. For instance, a DNA fingerprinting study in 22 tigers from a wild population in India revealed that its level of genetic variability (22.65%) did not differ from levels analysed in museum skin samples collected between 50 and 125 years ago (21.01%). This may indicate that the low genetic variability found in this subspecies was not caused by the population bottleneck of the beginning of the 20th century (Shankaranarayanan *et al.* 1997).

In a study on the effects of inbreeding in captive Indian tigers, semen was collected for analysis of spermatozoa quality and fertilizing ability, one

of the measures of inbreeding depression in mammals. The majority of samples fell within the estimated optimal values for the species, suggesting that high homozygosity does not cause inbreeding depression on the species (Shivali, Jayaprakash & Patil 1998). This type of research, however, is extremely new and much work is still needed to get to more accurate results.

3.8. Conclusion.

Captive tigers seem to preserve natural reproductive parameters, and research with these populations can prove very valuable to the study of wild populations. Studbooks can provide reliable data, and individual institutional records can help to unravel factors that affect the reproduction on the species.

Conservation efforts should integrate research in both captivity and the wild, in a complementary way, since certain factors were not yet researched in wild populations due to the obstacles of data collection; this information can be crucial for the maintenance and protection of surviving wild populations.

It is important to stress the urgent need of a more collaborative interchange of information between the several groups participating in the conservation of tigers, to preserve remaining populations in the wild, once the reintroduction of large predators can face many challenges. A multidisciplinary approach can facilitate the understanding of the causes of species decline and, with the help and participation of the human populations surrounding tiger reserves, achieve more positive results towards the persistence of the species in the wild.

Chapter 4

Biological factors affecting infant survival in captive carnivores

Abstract

The proportion of infant mortality in 98 species of captive carnivores was calculated from the *International Zoo Yearbooks* (Vols. 26-35). Infant mortality in carnivore species in captivity was significantly affected by the developmental characteristics of the species such as the age when young open their eyes, gestation length and weaning age. Specific body weight did not have an effect on infant mortality in captivity as it had on the juvenile mortality of carnivores in the wild. Species with delayed implantation of embryos did not have lower infant mortality in captivity. Declining species presented higher infant mortality than abundant ones, but the rates of juvenile mortality differ greatly between wild and captive populations. Species from temperate or cold climate had higher infant mortality when kept outside of their natural latitudinal ranges, although this effect was not significant for tropical species, even those with restricted distribution. Infant mortality in captive carnivores was also significantly affected by the specific type of diet and zonation, but there were not significant effects of activity patterns or habitat vegetation of the species on infant mortality in captivity.

4.1. Introduction.

The evolution of species is driven by adaptation to habitats. Some species spread through large areas, encompassing more than one ecosystem; others became highly specialised in only one, sometimes frail, habitat. Species that occupy different habitats must not only have flexible strategies to explore resources, but also be able to reproduce in the most varied conditions. Highly specialised ones, however, are restricted to places that present optimal conditions.

The process of reproduction has a complex structure that could be influenced by many factors, whether biological or environmental (cf. Ch. 1). For captive reproduction programmes, it may be tempting then to try and

control enclosure conditions. Given the diversity of species and their particular demands, it is unlikely however that there is a panacea-like compilation of guidelines to be developed, and many programmes cannot afford experimental failures.

Although some research has been made on breeding success of mammals in captivity, and the effects of biological factors on infant mortality, most of them are focused on primates (e. g. Birrell et al. 1996; Courtenay 1988; Debyser 1995a; Debyser 1995b; Debyser 1995c; Mooney & Lee 1999). In the order Carnivora, this aspect has been researched mainly in the cheetah *Acinonyx jubatus* (Beekman et al. 1999; Benzon & Smith 1974; Marker & O'Brien 1989; Wielebnowski 1996), arctic foxes *Alopex lagopus* (Bakken 1992; Bakken 1993; Bakken 1994; Ilukha, Harri & Rekilä 1997) and black-footed ferrets *Mustela nigripes* (Biggins et al. 1998; Thorne & Oakleaf 1991). Species are not usually randomly chosen for this type of research. The captive population of the cheetah is one of the oldest and better recorded of all captive carnivores, and databases yield decades of breeding information (Marker-Kraus 1997). Arctic foxes are farmed for their fur, and higher breeding success leads to more profit (Bakken 1992). Finally, black-footed ferrets, being extinct in the wild, depend on low mortality rates to produce individuals for reintroduction programmes (Thorne & Oakleaf 1991). Also, the biology of these species was already well described (cf. cited authors). For other species, especially those with many, at present, unknown biological characteristics, research is extremely scant.

Conservation programmes have been developed for several species of carnivores, and those depending on captive populations are listed in Table 4.1. It is important to notice that the Species Survival Commission of the World Conservation Union, in its technical guidelines for *ex situ* conservation programmes (IUCN 2002a), decided that all species classified as Critically Endangered (CR) or Extinct in the Wild (EW) should be subjected to *ex situ* conservation in a way to restore wild populations, but many resources have been applied in programmes for species that are not listed. Some critically

endangered species, such as the Iberian lynx *Lynx pardina* and the Malabar civet *Viverra civettina*, do not have a breeding captive population until present. The IUCN also points out that the investment on *ex situ* programmes should take into consideration research results, and many species were not researched properly yet.

4.1.1. Hypotheses to be tested

Carnivore life history traits have been used in the understanding of complex biological processes such as energy expenditure in reproduction or extinction risk (Oftedal & Gittleman 1989; Purvis *et al.* 2000). Here I use multiple datasets to predict, through statistical models, the possible effects of the natural history of the species on infant mortality in captivity.

If conditions in captivity are providing all the resources for a species, then the specific proportion of infant mortality in captivity should be lower than in the wild, because the young do not risk dying of starvation (Kauhala & Helle 1995; Rogers 1987; Sklepkovych 1989), predation (Ralls & White 1995; Steiger *et al.* 1989; Van Heerden *et al.* 1995) or hunting (Lode 1995; Payne & Root 1986; Takeuchi & Koganezawa 1994).

Due to the great diversity of life history traits of the Carnivora (Gittleman 1986a, 1986b), it is expected that the variance in the proportion of infant mortality between species will indicate which are thriving in captivity (Debyser 1995a, 1995b, 1995c) and which biological parameters have an influence on infant survival (Birrell *et al.* 1996; Courtenay 1988; Mooney & Lee 1999). As altricial young are more fragile than precocial ones (Hayssen 1984; Hrdy 1979; Packer & Pusey 1984; Wolff 1997; Wolff & Peterson 1998), it is possible that species with slower development were more prone to lose infants. Arctic species are affected by photoperiod (Curlewis 1992; Jallageas *et al.* 1994; Lincoln 1998) and may present higher infant mortality when housed outside the species' range of distribution.

Infant mortality can be directly related to resource availability, through competition, predation and curtailed maternal investment (Clutton-Brock 1991), and high infant mortality in captivity may indicate that optimal conditions were not provided by the institutions (Debyser 1995a; Marker-Kraus 1997; Promislow & Harvey 1991). Species with elaborate nutrition, such as insectivores (Ashton & Jones 1979; Donoghue & Langenberg 1994; Price *et al.* 1999), and those with complex environmental needs, which need multi-environment enclosures (Carlstead & Shepherdson 1994; Lyons, Young & Deag 1997), are expected to breed poorly in captive conditions.

Table 4.1: Conservation programmes involving captive populations of carnivores. IUCN categories are from version 3.1 (IUCN 2001). Types of programmes: Managed captive populations = individuals tracked by studbooks and breeding depends on space availability; Captive breeding = extensive breeding loans with the purpose of enhancing captive population; Reintroduction = extensive captive breeding aiming reintroduction of new individuals in the wild.

Species	IUCN Status (IUCN 2002b)	Type of programme	Reference
<i>Acinonyx jubatus</i>	VU C2a(i)	Managed captive populations, captive breeding	Balmford, Mace & Leader-Williams 1996
<i>Ailuropoda melanoleuca</i>	EN B1+2c, C2a	Managed captive populations, captive breeding	Balmford, Mace & Leader-Williams 1996; Peng, Jiang & Hu 2001
<i>Ailurus fulgens</i>	EN C2a	Managed captive populations	Balmford, Mace & Leader-Williams 1996
<i>Amblonyx cinereus</i>	LR/nt	Managed captive populations	Balmford, Mace & Leader-Williams 1996
<i>Canis lupus</i>	The subspecies reintroduced are not listed	Managed captive populations, captive breeding, reintroduction	Balmford, Mace & Leader-Williams 1996; Berger <i>et al.</i> 2001
<i>Canis rufus</i>	CR D	captive breeding, reintroduction	Moore & Smith 1990; Woodroffe & Ginsberg 1997; Gittleman & Gompper 2001
<i>Chrysocyon brachyurus</i>	LR/nt	Managed captive populations	Balmford, Mace & Leader-Williams 1996
<i>Enhydra lutris</i>	EN A1ace	captive breeding, reintroduction	Gittleman & Gompper 2001
<i>Lycaon pictus</i>	EN C1	Managed captive populations, captive breeding, reintroduction	Balmford, Mace & Leader-Williams 1996, Gittleman & Gompper 2001
<i>Mustela nigripes</i>	EW	Managed captive populations, captive breeding, reintroduction	Thorne & Oakleaf 1991; Balmford, Mace & Leader-Williams 1996; Gittleman & Gompper 2001
<i>Neofelis nebulosa</i>	VU C2 a(i)	Managed captive populations	Balmford, Mace & Leader-Williams 1996
<i>Panthera leo persica</i>	CR C2a(ii)	Managed captive populations	Balmford, Mace & Leader-Williams 1996
<i>Panthera tigris</i>	EN C2a(i)	Managed captive populations	Balmford, Mace & Leader-Williams 1996
<i>Puma concolor coryi</i>	CR D	captive breeding, reintroduction	Gittleman & Gompper 2001
<i>Speothos venaticus</i>	VU C2a	Managed captive populations	Balmford, Mace & Leader-Williams 1996
<i>Ursus arctos</i>	Not listed	captive breeding, reintroduction	Berger <i>et al.</i> 2001, Gittleman & Gompper 2001

4.2. Methods.

4.2.1. The dataset: species and variables used.

The dataset used in this chapter was described in section 2.2.2.1 and is presented in Appendix 1. From the 98 species of carnivores that bred in zoos between 1986 and 1996, 85 species were analysed using biological, ecological and geographical data compiled by Gittleman (1983, 1986a, 1986b), Mead (1989) and Wilson and Reeder (1993), as predictors to infant mortality in captivity. The conservation status of the species was extracted from IUCN (2002b). Table 4.2 summarises the variables derived from each one of the sources.

Table 4.2: Summary of the biological, ecological and geographical variables, and their sources, used in statistical analysis in this chapter.

Type of data	Variables	Source	
Developmental Biology	Gestation length, litter growth rate, age of young opening the eyes, weaning age and age of independence	Gittleman	1986a, 1986b
Ecological	Type of diet, type of vegetation of habitat, type of preferred zonation, activity pattern	Gittleman	1986a, 1986b
Juvenile mortality	Juvenile mortality in the wild	Gittleman 1983	
Embrionic diapause	Presence and length of delayed implantation of embryo	Mead 1989	
Geographical	Northernmost and southernmost latitudes of species distribution	Wilson & Reeder	1993
Conservation status	Decline of wild populations of the species	IUCN 2002b	
Infant mortality in captivity	Median proportion of infant mortality in captivity	IZY Vols. 26-35	

Not all variables were available to the 98 species presented in Appendix 1. Table 4.3 displays which species were included in each one of the statistical analyses of this chapter.

Table 4.3: Species included in the statistical analysis using different groups of variables collected from published databases (Gittleman 1983, 1986a, 1986b; Mead 1989; Wilson & Reeder 1993). DB = developmental biology; ECO = ecological; JM = juvenile mortality; ED = Embryonic diapause; LAT = geographical. Specific variables are displayed in Table 4.2.

Family	Species	DB	ECO	JM	ED	LAT
Canidae	<i>Alopex lagopus</i>	•	•			•
Canidae	<i>Canis aureus</i>		•			
Canidae	<i>Canis lupus</i>	•	•	•		
Canidae	<i>Canis mesomelas</i>		•			
Canidae	<i>Cerdocyon thous</i>		•			
Canidae	<i>Chrysocyon brachyurus</i>		•			•
Canidae	<i>Cuon alpinus</i>		•			
Canidae	<i>Lycaon pictus</i>	•	•			•
Canidae	<i>Nyctereutes procyonoides</i>		•			
Canidae	<i>Otocyon megalotis</i>		•			
Canidae	<i>Speothos venaticus</i>		•			
Canidae	<i>Urocyon cinereoargenteus</i>		•	•		
Canidae	<i>Vulpes rueppelli</i>		•			
Canidae	<i>Vulpes velox</i>		•			
Canidae	<i>Vulpes vulpes</i>	•	•	•		•
Canidae	<i>Vulpes zerda</i>	•	•			•
Felidae	<i>Acinonyx jubatus</i>		•			
Felidae	<i>Caracal caracal</i>	•	•			•
Felidae	<i>Felis chaus</i>	•	•			•
Felidae	<i>Felis margarita</i>		•			
Felidae	<i>Felis nigripes</i>		•			
Felidae	<i>Felis silvestris</i>	•	•			
Felidae	<i>Herpailurus yaguarondi</i>		•			
Felidae	<i>Leopardus pardalis</i>				•	
Felidae	<i>Leopardus tigrinus</i>		•			
Felidae	<i>Leptailurus serval</i>		•			•
Felidae	<i>Lynx lynx</i>	•	•	•		•
Felidae	<i>Lynx rufus</i>	•	•	•		•
Felidae	<i>Oncifelis geoffroyi</i>		•			
Felidae	<i>Panthera leo persica</i>	•	•	•		
Felidae	<i>Panthera onca</i>	•	•			
Felidae	<i>Panthera pardus</i>	•	•			
Felidae	<i>Panthera tigris</i>	•	•	•		
Felidae	<i>Prionailurus bengalensis</i>	•				
Felidae	<i>Prionailurus viverrinus</i>		•			
Felidae	<i>Puma concolor</i>		•			
Felidae	<i>Uncia uncia</i>		•			•

Family	Species	DB	ECO	JM	ED	LAT
Herpestidae	<i>Atilax paludinosus</i>		•			
Herpestidae	<i>Cynictis penicillata</i>		•			
Herpestidae	<i>Helogale parvula</i>		•			
Herpestidae	<i>Mungos mungo</i>		•			•
Herpestidae	<i>Suricata suricatta</i>	•	•			•
Hyaenidae	<i>Crocuta crocuta</i>	•	•	•		
Hyaenidae	<i>Hyaena hyaena</i>	•	•			
Hyaenidae	<i>Proteles cristatus</i>		•			
Mustelidae	<i>Amblonyx cinereus</i>				•	
Mustelidae	<i>Eira barbara</i>				•	
Mustelidae	<i>Enhydra lutris</i>	•	•		•	
Mustelidae	<i>Gulo gulo</i>	•	•		•	
Mustelidae	<i>Ictonyx striatus</i>	•	•		•	
Mustelidae	<i>Lutra canadensis</i>	•	•	•	•	
Mustelidae	<i>Lutra lutra</i>	•	•		•	
Mustelidae	<i>Martes flavigula</i>		•		•	
Mustelidae	<i>Martes foina</i>				•	
Mustelidae	<i>Martes martes</i>		•		•	
Mustelidae	<i>Martes zibellina</i>	•	•		•	
Mustelidae	<i>Meles meles</i>	•	•		•	
Mustelidae	<i>Mephitis mephitis</i>	•	•	•	•	
Mustelidae	<i>Mustela erminea</i>		•	•	•	
Mustelidae	<i>Mustela eversmanni</i>				•	
Mustelidae	<i>Mustela lutreola</i>	•	•		•	
Mustelidae	<i>Mustela nigripes</i>				•	
Mustelidae	<i>Mustela nivalis</i>	•	•		•	
Mustelidae	<i>Mustela putorius</i>		•		•	
Mustelidae	<i>Mustela vison</i>		•		•	
Mustelidae	<i>Pteronura brasiliensis</i>				•	
Mustelidae	<i>Vormela peregusna</i>		•		•	
Procyonidae	<i>Ailurus fulgens</i>		•	•	•	•
Procyonidae	<i>Bassariscus astutus</i>	•	•			
Procyonidae	<i>Nasua nasua</i>					•
Procyonidae	<i>Potos flavus</i>		•			•
Procyonidae	<i>Procyon lotor</i>	•	•	•		
Ursidae	<i>Helarctos malayanus</i>		•		•	
Ursidae	<i>Melursus ursinus</i>		•		•	
Ursidae	<i>Tremarctos ornatus</i>				•	•
Ursidae	<i>Ursus americanus</i>	•	•	•	•	•
Ursidae	<i>Ursus arctos</i>	•	•	•	•	
Ursidae	<i>Ursus maritimus</i>		•	•	•	•
Ursidae	<i>Ursus thibetanus</i>	•	•		•	•
Viverridae	<i>Arctictis binturong</i>		•			•
Viverridae	<i>Civettictis civetta</i>		•			
Viverridae	<i>Genetta genetta</i>	•	•			
Viverridae	<i>Genetta tigrina</i>	•	•			
Viverridae	<i>Paguma larvata</i>		•			
Viverridae	<i>Paradoxurus hermaphroditus</i>		•			

Although tests did not point the need for phylogenetic analysis of this data (section 2.3.4), it is important to remember that there can be some bias towards certain taxa in different analyses.

4.2.2. Reproductive biology and infant survival.

To choose which variables were more adequate to test the hypothesis that the biology of species affects infant survival of carnivores in captivity, a stepwise regression (forward method) was used on the biological database of carnivores (Gittleman 1983, 1986a, 1986b). Biological aspects of the species that are related with the level of development in which the young are born and the post natal growth (gestation length, litter growth rate, age of opening the eyes, weaning age and age of independence, all transformed by natural log) were tested against the median proportion of infant mortality (*I*) for the captive populations published on the *International Zoo Yearbooks* between 1986 and 1996 (equation 2.1). Also, the same variables were tested by the “best subsets” regression method, in a way to identify the best model to be applied in the multiple regressions. These variables were used in a fully factorial general linear model using the software MINITAB v.13, and all the p-values presented in this chapter are adjusted for multiple analyses.

4.2.3. Delayed implantation.

Information on delayed implantation (DI) from several species, from Mead (1989), was transformed to codes. The information available was presented as an objective measure for only two species (in months of delay), but for all others it was presented in a subjective form (absent, suspected, unknown duration, short duration and long duration). It was then considered that the DI length of those species represented in months was equivalent to ‘long duration’, for they consist in more than 70% of the total gestation time.

The code then was as follows: 0 = absent; 1 = suspected; 2 = unknown duration; 3 = short duration and 4 = long duration. This variable was then tested against the value *I*, through an analysis of variance (ANOVA).

4.2.4. Captive infant mortality in declining species.

The IUCN Red List categorises species according to subtle threat levels, such as 'lower risk, least concern', 'critically endangered' and 'extinct in the wild'. Some species are classified as 'data deficient'. For this test, due to the small sample size, there are not degrees of freedom enough to allow the analysis with the full 5-level IUCN code used by Purvis *et al.* (2000). A two-level code was then used, assigning a value of 1 if the species is listed as 'lower risk' or higher, or 0 if not listed. Data deficient species were not considered. Again, the dependent variable in the analysis of variance was the value *I*.

4.2.5. Photoperiod and latitudes.

Based in this influence of latitude in the biology of temperate species, specific infant mortality was tested against the latitude of the zoological institutions. Information on the distribution of species was collected from Wilson and Reeder (1993). The extreme points of latitudinal distribution of each species were found in maps, and the latitude of each locality was found with the aid of a geographic positioning software, Earth Explorer 2.5 (Motherplanet Inc.). The species considered for this test were those with a range of latitudinal distribution smaller than 50 degrees (for example, from 20° N to 50° N), and that were kept in more than 30 institutions. Species were considerate Arctic if the lower latitude of distribution was above 23°, therefore having no population between the Tropics, and Tropical if many populations were found in this area. The list of species can be seen in Table 4.3. Code

values were attributed to institutions, depending whether they were outside (0) or inside (1) the natural range of the species.

4.2.6. *Nutrition and zonation.*

Information on type of diet, activity patterns, type of preferred vegetation and zonation from Gittleman (1986a, 1986b) was tested against a coded value of infant mortality in captivity through a logistic regression (LOGIT). The response variable on a logistic regression has to be categorical, so to transform the dependent variable into a categorical variable, the upper and lower quartiles of *I* values for all the 98 species of carnivores in captivity was calculated, together with the median *I* for the order. If the specific *I* fell below the lower quartile of the order, it was coded 1. If it fell between the lower and upper quartile for the order, it was coded 2; and it was coded 3 if it fell above the upper quartile for the order.

Diet types were piscivorous; herbivorous; carnivorous; omnivorous; and insectivorous. Activity patterns were nocturnal; diurnal; crepuscular; nocturnal and crepuscular; and arrhythmic. Habitat vegetation categories were open grassland; forest; grassland and woodland; dense brush or scrub; desert; woodland; and aquatic. Types of zonation were aquatic; terrestrial occasionally arboreal; arboreal; terrestrial; and arboreo-terrestrial.

4.3. Results.

4.3.1. Reproductive biology and infant survival.

The variables that seem to have some influence on juvenile mortality rates are age of opening eyes (OE), gestation length (GL) and, with a weaker response, weaning age (WA). Gestation length values correlate both with weaning age (regression: $F_{1, 48} = 12.10$, $p = 0.001$) and with the age of opening eyes (regression: $F_{1, 47} = 10.45$, $p = 0.002$).

The variables OE and GL seem to have a linear effect on juvenile mortality rates in captivity, while WA almost presents a quadratic effect (GLM on transformed data: OE: $F_{1, 33} = 5.36$, $p = 0.027$; GL: $F_{1, 33} = 5.29$, $p = 0.028$; WA^2 : $F_{2, 33} = 3.48$, $p = 0.071$). However, this model will only explain 17.32% of the variance of infant mortality rates. Figure 4.1 shows the plotted the raw data used in this model.

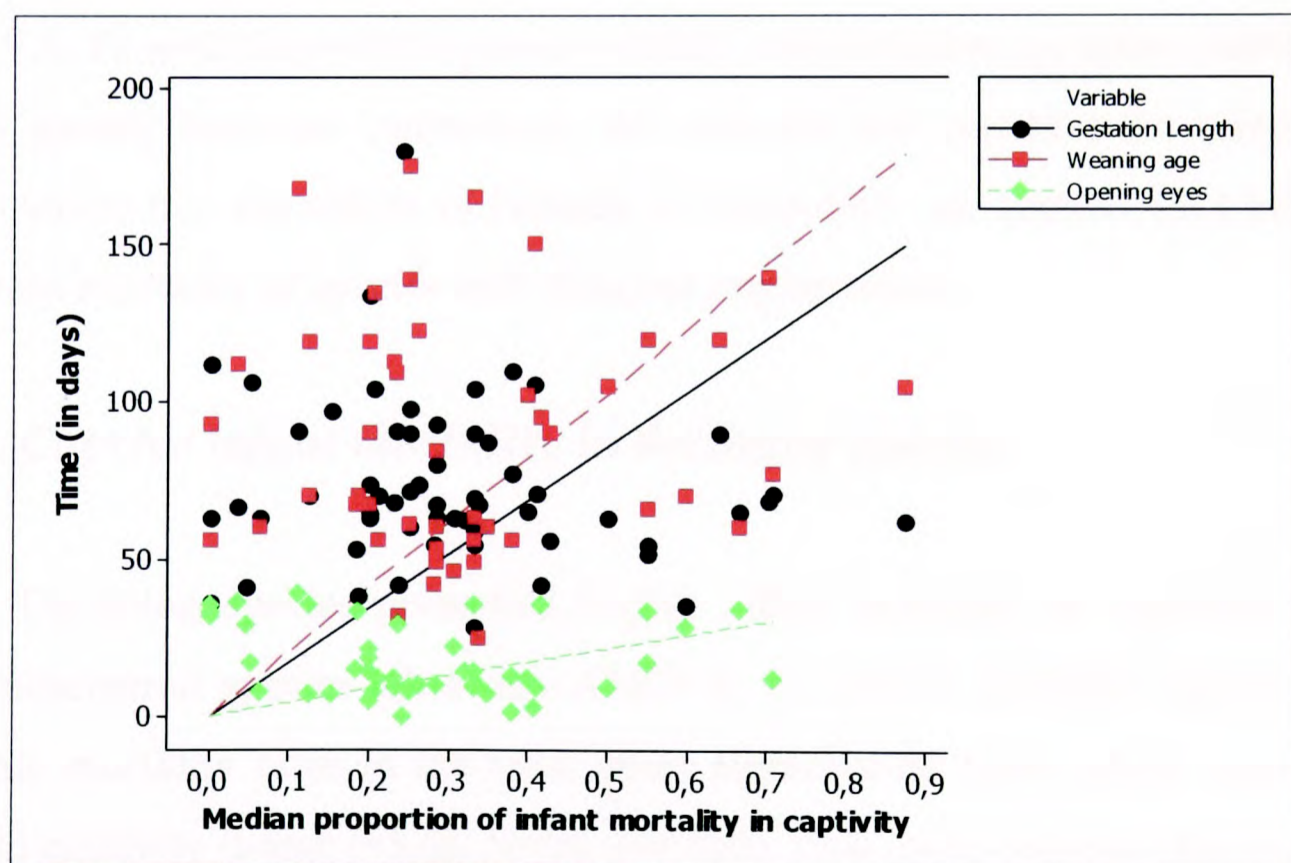


Figure 4.1: Developmental biology data of carnivore species plotted against the median proportion of infant mortality in captivity. Each colour represents a variable, and each dot represents a species. The species have values in all variables. Lines represent expected values.

Specific female body weight did not have a significant effect on the proportion of infant mortality in captive carnivores (One-way ANOVA: $F_{1,67} = 0.256$, $p = 0.5$) as it had on the juvenile mortality of carnivores in the wild presented by Gittleman 1983 (One-way ANOVA: $F_{1,14} = 12.72$, $p = 0.003$).

The possible, if weak, effect of these factors may indicate a tendency of higher mortality rates in altricial species, although a more complete database would be needed to enhance test power. It is important to remark, however, that many other factors may be affecting infant mortality, and it is unlikely that there will be a model to explain a much higher percentage of the variance.

4.3.2. Delayed implantation.

Overall, species with delayed implantation do not seem to be performing in captivity differently from those without this mechanism, although a larger sample size is needed, to increase test power (One-way ANOVA: $F_{4,25}=0.30$; $p=0.876$; $\text{power}=0.8225$). Nevertheless, as infant mortality varies greatly between institutions, this test did not consider the effects of photoperiod (for the effects of latitude of institution, see section 4.3.4 below) on infant mortality of species with delayed implantation.

4.3.3. Captive infant mortality in declining species.

Declining species presented higher infant mortality in captivity than non-endangered species (One-way ANOVA: $F_{1,96}=7.62$, $p=0.007$; Figure 4.1). Juvenile mortality rates in the wild differ significantly from infant mortality rates in captivity (t-test: $t=3.48$, $N=16$, $p=0.003$). This could indicate that factors affecting these values in the wild may be exacerbated by, or different to those of, a restricted environment.

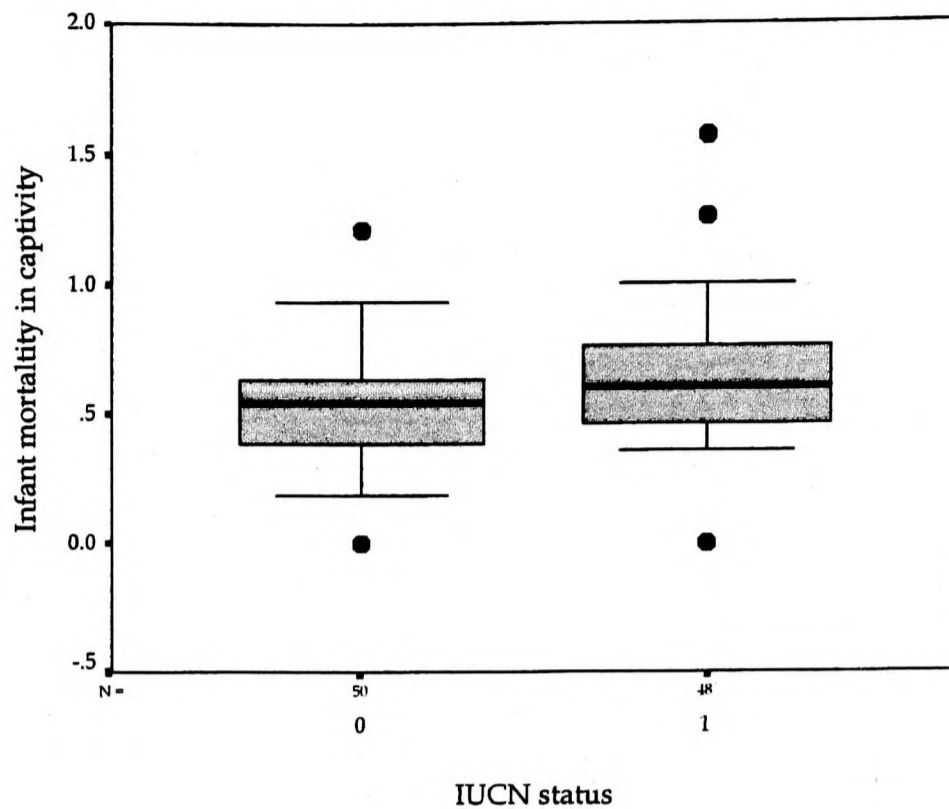


Figure 4.2: Infant mortality in captivity of non-endangered (N=49) and endangered (N=49) species of carnivores.

4.3.4. Photoperiod and latitudes.

Infant mortality was significantly higher when restricted distribution Arctic species (lower latitude of distribution $< 23^\circ$) were kept in institutions outside their natural range of distribution (One-way ANOVA: $F_{1, 812}=4.84$, $p < 0.03$), as it can be seen in Figure 4.2. Species with restricted distribution, but primarily tropical or subtropical, were also tested, and there was no effect of latitude in infant survival (One-way ANOVA: $F_{1, 756}=1.10$, $p > 0.1$; power = 0.96).

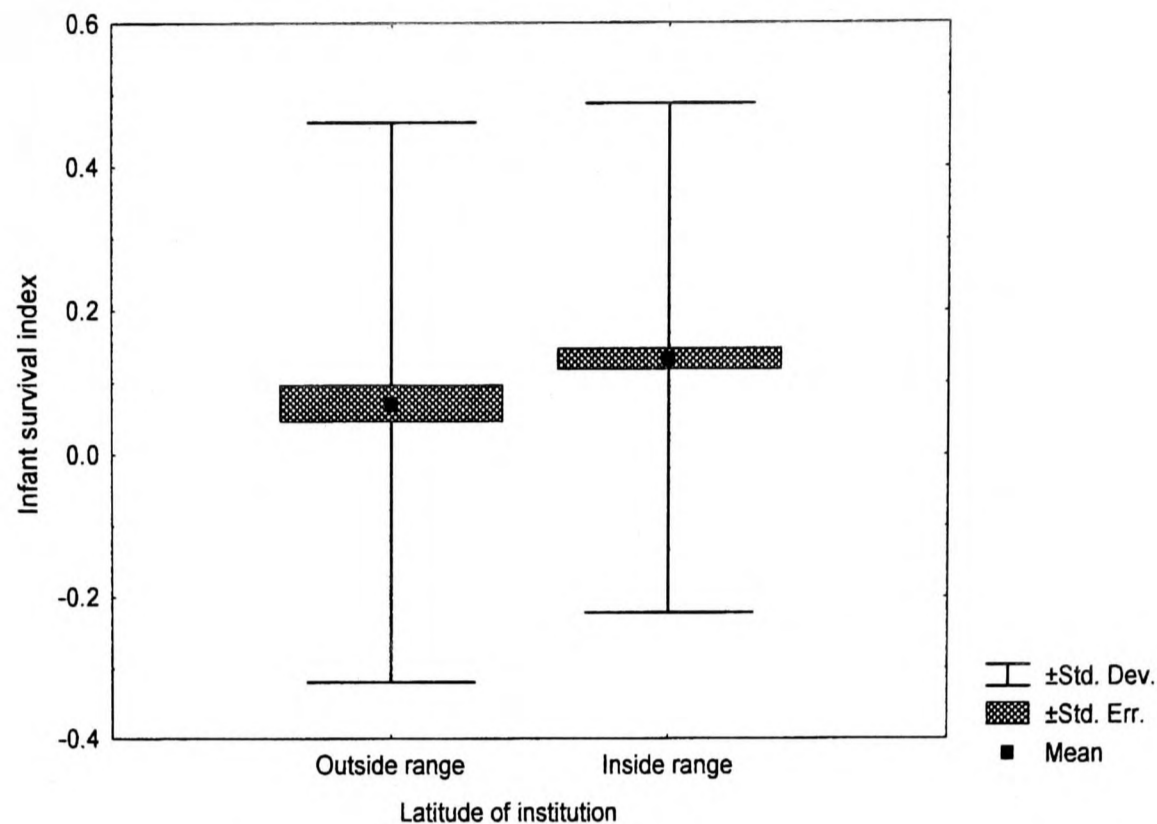


Figure 4.3: Index of infant survival of captive low distribution carnivores kept in institutions outside (N=240) and inside (N=572) their natural range of distribution.

4.3.5. Nutrition and Zonation.

There was a significant effect of type of diet (LOGIT: $Z_{64} = -1.97$, $p = 0.05$, odds ratio = 0.52) and zonation (LOGIT: $Z_{70} = -2.26$, $p = 0.02$, odds ratio = 0.46) on infant mortality in captivity. The type of diet of species with higher infant mortality was insectivorous, and piscivorous species performed better (Figure 4.3). Aquatic species suffered smaller infant mortality than all other types of zonation, and arboreal-terrestrial species presented higher proportion of young mortality (Figure 4.4).

There were not statistically significant effects of activity patterns (LOGIT: $Z_{64} = -1.51$, $p = 0.13$, odds ratio = 0.74) or habitat vegetation (LOGIT: $Z_{66} = -0.95$, $p = 0.34$, odds ratio = 0.91) of the species on infant mortality in captivity.

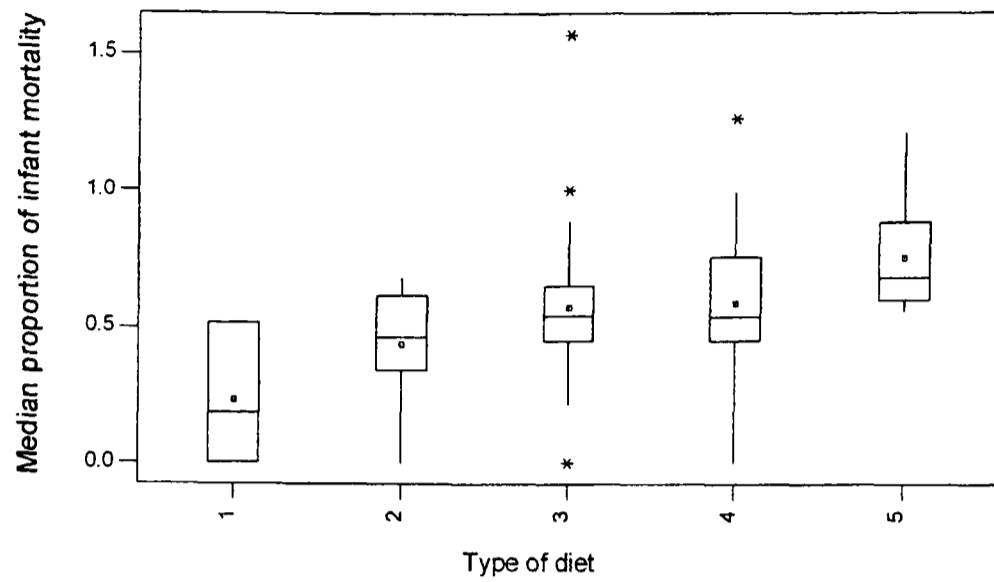


Figure 4.4: Median proportion of infant mortality according to the type of diet: 1= piscivorous (N=3); 2= Herbivorous (N=7); 3= carnivorous (N=28); 4= omnivorous (N=21); 5=insectivorous (N=9).

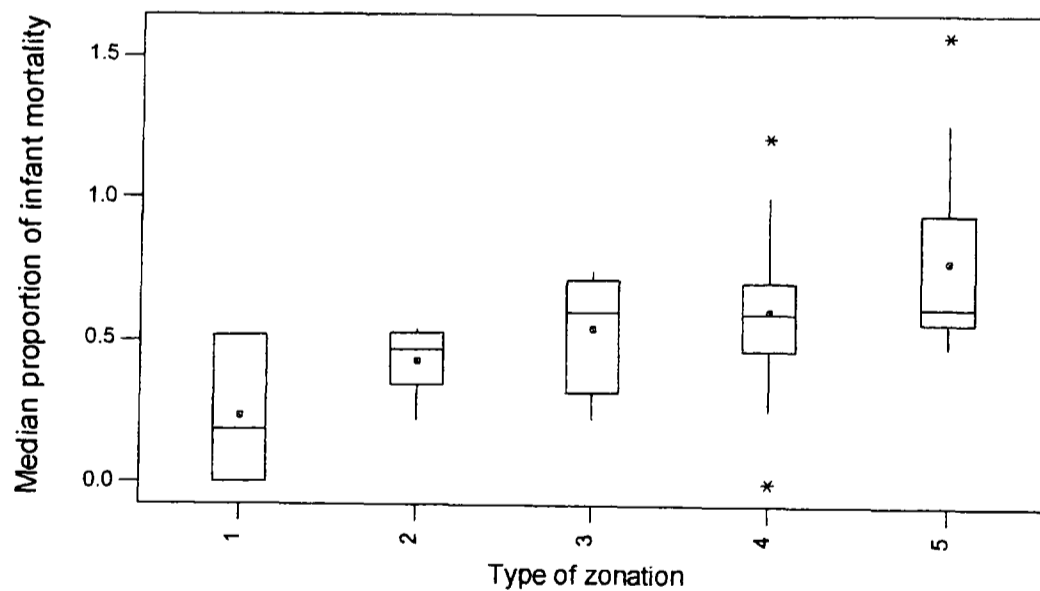


Figure 4.5: Median proportion of infant mortality according to the type of zonation. 1= aquatic (N=3); 2= terrestrial occasionally arboreal (N=7); 3=arboreal (N=4); 4= terrestrial (N=50); 5= arboreo-terrestrial (N=10).

4.4. Discussion.

4.4.1. Reproductive biology and infant survival.

The biology of reproduction is diverse within the Order Carnivora. As a result of the large morphological, ecological and behavioural variation in the order, some aspects of the breeding biology of carnivores can present extremely different values. For example, birth weight ranges from three grams to almost two kilos, litter size from one to eight cubs and lactation period from 30 to 730 days (Gittleman 1986). Databases containing specific information can allow a better understanding of dependent processes, such as the energy output during reproduction (Oftedal & Gittleman 1989), or supply models that may predict variation of certain phenomena, such as extinction risk in endangered species (Purvis *et al.* 2000).

Mortality rates, clustered by age or sex, have been suggested as a valid tool in the study of evolution and ecology of mammals (Promislow & Harvey 1991). Infant mortality rates (the proportion of young that died before certain age) are frequently used as an alternative to the traditional measure of breeding success (number of young per mature female per year), especially in large database research, where the number of females is not always available (cf. Courtenay 1988; Debyser 1995a, 1995b, 1995c). One advantage of using proportion of infant mortality instead of female productivity is that the latter method usually counts all females of the population studied, regardless of the fact that some females do not reproduce, either because of reproductive suppression or, in captive populations, because they are under contraceptive medication or do not have breeding opportunities.

In captive populations, infant mortality *per se* is a problem, and can jeopardise captive breeding programmes (cf. Chapter 1). Even though there is yet no detailed research on the most common causes of infant deaths in

captive carnivores³, diseases are reported to be the cause of death in many occasions (e. g. Geidel & Gensch 1976; Jayewardene 1975; Kitchener 1968; Olbricht & Sliwa 1997; Sausman 1997). Although the actual influence of maternal infanticide is not yet known, cannibalism and maternal neglect are profusely reported in several species (e. g. Coimbra-Filho 1966; Hess 1971; Leslie 1971; Michalowski 1971; Murphy 1966; Scheffel & Hemmer 1975). In captive cheetah, most of the mortality before 6 months of age results from maternal infanticide or neglect (Marker-Kraus 1997).

One aspect that may facilitate both the onset of diseases and the occurrence of infanticide⁴ is the level of altriciality of the young. Carnivores may be considered altricial if compared to ungulates (Grand 1992), but there is great variation within the order (Gittleman 1986). Having in mind that altricial young are more prone to die of exposure, contract diseases and be eaten by the dam (or by other adults) than precocial ones (Hayssen 1984; Hrdy 1979; Packer & Pusey 1984; Wolff 1997; Wolff & Peterson 1998), it is possible that species with more immature young will present higher levels of infant mortality in captivity, as it was found in this research.

4.4.2. Delayed implantation.

Delayed implantation (DI) is a mechanism through which an embryo is kept in diapause after fertilisation, allowing the gestation length to be longer than the expected for the female body weight. This pause can last from a few days to 10 months, and occurs in several species of four families of terrestrial carnivores (Mustelidae, Ursidae, Ailuropodidae and Ailuridae). The ecological importance of delayed implantation is still controversial, and in need of further investigation. It was suggested, though, that DI could favour litter survival, either through the synchronisation of births, in species with alloparental care of the young, or allowing the female to give birth in the

³ A detailed analysis of the causes of infant death in captive carnivores can be seen on Chapter 5.

⁴ For the aetiology of infanticide and its relation to altriciality, see Chapter 6.

beginning of spring, thus giving time for the young to be independent through winter (Mead 1989). Delayed implantation can also be affected by photoperiod, as in European badgers *Meles meles* (Woodroffe & MacDonald 2000), which varies with latitude and, consequently, with the zoological institution. The lack of a significant effect of delayed implantation in this research can be due to the stronger effects of other factors and may reflect the homogeneity of resources throughout the year for captive carnivores. Information on this aspect for a larger number of species, or more detailed data on the species with known delayed implantation, is needed to rule out any effect of this biological characteristic on infant mortality in captive carnivores.

4.4.3. Captive infant mortality in declining species.

Declining species are often subjects of conservation programmes in zoos, and much research is realised in institutions for the validation of biochemical tests, most commonly non-invasive techniques of monitoring the breeding status of individuals (Brown, Terio & Graham 1996; Graham *et al.* 1993; Monfort *et al.* 1997; Schwarzenberger *et al.* 1996; Whitten, Brockman & Staviski 1998). It was suggested that species listed on the IUCN Red List have particular biological aspects that increase the chances of a population not being able to sustain itself, such as a slow life history and small geographical range size. Furthermore, there can be an effect of human activity, speeding up the process of extinction. Juvenile mortality was not identified as one of the predicting factors (Purvis *et al.* 2000).

As much effort is put into the breeding success of declining species of carnivores in zoological institutions, and juvenile mortality does not appear to be a threat for these species in the wild, infant mortality in captivity should be homogeneous between endangered and non-endangered species, or even higher in the latter. As the results showed that endangered species, although

not presenting higher juvenile mortality in the wild, are performing worse in captivity, it may be the case that captivity conditions and husbandry protocols are influencing the breeding success in institutions.

4.4.4. Photoperiod and latitudes.

Changes in photoperiod are believed to be responsible for the regulation of several seasonal phenomena in mammals, especially in species from cold and temperate climates (Lincoln 1998). An example of how crucial photoperiodic regulation is for some species can be found in the lesser mouse lemur (*Microcebus murinus*). In this species, exposure to daylight for periods shorter than 12 hours leads to a general reduction of activity, sexual or behavioural, and the reverse happens when daylight lasts more than 12 hours. Individuals kept in a regimen of five months of long photoperiod followed by three months of short photoperiod ('accelerated seasonal rhythm') had a significant reduction in the number of months of their life span, although still living the same number of seasonal cycles (average of 5, with maximum survival of 9-10 cycles) as the individuals kept in normal photoperiod, although there was no effect on breeding success (Perret 1997).

Photoperiodic control of reproduction occurs in several species. For example, in the brush tail possum (*Trichosurus vulpecula*), changing photoperiod from long to short-day can hasten the onset of breeding activity, and it is advised for institutions involved in conservation to do so (Gemmell & Sernia 1995). Species that suffer this type of breeding control and are kept in institutions outside their natural distribution range can show inversion of season, as it is seen in Himalayan tahrs (*Hemitragus jemlahicus*). Captive specimens kept in the southern hemisphere have their breeding season six months apart from those kept in the northern hemisphere (Pare, Barrette & Prescott 1996). For some captive North Pacific pinnipeds, such as California sea lions (*Zalophus californianus*) and Pacific harbour seals (*Phoca vitulina*

richardsi), birth timing seems to be tightly regulated by photoperiod, and varies with the latitude of the institutions in which they are kept. In California sea lions, shorter birthing periods occur at higher latitudes. The effect is as subtle as a subtraction of 0.6 days in the pupping season for each degree of latitude added (Temte 1993).

Photoperiodic breeding control occurs through the action of pineal-secreted hormone melatonin, which is released during dark hours (Bittman 1993). The length of nocturnal melatonin secretion reflects changes in the photoperiod, and regulates the pulsatile secretion of gonadotrophin-releasing hormone (GnRH) from the hypothalamus, inducing changes in luteinizing hormone (LH) secretion. LH is responsible for the alternation of occurrence of ovulation in females, and influences sperm production in males (Malpaux, Thiery & Chemineau 1999). In mink (*Mustela vison*), seasonal variations in photoperiod interfere with the pulse frequency of LH release (Jallageas *et al.* 1994).

Other important hormonal change caused by photoperiod concerns the release of prolactin (Curlewis 1992), peptide that stimulates milk protein synthesis and growth of mammary glands, and that seems to play a crucial role eliciting maternal behaviour (Randall, Burggreen & French 1997). There is evidence of the action of prolactin in eliciting nesting behaviour in wolves *Canis lupus* (Mech *et al.* 1996), domestic pigs (Boulton *et al.* 1997; Lawrence *et al.* 1994), and domestic rabbits (Gonzalez-Mariscal *et al.* 2000). It was suggested the involvement of low levels of prolactin with lack of nursing behaviour, and infanticide (McCarthy, Curran & Siegel 1994; Peters, Sist & Kristal 1991). Thus, inadequate photoperiods could eventually lead to higher infant mortality, through poor nutrition and lack of maternal care.

In this research, infant mortality was higher when Arctic species (*Alopex lagopus*, *Vulpes vulpes*, *Lynx lynx*, *L. rufus*, *Uncia uncia*, *Ailurus fulgens* and *Ursus maritimus*) were housed in institutions located in subtropical or tropical areas. Tropical species did not present higher infant mortality when housed in

temperate areas. Photoperiodic-controlled environments have been used recently to manipulate reproduction in captive Palla's cats *Otocolobus manul* (Brown *et al.* 2002), black-footed ferrets *Mustela nigripes* and Siberian polecats *Mustela eversmanii* (Branvold, Biggins & Wimsat 2003), and results point out the urge to develop specific protocols to be followed by institutions involved in *ex situ* breeding programmes. The results of this research may indicate that artificially controlled photoperiods are extremely necessary for Arctic species housed outside of their range to breed successfully in captivity.

4.4.5. Nutrition and zonation.

To provide appropriate accommodation and nutrition to wild species is a constant concern for institutions. Inadequate nutrition can lead to severe health problems, and many physiological and behavioural problems can emerge from inappropriate enclosures (cf. Chapter 1).

The results found in this research point out to the fact that species with complex ecological demands, such as a diet of insects and a elaborate environment, have higher infant mortality in captivity than species with less complex demands, such as a diet of fish or aquatic habits. This may indicate the level of difficulty in the husbandry of these species and that the institutions are not yet giving appropriate conditions for young to thrive.

Nutritional deficiencies are usually a more immediate threat to the health and welfare of captive mammals. Highly carnivorous species, such as many felids, often do not receive enough calcium and vitamins from clean meat, i.e. with no cartilage, skin or fat, needing to eat whole carcasses to maintain health (Ashton & Jones 1979). Herbivorous and insectivorous species, whose low-calcium diet depends on food diversity, can suffer malnutrition because of the difficulty of providing a diverse and palatable diet in captivity (Donoghue & Langenberg 1994). In this research, nine insectivorous species (*Cynictis penicillata*, *Helogale parvula*, *Meles meles*, *Mephitis mephitis* *Mungos mungos*,

Otocyon megalotis, *Proteles cristatus*, *Suricata suricatta* and *Vormela peregusna*) presented higher levels of infant mortality. Insectivorous species require high levels of protein in captivity, and most of the dry food pellets in offer do not have appropriate protein contents. Institutions can usually raise one or two species of insects as live prey for insectivores, but this works more as a behavioural stimulus than a nutritional supplement (Price et al. 1999). Many institutions do not meet the nutritional requirements of many species with complex diets because of the lack of information on their natural feeding habit (Dierenfeld 1997) and this may be reflected in the results.

Carnivore species that occupy both terrestrial and arboreal environments also bred poorly in captivity. Seven species used in these analyses (*Bassariscus astutus*, *Leopardus pardalis*, *L. tigrinus*, *Martes flavigula*, *M. martes*, *Oncifelis geoffroyi* and *Prionailurus viverrinus*) are small and very mobile, requiring platforms and branches in the enclosure, while two (*Helarctos malayanus* and *Melursus ursinus*) have larger sizes, making the provision of climbing areas more difficult.

The conditions of accommodation can affect captive animals in many aspects. For example, most felids are terrestrial and use taller vegetation as shelter or observation points. In captivity, platforms or other elevated structures are usually the most utilised area of the enclosure. Individuals with access to these structures spent more time resting or observing the surroundings than those in flat enclosures (Lyons, Young & Deag 1997). It was suggested that enclosure type and conditions could indirectly influence reproduction, through modulation of stress, social maintenance, and occurrence of play behaviour (Carlstead & Shepherdson 1994).

The conditions of captivity can play a role in the reproductive output of these species, and it is important that specific demands are reached by the institutions in a way to reduce infant mortality. The costs of providing such conditions to highly-demanding species should be taken into account in the design of any captive breeding programme.

4.5. Conclusion.

Biological characteristics of species can serve as predictors of breeding success in captivity. Species with complex environmental and biological demands, which are difficult to be reproduced in constrained spaces and limited resources, presented higher infant mortality rates even when this is not true for wild populations. The actual effect of husbandry techniques is unclear, but differences on performance of the same species from one institution to the other suggest that some protocols may be more efficient than others. However, the permanence and homogeneity of resources in captive conditions may affect positively the reproductive output of species with delayed implantation, once the diapause would not be needed in these conditions.

If the aim of institutions is to drastically reduce infant mortality in captive carnivores, it may be necessary to review husbandry protocols, especially those concerning housing and dietary demands of the species. Special attention should be given to photoperiodic control in the enclosures of Arctic species, when housed in lower latitudes, and to the nutritional and spatial demands of the species. Some species, however, will be naturally more prone to die in younger ages due to their slower development, because they have a longer period in which they are exposed to several factors, such as diseases and conspecific aggression that may cause death.

Chapter 5

Causes of mortality in young captive carnivores.

Abstract

Data collected from published papers served to the analysis of the causes of infant deaths in captivity for 29 species of carnivores. In this dataset, 17% of young deaths had unknown causes. The three main causes of death are related to inadequate maternal behaviour (maternal cannibalism, infanticide and neglect). Around 9% of young died under hand-hearing attempts, and in the majority of cases the young were removed from the mother after displays of aggression or abandonment. In general, young died in the first days *postpartum* and mortality decreased substantially after one week. Perinatal mortality was caused by stillbirths and inadequate behaviour, and all deaths that occurred after 30 days were caused by infectious diseases. There is no statistically significant evidence that the median age of death of infants or their cause of death are related to the taxonomic family of the species. Species with larger home ranges seem to be slightly more prone to lose young due to infectious diseases. There was no evidence that occurrence of stillbirths was affected by the zonation of the species, although a larger sample would be needed to increase power. Stillbirths were significantly more frequent in species with piscivorous and carnivorous diets than in vegetarian, omnivorous or insectivorous species. Mortality caused by inadequate maternal behaviour, such as by infanticide or neglect, was not connected to the type of habitat preferred by the species, but it was significantly higher in species which prey on small and very small animals. Species with faster development, i.e. with shorter lactation and with young that open their eyes early have higher proportion of mortality caused by the dam, indicating the interruption of maternal investment in captivity. While diseases and stillbirths can be greatly reduced by prophylactic vaccination and proper nutrition, adequate maternal behaviour depends on the welfare of nursing females, which can only be reached by the provision of adequate conditions for each species.

5.1. Introduction.

As seen in the previous chapters, infant mortality in captive carnivores may represent a drawback to breeding programmes aimed at the conservation of declining species. Mortality of young captive carnivores is reportedly caused by several factors, from infectious diseases to inadequate maternal

behaviour, and research on the subject is usually descriptive rather than analytical (e.g. Marker & O'Brien 1989; Munson 1993; Maia & Gouveia 2002).

The investigation of the causes of infant mortality can be an important tool for understanding local threats to wild populations. For example, in one study on the causes of death of harbour seal pups in three regions in Washington, USA, several differences between populations were found. Neonatal mortality ranged from 12% to 26%, and the primary causes of death were predation by coyotes *Canis latrans*, premature parturition or starvation, depending on the location (Steiger *et al.* 1989). This type of information can help to find better solutions for the management of wild populations.

In this chapter, information on known causes of death in captive young carnivores is described and analysed in the search for factors that, if taken into consideration, could help alleviate the incidence of infant mortality of these species in captivity.

5.2. Causes of death of young wild carnivores.

It can be very difficult to determine the cause of death of carnivores in the wild. In a three-year study of 10 packs of African hunting dogs *Lycaon pictus* from the Kruger National Park, the cause of death was established only in a small number of cases: the two main causes found were predation by lions *Panthera leo* and disease, responsible for, respectively, 32.3 and 9.7% of the mortality in both adults and juveniles; several pathogens were found in over 95% of captured animals, and infant mortality exceeded 70% (Van Heerden *et al.* 1995).

To determine the mortality in neonates (up to 30 days of age) in the wild is very difficult for certain species, in special those of small size and that nest in burrows. For instance, in a study on breeding success of the meerkat *Suricata suricatta*, infant mortality was calculated from the time of emergence of the burrow until 6 months of age, since it is very difficult to observe the

actual number of young born in this species (Clutton-Brock *et al.* 1999). Other shy species may pose the same challenge. Gittleman (1983) compiled juvenile mortality data in wild carnivores, which is available for only 19 species, mostly terrestrial species with medium to large body sizes.

Research on the mortality of young carnivores in the wild has been yielding valuable information on the effect of ecological factors on population numbers. Resource competition, predation and human intervention appear to have a strong effect on infant mortality of young wild carnivores.

The availability of resources is related to juvenile mortality in several species of wild carnivores. In several populations of raccoon-dogs *Nyctereutes procyonoides* in Finland, the availability of berries is directly related to juvenile survival through autumn and winter (Kauhala & Helle 1995). Starvation was found to be the main cause of death of young wild Arctic foxes *laopex lagopus* in Sweden, and it was related to the decrease in numbers of microtine rodents, their primary prey (Sklepkovych 1989). Food scarcity was also responsible for most of black bears *Ursus americanus* cub deaths in Minnesota, USA (Rogers 1987).

Predation is a common cause of infant mortality in many ecosystems. Large canids, such as the coyote *Canis latrans* and introduced red foxes *Vulpes vulpes*, account for most of the few known deaths of juvenile and adult San Joaquin kit foxes *Vulpes macrotis mutica* in California (Ralls & White 1995). In Poland, reported causes of death of young wild badgers *Meles meles* were predation by raptorial birds and large carnivores, and in smaller scale road accidents and viral epidemics, especially rabies (Ruprecht 1996).

Intra-specific aggression seems to be the reason why female grizzly bears that harvest spawning trout at Yellowstone National Park, USA, lose more dependent cubs; high concentration of bears in trout-spawning areas increase the frequency of aggressive encounters with unrelated adults (Mattson & Reinhart 1995). Infanticide followed by cannibalism was registered in Alaskan brown bears *Ursus arctos*, though unrelated adults killed only four

cubs in 19 summers of observations (Hessing & Aumiller 1994). Infanticide by related adults and juveniles followed by cannibalism was also recorded in Arctic foxes, when there was a drastic reduction in prey numbers in Sweden, but it was an isolated event in five years of observations (Sklepkovych 1989).

Besides resource competition and predation, other ecological factors can lead to the loss of infants and adults alike. For example, the dens of polar bears occasionally collapse in exceptionally warm weather, leading to the death of the mother and cubs (Clarkson & Irish 1991). With the rise in global temperature, these events can become much more frequent because of the higher levels of rain in late winter; together with the increased nutritional stress, this would have a devastating effect to the polar bear *Ursus maritimus* populations, especially for those in the southern boundaries of the range, such as Hudson Bay (Stirling & Derocher 1993).

Anthropical activities, such as trapping, shooting and road accidents, are responsible for many juvenile deaths in species such as the red fox *Vulpes vulpes* in central Japan (Takeuchi & Koganezawa 1994), the introduced American mink *Mustela vison* and native polecats *M. putorius* in France (Lode 1995), European lynx *Lynx lynx* in Poland and Belarus (Jedrzejewski *et al.* 1996), and European badgers in Poland (Ruprecht 1996). Trapping or shooting can cause up to 98% of infant mortality in harvested populations, such as the raccoon *Procyon lotor* populations in Wisconsin, USA (Payne & Root 1986).

Captive populations are safe from many of the causes of mortality of wild carnivores, such as predation, lack of resources or hunting, so it is expected that infant mortality in captivity reach lower proportions than those found in wild populations. In fact, from 16 species that had known proportion of mortality both in the wild and in captivity, only the polar bear *Ursus maritimus* and the spotted hyaena *Crocuta crocuta* presented higher infant mortality in captivity than in the wild, and at least half of the institutions housing brown bears *Ursus arctos* and Canadian otters *Lutra canadensis* did not report a dead young from 1986 to 1996 (Table 5.1).

Table 5.1: Juvenile mortality in the wild (Gittleman 1983) and median proportion of infant mortality in captivity (IZY Vols. 26-35) in 16 species of carnivores.

Family	Species	Juvenile Mortality (Gittleman 1983)	Infant mortality in captivity
Canidae	<i>Canis lupus</i>	0.44	0.2
Canidae	<i>Vulpes vulpes</i>	0.76	0.33
Canidae	<i>Urocyon cinereoargenteus</i>	0.68	0.28
Ursidae	<i>Ursus arctos</i>	0.18	0
Ursidae	<i>Ursus americanus</i>	0.28	0.11
Ursidae	<i>Ursus maritimus</i>	0.20	0.67
Procyonidae	<i>Ailurus fulgens</i>	0.52	0.20
Procyonidae	<i>Procyon lotor</i>	0.42	0.20
Mustelidae	<i>Mustela erminea</i>	0.83	0.12
Mustelidae	<i>Mephitis mephitis</i>	0.66	0.31
Mustelidae	<i>Lutra canadensis</i>	0.46	0
Hyaenidae	<i>Crocuta crocuta</i>	0.16	0.38
Felidae	<i>Lynx lynx</i>	0.32	0.23
Felidae	<i>Lynx rufus</i>	0.53	0.06
Felidae	<i>Panthera leo</i>	0.67	0.40
Felidae	<i>Panthera tigris</i>	0.57	0.33

5.3. Causes of death of young captive carnivores.

In captivity, under generally controlled conditions, many external causes of mortality in neonates are eliminated, and the remaining causes can be determined more readily. For example, in Prague Zoo, Czech Republic, the reproduction of Pallas' cats *Otocolobus manul* was registered twice, but both litters died within two months due to parasitic or bacterial diseases (Volf 1999). In Denver Zoological Gardens, USA, four wild-caught adults Pallas' cats tested positive for *Toxoplasma gondii* antibodies, and six young born to them died of toxoplasmosis-related conditions (Kenny *et al.* 2002). In a survey of North American zoos, diseases were also responsible for a large number of deaths of young captive cheetahs *Acinonyx jubatus* (Munson 1993). Another survey on cheetahs, this time covering over 100 years of records, suggested a larger participation of behavioural aspects on infant mortality: several reports of death "by devouring" or "weakness" may point out to events of maternal infanticide and neglect (Marker & O'Brien 1989). A similar record research on

the maned wolf *Chrysocyon brachyurus* studbook revealed parental involvement in 67% of pup deaths, while infectious diseases were responsible for only 9% of the mortality (Maia & Gouveia 2002).

5.3.1. *Hypotheses to be tested.*

The causes of death of young carnivores in captivity are described in this chapter, and factors that can make certain species more prone to particular ailments are considered.

As seen in Chapter 4, life history traits can predict breeding success in captivity: altricial species that demand complex husbandry practices are more prone to lose young than easily manageable ones. There are indications that maternal infanticide is responsible for many infant deaths in captive carnivores (Bakken 1994; Laurenson 1993; Maia & Gouveia 2002; Marker & O'Brien 1989). As young in captivity are safe from most of the risks found in the wild, it is expected that inadequate maternal behaviour accounts for a large proportion of infant deaths in captivity. If so, most deaths shall occur in the first few weeks after birth, especially in species in which the young develop at a faster pace, due to the high costs of maternal care (Clutton-Brock 1991; Hrdy 1979; Labov *et al.* 1985). Species with large home ranges are heavily affected by captivity (Clubb & Mason 2003) and may be predisposed to contract diseases through poor nutrition or poor welfare.

5.4. *Methods.*

The dataset used for this analysis was the bibliographical dataset collected from 141 published papers (Appendix 4, described in section 2.2.5), which served for the analysis of causes of death in 29 species. Relative frequencies of known causes of death were calculated for each female and the median proportions were calculated for the species, being used as dependent variables in regressions. Average litter sizes, average age of breeding and average age of death of young were calculated for each female. Multiple analyses were performed using variables chosen through stepwise regression, forward method, from the datasets compiled by Gittleman (1983, 1986a, 1986b): juvenile mortality, home range sizes, age when young open their eyes, weaning age, type of vegetation in the habitat, type of diet and type of zonation. All proportional values were transformed by the arcsine of the square-root and other continuous variables were log-transformed. Table 5.2 displays the species used in the analysis of this chapter.

Table 5.2: Species used in the analysis of causes of death of young captive carnivores, with number of litters, dams, zoos holding the species, young born and young dead. The data was collected from 141 published papers (Appendix 4).

Family	Species	Litters	Dams	Zoos	Young born	Young dead
Canidae	<i>Canis lupus</i>	9	2	2	33	14
Canidae	<i>Cerdocyon thous</i>	6	4	2	25	9
Canidae	<i>Chrysocyon brachyurus</i>	20	13	8	49	19
Canidae	<i>Vulpes zerda</i>	10	7	3	23	8
Canidae	<i>Lycaon pictus</i>	4	2	2	24	8
Felidae	<i>Acinonyx jubatus</i>	22	15	12	66	17
Felidae	<i>Caracal caracal</i>	5	4	2	14	6
Felidae	<i>Felis margarita</i>	10	2	2	42	24
Felidae	<i>Felis nigripes</i>	8	3	2	13	8
Felidae	<i>Felis silvestris</i>	29	7	3	107	58
Felidae	<i>Leopardus tigrinus</i>	7	3	2	9	3
Felidae	<i>Lynx lynx</i>	10	7	3	23	8
Felidae	<i>Neofelis nebulosa</i>	15	5	4	31	14
Felidae	<i>Oncifelis geoffroyi</i>	15	5	3	34	17
Felidae	<i>Prionailurus bengalensis</i>	8	2	2	21	7
Felidae	<i>Uncia uncia</i>	11	5	4	28	17
Felidae	<i>Panthera tigris</i>	9	7	5	26	25
Herpestidae	<i>Helogale parvula</i>	14	3	2	51	20
Hyaenidae	<i>Crocuta crocuta</i>	4	3	3	6	2
Mustelidae	<i>Amblonyx cinereus</i>	11	3	3	28	21
Mustelidae	<i>Enhydra lutris</i>	9	5	4	9	9
Mustelidae	<i>Pteronura brasiliensis</i>	12	2	1	33	26
Mustelidae	<i>Meles meles</i>	2	2	1	3	1
Procyonidae	<i>Ailurus fulgens</i>	9	5	3	16	8
Ursidae	<i>Ailuropoda melanoleuca</i>	11	7	5	15	10
Ursidae	<i>Helarctos malayanus</i>	12	5	3	12	1
Ursidae	<i>Tremarctos ornatus</i>	11	4	4	17	12
Ursidae	<i>Ursus maritimus</i>	11	4	4	21	16
Viverridae	<i>Arctictis binturong</i>	13	5	5	33	12

5.5. Results.

5.5.1. Causes of death and age of young.

In this dataset, the cause of death was unknown in an average of 17% of young. Throughout 25 species, the three main causes of death are related to inadequate maternal behaviour (maternal cannibalism, infanticide and neglect), as it can be seen in Figure 5.1. Around 9% of young died under hand-rearing attempts, and in the majority of cases the young were removed from the mother after displays of aggression or abandonment. Most reports did not provide the exact date of removal of the young for hand-rearing.

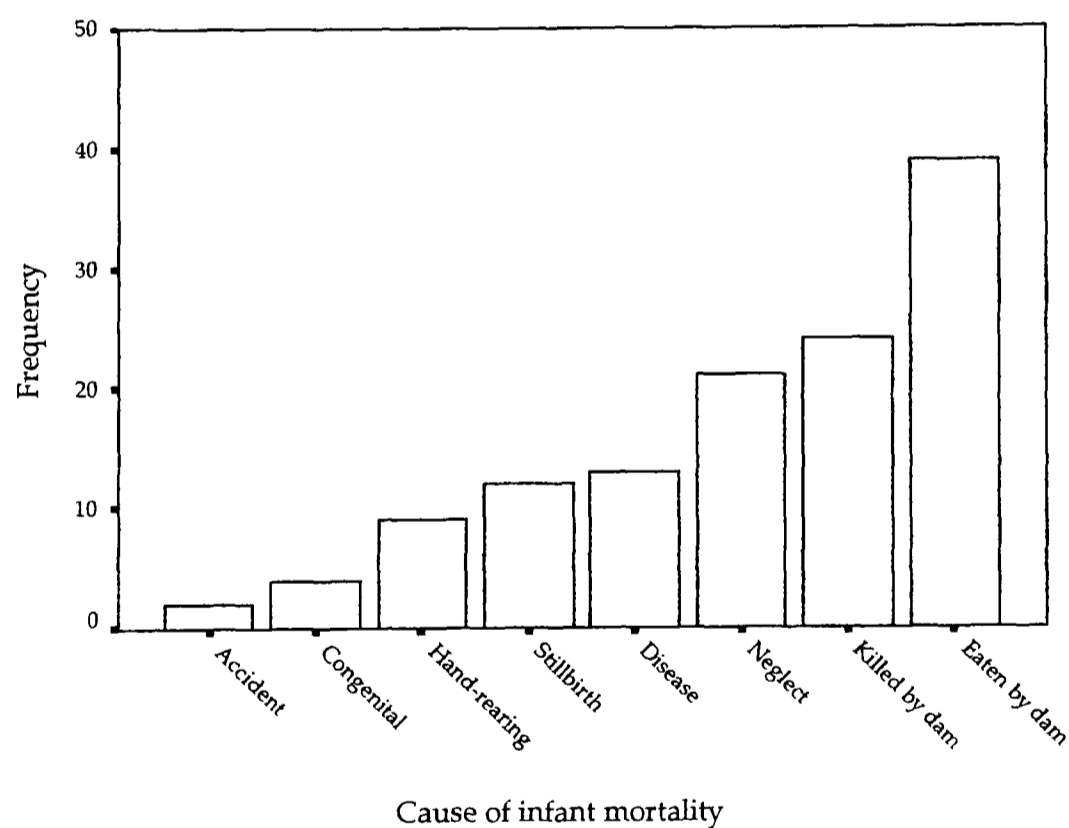


Figure 5.1. Absolute frequencies of cause of death in young captive carnivores (N = 364 young of 29 species).

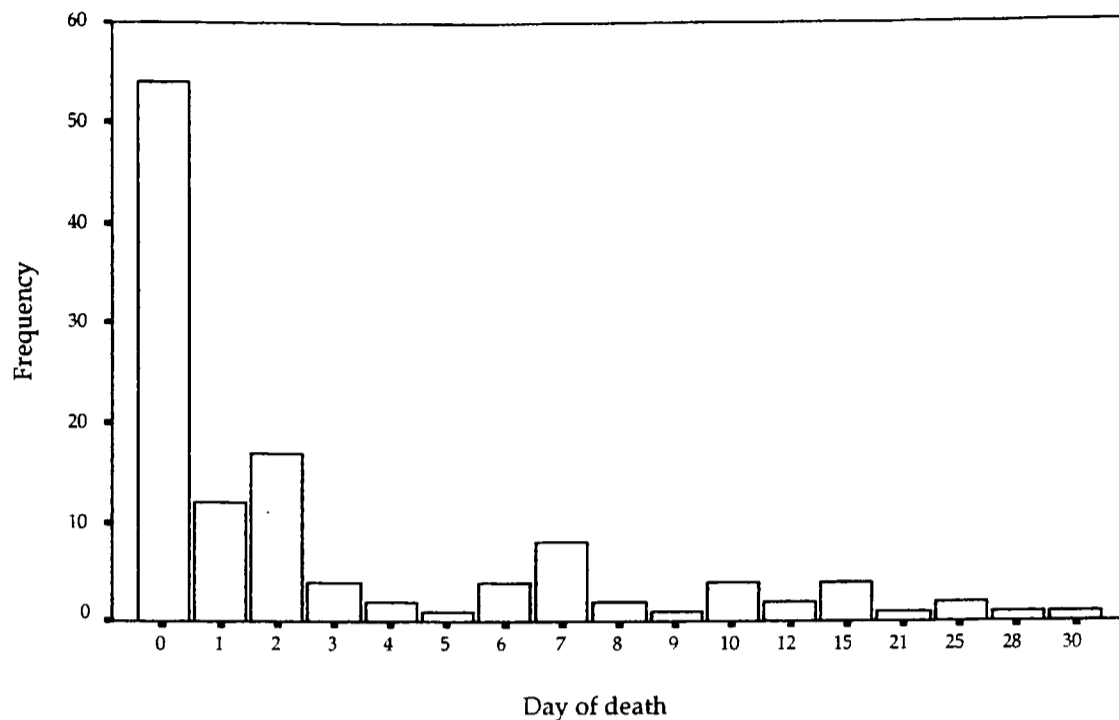


Figure 5.2. Absolute frequencies of age (in days) of death of young captive carnivores (N = 364 young of 29 species).

In general, young died in the first days *postpartum* and mortality decreased substantially after one week (Figure 5.2). *Postpartum* mortality was mainly caused by stillbirths, and inadequate maternal behaviour accounted for deaths up to two weeks of age (Figure 5.3); all deaths that occurred after 30 days were caused by infectious diseases (One-way ANOVA: $F_{3,60} = 10.66$, $p = 0.000$). Reports did not specify when the young were removed for hand-rearing.

There is no statistically significant evidence that the median age of death of infants (One-way ANOVA: $F_{5,74} = 0.98$, $p = 0.44$, power = 0.71) or the cause of death (LOGIT: $Z_{126} = -1.79$, $p = 0.21$, odds ratio = 0.66) are related to the taxonomic family of the species.

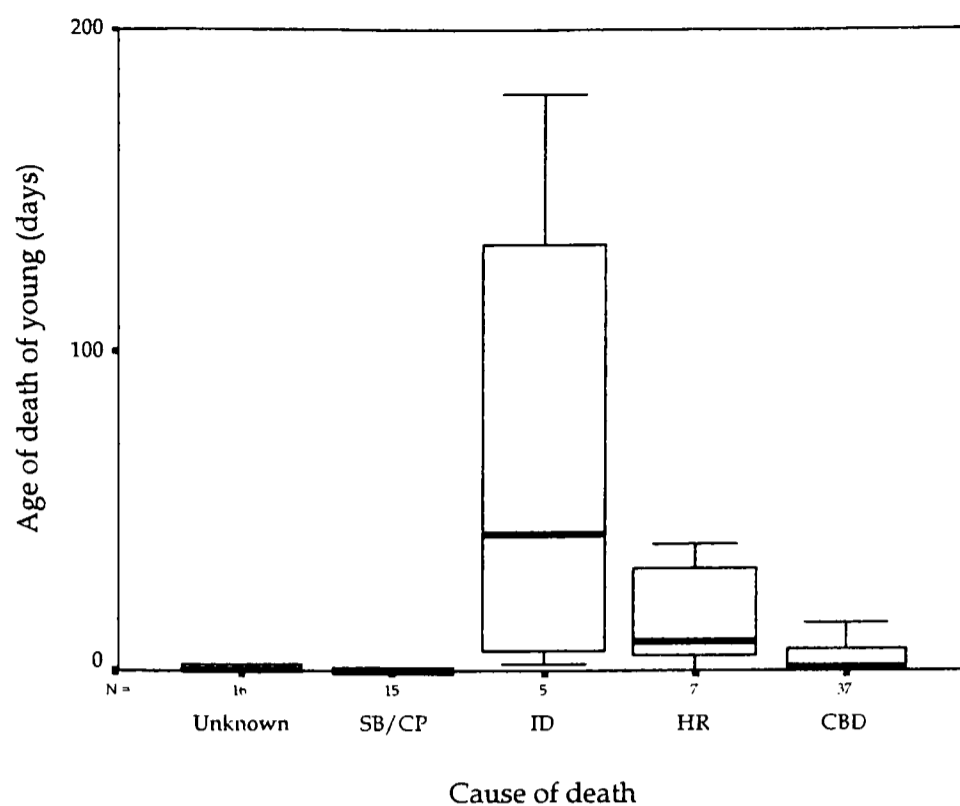


Figure 5.3: Age of death of young captive carnivores in days according to the cause of death, N = 364 young of 29 species. SB = stillbirth; CP = congenital problems; ID = infectious diseases; HR = under hand-rearing; CBD = maternal involvement (infanticide or neglect).

5.5.2. *Ecological factors as predictors of the cause of death of young captive carnivores.*

In captivity, carnivore species with larger home ranges seem to be slightly more prone to lose young to infectious diseases, and the regression explains 50% of the relation (Regression: $F_{9} = 6.07$, $p = 0.06$, $R^2 \text{ adj.} = 50\%$, figure 5.4). There was no evidence that occurrence of stillbirths was affected by the zonation of the species, although a larger sample would be needed to increase power (One-way ANOVA: $F_{5, 14} = 2.38$, $p = 0.09$, power = 0.26). However, stillbirths were significantly more frequent in species with piscivorous and carnivorous diets than in vegetarian, omnivorous or insectivorous species (One-way ANOVA: $F_{4, 12} = 3.47$, $p = 0.04$).

Mortality caused by inadequate maternal behaviour, such as by infanticide or neglect, was not connected to the type of habitat (open land, sparse vegetation, dense vegetation or aquatic) preferred by the species (One-

way ANOVA: $F_{3,16} = 0.89$, $p = 0.4$), but it was significantly higher in species which prey on small and very small animals (One-way ANOVA: $F_{2,14} = 7.20$, $p = 0.05$).

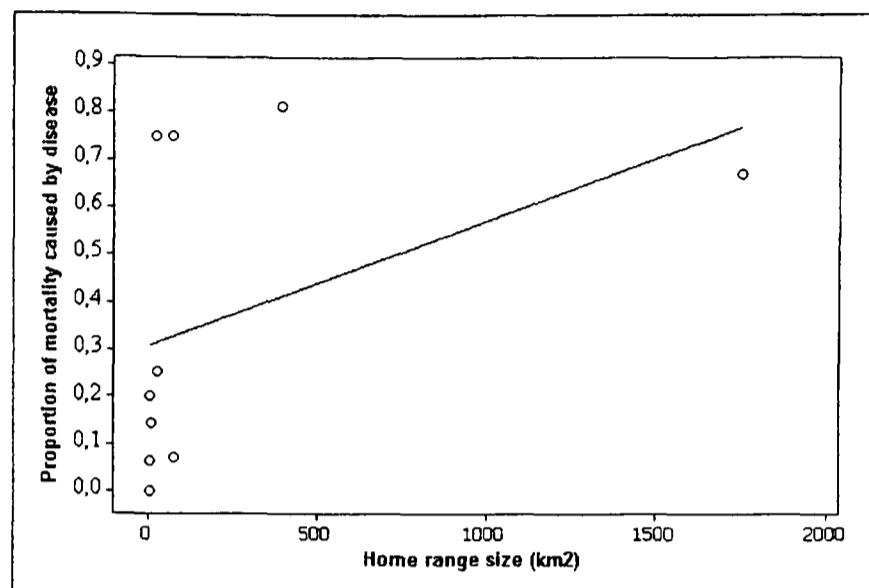


Figure 5.4: Graph of home range sizes (in km²) versus the occurrence of fatal infectious diseases in infants of 10 species of carnivores (data not transformed).

Weaning age (WA) seemed to have a negative effect on the proportion of mortality caused by the dam in this dataset, and a weak effect of the age in which the young open their eyes (OE) was also detected; the model explains 42% of the variance (GLM on transformed data: WA: $F_{1,13} = 609.15$, $p = 0.032$; OE: $F_{1,10} = 6.68$, $p = 0.073$, Figure 5.5).

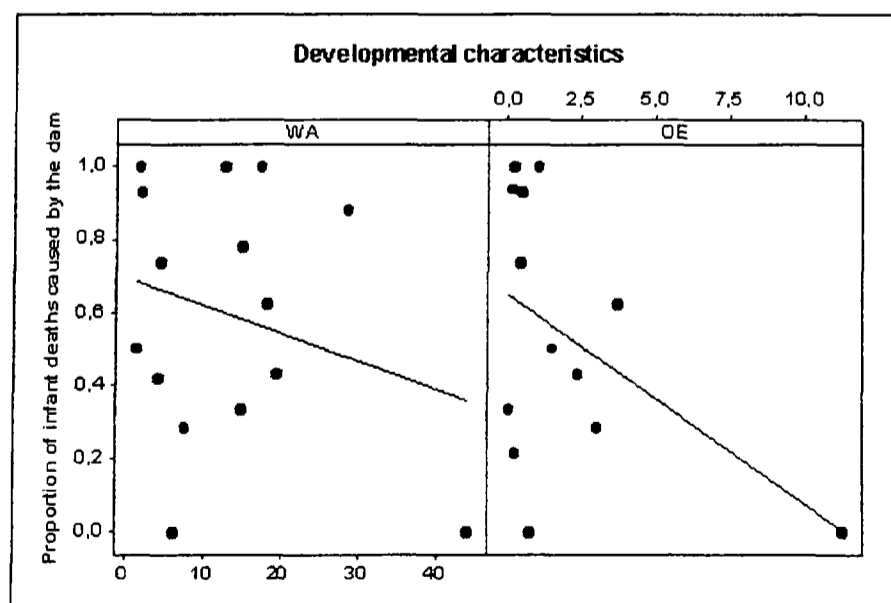


Figure 5.5: Scatterplots of developmental characteristics (WA = weaning age in weeks; OE = age of opening eyes in days) of 15 species of carnivores in relation to the occurrence of infant deaths caused by the dam in captivity. The data plotted was not transformed.

5.6. Discussion.

It seems that infant mortality in captive carnivores is frequently caused by inadequate maternal behaviour, so its several aspects will be analysed in detail in Chapter 6.

In this dataset, the young that survived birth were the most vulnerable to all causes of death during the first two weeks. In the wild, the observation of young carnivores is the most difficult during this period, and therefore most researches do not analyse this aspect. In captivity, however, similar results to this research were found: a survey on the mortality of captive African hunting dogs *Lycaon pictus* from 1983 to 1995 showed that the majority of infant deaths (57.5%) occurred before the end of the first week, decreasing to 14.9% between two weeks and 1 year of age (Van Heerden *et al.* 1996).

The actual number of stillbirths and the factors that may influence it in the wild is largely unknown. However, the phenomenon has been recorded frequently in captive carnivores. In farmed blue foxes, stillbirths occurred in 5.9% of the births, and 11.4% of the young died of infectious diseases before weaning (Ilukha, Harri & Rekila 1997). In domestic dogs of the Boxer breed, stillbirths and infections accounted for the majority of deaths (van der Beek *et al.* 1999).

The analysis of the present dataset suggested a relation between the type of diet of species and the occurrence of stillbirths. Nutrition can be an important factor to the health of young carnivores. An experiment compared weight gain and growth rates of young caged mink fed with products made with different technologies of preservation; kits fed with food prepared with a new technique of preserving raw animal by-products through fermentation decreased in weight and had lower growth rates compared to the ones fed with the traditional products (dried pellets) or fresh animal products (Urlings *et al.* 1993). In foxes, taurine deficiency causes the heart to dilate, and may lead

to decreased breeding success and even death, being the effect more pronounced in younger animals (Moise *et al.* 1991).

Nutritional deficiencies can also facilitate stillbirths by depressing immune responses and allowing the onset of infections. For example, canine herpesvirus and feline herpesvirus-1 cause abortion or neonatal deaths in domestic dogs and cats, and may infect, many times undetected, wild related species (Smith 1997). Experiments with mink *Mustela vison* and ferrets *Mustela putorius* revealed that females contaminated by *Campylobacter jejuni* either aborted the kits or had stillbirths, even when they survived previous infections with the same bacteria, mainly because of the development of severe placentitis (Bell & Manning 1990).

Certain diseases are widespread in domestic animals but can affect wild carnivores, leading to lower breeding success. In farmed silver foxes *Vulpes vulpes*, the presence of parvoviral antibodies was found to cause small litter size and to facilitate the infection of cubs by other microorganisms; treating affected females with adequate antibiotics significantly reduced the number of abortions and neonatal deaths (Mizak, Rzezutka & Matras 1998).

Diseases were the only cause of death after the critical period of 30 days. Many pathogens can cause septicaemia in young carnivores, but very little is known about the epidemiological mechanisms in wild species, even in captivity. All research in this area has been done in domestic species and those farmed for commercial purposes.

In domestic dogs, neonatal mortality is usually related to septicaemia caused by pathogenic microorganisms such as staphylococci and streptococci, which cause sudden death within the first week after birth, frequently without any previous symptom. Researchers developed a metaphylactic protocol to identify the presence of any pathogen in vaginal smears of the mother and the facilities where the animals are, 20 days after birth, and appropriate antibiotics are given to the mothers for 7 days before the calculated date of birth; puppies receive the same treatment for 3 days after birth, and mortality from

septicaemia was almost null in treated animals (Miljkovic *et al.* 1995). Research in Nigeria connected infant sudden death in domestic dogs to acute systemic infections by the Infectious Canine Hepatitis (ICH) virus, which killed puppies up to 12 weeks old without previous symptoms (Ojeh *et al.* 1989). It was suggested that inbreeding levels in domestic puppies had a very strong effect on the occurrence of neonatal infections (van der Beek *et al.* 1999).

Microorganisms can lead to asymptomatic deaths, and viral infections can affect infant survival even before they can be detected by any tests, and therefore may not be diagnosed as the cause of death in many occasions. Experiments with domestic cats showed that early infection with FIV, which is transmitted from the mother to kittens on birth or through nursing, may lead to neurological damage and poor growing within 12 weeks of birth, even before the full development of immunodeficiency (Johnston *et al.* 2002). Also, weanling kittens infected with feline leukaemia virus, which is widespread in domestic cats and eventually causes death, consumed and spent less energy, lost weight and developed a permanent growth impairment as early as the 4th day after inoculation, before the phase in which it can be detected by tests and much before the development of the bone marrow stage of the infection (Hartke *et al.* 1995).

Nowadays, most wild species can be vaccinated against domestic species common diseases, and there are alternative therapies for diseases that do not have specific vaccines. For example, one of the main causes of death in captive newborn mink kits is pneumonitis, or Aleutian disease, caused by the Aleutian Disease Virus (ADV). Although an ADV epidemic can be responsible for a large number of deaths in farmed mink, it can be controlled by the administration of antibody-specific gamma globulin, which is particularly effective in highly infected individuals (Aasted, Alexandersen & Hansen 1988).

In this dataset, there is a weak effect of the species average home range size in the frequency of infant mortality caused by diseases. This could be related to the effect of housing conditions in the health of individuals. Housing

conditions are known to affect breeding success in carnivores (Carlstead & Sheperson 1994) and improper keeping can lead to poor welfare (Carlstead 1996; Clubb & Mason 2004). It was also found that species with large home ranges are more prone to develop stereotypies and breed poorly (Clubb & Mason 2003). Stereotypies not only reduce general welfare of the individuals but can lead to sometimes severe friction wounds (Spendrup & Larsson 1998).

Some characteristics of the enclosure, such as type of flooring and access to hygiene can also affect the health of the animals. Necropsies and histopathologic analysis showed that most red wolf *Canis rufus* pups had infected lesions in their feet, caused by the combination of inadequate substrate and poor hygiene (Acton, Munson & Waddell 2000). Depending on the design of the enclosure, perfect hygiene cannot be performed and parasites can be a permanent problem, re-infecting treated animals and infecting young at birth (Fowler 1996).

The influence of inadequate maternal behaviour in captive carnivores has been noted in other studies, but results can be contradictory. For instance, in one study maternal infanticide was found to be the main cause of mortality in farmed silver fox cubs, occurring in 75.9% of the deaths (Braastad & Bakken 1993); other research showed that only 0.3% of the infant deaths in silver foxes were caused by the dam (Ilukha, Harri & Rekila 1997). Infanticide in this species is highly related to social competition: infanticidal females that were removed from the view of conspecifics raised their young normally (Bakken 1992, Bakken 1993), showing that husbandry decisions may be crucial to the occurrence of maternal infanticide.

It can be difficult to record deaths by maternal aggression when the event was not observed, and maternal neglect can be only assumed in *post-mortem* examination when the young display signs of starvation and general lack of care. In captive African hunting dogs, the cause of death could not be determined in the majority of cases of neonatal deaths, but it was supposed that around 42% of neonatal mortality was caused by exposure. Around 15%

of the dead animals, including both juveniles and adults, presented signs of trauma caused by conspecifics (Van Heerden *et al.* 1996).

It has been suggested that carnivores are more prone to commit infanticide due to their carnivorous diet and altricial young (Packer & Pusey 1984). In this dataset, species that prey on small and very small animals presented higher proportion of deaths caused by of inadequate maternal behaviour, independent of which form it takes. Analysis also suggests the influence of prey size in active maternal infanticide, as will be discussed in Chapter 6.

In Chapter 4 it was suggested that species with altricial young present higher levels of infant mortality in captivity, because species with slower development are exposed to all causes of mortality, including those cause by the dam, for a longer period. In fact, this study suggests that species with faster development, i.e. that open their eyes and wean earlier, are more prone to die due to inadequate maternal behaviour, and that most of these deaths occur in the first weeks after birth. These results may reflect the mechanism of brood reduction, where maternal investment is reduced or interrupted to enhance either the fitness of the surviving young or the female's own capability for further breeding (Clutton-Brock 1991). Maternal investment is higher during lactation and young can especially demanding in small felids (Ofstedal & Gittleman 1989); maternal infanticide and neglect were the main causes of death, in this research, in captive populations of *Caracal caracal* (100%), *Oncifelis geoffroyi* (88%), *Leopardus tigrinus* (67%), *Felis margarita* (42%) and *Felis nigripes* (38%). This type of strategy is commonly used when resources are low or there is the imminent danger of losing the brood (Clutton-Brock 1991), conditions which supposedly do not exist in captive conditions. It is possible that some species perceive captive environments as sub-optimal and react by interrupting reproductive investment, but further research, considering each species' characteristics, would be needed to fully understand the adaptive strategies involved.

5.7. Conclusion.

Infant mortality in captive carnivores occurs usually within a critical period before 30 days of age; after this period, the young are still vulnerable to infectious diseases, but many of those can be prevented through vaccination of other prophylactic measures. Certain characteristics of the species can make them more prone to some ailments, through poor nutrition or inadequate housing. Species that predate on small animals and have high-maintenance young have a tendency to display inadequate maternal behaviour, such as active infanticide and abandonment, and may perceive captive conditions as sub-optimal, thus interrupting maternal investment. Further research is necessary to understand the adaptive value of this strategy for each species and to develop possible solutions to reduce infant mortality in captivity.

Chapter 6:

Maternal infanticide in captive carnivores

Abstract

Most of infant mortality in many captive carnivores is caused by inadequate maternal behaviour. A dataset collected from 141 published papers yield information on the frequencies of three types of maternal behaviour fatal to infants (cannibalism, active infanticide and abandonment) in 29 species of carnivores. These values were tested against life history traits that could be predisposing species to perform these behaviours more frequently. Species that prey on larger animals tended to kill but not consume the young, but there was no significant evidence that species with smaller prey do consume the young. Species with slower growing litters showed a higher incidence of active infanticide without cannibalism.

Species weaning age and the age in which the young open their eyes did not influence the occurrence of active infanticide, followed or not by cannibalism. The proportion of passive infanticide reported for each species was strongly influenced by interbirth interval, but there was no significant effect of weaning age. Overall, wild-caught females presented slightly higher levels of maternal infanticide, passive or active, than captive-born ones independent of the species. Throughout the order Carnivora, there was a slight effect from the origin of the female (if wild-caught or captive born) in the type of infanticide: wild-caught females performed more passive infanticide and captive-born performed more active infanticide. The activity pattern of the species had a significant effect in the type of infanticide performed, with arrhythmic species presenting higher levels of active infanticide and diurnal species neglecting young more often. Overall, there was no statistically significant effect of the presence of the male in the incidence of maternal infanticide in species with exclusive maternal care (i.e. when the male does not care for the young), but a larger sample would be needed to increase test power. Brood reduction strategies seem to differ greatly between species of carnivores in captivity, but the decision on curtailing maternal investment may depend on the female's welfare, which can be affected by individual histories and husbandry practices.

6.1. Introduction.

Maternal behaviour seems to play a crucial role in the survival of captive-born carnivores, for most of the reported infant mortality is caused by maternal aggression or neglect, especially in species with expensive maternal care, such as small felids (Chapter 5). To control the impact of these phenomena in captive breeding, it is necessary to understand the adaptive role of infanticide and the conditions in which it is more prone to occur.

Infanticide, by related or unrelated adults, has different adaptive values depending on specific social and environmental conditions, as can be seen in Table 6.1; in the wild, maternal infanticide can occur under many conditions, as long as social or ecological pressures drive the females to opt for it (Hrdy 1979).

Maternal infanticide occurs when the female kills her own offspring, and was recorded in the wild in some species of the Order Carnivora, such as lions *Panthera leo* (Rudnai 1973), polar bears *Ursus maritimus* (Van Keullen-Kromhout 1976) and red foxes *Vulpes vulpes* (MacDonald 1980). It has been suggested that females may use this strategy to avoid further investment in a litter that will probably not survive (Hrdy 1979; Hausfater & Hrdy 1984). Certain species use maternal infanticide largely as a means of manipulating litter size or sex ratio (Labov *et al.* 1985; Wolff 1997).

The role of inadequate maternal behaviour in parental manipulation has been discussed only in recent years. Hrdy (1979) and Bakken (1994) pointed out that parental manipulation might occur at any level during the breeding process, from the destruction of gametes and abortion to the neglect or killing of young, the last two being caused by inadequate maternal behaviour. Maternal infanticide presents, then, both passive (neglect) and active (killing) expressions. The mother may ignore the young and allow them to die of starvation or hypothermia, or kill her young by direct action, such as biting or removing the young from the nest (Hayssen 1984). Both mechanisms can be

applied in maternal litter manipulation - where selected youngsters are killed while others are raised normally - or under severe lack of resources or threat of predation - through the termination of the whole litter (Clutton-Brock 1991). The benefits of these strategies may depend on three factors: a) the ability of the parents to produce another litter in the same season; b) the cost of having another litter to the parent's reproductive value; and c) on the probability of these youngsters, raised under adverse conditions, to breed when adults (Hayssen 1984).

The female will, in many cases, reabsorb nutrients by eating the young (Hrdy 1979). Some authors call the cannibalisation of the offspring Cronism, in reference to mythological Greek god Kronos, who ate all his children (In MacFarland 1981: 55-58). There is evidence in nature, although scant, of maternal infanticide followed by cannibalisation. For example, remains of partially eaten cubs were found in the frozen floor of a den of polar bears (*Ursus maritimus*) in the wild (Van Keulen-Kromhout 1976). As seen in Chapter 5, many Carnivora species have nests in burrows, are nocturnal and/or are extremely cautious of humans, and there are not many observations of these behaviours in the wild. Packer & Pusey (1984) reviewed the occurrence of infanticide in carnivores, with special focus in male infanticide strategies. There are some observations of infanticide in wild large carnivores, such as lions and brown bears, but it is related only to litter abandonment; active maternal infanticide was not mentioned in this research, but the authors point out for the lack of data on nocturnal and solitary species (Packey and Pusey 1984).

Until the 1980's, maternal infanticide was generally regarded as an abnormal or "unnatural" behaviour displayed only under the stress of captivity or extreme crowding (Calhoun 1962; Lorenz 1966; Labov *et al.* 1985), mostly due to the lack of research in the wild. For example, one of the causes of maternal infanticide in captivity was thought to be the low level of domestication in some species, such as silver foxes *Vulpes vulpes* (Bakken

1994). The occurrence of infanticide in domestic species such as rabbits *Oryctolagus cuniculus* (Sambraus 1985), pigs *Sus scrofa* (Lammers & De Lange 1986), and cats *Felis catus* (Feldman 1993) seems to disprove this hypothesis.

Infanticide is said to be more prone to occur among carnivores, perhaps because of their predatory feeding habits and altricial young (Packer & Pusey 1984). In captivity, despite the fact that there are many published records of maternal infanticide for almost all families of the Order Carnivora, the subject was mostly studied in commercially exploited species (e.g. Bakken 1994; Pyykonen *et al.* 1998; Korhonen & Harri 1988). Although potentially useful, records from zoological institutions have been under used in the research on breeding success, in particular referring to the impact of inadequate maternal behaviour in *ex situ* conservations programmes.

In this chapter, an overview of the adaptive value and occurrence of maternal infanticide in captivity is made. Statistical analysis of data from captive populations is used to tests the hypotheses that life history traits can predict the type of maternal infanticide displayed by a species under sub-optimal conditions, and that the characteristics of individual females, such as the rearing method or place of origin, can affect the female's decision to interrupt maternal investment.

Table 6.1: Classes of infanticide (Hrdy, 1979).

Classes	Types	Function/ Methods	Advantages
<i>Exploitation</i>	1) As a food resource 2) As a buffer protection 3) As a "mother toy"	1) Cannibalism 2) Using as a distraction 3) Excessive or inadequate alloparenting	1) Food; protein 2) Escape; defence 3) Possibly training
<i>Resource competition</i>	1) Directly related to rearing 2) Indirectly related to rearing 3) Milk "theft" 4) Burrow "theft" 5) Xenophobia 6) Brood reduction	1) Nest sites 2) Parental food supply 3) Killing new-borns from other females 4) Killing new-borns from other females 5) Killing infants from an unrelated or alien female 6) Killing own young passively (exposure, starvation)	1) & 2) Increase in the average the access to resources to the killer & descendants 3) The female will nurse the killer's young 4) The killer will take the mother's burrow 5) Increases the opportunities of killer's lineage 6) Allocation of parental efforts to one young that is more prone to survive.
<i>Parental manipulation</i>	Destruction of offspring: 1) Imperfect or debilitated 2) Non-defective	1) Premature; defectives 2) a) Under poor conditions b) In the presence of danger c) When there is a previous offspring	1) The cost is too high for a doubtful outcome. 2) a) & b) The risks are so high that the effort is not justified. c) Allocation of resources after a high investment in a stronger litter.
<i>Sexual selection</i>	Performed often by males, some forms by females.	1) Killing other male's offspring 2) Killing other female's offspring 3) Reabsorbing fetuses in the presence of strange males	1) Eliminates the rival's offspring and reduces the interval for another (the killer's) 2) Eliminates the rival's offspring and increases the chance of male's services 3) Eliminates an offspring that would be eventually killed ('Bruce effect').
<i>Social pathology</i>	Considered maladaptive and typical of human-disturbed locations (zoos and urban areas). Abnormal.	1) Disturbed by noise, light, and/or smells, captive females devour their offspring right after birth. 2) In poor conditions or in the lack of resources, an increase in aggressiveness lead to cannibalism, and infants of all ages are more vulnerable.	1) It could be a reflex of the selected mechanisms of parental manipulation discussed above, which are still at work in captivity. 2) Self-preservation mechanisms that could save one's life in extreme situations.

6.2. *The adaptive value of maternal infanticide.*

According to Hrdy (1979), maternal infanticide, passive or active, can be explained either as a tool for maternal manipulation (including the manipulation of sex ratio) or as a social pathology triggered by conditions of captivity. Wolff (1997) suggested that there are aspects that predict which species will commit infanticide: territorial females, altricial, non-mobile young and risk of predation are common to all that use this strategy, which is the case of many rodents and most terrestrial carnivores.

Mothers can cull young as a form of manipulation of litter size or sex ratio. In rodents, females that have very large litters may kill and commonly eat some young, thus reducing the number of young to the average of the species and nourishing the survivors properly (Labov *et al.* 1985). In many species, male pups are usually more demanding and at higher risk of being culled in harsher conditions (Labov *et al.* 1985). Maternal manipulation of the sex ratio can occur through differential abortion of the two sexes or a tendency to neglect, or respond less, to the begging and demands of the most costly sex (Eshel & Sansone 1994).

Infanticide occurs in species whose populations can be regulated by intrinsic, or behavioural, mechanisms. Territoriality, gender segregation, dispersal and reproductive suppression are also intrinsic regulation mechanisms that can reduce population growth before resource limitation (Wolff 1997).

Experiments with silver foxes showed that primiparous vixens had a higher percentage of infanticide when compared to multiparous ones, and those infanticidal females, when kept in the next year on visual and spatial isolation from the others, performed adequate maternal behaviour (Bakken 1992; Bakken 1993). This evidence links inadequate maternal behaviour, in this species, to social competition, rather than to any other mechanism (Braastad & Bakken 1993).

Whatever is the adaptive value of maternal infanticide, it is a very extreme strategy used sparingly in the wild, and its occurrence in captivity is generally viewed as social pathology of the individual. However, the frequency in which it is performed by captive carnivores and the fact that infanticidal females may, when housed in better conditions, perform adequate maternal behaviour, may indicate that females perceive the conditions in captivity as sub-optimal.

6.3. Maternal infanticide in captivity.

Many captive carnivore females present behavioural problems in the care of young. Published reports are common and many institutions remove the young of certain species for hand-rearing right after birth. Although hand-rearing has been reported to in several species (see Appendix 4), there are official guidelines in which species shall be preferentially removed to be raised by hand. The decision of removing young from the care of the female is largely at the discretion of the institution's staff and is usually based in personal experiences and anecdotal reports of infanticide, with little empirical support (Wharton & Mainka 1997).

Poor maternal care can seriously threaten the success of a captive breeding programme. Certain species do not respond well to hand-rearing, either through intolerance to milk replacements or through the lack of a proper behavioural development, especially in species with a sensitive socialisation phase (cf. Chapter 1). There is abundant evidence of maternal infanticide in zoo-housed carnivores, with examples in several families of the order (Chapter 5).

Although there is very little information to compare the occurrence of maternal infanticide in the wild and in captivity, the frequency with which it occurs in captive conditions is remarkable. For example, in captive cheetahs (*Acinonyx jubatus*), cub abandonment is said to be very common, but it was

only observed in the wild in very rare occasions, when prey are virtually non-existent (Laurenson 1993).

Zoological institutions can be an excellent source of information in the research of the causes of maternal infanticide. Data such as proportion of deaths caused by the dam, age of killer and offspring and husbandry routines are essential for the understanding of the phenomenon and are scarce in the wild (Braastad & Bakken 1993).

6.3.1. Husbandry techniques and maternal infanticide.

To unravel the factors behind the high frequency of maternal infanticide in zoos, the conditions in which the animals are kept must be looked at. Noise level, the proximity and quantity of visitors and the design of the enclosure are some of the aspects of captivity that seemed most likely to lead to episodes of infanticide.

The effect of husbandry on captive carnivore females was already evident and had led to the practical recommendation, among fur farmers, of removing infanticidal females from the stock, because they would continue to kill offspring while kept in the same conditions (Bakken 1994). Experiments showed, however, that these females can raise offspring when put in visual isolation from other females (Bakken 1994; Ilukha, Harri & Rekilä, 1997).

Noise levels were also known to be a source of disturbance to animals in exhibition. For example, bears seem to prefer smaller mothering dens than those provided by zoos. Small round dens are insulated from outer noises and allow the mother to hear the cubs (Van Keullen-Kromhout 1978). Female red pandas seem to be very disturbed by human presence and noise levels, and keep moving the cubs from one place to another, but the problem can be solved by roping the area and providing multiple nest boxes (Roberts 1975). Felids are said to require seclude and quiet to have cubs, and mothers could be

prone to neglect or eat cubs when disturbed or stressed (Laurenson, Wielebnowski & Caro 1995).

High noise levels seem to disturb polar bears more than other ursids, maybe because of their natural silent environment. On all occasions polar bears bred successfully in captivity, noise was carefully avoided. Although regular noise levels do not seem to disturb brown bears, one female killed her cubs after an incident where people invaded the zoo and used iron sticks to bang the bars of her cage (Van Keulen-Kromhout 1976).

Breeding or cubbing dens are an important aspect to be design in an enclosure. In nature, some species move their young to post-natal dens, such as the raccoon *Procyon lotor* (Judson, Clark & Andrews 1994), or abandon their previous cubbing dens when threatened, even when this movement may affect the female's breeding success (Swenson *et al.* 1997). One indication of the importance of multiple cubbing dens is perhaps the act, performed by many captive carnivores, of moving the young around the enclosure. Unsuccessful mothers usually perform cub-carrying (Foreman 1997; Callahan & Dulaney 1997), an act once mistaken as the female's desire of displaying the offspring to the visitors (Leslie 1971). Female bears sometimes do not accept a breeding den in a zoo, and behave restlessly after the cubs are born, carrying them around and eventually neglecting or eating them (Van Keulen-Kromhout 1976). In one event, a female spotted hyaena *Crocuta crocuta* started carrying her cub when one of the five lions that were moved through the service corridor of the enclosures pounded on the door of the cubbing den (Kinsey & Kreider 1990). Excessive cub-carrying is regarded as a warning signal of potential inadequate maternal behaviour, and for some species, such as snow leopards *Uncia uncia*, it is recommended to remove the cubs immediately after the display of this behaviour by the female (Wharton & Mainka 1997).

Providing the female with safe dens and the adequate requirements for the species can be successful. A good example occurred at Buffalo Zoo. A study about the cause of poor mothering of a female spectacled bear *Tremarctos*

ornatus tested several depths of bedding for the cubbing den. There is no information on the maternal behaviour of this species in the wild, but this observation in captivity showed that this female nursed her cubs not sitting or laying alongside, like other ursids, but laying on top of them. Previous litters were crushed to death, but with the bedding at 1 m of depth, the female prepared an oval nest with a deep middle depression where the cub was resting. The female then lay down on top of the nest to feed the young (Aquilina 1981).

Disturbances during parturition and early rearing are thought to elicit neglect of the litter or infanticide in some females. A strict hands-off policy during these times may result in a higher survival rate, especially with younger, less experienced females, which should give birth in familiar surroundings; the male rather than the female should be moved to another location during parturition and early rearing (Foreman 1997). For example, female giant otters *Pteronura brasiliensis* react to disturbance by lying flat on the floor, covering their teats and preventing the cubs from feeding; frequent repetition of this behaviour could be critical (Hagenbeck & Wunneman 1992). At Carl Hagenbecks Tierpark, in Germany, a female managed to rear two cubs after all sources of disturbance, including staff, were removed, and several nest boxes were provided for the female, and the institution recommends males to be separated from the females when cubs are present (Hagenbeck & Wunneman 1992).

There is a lack of information on many important aspects of the breeding biology of the species in the wild. This may have led to repeated mistakes in the management of more demanding species. For example, since the 1920s the maned wolf *Chrysocyon brachyurus* had been considered completely solitary, and there are not many observations of breeding in the wild (Rosenthal & Dunn 1995). Although paternal care is common among the Canidae, the role of male maned wolves in the caring of the young is unknown. Because of high juvenile mortality in mother-reared pups, the species survival plan used to

recommend that the offspring should be removed for hand-rearing (Rosenthal & Dunn 1995).

A short experiment, however, suggested that males spend as much time as the females in the company of the young, sharing the duty of “guarding” the pups; pup survival up to 6 months increased if the sire was introduced right after birth and kept with the dam (Bestelmeyer 1999). Because of similar experiences in other institutions, the Species Survival Plan for the maned wolf now recommends that zoos either leave the male with the females and pups from the time of the birth or introduce the male shortly after birth (Bestelmeyer 1999).

Information from captive populations of carnivores may provide clues to which species are more prone to commit maternal infanticide, and from this result it can be possible to work towards husbandry protocols aiming to reduce female disturbance and providing optimal conditions for young to thrive.

6.3.2. The impact of maternal infanticide in conservation programmes.

Captive populations of rare carnivore species are often the subjects of conservation programmes, and institutions taking part in conservation efforts frequently keep records of births and causes of mortality (Chapter 2).

One of the few species to have some of these records analysed was the cheetah *Acinonyx jubatus*. Analysis of data from 3 breeding centres in North America showed that 48.5% of 33 cub deaths were caused by abnormal maternal behaviour (Laurenson, Wielebnowski & Caro 1995). Another study, using records from 1829 to 1994, showed that 44% of deaths in captive cheetah before 6 months of age (n=364) were caused by aggression, trauma, maternal neglect, exposure or being eaten by the dam (Marker-Kraus 1997).

Less extensive research in other species also shows the relatively high proportion of deaths caused by mothers. Records from 8 institutions that keep rusty-spotted cats, collected between 1976 and 1994, showed that in 11 out of 17 registered deaths the young had been killed and eaten by the mother (Dmoch 1997). At Cincinnati Zoo, mortality in Pampas cat *Oncifelis colocolo* kittens is primarily a result of injuries inflicted by the dam (Callahan & Dulaney 1997).

Although subject to intensive breeding programmes, the giant panda *Ailuropoda melanoleuca* poses a challenge for institutions. From the seven young born in 1989, only one was hand-reared to sexual maturity, and inadequate maternal behaviour caused all but one death (Bingxing 1990).

Some records are not explicit about the cause of death in the young. Analysing data from leopards *Panthera pardus*, Shoemaker (1982) found a high number of deaths caused by “weakness” or “by devouring”, which could mean that females were neglecting or eating their cubs.

Usually, when faced with a frequently infanticidal species, institutions remove young for hand-rearing right after birth, or keep mother and young under observation to remove the young if its weight drops, which is the case with the cheetahs in Wassenaar Wildlife Breeding Centre (Beekman *et al.* 1997). The effects of hand-rearing in the behaviour of the adult individual are controversial, and must be specifically researched. An experiment with domestic cats suggested that hand-reared females have a much lower breeding success than mother-reared ones (Mellen 1992).

The high proportion of maternal infanticide found in Chapter 5 indicates that this behaviour have an impact in potential captive breeding programmes, reducing the output of new individuals and perhaps compromising their breeding capability through the possible long term effects of hand-rearing.

6.4. Hypotheses to be tested.

Statistical analysis of data from captive populations is used to test the hypotheses that life history traits can predict the type of maternal infanticide displayed by a species. As maternal investment is usually curtailed in times of scarce resources, when the welfare of the breeding female is low (Clutton-Brock 1991; Hrdy 1979; Hayssen 1984; Labov *et al.* 1985; Wolff 1997), it is possible that captive female carnivores are behaving as they would be under sub-optimal conditions. Active infanticide is more likely to occur during lactation, when energy output from the mother is higher (Clutton-Brock 1991; Oftedal & Gittleman 1989), and is probably more frequent in species with more demanding young (Oftedal & Gittleman 1989; Packer & Pusey 1984). Passive infanticide is more likely to occur when maternal investment is smaller (Clutton-Brock 1991), so species with lower mother-infant energy transfer are supposedly more prone to abandon their cubs. The welfare of carnivores is highly affected by captivity (Clubb & Mason 2003), thus it is expected that the characteristics of individual females, such as the rearing method or place of origin, can affect the female's decision to interrupt maternal investment. Also, husbandry protocols and housing conditions can affect directly the welfare of nursing females (Carlstead & Shepherdson 1994; Bakken *et al.* 1999), but can be adjusted to minimise the impact of inadequate maternal behaviour in the breeding success of captive carnivores.

6.5. *Methods.*

The dataset used for this analysis was the bibliographical dataset collected from published papers (described in section 2.2.5), which served for the analysis of maternal infanticide in 29 species. Data collected directly from zoological institutions (described in section 2.2.6) did not present detailed data on causes of infant mortality and were not suitable for the analysis.

Relative frequencies of the type of infanticide (killed by dam, eaten by dam or neglect) were calculated for each species, using the median of these proportions for each female, and used as dependent variables in regressions. Also, the type of infanticide (if active or passive) was used as a dependent variable to check the effect of categorical individual characteristics of the females through a binary logistic regression. A summary of the species used in the analysis and the sample sizes, together with the relative frequencies of infanticide, is displayed in Table 6.2.

Average litter sizes and the proportion of infant mortality were also calculated for each female. The origin of the females, as reported in the published records, was included in one analysis. There was some information on housing conditions, such as cage area and number of dens, but they were not used in the analysis of maternal infanticide because sample sizes were not large enough, after adjusting for species and females. Multiple analyses on the effects of life history traits were performed using the variables chosen through stepwise regression from the dataset compiled by Gittleman (1986a, 1986b, 1989). All proportional values were transformed by the arcsine of the square-root and other continuous variables were log-transformed.

Table 6.2: Species used in the analysis of the types of maternal infanticide in captive carnivores, with number of litters, dams, zoos holding the species, proportion of active infanticide (KBD), proportion of infanticide followed by cannibalism (EBD) and proportion of passive infanticide (NEG). The data was collected from 141 published papers (Appendix 4).

Family	Species	Litters	Dams	Zoos	KBD	EBD	NEG
Canidae	<i>Canis lupus</i>	9	2	2	0,9286	0,3571	0,5714
Canidae	<i>Cerdocyon thous</i>	6	4	2	0,7778	0	0,4445
Canidae	<i>Chrysocyon brachyurus</i>	20	13	8	0,7368	0,7368	0
Canidae	<i>Vulpes zerda</i>	10	7	3	0	0	0
Canidae	<i>Lycaon pictus</i>	4	2	2	1	0,3529	0,3529
Felidae	<i>Acinonyx jubatus</i>	22	15	12	1	1	0
Felidae	<i>Caracal caracal</i>	5	4	2	0,4167	0,4167	0
Felidae	<i>Felis margarita</i>	10	2	2	0,375	0,25	0
Felidae	<i>Felis nigripes</i>	8	3	2	0,4286	0,2857	0,1428
Felidae	<i>Felis silvestris</i>	29	7	3	0,67	0,67	0
Felidae	<i>Leopardus tigrinus</i>	7	3	2	0	0	0
Felidae	<i>Lynx lynx</i>	10	7	3	0,8571	0,3571	0,2142
Felidae	<i>Neofelis nebulosa</i>	15	5	4	0,8823	0,7647	0,1176
Felidae	<i>Oncifelis geoffroyi</i>	15	5	3	0,2857	0,2857	0
Felidae	<i>Prionailurus bengalensis</i>	8	2	2	0,2143	0,2143	0
Felidae	<i>Uncia uncia</i>	11	5	4	0,8	0,8	0
Felidae	<i>Panthera tigris</i>	9	7	5	0,9048	0,0476	0
Herpestidae	<i>Helogale paroula</i>	14	3	2	0,3333	0	0
Hyaenidae	<i>Crocuta crocuta</i>	4	3	3	0,1538	0	0
Mustelidae	<i>Amblonyx cinereus</i>	11	3	3	0,62	0,5	0
Mustelidae	<i>Enhydra lutris</i>	9	5	4	0,5	0	0,5
Mustelidae	<i>Pteronura brasiliensis</i>	12	2	1	1	0	0
Mustelidae	<i>Meles meles</i>	2	2	1	0,8333	0,8333	0
Procyonidae	<i>Ailurus fulgens</i>	9	5	3	0,9375	0,125	0
Ursidae	<i>Ailuropoda melanoleuca</i>	11	7	5	0,4167	0	0
Ursidae	<i>Helarctos malayanus</i>	12	5	3	0,9286	0,3571	0,5714
Ursidae	<i>Tremarctos ornatus</i>	11	4	4	0,7778	0	0,4445
Ursidae	<i>Ursus maritimus</i>	11	4	4	0,7368	0,7368	0
Viverridae	<i>Arctictis binturong</i>	13	5	5	0	0	0

6.6. Results.

6.6.1. Biological factors.

Species that prey on larger animals tended to kill but not consume the young (One-way ANOVA: $F_{2,14} = 7.54$, $p = 0.04$), but there was no significant evidence that species with smaller prey do consume the young (One-way ANOVA: $F_{2,7} = 3.8$, $p = 0.1$, power = 0.47). Species with slower growing litters showed a higher incidence of active infanticide without cannibalism (Regression: $F_{1,9} = 1.18$, $p = 0.05$, R^2 adj. = 28%), as it can be seen in Figure 6.1.

In this dataset, there was an effect of weaning age (WA) and age in which young open their eyes (OE) on the incidence of deaths caused by the dam in each species (section 5.5.2), but these variables did not influence the occurrence of active infanticide, followed or not by cannibalism (GLM on transformed data: WA: $F_{1,12} = 1.47$, $p = 0.4$; OE: $F_{1,14} = 1.18$, $p = 0.6$, R^2 adj. = 40%). The proportion of passive infanticide reported for each species was strongly influenced by interbirth interval (Regression: $F_{1,12} = 11.68$, $p = 0.005$, R^2 adj. = 45%, Figure 6.2) but there was no significant effect of weaning age (Regression: $F_{1,13} = 1.38$, $p > 0.2$, R^2 adj. = 10%).

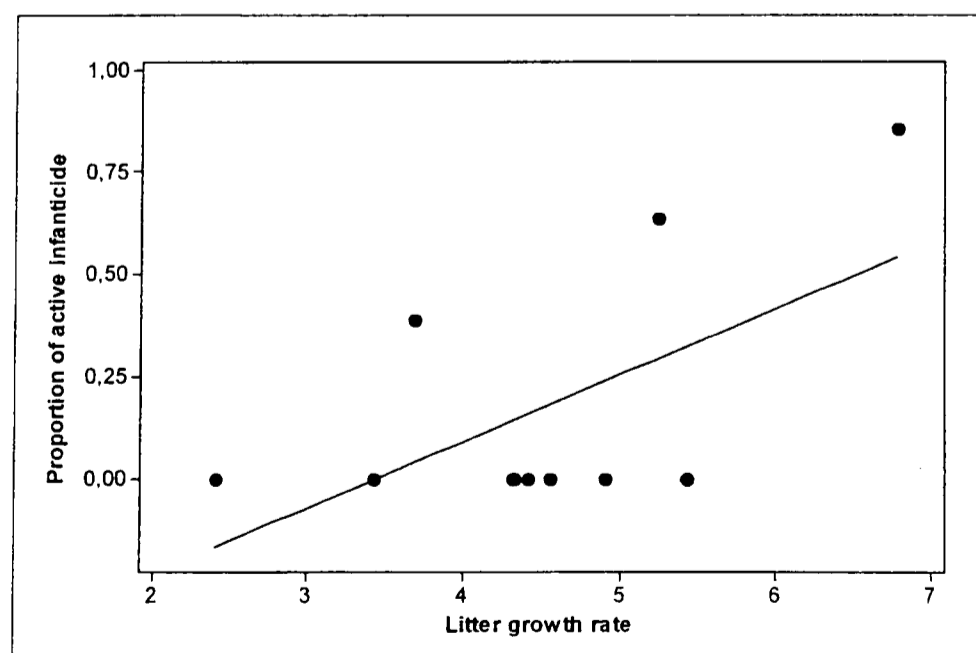


Figure 6.1: Regression plot of the effect of specific litter growth rate on the proportion of infant deaths caused by maternal active infanticide not followed by cannibalism, in 10 species of carnivores. The data plotted was transformed.

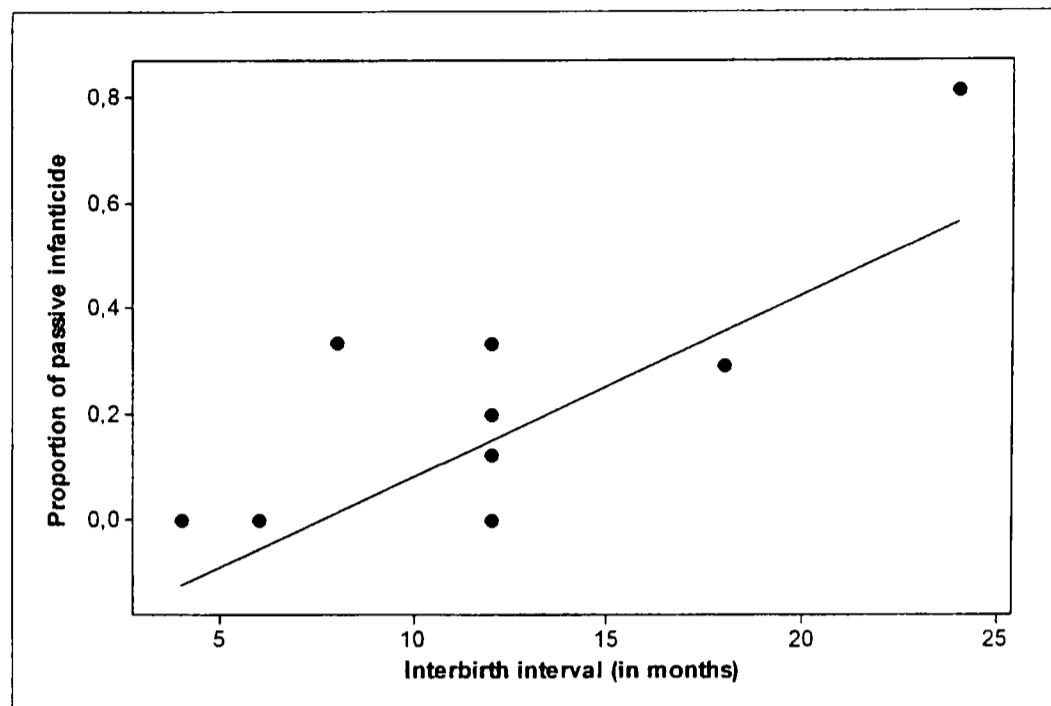


Figure 6.2: Regression plot of the effect of interbirth interval in the proportion of infant deaths caused by passive infanticide in 9 species of captive carnivores. The data plotted is not transformed.

6.6.2. Individual differences and ecological factors.

Overall, wild-caught females presented slightly higher levels of maternal infanticide, passive or active, than captive-born ones independent of the species (t-test: $t = 1.84$, $p = 0.06$, $df = 63$, Figure 6.3).

Throughout the order Carnivora, there was a slight effect from the origin of the female (if wild-caught or captive born) in the type of infanticide, although this is undetectable if other factors are added to the model: wild-caught females performed more passive infanticide (PI) and captive-born performed more active infanticide (AI), as it can be seen in Table 6.3. In the complete model, only the activity pattern of the species had a significant effect in the type of infanticide performed, with arrhythmic species presenting higher levels of AI and diurnal species neglecting young more often.

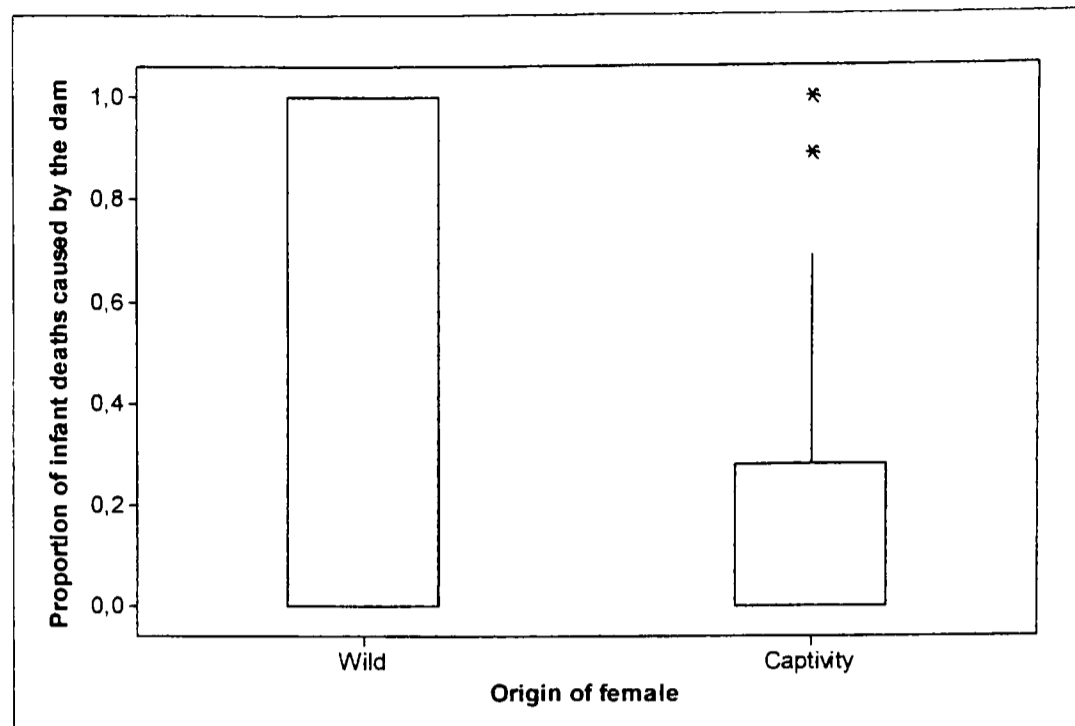


Figure 6.3: Proportion of mortality caused by inadequate maternal behaviour in wild-caught (N = 60) and captive-born (N = 27) females of 29 species of carnivores in captivity.

Overall, there was no statistically significant effect of the presence of the male in the incidence of maternal infanticide in species with exclusive maternal care (i.e. when the male does not care for the young), but a larger sample would be needed to increase test power (One-way ANOVA: $F_{1, 250} = 0.02$, $p = 0.89$, power = 0.27).

Table 6.3: Logistic regression results with categorical variables of female characteristics and specific ecological requirements of captive carnivores; the dependent variable is the type of maternal infanticide (passive or active).

Variable	Model 1		Model 2		Model 3	
	B	Wald	B	Wald	B	Wald
Constant	-0.56	2.39	12.74	0.11	12.56	0.11
ORIGIN	4.19	0.04*	-8.12	0.04	-8.15	0.05
REARING	-	-	-1.55	1.11	-1.49	1.03
ACTIVITY	-	-	-1.76	4.55*	-1.79	4.46*
DIET	-	-	-	-	0.11	0.082
Model Chi-Square [df]	6.337 [2]*		12.564 [3]**		12.646 [4]**	
Block Chi-Square [df]	-		6.227 [1]		0.082 [1] *	
% corrected prediction	63.6		78.8		78.8	
Nagelkerk's R ²	0.24		0.43		0.44	

* $p < 0.05$; ** $p < 0.01$

6.7. Discussion.

6.7.1. Husbandry requirements and individual differences affecting maternal infanticide in captive carnivores.

In this dataset, wild-caught females performed infanticide more often than captive-bred ones, though the latter were more prone to actively kill the young, while the former abandoned the cubs more often. The effect of the environment on encaged animals has been discussed in other studies, and some researchers affirm that stress from captivity can increase the occurrence of abnormal behaviour (Carlstead & Shepherdson 1994; Bakken *et al.* 1999). It is possible that wild-caught individuals are more sensitive to the conditions in captivity and therefore perform more infanticide. Carlstead and Shepherdson (1994) suggest some techniques that can be useful to reduce stress in captivity, and were useful to raise success breeding in captivity, focused basically on three aspects: special perception, such as providing platforms and windows to expand the view as well as places to “hide” from the public; feeding behaviour, to avoid pre-feeding stereotypical behaviour and to increase foraging time; and welfare, making sure that individuals have ways to control microclimate and are always interested in the environment.

A controversial suggestion, though, is that the animals should be trained for husbandry, to improve their relationship with human keepers. In a later work, Carlstead (1996) supports the idea that unconscious selection to tameness may lead to dangerous changes in natural behaviour; for instance, in malamute pups *Canis lupus lycaon*, unrestrained aggression and the absence of some threat displays seem to be result of a selection for neoteny. Inadequate environment or hand-rearing could lead to abnormal behaviour in species with sensitive periods (i.e., imprinting, socialisation) (Carlstead 1996). Unfortunately, this subject is not yet well researched.

It has been suggested that human interference, especially those related to research efforts in the wild, can increase the occurrence of cub abandonment in black bears *Ursus americanus* (Goodrich & Berger 1994), brown bears *Ursus arctos* (Swenson *et al.* 1997), and grey wolves *Canis lupus* (Ballard, Whitman & Gardner 1987). This could be reflected in the tendency of wild-caught females abandoning their litters in captivity.

Individual differences in the response to stress can predict which females will be more prone to commit infanticide. A study was performed to expose farmed silver foxes to stimuli that trigger stress-induced hyperthermia (SIH), phenomenon regulated by the HP axis, while measuring the time and intensity of their responses. Infanticidal females were more sensitive to stressors and responded quicker to the stimuli, while non-infanticidal ones had weaker responses. It was suggested that infanticidal behaviour could be predicted, and eliminated from the stock, by selecting breeders with lower levels of SIH (Bakken *et al.* 1999).

In the wild, threatened females will kill or leave their offspring to die after perceiving that there is no safe place to hide the young, or when there is only one surviving cub on a litter, because the risk of raising a single cub is too high, and it is more profitable try again (Packer & Pusey 1984).

There is a lack in research on the factors that can make species more prone to one or other type of infanticide. In this dataset, it was found that species with arrhythmic pattern of activity, i.e. that can be equally active during the day and the night, such as the grey wolf, performed active infanticide more frequently, and diurnal ones were more prone to abandon the litter. Species with diurnal activity are frequently at higher risk of predation than nocturnal or arrhythmic ones, as happens with baboons *Papio sp.* (Cowlshaw 1994) and striped skunks *Mephitis mephitis* (Lariviere & Messier 1997). Some species, such as the red-bellied lemur *Lemur rubriventer*, may have adjusted their activity pattern from exclusive diurnal to arrhythmic, or cathemeral, as a strategy to avoid predation by diurnal raptors (Overdorff

1988). If diurnal species are at higher risk of predation, leaving the litter alive may serve as a “buffer protection” (Hrdy 1979), as the young can be found by predators that could otherwise prey on the female; also, killing the young could attract predators through auditory or olfactory cues.

Unfortunately, it was not possible to calculate the effect of female age in maternal infanticide, once the data was available for very few females. Also, as discussed in Chapter 2, the dataset did not have reliable data to analyse sex ratio. Further research is recommendable to elucidate these points.

6.7.2. Biological factors affecting maternal infanticide in captive carnivores.

In Chapter 5, it was found that species that prey on small or very small animals perform more infanticide, active or passive, than those with larger prey (Section 5.5.2). Analysing infanticidal events separately, it was found that species with larger prey tend to actively kill the young, but not consume them. Prey size, in carnivores, has several ecological and morphological correlates, such as home range size, limb length and limb dexterity (Gittleman 1986a, 1986b; Harris & Steudel, 1997; Iwaniuk, Pellis & Whishaw 1999). Killing techniques, adapted to the type and size of prey, are the result of craniodental adaptations and differ greatly between taxa. Solitary predators, such as felids, usually kill prey with one single deep bite, while canids and hyaenids, social hunters, tend to kill with multiple shallow bites (Biknevivius & Van Valkenburgh 1996). In this dataset, the species that did perform more active infanticide without cannibalism were both group-hunting grey wolves, wild dogs and spotted hyaenas (prey “nibblers”), and the solitary tigers and cheetahs (prey “crushers”). This indicates the possible influence of feeding preference in this result: infants would be too small to consume out of starvation times. Further research is needed to look into the species that do

consume the young and the possible connection with prey size and feeding preference.

In this research, active infanticide was also more frequent in species with higher litter growth rates (LGR). This is predictable once LGR can be a measure of maternal energy output, through energy transfer in lactation (Oftedal & Gittleman 1989), and females are more likely to withdraw maternal investment, partially or totally, to a litter through infanticide when they perceive sub-optimal conditions (Clutton-Brock 1001; Hayssen 1984; Hrdy 1979; Packer & Pusey 1984; Wolff 1997).

In captivity, the present research showed that species that open the eyes and wean at later ages are more prone to die from all causes (Chapter 4), while species that open the eyes and wean earlier, and therefore are highly demanding on the first weeks after birth, are more susceptible to be killed as a result of inadequate maternal behaviour (Chapter 5). These factors, however, did not predict the occurrence of the type of infanticide, i.e. if active or passive, suggesting that the decision of abandon or kill the young depends on other factors. Altriciality is thought to be a factor predicting the occurrence of infanticide in carnivores (Packer & Pusey 1984); together with territorial females, it is a condition for a species to perform infanticide as a intrinsic population regulator, but all Carnivora are altricial if compared, for example, with Artiodactyla species (Wolff 1997). Altricial young represent, at birth, a low maternal investment if compared to precocial ones; fast-developing species, however, represent a very high maternal investment (Zavelloff & Boyce 1986). It is possible that, among Carnivora species that perform infanticide, fast development, rather than altriciality, predicts frequency of infanticide, at least in captive populations.

Frequency of reproduction, reflected in the length of the interval between births in the species, had a strong effect in the occurrence of passive infanticide. Species that presented high frequency of neglect were grey wolves, Malayan sun-bears, spectacled bears, crab-eating foxes and wild dogs. Grey

wolves and wild dogs are communal carers (Gittleman 1986a), dividing the energetic expenditure of raising the young between many. It is known that young desertion is more likely to be performed when the investment by the deserter is low (Clutton-Brock 1991). Brood desertion is common in ursids, and females are known to abandon a cub if it is the only survivor from a litter, once it is more profitable to raise a whole litter than to nurse the survivor (Clutton-Brock 1991). Other factors, however, may be involved in the interruption of maternal care. For example, in vervet monkeys (*Cercopithecus aethiops sabaesus*), infant mortality due to poor maternal care or abandonment was higher for females in poor body condition, while females in prime condition rejected their own young, that survived through alloparental care, to shorten the interval between conceptions (Fairbanks & MacGuire 1995). Unfortunately, there was not enough data in this study that allowed to check the influence of individual characteristics of the females, such as origin and housing conditions, in the occurrence of maternal infanticide.

6.8. Conclusion.

Maternal infanticide in captive carnivores is related to the level of maternal investment demanded by each species, with fast-developing young being more prone to be killed than slow-developing ones. The type of infanticide performed is also predicted by species characteristics: species that hunt large prey killed the young but did not eat them, while species with long intervals between conceptions tended to withdraw maternal care instead of actively killing the young. Each form of maternal manipulation of the litters will reflect the species' adaptation to balance maternal investment and resource availability within their environmental and social settings.

As the decision of interrupting maternal investment depends on the perception of the mother on resource availability, the welfare of the individual is crucial to successful breeding. The high frequency of maternal infanticide in captive populations of carnivores suggests that females perceive captivity conditions as sub-optimal, which can seriously damage the output of adult individuals by *ex situ* breeding programmes. This effect seems to be stronger in wild-caught females, unused to human manipulation, than in tamer captive-born ones. Further research is needed, however, to fully understand the influence of husbandry techniques and housing conditions in species prone to commit maternal infanticide.

Chapter 7

Increasing juvenile survival in captive carnivores:

Practical considerations

Abstract

There are few studies on the impact of husbandry practices in the breeding success of captive carnivores. Questionnaires were sent to institutions asking for details of practices and housing conditions of nine species of carnivores, and 16 institutions provided data on 148 litters from 63 females. Overall the order Carnivora, institutions that perform research presented a higher proportion infant mortality than institutions that do not perform any kind of research activity. The distance travelled by the female was positively correlated with the proportion of infant mortality, while the number of dens and the available area for each female (adjusted for each species) were negatively correlated with the proportion of infant mortality. Transferring the females from one institution to other, or from the wild to any institution, can affect infant mortality even when not considering distances; the latitude of the zoo or the number of keepers in contact with the animals do not seem to affect infant survival. In this dataset, the two hand-reared females had a lower proportion of infant mortality than mother-reared ones, although a larger sample would be needed. Also, institutions that have scientific staff and run research programmes had higher infant mortality overall, independently of their latitude or management system, which may indicate an effect of human interference, although further research is needed. It is recommended that breeding loans between institutions do not include females, and that direct manipulation of animals in studies are avoided until further understanding of the impact of these practices on breeding success.

7.1. Introduction.

Captive carnivores pose a challenge for conservationists and institutions alike, presenting many problems that range from diseases to poor welfare and unsuccessful breeding (Chapter 1). Available databases of captive populations are rich sources of information that can help determine which factors can affect

breeding success and the real potential of these populations in conservation programmes (Chapter 2). Some species, such as tigers *Panthera tigris*, seem to preserve in captivity the same reproductive parameters, such as litter size, lack of seasonality and breeding age, seen in wild animals, making captive individuals extremely useful in the research of reproductive biology, that can be applied in evolutionary and physiological studies of the order Carnivora (Chapter 3).

Specific reproductive characteristics can make some species more prone to lose young in captivity than others, and these factors must be taken into consideration when developing *ex situ* conservation programmes; for example, the effect of photoperiod in Arctic species should be taken into consideration when housing these species closer to the Tropics (Chapter 4). Infant mortality in captivity seems to be primarily caused by inadequate maternal behaviour, which is more frequent in fast developing species and wild-caught females; proper husbandry and well-designed enclosures can reduce the other causes of death (Chapter 5). Maternal infanticide, either passive or active, is also affected by biological and ecological characteristics of the species, but may be the result of poor maternal welfare as it is more frequent in wild-caught females (Chapter 6). It is important thus to research the effect of husbandry protocols and housing conditions in the breeding success of captive carnivores. Record research in the subject poses some problems: for example, data collection has to be restricted, once the many factors present in captive environments cannot be all accounted for, and the detail and reliability of the data varies between institutions.

In this chapter, an overview of studies relating husbandry and housing conditions to the welfare and reproductive success of captive carnivores is done. Records from institutions were used in the attempt of determining the effect of some of these conditions in the proportion of infant mortality in captivity, taking into consideration the results found in previous chapters.

7.2. Welfare and breeding success in captive carnivores.

Research has suggested that breeding success is strongly affected by the female's welfare, and non-invasive techniques to assess the adaptation of individuals to husbandry protocols can be very useful in reducing the impact of stress in reproduction (Wielebnowski 1999). Environmental enrichment techniques have been tested and applied in several species, such as the giant panda *Ailuropoda melanoleuca* (Swaisgood *et al.* 2001), the bush dog *Speothos venaticus* (Ings, Waran & Young 1997) and the kinkajou *Potos flavus* (Blount & Taylor 2000) with the aim of enhancing well-being and reducing unwanted behaviours, such as stereotypical movements, with positive results. Poor welfare and unsuccessful breeding attempts were connected in a study with captive clouded leopards *Neofelis nebulosa* in North American zoos, where behavioural problems, such as stereotypies and excessive aggressiveness are regarded as indicators of chronic stress, which can be detected by non-invasive corticoid monitoring (Wielebnowski *et al.* 2002).

Research with captive animals can point towards solutions to improve the welfare of individuals, and consequently increase breeding success. For example, a study on captive sloth bears *Ursus ursinus* showed that variation on physical and social captive environments affects activity patterns in this species, including the occurrence of stereotypies, and further studies may be helpful to identify which factors can be controlled to improve reproductive success and welfare (Forthman & Bakeman 1992).

In the subject of captive breeding, new techniques focus on the identification of oestrus, once it is difficult to perceive in some species and successful breeding frequently depends on the introduction of the male to the female while she is liable to accept mating. One example is the study of behavioural and physiological cues of oestrus in the cheetah *Acinonyx jubatus*, which uses laboratory techniques, such as cytology, non-invasive hormonal

analysis, and ethological observations to determine the precise moment when females come into oestrus (Graham *et al.* 1995; Brown *et al.* 1996; Bircher & Noble 1997). Cheetahs are said to have a “silent” oestrus, which has led to unsuccessful introductions of males to females, but non-invasive endocrinological studies have yielded good results in correlating physiological values with behavioural displays that can indicate the actual reproductive status of the females (Wielebnowski & Brown 1998). Oestrus is also difficult to determine in giant pandas, and researchers had found that there are chemical cues in the female’s urine that trigger *flehmen* displays in males, the knowledge of which can be used to detect heat in females without the need of laboratorial analyses (Swaisgood, Lindburg and Zhang 2002).

The introduction of new males to females is crucial in captive breeding, and the welfare of the individuals has to be taken into consideration. Introduction protocols have been developed aiming the reduction of aggressive episodes that frequently lead to the injury of one of the animals, as it happens with clouded leopards *Neofelis nebulosa* (Law & Tatner 1998).

Research in zoological institutions has yielded valuable results in the diagnosis of pregnancy through non-invasive methods, which avoid direct manipulation of the animals. For example, it is difficult to determine early pregnancy in giant pandas without collecting blood samples of the females; a study, however, validated a method of diagnosing early pregnancy through urinary steroid concentrations and also showed that the species has delayed implantation, which can lead to mistaken calculations of gestation length (Chaudhuri *et al.* 1988).

It has been suggested that captive wild animals should be trained for husbandry, although it was pointed out that selecting for tameness might lead to the loss of important natural behaviours in certain species (Carlstead 1996). Although the subject was never researched, reports on the breeding success of tamed females suggest that closer interactions with known keepers reduce stress and promote adequate maternal behaviour in certain species, such as

cheetahs (Florio & Spinelli 1967; Florio & Spinelli 1968; Tong 1974), giant pandas (Bingxing 1990; Zhang *et al.* 2000) and clouded leopards (Fellner 1965). The effect of the origin of the female on the frequency of inadequate maternal behaviour in captive carnivores (Chapter 6) seems to support this view, although further research is necessary. Nevertheless, it is a safer policy for institutions to give preference to captive-born females, used to husbandry routines, as founders in *ex situ* programmes, and invest in the use of non-invasive monitoring techniques in breeding research, to minimise the stress in the individuals.

7.3. Hand-rearing, fostering and inadequate maternal behaviour.

Most institutions prefer to let the mothers to raise their cubs, and put up a policy of very little interference while there are young under care, but sometimes it is necessary to remove the infants for hand-rearing, usually after episodes of neglect or aggressive behaviour (Edwards & Hawes 1997). Removal of the young right after birth is recommended by the species survival plan for the maned wolf *Chrysocyon brachyurus* when the female seems agitated or is weakened (Rosenthal & Dunn 1995). Other reasons for the removal of the young include unsuitable enclosures with no dens; manipulation of group structure; neonatal illnesses; taming of individuals for educational purposes (making them accustomed to direct handling); and elimination of parasitic infections that would otherwise persist in the group (Read & Meier 1996).

Hand-rearing techniques have been tried and tested in many species and protocols have been established for a number of species, such as brown hyaenas *Parahyaena brunnea* (Volf 1996), otters *Lutra lutra* (Sikora 1996), Mexican margays *Leopardus wiedii glaucula* (Edwards & Hawes 1997), cheetahs *Acinonyx jubatus* (Bircher & Noble 1997) and ferrets *Mustela putorius* (Manning & Bell 1990), among others. In any case, hand-reared young are prone to

develop certain ailments, especially related to improper nutrition, as it was noticed in hand-reared polar bear *Ursus maritimus* cubs which developed rickets; nutritional supplements were introduced, but one of the cubs was left with a permanent femur deformity (Kenny, Irlbeck & Eller 1999).

Other option is to give the neonates to foster mothers, usually of the same species, or a closely related one. For example, Asian golden cats *Catopuma temmincki* kits were successful reared by a domestic cat, although the fostered young had half of the weight of mother-reared ones before weaning (Louwman & Oyen 1968). Sometimes fostering can lead to unwanted results: a neglect giant panda cub was given to a foster giant panda dam that had a 3 weeks-old cub; the dam killed the foster young some days after introduction, and in the attack stamped her own young to death (Bingxing 1990).

Removing the young from their mothers is a decision to be taken only after analysing each individual case in detail, and realising that the young would certainly perish if left with the dam (Read & Meier 1996). However, since sometimes the young have to be removed, it is necessary to develop safer protocols that increase the survival of hand-reared or fostered captive carnivores.

7.3.1. Hypotheses to test.

Infant mortality is, overall, predicted by the level of altriciality of the species, but can be affected by institutional latitude in Arctic species (Chapter 4). Inadequate maternal behaviour was responsible for a significant proportion of infant deaths and is more common in species with fast development of the young (Chapter 5), but as this phenomenon is based on the female's decision to continue or curtail maternal investment, it can be triggered if the female perceive captivity conditions as sub-optimal, as it seems to happen more often in wild-caught individuals (Chapter 6). The high variance in the proportion of infant mortality between females and

institutions suggests the influence of factors such as institutional latitude (Lincoln 1998), husbandry practices (such as number of keepers and if the animals are subject to research) (Bingxing 1990; Florio & Spinelli 1967; Florio & Spinelli 1968; Mellen 1991; Tong 1974; Wielebnowski *et al.* 2002; Zhang *et al.* 2000), available area and nesting places (Clubb & Mason 2003; Roberts 1975; Van Keullen-Kromhout 1976), and the origin and the number of translocations females were subjected to, which can lead to stress and reduce breeding success (Forthman & Bakeman 1992; Ings, Waran & Young 1997; Swaisgood *et al.* 2001; Wielebnowski 1999).

7.4. Methods.

For this chapter, the dataset used in the search of husbandry factors affecting breeding success of captive carnivores was collected through electronic questionnaires (Appendix 5) sent directly to zoological institutions in Europe, North America and Australia (as described in section 2.2.5.1). From this, the proportion of infant mortality was calculated for each female, and paired with each individual's characteristics (such as origin and rearing) and housing conditions. In the general analysis, which involved all 9 species, the individual available area was calculated dividing the size of the enclosure by the number of individuals kept, and this result was divided by the average body weight for each species, in a way to adjust the value for the size of the animals. There were data on the origin and rearing of only 35 females. Table 7.1 summarises the data collected from the zoological institutions (Appendix 6). Multiple regression analyses were done using the arcsine of the square root of the infant mortality for each female. Cage areas were log-transformed.

Table 7.1: Species, number of litters, number of collections keeping the species and number of females summarised from the data collected through questionnaires from zoological institutions.

Family	Species	Number of litters	Number of collections	Number of females
Mustelidae	<i>Amblonyx cinerea</i>	8	2	2
Procyonidae	<i>Ailurus fulgens</i>	11	3	7
Canidae	<i>Chrysocyon brachyurus</i>	15	4	9
Canidae	<i>Canis lupus</i>	13	5	5
Procyonidae	<i>Nasua nasua</i>	11	1	6
Felidae	<i>Panthera tigris</i>	22	6	9
Herpestidae	<i>Suricata suricatta</i>	62	7	19
Felidae	<i>Uncia uncia</i>	3	1	1
Ursidae	<i>Ursus maririmus</i>	3	2	2

For the binary logistic regression, the proportion of infant mortality was transformed in a dummy variable: values below the median for the sample were coded 1, and above, 2.

To calculate the distance travelled by the female, information on the individuals transfer between institutions or between the local of capture and the institutions was transformed in kilometres. These values were found with the aid of the software Earth Explorer (Motherearth Inc.). Otherwise, the number of transfers to which the females was subjected was used in a logistic regression model.

The number of dens was given by the institutions and represents the number of nesting places present in the enclosure. The number of keepers represents the number of persons responsible for *in loco* husbandry, such as feeding, capturing (for changing enclosures, for example) and cleaning the cages. It is a common practice of institutions to delegate some enclosure to the care of particular staff, for it is believed, anecdotally, that the animals suffer less stress, and are more manageable, if they are used to the keeper.

To research the impact of animal manipulation on the breeding performance of captive carnivores, the datasets used were collected from the *International Zoo Yearbooks* (see section 2.2.2; Appendices 1 and 2). Relative institutional performance indices, Z , were calculated (equation 2.3) using the proportion of infant mortality for each species and each zoo, and institutions

were rated according to the proportion of the species that were breeding above or below the median proportion of mortality for the species (*I*, Appendix 1). Institutions were rated as “performing research” if the *IZY* related them as having research departments, academically qualified staff (such as PhDs and MScs) or were involved in conservation programmes. For the general linear model, categorical variables were used for the latitude of the zoos (tropical or temperate) , since some species have photoperiodic control of reproduction, and the management regimen (private, governmental or zoological society/charity), because only 17% of private zoos develop research programmes, while 35% of non-private zoos employ highly qualified staff and perform some type of research.

7.5. Results.

Overall the order Carnivora, institutions that perform research presented a higher proportion infant mortality than institutions that do not perform any kind of research activity; there was no significant effect of the latitude of the institution or the management regimen (Table 7.2, Figure 7.1).

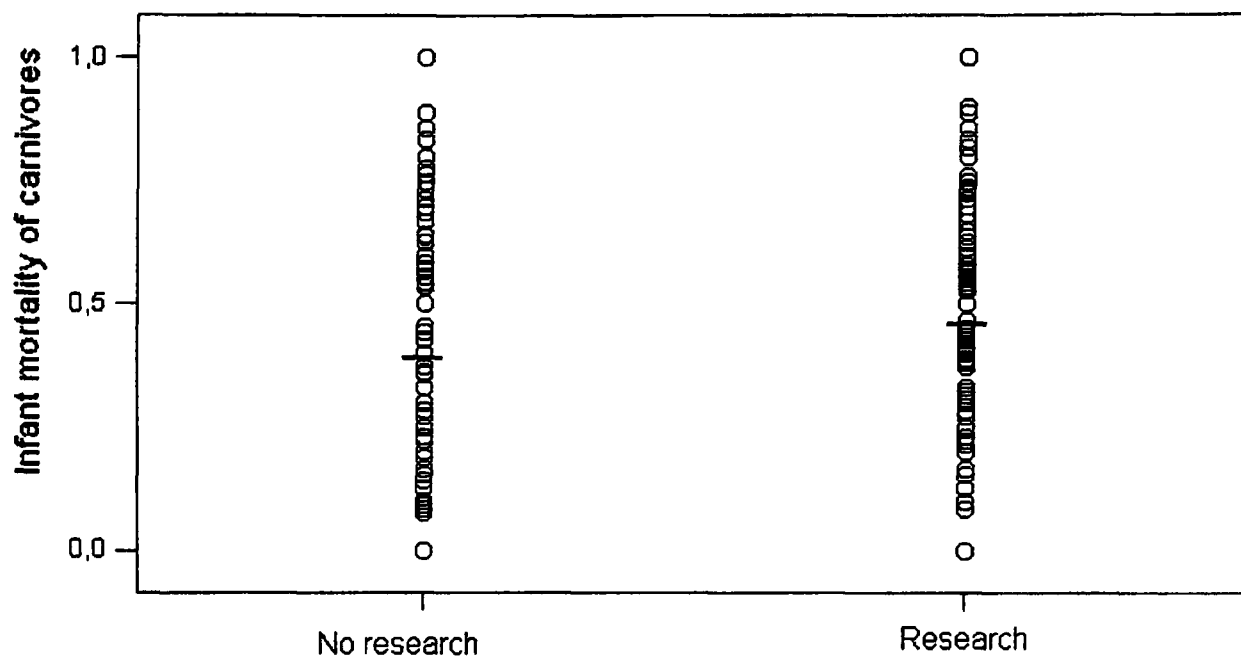


Figure 7.1: Relative proportion of infant mortality in 98 species carnivores housed in 461 zoological institutions with (N = 151) and without research facilities (N = 310). Solid lines represent means.

Table 7.2: General linear model (GLM) of the effect of latitude of the zoo (LAT), presence/absence of researchers (RES) and management regimen of the institution (MAN) on the institution's success in breeding carnivores.

Variable	DF	F	P
LAT	1	0.26	0.611
RES	1	6.31	0.012
MAN	2	0.06	0.942

The distance travelled by the female was positively correlated with the proportion of infant mortality (Figure 7.2), while the number of dens and the available area for each female (adjusted for each species) were negatively correlated with the proportion of infant mortality (Table 7.3, Figure 7.3). The multiple regression model with all three predictors produced R^2 (adj.) = 21%, $F_{3,63} = 5.384$, $p = 0.008$.

Table 7.3: Correlations and results from the regression analysis of distance travelled by the dam (DTD), number of dens in the dam's enclosure (NAD) and relative available area (AIA) on the proportion of infant mortality (PIM) in captive carnivores.

Variable	Correlation with PIM	B	β
DTD	2.403	0.270	0.341*
NAD	-2.748	-0.485	-0.390**
AIA	-2.146	-1.174	-0.245*

* $p < 0.05$; ** $p < 0.001$

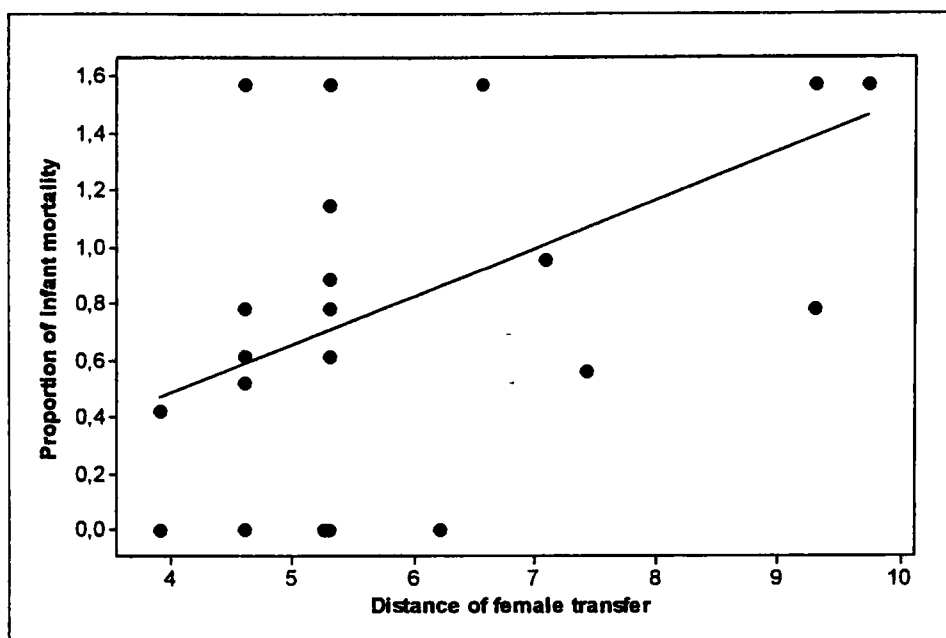


Figure 7.2: Scatterplot of the median proportion of mortality in 66 litters from 45 females of 9 species of captive carnivores in relation to the distance travelled by the female (log-transformed).

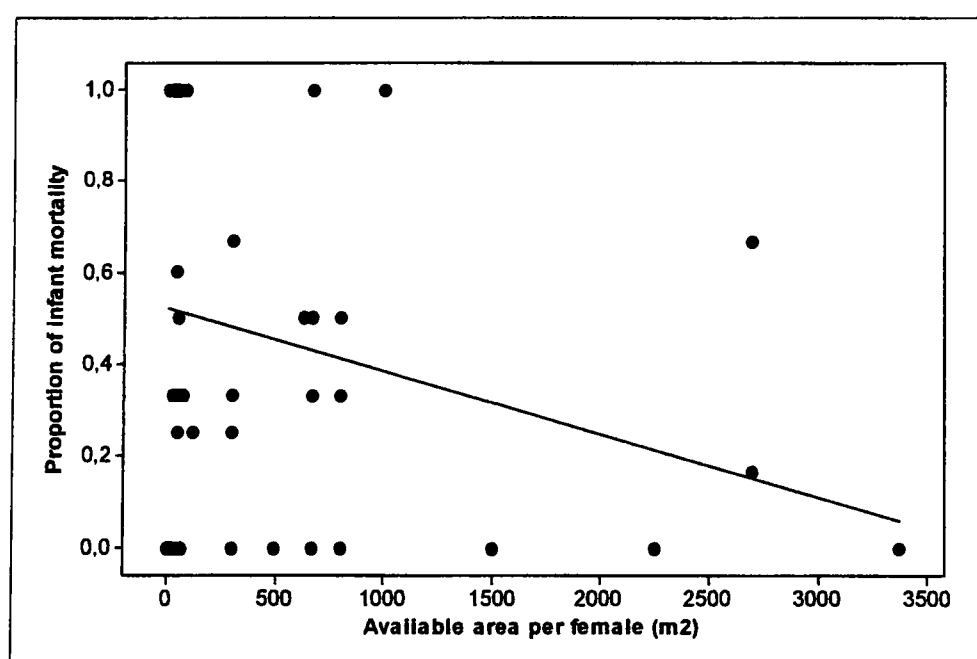


Figure 7.3: Scatterplot of the proportion of mortality in 74 litters of 39 females of 9 species of captive carnivores in relation to the relative available area per female (m²).

Transferring the females from one institution to other, or from the wild to any institution, can affect infant mortality even when not considering distances; the latitude of the zoo or the number of keepers in contact with the animals do not seem to affect infant survival (Table 7.4).

Table 7.4: Logistic regression results with categorical variables related to husbandry practices on captive carnivores; the dependent variable is a binary measure of infant mortality per female (148 litters from 60 females of 9 species).

Variable	Model 1		Model 2		Model 3	
	B	Wald	B	Wald	B	Wald
Constant	- 0.16	0.319	1.813	2.953*	2.232	1.199
LATITUDE OF ZOO /SPECIES RANGE	- 0.022	0.002	- 0.733	1.483	- 0.817	1.033
DAM TRANSFER	-	-	- 1.152	3.791**	- 1.234	2.898*
No. OF KEEPERS	-	-	-	-	- 0.049	0.04
Model Chi-Square [df]	0.002 [1]		4.116 [2]*		0.040 [3]	
Block Chi-Square [df]	0.002		4.113 [1]**		0.040 [1]	
% corrected prediction	54.2		59		59	
Nagelkerk's R²	0.02		0.15		0.15	

* p<0.1; ** p<0.05.

In this dataset, the two hand-reared females had a lower proportion of infant mortality than mother-reared ones, although a larger sample would be needed; with this sample, a one-way ANOVA test provided $F_{1, 34} = 3.82$, $p = 0.059$ (Figure 7.4).

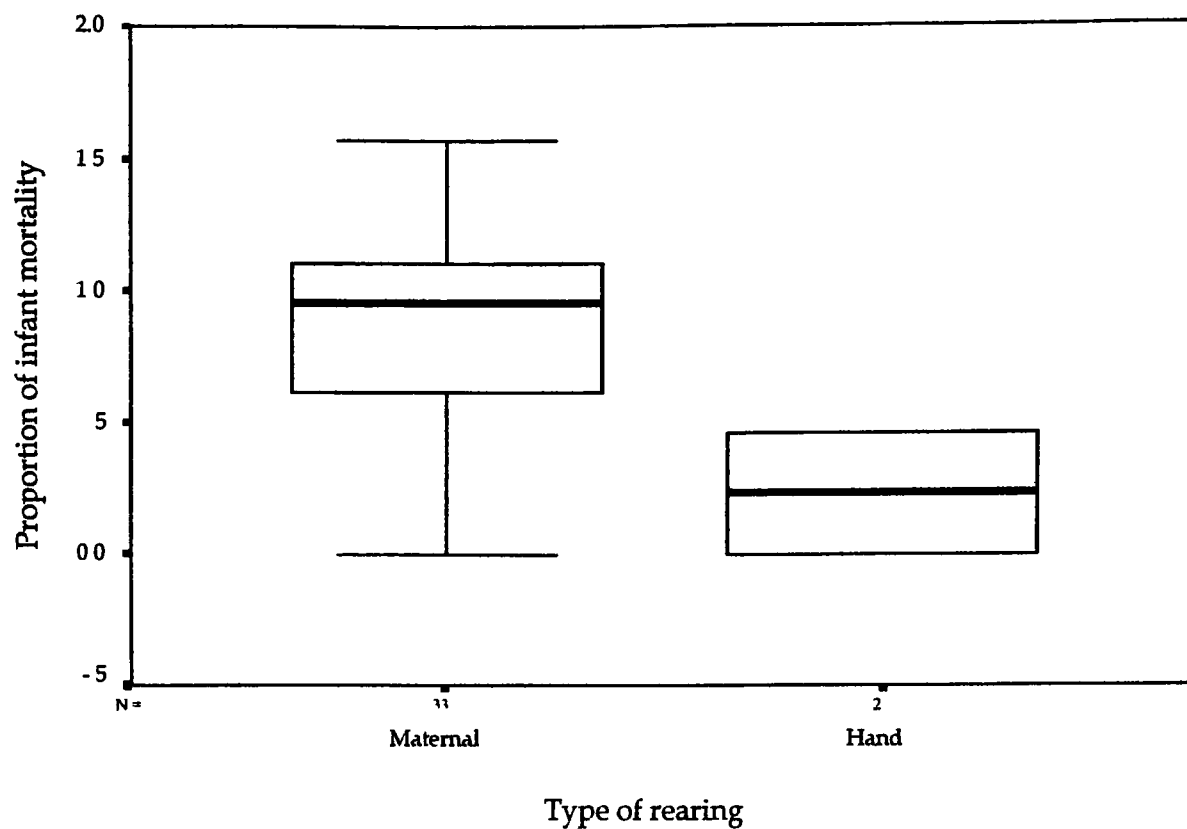


Figure 7.4: Proportion of infant mortality in mother-reared and hand-reared female captive carnivores.

7.6. Discussion.

7.6.1. Housing conditions.

There is a significant effect of the presence of researchers in the breeding success of captive carnivores, with females housed in institutions performing research presenting higher infant mortality. Research in zoological institutions has been growing since the beginning of the 1980s (Benirschke 1987; Finlay & Maple 1986; Stoinski, Lukas & Maple 1998) and it generally involves the direct observation and manipulation of animals or, at least, changes in the environment (Appleby 1997; Robinson 1998). Recently, researchers have been pointing out the need for non-invasive techniques for metabolical monitoring of stress (Wielebnowski 1999; Wielebnowski *et al.* 2002) and breeding (Graham *et al.* 1995; Brown *et al.* 1996; Bircher & Noble 1997), and also for the need to develop reliable ethological methods to assess welfare (Dawkins 2003), to reduce the stress caused by capture and blood-sampling in the individuals. It

is generally agreed that direct manipulation and other environmental stressors affect the individual's welfare and can reduce breeding success (Bakken *et al.* 1999), but some researchers are calling for the use of more invasive research tools for captive non-domestic species in captivity, disregarding the findings that do not recommend this approach (Goodrowe 2003). This research, however, found that the impact of research, even non-invasive ones, in the breeding success of captive carnivores can be stronger than previously thought, although a larger sample size would be needed to better support this view.

In this research, there was a significant effect of some aspects of housing in the proportion of infant mortality. Although there are other aspects of the enclosure that should be researched, in this dataset only the number of dens and the available individual area were added to the model, both presenting strong effects on the dependent variable.

Housing conditions are known to affect the welfare and breeding success in many species of carnivores, but different species have different demands in captivity and results vary. For example, clouded leopards are sensitive to husbandry and housing conditions: faecal corticoid measurements were lower in individuals with higher enclosures and those with few keepers that spent more time weekly interacting with them; animals with multiple keepers and those in public exhibition, especially the ones housed in the proximity of potential predators (Wielebnowski *et al.* 2002). A study on captive leopards *Panthera pardus* achieved different results: off-exhibit animals had higher levels of stereotypical behaviour than animals in exhibition, but off-exhibit enclosures were indoors, while exhibition enclosures were larger and mostly outdoors (Mallapur & Chellam 2002).

In this dataset, the number of dens correlated negatively with the proportion of mortality. The type and number of dens had been correlated with levels of cortisol, but again specific preferences may differ. A study with farmed silver foxes *Vulpes vulpes* and blue foxes *Alopex lagopus* determined that

these two species have different preferences regarding the design of nest boxes, although urinary cortisol levels, used as a measure of stress, did not vary significantly as long as a nest box was provided (Jeppesen, Pedersen & Heller 2000). Other study showed that cortisol levels were significantly lower in female silver foxes that were provided with a nest box than those that were kept in wire mesh cages with no nest box, suggesting that stress levels can be connected with the presence of a secluded space for nesting (Jeppesen & Pedersen 1991). It would be interesting to research the preferred design of nest boxes in other wild species of carnivores that are not commercially exploited.

Housing conditions also seem to affect leopard cats *Prionailurus bengalensis*, which can be detected through non-invasive monitoring, and the welfare of individuals can be ameliorated through environmental enrichment (Carlstead *et al.* 1993). Simple enrichment techniques, such as the placement of scents and new objects in the enclosure, turning the environment more complex, and feeding devices that increase foraging time, had also very positive results in the behaviour of encaged African lions *Panthera leo* (Powell 1995).

The lack of effect of the number of keepers may be caused by the small variation of this characteristic in the dataset. Further research with larger sample sizes is recommended.

7.6.2. *Individual history.*

This dataset showed a very strong positive correlation of the distance travelled by the dam and the proportion of infant mortality, and this result was confirmed even when the actual distance was not considered, suggesting that any transfer of females can increase infant mortality. Inter-zoo breeding loans are common, especially of species that are under captive breeding programmes, and both males and females can travel distances as far as from Beijing to London (Block & Perkins 1996). There are no further studies on the effect of this practice in breeding success, but this result suggest that transferring females can have an impact in the success of captive breeding programmes and should be researched using a larger sample.

A larger sample would be necessary also to identify the real impact of rearing techniques in infant mortality. The low proportion of infant mortality in this research may reflect the relative facility of keeping these females, once hand-reared carnivores tend to prefer the company of humans to their own species (Read & Meier 1996). On the other side, hand-reared animals can present some behavioural problems. For example, in sloth bears not only the enclosure type and the composition of the captive group affected activity, but also the individual's rearing history: stereotypical behaviour, including self-directed activities, was more common in hand-reared animals than in mother-reared ones (Forthman & Bakeman 1992). Hand-rearing also seems to have an effect on the development of young giant pandas *Ailuropoda melanoleuca*: hand-reared or partially hand-reared cubs grew slower than mother-reared cubs up to 6 months of age, although there was no significant difference after that age (Zhang *et al.* 1996). Response to hand-reared can also have the opposite effect in other species: a study revealed that the use of highly energetic milk replacements in hand-reared brown hyaena pups led to a faster weight gain on those, when compared to mother-reared ones, although the growth rate was equalised between groups after weaning (Volf 1998).

Apart from differences in housing and rearing, individuals can cope differently with levels of stress, as it was shown in a study on beech martens *Martes foina*; according to the type of cortical response, it was suggested that individuals can be classified in high activity individuals (Type A), which are more prone to display stereotypical behaviour and cope poorly in captivity, and low activity individuals (Type B), which would cope better with captive conditions (Hansen & Damgaard 1993).

Unfortunately, the low level of response from the institutions did not allow more robust data analyses. It is possible, however, that husbandry techniques are affecting the breeding success of captive carnivores in a much broader way than it was considered until now, and further analysis could elucidate which species are more sensitive to hand-rearing and translocations.

7.7. Conclusion.

Certain aspects of husbandry and housing conditions of captive carnivores can affect the reproductive output of females. If a species is already prone to higher levels of infant mortality, either by inadequate maternal behaviour or the tendency to stillbirths and exposure to infectious diseases, the conditions in which they are kept can increase the frequency of infant deaths. However, further research is needed to determine safely these factors, once there are many uncontrolled factors in the institutions that have the potential to affect the individuals' welfare and, ultimately, the breeding success of these individuals. Certain common practices of institutions, such as transferring females in breeding loans to other institutions, may have to be reviewed.

We cannot forget that the potential of captive populations is varied. For example, many species are at the verge of extinction, and conservation efforts that last for more than 5 generations of the species have not improved its status. It is possible that some of these species will only be found in captivity, and all the human knowledge will be therefore based on zoo specimens. Important factors come from this possibility: there is not enough research on several species in the wild, and biological parameters are unknown in many. Also, many ethograms are based in captive observations. Unless research in the wild, sometimes of the most basic descriptive nature, is immediately done, many adaptive phenomena will disappear before they can be fitted into the evolutionary chain.

The lack of knowledge in the wild also impairs the correct development of environmental enrichment techniques. Many of them are based on the premise of facilitating natural behaviour. Obviously, if the natural behaviour of the species is unknown, there is no parameter on which to base the results of the experiments.

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Appendices

Appendix 1 (3 pages): Data on 98 species of carnivores in captivity, collected from the *International Zoo Yearbooks* Vols. 26-35. Meanings and abbreviations: IUCN Red List = status of the species in the IUCN Red List 2002; Max. Latitude North = Northernmost latitude of distribution of the species (Wilson & Reeder 1993); Max. Latitude South = Southernmost latitude of distribution of the species (Wilson & Reeder 1993); No. of Zoos = Number of zoological institutions keeping the species; Infant Mortality = median proportion of infant mortality of the species in captivity.

Appendix 2 (32 pages): Data on 535 zoological institutions collected from the *International Zoo Yearbooks*. Meanings and abbreviations: Zoo code = short name for the institutions; Carnivora sp. = Number of species of carnivores kept by the institution; Underbred sp. = Number of species of carnivores in the institution that have proportion of infant mortality higher than the median for the species (Appendix 1); Index = Institutional breeding performance, calculated by Equation 2.3; Attendance = Total annual attendance of the institution; SpMammals = Total of mammalian species kept by the institution; Total Sp = Total of species, including invertebrates, kept by the institution; IndMammal = Total number of mammalian individuals in the institution.

Appendix 3 (16 pages): Data on 249 litters from 126 female tigers (*Panthera tigris*) housed in 116 institutions, collected from the International Tiger Studbook. Meanings and abbreviations: M+ = Number of males born; F+ = Number of females born; U+ = Number of young of undetermined sex born; M- = Number of young males dead; F- = Number of young females dead; U- = Number of young of undetermined sex dead; Rearing: M = Maternal, H = Hand-rearing, U = unknown; Inbreeding = inbreeding level of the litter.

Appendix 4 (22 pages): Data on 447 litters born from 212 females of 69 species of captive carnivores, collected from 141 papers and reports. Meanings and abbreviations: Origin = Place of birth of the dam; Age = Age of the dam at parturition; Infant mortality = "Young dead" divided by "litter size"; Dens = Number of dens available to the female; Male separated? = Presence (No) or absence (Yes) of the male during nursing.

Appendix 5 (2 pages): Sample of questionnaires sent to zoological institutions. Page 1 = Keys and samples; Page 2: Form to be filled by the institutions.

Appendix 6 (10 pages): Data on 148 litters born from 63 females of 9 species of carnivores housed in 16 institutions. Meanings and abbreviations: Travel (km) = Distance in kilometres travelled by the female between capture site to the institutions (for wild-caught females) or between translocations due to breeding loans or trade (for captive-born); Origin = Local of birth of the dam; Ind/cage = Number of adult individuals in the enclosure; no of dens = number of dens available to the female; no of keepers = number of persons in direct contact with the animals.

Appendix 1, page 1

Family	Genus	Species	IUCN Red List	Max. latitude North	Max. latitude South	No. of zoos	Infant mortality
Canidae	<i>Alopex</i>	<i>lagopus</i>		80 N	60 N	40	0.18466
Canidae	<i>Canis</i>	<i>aureus</i>				22	0.25865
Canidae	<i>Canis</i>	<i>familiaris</i>		World	World	34	0.08846
Canidae	<i>Canis</i>	<i>lupus</i>	EW, LR, VU	78 N	20 S	154	0.2
Canidae	<i>Canis</i>	<i>mesomelas</i>				11	0.25
Canidae	<i>Canis</i>	<i>rufus</i>	CR			16	0.18871
Canidae	<i>Cerdocyon</i>	<i>thous</i>				7	0.42857
Canidae	<i>Chrysocyon</i>	<i>brachyurus</i>		5 S	45 S	70	0.5
Canidae	<i>Cuon</i>	<i>alpinus</i>				11	0.32
Canidae	<i>Lycan</i>	<i>pictus</i>		15 N	30 S	52	0.70894
Canidae	<i>Nyctereutes</i>	<i>procyonoides</i>		50 N	22 S	57	0.27586
Canidae	<i>Otocyon</i>	<i>megalotis</i>				10	0.875
Canidae	<i>Speothos</i>	<i>venaticus</i>	VU			29	0.66667
Canidae	<i>Urocyon</i>	<i>cinereoargenteus</i>				13	0.28571
Canidae	<i>Vulpes</i>	<i>corsac</i>				16	0.43922
Canidae	<i>Vulpes</i>	<i>rueppelli</i>				4	0.17046
Canidae	<i>Vulpes</i>	<i>velox</i>	LR			6	0.18182
Canidae	<i>Vulpes</i>	<i>vulpes</i>		60 N	26 N	68	0.33333
Canidae	<i>Vulpes</i>	<i>zerda</i>		30 N	15 N	46	0.55
Felidae	<i>Acinonyx</i>	<i>jubatus</i>	VU	40 N	34 S	54	0.23476
Felidae	<i>Caracal</i>	<i>caracal</i>		40 N	34 S	48	0.26136
Felidae	<i>Catopuma</i>	<i>temmincki</i>	LR			9	0
Felidae	<i>Felis</i>	<i>chaus</i>		50 N	5 N	46	0.4
Felidae	<i>Felis</i>	<i>margarita</i>	LR			13	0.2
Felidae	<i>Felis</i>	<i>nigripes</i>				16	0.13816
Felidae	<i>Felis</i>	<i>silvestris</i>	VU	58 N	34 S	74	0.28348
Felidae	<i>Herpailurus</i>	<i>yaguarondi</i>	EN			16	0.16667
Felidae	<i>Leopardus</i>	<i>pardalis</i>	EN	37 N	33 S	59	0.28571
Felidae	<i>Leopardus</i>	<i>tigrinus</i>	LR			4	0.90909
Felidae	<i>Leopardus</i>	<i>wiedii</i>	LR			13	0.25
Felidae	<i>Leptailurus</i>	<i>serval</i>	EN	30 N	5 N	100	0.41144
Felidae	<i>Lynx</i>	<i>canadensis</i>				18	0.19091
Felidae	<i>Lynx</i>	<i>lynx</i>		78 N	32 N	109	0.23077
Felidae	<i>Lynx</i>	<i>rufus</i>		52 N	45 N	40	0.0625
Felidae	<i>Neofelis</i>	<i>nebulosa</i>	VU			23	0.19149

Family	Genus	Species	IUCN Red List	Max. latitude North	Max. latitude South	No. Of zoos	Infant mortality
Felidae	<i>Oncifelis</i>	<i>geoffroyi</i>				21	0.33333
Felidae	<i>Otocolobus</i>	<i>manul</i>	LR			3	0.90909
Felidae	<i>Panthera</i>	<i>leo</i>	EN			22	0.4069
Felidae	<i>Panthera</i>	<i>onca</i>	LR	35 N	35 S	104	0.20526
Felidae	<i>Panthera</i>	<i>pardus</i>	EN, CR	55 N	10 S	85	0.25
Felidae	<i>Panthera</i>	<i>tigris</i>	EN, CR	62 N	10 S	230	0.33333
Felidae	<i>Prionailurus</i>	<i>bengalensis</i>	EN	57 N	5 S	64	0.33809
Felidae	<i>Prionailurus</i>	<i>rubiginosus</i>	DD			5	0.33333
Felidae	<i>Prionailurus</i>	<i>viverrinus</i>	LR			19	0.28571
Felidae	<i>Puma</i>	<i>concolor</i>	CR	55 N	40 S	102	0.25
Felidae	<i>Uncia</i>	<i>uncia</i>	EN	60 N	27 N	86	0.15152
Herpestidae	<i>Atilax</i>	<i>paludinosus</i>				6	0.51731
Herpestidae	<i>Cryptoprocta</i>	<i>ferox</i>	VU			2	0.32738
Herpestidae	<i>Cynictis</i>	<i>penicillata</i>				6	0.28205
Herpestidae	<i>Helogale</i>	<i>parvula</i>				19	0.55556
Herpestidae	<i>Mungos</i>	<i>mungo</i>		10 N	30 S	33	0.33333
Herpestidae	<i>Suricata</i>	<i>suricata</i>		4 S	34 S	109	0.37931
Hyanidae	<i>Crocuta</i>	<i>crocuta</i>	LR			16	0.38095
Hyanidae	<i>Hyaena</i>	<i>hyaena</i>				24	0.34848
Hyanidae	<i>Proteles</i>	<i>cristatus</i>	DD			8	0.64103
Mustelidae	<i>Amblyonyx</i>	<i>cinereus</i>	LR	28 N	10 S	57	0.15385
Mustelidae	<i>Eira</i>	<i>barbara</i>	VU			7	0.33333
Mustelidae	<i>Enhydra</i>	<i>lutris</i>				16	0.24265
Mustelidae	<i>Gulo</i>	<i>gulo</i>	VU			8	0.6
Mustelidae	<i>Ictonyx</i>	<i>striatus</i>				2	0
Mustelidae	<i>Lontra</i>	<i>longicaudis</i>				2	0.65
Mustelidae	<i>Lutra</i>	<i>canadensis</i>				12	0
Mustelidae	<i>Lutra</i>	<i>lutra</i>		55 N	10 S	31	0.03371
Mustelidae	<i>Martes</i>	<i>flavigula</i>	EN			7	1
Mustelidae	<i>Martes</i>	<i>foina</i>				9	0
Mustelidae	<i>Martes</i>	<i>martes</i>				8	0.33333
Mustelidae	<i>Martes</i>	<i>zibellina</i>				3	0.33333
Mustelidae	<i>Meles</i>	<i>meles</i>				24	0.41667
Mustelidae	<i>Mephitis</i>	<i>mephitis</i>				27	0.30769
Mustelidae	<i>Mustela</i>	<i>erminea</i>				3	0.125

Family	Genus	Species	IUCN Red List	Max. latitude North	Max. latitude South	No. Of zoos	Infant mortality
Mustelidae	<i>Mustela</i>	<i>eversmannii</i>	VU			15	0.16667
Mustelidae	<i>Mustela</i>	<i>lutreola</i>	EN			4	0.18546
Mustelidae	<i>Mustela</i>	<i>nigripes</i>	EW			6	0.48849
Mustelidae	<i>Mustela</i>	<i>nivalis</i>				4	0.2381
Mustelidae	<i>Mustela</i>	<i>putorius</i>				29	0.04651
Mustelidae	<i>Mustela</i>	<i>vison</i>				12	0
Mustelidae	<i>Pteronura</i>	<i>brasiliensis</i>	VU			3	1
Mustelidae	<i>Vormela</i>	<i>peregusna</i>	VU			5	0.4
Procyonidae	<i>Ailurus</i>	<i>fulgens</i>	EN	32 N	25 N	83	0.2
Procyonidae	<i>Bassariscus</i>	<i>astutus</i>				4	0.55
Procyonidae	<i>Nasua</i>	<i>narica</i>				25	0.2
Procyonidae	<i>Nasua</i>	<i>nasua</i>		10 N	35 S	114	0.29151
Procyonidae	<i>Potos</i>	<i>flavus</i>		23 N	20 S	30	0.05
Procyonidae	<i>Procyon</i>	<i>cancrivorus</i>				11	0.33333
Procyonidae	<i>Procyon</i>	<i>lotor</i>		50 N	6 S	102	0.2
Ursidae	<i>Helarctos</i>	<i>malayanus</i>				23	0.4
Ursidae	<i>Melursus</i>	<i>ursinus</i>	VU			15	0.33333
Ursidae	<i>Tremarctos</i>	<i>ornatus</i>	VU	10 N	20 S	32	0.31667
Ursidae	<i>Ursus</i>	<i>americanus</i>		65 N	22 N	33	0.11111
Ursidae	<i>Ursus</i>	<i>arctos</i>		70 N	32 N	121	0
Ursidae	<i>Ursus</i>	<i>maritimus</i>	LR	80 N	60 N	63	0.66667
Ursidae	<i>Ursus</i>	<i>thibetanus</i>	VU, CR	50 N	5 N	54	0.125
Viverridae	<i>Arctictis</i>	<i>binturong</i>	VU	25 N	10 S	31	0.33333
Viverridae	<i>Civettictis</i>	<i>civetta</i>	CR			4	0.70175
Viverridae	<i>Genetta</i>	<i>genetta</i>	VU			21	0.25
Viverridae	<i>Genetta</i>	<i>tigrina</i>				5	0.21053
Viverridae	<i>Paguma</i>	<i>larvata</i>				9	0.3
Viverridae	<i>Paradoxurus</i>	<i>hermaphroditus</i>	VU			15	0.45833

A	B	C	D	E	F
Zoo code	Official name	City	Country	Carnivora sp	Underbred sp
1					
2	Aachener Tierpark	Aachen	Germany	7	3
3	Aalborg Zoologiske Have	Aalborg	Denmark	9	5
4	Abilene Zoological Gardens	Abilene, Texas	USA	1	0
5	Al Ain Zoo and Aquarium	Abu Dhabi	U of Arab Emirates	6	0
6	Cleland Wildlife Park	Adelaide	Australia	12	4
7	Parco Faunistico 'La Torbiera'	Agrate Conturbia	Italy	14	5
8	Kamla Nehru Zoological Gardens	Ahmedabad	India	1	0
9	Zoo Ahtari	Ahtari	Finland	2	0
10	Omiyama Zoo	Akita	Japan	7	4
11	Albuquerque Biological Park	Albuquerque, New Mexico	USA	6	0
12	Alexandria Zoological Park	Alexandria, Louisiana	USA	3	0
13	Drusillas Zoo Park	Alfriston	England	3	3
14	Fonden Bornholms Dyre-og Naturpark	Allinge	Denmark	3	0
15	Alma-Atinskii Zoopark	Alma-Ata	Kazakhstan	19	13
16	Amagi Wild Boar Park	Amagi-Yugashima	Japan	1	0
17	Dierenpark Amersfoort	Amersfoort	Netherlands	9	8
18	Parc Zoologique de la Ville d'Amiens	Amiens	France	1	0
19	Parc Zoologique du Bois de Coulange	Amneville	France	11	7
20	Stichting Koninklijk Zoologisch Genootschap Natura Artis Magistra	Amsterdam	Netherlands	15	6
21	Ankara Zoo	Ankara	Turkey	3	0
22	Antwerp Zoo	Antwerp	Belgium	4	1
23	Stichting Apenheul	Apeldoorn	Netherlands	1	0
24	Burgers Zoo and Safari	Arnhem	Netherlands	13	4
25	Tenerife Zoo	Los Cristianos (form Arona)	Canary Islands	1	1
26	Asahiyama Zoo	Asahikawa	Japan	6	5
27	North Carolina Zoological Park	Asheboro, North Carolina	USA	2	1
28	Turkmenistan Dovlet Zoology Bogy	Ashkhabad	Turkmenistan	4	1
29	Zoo Atlanta/Atlanta-Fulton County Park	Atlanta, Georgia	USA	1	1
30	Auckland Zoological Park	Auckland	New Zealand	3	2
31	Zoologischer Garten Augsburg	Augsburg	Germany	6	2
32	Sea World Of Ohio	Aurora, Ohio	USA	1	1
33	Monde Sauvage Safari	Aywaille	Belgium	4	1
34	California Living Museum (CALM)	Bakersfield, California	USA	1	1
35	Baku Zoo	Baku	Azerbaijan	6	4

Appendix 2, page 2

	G		H		I	J	K		L		M		N		O		P		Q		R		
	Index	Latitude	Latitude	Longitude			No. Of vets	Research	Staff	Size (ha)	Attendance	SpMammals	Total Sp	IndMamm	Total indiv	Management							
1																							
2	0.428571	50n47			3	No	12	9	164239	39	239	209	1332	Private									
3	0.555556	57n03			1	No	37	16	310000	40	251	129	698	Private									
4	0	32n27			1	No	13	6	142401	25	158	76	491	City/Zool. society									
5	0	24n28			1	No	190	450	290000	104	403	1061	7764	City/National									
6	0.333333	34s55			1	No	18	26	109742	109	98	529	732	National									
7	0.357143	45n34			3	No	5	10	58000	30	125	121	545	Private									
8	0	23n02				No	200	12	2465000	50	450	215	2250	City									
9	0	62n34				No	17	60	179698	29	136	59	290	Private									
10	0.571429	39n43			1	No	16	8	173102	41	228	110	515	City									
11	0	35n05			1	No	88	25	490974	57	267	302	1068	City/Zool. society									
12	0	31n19			1	No	7	9	125000	33	92	107	387	City									
13	1	50n48			1	No	8	15	145972	39	150	103	369	Private									
14	0	55n16				No																	
15	0.684211	43n15			1	Yes	220	22	976977	90	336	249	1522	City									
16	0	34n53			1	Yes	15	33	369808	3	120			Private									
17	0.888889	52n09			2	No	27	18	190000	43	102	305	823	Private									
18	0	49n54				No	21	7	92000	41	127	89	252	City									
19	0.636364	49n22				No																	
20	0.4	52n22			1	No	150	10	1007310	129	1103	763	6173	Zool. society									
21	0	39n56			2	No	54	12	622800	37	119	160	1223	National									
22	0.25	51n13			2	Yes	209	10	935166	142	774	799	5260	Zool. society/National									
23	0	52n13			1	No	15	12	460000	17	23	294	322	Private									
24	0.307692	51n59			1	Yes	50	44	1064000	67	261	382	1535	Private									
25	1	28n0				No																	
26	0.833333	43n46			1	No	17	14	509245	42	181	128	594	City									
27	0.5	35n42			2	Yes	176	121	520707	54	151	317	816	State/Zool. society									
28	0.25	37n57			1	No	40	32	133800	34	98	83	403	City									
29	1	33n45			1	No	51	12	567500	38	264	77	800	City/Zool society									
30	0.666667	36s52				Yes	45	19	333000	51	195	305	1249	City									
31	0.333333	48n23			1	No	45	22	457773	70	366	380	1933	City									
32	1	41n19			1	No	100	32	1300000	9	1550	57	4117	Private									
33	0.25	50n28				No																	
34	1	35n22			1	No	2	52	27310	20	63	57	184										
35	0.666667	40n23			1	No	42	45	297186	32	143	120	792	City									

A	B	C	D	E	F
Zoo code	Official name	City	Country	Carnivora sp	Underbred sp
36					
37	Curraugh Wildlife Park	Ballaugh	Isle of Man	3	3
38	Baltimore Zoo	Baltimore, Maryland	USA	3	1
39	Kebun Binatang Bandung/Yayasan Margasatwa Tamansari	Bandung	Indonesia	3	1
40	Dusit Zoo	Bangkok	Thailand	7	0
41	Banham Zoo	Banham (Norwich)	England	11	7
42	Parc Zoologic de Barcelona SA	Barcelona	Spain	7	5
43	Servicio Autónomo Parque Zoológico y Botánico Baranda	Barquisimeto	Venezuela	11	6
44	Zoologischer Garten Basel	Basle	Switzerland	12	2
45	Greater Baton Rouge Zoo	Baton Rouge, Louisiana	USA	12	1
46	Howletts Wild Animal Park	Bekesbourne	England	13	2
47	Belfast Zoological Gardens	Belfast	Northern Ireland	5	2
48	Belize Zoo & Tropical education Center	Belize City	Belize	2	0
49	Fundação Zoo-Botânica de Belo Horizonte	Belo Horizonte, MG	Brazil	7	3
50	Tierpark Berlin - Friedrichsfelde	Berlin	Germany	29	16
51	Zoologischer Garten und Zoo-Aquarium Berlin	Berlin	Germany	30	21
52	Tierpark Daehloelzi	Berne	Switzerland	15	7
53	West Midland Safari and Leisure Park Ltd	Bewdley	England	3	2
54	Nandankanan Zoological Park	Bhubaneswar	India	5	3
55	Ross Park Zoo	Binghamton, New York	USA	4	2
56	Birmingham Nature Centre	Birmingham	England	5	2
57	Dakota Zoo	Bismarck, North Dakota	USA	8	3
58	National Zoological Park	Bissau	Guinea-Bissau	1	0
59	Blackpool Zoopark	Blackpool	England	3	2
60	Blair Drummond Safari and Leisure Park	Blair Drummond	Scotland	1	0
61	Bloemfontein Zoo	Bloemfontein	South Africa	7	4
62	Miller Park Zoo	Bloomington, Illinois	USA	3	1
63	Tierpark + Fossilium Bochum	Bochum	Germany	5	1
64		Bogor	Indonesia	3	0
65	Zoologicka Zahrada Bojnice	Bojnice	Slovakia	12	9
66	Bolsherech'yenskii Zoopark	Bolsherech'ye	Russia	13	1
67	Veermata Jijabai Bhosle Udyan - Zoo	Bombay	India	2	1
68	Thaba-Ya-Batho Zoological Gardens	Bon Accord	South Africa	2	0
69	Boras Djurpark	Boras	Sweden	3	1
70	Jardim Zoológico de Brasília	Brasilia, DF	Brazil	7	6

	G		H		I	J	K	L	M	N		O		P		Q	R
	Index	Latitude	Latitude	Size (ha)						Attendance	SpMammals	Total Sp	IndMamm	Total indiv	Management		
36																	
37	1	54n20		8	40024	19	92	117	1073	State							
38	0.333333	39n17	1	85	329052	65	168	310	908	City/Zool. society							
39	0.333333	6s54								Zool. society/State/City							
40	0	13n45	2	172	2311666	82	260	323	1378	National							
41	0.636364	52n38	1	12	95000	44	340	138	668	Private							
42	0.714286	41n23		128	1000000	111	516	624	8637	City							
43	0.545455	10n04	2	75	441317	63	129	255	1281	National							
44	0.166667	47n33	1	71	885175	73	576	506	4386	Private							
45	0.083333	30n27	1	41	270000	78	206	366	858	City							
46	0.153846	51n17	2	47	124225	46	357	52	411	Charity							
47	0.4	54n35	1	42	192000	47	325	107	610	City							
48	0	17n30															
49	0.428571	19s4901	4	115	993307	55	150	282	1104	City							
50	0.551724	52n30	12	449	2515301	210	2199	959	6650	City							
51	0.7	52n30	2	40	2663165	266	1403	1785	12203	Private							
52	0.466667	46n57	2	25	146670	54	152	241	783	City							
53	0.666667	52n22	1	8	360000	35	400	36	405	Private							
54	0.6	20n14	2	110	954335	53	494	154	1616	National							
55	0.5	42n06	1	20	106000	23	63	55	145	City/Zool. society							
56	0.4	52n30		4	160000	30	193	73	280	City							
57	0.375	46n49	2	6	125000	59	142	261	657	Private/Zool. society							
58	0	11n51															
59	0.666667	53n50	1	33	308048	36	185	111	579	City							
60	0	56n07	1	9	131684	23	161	27	208	Private							
61	0.571429	29s12		28	107480	78	146	384	1246	City							
62	0.333333	40n29	2	5	101834					City/Zool. society							
63	0.2	51n28	2	18	231045	18	101	247	1093	Zool. society							
64	0	6s35															
65	0.75	48n47	1	62	531000	67	308	262	1439	City							
66	0.076923	55n0	1	39		67	141	130	385	City							
67	0.5	18n58	2	61	2999200	35	185	131	985	City							
68	0	25s38															
69	0.333333	57n43	1	20	280000	45	142	230	685	City							
70	0.857143	15s4647	1	108	366675	35	129	134	801	National							

	A	B	C	D	E	F
	Zoo code	Official name	City	Country	Carnivora sp	Underbred sp
71	Bratisla	Zoologicka Zahrada Bratislava	Bratislava	Slovakia	11	3
72	Bremerha	Zoo Am Meer	Bremerhaven	Germany	3	0
74	Bridgepor	Beardsley Zoological Gardens	Bridgeport, Connecticut	USA	2	2
75	Bridgeto	Cohanzick Zoo	Bridgeton, New Jersey	USA	1	0
76	Bristol	Bristol Zoo Gardens	Bristol	England	4	1
77	Brno	Zoologicka Zahrada Mesta Brna	Brno	Czech Republic	16	8
78	Brownsvi	Gladys Porter Zoo	Brownsville, Texas	USA	4	1
79	Buchares	Gradina Zoologica Bucuresti	Bucharest	Romania	9	6
80	Budapest	Zoological and Botanical Garden of the City of Budapest	Budapest	Hungary	22	9
81	BuenosAi	Jardin Zoológico de la Ciudad de Buenos aires	Buenos Aires	Argentina	8	2
82	Buffalo	Buffalo Zoological Gardens	Buffalo, New York	USA	11	6
83	Bungay	Otter Trust	Bungay	England	2	0
84	Burford	Cotswold Wildlife Park	Burford	England	8	8
85	Bydgoszc	Forest Park of Culture and Rest-The Garden of polish Fauna	Bydgoszcz	Poland	6	4
86	Calcutta	Zoological Garden Alipore	Calcutta	India	4	0
87	Calgary	Calgary Zoo, Botanical Gardens & Prehistoric Park	Calgary	Canada	12	1
88	Cali	Zoológico de Cali	Cali	Colombia	12	6
89	Cambridg	Cambridge Zoo	Cambridge	Canada	1	0
90	Catskill	Catskill Game Farm Inc	Catskill, New York	USA	1	0
91	Chandiga	M C Zoological Park Chhart-Bir	Chandigarh	India	1	0
92	Chard	Cricknet St Thomas Wildlife Park	Chard	England	6	3
93	Chemnitz	Tierpark Chemnitz	Chemnitz	Germany	3	0
94	Cherkass	Cherkassy Zoo	Cherkassy	Ukraine	5	3
95	Chessing	Chessington World of Adventure	Chessington	England	7	4
96	Chester	Chester Zoo	Chester	England	19	6
97	Chiba	Chiba Zoological Park	Chiba	Japan	4	1
98	ChicagoB	Chicago Zoological Park (Brookfield Zoo)	Chicago, Illinois	USA	16	9
99	ChicagoL	Lincoln Park Zoological Gardens	Chicago, Illinois	USA	4	1
100	Chiengma	Chiengmai Zoo	Chiengmai (Chiang Mai)	Thailand	5	0
101	Chimkent	Chimkent Zoo	Chimkent	Kazakhstan	7	4
102	Chomutov		Chomutov	Czech Republic	6	3
103	Chonbury	Khao Kheow Open Zoo	Chon Buri	Thailand	2	1
104	ChristOP	Orana Park Wildlife Trust	Christchurch	New Zealand	1	1
105	Cincinnati	Cincinnati Zoo & Botanical Garden	Cincinnati, Ohio	USA	26	10

	G		H	I	J	K	L	M		N		O		P	Q		R
	Index	Latitude	No. Of vets	Research	Staff	Size (ha)	Attendance	SpMammals	Total Sp	IndMamm	Total indiv	Management					
71	0.272727	48n09		No	100	95	360000	87	356	226	887	City					
72	0	53n33		No	20	0.6	330358	33	116	251	1603	City					
73	1	41n10	1	No	20	13	200000	24	74	92	244	City/Zool. society					
74	0	39n26	2	No		2.4	1000000	24	84	52	171	City/Zool. society					
75	0.25	51n27	2	Yes	86	5	500000	67	444	205	1039	Zool. society					
76	0.5	49n12	1	No	71	65	380000	48	196	297	1608	City					
77	0.25	25n54	3	No	72	12.5	234210	77	339	444	1494	City/Zool. society					
78	0.666667	44n26		No													
79	0.409091	47n30	1	Yes	183	12	1583943	130	445	571	3635	City					
80	0.25	34s36	1	Yes	241	18	3000000	94	363	501	2058	City					
81	0.545455	42n53	1	No	63	9	424228	72	272	612	1441	City/Zool. society					
82	0	52n28		No	4	9	41141	4	48	27	548	Charity					
83	1	51n49	1	No	30	48	332010	42	246	279	1170	Private					
84	0.666667	53n08		No													
85	0	22n32	3	Yes	235	18	2162068	64	487	291	2841	State					
86	0.083333	51n03	2	Yes	96	116	810237	84	334	371	1400	Zool. society/National					
87	0.5	3n27	1	No	28	20	10062	39	172	101	377	City					
88	0	43n22	1	No		110	400000	55	602	140	1052	Private					
89	0	42n13	1	No	33	373	262008	85	134	1118	1337	Private					
90	0	30n44		No	163	202	204238	47	238	112	658	State/National					
91	0.5	50n53	1	No	18	16	270000	36	138	129	493	Private					
92	0	50n50		No													
93	0.6	49n26	1	No	42	6	230000	47	111	117	356	City					
94	0.571429	51n21	2	No	80	14	546403	49	156	161	432	Private					
95	0.315789	53n12	3	Yes	138	45	731838	99	764	545	5141	Zool. society					
96	0.25	35n36	1	No	34	20	510761	70	395	135	639	City					
97	0.5625	41n51	5	Yes	306	81	1838954	147	445	1238	2598	Zool. society					
98	0.25	41n51	3	Yes	96	14	4500000	116	389	968	1845	City/Zool. society					
99	0	18n47	1	No	123	85	392035	72	220	324	1110	National					
100	0.571429	42n18	1	Yes	212	54	272464	68	202	190	1845	City					
101	0.5	50n28		No													
102	0.5	13n22	1	No	49	500	375092	38	205	518	4395	National					
103	1	43s32	1	No	10	24	120000	22	42	145	230	Zool. society					
104	0.384615	39n10	2	Yes	110	22	1200000	133	704	639	2266	City/Zool. society					

	A	B	C	D	E	F
	Zoo code	Official name	City	Country	Carnivora sp	Underbred sp
106	Cleveland	Cleveland Metroparks Zoological Park	Cleveland, Ohio	USA	10	4
107	Colchest	Colchester Zoo	Colchester	England	15	3
109	Cologne	Aktiengesellschaft Zoologischer Garten Köln	Cologne	Germany	14	7
110	Colombo	National Zoological Gardens	Colombo	Sri Lanka	11	7
111	Colorado	Cheyenne Mountain Zoological Park	Colorado Springs, Colorado	USA	6	3
112	Columbia	Riverbanks Zoological Park	Columbia, S Carolina	USA	5	1
113	Columbus	Columbus Zoological Gardens & A C Johnson Aquarium	Columbus, Ohio	USA	5	1
114	ColwynB	Zoological Society of Wales, The Welsh Mountain Zoo & Botanical Gardens	Colwyn Bay	Wales	7	5
115	Copenha	Copenhagen Zoo	Copenhagen	Denmark	9	2
116	Cottbus	Tierpark Cottbus	Cottbus	Germany	1	0
117	Courzien	Parc Animalier de Courzieu	Courzieu	France	1	1
118	CpTownWB	World of Birds Wildlife Sanctuary	Cape Town	South Africa	3	1
119	Curitiba	Zoológico de Curitiba	Curitiba	Brazil	5	3
120	Dallas	Dallas Zoo	Dallas, Texas	USA	2	1
121	Darjeelin	Padmaja Naidu Hailayan Zoological Park	Darjeeling	India	3	3
122	Darmstad	Vivarium Darmstadt	Darmstadt	Germany	3	1
123	Darwin	Territory Wildlife Park	Darwin	Australia	1	0
124	Debrecen	Nagyerdő Kulturpark Allat-Esnovénykertje	Debrecen	Hungary	6	3
125	Decin	Zoologicka Zahrada Decin	Decin	Czech Republic	3	0
126	Delhi	National Zoological Park	Delhi	India	2	1
127	Denver	Denver Zoological Gardens	Denver, Colorado	USA	11	5
128	Detroit	Detroit Zoological Park & Belle Isle Zoo & Aquarium	Detroit, Michigan	USA	4	0
129	Doha	Al-Wabra (Doha Zoological Gardens)+B242	Doha	Qatar	3	2
130	Dortmund	Tierpark Dortmund	Dortmund	Germany	24	10
131	DozeLaFo	Zoo de Doue	Doué la Fontaine	France	6	5
132	Dresden	Zoo Dresden	Dresden	Germany	13	3
133	Dubai	Dubai Zoo	Dubai	U Arab Emirates	6	1
134	Dubbo	Western Plains Zoo	Dubbo	Australia	3	2
135	Dublin	Zoological Society of Ireland	Dublin	Republic of Ireland	6	3
136	Dudley	Dudley and West Midlands Zoological Society	Dudley	England	6	1
137	Duisburg	Zoo Duisburg	Duisburg	Germany	11	3
138	Duluth	Lake Superior Zoological Gardens	Duluth, Minnesota	USA	3	1
139	Dushanbe	Dushanbe Zoo	Dushanbe	Tadzhikistan	6	3
140	DvurKral	Zoologicka Zahrada Dvur Kralove nad Labem	Dvur Králové	Czech Republic	16	10

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	G	H	I	J	K	L	M	N	O	P	Q	R
	Index	Latitude	No. Of vets	Research	Staff	Size (ha)	Attendance	SpMammals	Total Sp	IndMamm	Total indiv	Management
106												
107	0.4	41n30	3	No	70	50	847093	92	464	519	3125	Private
108	0.2	51n54	3	No	30	12	285000	83	194	201	717	Private
109	0.5	50n56	1	Yes	116	20	967730	98	693	637	5936	City
110	0.636364	6n56	1	Yes	199	22	980741	129	551	466	5037	National
111	0.5	38n50	1	No	60	41	341507	52	147	301	647	C/Zool. society
112	0.2	34n0	1	No	67	54	410000	47	199	192	744	City
113	0.2	39n58	1	No	86	40	725000	72	640	305	7260	City
114	0.714286	53n18	1	No	21	15	185000	30	229	110	629	Private/Zool. society
115	0.222222	55n40		No	78	11	1001696	73	615	279	2015	Private
116	0	51n45		Yes	40	25	261210	48	261	184	1013	City
117	1	45n45		No	3	20	70000	10	30	35	85	Private
118	0.333333	33s55		No	16	3.2	100000	20	375	123	3213	Private
119	0.6	25s2540	1	No	80	12	500000	51	295	303	1832	City
120	0.5	32n47	2	Yes	92	40	601809	65	401	308	1519	City/Zool. society
121	1	27n02		No								
122	0.333333	49n53	1	No	22	3.6	195229	27	122	197	925	City
123	0	12s28		No								
124	0.5	47n32		No	17	6.5	365176	73	218	120	559	City
125	0	50n48	2	No	22	7	79975	35	95	98	295	City
126	0.5	28n40	1	Yes	268	74.5	1688200	66	361	167	1557	National
127	0.454545	39n44	1	No	70	21	1031948	103	339	418	1463	City/Zool. society
128	0	42n20	1	No	107	51	800048	56	394	408	1919	City/Zool. society
129	0.666667	25n17	3	No	130	75	175019	52	122	378	723	City
130	0.416667	51n31	1	Yes	62	27	610877	87	664	282	2188	City
131	0.833333	47n13		No	14	10	228000	35	220	90	530	Private
132	0.230769	51n03	1	Yes	114	13	1073000	81	321	487	2330	City
133	0.166667	25n18		No	17	1.7						
134	0.666667	32s15	1	Yes	37	850	214000	58	98	678	813	
135	0.5	53n20	1	Yes	73	12	665803	63	242	199	864	Zool. society
136	0.166667	52n30	1	Yes	55	19	200000	63	179	198	1166	Private
137	0.272727	51n25	1	Yes	80	16	759340	145	855	573	3876	Private/Zool. society
138	0.333333	46n47		No	10	5	96000	50	138	199	573	City
139	0.5	38n35	1	Yes	116	8	913430	96	328	441	2044	City
140	0.625	50n26	2	Yes	248	68	619280	77	627	266	1426	National

	A	B	C	D	E	F
	Zoo code	Official name	City	Country	Carnivora sp	Underbred sp
141	EastLond	Queen's Park Zoological Gardens	East London	South Africa	8	4
142	Eatonvil	Northwest Trek Wildlife Park	Eatonville, Washington	USA	1	0
143	Eberswal	Tierpark Eberswalde	Eberswalde	Germany	2	0
144	Edinburg	Royal Zoological Society of Scotland	Edinburgh	Scotland	14	10
145	Edmonton	Valley Zoo	Edmonton	Canada	4	1
146	Eichberg	Zoologische Station Eichberg	Eichberg	Switzerland	2	1
147	Ekaterin	Ekaterinburg Zoo	Ekaterinburg	Russia	14	5
148	El Paso	El Paso Zoo	El Paso, Texas	USA	1	1
149	Emmen	Noorder Dierenpark/Zoo	Emmen	Netherlands	7	6
150	Erfurt	Thuringer Zoopark Erfurt	Erfurt	Germany	7	2
151	Erie	Erie Zoological Gardens	Erie, Pennsylvania	USA	1	1
152	Eskilstu	Parken Zoo Ab	Eskilstuna	Sweden	6	2
153	Evansvil	Mesker Park Zoo & Botanic Garden	Evansville, Indiana	USA	2	1
154	F. Worth	Fort Worth Zoological Park & JR Record Aquarium	Fort Worth, Texas	USA	3	2
155	FortWayn	Fort Wayne Children's Zoo	Fort Wayne, Indiana	USA	4	3
156	Fota	Fota Wildlife Park	Fota Island, Carringtwhill	Republic of Ireland	3	1
157	Frankfur	Zoologischer Garten der Stadt Frankfurt am Main	Frankfurt am Main	Germany	18	8
158	Fréjus	Parc Zoologique de Fréjus	Fréjus	France	2	0
159	Fresno	Chaffee Zoological Gardens of Fresno	Fresno, California	USA	5	0
160	FrontRoy	Conservation and Research Center	Front Royal, Virginia	USA	2	0
161	Froson	Froso-Zoo	Frösön-Östermund	Sweden	2	0
162	Fujisawa	Enoshima Aquarium	Fujisawa	Japan	1	0
163	Fukuoka	Fukuoka Municipal Zoo	Fukuoka	Japan	6	5
164	Gainesvi	Santa Fé Community College Teaching Zoo	Gainesville, Florida	USA	1	0
165	GardenCt	Lee Richardson Zoo	Garden City, Kansas	USA	2	1
166	Gauhati	Assam State Zoo and Botanical Garden	Gauhati	India	6	2
167	Gdansk	Municipal Zoological Garden of the Sea Coast	Gdansk	Poland	6	5
168	Gelsenki	Ruhr Zoo Betriebsgesellschaft	Gelsenkirchen	Germany	7	5
169	Ghent	Zoological Park Gent	Gent	Belgium	4	1
170	Give	Givskud Zoo	Give	Denmark	1	0
171	Glasgow	Glasgow Zoo	Glasgow	Scotland	3	1
172	Goldau	Natur und Tierpark Goldau	Goldau	Switzerland	2	0
173	Gorlitz	Naturschutz-Tierpark Gorlitz	Gorlitz	Germany	5	3
174	Gossau	Walter Zoo	Gossau	Switzerland	2	1

	G	H	I	J	K	L	M	N	O	P	Q	R
	Index	Latitude	No. Of vets	Research	Staff	Size (ha)	Attendance	SpMammals	Total Sp	IndMamm	Total Indiv	Management
141												
142	0.5	33s0		No	19	4	76654	56	193	412	1363	City
143	0	46n52	1	No	16	270	136000	22	34	171	471	City
144	0	52n50		No								
145	0.714286	55n57	1	Yes	94	32	446030	64	319	175	1371	Zool. society/City
146	0.25	53n33	1	No	17	31	231000	39	122	107	360	City/National
147	0.5	47n21		No								
148	0.357143	56n51		No								
149	1	31n46	2	No	23	2	249175	26	116	76	382	City/Zool. society
150	0.857143	52n47	1	Yes	60	12	1000000	64	369	783	5534	City/Private
151	0.285714	50n58	1	Yes	71	25	400000	119	773	206	1080	City
152	1	42n08	1	No	15	6	225000	44	69	168	268	City/Zool society
153	0.333333	59n22	1	No	10	11	343000	24	70	116	404	City
154	0.5	37n58	3	No	26	27	140000	52	122	150	419	City/Zool society
155	0.666667	32n44	1	No	58	16	717079	60	781	291	3700	City/Zool. society
156	0.75	41n08	1	No	10	15	403973	37	92	170	450	City/Zool. society
157	0.333333	51n54	1	No	8	28	174000	24	199	68	449	Zool. society
158	0.444444	50n07	1	No	157	11	2330884	122	1188	617	5649	City
159	0	43n26	1	No	12	15	140000	25	150	90	430	Private
160	0	36n45	2	No	25	6	457514	51	169	124	450	City
161	0	38n55	4	Yes	320	1274	3300000	132	469	1345	4173	National
162	0	63n11	1	No	35	26	132000	45	234	142	979	Private
163	0	35n21	1	No	38	0.7	583082	13	52	355	10899	Private
164	0.833333	35n34	2	No	38	10.3	1197803	65	324	203	1068	City
165	0	29n39	1	No	5	5.6	15000	13	86	42	253	State
166	0.5	37n58	1	No	16	19	230000	35	106	159	660	City
167	0.333333	26n11		No	123	130	606326	58	466	134	1101	State
168	0.833333	54n23	1	Yes	98	112	451766	76	176	290	784	City
169	0.714286	51n31		Yes	52	22	268000	110	516	222	1061	Private
170	0.25	51n03		No								
171	0	55n51	1	No	16	90	250000	31	365	112	736	
172	0.333333	55n53		No	24	20	150000	34	117	101	297	Zool. society/City
173	0	47n03		No	8	26	156000	22	86	199	556	Zool. society
174	0.6	51n09		No								
175	0.5	47n25	1	No	12	5	112000	39	100	144	382	Private

	A	B	C	D	E	F
	Zoo code	Official name	City	Country	Carnivora sp	Underbred sp
176	Granby	Granby Zoo	Granby	Canada	14	10
177	GrandRa	John Ball Zoological Gardens	Grand Rapids, Michigan	USA	6	3
179	Greenville	Greenville Zoo	Greenville, South Carolina	USA	3	2
180	Grodno	Grodno Zoo	Grodno	Belorussia	9	3
181	Gt.Witch	Norfolk Wildlife Centre and Country Park	Great Witchingham	England	7	2
182	Gt.Yarmo	Thrigby Hall Wildlife Gardens	Great Yarmouth	England	6	3
183	Guernsey	Zoological Trust of Guernsey	Guernsey	Channel Islands-UK	1	0
184	Haifa	Haifa Educational Zoo (of the Biological Institute)	Haifa	Israel	14	5
185	HalfwayH	Transvaal Snake Park	Halfway House	South Africa	2	1
186	Halle	Zoologischer Garten Halle	Halle	Germany	21	8
187	Hamamats	Hamamatsu Municipal Zoo	Hamamatsu	Japan	2	1
188	Hamburg	Carl Hagenbeck Tierpark	Hamburg	Germany	7	0
189	Hamilton	Hamilton Zoological Gardens	Hamilton	New Zealand	1	1
190	Hanoi	Hanoi Zoological Gardens	Hanoi	Vietnam	3	0
191	Hanover	Zoo Hannover	Hanover	Germany	5	1
192	Hastings	Ridgeway Trust for Endangered Cats	Hastings	England	3	1
193	Havana	Havana Zoological Garden	Havana	Cuba	5	1
194	Hayle	Paradise Park	Hayle	England	2	1
195	Healesvi	Healesville Sanctuary	Healesville	Australia	1	0
196	Heidelbe	Tiergarten Heidelberg	Heidelberg	Germany	12	0
197	Helsinki	Helsinki Zoo	Helsinki	Finland	11	0
198	Hershey	Zooamerica North American Wildlife Park	Hershey, Pennsylvania	USA	3	2
199	Higashi	Atagawa Tropical & Alligator Garden	Higashi-Izu	Japan	1	0
200	Higashim	Saitama Children's Zoo (?)	Higashi-Matsuyama	Japan	1	0
201	Hilvaren	Safaripark Beekse Bergen	Hilvarenbeek	Netherlands	6	3
202	Hiroshim	Asa Zoological Park	Hiroshima	Japan	8	1
203	Hodenhag	Serengeti Safaripark Hodenhagen	Hodenhagen	Germany	4	0
204	Hoedspru	Hoedspruit Research and Breeding Centre for Endangered Species	Hoedspruit	South Africa	2	0
205	Hof	Zoologischer Garter Hof, Naturkunde am Theresienstein	Hof/Saale	Germany	3	1
206	Holyrood	Salmoner Nature Park	Holyrood	Canada	3	2
207	HongKong	Hong Kong Zoological and Botanical Gardens	Hong Kong	China	2	2
208	Honolulu	Honolulu Zoo	Honolulu, Hawaii	USA	3	2
209	Hoor	Stiftelsen Skanes Djurpark (Scania Zoo Foundation)	Höör	Sweden	5	1
210	Houston	Houston Zoological Gardens	Houston, Texas	USA	11	3

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	G	H	I	J	K	L	M	N	O	P	Q	R
	Index	Latitude	No. Of vets	Research	Staff	Size (ha)	Attendance	SpMammals	Total Sp	IndMamm	Total indiv	Management
176	0.714286	45n24	1	No	28	18	328824	60	196	201	720	Zool. society
177	0.5	42n58		No	15	5.6	340500	41	165	154	555	City
178	0.666667	34n51	1	No	12	5.6	123000	15	106	34	361	City
180	0.333333	53n41	1	No	80	8.2	456935	83	306	1425	3108	City
181	0.285714	52n38		No	9	16	57793	28	192	92	657	Charity
182	0.5	52n37		No	4	4	58766	20	47	73	165	Private
183	0	49n30	1	No	4	1.2	58000	20	105	31	173	Charity
184	0.357143	32n49	1	No	40	3	200000	41	202	164	701	Zool. society/City/State
185	0.5	25s45	1	No	8	0.8	150000	1	107	4	392	Private
186	0.380952	51n29	1	Yes	85	8.5	509214	103	360	383	1803	City
187	0.5	34n42	3	No	35	14	654576	58	202	124	476	City
188	0	53n33	1	No	90	27	800000	67	471	342	1752	Private
189	1	37s47		No								
190	0	21n02		No								
191	0.2	52n24	2	Yes	106	21	786323	109	542	238	1205	City/Zool. society
192	0.333333	50n51		No								
193	0.2	23n08		No	146	23	2364000	83	562	228	3042	National
194	0.5	50n12		No	15	7	100000	8	60	133	560	Private
195	0	37s40	1	Yes	47	175	280696	48	175	501	1477	State
196	0	49n25		No	35	11	313776	84	412	319	1260	City/Private
197	0	60n10	2	Yes	83	22	343216	61	541	166	1014	City
198	0.666667	40n17	1	No	10	4.5	514111	21	65	89	212	Private
199	0	34n48		No	60	3.3	600000	4	26	69	487	Private
200	0	36n02	4	No	43	80	926423	20	297	88	795	City
201	0.5	51n29	1	Yes	20	135	340000	63	125	395	871	Private
202	0.125	34n24	2	No	63	23	547001	51	459	150	1093	City
203	0	52n46		No	15	130	250000	34	370	42	405	Private
204	0	24s20		No	29	6000	9200					National
205	0.333333	50n18	1	No	6	1	40000	18	90	107	823	
206	0.666667	47n26	3	No	8	40	35000	15	42	57	188	State
207	1	22n17	1	Yes	51	5.4		18	77	333	1223	City
208	0.666667	21n18		No	59	17	775095	46	260	168	875	City
209	0.2	55n56		No								
210	0.272727	29n46	3	Yes	110	20	2021853	106	627	445	3066	City

	A	B	C	D	E	F
	Zoo code	Official name	City	Country	Carnivora sp	Underbred sp
211	Hoyersw	Zoo Hoyerswerda	Hoyerswerda	Germany	7	1
212	Hunnesbo	Nordic Ark	Hunnebostrand	Sweden	5	1
213	Huntingd	Hamerton Wildlife Park	Huntingdon	England	3	1
214	Hyderabad	Nehru Zoological Park	Hyderabad	India	6	4
215	Indianap	Indianapolis Zoological Park	Indianapolis, Indiana	USA	1	0
216	Innsbruc	Alpenzoo	Innsbruck	Austria	7	1
217	Jackson	Jackson Zoological Park	Jackson, Mississippi	USA	4	0
218	Jacksonv	Jacksonville Zoological Gardens	Jacksonville, Florida	USA	2	1
219	Jaipur	Zoological Garden Jaipur	Jaipur	India	1	1
220	Jakarta	Ragunan Zoo	Jakarta	Indonesia	4	0
221	Jalisco	Zoológico Guadalajara	Jalisco	Mexico	8	6
222	Jerez	Zoo Jerez	Jerez de la Frontera	Spain	4	3
223	Jersey	Jersey Wildlife Preservation Trust	Jersey	Channel Islands-UK	2	1
224	Jerusale	Tisch Family Zoological Gardens	Jerusalem	Israel	6	1
225	Jihlava	Zoologicka Zahrada Jihlava	Jihlava	Czech Republic	10	0
226	Johannes	Johannesburg Zoological Gardens	Johannesburg	South Africa	14	6
227	Junagadh	Sakkarbaug Zoo Junagadh	Junagadh	India	5	2
228	Kagoshim	Hirakawa Zoological Park	Kagoshima	Japan	9	8
229	Kalining	Kaliningrad Zoo	Kaliningrad	Russia	12	6
230	Kamloops	Kamloops Wildlife Parks	Kamloops	Canada	2	1
231	Kamogawa	Kamogawa Sea World	Kamogawa	Japan	1	0
232	Kanpur	Kanpur Zoological Park	Kanpur	India	3	1
233	Kansas	Kansas City Zoological Gardens	Kansas City, Missouri	USA	6	2
234	Karagand	Karaganda Zoo	Karaganda	Kazakhstan	6	4
235	Karlsruh	Zoologischer Garten Karlsruhe	Karlsruhe	Germany	9	3
236	Katowice	Slaski Ogród Zoologiczny	Katowice	Poland	5	2
237	Kaunas	Lietuvos Zoologijos Sodas	Kaunas	Lithuania	20	9
238	KawasaAq	Yomiuri Land Marine Aquarium	Kawasaki	Japan	1	0
239	Kawasaki	Yumemigasaki Zoological Park	Kawasaki	Japan	2	0
240	Kazan	Kazan Zoo Botsad	Kazan	Russia	8	5
241	Kessingl	Suffolk Wildlife Park	Kessingland	England	3	3
242	Kharkov	Kharkov Zoo	Kharkov	Ukraine	14	7
243	Kiev	Kiev Zoo	Kiev	Ukraine	12	9
244	Kincraig	Royal Zoological Society of Scotland-Highland Wildlife Park	Kincraig	Scotland	7	1

	A	B	C	D	E	F
	Zoo code	Official name	City	Country	Carnivora sp	Underbred sp
246	Kishinev	Kishinev Zoo	Kishinev	Moldavia	8	2
247	Kitakyus	Green Park/Itozu Zoo Park	Kitakyushu	Japan	3	1
248	Knoxvill	Knoxville Zoological Gardens	Knoxville, Tennessee	USA	6	3
249	Kobe	Kobe Oji Zoo	Kobe	Japan	10	5
250	Kobe Aq	Suma Aqualife Park	Kobe	Japan	1	0
251	Kosice	Zoologicka Zahrada Kosice	Kosice	Slovakia	3	1
252	Kraaifont	Tygerberg Zoopark	Kraaifontein	South Africa	6	4
253	Krakow	Local Park and Zoological Gardens Foundation	Kraków	Poland	18	10
254	Krefeld	Krefelder Zoo	Krefeld	Germany	14	7
255	Kronberg	Opel-Zoo	Kronberg	Germany	4	3
256	KualaLum	Zoo Negara Malaysia	Kuala Lumpur	Malaysia	10	7
257	Kumamoto	Kumamoto Zoological and Botanical Garden	Kumamoto	Japan	6	6
258	Kushiro	Kushiro Zoo	Kushiro	Japan	3	0
259	Kuwait	Kuwait Zoo	Kuwait City	Kuwait	5	5
260	Kyoto	Kyoto Municipal Zoo	Kyoto	Japan	8	4
261	La Chaux	Parc d'Acclimatation du Bois du Petit Chateau	La Chaux-de-Fonds	Switzerland	3	1
262	La Fleche	Zoo de la Fleche	La Flèche	France	3	0
263	La Plata	Zoológico La Plata	La Plata	Argentina	3	3
264	LakeArie	Claws'n'Paws Wild Animal Park	Lake Ariel, Pennsylvania	USA	6	4
265	LakeMonr	Central Florida Zoological Park	Lake Monroe, Florida	USA	2	0
266	Landau	Zoo Landau	Landau	Germany	1	0
267	Le Vaud	La Garenne	Le Vaud	Switzerland	4	3
268	Legal	Alberta Wildlife Park	Legal	Canada	2	0
269	Leipzig	Zoologischer Garten Leipzig	Leipzig	Germany	15	10
270	Liberec	Zoologicka Zahrada Liberec	Liberec	Czech Republic	5	3
271	Lignano	Parco Zoo 'Punta Verde'	Lignano Sabbiadoro	Italy	6	0
272	Lima	Patronato Nacional del Parque de Las Leyendas	Lima	Peru	6	6
273	Lincoln	Folsom Children's Zoo and Botanical Gardens	Lincoln, Nebraska	USA	1	0
274	Lipetsk	Lipetsk Zoo	Lipetsk	Russia	8	1
275	Lisbon	Jardim Zoológico de Lisboa	Lisbon	Portugal	10	7
276	Lisieux	Cerza Lisieux	Lisieux	France	2	2
277	Litchfie	Wildlife World Zoo	Litchfield Park, Arizona	USA	6	2
278	LittleRo	Little Rock Zoo	Little Rock, Arkansas	USA	11	4
279	Ljubljana	Zooloski vrt Mesta Ljubljane	Ljubljana	Slovenia	10	3

	A	B	C	D	E	F
	Zoo code	Official name	City	Country	Carnivora sp	Underbred sp
281	Lodz	Miejski Ogród Zoologiczny w Lodzi	Lódz	Poland	13	7
282	LondonZS	Zoological Society of London	London	England	9	3
283	LongLeat	Longleaf Safari Park (Lions of Longleaf)	Warminster	England	1	0
284	LosAngel	Los Angeles Zoo	Los Angeles, California	USA	11	6
285	Louisvil	Louisville Zoological Garden	Louisville, Kentucky	USA	5	1
286	Lufkin	Ellen Trout Park Zoo	Lufkin, Texas	USA	3	2
287	Lycsele	Lycsele Djurpark/Zoo	Lycsele	Sweden	7	3
288	Lympne	Port Lympne Wild Animal Park	Lympne	England	13	3
289	Lyons	Jardin Zoologique de la Ville de Lyon	Lyon	France	4	2
290	Mabletho	Animal Gardens	Mablethorpe	England	1	0
291	Madison	Henry Vilas Zoo	Madison, Wisconsin	USA	3	2
292	Madras	Arignar Anna Zoological Park	Madras	India	6	3
293	Madrid	Zoo-Aquarium de la Casa de Campo	Madrid	Spain	10	5
294	Magdebur	Zoologischer Garten Magdeburg	Magdeburg	Germany	11	3
295	Malton	Flamingo Land	Malton	England	2	0
296	Manhatta	Sunset Zoological Park	Manhattan, Kansas	USA	3	1
297	Marwell	Marwell Zoological Park	Marwell, Winchester	England	13	7
298	Matlock	Riber Castle Wildlife Park	Matlock	England	6	3
299	Matsushi	Matsushima Aquarium	Matsushima	Japan	1	1
300	Mayaguez	Zoorico P R. Zoological Gardens	Mayaguez	Puerto Rico	4	1
301	Mechelen	Dierenpark Planckendael	Mechelen	Belgium	7	3
302	Medellin	Parque Zoológico Santa Fé	Medellin	Colombia	6	4
303	Melaka	Zoo Melaka	Melaka	Malaysia	2	0
304	Melbourn	Royal Melbourne Zoological Gardens	Melbourne	Australia	17	9
305	Memphis	Memphis Zoo and Aquarium	Memphis, Tennessee	USA	3	1
306	MexicoSJ	San Juan de Aragon Zoological Park	Mexico City	Mexico	5	3
307	MiamiMZ	Miami Metrozoo	Miami, Florida	USA	8	1
308	Mihama	Minamichita Beach Land Aquarium	Mihama-cho	Japan	1	1
309	Milwauk	Milwaukee County Zoo	Milwaukee, Wisconsin	USA	9	2
310	Minneapo	Minnesota Zoological Garden	Minneapolis/St Paul, Minnesota	USA	10	4
311	Misaki	Misaki Park Zoo and Aquarium	Misaki	Japan	2	2
312	MiyajjAq	Miyajima Public Aquarium	Miyajima	Japan	1	1
313	Miyajjima		Miyajima	Japan	1	1
314	Miyazaki	Phoenix Zoo	Miyazaki	Japan	3	0

	G		H		I	J	K		L		M		N		O		P		Q		R		
	Index	Latitude	Latitude	Size (ha)			No. Of vets	Research	Staff	Size (ha)	Attendance	SpMammals	Total Sp	IndMamm	Total indiv	Management							
281																							
282	0.538462	51n46	51n46	16.5	4	Yes	100	425770	99	344	644	2944	City										
283	0.333333	51n30	51n30	15	5	Yes	368	1225000	142	1255	552	2794	Zool. society										
284	0	51n13	51n13	81		No	30	400000	40	233	49	278	Private										
285	0.545455	34n03	34n03	32	3	Yes	150	1631065	136	518	846	2083	City/Zool society										
286	0.2	38n15	38n15	30	2	No	72	400000	50	213	191	639	City										
287	0.666667	31n20	31n20	6.4	3	Yes	10	160000	30	182	80	537	City										
288	0.428571	64n36	64n36	40	2	No	6	125000	14	31	141	370	City										
289	0.230769	51n05	51n05	160	2	No	67	120000	47	342	48	343	Charity										
290	0.5	45n45	45n45	6		No	18		39	183	131	759	City										
291	0	53n21	53n21	0.8	1	No	4	35000	22	78	83	338	Private										
292	0.666667	43n04	43n04	8	3	No	15	750000	49	190	223	802	City										
293	0.5	13n05	13n05	510	1	Yes	108	768278	47	298	123	776	State										
294	0.5	40n24	40n24	20	3	Yes	95	944478	130	229	561	1641	Private										
295	0.272727	52n07	52n07	20	2	No	65	483965	68	321	195	951	City										
296	0	54n08	54n08	40	2	No	50	800000	41	235	155	1011	Private										
297	0.333333	39n11	39n11			No																	
298	0.538462	51n04	51n04	40	1	No	41	234808	70	469	125	661	Charity										
299	0.5	53n08	53n08	16		No	7	150000	20	100	47	300	Private										
300	1	38n22	38n22	0.7		No	30	800000	7	32	334	11312	Private										
301	0.25	18n12	18n12	12	1	No	76	242580	39	102	128	518	National										
302	0.428571	51n02	51n02	40	2	Yes	39	271288	53	135	287	687	Zool. society/National										
303	0.666667	6n15	6n15	4	1	No	61	650000	61	286	239	1181	City										
304	0	2n12	2n12			No																	
305	0.529412	37s49	37s49	30	1	Yes	180	1114261	109	382	689	3392	State										
306	0.333333	35n09	35n09	14.5	1	Yes	65	541185	73	399	284	1904	City										
307	0.6	19n24	19n24	37	3	Yes	50	1200000	55	115	504	1451	City										
308	0.125	25n46	25n46	283	2	Yes	136	814452	76	262	317	1120	City/Zool society										
309	1	34n46	34n46			No																	
310	0.222222	43n03	43n03	72	3	Yes	104	1654818	118	414	325	2084	City/Zool society										
311	0.4	44n59	44n59	195	3	Yes	150	883960	70	180	428	880	State/Zool. society										
312	1	35n08	35n08	10	1	No	61	629223	22	204	170	1489	Private										
313	1	34n18	34n18			No																	
314	1	34n18	34n18	0.8		No	18	710572	6	338	25	12873	City										
315	0	35n56	35n56	13	1	No	44						Private										

	A	B	C	D	E	F
	Zoo code	Official name	City	Country	Carnivora sp	Underbred sp
316	Monroe	Louisiana Purchase Gardens and Zoo	Monroe, Louisiana	USA	3	1
317	MontePPR	Jardin Zoológico Parque Dolores Pereira de Rossell y Parque Lecoq	Montevideo	Uruguay	3	3
318	Montgome	Montgomery Zoo	Montgomery, Alabama	USA	5	3
319	Montpell	Parc Zoologique de Lunaret	Montpellier	France	7	3
320	Montreal	Biodome de Montreal	Montreal	Canada	1	0
321	Morelia	Parque Zoológico 'Benito Juárez'	Morelia	Mexico	4	2
322	Moscow	Moscow Zoo	Moscow	Russia	14	3
323	Mulhouse	Parc Zoologique et Botanique de la Ville de Mulhouse	Mulhouse	France	14	8
324	Munich	Munchener Tierpark Hellbrunn AG	Munich	Germany	16	3
325	Munster	Westfälischer Zoologischer Garten Munster	Munster	Germany	12	6
326	Murrells	Brookgreen Gardens	Murrells Inlet, S Carolina	USA	2	1
327	Mysore	Sri Chamarajendra Zoological Gardens	Mysore	India	2	1
328	Nagano	Nagano Municipal Chausuyama & Jogyama Branch Zoo	Nagano	Japan	1	0
329	NagasaAq	Nagasaki Aquarium	Nagasaki	Japan	2	1
330	Nagoya	Nagoya Higashiyama Zoo	Nagoya	Japan	17	13
331	Nairobi	Institute of Primate Research (?)	Nairobi	Kenya	1	0
332	Nalchik	Nalchik Zoo	Nalchik	Russia	6	3
333	Nanyuki	Mount Kenya Game Ranch	Nanyuki	Kenya	2	0
334	Naples	Giardino Zoologico di Napoli	Naples	Italy	7	0
335	Neath	Pencynor Wildlife Park	Neath	Wales	3	1
336	Neumuns	Tierpark Neumunster	Neumunster	Germany	7	6
337	Neuwied	Zoo Neuwied	Neuwied	Germany	9	3
338	NewOrlea	Audubon Park and Zoological Garden	New Orleans, Louisiana	USA	5	3
339	Nihonmat	Tohoku Safari Park	Nihommatsu	Japan	4	3
340	Nikolaev	Nikolaev Zoo	Nikolajev	Ukraine	21	16
341	Noichi	Noichi Zoological Park of Koichi Prefecture	Noichi	Japan	1	0
342	Nordhorn	Tierpark Nordhorn	Nordhorn	Germany	1	1
343	Norrkopi	Kolmardens Djurpark	Norrköping	Sweden	4	1
344	Novosibi	Novosibirsk Zoo	Novosibirsk	Russia	26	4
345	Numazu	Izo-Mito Sea Paradise	Numazu	Japan	1	0
346	Nurembe	Tiergarten der Stadt Nurnberg	Nürnberg	Germany	10	6
347	NYBronx	Bronx Zoo & Wildlife Conservation Park	New York, NY	USA	10	1
348	NYCNP	Central Park Wildlife Center	New York, NY	USA	1	0
349	NYStaten	Staten Island Zoo	New York, NY	USA	1	1

	G	H	I	J	K	L	M	N	O	P	Q	R
	Index	Latitude	No. Of vets	Research	Staff	Size (ha)	Attendance	SpMammals	Total Sp	IndMamm	Total indiv	Management
316												
317	0.333333	32n31		No	24	32	100000	64	245	320	898	City
318	1	34s53		No								
319	0.6	32n22	1	No	15	2.4	148281	29	141	144	563	City/Zool. society
320	0.428571	43n36	3	No	37	80	300000	48	263	132	613	City
321	0	45n31	2	No	25	1	319322	21	109	92	373	City
322	0.5	19n42	2	Yes	101	23	1133402	94	259	340	2463	State
323	0.214286	55n45	3	Yes	333	17	3508870	119	825	513	5282	City
324	0.571429	47n45		No	47	25	279512	81	454	233	1251	City
325	0.1875	48n08	1	No	104	36	1282236	112	831	377	2480	Private/City
326	0.5	51n57	1	Yes	82	30	812050	64	362	350	1949	Private/City/Zool. society
327	0.5	33n33	1	No	7	13	176000	5	13	68	104	Private
328	0.5	12n18		No	128	100	1810000	60	322	146	885	State
329	0	36n39	1	No	34	17	299061	36	107	265	561	City
330	0.5	32n48		No								Private
331	0.764706	35n10	10	No	50	30	3200000	117	470	449	3144	City
332	0	1s17	2	No	120	200	0	8	8	480	480	National
333	0.5	43n29	1	No	30	7.3	210000	61	115	146	278	City
334	0	0n01	1	No	41	490	26400	29	31	910	1020	Private
335	0	40n51	2	No	27	331	650000	83	321	274	1344	Zool. society/State/City
336	0.333333	51n40	2	No	16	6	290000	34	164	179	5762	Private
337	0.857143	54n04	2	No	25	24	113676	59	237	170	657	Zool. society/City/State
338	0.333333	50n25	1	No	10	12	100000	46	314	101	670	City/State
339	0.6	29n57	1	Yes	189	22	915500	83	372	356	1425	City
340	0.75	37n35		No								
341	0.761905	46n58	1	Yes	142	23	499650	96	335	304	2147	City
342	0	33n33		No								
343	1	52n27		No								
344	0.25	58n36	1	Yes	85	270	1300000	47	65	627	786	Private
345	0.153846	55n02	1	Yes	102	0.8	1571000	101	292	327	1010	City
346	0	35n06	1	Yes	105	1.6	871930	17	242	125	5606	Private
347	0.6	49n27	1	No	110	63	760188	91	687	345	2024	City/Zool. society
348	0.1	40n43	2	Yes	470	106	2077884	139	648	2056	5162	Zool. society
349	0	40n43		No								
350	1	40n43	1	No	32	3.6	279326	33	274	107	577	Private/Zool. society

	A	B	C	D	E	F
	Zoo code	Official name	City	Country	Carnivora sp	Underbred sp
351	Obihiro	Obihiro Zoo	Obihiro	Japan	3	0
352	Obregon	Parque de la Naturaleza de Cabarceno	Obregon	Spain	5	3
353	Obterre	Parc de la Haute-Touche	Obterre	France	2	1
354	Odense	Odense Zoo	Odense	Denmark	7	4
355	Oita	Marine Palace	Oita	Japan	1	0
356	Oklahoma	Oklahoma City Zoological Park	Oklahoma City, Oklahoma	USA	8	5
357	OklahoOC	Oakhill Center for Rare and Endangered Species	Oklahoma City, Oklahoma	USA	5	2
358	Olomouc	Zoologicka Zahrada Olomouc	Olomouc	Czech Republic	16	7
359	Omaha	Omaha's Henry Doorly Zoological Gardens	Omaha, Nebraska	USA	4	1
360	Opole	Ogrod Zoologiczny w Opole	Opole	Poland	9	6
361	Orlando	Sea World of Florida	Orlando, Florida	USA	1	0
362	Osaka	Osaka Municipal Tennoji Zoological Garden	Osaka	Japan	10	6
363	Osnabruc	Zoo Osnabruck	Osnabruck	Germany	9	3
364	Ostrava	Zoologicka Zahrada Ostrava	Ostrava	Czech Republic	19	11
365	Ozoir	Parc Zoologique du Bois d'Atilly	Ozoir-la-Ferrière	France	1	0
366	Paignton	Paignton Zoological & Botanical Gardens	Paignton	England	3	1
367	PalmDes	Living Desert	Palm Desert, California	USA	7	2
368	Paramus	Bergen County Zoological Park	Paramus, New Jersey	USA	2	0
369	ParisMen	Menagerie du Jardin des Plantes	Paris	France	4	3
370	ParisPZ	Parc Zoologique de Paris	Paris	France	3	2
371	Pastreng	Parco Natura Viva	Pastrengo	Italy	7	4
372	Peaugres	Safari de Peaugres	Peaugres	France	6	2
373	Pecs	Mecseki Kulturpark Zoo	Pecs	Hungary	6	4
374	Penza	Penza Zoo	Penza	Russia	14	9
375	Perm	Perm Zoo	Perm	Russia	15	11
376	Perth	Perth Zoological Gardens	Perth	Australia	5	3
377	Peterbor	Riverview Park and Zoo	Peterborough	Canada	1	1
378	Philadel	Philadelphia Zoological Garden	Philadelphia, Pennsylvania	USA	5	1
379	Phoenix	Phoenix Zoo	Phoenix, Arizona	USA	7	3
380	Pilsen	Zoologicka a Botanicka Zahrada Mesta Plzne	Pilsen	Czech Republic	12	5
381	Piriapol	Estacion de la Fauna Autoctona	Piriapolis	Uruguay	2	0
382	Pistoia	Giardino Zoologico di Pistoia	Pistoia	Italy	7	3
383	Pittsbur	Pittsburgh Zoo	Pittsburgh, Pennsylvania	USA	2	0
384	Pl.duTou	African Safari	Plaisance du Touch	France	3	2

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	Index	Latitude	No. Of vets	Research	Staff	Size (ha)	Attendance	SpMammals	Total Sp	IndMamm	Total indiv	Management
351												
352	0	42n55	2	No	16	7	214082	47	100	303	793	City
353	0.6	43n21		No								
354	0.5	47n13		No								
355	0.571429	55n24		No	15	15	175000	34	118	76	233	Private
356	0	33n14		No	58	05						Private
357	0.625	35n28	1	Yes	119	56	665020	84	456	328	1887	City
358	0.4	35n28		No								
359	0.4375	49n36	1	Yes	47	44	277199	68	376	191	1246	National
360	0.25	41n16	2	No	47	45	590000	59	197	380	872	Zool. society
361	0.666667	50n41	1	No	43	18	117000	38	145	143	1112	National
362	0	28n32	3	No	700	40	4350000	14	402	140	9633	Private
363	0.6	34n40	5	No	87	107	1138800	110	326	441	1110	City
364	0.333333	52n16	2	No	36	21	320000	73	309	300	1842	Zool. society/City
365	0.578947	49n50	2	Yes	101	104	408245	72	363	182	1183	City
366	0	48n46	1	No	6	15	100000	50	250	210	730	Private
367	0.333333	50n26	1	Yes	70	40	354699	65	253	335	1341	Charity
368	0.285714	33n43	1	No	17	480	120000	24	108	73	744	Private
369	0	40n57	1	No	16	2	430000	21	61	73	189	City
370	0.75	48n52	1	Yes	66	6.5	413325	73	310	263	1068	National
371	0.666667	48n52	1	Yes	129	17	878107	86	433	165	948	National
372	0.571429	45n29		No								Private
373	0.333333	45n19		No	17	70	215000					Private
374	0.666667	46n05		Yes	68	6.5	397000	93		234		City
375	0.642857	53n13	1	No	43	10.5	164440	38	98	97	315	City
376	0.733333	58n0	1	No	74	2.2	597130	53	152	133	544	City
377	0.6	31s57	1	Yes	69	18	420480	102	329	512	1688	State
378	1	44n18		No	4	21	277000	14	41	43	311	City
379	0.2	39n57	2	Yes	155	15	1323530	112	459	439	1509	Private/Zool. society
380	0.428571	33n27	2	No	125	50	880007	72	293	438	2252	Zool. society
381	0.416667	49n45	1	Yes	47	65	131000	43	173	169	604	City
382	0	34s54	3	No	10	86	70000	12	148	58	551	City
383	0.428571	43n55	2	No	23	7	346000	59	221	149	604	Private
384	0	40n26	1	No	50	30	450000	38	299	154	3450	City
385	0.666667	43n36	2	No	8	5.5	90000	28	71	56	188	Private

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A		B		C		D		E		F	
Zoo code		Official name		City		Country		Carnivora sp		Underbred sp	
386	Plock	Miejski Ogród Zoologiczny		Plock		Poland		4		2	
387	PlymWLP	Dartmoor Wildlife Park & West Country Falconry Centre		Plymouth		England		4		1	
388	PortEiz	Port Elizabeth Museum, Oceanarium, Snake Park & Tropical House		Port Elizabeth		South Africa		1		1	
390	Portland	Metro Washington Park Zoo		Portland, Oregon		USA		3		0	
391	PortofSp	Emperor Valley Zoo		Port-of-Spain		Trinidad		3		1	
392	Potgiest	Potgietersrus Breeding Centre		Potgietersrus		South Africa		4		0	
393	Poznan	Ogród Zoologiczny w Poznaniu		Poznan		Poland		18		9	
394	Prague	Zoologicka Zahrada Praha		Prague		Czech Republic		15		8	
395	Prescot	Knowsley Safari Park		Prescot		England		1		1	
396	Pretoria	National Zoological Gardens of South Africa		Pretoria		South Africa		16		6	
397	Providen	Roger Williams Park Zoo		Providence, Rhode Island		USA		4		2	
398	PuertoDLC	Loro Parque		Puerto de la Cruz		Spain		1		0	
399	Pune	Peshawe Park Zoological Garden		Pune		India		2		1	
400	PuntoFij	Gustavo Rivera Zoo		Punto Fijo		Venezuela		4		1	
401	Quebec	Jardin Zoologique du Quebec		Quebec		Canada		5		3	
402	Rabat	Parc Zoologique National de Rabat		Rabat		Morocco		10		3	
403	RamatGa	Zoological Center Tel Aviv Ramat-Gan Ltd		Ramat-Gan		Israel		15		9	
404	Ranua	Ranua Zoo		Ranua		Finland		9		3	
405	Rheine	Tierpark Rheine		Rheine		Germany		1		1	
406	Rhenen	Ouwehands Dierenpark		Rhenen		Netherlands		15		7	
407	Riga	Rigas Zoologiskais Darzs		Riga		Latvia		14		8	
408	Rio	Fundação Jardim Zoológico da Cidade do Rio de Janeiro-RIOZOO		Rio de Janeiro, RJ		Brazil		9		9	
409	Roanoke	Mill Mountain Zoo		Roanoke, Virginia		USA		2		1	
410	Rocheste	Seneca Park Zoo		Rochester, New York		USA		5		2	
411	Romanech	Touroparc		Romaneche-Thorins		France		7		3	
412	Rome	Giardino Zoologico I Museo di Zoologia del Comune di Roma		Roma		Italy		11		8	
413	Rostock	Zoologischer Garten Rostock		Rostock		Germany		9		8	
414	Rostov	Rostov-on-Don Zoo		Rostov-on-Don		Russia		20		13	
415	Rotterda	Royal Rotterdam Zoological and Botanical Gardens		Rotterdam		Netherlands		21		9	
416	Royan	Zoo de la Palmyre		Royan		France		6		2	
417	Saarbruc	Zoologischer Garten der Landeshauptstadt Saarbrücken		Saarbrücken		Germany		1		0	
418	Sababurg	Tierpark Sababurg		Sababurg		Germany		2		0	
419	Sacramen	Sacramento Zoo		Sacramento, California		USA		6		1	
420	SaintPet	Barbados Primate Research Center and Wildlife Reserve		St Peter		Barbados		1		1	

	A	B	C	D	E	F
	Zoo code	Official name	City	Country	Carnivora sp	Underbred sp
421	Salisbury	Salisbury Zoological Park	Salisbury, Maryland	USA	1	0
422	SaltLake	Utah's Hogle zoological Gardens	Salt Lake City, Utah	USA	8	3
423	Salvador	Parque Zoológico Getúlio Vargas	Salvador, BA	Brazil	1	0
424	Salzburg	Salzburger Tiergarten Hellbrunn	Salzburg	Austria	10	3
425	San Jose	Happy Hollow Zoo	San José, California	USA	1	0
426	SanAnton	San Antonio Zoological Gardens and Aquarium	San Antonio, Texas	USA	16	11
427	SanAntSW	Sea World of Texas	San Antonio, Texas	USA	2	2
428	SanDiego	San Diego Zoological Garden	San Diego, California	USA	34	11
429	SanFranc	San Francisco Zoological Gardens	San Francisco, California	USA	4	1
430	SantaBar	Santa Barbara Zoological Gardens	Santa Barbara, California	USA	3	1
431	Santilla	Santillana del Mar Zoo	Santillana del Mar	Spain	5	2
432	SantoDom	Parque Zoológico Nacional (ZOODOM)	Santo Domingo	Dominican Republic	3	1
433	SaoPaulo	Fundação Parque Zoológico de São Paulo	São Paulo, SP	Brazil	13	4
434	Sapporo	Sapporo Maruyama Zoo	Sapporo	Japan	10	1
435	Sapucaia	Fundação Zoo-Botânica do Rio Grande do Sul-Parque Zoológico	Sapucaia do Sul, RS	Brazil	6	0
436	Schwerin	Zoologischer Garten Schwerin	Schwerin	Germany	11	9
437	SDiegoSW	Sea World of California	San Diego, California	USA	3	2
438	SDiegoWA	San Diego Wild Animal Park	San Diego, California	USA	2	0
439	SeattleW	Woodland Park Zoological Gardens	Seattle, Washington	USA	13	3
440	Sendai	Yagiya Zoological Park	Sendai	Japan	4	2
441	Seoul	Seoul Grand Park Zoo	Seoul	South Korea	2	0
442	SeoulCGP	Children's Grand Park	Seoul	South Korea	4	0
443	Servion	Zoo de Servion	Servion	Switzerland	2	2
444	Seversk	Seversk Zoo	Seversk	Russia	7	6
445	Shaldon	Shaldon Wildlife Trust	Shaldon	England	3	1
446	Shanghai	Shanghai Zoological Gardens	Shanghai	China	3	0
447	Shiraham	Adventure World	Shirahama	Japan	9	5
448	Shizuoka	Shizuoka Municipal Nihondaira Zoo	Shizuoka	Japan	4	2
449	Shubenac	Provincial Wildlife Park	Shubenacadie	Canada	2	0
450	Sigean	Reserve Africaine de Sigean	Sigean	France	2	2
451	Singapor	Singapore Zoological Gardens/Night Safari	Singapore	Singapore	23	9
452	SiouxFal	Great Plains Zoo and Museum	Sioux Falls, South Dakota	USA	5	2
453	Sleopold	Parque Zoológico do Rio Grande do Sul	São Leopoldo, RS	Brazil	5	1
454	Sofia	Sofia Zoological Gardens	Sofia	Bulgaria	10	7

	G	H	I	J	K	L	M	N	O	P	Q	R
	Index	Latitude	No. Of vets	Research	Staff	Size (ha)	Attendance	SpMammals	Total Sp	IndMamm	Total indiv	Management
421												
422	0	38n22	1	No	7	5	150000	22	86	137	408	City
423	0.375	40n46	1	No	46	20	648391	82	303	299	1086	State/Zool. society
424	0	12s5816	5	No	90	8	500000	33	148	201	1046	National
425	0.3	47n48	1	No	24	20	300000	57	114	267	614	Zool. society/National
426	0	37n20	1	No	7	1.2	238539	21	63	134	239	City
427	0.6875	29n25	1	No	168	16	985932	126	647	789	3268	City/Zool. society
428	1	29n25		No								
429	0.323529	32n43	2	Yes	624	41	3352000	227	845	1268	3568	City/Zool society
430	0.25	37n47	2	No	76	44	1151140	115	325	532	983	City/Zool. society
431	0.333333	34n25		No	24	32	282073	34	103	155	435	CHARITY
432	0.4	43n21		No								
433	0.333333	18n28	2	Yes	163	100	239844	37	154	129	1031	National
434	0.307692	23s32	1	Yes	380	65	2550575	138	410	769	2270	State
435	0.1	43n03	2	No	40	22.4	837507	68	224	388	7022	City
436	0	29s50		No								
437	0.818182	53n38		Yes	54	15	320064	47	197	125	764	City
438	0.666667	32n43	3	Yes	750	24	3012179	16	621	146	14912	Private
439	0	32n43	3	Yes	221	728	1219019	126	341	1522	2374	Zool. society
440	0.230769	47n36	2	No	87	37	786401	84	247	326	4853	City
441	0.5	38n15	2	No	51	14.6	612730	57	190	216	802	City
442	0	37n33	3	Yes	325	900	3045701	180	390	1088	4335	City
443	0	37n33		No	20	3.5						City
444	1		2	No	6	5.5	110000	37	100	125	345	Private
445	0.857143	44n51		No								
446	0.333333	50n33	1	No	4	0.6	16000	25	100	63	177	Charity
447	0	31n14		No	585	75	4200000	95	445	320	3801	National
448	0.555556	33n40	4	No	178	100	768218	66	113	742	1181	Private
449	0.5	34n58	1	No	34	11	700000	69	211	360	934	City
450	0	45n11		No								State
451	1	43n02	1	No	29	80	306386	27	263	129	1297	Private
452	0.391304	1n16	1	Yes	131	28	1112000	82	168	510	1443	National
453	0.4	43n33	1	No	13	8	213606	30	61	145	365	City/Zool. society
454	0.2	29s4537	1	Yes	161	153	621524	64	223	418	2561	State
455	0.7	42n41	2	Yes	131	23	685996	78	228	908	2937	City

	A	B	C	D	E	F
	Zoo code	Official name	City	Country	Carnivora sp	Underbred sp
456	Sorocaba	Parque Zoológico Municipal 'Quinzinho de Barros'	Sorocaba, SP	Brazil	10	9
457	Southpor	Southport Zoo and Conservation Trust	Southport	England	5	4
458	Springe	Staatliches Forstamt Saupark - Wisentgehege Springe	Springe	Germany	5	1
460	Springfi	Dickerson Park Zoo	Springfield, Missouri	USA	3	2
461	St. Paul	St. Paul's Como Zoo	St Paul, Minnesota	USA	1	1
462	St.Ettienn	Espace Zoologique de St Martin La Plaine	St. Etienne	France	13	8
463	St.Felici	Zoo de St Felicien	St Felicien	Canada	6	1
464	St.Louis	St. Louis Zoological Park	St Louis, Missouri	USA	12	9
465	St.Peters	Sankt Peterburg Zoo	St Petersburg	Russia	34	25
466	StAignan	Zooparc de Beauval	St Aignan sur Cher	France	2	0
467	StGallen	Wildpark Peter+Paul	St Gallen	Switzerland	2	0
468	StockhAq	Skansen-Akvariet	Stockholm	Sweden	1	1
469	Stockhol	Skansen Foundation, Biological Department	Stockholm	Sweden	6	3
470	Strasbourg	Strasbourg Zoo de la L'Orangerie	Strasbourg	France	2	0
471	Straubin	Tiergarten Straubing	Straubing	Germany	2	0
472	Studen	Zoo Seeteufel	Studen	Switzerland	1	0
473	Stuttgart	Wilhelma Zoologisch-Botanischer Garten	Stuttgart	Germany	12	6
474	Sverdlov	Sverdlovskii Zoopark	Sverdlovsk	Russia	10	5
475	Sydney	Taronga Zoo	Sydney	Australia	4	0
476	Syracuse	Burnet Park Zoo	Syracuse, New York	USA	5	3
477	Tacoma	Point Defiance Zoo and Aquarium	Tacoma, Washington	USA	1	0
478	Taipei	Taipei Zoo	Taipei	Taiwan	15	5
479	Takamats	Ritsurin Park Zoo	Takamatsu	Japan	2	2
480	Takarazu	Takarazuka Zoological and Botanical Gardens	Takarazuka	Japan	9	7
481	Tallin	Tallinn Zoo	Tallin	Estonia	22	12
482	TampaLP	Lowry Park Zoological Garden	Tampa, Florida	USA	5	2
483	Tamworth	Drayton Manor Park Zoo	Tamworth	England	2	1
484	Tarragon	Aqualeon Parc de la Natura (?)	Tarragona	Spain	1	0
485	Tashkent	Tashkent Zoo	Tashkent	Uzbekistan	10	8
486	Tbilisi	Tbilisi Zoopark	Tbilisi	Georgia	2	0
487	Tel Aviv	Meir Segal's Garden for Zoological Research	Tel Aviv	Israel	8	1
488	Termez	Termez Zoo	Termez	Uzbekistan	2	0
489	Thefford	Kilverstone Wildlife Park	Thefford	England	6	3
490	Thoiry	Parc Zoologique de Thoiry	Thoiry	France	1	0

456	G		H		I	J	K	L		M		N		O		P		Q		R
	Index		Latitude					No. Of vets	Research	Staff	Size (ha)	Attendance	SpMammals	Total Sp	IndMamm	Total indiv	Management			
457	0.9		23S30		3	Yes	50	20	420000	55	239	189	1161	City						
458	0.8		53n39			No	6	1.8	82860	34	101	166	627	Private						
459	0.2		52n12			No	6	46	170000	20	230	62	419	State						
460	0.666667		37n13		2	Yes	22	44	225643	37	137	127	435	City/Zool. society						
461	1		44n57		2	No	15	4.5	850000	37	107	162	361	City/Zool. society						
462	0.615385		45n32		1	Yes	5	8	78416	30	120	61	285	Private						
463	0.166667		48n26		1	No	22	385	127783			123	537	Zool. society						
464	0.75		38n38		2	Yes	141	33	2521000	73	613	361	2346	City/Zool. society						
465	0.735294		59n55		1	Yes	165	5.5	1365100	144	423	429	1555	City						
466	0		47n16			No														
467	0		47n25		3	No	2	6.3		8		84		Private						
468	1		59n20		1	Yes	11	1.6	500000	37	334	163	6054	Private						
469	0.5		59n20		1	No	22	33	1749920	26	62	156	451	Zool. society						
470	0		48n35		2	No	3			11	81	48	344	Private						
471	0		48n53			No														
472	0					No	12	6.5	200000	15	86	175	564	Private						
473	0.5		48n46		2	Yes	174	27	1500000	124	784	1007	9497	State/Zool. society						
474	0.5		56n51		1	No	85	1.2	881000	37	126	98	398	City						
475	0		33s52		1	Yes	182	28	911867	110	708	692	3370	State						
476	0.6		43n03		1	Yes	47	24	480000	60	265	235	935	City/Zool. society						
477	0		47n15		1	No	41	27	380199	36	231	308	3704	City						
478	0.333333		25n03		3	No	264	182	3000000	114	273	713	1857	City						
479	1		34n20		2	No	14	0.9	352266	54	116	138	317	Private						
480	0.777778		35n50		1	No	100	16	2467202	45	139	236	675	Private						
481	0.545455		59n25		1	Yes	162	85	530098	108	440	709	4505	City						
482	0.4		27n57		2	No	101	81	3000000	81	337	854	2821	Private						
483	0.5		52n38		2	No	9	6	800000	28	91	67	270	Private						
484	0		41n07			No														
485	0.8		41n20		1	Yes	133	3.8	771000	69	198	194	1075	City						
486	0		41n43		1	Yes	113	6.4	425235	59	213	142	1118	City						
487	0.125		32n04		1	Yes	15	2.8	10000	51	670	309	2492	Zool. society						
488	0		37n14		1	No	92	8	500915	97	256	720	2019	City						
489	0.5		52n25		3	No	29	25	155000	69	359	128	672	Private						
490	0		48n52		2	No	39	150	510000	44	560	84	733	Private						

	A	B	C	D	E	F
	Zoo code	Official name	City	Country	Carnivora sp	Underbred sp
491	Tokuyama	Tokuyama Municipal Zoo	Tokuyama	Japan	9	4
492	TokyoAq	Sunshine International Aquarium	Tokyo	Japan	1	0
493	TokyoIPZ	Inokashira Park Zoo	Tokyo	Japan	4	1
494	TokyoTam	Tama Zoological Park	Tokyo	Japan	6	4
495	TokyoUen	Ueno Zoological Gardens	Tokyo	Japan	7	5
496	Toledo	Toledo Zoological Society	Toledo, Ohio	USA	5	1
497	Tomsk		Tomsk	Russia	8	3
498	Toronto	Toronto Zoo	Toronto	Canada	11	6
499	Toyohash	Toyohashi Zoo and Botanical Park	Toyohashi	Japan	5	3
500	Tregomeu	Jardin Zoologique de Bretagne	Tregomeur	France	1	1
501	Tripoli	Tripoli Zoo	Tripoli	Libya	10	9
502	TucsonAD	Arizona-Sonora Desert Museum	Tucson, Arizona	USA	5	3
503	TucsonRP	Reid Park Zoo	Tucson, Arizona	USA	2	2
504	Tulsa	Tulsa Zoological Park	Tulsa, Oklahoma	USA	6	4
505	Tunis	Parc Zoologique de la Ville de Tunis	Tunis	Tunisia	7	5
506	Tuxtla	Zoológico Regional Miguel Alvarez del Toro	Tuxtla Gutierrez	Mexico	8	1
507	Twycross	Twycross Zoo	Twycross	England	4	2
508	Tyler	Caldwell Zoo	Tyler, Texas	USA	5	3
509	Ueckermu	Tierpark Ueckermunde	Ueckermunde	Germany	2	0
510	Usti	Zoologicka Zahrada Usti nad Labem	Usti nad Labem	Czech Republic	11	7
511	Utica	Utica Zoo	Utica, New York	USA	2	2
512	Valbremb	Parco Faunistico Le Cornello	Valbrembo	Italy	1	0
513	Valencia	Jardin Zoológico de Valencia	Valencia	Spain	2	1
514	VancouAq	Vancouver Aquarium (in Stanley Park)	Vancouver	Canada	1	1
515	Verona	Giardino Zoologico di Verona	Verona	Italy	1	0
516	Veszprem	Kittenberger Zoo Veszprem	Veszprem	Hungary	8	3
517	Vienna	Tiergarten Schoenbrunn	Vienna	Austria	10	5
518	Vigo	Organismo Autónoma Municipal 'Vigo Zoo'	Vigo	Spain	2	0
519	Villiers	Zoorama Europeen de la Foret de Chize	Villiers en Bois	France	7	0
520	Waco	Cameron Park Zoo	Waco, Texas	USA	2	0
521	Warsaw	Miejski Ogród Zoologiczny w Warszawie	Warsaw	Poland	23	17
522	WashNZZP	National Zoological Park	Washington, DC	USA	15	5
523	Wassenaar	Wassenaar Wildlife Breeding Centre	Wassenaar	Netherlands	6	1
524	Wellingt	Wellington Zoological Gardens	Wellington	New Zealand	6	6

	G	H	I	J	K	L	M	N	O	P	Q	R
	Index	Latitude	No. Of vets	Research	Staff	Size (ha)	Attendance	SpMammals	Total Sp	IndMamm	Total indiv	Management
491												
492	0.444444	34n03	2	Yes	27	5	294897	48	156	198	832	City
493	0	35n42		No	30	0.6	1451913	8	463	25	17694	Private
494	0.25	35n42	1	No	32	12	873000	30	245	216	5028	City
495	0.666667	35n42	2	Yes	100	53	1441960	66	208	453	1614	City
496	0.714286	35n42	2	Yes	106	14	6120245	86	840	454	12816	City
497	0.2	41n40	1	No	70	12	615000	60	393	204	1879	Private/Zool. society
498	0.375	56n30		No								
499	0.545455	43n39	2	Yes	229	287	1327426	107	1043	487	3831	City/Zool. society
500	0.6	34n46	3	No	21	12	365869	35	113	158	579	City
501	1	48n24		No	2	10	75000	34		63	400	Private
502	0.9	32n54		No								
503	0.6	32n13	3	No	95	75	567873	28	290	88	4776	Private
504	1	32n13	2	No	20	6.5	387783	33	86	83	214	City/Zool. society
505	0.666667	36n09	1	No	55	27	300000	61	316	194	1309	City
506	0.714286	36n48	2	No	90	13	600000	70	106	304	1634	City
507	0.125	16n45	2	Yes	93	30	400000	28	172	196	646	State
508	0.5	52n38	1	No	45	10	380500	56	350	132	669	Private
509	0.6	32n21	1	Yes	85	14	390611	51	242	209	893	Private
510	0	53n44		No								
511	0.636364	50n40	1	Yes	78	26	150650	66	302	216	1774	City
512	1	43n06	1	No	23	32	65000	40	116	124	263	City/Zool society
513	0			No								
514	0.5	39n28		No								
515	1	49n16	1	Yes	60	1	876825	5	34	544	6808	Zool. society/National
516	0	45n27		No	7	3	100000	22	73	69	207	City
517	0.375	47n06	1	No	37	28	422318	66	306	140	613	City
518	0.5	48n13	1	Yes	88	12	639702	92	764	319	5580	National
519	0	42n14	1	No	18	4.5	102200	14	134	35	433	City
520	0	46n10	1	No	7	25	63173	42	229	137	539	Private
521	0	31n33	1	No	13	4	80000	43	171	156	519	City/Zool. society
522	0.73913	52n15	4	Yes	122	39.5	801000	77	240	475	2407	City
523	0.333333	38n54	4	Yes	320	67	3300000	132	469	1345	4173	National
524	0.166667	52n09	1	Yes	3	19	0	9	79	54	334	Private
525	1	41s18	1	No	27	12	164562	42	145	166	691	City

	A	B	C	D	E	F
	Zoo code	Official name	City	Country	Carnivora sp	Underbred sp
526	Wheeling	Oglebay's Good Zoo	Wheeling, West Virginia	USA	1	1
527	Whipsnad	Zoological Society of London	Whipsnade	England	8	4
528	Wichita	Sedgwick County Zoo	Wichita, Kansas	USA	4	1
530	Widdingt	Mole Hall Wildlife Park	Widdington	England	3	1
531	Windsor	Windsor Safari Park	Windsor	England	1	0
532	Winnipeg	Assiniboine Park Zoo	Winnipeg	Canada	11	1
533	Winston	Wildlife Safari	Winston, Oregon	USA	1	1
534	Woburn	Woburn Safari Park	Woburn	England	4	2
535	WPBeach	Dreher Park Zoological Gardens	West Palm Beach, Florida	USA	2	1
536	Wroclaw	Miejski Ogród Zoologiczny	Wroclaw	Poland	10	8
537	Wuppert	Zoologischer Garten Wuppertal	Wuppertal	Germany	12	0
538	Yangon	Yangon Zoological Gardens	Yangon	Myanmar	3	0
539	Yerevan	Yerevan Zoo	Yerevan	Armenia	6	6
540	Yokohama	Kanazawa Zoological Gardens of Yokohama	Yokohama	Japan	5	3
541	Yongin-k	Yong-in Everland Zoological Gardens	Yongin-kun	South Korea	5	0
542	Yulee	White Oak Conservation Center	Yulee, Florida	USA	3	0
543	Zagreb	Zooloski vrt Zagreb	Zagreb	Croatia	3	1
544	Zamosc	Ogród Zoologiczny im Stefana Milera	Zamosc	Poland	5	1
545	Zhlobin	Zhlobin Zoo	Zhlobin	Belorussia	6	1
546	Zlin	Zoological Garden And Chateau Lesna-Zlin	Zlin	Czech Republic	10	6
547	Zurich	Zoologischer Garten Zurich	Zurich	Switzerland	12	7
548	ZurichWP	Wildpark Langenberg der Stadt Zurich	Zurich	Switzerland	3	0

	G	H	I	J	K	L	M	N	O	P	Q	R
	Index	Latitude	No. Of vets	Research	Staff	Size (ha)	Attendance	SpMammals	Total Sp	IndMamm	Total indiv	Management
526												
527	1	40n04	1	No	16	26	170000	16	81	72	445	City
528	0.5	51n54	2	Yes	102	260	383000	61	1460	172	2414	Zool. society
529	0.25	37n42	1	No	55	86	349081	59	276	316	1402	City/Zool. society
530	0.333333	51n53	3	No	5	10		18	111	54	325	Private
531	0	51n29	2	No	75	56	750000	43	386	106	5664	Private
532	0.090909	49n53	1	No	50	40	653825	71	434	221	1129	City
533	1	43n07	1	No	23	72	152572	45	90	267	528	Private
534	0.5	52n01	1	No	24	142	500000	23	197	32	249	Private
535	0.5	26n43	1	No	21	12	158000	32	95	95	286	Zool. society
536	0.8	51n06	2	No	118	35	772000	117	667	502	5216	City
537	0	51n16	1	No	85	20	712606	72	436	514	3490	City
538	0	16n47		No								
539	1	40n11	1	Yes	87	11.2	500020	84	287	828	3350	City
540	0.6	35n27	4	No	36	9.4	822755	65	251	448	1205	City
541	0	37n33	3	No	48	15	2500000	70	167	621	3247	Private
542	0	30n38	2	Yes	9	60	0	24	54	257	428	Private
543	0.333333	45n48		No								
544	0.2	50n44		No								
545	0.166667	52n54		No								
546	0.6	49n13		No								
547	0.583333	47n23	1	No	69	12	548151	65	242	359	1613	Private
548	0	47n23		No	8	80						City

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	A	B	C	D	E	F	G	H	I	J
	Zoo	Date	Sire	Dam	Subspecies	Place of birth(dam)	Rearing (dam)	No of transfers	Age of dam	M+
1										
2	Bochum	26/02/91	263	145	Sumatran					1
3	Berlin ZG	01/08/88	358	244	Sumatran					0
4	Berlin ZG	09/12/88	358	244	Sumatran					0
5	Berlin ZG	11/02/88	358	244	Sumatran					1
6	Melbourne	25/07/90	365	296	Sumatran					1
7	Wroclaw	31/03/93	500	321	Sumatran					2
8	Berlin ZG	29/04/87	358	323	Sumatran					3
9	Berlin ZG	25/03/90	358	323	Sumatran					1
10	Berlin ZG	25/06/92	358	323	Sumatran					1
11	Berlin ZG	01/06/93	358	323	Sumatran					1
12	Dortmund	01/07/88	280	339	Sumatran					3
13	Frankfurt	30/01/87	362	364	Sumatran	Frankfurt	Unknown	0	7	2
14	Berlin TP	26/05/95	718	384	Sumatran	Berlin TP	Unknown	0	15	0
15	Brno	06/10/91	582	452	Sumatran				8	2
16	Brno	10/06/91	582	452	Sumatran				8	1
17	Ostrava	01/10/87	444	453	Sumatran	Ostrava	Unknown	1	4	0
18	Praha	29/11/94	616	453	Sumatran	Ostrava	Unknown	1	11	1
19	Praha	31/07/95	616	453	Sumatran	Ostrava	Unknown	1	12	1
20	Berlin TP	30/03/90	46	472	Sumatran				7	2
21	Warsaw	27/03/87	500	497	Sumatran	Frankfurt	Unknown	1	3	2
22	Warsaw	15/03/89	500	497	Sumatran	Frankfurt	Unknown	1	5	1
23	Warsaw	18/07/90	500	497	Sumatran	Frankfurt	Unknown	1	6	0
24	Warsaw	22/10/96	500	497	Sumatran	Frankfurt	Unknown	1	12	1
25	Jakarta	01/09/89	535	528	Sumatran				4	2
26	Sd- Wlap	29/11/89	314	536	Sumatran				9	1
27	Yarmouth	09/04/94	674	547	Sumatran	Krefeld	Unknown	1	11	3
28	Yarmouth	11/11/96	674	547	Sumatran	Krefeld	Unknown	1	4	1
29	Arnhem	07/07/89	545	554	Sumatran	Dortmund	Unknown	2	6	3
30	Arnhem	21/07/91	545	554	Sumatran	Dortmund	Unknown	2	4	0
31	Rhenen	21/08/89	553	556	Sumatran				7	1
32	Rhenen	26/06/92	553	556	Sumatran				6	1
33	Atlanta	05/07/91	526	573	Sumatran				7	0
34	London RP	10/02/94	592	626	Sumatran	Bremerhaven	Unknown	2	7	1
35	London RP	19/06/94	592	626	Sumatran	Bremerhaven	Unknown	2	7	1

Appendix 3, page 2

	L		M	N		O	P		Q	R	S		T	U
	U+		Litter size	M-		F-	U-		Young dead	Age of death (days)	Cause of death	Rearing	Inbreeding	
1	0	0	1	1	0	0	0	0	1	1	Unknown	M	0.09370	
2	0	0	1	0	1	0	0	0	1	9	Unknown	U	0.01560	
3	0	0	1	0	1	0	0	0	1	0	Unknown	U	0.01560	
4	0	0	1	1	0	0	0	0	1	34	Euthanasia	U	0.01560	
5	1	0	3	0	0	0	1	1	1	0	Unknown	M	0.03120	
6	0	0	2	1	0	0	0	1	1	5	Unknown	H	0.02740	
7	0	0	3	1	0	0	0	1	1	2	Unknown	U	0.01560	
8	0	0	3	0	1	0	0	1	1	0	Unknown	M	0.01560	
9	1	0	2	1	0	0	0	1	1	0	Stillbirth	M	0.01560	
10	0	0	1	1	0	0	0	1	1	0	Unknown	M	0.01570	
11	0	0	3	1	0	0	0	1	1	0	Unknown	U	0.25000	
12	0	0	2	2	0	0	0	2	2	3,4	Unknown	U	0.03120	
13	1	0	2	0	0	0	1	1	1	2	Unknown	M	0.00000	
14	0	0	2	2	0	0	0	2	2	0,0	Unknown	M	0.28130	
15	0	0	2	1	1	0	0	2	2	5,5	Unknown	U	0.28130	
16	0	0	2	0	2	0	0	2	2	1,2	Unknown	U	0.43750	
17	0	0	2	0	1	0	0	1	1	4	Unknown	H	0.10930	
18	0	0	1	1	0	0	0	1	1	0	Unknown	M	0.10930	
19	1	0	3	1	0	0	1	2	2	1	Unknown	M	0.32810	
20	0	0	3	2	1	0	0	3	3	0,0,3	Unknown	U	0.06250	
21	0	0	2	0	1	0	0	1	1	91	Unknown	U	0.06250	
22	0	0	2	0	2	0	0	2	2	1,2	Unknown	M	0.06250	
23	0	0	1	1	0	0	0	1	1	66	Unknown	M	0.06250	
24	0	0	4	0	2	0	0	2	2	N/A	Unknown	U		
25	0	0	1	1	0	0	0	1	1	0	Unknown	U	0.00000	
26	0	0	3	1	0	0	0	1	1	15	Unknown	M	0.02350	
27	0	0	3	1	1	0	0	2	2	0,38	Stillbirth/Hypothermia	M	0.02340	
28	0	0	3	1	0	0	0	1	1	2	Unknown	M	0.09370	
29	2	0	2	0	0	0	1	1	1	5	Unknown	M	0.09370	
30	0	0	2	1	1	0	0	2	2	0,0	Stillbirth	M		
31	0	0	2	0	1	0	0	1	1	18	Unknown	U	0.39060	
32	0	0	1	0	1	0	0	1	1	6	Unknown	M	0.00000	
33	0	0	2	1	1	0	0	2	2	0,1	Unknown	M	0.09380	
34	0	0	2	1	1	0	0	2	2	0,0	Unknown	M	0.09380	
35	0	0	2	1	1	0	0	2	2	0,0	Unknown	M	0.09380	

	A	B	C	D	E	F	G	H	I	J	K
	Zoo	Date	Sire	Dam	Subspecies	Place of birth(dam)	Rearing (dam)	No of transfers	Age of dam	M+	F+
36											
37	London RP	24/10/94	592	626	Sumatran	Bremerhaven	Unknown	2	9	0	2
38	London RP	05/07/96	592	626	Sumatran	Bremerhaven	Unknown	2	3	1	0
39	Kuala Lumpur	22/05/91	674	671	Sumatran				3	3	1
40	Kuala Lumpur	27/09/91	674	671	Sumatran				5	3	1
41	Dollone	17/10/93	609	697	Sumatran				7	0	0
42	Dudley	15/06/96	827	702	Sumatran	Yarmouth	Unknown	2	4	0	0
43	Rheine	04/11/93	523	707	Sumatran	Berlin TP	Unknown	N/A	4	0	1
44	Rheine	27/03/93	523	707	Sumatran	Berlin TP	Unknown	N/A	5	0	0
45	Rheine	18/06/94	523	707	Sumatran	Berlin TP	Unknown	N/A	4	0	2
46	Atlanta	30/09/93	526	719	Sumatran				2	1	0
47	Amneville	25/08/92	615	732	Sumatran	Berlin ZG	Unknown	3	2	2	0
48	Phoenix	24/03/92	762	761	Sumatran				4	0	0
49	Lisbon	03/12/94	362	790	Sumatran	Rhenen	Unknown	2	5	0	1
50	Lisbon	18/04/95	362	790	Sumatran	Rhenen	Unknown	2	5	0	2
51	Lisbon	30/08/95	362	790	Sumatran	Rhenen	Unknown	2	6	1	1
52	Lisbon	17/01/96	362	790	Sumatran	Rhenen	Unknown	2	4	1	2
53	Berlin ZG	16/07/96	358	797	Sumatran	Berlin TP	Unknown	1	2	1	2
54	Bandung	03/12/96	942	953	Sumatran					2	0
55	Hamburg	05/04/91	3309	1064	Siberian					0	0
56	Zurich	19/05/87	1228	1106	Siberian					0	2
57	Zurich	07/01/89	1228	1106	Siberian					2	0
58	Leipzig	25/05/87	900	1126	Siberian					2	1
59	Amersfoort	02/01/90	1314	1209	Siberian					1	0
60	Amersfoort	10/05/90	1314	1209	Siberian					0	2
61	Emmen	27/11/88	2678	1275	Siberian					1	0
62	Halle	14/08/89	1125	1355	Siberian				11	1	3
63	Nikolaev	28/04/88	1611	1419	Siberian				11	1	2
64	Kyiv Zoo	23/06/88	2500	1580	Siberian				12	1	1
65	Kyiv Zoo	28/05/89	3260	1580	Siberian				10	2	2
66	Veszprem	30/04/88	1630	1635	Siberian				9	1	1
67	Whipsnade	10/03/87	1534	1652	Siberian	Marwell	Unknown	3	11	0	0
68	Whipsnade	12/04/89	1534	1652	Siberian	Marwell	Unknown	3	12	0	1
69	Whipsnade	17/08/90	1534	1652	Siberian	Marwell	Unknown	3	14	0	1
70	Whipsnade	08/08/92	1534	1652	Siberian	Marwell	Unknown	3	16	0	1

	L	M	N	O	P	Q	R	S		T	U
								Young dead	Cause of death		
	U+	Litter size	M-	F-	U-	Young dead	Age of death (days)		Cause of death	Rearing	Inbreeding
36	0	2	0	2	0	2	0, 13		Unknown	H	0.09370
37	1	2	1	0	1	2	0, 1		Stillbirth/Unknown	M	0.09370
38	0	4	1	1	0	2	0, 2		Unknown	U	
39	1	5	0	0	1	1	5		Unknown	M	
40	3	3	0	0	3	3	0, 0, 0		Stillbirth	M	0.08410
41	2	2	0	0	2	2	5, 5		Unknown	M	0.06730
42	1	2	0	1	1	2	5, 13		Unknown	M	0.04680
43	1	1	0	0	1	1	0		Stillbirth	U	0.04680
44	0	2	0	2	0	2	3, 11		Unknown	M	0.04680
45	0	1	1	0	0	1	1		Unknown	U	
46	0	2	2	0	0	2	4, 4		Unknown	U	0.28900
47	3	3	3	0	0	3	2, 2, 2		Unknown	U	
48	2	3	0	1	2	3	0, 0, 1		Unknown	M/H	0.07810
49	0	2	0	2	0	2	2, 3		Unknown	H	0.07810
50	0	2	1	1	0	2	33, 48		Congenital Malformation	H	0.07810
51	0	3	1	0	0	1	11		Killed by dam	M	0.07810
52	0	3	1	1	0	2	10, 13		Unknown	U	0.02340
53	0	2	2	0	0	2	2, 17		Unknown	U	
54	2	2	0	0	2	2	0		Unknown	U	
55	0	2	0	1	0	1	115		Unknown	U	
56	0	2	1	0	0	1	60		Unknown	U	
57	0	3	1	0	0	1	1		Unknown	U	
58	0	1	1	0	0	1	0		Unknown	U	
59	0	2	0	1	0	1	1		Unknown	U	
60	0	1	1	0	0	1	1		Unknown	U	
61	0	4	0	1	0	1	1		Unknown	U	
62	0	3	0	1	0	1	0		Unknown	U	
63	0	2	0	1	0	1	29		Unknown	H	
64	0	4	2	2	0	4	3, 3, 4, 7		Unknown	U	
65	4	2	1	1	0	2	50, 54		Unknown	U	
66	4	4	0	0	2	2	8, 8		Unknown	U	
67	2	3	0	1	0	1	35		Unknown	U	
68	0	1	0	1	0	1	38		Unknown	U	
69	0	1	0	1	0	1	22		Unknown	U	
70	0	1	0	1	0	1			Unknown	U	

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	A	B	C	D	E	F	G	H	I	J	K
	Zoo	Date	Sire	Dam	Subspecies	Place of birth(dam)	Rearing (dam)	No of transfers	Age of dam	M+	F+
71											
72	Whipsnade	15/06/94	1534	1652	Siberian	Marwell	Unknown	3	9	0	1
73	Hilvarenberg	06/12/88	1668	1669	Siberian				10	0	1
74	Hilvarenberg	09/04/89	1668	1669	Siberian				9	1	1
75	Houston P	03/09/88	1694	1683	Siberian				11	1	0
76	Kyoto	31/07/90	1501	1687	Siberian				12	0	1
77	Kyoto	12/06/91	1501	1687	Siberian				8	0	1
78	Sacramento	26/01/87	1861	1730	Siberian				7	1	1
79	Kaunas	28/08/87	930	1761	Siberian				8	1	0
80	Kaunas	19/10/88	2501	1767	Siberian				8	1	2
81	Buffalo	10/12/88	1853	1990	Siberian				7	2	1
82	Philadelphia	16/02/87	1851	1994	Siberian				7	1	1
83	Moscow	15/05/87	1815	2056	Siberian				7	1	1
84	Tallin	08/04/87	3116	2075	Siberian	Wild	Mother	2	7	0	0
85	Tallin	24/06/87	3116	2075	Siberian	Wild	Mother	2	8	3	1
86	Tallin	10/07/88	3116	2075	Siberian	Wild	Mother	2	8	2	0
87	Tallin	20/03/88	3116	2075	Siberian	Wild	Mother	2	10	2	1
88	Tallin	16/06/90	3116	2075	Siberian	Wild	Mother	2	10	2	0
89	Tallin	27/12/90	3116	2075	Siberian	Wild	Mother	2	11	1	1
90	Tallin	08/09/91	3116	2075	Siberian	Wild	Mother	2	12	3	1
91	Tallin	27/06/92	3116	2075	Siberian	Wild	Mother	2	14	1	2
92	Tallin	12/05/94	3116	2075	Siberian	Wild	Mother	2	9	0	2
93	Leipzig	22/05/90	3000	2086	Siberian	Leipzig	Unknown	0	11	4	2
94	Leipzig	10/05/92	3000	2086	Siberian	Leipzig	Unknown	0	13	3	4
95	Leipzig	29/05/94	3000	2086	Siberian	Leipzig	Unknown	0	6	0	2
96	Belo Horizonte	31/01/89	2792	2099	Siberian				6	2	2
97	Nagoya	07/06/87	736	2152	Siberian				7	1	1
98	Rotterdam	15/04/87	1517	2156	Siberian				11	0	1
99	Lympne	27/03/88	2131	2177	Siberian				5	0	3
100	Lympne	08/08/92	2131	2177	Siberian				6	2	1
101	Leipzig	29/05/87	2221	2220	Siberian				5	2	2
102	Leipzig	03/05/88	2221	2220	Siberian				10	1	1
103	Leipzig	17/04/87	900	2222	Siberian				10	2	1
104	Denver	02/03/92	3118	2225	Siberian				9	1	1
105	Denver	08/10/92	3118	2225	Siberian				5	0	0

	L	M	N	O	P	Q	R	S	T	U
	U+	Litter size	M-	F-	U-	Young dead	Age of death (days)	Cause of death	Rearing	Inbreeding
71	1	2	0	1	1	2	2, 2	Unknown	M	0.08590
72	0	1	0	1	0	1	1	Unknown	U	
73	0	2	1	1	0	2	1, 1	Unknown	U	
74	0	1	1	0	0	1	57	Unknown	U	
75	0	1	0	1	0	1	0	Unknown	U	
76	0	1	0	1	0	1	0	Unknown	U	
77	1	3	0	0	1	1	0	Unknown	U	
78	0	1	1	0	0	1	0	Unknown	U	
79	0	3	0	1	0	1	2	Unknown	U	
80	0	3	1	0	0	1	0	Unknown	U	
81	1	3	0	0	1	1	0	Unknown	U	
82	0	2	1	1	0	2	75, 75	Unknown	U	
83	1	1	0	0	1	1	0	Unknown	U	
84	0	4	3	1	0	4	0, 0, 0, 0	Unknown	U	
85	0	2	2	0	0	2	0, 0	Unknown	U	
86	0	3	2	1	0	3	0, 0, 0	Unknown	U	
87	0	2	2	0	0	2	1, 1	Unknown	U	
88	0	2	1	1	0	2	0, 0	Unknown	U	
89	0	4	3	1	0	4	0, 0, 0, 0	Unknown	U	
90	0	3	1	1	0	2	0, 0	Unknown	U	
91	0	2	0	2	0	2	0, 0	Unknown	M	0.00000
92	0	6	1	1	0	2	1, 1	Unknown	U	
93	0	7	2	2	0	4	0, 6, 25, 81	Unknown	U	
94	0	2	0	2	0	2	0, 0	Stillbirth	M	0.00000
95	0	4	0	2	0	2	15, 24	Unknown	U	
96	0	2	1	1	0	2	0, 33	Unknown	U	
97	0	1	0	1	0	1	0	Unknown	U	
98	0	3	0	1	0	1	9	Unknown	U	
99	0	3	0	1	0	1	32	Unknown	U	
100	1	5	1	0	1	2	3, 3	Unknown	U	
101	0	2	1	1	0	2	0, 0	Unknown	U	
102	0	3	0	1	0	1	2	Unknown	U	
103	0	2	1	1	0	2	2, 2	Euthanasia	U	
104	1	1	0	0	1	1	0	Stillbirth	M	

	L	M	N	O	P	Q	R	S	T	U
	U+	Litter size	M-	F-	U-	Young dead	Age of death (days)	Cause of death	Rearing	Inbreeding
141	0	3	2	1	0	3	1, 1, 1	Unknown	U	
142	0	3	1	2	0	3	0, 4, 7	Unknown	U	
143	0	3	2	1	0	3	0, 0, 3	Unknown	U	
144	0	3	2	1	0	3	1, 1, 1	Unknown	U	
145	0	1	0	1	0	1	0	Unknown	U	
146	1	1	0	0	1	1	4	Unknown	U	
147	0	1	0	1	0	1	3	Unknown	U	
148	0	3	0	1	0	1	70	Unknown	U	
149	0	3	2	0	0	2	0, 0	Unknown	U	
150	0	2	1	1	0	2	0, 0	Unknown	U	
151	0	2	1	1	0	2	0, 0	Unknown	U	
152	0	3	2	1	0	3	1, 1, 1	Unknown	U	
153	0	4	1	1	0	2	0, 0	Unknown	U	
154	0	3	1	0	0	1	2	Unknown	U	
155	1	3	0	0	1	1	0	Unknown	U	
156	0	4	0	1	0	1	0	Unknown	U	
157	0	2	0	1	0	1	0	Unknown	U	
158	0	4	0	1	0	1	0	Unknown	U	
159	0	2	0	1	0	1	23	Unknown	U	
160	0	2	0	1	0	1	0	Unknown	M	0 00000
161	0	3	1	0	0	1	53	Unknown	U	
162	0	3	2	1	0	3	0, 0, 0	Unknown	U	
163	0	3	0	1	0	1	0	Unknown	U	
164	0	1	0	1	0	1	0	Unknown	U	0 00000
165	0	5	2	0	0	2	0, 0	Unknown	U	
166	4	4	0	0	4	4	0, 0, 0, 0	Unknown	U	
167	0	6	2	1	0	3	0, 0, 0	Euthanasia	U	
168	0	3	1	0	0	1	5	Unknown	M	0.21100
169	1	3	0	2	1	3	1, 8, 9	Unknown	U	
170	0	3	1	0	0	1	17	Unknown	U	
171	0	3	2	1	0	3	10, 10, 14	Unknown	U	
172	0	4	1	0	0	1	1	Infection	U	
173	1	3	0	0	1	1	0	Unknown	U	
174	1	3	0	0	1	1	1	Unknown	U	
175	0	1	1	0	0	1	0	Unknown	U	

	A		B		C		D		E		F		G		H		I		J		K	
	Zoo	Date	Sire	Dam	Subspecies	Place of birth(dam)	Rearing (dam)	No of transfers	Age of dam	M+	F+											
176	Bekesbourne	03/08/91	2927	2910	Siberian				6	2	2											
177	Bekesbourne	19/08/92	2927	2910	Siberian				7	2	5											
179	Duisburg	13/06/93	3545	2923	Siberian	Munster		1	8	1	3											
180	Duisburg	25/06/94	3545	2923	Siberian	Munster		1	6	1	0											
181	Duisburg	13/12/92	3545	2924	Siberian	Munster		1	7	4	0											
182	Duisburg	09/05/93	3545	2924	Siberian	Munster		1	4	1	3											
183	Aalborg	08/08/90	3184	2955	Siberian	Stuttgart		1	5	0	1											
184	Aalborg	12/05/91	3184	2955	Siberian	Stuttgart		1	5	0	0											
185	Aalborg	30/11/91	3184	2955	Siberian	Stuttgart		1	5	1	1											
186	Koln	10/05/90	3030	2956	Siberian				4	0	3											
187	Koln	27/03/91	3030	2956	Siberian				5	2	2											
188	Koln	20/09/93	3030	2956	Siberian				7	0	0											
189	Osaka	16/04/92	3144	2994	Siberian				6	1	1											
190	Osaka	19/08/92	3144	2994	Siberian				6	4	1											
191	Osaka	20/05/93	3144	2994	Siberian				7	3	1											
192	Kaunas	20/09/87	2500	3016	Siberian				2	1	1											
193	Kaunas	01/09/88	2501	3016	Siberian				3	0	0											
194	Kaunas	20/04/88	2501	3016	Siberian				3	1	2											
195	Kaunas	22/04/89	2501	3016	Siberian				4	0	0											
196	Kaunas	19/05/91	2501	3016	Siberian				6	1	1											
197	Kaunas	10/05/91	2501	3020	Siberian	Kaunas		0	5	0	1											
198	Kaunas	13/09/91	2501	3020	Siberian	Kaunas		0	5	1	1											
199	Kaunas	14/05/93	2501	3020	Siberian	Kaunas		0	7	4	0											
200	Emmen	08/10/89	2678	3028	Siberian	Leipzig		N/A	2	1	2											
201	Emmen	02/06/92	3257	3028	Siberian	Leipzig		N/A	5	2	1											
202	Emmen	04/06/90	2678	3035	Siberian				3	1	2											
203	Emmen	04/06/91	3257	3035	Siberian				4	2	1											
204	Emmen	21/05/92	3257	3035	Siberian				5	2	0											
205	Emmen	03/05/93	3257	3035	Siberian				6	1	1											
206	Darjeelin	12/12/93	2077	3043	Siberian				6	1	2											
207	Darjeelin	30/07/93	2077	3043	Siberian				6	2	1											
208	Edinburgh	07/07/91	2308	3073	Siberian	Marwell		2	4	1	3											
209	Kyiv Zoo	07/09/89	3260	3155	Siberian	Tallin		3	4	2	2											
210	Kyiv Zoo	06/09/90	3260	3155	Siberian	Tallin		3	5	4	0											

	L	M	N		O		P		Q	R	S	T	U
			U+	Litter size	M-	F-	U-	Young dead					
176	0	4	0	1	0	0	1	0	1	3	Unknown	U	
177	0	7	0	1	0	0	1	0	1	5	Unknown	U	
178	0	4	0	2	0	0	2	0	2	29, 58	Euthanasia	M	
180	0	1	1	0	0	0	0	0	1	31	Unknown	M	0 00000
181	0	4	4	0	0	0	0	0	4	0, 0, 0, 0	Unknown	M	
182	0	4	0	1	0	0	1	0	1	5	Infection	H	
183	0	1	0	1	0	0	1	0	1	2	Unknown	M	
184	1	1	0	0	1	0	0	1	1	0	Stillbirth	U	
185	0	2	1	1	0	0	1	0	2	0, 0	Stillbirth	U	
186	0	3	0	3	0	0	3	0	3	1, 1, 1	Unknown	U	
187	0	4	1	1	0	0	1	0	2	70, 74	Unknown	U	
188	3	3	0	0	3	0	0	3	3	6, 21, 46	Unknown	M	
189	0	2	1	0	0	0	0	0	1	1	Unknown	U	
190	0	5	4	0	0	0	0	0	4	1, 1, 2, 7	Unknown	U	
191	0	4	1	1	0	0	1	0	2	1, 1	Unknown	U	
192	0	2	1	1	0	0	1	0	2	0, 0	Unknown	U	
193	4	4	0	0	4	0	0	4	4	0, 0, 0, 8	Unknown	U	
194	0	3	1	1	0	0	1	0	2	0, 1	Unknown	U	
195	1	1	0	0	1	0	0	1	1	0	Unknown	U	
196	0	2	1	1	0	0	1	0	2	0, 0	Unknown	U	
197	2	3	0	1	2	0	1	2	3	2, 2, 2	Unknown	U	
198	0	2	1	1	0	0	1	0	2	0, 0	Unknown	U	
199	2	6	3	0	0	0	0	0	3	8, 9, 10	Unknown	H	
200	0	3	1	2	0	0	2	0	3	0, 0, 2	Unknown	U	
201	0	3	2	1	0	0	1	0	3	62, 62, 63	Unknown	M	
202	3	6	0	0	3	0	0	3	3	2, 2, 2	Unknown	U	
203	0	3	2	1	0	0	1	0	3	0, 0, 0	Unknown	U	
204	1	3	1	0	1	0	0	1	2	5, 7	Unknown	M	
205	1	3	1	1	0	0	1	0	2	0, 8	Unknown	M	
206	0	3	1	1	0	0	1	0	2	4, 5	Unknown	U	
207	0	3	1	1	0	0	1	0	2	0, 0	Unknown	U	
208	2	6	1	0	2	0	0	2	3	0, 0, 7	Unknown	U	
209	0	4	1	2	0	0	2	0	3	5, 9, 10	Unknown	U	
210	0	4	1	0	0	0	0	0	1	0	Unknown	U	

	A	B	C	D	E	F	G	H	I	J	K
	Zoo	Date	Sire	Dam	Subspecies	Place of birth(dam)	Rearing (dam)	No of transfers	Age of dam	M+	F+
211											
212	Geselkirchen	21/07/94	3920	3200	Siberian	Edinburgh	Unknown	2	6	1	3
213	Munster	20/05/92	2492	3259	Siberian	Berlin TP	Unknown	1	4	0	0
214	Kyiv Zoo	13/11/88	2500	3261	Siberian				1	2	1
215	Kyiv Zoo	26/05/89	2500	3261	Siberian				2	2	2
216	Kyiv Zoo	29/05/92	2500	3261	Siberian				5	1	2
217	Tallin	13/01/92	3116	3269	Siberian				4	0	2
218	Rostov	26/04/92	2028	3270	Siberian	Wild	Mother	1	5	1	0
219	Rostov	08/05/93	2028	3270	Siberian	Wild	Mother	1	6	0	2
220	Rostov	21/09/93	2028	3270	Siberian	Wild	Mother	1	6	1	0
221	Ekaterinburg	18/09/89	3288	3289	Siberian				1	3	2
222	Tucson	21/03/92	3021	3343	Siberian				4	1	1
223	Ekaterinburg	23/07/88	3288	3348	Siberian				1	2	2
224	Kaliningrad	02/07/92	3900	3350	Siberian	Wild	Mother	1	6	2	2
225	Kaliningrad	24/04/93	3900	3350	Siberian	Wild	Mother	1	7	1	0
226	Kaliningrad	11/05/95	3900	3350	Siberian	Wild	Mother	1	9	2	2
227	Riga	15/06/92	3536	3366	Siberian	Wild	Mother	2	4	0	2
228	Riga	27/05/93	3536	3366	Siberian	Wild	Mother	2	5	1	2
229	Riga	03/04/94	3536	3366	Siberian	Wild	Mother	2	6	1	0
230	Riga	04/09/95	3536	3366	Siberian	Wild	Mother	2	7	2	1
231	Munich	23/06/91	3260	3368	Siberian				3	0	1
232	Munich	23/10/91	3260	3368	Siberian				3	1	1
233	Chicago BR	30/06/92	2227	3411	Siberian				3	0	1
234	Praha	08/04/96	3104	3486	Siberian				7	2	2
235	Zurich	17/08/92	3373	3501	Siberian	Wild	Mother	3	3	0	0
236	Zurich	17/12/92	3373	3501	Siberian	Wild	Mother	3	3	1	0
237	Kyiv Zoo	02/06/92	3514	3513	Siberian	Wild	Mother	1	5	2	1
238	Kyiv Zoo	03/02/92	3514	3513	Siberian	Wild	Mother	1	5	2	1
239	Emmen	05/05/93	3257	3566	Siberian				3	1	2
240	Emmen	16/05/93	3257	3567	Siberian	Emmen	Unknown	1	3	1	1
241	Osnabruck	04/07/93	3455	3577	Siberian				3	6	0
242	Berlin ZG	10/05/93	2719	3598	Siberian	Budapest	Unknown	1	3	2	1
243	Berlin ZG	15/09/93	2719	3598	Siberian	Budapest	Unknown	1	3	2	2
244	Berlin ZG	05/11/95	2719	3598	Siberian	Budapest	Unknown	1	5	1	2
245	Whipsnade	20/10/94	1534	3636	Siberian	Whipsnade	Mother	2	4	1	2

	L	M	N	O	P	Q	R	S	T	U
	U+	Litter size	M-	F-	U-	Young dead	Age of death (days)	Cause of death	Rearing	Inbreeding
211	0	4	0	1	0	1	15	Unknown	U	0 29390
212	3	3	0	0	3	3	2, 2, 5	Unknown	U	
213	0	3	2	1	0	3	0, 0, 0	Killed by dam	M	
214	0	4	1	1	0	2	3, 8	Unknown	U	
215	0	3	1	1	0	2	0, 1	Killed by dam	M	
216	0	2	0	2	0	2	0, 0	Unknown	U	
217	0	1	1	0	0	1	2	Unknown	U	
218	0	2	0	2	0	2	0, 4	Killed by dam	M	
219	0	1	1	0	0	1	19	Unknown	H	0.00000
220	1	5	2	1	0	3	0, 34, 46	Unknown	U	
221	0	3	1	0	0	1	25	Unknown	U	
222	0	4	1	0	0	1	2	Unknown	U	
223	0	4	2	2	0	4	0, 0, 0, 0	Stillbirth	M	
224	0	1	1	0	0	1	0	Unknown	M	
225	0	4	2	1	0	3	1, 1, 1	Killed by dam	M	0.00000
226	0	2	0	1	0	1	82	Unknown	U	
227	0	3	1	2	0	3	0, 0, 0	Killed by dam	M	
228	0	1	1	0	0	1	1	Killed by dam	M	0.00000
229	0	3	2	1	0	3	1, 1, 7	Unknown	H	0.00000
230	0	1	0	1	0	1	8	Unknown	U	
231	0	2	1	1	0	2	0, 11	Unknown	U	
232	0	1	0	1	0	1	10	Unknown	U	0 00000
233	0	4	2	1	0	3	32, 37, 40	Unknown	U	
234	1	1	0	0	1	1	0	Unknown	U	
235	0	1	1	0	0	1	0	Unknown	U	
236	0	3	2	1	0	3	0, 0, 6	Killed by dam	M	
237	0	3	2	1	0	3	0, 0, 0	Killed by dam	M	
238	0	3	1	0	0	1	12	Unknown	M	
239	1	3	1	1	1	3	0, 1, 4	Unknown	M	
240	0	6	1	0	0	1	0	Unknown	U	
241	0	3	2	1	0	3	0, 0, 0	Unknown	M	
242	0	4	2	2	0	4	0, 0, 0, 0	Unknown	M	
243	0	3	1	2	0	3	0, 0, 0	Euthanasia	M	0 05820
244	0	3	1	2	0	3	0, 0, 3	Unknown	M	0 30070
245	0	3	1	2	0	3				

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	A	B	C	D	E	F	G	H	I	J	K
	Zoo	Date	Sire	Dam	Subspecies	Place of birth(dam)	Rearing (dam)	No of transfers	Age of dam	M+	F+
246											
247	Perm	06/05/90	3654	3655	Siberian				1	1	1
248	Perm	13/09/90	3654	3655	Siberian				1	2	1
249	Perm	14/04/91	3654	3655	Siberian				2	3	1
250	Perm	16/08/91	3654	3655	Siberian				2	2	2
251	Madrid Zoo	27/12/94	3682	3686	Siberian				4	2	1
252	Dvur Kralove	08/10/95	3832	3697	Siberian	Warsaw	Unknown	1	4	1	1
253	Leipzig	30/04/95	3821	3800	Siberian	Leipzig	Unknown	0	3	2	1
254	Leipzig	09/06/96	3821	3800	Siberian	Leipzig	Unknown	0	4	3	0
255	Moscow	01/03/96	4125	3815	Siberian	Wild	Mother	2	5	2	1
256	Praha	16/05/96	3485	3883	Siberian	Riga	Unknown	1	4	0	2
257	St Petersburg	19/06/96	4003	3953	Siberian	Wild	Mother	4	4	2	3

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	A	B	C	K	L	M	N	O	P	Q	R
	Family	Genus	Species	Zoo	Dam	Origin	Rearing	Age	Litter size	Young dead	Infant Mortality
1	Canidae	Alopex	lagopus	London RP	A				5	0	0 00
2	Canidae	Alopex	lagopus	London RP	B				4	0	0 00
3	Canidae	Canis	familians	London RP	A				1	1	1 00
4	Canidae	Canis	latrans	London RP	A				3	0	0 00
5	Canidae	Canis	latrans	London RP	A				7	0	0 00
6	Canidae	Canis	lupus	Dresden	A	Captivity	Parental	3	5	5	1 00
7	Canidae	Canis	lupus	Dresden	A	Captivity	Parental	4	4	0	0 00
8	Canidae	Canis	lupus	Dresden	A	Captivity	Parental	5	3	0	0 00
9	Canidae	Canis	lupus	Dresden	A	Captivity	Parental	6	4	4	1 00
10	Canidae	Canis	lupus	Dresden	A	Captivity	Parental	7	4	4	1 00
11	Canidae	Canis	lupus	Dresden	A	Captivity	Parental	8	1	0	0 00
12	Canidae	Canis	lupus	Jaipur	A	Wild		3	4	0	0 00
13	Canidae	Canis	lupus	Jaipur	A	Wild		6	3	1	0 33
14	Canidae	Canis	lupus	Jaipur	A	Wild		7	5	0	0 00
15	Canidae	Canis	lupus	Rio	A	Wild	Parental		3	3	1 00
16	Canidae	Cercocyon	thous	WNZP	1	Wild	Hand	1	3	0	0 00
17	Canidae	Cercocyon	thous	WNZP	1	Wild	Hand	1	5	2	0 40
18	Canidae	Cercocyon	thous	WNZP	2	Wild	Hand	1	4	0	0 00
19	Canidae	Cercocyon	thous	WNZP	2	Wild	Hand	2	4	4	1 00
20	Canidae	Cercocyon	thous	WNZP	3	Captivity	Parental	1	6	0	0 00
21	Canidae	Cercocyon	thous	WNZP	3	Captivity	Parental		4	4	1 00
22	Canidae	Chrysocyon	brachyurus	Frankfurt	A	Wild	Parental		3	3	1 00
23	Canidae	Chrysocyon	brachyurus	Frankfurt	A	Wild	Parental		2	2	1 00
24	Canidae	Chrysocyon	brachyurus	Frankfurt	A	Wild	Parental		2	2	1 00
25	Canidae	Chrysocyon	brachyurus	Brasilia	A	Wild	Parental	3	1	1	1 00
26	Canidae	Chrysocyon	brachyurus	Brasilia	B	Wild	Parental	3	1	1	1 00
27	Canidae	Chrysocyon	brachyurus	Brasilia	C	Wild	Parental	3	3	3	1 00
28	Canidae	Chrysocyon	brachyurus	Los Angeles	A	Wild	Parental	4	2	2	1 00
29	Canidae	Chrysocyon	brachyurus	Los Angeles	A	Wild	Parental	5	5	0	0 00
30	Canidae	Chrysocyon	brachyurus	WNZP	50	Wild	Parental		2	0	0 00
31	Canidae	Chrysocyon	brachyurus	WNZP	50	Wild	Parental		2	2	1 00
32	Canidae	Chrysocyon	brachyurus	WNZP	50	Wild	Parental		2	0	0 00
33	Canidae	Chrysocyon	brachyurus	WNZP	53	Wild	Parental		1	1	1 00
34	Canidae	Chrysocyon	brachyurus	WNZP	53	Wild	Parental		1	0	0 00
35	Canidae	Chrysocyon	brachyurus	Chicago LP	A				3	0	0 00
36	Canidae	Chrysocyon	brachyurus	Chicago LP	A				1	0	0 00
37	Canidae	Chrysocyon	brachyurus	Belo Horizonte	Luana				3	0	0 00
38	Canidae	Chrysocyon	brachyurus	Belo Horizonte	Xuxa				4	0	0 00
39	Canidae	Chrysocyon	brachyurus	Houston	1029		Parental	5	4	0	0 00
40	Canidae	Chrysocyon	brachyurus	Fossil Rim	1191		Hand	4	2	0	0 00
41	Canidae	Chrysocyon	brachyurus	Fossil Rim	1285		Parental	4	3	0	0 00
42	Canidae	Cuon	alpinus	Moscow	Ozornitsa	Captivity		2	3	3	1 00
43	Canidae	Cuon	alpinus	Moscow	Reta	Wild	Parental	10	2	2	1 00
44	Canidae	Cuon	alpinus	Duisburg	A	Wild	Hand	4	5	1	0 20

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	S	T	U	V	W	X	Y
	Cause of death	Age of death(Days)	Cage area (m2)	Ind/cage	Dens	Male separated?	Reference
1				6	5	No	Kleiman 1968
2				6	5	No	Kleiman 1968
3							Kleiman 1968
4	Eaten by dam	0					Kleiman 1968
5				1		Yes	Kleiman 1968
6				1		Yes	Kleiman 1968
7	Eaten by dam	5		2		Yes	Gensch 1968
8				2		No	Gensch 1968
9				2		No	Gensch 1968
10	Killed by others	0		5		No	Gensch 1968
11	Killed by dam	0		2		No	Gensch 1968
12				2		No	Gensch 1968
13			20	2	3	No	Yadav 1968
14	Enteritis	129	20	2	3	No	Yadav 1968
15			20	2	3	No	Yadav 1968
16	Neglect	2	8	2	1		Coimbra-Filho 1966
17			57	3	1	Yes	Brady 1978
18	Not Reported	0	57	3	1	No	Brady 1978
19			57	3	1	No	Brady 1978
20	Killed by others	0	57	10	2	No	Brady 1978
21			57	4	2	No	Brady 1978
22	Eaten by dam	10	248	2	1	Yes	Faust & Sherpner 1967
23	Eaten by dam	10	248	2	1	Yes	Faust & Sherpner 1967
24	Eaten by dam	10	248	2	1	Yes	Faust & Sherpner 1967
25	Eaten by dam	1	150		1	Yes	Silveira 1968
26	Eaten by dam	1	400		1	Yes	Silveira 1968
27	Eaten by dam	0	150		1	Yes	Silveira 1968
28	Heart defect	0		2		No	Acosta 1972
29				2		No	Acosta 1972
30			20	2	2		Brady & Ditton 1978
31	Not Reported	0	20	2	2		Brady & Ditton 1979
32			20	2	2		Brady & Ditton 1979
33	Not Reported	0	20	2	2		Brady & Ditton 1979
34			20	2	2		Brady & Ditton 1979
35						Yes	Rosenthal & Dunn 1995
36						Yes	Rosenthal & Dunn 1995
37			1100	2	1	Yes	Veado 1997
38			338	2	3	No	Veado 1997
39						No	Bestelmeyer 1999
40						Yes	Bestelmeyer 1999
41						No	Bestelmeyer 1999
42	Killed by dam	6	12	2	1		Sosnovskii 1967
43	Killed by dam	2	12	2	1		Sosnovskii 1967
44	Weakness	3	35	3	1	No	Gewalt 1978

	A	B	C	K	L	M	N	O	P	Q	R
	Family	Genus	Species	Zoo	Dam	Origin	Rearing	Age	Litter size	Young dead	Infant Mortality
45	Canidae	Cuon	alpinus	Duisburg	B	Wild	Hand	4	4	4	1 00
46	Canidae	Duscicyon	vetulus	Rio	A	Wild	Parental		4	4	1 00
47	Canidae	Vulpes	zerda	Strasbourg	A				2	2	1 00
48	Canidae	Vulpes	zerda	Strasbourg	B				6	1	0 17
49	Canidae	Vulpes	zerda	Melbourne	A	Captivity			3	2	0 67
50	Canidae	Vulpes	zerda	Pittsburgh	A				3	3	1 00
51	Canidae	Vulpes	zerda	Pittsburgh	A				3	3	1 00
52	Canidae	Vulpes	zerda	Pittsburgh	A				3	3	0 33
53	Canidae	Vulpes	zerda	Pittsburgh	A				2	1	0 50
54	Canidae	Vulpes	zerda	Pittsburgh	A				2	1	0 50
55	Canidae	Lycyaon	pictus	Nairobi	A			2	4	4	1 00
56	Canidae	Lycyaon	pictus	Amsterdam	A			2	2	2	1 00
57	Canidae	Lycyaon	pictus	Amsterdam	A			2	8	3	0 38
58	Canidae	Lycyaon	pictus	Amsterdam	A			3	9	0	0 00
59	Canidae	Otocyon	megalotis	Utica USA	A				5	5	1 00
60	Canidae	Speothos	venaticus	Chicago LP	A	Wild	Parental	3	3	3	1 00
61	Canidae	Speothos	venaticus	Los Angeles	A				6	2	0 33
62	Canidae	Speothos	venaticus	Frankfurt	A	Wild	Parental	3	6	5	0 83
63	Canidae	Speothos	venaticus	Frankfurt	A	Wild	Parental	4	6	5	0 83
64	Canidae	Speothos	venaticus	Frankfurt	A	Wild	Parental	5	6	2	0 33
65	Canidae	Vulpes	corsac	Berlin TP	A				5	0	0 00
66	Canidae	Vulpes	corsac	Berlin TP	A				5	0	0 00
67	Canidae	Vulpes	corsac	Berlin TP	A				6	0	0 00
68	Canidae	Vulpes	corsac	Berlin TP	A				3	0	0 00
69	Canidae	Vulpes	corsac	Berlin TP	B				3	1	0 33
70	Felidae	Acinonyx	jubatus	Oklahoma City	A			2	3	3	1 00
71	Felidae	Acinonyx	jubatus	Oklahoma City	A			2	3	3	1 00
72	Felidae	Acinonyx	jubatus	Private	Beauty	Wild	Parental	4	1	0	0 00
73	Felidae	Acinonyx	jubatus	Private	Beauty	Wild	Parental	4	3	0	0 00
74	Felidae	Acinonyx	jubatus	Whipsnade	Juanita	Captivity	Parental	3	3	0	0 00
75	Felidae	Acinonyx	jubatus	Whipsnade	Juanita	Captivity	Parental	4	3	0	0 00
76	Felidae	Acinonyx	jubatus	Whipsnade	Juanita	Captivity	Parental	6	2	0	0 00
77	Felidae	Acinonyx	jubatus	Montpellier	A	Wild	Parental	4	3	0	0 00
78	Felidae	Acinonyx	jubatus	Arnhem	A				2	2	1 00
79	Felidae	Acinonyx	jubatus	Krefeld	A				4	2	0 50
80	Felidae	Acinonyx	jubatus	Montpellier	A	Wild	Parental	6	4	0	0 00
81	Felidae	Acinonyx	jubatus	Philadelphia	A				3	3	1 00
82	Felidae	Acinonyx	jubatus	Philadelphia	A				2	2	1 00
83	Felidae	Acinonyx	jubatus	Rome	A				3	0	0 00
84	Felidae	Acinonyx	jubatus	San Diego	A				3	2	0 67
85	Felidae	Acinonyx	jubatus	Whipsnade	Juanita	Captivity	Parental	7	3	0	0 00
86	Felidae	Acinonyx	jubatus	Toledo	B	Wild	Parental	3	4	0	0 00
87	Felidae	Acinonyx	jubatus	Whipsnade	Janica	Captivity	Parental	4	1	0	0 00
88	Felidae	Acinonyx	jubatus	Beekse BSP	Angela			4	5	0	0 00

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	S	T	U	V	W	X	Y
	Cause of death	Age of death(Days)	Cage area (m2)	Ind/cage	Dens	Male separated?	Reference
45	Eaten by dam	0	35	3	1	No	Gewalt 1978
46	Neglect	1	8	2	1		Coimbra-Filho 1966
47	Neglect	2	2	1	1	Yes	Gangloff 1972
48	Toxocara infection	38	7	1	2	Yes	Gangloff 1972
49	Neglect/Hand-rearing	20		2		No	Weiber 1976
50	Killed by dam	2				No	Thomas & Philips 1978
51	Killed by dam	2				No	Thomas & Philips 1978
52	Killed by dam	1				No	Thomas & Philips 1978
53	Hand-rearing	5				No	Thomas & Philips 1978
54	Eaten by dam	6		2	1		Cade 1967
55	Eaten by dam	1	90	2	2	No	Dekker 1968
56	Pneumonia	8	90	2	2	Yes	Dekker 1968
57	Neglect	2	90	2	3	No	Dekker 1968
58	Congenital defect/Hand-rearing	12	4	2	1	No	Rosenberg 1971
59	Neglect	7		2		Yes	Kitchener 1971
60	Eaten by dam/pneumonia	4		1	2	Yes	Collier & Emerson 1973
61	Neglect	2		2	2	Yes	Jantschke 1973
62	Stilbirth/enteritis	0		2	2	Yes	Jantschke 1973
63				5			Dathe 1961
64				5			Dathe 1961
65				5			Dathe 1961
66				5			Dathe 1961
67				5			Dathe 1961
68				5			Dathe 1961
69	Eaten by dam			2	1	No	Thomas 1965
70	Killed by others	0	56	2	1	No	Thomas 1965
71	Neglect/Hand-rearing	0, 0, 10	55	2	1	No	Thomas 1965
72			200	1		Yes	Fiorio & Spinelli 1967
73					1	No	Fiorio & Spinelli 1968
74			150	2	2	Yes	Manton 1970
75			150	2	2	Yes	Manton 1970
76			150	2	2	Yes	Manton 1971
77			1200	3		Yes	Vallat 1971
78	Eaten by dam	2					Rawlins 1972
79	Eaten by dam	2	90				Rawlins 1972
80			1200	3		Yes	Rawlins 1972
81	Killed by dam/Hand-rearing	3					Rawlins 1972
82	Hand-rearing	90					Rawlins 1972
83			150	3		No	Rawlins 1972
84	Eaten by dam	0	2000	3		No	Rawlins 1972
85			150	2	2	Yes	Rawlins 1972
86			1748	4	1	Yes	Skeldon 1973
87			540	3	1	Yes	Manton 1974
88			4000	10	1	Yes	Tong 1974

	A	B	C	K	L	M	N	O	P	Q	R
	Family	Genus	Species	Zoo	Dam	Origin	Rearing	Age	Litter size	Young dead	Infant Mortality
89	Felidae	Acinonyx	Jubatus	Fota	FWP35	Captivity	Parental	4	4	0	0 00
90	Felidae	Acinonyx	Jubatus	Fota	FWP37	Captivity	Parental	4	5	0	0 00
91	Felidae	Acinonyx	Jubatus	Fota	FWP75	Wild	Parental	5	2	0	0 00
92	Felidae	Catopuma	temminckii	Wassenaar	A	Wild	Parental	3	1	0	0 00
93	Felidae	Catopuma	temminckii	Wassenaar	A	Wild	Parental	3	1	0	0 00
94	Felidae	Catopuma	temminckii	Melbourne	Cassandra	Captivity			1	1	1 00
95	Felidae	Catopuma	temminckii	Melbourne	Cassandra	Captivity			2	0	0 00
96	Felidae	Catopuma	temminckii	Melbourne	Cassandra	Captivity			1	1	1 00
97	Felidae	Catopuma	temminckii	Melbourne	Cassandra	Captivity			1	0	0 00
98	Felidae	Catopuma	temminckii	Melbourne	Cassandra	Captivity			1	0	0 00
99	Felidae	Catopuma	temminckii	Melbourne	Cassandra	Captivity			1	0	0 00
100	Felidae	Catopuma	temminckii	Melbourne	Cassandra	Captivity			1	0	0 00
101	Felidae	Catopuma	temminckii	Melbourne	Cassandra	Captivity			1	1	1 00
102	Felidae	Catopuma	temminckii	Melbourne	Cassandra	Captivity			1	0	0 00
103	Felidae	Catopuma	temminckii	Melbourne	Cassandra	Captivity			1	0	0 00
104	Felidae	Catopuma	temminckii	Melbourne	Cassandra	Captivity			1	0	0 00
105	Felidae	Catopuma	temminckii	Melbourne	Cassandra	Captivity			1	0	0 00
106	Felidae	Catopuma	temminckii	Melbourne	Cim	Captivity			1	0	0 00
107	Felidae	Catopuma	temminckii	Melbourne	Cim	Captivity			1	0	0 00
108	Felidae	Catopuma	temminckii	Melbourne	Indra	Captivity			1	0	0 00
109	Felidae	Catopuma	temminckii	Melbourne	Indra	Captivity		4	2	0	0 00
110	Felidae	Catopuma	temminckii	Melbourne	Marigold	Captivity		4	1	0	0 00
111	Felidae	Catopuma	temminckii	Melbourne	Marigold	Captivity		5	2	0	0 00
112	Felidae	Catopuma	temminckii	Melbourne	Marigold	Captivity		5	1	0	0 00
113	Felidae	Catopuma	temminckii	Melbourne	Marigold	Captivity		6	1	1	1 00
114	Felidae	Catopuma	temminckii	Melbourne	Marigold	Captivity		6	1	1	1 00
115	Felidae	Felis	bengalensis	Berlin TP	Sura	Wild	Parental	4	3	0	0 00
116	Felidae	Felis	bengalensis	Berlin TP	Sura	Wild	Parental	5	3	2	0 67
117	Felidae	Felis	bengalensis	Berlin TP	Sura	Wild	Parental	6	3	0	0 00
118	Felidae	Felis	bengalensis	Berlin TP	Sura	Wild	Parental	7	3	2	0 67
119	Felidae	Felis	bengalensis	Berlin TP	Sura	Wild	Parental	7	3	3	1 00
120	Felidae	Felis	bengalensis	Berlin ZG	A			6	1	0	0 00
121	Felidae	Felis	bengalensis	Berlin ZG	A			7	3	0	0 00
122	Felidae	Felis	bengalensis	Berlin ZG	A			8	2	0	0 00
123	Felidae	Caracal	caracal	Mysore	A	Captivity	Parental	1	1	0	0 00
124	Felidae	Caracal	caracal	Mysore	A	Captivity	Parental	2	1	0	0 00
125	Felidae	Caracal	caracal	Brno	1	Wild	Parental	5	6	2	0 33
126	Felidae	Caracal	caracal	Brno	2	Wild	Parental	5	4	4	1 00
127	Felidae	Caracal	caracal	Nairobi	A		Hand	2	2	0	0 00
128	Felidae	Felis	margarita	Brookfield	Br2	Wild	Parental		4	4	1 00
129	Felidae	Felis	margarita	Brookfield	Br2	Wild	Parental		4	0	0 00
130	Felidae	Felis	margarita	Brookfield	Br2	Wild	Parental		4	4	1 00
131	Felidae	Felis	margarita	Brookfield	Br2	Wild	Parental		4	0	0 00
132	Felidae	Felis	margarita	Brookfield	Br2	Wild	Parental		2	2	1 00

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	S	T	U	V	W	X	Y
	Cause of death	Age of death(Days)	Cage area (m2)	Ind/cage	Dens	Male saparated?	Reference
89							
90			405	1	3	Yes	O'Donovan et al/ 1993
91			1515	1	2	Yes	O'Donovan et al. 1993
92			406	1	2	Yes	O'Donovan et al/ 1993
93				2		No	Louwman & Van Oyen 1968
94				2		No	Louwman & Van Oyen 1968
95	Not Reported		50		1		Brocklehurst 1997
96			50		1		Brocklehurst 1997
97	Not Reported		50		1		Brocklehurst 1997
98			50		1		Brocklehurst 1997
99			50		1		Brocklehurst 1997
100			50		1		Brocklehurst 1997
101	Not Reported		50		1		Brocklehurst 1997
102			50		1		Brocklehurst 1997
103			50		1		Brocklehurst 1997
104			50		1		Brocklehurst 1997
105			50		1		Brocklehurst 1997
106			50		1		Brocklehurst 1997
107			50		1		Brocklehurst 1997
108			50		1		Brocklehurst 1997
109			50		1		Brocklehurst 1997
110			50		1		Brocklehurst 1997
111			50		1		Brocklehurst 1997
112			50		1		Brocklehurst 1997
113	Not Reported		50		1		Brocklehurst 1997
114	Not Reported		50		1		Brocklehurst 1997
115				2		No	Dathe 1968
116	Enteritis	110		2		No	Dathe 1968
117				2		No	Dathe 1968
118	Eaten by dam	7		2		No	Dathe 1968
119	Pneumonia	7		2		No	Dathe 1968
120			4	2	1	No	Frese 1980
121			4	2	1	No	Frese 1980
122			4	2	1	No	Frese 1980
123			9	2	1	Yes	Gowda 1967
124			9	2	1	No	Gowda 1967
125	Eaten by dam	0		3		Yes	Kralik 1967
126	Eaten by dam	0		3		Yes	Kralik 1967
127				1		Yes	Cade 1968
128	Eaten by dam	0					Hemmer 1976
129							Hemmer 1976
130	Eaten by dam	0					Hemmer 1976
131							Hemmer 1976
132	Eaten by dam	0					Hemmer 1976

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	A	B	C	K	L	M	N	O	P	Q	R
	Family	Genus	Species	Zoo	Dam	Origin	Rearing	Age	Litter size	Young dead	Infant Mortality
133	Felidae	Felis	<i>margarita</i>	Brookfield	Br2	Wild	Parental		4	0	0.00
134	Felidae	Felis	<i>margarita</i>	Brookfield	Br2	Wild	Parental		4	0	0.00
135	Felidae	Felis	<i>margarita</i>	Private	Sch3	Wild	Parental	3	4	2	0.50
136	Felidae	Felis	<i>margarita</i>	Private	Sch3	Wild	Parental	3	8	8	1.00
137	Felidae	Felis	<i>margarita</i>	Private	Sch3	Wild	Parental	4	4	4	1.00
138	Felidae	Felis	<i>margarita</i>	Wuppertal	Brout	Wild	Parental	5	1	0	0.00
139	Felidae	Felis	<i>nigripes</i>	Wuppertal	Brout	Wild	Parental	6	2	0	0.00
140	Felidae	Felis	<i>nigripes</i>	Wuppertal	Brout	Wild	Parental	2	2	2	1.00
141	Felidae	Felis	<i>nigripes</i>	Wuppertal	Buster	Captivity	Parental	3	1	1	1.00
142	Felidae	Felis	<i>nigripes</i>	WNZP	A						
143	Felidae	Felis	<i>nigripes</i>	WNZP	A			4	2	0	0.00
144	Felidae	Felis	<i>nigripes</i>	WNZP	A			5	2	2	1.00
145	Felidae	Felis	<i>nigripes</i>	WNZP	A			5	2	2	1.00
146	Felidae	Felis	<i>nigripes</i>	WNZP	A			6	1	1	1.00
147	Felidae	Felis	<i>pardalis</i>	Tulsa	A			2	1	0	0.00
148	Felidae	<i>Leptailurus</i>	<i>serval</i>	Mole Hall WP	A	Wild	Parental		3	1	0.33
149	Felidae	<i>Leptailurus</i>	<i>serval</i>	Mole Hall WP	A	Wild	Parental		2	0	0.00
150	Felidae	Felis	<i>silvestris</i>	Berne	Celine	Wild	Parental	2	5	0	0.00
151	Felidae	Felis	<i>silvestris</i>	Berne	Celine	Wild	Parental	3	4	1	0.25
152	Felidae	Felis	<i>silvestris</i>	Berne	Celine	Wild	Parental	4	8	2	0.25
153	Felidae	Felis	<i>silvestris</i>	Berne	Celine	Wild	Parental	5	6	2	0.33
154	Felidae	Felis	<i>silvestris</i>	Berne	Celine	Wild	Parental	7	6	0	0.00
155	Felidae	Felis	<i>silvestris</i>	Berne	Celine	Wild	Parental	8	5	0	0.00
156	Felidae	Felis	<i>silvestris</i>	Berne	Celine	Wild	Parental	1	2	0	0.00
157	Felidae	Felis	<i>silvestris</i>	Berne	Eliane	Wild	Parental	3	1	1	1.00
158	Felidae	Felis	<i>silvestris</i>	Berne	Eliane	Wild	Parental	4	3	0	0.00
159	Felidae	Felis	<i>silvestris</i>	Berne	Eliane	Wild	Parental	5	3	3	1.00
160	Felidae	Felis	<i>silvestris</i>	Berne	Eliane	Wild	Parental	6	1	0	0.00
161	Felidae	Felis	<i>silvestris</i>	Berne	Sabine	Wild	Parental	2	5	0	0.00
162	Felidae	Felis	<i>silvestris</i>	Berne	Sabine	Wild	Parental	3	6	0	0.00
163	Felidae	Felis	<i>silvestris</i>	Berne	Sabine	Wild	Parental	3	3	0	0.00
164	Felidae	Felis	<i>silvestris</i>	Berne	Tatra	Captivity		2	5	0	0.00
165	Felidae	Felis	<i>silvestris</i>	Berne	Tatra	Captivity		3	4	2	0.50
166	Felidae	Felis	<i>silvestris</i>	Prague	A	Wild	Parental	2	4	4	1.00
167	Felidae	Felis	<i>silvestris</i>	Prague	A	Wild	Parental	2	3	0	0.00
168	Felidae	Felis	<i>silvestris</i>	Prague	A	Wild	Parental	3	5	0	0.00
169	Felidae	Felis	<i>silvestris</i>	Prague	A	Wild	Parental	3	1	1	1.00
170	Felidae	Felis	<i>silvestris</i>	Prague	A	Wild	Parental	4	3	3	1.00
171	Felidae	Felis	<i>silvestris</i>	Prague	A	Wild	Parental	4	5	0	0.00
172	Felidae	Felis	<i>silvestris</i>	Prague	A	Wild	Parental	5	1	1	1.00
173	Felidae	Felis	<i>silvestris</i>	Prague	A	Wild	Parental	5	2	0	0.00
174	Felidae	Felis	<i>silvestris</i>	Aberdeen	A	Wild	Parental		3	0	0.00
175	Felidae	Felis	<i>silvestris</i>	Aberdeen	A	Wild	Parental		3	0	0.00
176	Felidae	Felis	<i>silvestris</i>	Aberdeen	B	Wild	Parental		4	0	0.00

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	S	T	U	V	W	X	Y
	Cause of death	Age of death(Days)	Cage area (m2)	Ind/cage	Dens	Male saparated?	Reference
133							Hemmer 1976
134							Hemmer 1976
135							Hemmer 1976
136	Not Reported	0					Hemmer 1976
137	Stillbirth	0					Hemmer 1976
138	Stillbirth	0					Hemmer 1976
139				1	2	Yes	Leyhausen & Tonkin 1966
140				1	2	Yes	Leyhausen & Tonkin 1966
141	Not Reported	66		1	2	Yes	Leyhausen & Tonkin 1966
142	Enteritis under Hand-rearing	3		2			Armstrong 1975
143				2			Armstrong 1975
144	Eaten by dam	8		2			Armstrong 1975
145	Panleucopenia	180		2			Armstrong 1975
146	Drowned	58		2			Armstrong 1975
147			20	2	2	Yes	Dunn 1974
148	Neglect	4		2		Yes	Johnstone 1977
149				2		Yes	Johnstone 1977
150				3	5	No	Meyer-Holzappel 1968
151	Stillbirth	0		3	5	No	Meyer-Holzappel 1968
152	Stillbirth	0		3	5	No	Meyer-Holzappel 1968
153	Stillbirth	0		3	5	No	Meyer-Holzappel 1968
154				3	5	No	Meyer-Holzappel 1968
155				3	5	No	Meyer-Holzappel 1968
156				3	5	No	Meyer-Holzappel 1968
157	Not Reported	0		3	5	No	Meyer-Holzappel 1968
158				3	5	No	Meyer-Holzappel 1968
159	Not Reported	2		3	5	No	Meyer-Holzappel 1968
160				3	5	No	Meyer-Holzappel 1968
161				3	5	No	Meyer-Holzappel 1968
162				3	5	No	Meyer-Holzappel 1968
163				3	5	No	Meyer-Holzappel 1968
164				2	5	No	Meyer-Holzappel 1968
165	Eaten by dam	0		2	5	No	Meyer-Holzappel 1968
166	Eaten by dam	0		2		No	Voif 1968
167				2		No	Voif 1968
168				2		No	Voif 1968
169	Not Reported	1		2		No	Voif 1968
170	Killed by dam	104		2		No	Voif 1968
171				2		No	Voif 1968
172	Not Reported	0		2		No	Voif 1968
173				2		No	Voif 1968
174			110	2	5	No	Leslie 1973
175			110	2	5	No	Leslie 1973
176			110	4	4	No	Leslie 1973

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	A	B	C	K	L	M	N	O	P	Q	R
	Family	Genus	Species	Zoo	Dam	Origin	Rearing	Age	Litter size	Young dead	Infant Mortality
177	Felidae	Felis	silvestris	Wuppertal	A	Wild	Parental		2	0	0.00
178	Felidae	Felis	silvestris	Wuppertal	B	Wild	Parental		4	1	0.25
179	Felidae	Felis	viverrina	Philadelphia	A	Wild	Parental	6	2	1	0.50
180	Felidae	Felis	viverrina	Philadelphia	A	Wild	Parental	8	2	0	0.00
181	Felidae	Felis	viverrina	Philadelphia	A	Wild	Parental	8	2	0	0.00
182	Felidae	Felis	viverrina	Sri Lanka NZ	A	Wild	Parental		2	1	0.50
183	Felidae	Herpailurus	yagouaroundi	Chester	A	Captivity			2	0	0.00
184	Felidae	Herpailurus	yagouaroundi	Private	A	Wild	Parental		1	1	1.00
185	Felidae	Herpailurus	yagouaroundi	Private	A	Wild	Parental		2	0	0.00
186	Felidae	Herpailurus	yagouaroundi	Private	A	Wild	Parental		4	0	0.00
187	Felidae	Herpailurus	yagouaroundi	Mainz	A	Wild	Parental	3	1	1	1.00
188	Felidae	Leopardus	geoffroyi	Mainz	B				3	2	0.67
189	Felidae	Leopardus	geoffroyi	Memphis	A			4	3	3	1.00
190	Felidae	Leopardus	geoffroyi	Memphis	A			4	3	0	0.00
191	Felidae	Leopardus	geoffroyi	Memphis	A			4	3	0	0.00
192	Felidae	Leopardus	geoffroyi	Memphis	A			4	3	0	0.00
193	Felidae	Leopardus	geoffroyi	Memphis	A			5	2	2	1.00
194	Felidae	Leopardus	geoffroyi	Memphis	A			5	2	0	0.00
195	Felidae	Leopardus	geoffroyi	Memphis	A			6	3	0	0.00
196	Felidae	Leopardus	geoffroyi	Memphis	A			6	2	0	0.00
197	Felidae	Leopardus	geoffroyi	Memphis	B			2	2	2	1.00
198	Felidae	Leopardus	geoffroyi	Memphis	B			2	3	3	1.00
199	Felidae	Leopardus	geoffroyi	Glasgow	A			5	2	2	1.00
200	Felidae	Leopardus	geoffroyi	Glasgow	A			6	1	0	0.00
201	Felidae	Leopardus	geoffroyi	Glasgow	A			7	2	2	1.00
202	Felidae	Leopardus	geoffroyi	Glasgow	A			7	2	0	0.00
203	Felidae	Leopardus	tigrinus	Private	A				1	1	1.00
204	Felidae	Leopardus	tigrinus	Private	A				2	0	0.00
205	Felidae	Leopardus	tigrinus	Private	A				2	2	1.00
206	Felidae	Leopardus	tigrinus	Private	A	Wild	Parental		1	0	0.00
207	Felidae	Leopardus	tigrinus	Private	A	Wild	Parental		1	0	0.00
208	Felidae	Leopardus	tigrinus	Private	A	Wild	Parental		1	0	0.00
209	Felidae	Leopardus	tigrinus	Private	B	Captivity	Hand	4	1	0	0.00
210	Felidae	Leopardus	wiedii	New Orleans	A	Wild	Parental	4	2	0	0.00
211	Felidae	Leopardus	wiedii	RTEC Hastings	A	Wild	Parental	3	1	1	1.00
212	Felidae	Leopardus	wiedii	RTEC Hastings	A	Wild	Parental	4	1	0	0.00
213	Felidae	Lynx	lynx	Magdeburg	Anna	Wild	Parental	5	3	3	1.00
214	Felidae	Lynx	lynx	Magdeburg	Anna	Wild	Parental	6	3	3	1.00
215	Felidae	Lynx	lynx	Magdeburg	Anna	Wild	Parental	8	3	0	0.00
216	Felidae	Lynx	lynx	Magdeburg	Cora	Captivity		3	2	0	0.00
217	Felidae	Lynx	lynx	Norfolk WVP	A	Wild	Parental	2	1	0	0.00
218	Felidae	Lynx	lynx	Ostrava	A	Wild	Parental		1	1	1.00
219	Felidae	Lynx	lynx	Ostrava	B	Wild	Parental		1	1	1.00
220	Felidae	Lynx	lynx	Ostrava	C	Wild	Parental		3	0	0.00

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	S	T	U	V	W	X	Y
	Cause of death	Age of death(Days)	Cage area (m2)	Ind/cage	Dens	Male separated?	Reference
177				3	0	Yes	Tonkin & Kohler 1981
178				3	0	Yes	Tonkin & Kohler 1981
179	Not Reported	2		1	1	Yes	Ulmer 1968
180	Eaten by dam	0		1	1	Yes	Ulmer 1968
181				1	1	Yes	Ulmer 1968
182				2	1	Yes	Jayewardene 1975
183	Pneumonia	7	182	1	4	Yes	Hulley 1976
184			20	1		No	Hulley 1976
185	Panleucopenia	90	5000			No	Hulley 1976
186			5000			No	Hulley 1976
187			5000			No	Hulley 1976
188	Eaten by dam	2	4	2	0		Scheffel & Hemmer 1975
189	Killed by dam	1	2	1	1		Scheffel & Hemmer 1975
190	Eaten by dam	12	2	2	0	Yes	Anderson 1977
191			6	2	2	Yes	Anderson 1977
192			6	2	2	Yes	Anderson 1977
193	Eaten by dam	0	6	2	2	Yes	Anderson 1977
194			6	2	2	Yes	Anderson 1977
195			6	2	2	Yes	Anderson 1977
196			6	3	2	Yes	Anderson 1977
197	Eaten by dam	0	6	3	2	Yes	Anderson 1977
198	Eaten by dam	0	6	3	2	Yes	Anderson 1977
199	Breech birth	0	10	2	5	Yes	Law & Boyle 1984
200			10	2	5	Yes	Law & Boyle 1984
201	Eaten by dam	0	10	2	5	Yes	Law & Boyle 1984
202			10	2	5	Yes	Law & Boyle 1984
203	Not Reported	3	200	6		No	Leyhausen & Tonkin 1966
204			200	6		No	Leyhausen & Tonkin 1966
205	Eaten by dam	0	200	6		No	Leyhausen & Tonkin 1966
206				2			Quillen 1981
207				2			Quillen 1981
208				2			Quillen 1981
209				2			Quillen 1981
210			17	2	1	Yes	Paintiff & Anderson 1980
211	Eaten by dam	50	40	2	4		Mansard 1997
212			40	2	4		Mansard 1997
213	Cat distemper	180	50	3	2		Burger 1966
214	Cat distemper	133	50	3	2		Burger 1966
215			50	3	2		Burger 1966
216			50	3	2		Burger 1966
217			20	2		Yes	Wayre 1969
218	Stillbirth	0		4		Yes	Kunc 1970
219	Stillbirth	0		4		Yes	Kunc 1970
220				4		Yes	Kunc 1970

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	A	B	C	K	L	M	N	O	P	Q	R
	Family	Genus	Species	Zoo	Dam	Origin	Rearing	Age	Litter size	Young dead	Infant Mortality
221	Felidae	Lynx	lynx	Ostrava	D	Wild	Parental		3	0	0.00
222	Felidae	Lynx	lynx	Ostrava	D	Wild	Parental		3	0	0.00
223	Felidae	Neofelis	nebulosa	Frankfurt	A	Captivity			3	0	0.00
224	Felidae	Neofelis	nebulosa	Frankfurt	A	Captivity			2	0	0.00
225	Felidae	Neofelis	nebulosa	Dallas	A			7	4	3	0.75
226	Felidae	Neofelis	nebulosa	Dresden	A			4	2	0	0.00
227	Felidae	Neofelis	nebulosa	Dublin	Rita	Wild	Parental	3	1	1	1.00
228	Felidae	Neofelis	nebulosa	Dublin	Rita	Wild	Parental	4	2	2	1.00
229	Felidae	Neofelis	nebulosa	Dublin	Rita	Wild	Parental	4	2	2	1.00
230	Felidae	Neofelis	nebulosa	Dublin	Rita	Wild	Parental	4	2	2	1.00
231	Felidae	Neofelis	nebulosa	Dublin	Rita	Wild	Parental	5	2	0	0.00
232	Felidae	Neofelis	nebulosa	Dublin	Rita	Wild	Parental	6	2	0	0.00
233	Felidae	Neofelis	nebulosa	Dublin	Sita	Wild	Parental	2	2	0	0.00
234	Felidae	Neofelis	nebulosa	Dublin	Sita	Wild	Parental	4	2	2	1.00
235	Felidae	Neofelis	nebulosa	Dublin	Sita	Wild	Parental	4	2	2	1.00
236	Felidae	Neofelis	nebulosa	Dublin	Sita	Wild	Parental	5	1	1	1.00
237	Felidae	Neofelis	nebulosa	Dublin	Sita	Wild	Parental	5	2	0	0.00
238	Felidae	Neofelis	nebulosa	Dublin	Sita	Wild	Parental	6	2	1	0.50
239	Felidae	Panthera	onca	Topeka	A			5	2	1	0.50
240	Felidae	Panthera	onca	Ostrava	A	Captivity		3	2	0	0.00
241	Felidae	Panthera	onca	Ostrava	A	Captivity		4	2	0	0.00
242	Felidae	Panthera	onca	Ostrava	B			3	1	0	0.00
243	Felidae	Panthera	pardus	Prague	A				1	0	0.00
244	Felidae	Panthera	tigris	London RP	A				3	3	1.00
245	Felidae	Panthera	tigns	London RP	A				2	2	1.00
246	Felidae	Panthera	tigris	London RP	A				3	3	1.00
247	Felidae	Panthera	tigris	London RP	A				2	2	1.00
248	Felidae	Panthera	tigris	London RP	A				3	3	1.00
249	Felidae	Panthera	tigris	London RP	A				3	3	1.00
250	Felidae	Panthera	tigris	London RP	A				3	3	1.00
251	Felidae	Panthera	tigris	Mysore	Rani				4	4	1.00
252	Felidae	Panthera	tigris	Whipsnade	A	Wild	Parental	8	4	3	0.75
253	Felidae	Uncia	uncia	St Louis	A	Captivity		6	4	3	0.75
254	Felidae	Uncia	uncia	St Louis	A	Captivity		7	3	2	0.67
255	Felidae	Uncia	uncia	St Louis	A	Captivity		8	4	2	0.50
256	Felidae	Uncia	uncia	Kaunas	Ramuna	Wild	Parental	3	2	0	0.00
257	Felidae	Uncia	uncia	Kaunas	Ramuna	Wild	Parental	4	2	2	1.00
258	Felidae	Uncia	uncia	Kaunas	Ramuna	Wild	Parental	8	4	1	0.25
259	Felidae	Uncia	uncia	Kaunas	Ruta	Wild	Parental	7	3	1	0.33
260	Felidae	Uncia	uncia	Kaunas	Ruta	Wild	Parental	8	2	2	1.00
261	Felidae	Uncia	uncia	Kaunas	Ruta	Wild	Parental	10	1	1	1.00
262	Felidae	Uncia	uncia	Dallas	A	Wild	Parental	4	2	0	0.00
263	Felidae	Uncia	uncia	Seattle WP	A	Wild	Parental	5	1	0	0.00
264	Herpestidae	Galidia	elegans	WNZP		Wild	Parental	3	1	1	1.00

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	S	T	U	V	W	X	Y
	Cause of death	Age of death(Days)	Cage area (m2)	Ind/cage	Dens	Male separated?	Reference
221				4		Yes	Kunc 1970
222				4		Yes	Kunc 1970
223				2	1		Fellner 1965
224				2	1		Fellner 1965
225				2			Fontaine 1965
226	Killed by dam/Infection	1		2			Geidel & Gensch 1976
227			11	2	2	No	
228	Neglect	15	20	1	2	Yes	Murphy 1976
229	Eaten by dam	0	20	1	2	Yes	Murphy 1976
230	Neglect	7	20	1	2	Yes	Murphy 1976
231			6	1	1	Yes	Murphy 1976
232			6	1	1	Yes	Murphy 1976
233			20	1	2	Yes	Murphy 1976
234	Eaten by dam	0	20	1	2	Yes	Murphy 1976
235	Enteritis	2	20	1	2	Yes	Murphy 1976
236	Neglect	1	20	1	2	Yes	Murphy 1976
237			6	1	1	Yes	Murphy 1976
238	Eaten by dam	28	20	1	2	Yes	Murphy 1976
239	Eaten by dam	0				Yes	Hunt 1967
240				1		Yes	Stehlik 1971
241				1		Yes	Stehlik 1971
242				1		Yes	Stehlik 1971
243				2			Dobroruka 1968
244	Killed by dam			1		Yes	Sadleir 1966
245	Killed by dam			1		Yes	Sadleir 1966
246	Killed by dam			1		Yes	Sadleir 1966
247	Stillbirth			1		Yes	Sadleir 1966
248	Stillbirth			1		Yes	Sadleir 1966
249	Killed by dam			1		Yes	Sadleir 1966
250	Killed by dam			1		Yes	Sadleir 1966
251	Breech birth	0		4			Gowda 1968
252	Neglect	0		2		Yes	Hughes 1977
253	Congenital defect	15		2		No	Frueh 1968
254	Pneumonia	6		2		No	Frueh 1968
255	Eaten by dam/pneumonia	6		2		No	Frueh 1968
256			40	2	1	Yes	Marma & Yunchis 1968
257	Not Reported	0	40	2	1	Yes	Marma & Yunchis 1968
258	Not Reported	2	40	2	1	Yes	Marma & Yunchis 1968
259	Not Reported	0	40	2	1	Yes	Marma & Yunchis 1968
260	Premature birth	0	40	2	1	Yes	Marma & Yunchis 1968
261	Eaten by dam	0	40	2	1	Yes	Marma & Yunchis 1968
262				2		Yes	Calvin 1969
263			110	2	1	Yes	Freeman 1975
264	Neglect	1		4		No	Larkin & Roberts 1979

	A	B	C	K	L	M	N	O	P	Q	R
	Family	Genus	Species	Zoo	Dam	Origin	Rearing	Age	Litter size	Young dead	Infant Mortality
265	Herpestidae	<i>Galidia</i>	<i>elegans</i>	WNZP		Wild	Parental	4	1	1	1 00
266	Herpestidae	<i>Galidia</i>	<i>elegans</i>	WNZP		Wild	Parental	4	1	0	0 00
267	Herpestidae	<i>Helogale</i>	<i>parvula</i>	Basle	A	Captivity			4	2	0 50
268	Herpestidae	<i>Helogale</i>	<i>parvula</i>	Basle	A	Captivity			4	4	1 00
269	Herpestidae	<i>Helogale</i>	<i>parvula</i>	Basle	A	Captivity			1	0	0 00
270	Herpestidae	<i>Helogale</i>	<i>parvula</i>	Basle	A	Captivity			4	0	0 00
271	Herpestidae	<i>Helogale</i>	<i>parvula</i>	Basle	A	Captivity			4	4	1 00
272	Herpestidae	<i>Helogale</i>	<i>parvula</i>	Basle	A	Captivity			4	4	1 00
273	Herpestidae	<i>Helogale</i>	<i>parvula</i>	Basle	A	Captivity			4	4	1 00
274	Herpestidae	<i>Helogale</i>	<i>parvula</i>	Basle	A	Captivity			4	4	1 00
275	Herpestidae	<i>Helogale</i>	<i>parvula</i>	Basle	A	Captivity			5	0	0 00
276	Herpestidae	<i>Helogale</i>	<i>parvula</i>	London RP	A	Captivity		2	4	0	0 00
277	Herpestidae	<i>Helogale</i>	<i>parvula</i>	London RP	A	Captivity		2	2	0	0 00
278	Herpestidae	<i>Helogale</i>	<i>parvula</i>	London RP	A	Captivity		4	2	2	1 00
279	Herpestidae	<i>Helogale</i>	<i>parvula</i>	London RP	A	Captivity		4	5	0	0 00
280	Herpestidae	<i>Helogale</i>	<i>parvula</i>	London RP	A	Captivity		4	4	0	0 00
281	Herpestidae	<i>Helogale</i>	<i>parvula</i>	London RP	B	Captivity		2	4	0	0 00
282	Hyaenidae	<i>Crocuta</i>	<i>crocuta</i>	Ibadan, Nigeria	A	Wild	Parental	7	2	0	0 00
283	Hyaenidae	<i>Crocuta</i>	<i>crocuta</i>	Ibadan, Nigeria	A	Wild	Parental	8	1	0	0 00
284	Hyaenidae	<i>Crocuta</i>	<i>crocuta</i>	Malton	A				2	1	0 50
285	Hyaenidae	<i>Crocuta</i>	<i>crocuta</i>	Denver	A				1	0	0 00
286	Hyaenidae	<i>Parahyaena</i>	<i>brunnea</i>	Okahandja	A	Wild	Parental	4	3	3	1 00
287	Hyaenidae	<i>Parahyaena</i>	<i>brunnea</i>	Okahandja	A	Wild	Parental	7	4	1	0 25
288	Hyaenidae	<i>Parahyaena</i>	<i>brunnea</i>	Okahandja	A	Wild	Parental	7	4	0	0 00
289	Mustelidae	<i>Amblyonyx</i>	<i>cinerea</i>	Aberdeen	Ovaltine			3	1	1	1 00
290	Mustelidae	<i>Amblyonyx</i>	<i>cinerea</i>	Aberdeen	Ovaltine			4	1	1	1 00
291	Mustelidae	<i>Amblyonyx</i>	<i>cinerea</i>	Aberdeen	Ovaltine			4	1	1	1 00
292	Mustelidae	<i>Amblyonyx</i>	<i>cinerea</i>	Aberdeen	Ovaltine			5	2	0	0 00
293	Mustelidae	<i>Amblyonyx</i>	<i>cinerea</i>	Aberdeen	Ovaltine			6	3	3	1 00
294	Mustelidae	<i>Amblyonyx</i>	<i>cinerea</i>	Chester	A	Wild	Parental	2	6	6	1 00
295	Mustelidae	<i>Amblyonyx</i>	<i>cinerea</i>	Chester	A	Wild	Parental	3	5	4	0 80
296	Mustelidae	<i>Amblyonyx</i>	<i>cinerea</i>	WNZP	55	Captivity		8	1	1	1 00
297	Mustelidae	<i>Amblyonyx</i>	<i>cinerea</i>	WNZP	55	Captivity		11	2	2	1 00
298	Mustelidae	<i>Amblyonyx</i>	<i>cinerea</i>	WNZP	55	Captivity		12	3	1	0 33
299	Mustelidae	<i>Amblyonyx</i>	<i>cinerea</i>	WNZP	55	Captivity		12	3	1	0 33
300	Mustelidae	<i>Arconyx</i>	<i>collaris</i>	Toronto Metro	A	Wild	Parental	2	2	1	0 50
301	Mustelidae	<i>Eira</i>	<i>barbara</i>	Krefeld	A			4	3	1	0 33
302	Mustelidae	<i>Eira</i>	<i>barbara</i>	Lincoln	A	Wild	Parental	3	2	0	0 00
303	Mustelidae	<i>Eira</i>	<i>barbara</i>	Lincoln	A	Wild	Parental	4	2	0	0 00
304	Mustelidae	<i>Enhydra</i>	<i>lutris</i>	La Jolla	A	Wild	Parental		1	1	1 00
305	Mustelidae	<i>Enhydra</i>	<i>lutris</i>	Sea World	A	Wild	Parental		1	1	1 00
306	Mustelidae	<i>Enhydra</i>	<i>lutris</i>	Seattle Aq	A				1	1	1 00
307	Mustelidae	<i>Enhydra</i>	<i>lutris</i>	Vancouver Aq	A				1	1	1 00
308	Mustelidae	<i>Enhydra</i>	<i>lutris</i>	Sea World	B	Wild	Parental		1	1	1 00

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	S	T	U	V	W	X	Y
	Cause of death	Age of death(Days)	Cage area (m2)	Ind/cage	Dens	Male separated?	Reference
265	Neglect	2		4		Yes	Larkin & Roberts 1979
266				4		Yes	Larkin & Roberts 1979
267						No	Ruedi 1984
268	Not Reported		16	2	10	No	Ruedi 1984
269	Eaten by dam		16	2	10	No	Ruedi 1984
270			16	2	10	No	Ruedi 1984
271			16	3	10	No	Ruedi 1984
272	Eaten by dam		16	7	10	No	Ruedi 1984
273	Eaten by dam		16	7	10	No	Ruedi 1984
274	Eaten by dam		16	7	10	No	Ruedi 1984
275			16	7	10	No	Ruedi 1984
276			14.5	8	1	No	Christie & Wheeler 1990
277			14.5	8	1	No	Christie & Wheeler 1990
278	Not Reported	7	14.5	8	1	No	Christie & Wheeler 1990
279			14.5	8	1	No	Christie & Wheeler 1990
280			14.5	8	1	No	Christie & Wheeler 1990
281			14.5	8	1	No	Christie & Wheeler 1990
282			7	2	1	Yes	Golding 1969
283			7	2	1	Yes	Golding 1969
284	Stillbirth	0	60	2	1	No	Reitz 1972
285			40	1	0	Yes	Kinsey & Kreider 1990
286	Eaten by dam	21					Schulz 1966
287	Underdevelopment	15					Schulz 1966
288							Schulz 1966
289	Neglect	7	20	2	1	No	Leslie 1970
290	Eaten by dam	0	20	2	1	No	Leslie 1970
291	Neglect	0	20	2	1	No	Leslie 1970
292			20	2	1	No	Leslie 1970
293	Neglect	1	100	2	1	No	Leslie 1970
294	Neglect	10	150	9	8	No	Timmis 1971
295	Neglect	2	6	1	1	Yes	Timmis 1971
296	Not Reported	49	120	3	2	No	Prima 1992
297	Inanition	96	120	3	2	No	Prima 1992
298	Inanition	100	120	3	2	No	Prima 1992
299	Not Reported		120	3	2	No	Prima 1992
300	Killed by others	3	13	2	2	No	Parker 1979
301	Enteritis	30	50	2	3	No	Encke 1968
302			8	2	1	Yes	Vaughn 1974
303			8	2	1	Yes	Vaughn 1974
304	Pneumonia	43					Antrim & Cornell 1980
305	Stillbirth	0					Antrim & Cornell 1980
306	Not Reported	2					Antrim & Cornell 1980
307	Stillbirth	0				No	Antrim & Cornell 1980
308	Neglect	1					Antrim & Cornell 1980

	A	B	C	K	L	M	N	O	P	Q	R
	Family	Genus	Species	Zoo	Dam	Origin	Rearing	Age	Litter size	Young dead	Infant Mortality
309	Mustelidae	<i>Enhydra</i>	<i>lutris</i>	Sea World		Wild	Parental		1	1	1 00
310	Mustelidae	<i>Enhydra</i>	<i>lutris</i>	Sea World		Wild	Parental		1	1	1 00
311	Mustelidae	<i>Enhydra</i>	<i>lutris</i>	Sea World		Wild	Parental		1	1	1 00
312	Mustelidae	<i>Enhydra</i>	<i>lutris</i>	Sea World		Wild	Parental		1	1	1 00
313	Mustelidae	<i>Enhydra</i>	<i>lutris</i>	Sea World		Wild	Parental		1	1	1 00
314	Mustelidae	<i>Gulo</i>	<i>gulo</i>	Colorado	A	Wild	Parental	2	2	0	0 00
315	Mustelidae	<i>Lutra</i>	<i>canadensis</i>	Norfolk WP	A	Wild	Parental	4	3	0	0 00
316	Mustelidae	<i>Lutra</i>	<i>canadensis</i>	Private	A	Wild	Parental		2	0	0 00
317	Mustelidae	<i>Lutra</i>	<i>canadensis</i>	Private	B	Captivity		4	5	1	0 20
318	Mustelidae	<i>Lutra</i>	<i>lutra</i>	Norfolk WP	A	Wild	Parental	3	2	0	0 00
319	Mustelidae	<i>Lutra</i>	<i>lutra</i>	Norfolk WP	B	Wild	Parental	3	1	0	0 00
320	Mustelidae	<i>Lutra</i>	<i>lutra</i>	Krefeld	A	Captivity		2	2	0	0 00
321	Mustelidae	<i>Lutra</i>	<i>lutra</i>	Krefeld	A	Captivity		3	4	0	0 00
322	Mustelidae	<i>Lutra</i>	<i>lutra</i>	Krefeld	A	Captivity		5	3	0	0 00
323	Mustelidae	<i>Lutra</i>	<i>lutra</i>	Krefeld	A	Captivity		7	3	0	0 00
324	Mustelidae	<i>Lutra</i>	<i>lutra</i>	Krefeld	B	Captivity	Parental	3	2	0	0 00
325	Mustelidae	<i>Lutra</i>	<i>perspicillata</i>	Jaipur	A	Captivity		4	2	0	0 00
326	Mustelidae	<i>Lutra</i>	<i>perspicillata</i>	Jaipur	A	Captivity		6	1	0	0 00
327	Mustelidae	<i>Lutra</i>	<i>perspicillata</i>	Jaipur	B	Captivity	Parental	4	2	0	0 00
328	Mustelidae	<i>Lutra</i>	<i>perspicillata</i>	Twycross	A				1	0	0 00
329	Mustelidae	<i>Lutra</i>	<i>perspicillata</i>	Delhi	Moti			7	5	5	1 00
330	Mustelidae	<i>Lutra</i>	<i>perspicillata</i>	Delhi	Radha			2	3	0	0 00
331	Mustelidae	<i>Lutra</i>	<i>perspicillata</i>	Delhi	Sonya			2	2	0	0 00
332	Mustelidae	<i>Lutra</i>	<i>perspicillata</i>	Delhi	Vimla			4	3	0	0 00
333	Mustelidae	<i>Meles</i>	<i>meles</i>	Wrocław	A	Wild	Parental	6	1	0	0 00
334	Mustelidae	<i>Meles</i>	<i>meles</i>	Wrocław	A	Wild	Parental	7	2	0	0 00
335	Mustelidae	<i>Mellivora</i>	<i>capensis</i>	Bekesbourne	A				1	0	0 00
336	Mustelidae	<i>Mellivora</i>	<i>capensis</i>	Bekesbourne	B				1	0	0 00
337	Mustelidae	<i>Mustela</i>	<i>nigripes</i>	Patuxent WRC	4	Wild	Parental	4	5	5	1 00
338	Mustelidae	<i>Mustela</i>	<i>nigripes</i>	Patuxent WRC	4	Wild	Parental	5	5	5	1 00
339	Mustelidae	<i>Pteronura</i>	<i>brasiliensis</i>	Hamburg	Alte			3	3	3	1 00
340	Mustelidae	<i>Pteronura</i>	<i>brasiliensis</i>	Hamburg	Alte			3	2	2	1 00
341	Mustelidae	<i>Pteronura</i>	<i>brasiliensis</i>	Hamburg	Alte			3	1	1	1 00
342	Mustelidae	<i>Pteronura</i>	<i>brasiliensis</i>	Hamburg	Alte			4	2	2	1 00
343	Mustelidae	<i>Pteronura</i>	<i>brasiliensis</i>	Hamburg	Alte			6	4	4	1 00
344	Mustelidae	<i>Pteronura</i>	<i>brasiliensis</i>	Hamburg	Alte			6	2	2	1 00
345	Mustelidae	<i>Pteronura</i>	<i>brasiliensis</i>	Hamburg	Alte			8	2	2	1 00
346	Mustelidae	<i>Pteronura</i>	<i>brasiliensis</i>	Hamburg	Ottlie			2	2	2	1 00
347	Mustelidae	<i>Pteronura</i>	<i>brasiliensis</i>	Hamburg	Ottlie			3	2	2	1 00
348	Mustelidae	<i>Pteronura</i>	<i>brasiliensis</i>	Hamburg	Ottlie			4	4	4	1 00
349	Mustelidae	<i>Pteronura</i>	<i>brasiliensis</i>	Hamburg	Ottlie			5	4	1	0 25
350	Mustelidae	<i>Pteronura</i>	<i>brasiliensis</i>	Hamburg	Ottlie			5	5	1	0 20
351	Procyonidae	<i>Ailurus</i>	<i>fulgens</i>	WNZP	A	Wild	Parental	4	2	2	1 00
352	Procyonidae	<i>Ailurus</i>	<i>fulgens</i>	WNZP	A	Wild	Parental	9	1	1	1 00

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	S	T	U	V	W	X	Y
	Cause of death	Age of death(Days)	Cage area (m2)	Indicage	Dens	Male separated?	Reference
309							
310	Neglect	3					Antrim & Cornell 1980
311	Stillbirth	0					Antrim & Cornell 1980
312	Neglect	10					Antrim & Cornell 1980
313	Stillbirth	0					Antrim & Cornell 1980
314				2			Davis 1967
315			28	2	1	Yes	Wayre 1967
316				2	1	Yes	Harris 1969
317	Drowned	78		2	1	Yes	Harris 1969
318			50	1	4	Yes	Wayre 1972
319			50	1	4	Yes	Wayre 1972
320			250	2	5	No	Vogt 1987
321			250	2	5	No	Vogt 1987
322			250	2	5	No	Vogt 1987
323			250	3	5	No	Vogt 1987
324			250	3	5	No	Vogt 1987
325			40	2		No	Yadav 1967
326			40	2		No	Yadav 1967
327			40	2		No	Yadav 1967
328			315	2	1	No	Badham 1973
329	Neglect	0	534	5	5	No	Desai 1974
330			534	5	5	No	Desai 1974
331			534	5	5	No	Desai 1974
332			534	5	5	No	Desai 1974
333			12	2	2	No	Gucwinska & Gucwinski 1968
334			12	2	2	No	Gucwinska & Gucwinski 1968
335				3	0	No	Johnstone-Scott 1975
336				3	5	Yes	Johnstone-Scott 1975
337	Eaten by dam	3	2	1	2	Yes	Hillman & Carpenter 1984
338	Neglect/Hand-rearing	2	2	1	2	Yes	Hillman & Carpenter 1984
339	Not Reported	4	180	5	5	No	Hagenbeck & Wunnemann 1992
340	Not Reported	1	180	5	5	No	Hagenbeck & Wunnemann 1992
341	Not Reported	13	180	5	5	No	Hagenbeck & Wunnemann 1992
342	Not Reported	2	180	5	5	No	Hagenbeck & Wunnemann 1992
343	Not Reported	3	180	5	5	No	Hagenbeck & Wunnemann 1992
344	Stillbirth	0	180	5	5	No	Hagenbeck & Wunnemann 1992
345	Stillbirth	0	180	5	5	No	Hagenbeck & Wunnemann 1992
346	Not Reported	0	180	5	5	No	Hagenbeck & Wunnemann 1992
347	Not Reported	3	180	5	5	No	Hagenbeck & Wunnemann 1992
348	Neglect	4	180	5	5	No	Hagenbeck & Wunnemann 1992
349	Stillbirth	0	180	5	5	No	Hagenbeck & Wunnemann 1992
350	Stillbirth	0	180	5	5	Yes	Hagenbeck & Wunnemann 1992
351	Unknown	120	45	2			Gray 1970
352	Pneumonia/Hand-rearing	40	45	2		Yes	Gray 1970

	A	B	C	K	L	M	N	O	P	Q	R
	Family	Genus	Species	Zoo	Dam	Origin	Rearing	Age	Litter size	Young dead	Infant Mortality
353	Procyonidae	<i>Ailurus</i>	<i>fulgens</i>	WNZP	A			3	2	0	0.00
354	Procyonidae	<i>Ailurus</i>	<i>fulgens</i>	WNZP	A			4	2	0	0.00
355	Procyonidae	<i>Ailurus</i>	<i>fulgens</i>	Krefeld	A	Wild	Parental	4	2	2	1.00
356	Procyonidae	<i>Ailurus</i>	<i>fulgens</i>	Krefeld	A	Wild	Parental	6	2	2	1.00
357	Procyonidae	<i>Ailurus</i>	<i>fulgens</i>	Krefeld	A	Wild	Parental	11	1	0	0.00
358	Procyonidae	<i>Ailurus</i>	<i>fulgens</i>	Front Royal	A	Captivity		2	2	0	0.00
359	Procyonidae	<i>Ailurus</i>	<i>fulgens</i>	Front Royal	B			2	2	1	0.50
360	Procyonidae	<i>Ailurus</i>	<i>fulgens</i>	ASDM USA	S1	Captivity			6	6	1.00
361	Procyonidae	<i>Nasua</i>	<i>narica</i>	ASDM USA	S2	Captivity			6	6	1.00
362	Procyonidae	<i>Nasua</i>	<i>narica</i>	ASDM USA	S3	Captivity			4	4	1.00
363	Procyonidae	<i>Nasua</i>	<i>narica</i>	ASDM USA	S4	Captivity			6	3	0.50
364	Procyonidae	<i>Nasua</i>	<i>narica</i>	Santa Barbara US	A			2	5	0	0.00
365	Procyonidae	<i>Nasua</i>	<i>narica</i>	Santa Barbara US	B			2	5	0	0.00
366	Procyonidae	<i>Potos</i>	<i>flavus</i>	Syracuse	A				1	0	0.00
367	Procyonidae	<i>Potos</i>	<i>flavus</i>	Delhi	A			3	1	1	1.00
368	Procyonidae	<i>Potos</i>	<i>flavus</i>	Delhi	A			7	1	0	0.00
369	Procyonidae	<i>Potos</i>	<i>flavus</i>	Barquisimeto	A				1	0	0.00
370	Procyonidae	<i>Potos</i>	<i>flavus</i>	Barquisimeto	A				1	0	0.00
371	Procyonidae	<i>Potos</i>	<i>flavus</i>	Barquisimeto	A				1	0	0.00
372	Procyonidae	<i>Potos</i>	<i>flavus</i>	Barquisimeto	A				1	0	0.00
373	Ursidae	<i>Ailuropoda</i>	<i>melanoleuca</i>	Xian	Dan Dan	Wild	Parental	2	2	2	1.00
374	Ursidae	<i>Ailuropoda</i>	<i>melanoleuca</i>	Xian	Dan Dan	Wild	Parental	5	1	0	0.00
375	Ursidae	<i>Ailuropoda</i>	<i>melanoleuca</i>	Chengdu	Lin Jin				2	2	1.00
376	Ursidae	<i>Ailuropoda</i>	<i>melanoleuca</i>	WNZP	Ling Ling				1	1	1.00
377	Ursidae	<i>Ailuropoda</i>	<i>melanoleuca</i>	Chengdu	Mei Mei				1	1	1.00
378	Ursidae	<i>Ailuropoda</i>	<i>melanoleuca</i>	Chengdu	Qeng Qeng	Captivity		7	1	0	0.00
379	Ursidae	<i>Ailuropoda</i>	<i>melanoleuca</i>	Fuzhon	Qing Qing				1	0	0.00
380	Ursidae	<i>Ailuropoda</i>	<i>melanoleuca</i>	Fuzhon	Qing Qing				2	2	1.00
381	Ursidae	<i>Ailuropoda</i>	<i>melanoleuca</i>	Fuzhon	Qing Qing				1	0	0.00
382	Ursidae	<i>Ailuropoda</i>	<i>melanoleuca</i>	Wolong	A	Wild	Parental	11	1	0	0.00
383	Ursidae	<i>Ailuropoda</i>	<i>melanoleuca</i>	Wolong	A	Wild	Parental	10	2	2	1.00
384	Ursidae	<i>Helarctos</i>	<i>malayanus</i>	Berlin TP	Tschita				1	0	0.00
385	Ursidae	<i>Helarctos</i>	<i>malayanus</i>	Berlin TP	Tschita				1	0	0.00
386	Ursidae	<i>Helarctos</i>	<i>malayanus</i>	Melbourne	A				1	0	0.00
387	Ursidae	<i>Helarctos</i>	<i>malayanus</i>	Berlin TP	Bonzo	Captivity			1	0	0.00
388	Ursidae	<i>Helarctos</i>	<i>malayanus</i>	Berlin TP	Evi	Captivity	Hand	8	1	1	1.00
389	Ursidae	<i>Helarctos</i>	<i>malayanus</i>	Berlin TP	Tschita				1	0	0.00
390	Ursidae	<i>Helarctos</i>	<i>malayanus</i>	Berlin TP	Tschita				1	0	0.00
391	Ursidae	<i>Helarctos</i>	<i>malayanus</i>	Berlin TP	Tschita				1	0	0.00
392	Ursidae	<i>Helarctos</i>	<i>malayanus</i>	Berlin TP	Tschita				1	0	0.00
393	Ursidae	<i>Helarctos</i>	<i>malayanus</i>	Forth Worth	A			6	1	0	0.00
394	Ursidae	<i>Helarctos</i>	<i>malayanus</i>	Forth Worth	A			7	1	0	0.00
395	Ursidae	<i>Helarctos</i>	<i>malayanus</i>	Forth Worth	A			7	1	0	0.00
396	Ursidae	<i>Ursus</i>	<i>maritimus</i>	St Paul	A			18	2	1	0.50

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	S	T	U	V	W	X	Y
	Cause of death	Age of death(Days)	Cage area (m2)	Ind/cage	Dens	Male separated?	Reference
353			65	2	2		Roberts 1975
354			65	2	2		Roberts 1975
355			40	2	5	Yes	Vogt et al 1980
356	Eaten by dam	3	40	2	5	Yes	Vogt et al 1980
357	Eaten by dam	0	40	2	5	Yes	Vogt et al 1980
358			40	2	5		Conway 1981
359			22	2	2		Conway 1981
360	Not Reported	4	22	2	2		Smith 1980
361	Killed by dam	0	96	4	1	Yes	Smith 1980
362	Killed by dam	0	2	4	0	Yes	Smith 1980
363	Killed by dam	0	96	4	1	Yes	Smith 1980
364	Stillbirth	0	96	4	1	Yes	Smith 1980
365			11	2	1	Yes	McToldridge 1969
366			11	2	1	Yes	McToldridge 1969
367			5	2	0	No	Clift 1967
368	Not Reported	0		2	1	No	Bhatia & Desai 1972
369				2	1	No	Bhatia & Desai 1972
370							Pernalette 1997
371							Pernalette 1997
372			11	2	1	No	Pernalette 1997
373	Neglect	3				Yes	Bingxing 1990
374						Yes	Bingxing 1990
375	Killed by dam/Killed by others	12				Yes	Bingxing 1990
376	Not Reported	1					Bingxing 1990
377	Killed by dam	15				Yes	Bingxing 1990
378						Yes	Bingxing 1990
379						Yes	Bingxing 1990
380	Not Reported	2				Yes	Bingxing 1990
381						Yes	Bingxing 1990
382				1		Yes	Zhang et al 2000
383	Disease while Hand-rearing	9		1		Yes	Zhang et al 2000
384				2		Yes	Dathe 1961
385				2		Yes	Dathe 1961
386							Weber 1969
387				2		Yes	Dathe 1970
388	Peritonitis/Hand-rearing	7		2		Yes	Dathe 1970
389				2		Yes	Dathe 1970
390				2		Yes	Dathe 1970
391				2		Yes	Dathe 1970
392				2		Yes	Dathe 1970
393			22	2	2	Yes	McCusker 1974
394			22	2	2	Yes	McCusker 1974
395			22	2	2	Yes	McCusker 1974
396	Neglect	2				Yes	Hess 1971

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	A	B	C	K	L	M	N	O	P	Q	R
	Family	Genus	Species	Zoo	Dam	Origin	Rearing	Age	Litter size	Young dead	Infant Mortality
397	Ursidae	<i>Ursus</i>	<i>maritimus</i>	Rochester	A				2	1	0.50
398	Ursidae	<i>Ursus</i>	<i>maritimus</i>	Topeka	A	Wild	Parental	9	2	2	1.00
399	Ursidae	<i>Ursus</i>	<i>maritimus</i>	Topeka	A	Wild	Parental	11	2	2	1.00
400	Ursidae	<i>Ursus</i>	<i>maritimus</i>	Topeka	A	Wild	Parental	12	2	1	0.50
401	Ursidae	<i>Ursus</i>	<i>maritimus</i>	Tulsa	A			6	2	2	1.00
402	Ursidae	<i>Ursus</i>	<i>maritimus</i>	Tulsa	A			7	1	1	1.00
403	Ursidae	<i>Ursus</i>	<i>maritimus</i>	Tulsa	A			9	2	2	1.00
404	Ursidae	<i>Ursus</i>	<i>maritimus</i>	Tulsa	A			10	2	2	1.00
405	Ursidae	<i>Ursus</i>	<i>maritimus</i>	Tulsa	A			11	2	2	1.00
406	Ursidae	<i>Ursus</i>	<i>maritimus</i>	Tulsa	A			12	2	0	0.00
407	Ursidae	<i>Ursus</i>	<i>maritimus</i>	Dresden	A				2	2	1.00
408	Ursidae	<i>Tremarctos</i>	<i>ornatus</i>	Dresden	A				1	0	0.00
409	Ursidae	<i>Tremarctos</i>	<i>ornatus</i>	Dresden	A				2	2	1.00
410	Ursidae	<i>Tremarctos</i>	<i>ornatus</i>	Jersey	A				2	0	0.00
411	Ursidae	<i>Tremarctos</i>	<i>ornatus</i>	Jersey	A				2	2	1.00
412	Ursidae	<i>Tremarctos</i>	<i>ornatus</i>	Calgary	Rooti	Wild	Parental	8	2	2	1.00
413	Ursidae	<i>Tremarctos</i>	<i>ornatus</i>	Calgary	Rooti	Wild	Parental	9	2	2	1.00
414	Ursidae	<i>Tremarctos</i>	<i>ornatus</i>	Calgary	Rooti	Wild	Parental	10	1	0	0.00
415	Ursidae	<i>Tremarctos</i>	<i>ornatus</i>	Buffalo	A	Wild	Parental	11	1	1	1.00
416	Ursidae	<i>Tremarctos</i>	<i>ornatus</i>	Buffalo	A	Wild	Parental	12	2	2	1.00
417	Ursidae	<i>Tremarctos</i>	<i>ornatus</i>	Buffalo	A	Wild	Parental	13	1	1	1.00
418	Ursidae	<i>Tremarctos</i>	<i>ornatus</i>	Buffalo	A	Wild	Parental	15	1	0	0.00
419	Ursidae	<i>Ursus</i>	<i>arctos</i>	Buffalo	A				2	0	0.00
420	Ursidae	<i>Ursus</i>	<i>arctos</i>	Houston	A	Wild	Parental	8	2	0	0.00
421	Viverridae	<i>Arctictis</i>	<i>binturong</i>	Dresden	A	Wild	Parental	2	2	0	0.00
422	Viverridae	<i>Arctictis</i>	<i>binturong</i>	Liberec	A	Captivity		3	3	0	0.00
423	Viverridae	<i>Arctictis</i>	<i>binturong</i>	Liberec	A	Captivity		3	3	1	0.33
424	Viverridae	<i>Arctictis</i>	<i>binturong</i>	Glasgow	A			3	2	2	1.00
425	Viverridae	<i>Arctictis</i>	<i>binturong</i>	Glasgow	A			4	3	3	1.00
426	Viverridae	<i>Arctictis</i>	<i>binturong</i>	Glasgow	A			4	1	1	1.00
427	Viverridae	<i>Arctictis</i>	<i>binturong</i>	Glasgow	A			5	3	1	0.33
428	Viverridae	<i>Arctictis</i>	<i>binturong</i>	Glasgow	A			5	1	0	0.00
429	Viverridae	<i>Arctictis</i>	<i>binturong</i>	WNZP	A	Wild	Parental	2	3	3	1.00
430	Viverridae	<i>Arctictis</i>	<i>binturong</i>	WNZP	A	Wild	Parental	3	3	0	0.00
431	Viverridae	<i>Arctictis</i>	<i>binturong</i>	Buffalo	A	Wild	Parental	2	2	0	0.00
432	Viverridae	<i>Arctictis</i>	<i>binturong</i>	Buffalo	A	Wild	Parental	3	3	0	0.00
433	Viverridae	<i>Arctictis</i>	<i>binturong</i>	Buffalo	A	Wild	Parental	4	4	1	0.25
434	Viverridae	<i>Arctogalidia</i>	<i>trivirgata</i>	Santa Cruz USA	A	Wild	Parental	3	3	0	0.00
435	Viverridae	<i>Atilax</i>	<i>paludinosus</i>	Berlin ZG	A			2	3	3	1.00
436	Viverridae	<i>Atilax</i>	<i>paludinosus</i>	Berlin ZG	A			2	3	2	0.67
437	Viverridae	<i>Cryptoprocta</i>	<i>ferox</i>	Montpellier	A	Wild	Hand	4	2	0	0.00
438	Viverridae	<i>Fossa</i>	<i>fossa</i>	WNZP	A	Wild	Parental	4	1	0	0.00
439	Viverridae	<i>Genetta</i>	<i>genetta</i>	Tucson	A				2	0	0.00
440	Viverridae	<i>Hemigalus</i>	<i>derbyanus</i>	Wassenaar	A			2	2	0	0.00

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	S	T	U	V	W	X	Y
	Cause of death	Age of death(Days)	Cage area (m2)	Ind/cage	Dens	Male separated?	Reference
397	Neglect	0				Yes	Michalowski 1971
398	Neglect	0				Yes	Wortman & Larue 1974
399	Hand-rearing	25				Yes	Wortman & Larue 1974
400	Pneumonia/Hand-rearing	120				Yes	Wortman & Larue 1974
401	Neglect	4	50	1	1	Yes	Nunley 1977
402	Neglect	2	50	1	1	Yes	Nunley 1977
403	Eaten by dam	0	50	1	1	Yes	Nunley 1977
404	Hand-rearing	24, 67	50	1	1	Yes	Nunley 1977
405	Neglect/Hand-rearing	3	50	1	1	Yes	Nunley 1977
406	Not Reported	0	50	1	2	Yes	Nunley 1977
407				2			Gensch 1965
408				2			Gensch 1965
409	Eaten by dam	0	267	2	4	Yes	Bloxam 1977
410				2	4	Yes	Bloxam 1977
411	Eaten by dam	2		2	0	No	Peel, Price & Karsten 1979
412	Eaten by dam	7	40	2	2	Yes	Peel, Price & Karsten 1979
413	Eaten by dam		40	2	2	Yes	Peel, Price & Karsten 1979
414	Eaten by dam	1		2	1	No	Aquilina 1981
415	Eaten by dam	2		2	1	No	Aquilina 1981
416	Eaten by dam	0		2	1	No	Aquilina 1981
417				1	2	Yes	Aquilina 1981
418				2	1	No	Freiheit 1969
419				2	0	No	Quick 1969
420				2	1	No	Gensch 1962
421			20	2	1	No	Bulir 1972
422			5	2	0	No	Bulir 1972
423	Not Reported	0	5	1	1	Yes	Kuschinski 1974
424	Not Reported	0	20	2	2	No	Kuschinski 1974
425	Stillbirth	0	20	2	2	No	Kuschinski 1974
426	Neglect	0	20	2	2	No	Kuschinski 1974
427	Hand-rearing	25	20	2	2	Yes	Kuschinski 1974
428			20	2	3	Yes	Kuschinski 1974
429	Neglect	5	16	2	1	No	Xanten, Kafka & Olds 1976
430			2	1	3	Yes	Xanten, Kafka & Olds 1976
431			9	1	1	Yes	Aquilina & Beyer 1979
432			9	3	1	Yes	Aquilina & Beyer 1979
433	Stillbirth	0	18	4	1	No	Aquilina & Beyer 1979
434			2	2	1	Yes	Batten & Batten 1966
435	Eaten by dam	0	8	2	0	No	Frese 1981
436	Killed by others	60	8	3	1	No	Frese 1981
437				2	1		Albignac 1975
438				2		No	Wemmer 1971
439				2		Yes	Flint 1975
440			6	2	1	Yes	Louwman 1970

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	A	B	C	K	L	M	N	O	P	Q	R
	Family	Genus	Species	Zoo	Dam	Origin	Rearing	Age	Litter size	Young dead	Infant Mortality
441	Viverridae	Paradoxurus	hermaphroditus	Dvur Kralove	A	Wild	Parental	3	3	0	0.00
442	Viverridae	Paradoxurus	hermaphroditus	Dvur Kralove	A	Wild	Parental	3	3	2	0.67
443	Viverridae	Paradoxurus	hermaphroditus	Dvur Kralove	A	Wild	Parental	4	3	0	0.00
444	Viverridae	Paradoxurus	hermaphroditus	Dvur Kralove	A	Wild	Parental	4	3	3	1.00
445	Viverridae	Prionodon	linsang	Wassenaar	A			2	2	0	0.00
446	Viverridae	Viverra	civetta	Jersey	Sierra Leone			2	2	2	1.00
447	Viverridae	Viverra	civetta	Jersey	Sierra Leone			3	3	0	0.00
448	Viverridae	Viverra	civetta	Jersey	Uganda			3	2	2	1.00
449	Viverridae	Viverra	civetta	Jersey	Uganda			4	4	1	0.25
450	Viverridae	Viverra	civetta	Jersey	Uganda			5	1	1	1.00
451	Viverridae	Viverra	civetta	Jersey	Uganda			6	2	1	0.50
452	Viverridae	Viverra	civetta	Jersey	Zoo-born	Captivity	Parental	2	3	3	1.00
453	Viverridae	Viverra	civetta	Jersey	Zoo-born	Captivity	Parental	3	2	2	1.00
454	Viverridae	Viverra	civetta	Jersey	Zoo-born	Captivity	Parental	4	1	1	1.00
455	Viverridae	Viverra	zibetha	Ahmedabad	A			2	3	0	0.00
456	Viverridae	Viverra	zibetha	Ahmedabad	A			3	3	0	0.00
457	Viverridae	Viverra	zibetha	Ahmedabad	A			3	3	0	0.00

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	S	T	U	V	W	X	Y
	Cause of death	Age of death(Days)	Cage area (m2)	Ind/cage	Dens	Male separated?	Reference
441				2		Yes	Dobroruka 1978
442		3		2		Yes	Dobroruka 1978
443	Hand-rearing/pneumonia			2		Yes	Dobroruka 1978
444		2		2		Yes	Dobroruka 1978
445	Neglect			2		Yes	Dobroruka 1978
446			6	2	1	Yes	Louwman 1970
447	Not Reported	2	54	2	2	No	Mallinson 1969
448			54	2	2	No	Mallinson 1969
449	Eaten by dam	0	54	2	5	No	Mallinson 1969
450	Stillbirth	0	54	2	5	No	Mallinson 1969
451	Eaten by dam	0	54	2	5	No	Mallinson 1969
452	Killed by dam	0	54	2	5	No	Mallinson 1969
453	Killed by dam	0	54	2	5	No	Mallinson 1969
454	Killed by dam	0	54	2	5	No	Mallinson 1969
455	Killed by dam	1	54	2	5	No	Mallinson 1969
456			16	1	1	Yes	David 1967
457			16	1	1	Yes	David 1967

Keys:		IZY data:		Individual litters:							
First part:											
Species											
YEAR											
M+	Males born										
F+	Females born										
U+	Unsexed										
T+	Total born										
M-	Males dead before 30 days										
F-	Females dead before 30 days										
U-	Unsexed dead b4 30 days										
T-	Total dead										
No. of litters	Please fill with the number of litters born in the year										
Samples:											
Species	YEAR	M+	F+	U+	T+	Young dead	D/D	Area	Ind/lenc	Dens	Keepers
<i>Amblyonyx cinereus</i>	1993	2	2	4	8						
		M-	F-	U-	T-						
		0	0	0	0						
Name/code of young	Date of birth	Sire	Dam	Rearing	Young dead	Cause of death	D/D	Area	Ind/lenc	Dens	Keepers
A12M, A13F	12 3 93	A01	A02	MOTHER	A12M	PREDATOR	20 3 93	230	7	3	1
A21M, A22M, A23F	20.4 93	A01	A03	MOTHER	A21M	HYPOTHERMIA	21 4 93	230	7	3	1
Parental details:											
Species	Name/code	Gender	Date of birth	Place of birth	Type of rearing	Notes					
<i>A. cinereus</i>	A01	M	10 2 91	Frankfurt Zoo	Hand	Tamed Presented in 30 8 91					
<i>A. cinereus</i>	A02	F	? 89	Wild (Asia)	Mother	Taken to Singapore Zoo in 8 2 90 Pres 21 5 91					
<i>A. cinereus</i>	A03	F	20 4 92	London Zoo	Mother	Pres in 1 3 93					

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	A	B	C	D	E	F	G	H	I
	Zoo	Species	Date of birth	Sire	Age	Dam	Age	Travel (km)	Origin
1									
2	AU1	<i>Suricata suricatta</i>	24.04.96	SS8m		SS11f		0	
3	AU1	<i>Suricata suricatta</i>	01.11.92	SS11m		SS14f		0	
4	AU1	<i>Suricata suricatta</i>	04.02.92	SS11m		SS14f		0	
5	AU1	<i>Suricata suricatta</i>	04.09.91	SS11m		SS14f		0	
6	AU1	<i>Suricata suricatta</i>	08.10.90	SS11m		SS14f		0	
7	AU1	<i>Suricata suricatta</i>	12.01.91	SS11m		SS14f		0	
8	AU1	<i>Suricata suricatta</i>	15.09.96	SS11m		SS14f		0	
9	AU1	<i>Suricata suricatta</i>	19.11.91	SS11m		SS14f		0	
10	AU1	<i>Suricata suricatta</i>	17.03.90	SS6m		SS6f		0	
11	AU2	<i>Chrysocyon brachyurus</i>	10.07.88	CB5m		CB5f		0	
12	AU2	<i>Chrysocyon brachyurus</i>	25.06.90	CB5m		CB6f		0	
13	AU2	<i>Chrysocyon brachyurus</i>	07.08.89	CB6m		CB7f		0	
14	AU2	<i>Chrysocyon brachyurus</i>	21.09.88	CB6m		CB7f		0	
15	AU2	<i>Chrysocyon brachyurus</i>	25.06.90	CB6m		CB7f		0	
16	EE1	<i>Ailurus fulgens</i>	15.07.96	AF3m	5	AF3f	5		
17	EE1	<i>Ailurus fulgens</i>	23.06.94	AF3m	3	AF3f	3		
18	EE1	<i>Ailurus fulgens</i>	26.06.97	AF3m	6	AF3f	6		
19	EE1	<i>Chrysocyon brachyurus</i>	11.12.92	CB7m	5	CB8f		11000	
20	EE1	<i>Chrysocyon brachyurus</i>	19.04.91	CB7m	4	CB8f		11000	
21	EE1	<i>Chrysocyon brachyurus</i>	31.01.95	CB8m		CB9f			
22	EE1	<i>Ursus maritimus</i>	08.11.96	UM2m		UM2f			
23	EE2	<i>Chrysocyon brachyurus</i>	04.03.89	CB1m	5	CB1f	4	1670	Captivity
24	EE2	<i>Chrysocyon brachyurus</i>	25.01.90	CB1m	6	CB1f	5	1670	Captivity
25	EE2	<i>Chrysocyon brachyurus</i>	10.12.92	CB2m~	3	CB1f	7	1670	Captivity
26	EE2	<i>Panthera tigris</i>	01.03.96	PT6m	3	PT6f	5		Wild
27	EE2	<i>Panthera tigris</i>	16.04.96	PT6m	3	PT7f	4		Wild
28	NL1	<i>Amblonyx cinerea</i>	03.06.96	AC1m	6	AC1f	8	50	
29	NL1	<i>Amblonyx cinerea</i>	03.12.94	AC1m	4	AC1f	6	50	
30	NL1	<i>Amblonyx cinerea</i>	04.06.93	AC1m	3	AC1f	5	50	
31	NL1	<i>Amblonyx cinerea</i>	22.08.95	AC1m	5	AC1f	7	50	
32	NL1	<i>Amblonyx cinerea</i>	25.09.89	AC2m	3	AC1f	1	50	
33	NL1	<i>Amblonyx cinerea</i>	28.04.94	AC3m	4	AC1f	6	50	
34	NL1	<i>Amblonyx cinerea</i>	29.06.92	AC3m	2	AC1f	4	50	
35	NL1	<i>Panthera tigris</i>	07.07.89	PT5m	4	PT5f	4	200	
36	NL1	<i>Panthera tigris</i>	15.08.92	PT5m	7	PT5f	7	200	

	J	K	L	M	N	O	P	Q	R
	Rearing	Litter size	no of dead	Cause of death	age of death	Cage size	ind/cage	no of dens	no of keepers
1	Parental	1	0						3
2	Parental	2	2	Eaten	36/50 days				3
3	Parental	4	1	Unknown	44 days				3
4	Parental	3	3	Eaten	5/8 days				3
5	Parental	3	3	Eaten		0			3
6	Parental	2	0						3
7	Parental	1	1	Unknown	1 day				3
8	Parental	1	1	Unknown	33 days				3
9	Parental	1	1	Eaten	6 days				3
10	Parental	5	5	Eaten	5 days			6	3
11	Parental	2	2	Eaten	90 days			6	3
12	Parental	3	1	Euthanasia	1 year +			6	3
13	Parental	2	2	Eaten	7 days			6	3
14	Parental	5	5	Eaten	90 days			6	3
15	Parental	1	1	Vanished	5 days	60	2	4	1
16		1	0			60	2	4	1
17		1	1	Eaten	2 1/2 months	60	2	4	1
18	Parental	2	2	Killed by dam	2 days				
19	Parental	4	2	Unknown	3 days				
20		1	1	Unknown	4 days				
21		3	3	Unknown		0			
22	Unknown	3	0						
23	Unknown	7	2	Unknown	7				
24	Unknown	2	0						
25	Parental	3	1	Unknown	17				
26	Parental	3	0						
27	Parental	6	4	Euthanasia (surplus)	0	13500	5	2	4
28	Parental	6	0			13500	9	2	4
29	Parental	6	0			13500	6	2	4
30	Parental	6	1	Stillborn	0	13500	5	2	4
31	Parental	1	0			13500	4	2	4
32	Parental	3	0			13500	9	2	4
33	Parental	7	0			13500	4	2	4
34	Unknown	3	1	Unknown	62 days	1600	2	2	4
35	Unknown	3	0			1600	2	2	4

	A	B	C	D	E	F	G	H	I
	Zoo	Species	Date of birth	Sire	Age	Dam	Age	Travel (km)	Origin
37									
38	NL1	<i>Panthera tigris</i>	21.07.91	PT5m	6	PT5f	6	200	
39	NL1	<i>Suncata suricatta</i>	01.09.91	SS3m		SS3f	3	200	
40	NL1	<i>Suricata suricatta</i>	01.09.92	SS3m		SS3f	4	200	
41	NL1	<i>Suricata suricatta</i>	02.01.90	SS3m		SS3f	15	200	
42	NL1	<i>Suricata suricatta</i>	03.06.90	SS3m		SS3f	2	200	
43	NL1	<i>Suricata suricatta</i>	06.01.91	SS3m		SS3f	3	200	
44	NL1	<i>Suricata suricatta</i>	06.06.91	SS3m		SS3f	3	200	
45	NL1	<i>Suricata suricatta</i>	10.04.93	SS3m		SS3f	5	200	
46	NL2	<i>Canis lupus</i>	15.05.88	CL2m		CL2f	8	0	
47	NL2	<i>Canis lupus</i>	15.05.89	CL2m		CL2f	9	0	
48	NL2	<i>Canis lupus</i>	01.06.95	CL5m		CL5f			
49	NL2	<i>Canis lupus</i>	23.05.94	CL5m		CL5f			
50	UK1	<i>Panthera tigris</i>	04.01.96	PT8m	9	PT9F	8		
51	UK1	<i>Panthera tigris</i>	08.03.95	PT8m	9	PT9F	8		
52	UK1	<i>Panthera tigris</i>	10.02.94	PT8m	8	PT9F	7		
53	UK1	<i>Panthera tigris</i>	19.06.94	PT8m	8	PT9F	7		
54	UK1	<i>Panthera tigris</i>	24.10.94	PT8m	8	PT9F	7		
55	UK2	<i>Ailurus fulgens</i>	11.06.92	AF1m	45	AF1f	25	17000	
56	UK2	<i>Ailurus fulgens</i>	20.06.96	AF2m	11	AF2f	3	500	
57	UK2	<i>Nasua nasua</i>	03.05.92	NN1m	2	NN1f	10	0	
58	UK2	<i>Nasua nasua</i>	10.04.94	NN1m	4	NN1f	12	0	
59	UK2	<i>Nasua nasua</i>	10.04.95	NN1m	5	NN2f	5	0	
60	UK2	<i>Nasua nasua</i>	13.04.96	NN1m	6	NN2f	6	0	
61	UK2	<i>Nasua nasua</i>	30.07.94	NN1m	4	NN2f	4	0	
62	UK2	<i>Nasua nasua</i>	06.04.95	NN1m	5	NN3f	3	0	
63	UK2	<i>Nasua nasua</i>	05.07.96	NN1m	6	NN4f	6	0	
64	UK2	<i>Nasua nasua</i>	20.04.90	NN2m	5	NN5f	8	0	
65	UK2	<i>Nasua nasua</i>	23.04.91	NN2m	6	NN5f	9	0	
66	UK2	<i>Nasua nasua</i>	17.08.88	NN2m	3	NN6f	8	0	
67	UK2	<i>Nasua nasua</i>	26.04.88	NN2m	3	NN6f	8	0	
68	UK2	<i>Panthera tigris</i>	06.02.89	PT2m	8	PT2f	7	0	
69	UK2	<i>Panthera tigris</i>	07.03.93	PT2m	12	PT2f	11	0	
70	UK2	<i>Panthera tigris</i>	08.10.91	PT2m	10	PT2f	9	0	
71	UK2	<i>Panthera tigris</i>	14.07.94	PT2m	13	PT2f	12	0	
72	UK2	<i>Panthera tigris</i>	17.03.96	PT2m	15	PT2f	14	0	

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	J	K	L	M	N	O	P	Q	R
	Rearing	Litter size	no of dead	Cause of death	age of death	Cage size	ind/cage	no of dens	no of keepers
37	Unknown	2	1	Eaten	5 days	1600	2	2	4
38	Parental	5	0			18	9	1	4
39	Parental	2	0			18	9	1	4
40	Parental	2	2	Vanished	10 days	18	9	1	4
41	Parental	2	2	Vanished	16/85 days	18	9	1	4
42	Parental	1	1	Exposure	1 day	18	9	1	4
43	Parental	2	0			18	9	1	4
44	Parental	2	2	Vanished	1 day	18	9	1	4
45	Parental	10	0			3000	10	2	4
46	Parental	9	6	Vanished	38-42 days	3000	10	2	4
47		4	1	Broken ribs/ruptured liver	?	2100	7	2	4
48		3	1	Vanished	6 days	2100	7	2	4
49		1	1	Hand-reared			2		
50		1	1	Stillborn			2		
51		2	2	Unknown			2		
52		2	2	Unknown			2		
53		2	2	Unknown			2		
54		2	2	Unknown	3-11 days		2		
55	Unknown	2	2	Unknown	4-10 days	45	2	3	3
56	Hand	1	0			45	2	3	3
57	Parental	4	1	Unknown	26 days	243	5	3	6
58	Parental	3	3	Vanished	15 days	243	6	3	6
59	Parental	5	3	Killed by others	9 days	243	6	3	6
60	Parental	3	3	Killed by others	6 days	243	9	3	6
61	Parental	2	0			243	6	3	6
62	Parental	3	3	Killed by others		243	6	3	6
63	Parental	3	1	Killed by others	8 days	243	9	3	6
64	Parental	4	1	Unknown	106 days	243	2	3	6
65	Parental	3	1	Unknown	3 days	243	4	3	6
66	Parental	7	7	Unknown	11-12 days	243	3	3	6
67	Parental	6	6	Vanished	0-35 days	243	3	3	6
68	Parental	4	2	Euthanased/stillborn	0-7 days	2000	3	8	2
69	Parental	2	0			2000	3	8	2
70	Parental	2	0			2000	3	8	2
71	Parental	3	0			2000	3	8	2
72	Parental	2	2	Killed by others	1 day	2000	2	8	2

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	A	B	C	D	E	F	G	H	I
	Zoo	Species	Date of birth	Sire	Age	Dam	Age	Travel (km)	Origin
73	UK2	<i>Panthera tigris</i>	25.05.90	PT2m	9	PT2f	8	0	
74	UK2	<i>Panthera tigris</i>	21.07.88	PT2m	7	PT3f	6	0	
75	UK2	<i>Panthera tigris</i>	01.05.93	PT3m		PT4f	8	0	
76	UK2	<i>Panthera tigris</i>	08.09.92	PT3m		PT4f	7	0	
77	UK2	<i>Panthera tigris</i>	27.08.95	PT4m	2	PT4f	10	0	
78	UK2	<i>Uncia uncia</i>	01.06.91	UU1m	4	UU1f	4	200	
79	UK2	<i>Uncia uncia</i>	02.05.95	UU1m	8	UU1f	8	200	
80	UK2	<i>Uncia uncia</i>	13.05.93	UU1m	6	UU1f	6	200	
81	UK3	<i>Panthera tigris</i>	20.05.89	PT1m	16	PT1f	4	0	
82	UK3	<i>Suncata suricatta</i>	11.08.88	SS10m	2	SS13f	2	100	
83	UK3	<i>Suricata suricatta</i>	12.03.88	SS10m	2	SS13f	2	100	
84	UK3	<i>Suricata suricatta</i>	14.02.89	SS10m	2	SS13f	2	100	
85	UK3	<i>Suricata suricatta</i>	15.07.89	SS10m	3	SS13f	3	100	
86	UK3	<i>Suricata suricatta</i>	17.07.90	SS10m	4	SS13f	4	100	
87	UK3	<i>Suricata suricatta</i>	22.04.90	SS10m	4	SS13f	4	100	
88	UK3	<i>Suricata suricatta</i>	22.04.91	SS10m	5	SS13f	5	100	
89	UK3	<i>Suricata suricatta</i>	28.05.88	SS10m	2	SS13f	2	100	
90	UK3	<i>Suricata suricatta</i>	16.05.93	SS1m	7	SS1f	3	100	
91	UK3	<i>Suricata suricatta</i>	24.07.94	SS2m	2	SS1f	4	100	
92	UK3	<i>Suricata suricatta</i>	29.04.95	SS2m	3	SS2f	5	100	
93	US1	<i>Amblyonyx cinerea</i>	23.10.88	AC4m	3	AC2f	3	1200	
94	US1	<i>Canis lupus</i>	15.04.93	CL3m		CL3f	8	0	
95	US1	<i>Canis lupus</i>	18.04.94	CL3m		CL3f	9	0	
96	US1	<i>Canis lupus</i>	19.04.92	CL3m		CL3f	7	0	
97	US1	<i>Canis lupus</i>	20.04.95	CL3m		CL3f	10	0	
98	US1	<i>Canis lupus</i>	21.04.96	CL3m		CL3f	11	0	
99	US1	<i>Panthera tigris</i>	30.06.92	PT7m	10	PT8f	7	700	
100	US1	<i>Suricata suricatta</i>	11.10.96	SS7m	7	SS10f	4	0	
101	US1	<i>Suricata suricatta</i>	18.07.96	SS7m	7	SS10f	4	0	
102	US1	<i>Suncata suricatta</i>	30.12.96	SS7m	7	SS10f	4	0	
103	US1	<i>Suricata suricatta</i>	04.01.88	SS12m		SS15f	5	200	
104	US1	<i>Suricata suricatta</i>	05.03.89	SS12m		SS15f	6	200	
105	US1	<i>Suricata suricatta</i>	23.03.88	SS12m		SS15f	5	200	
106	US1	<i>Suricata suricatta</i>	04.05.93	SS12m		SS16f	5	0	
107	US1	<i>Suricata suricatta</i>	15.02.93	SS12m		SS16f	5	0	
108	US1	<i>Suncata suricatta</i>							

	A	B	C	D	E	F	G	H	I
	Zoo	Species	Date of birth	Sire	Age	Dam	Age	Travel (km)	Origin
109									
110	US1	<i>Suricata suricatta</i>	10.06.93	SS12m		SS17f	4	0	
111	US1	<i>Suricata suricatta</i>	14.05.92	SS12m		SS17f	3	0	
112	US1	<i>Suricata suricatta</i>	16.09.92	SS12m		SS17f	3	0	
113	US1	<i>Suricata suricatta</i>	26.03.93	SS12m		SS17f	4	0	
114	US1	<i>Suricata suricatta</i>	27.02.92	SS12m		SS17f	3	0	
115	US1	<i>Suricata suricatta</i>	21.06.96	SS12m		SS18f	7	0	
116	US1	<i>Suricata suricatta</i>	04.11.95	SS7m	6	SS7f	7	0	
117	US1	<i>Suricata suricatta</i>	07.08.90	SS7m	1	SS7f	2	0	
118	US1	<i>Suricata suricatta</i>	21.09.95	SS7m	6	SS7f	7	0	
119	US1	<i>Suricata suricatta</i>	12.01.91	SS7m	2	SS8f	2	0	
120	US1	<i>Suricata suricatta</i>	03.11.96	SS7m	7	SS9f	4	0	
121	US1	<i>Suricata suricatta</i>	16.08.96	SS7m	7	SS9f	4	0	
122	US2	<i>Chrysocyon brachyurus</i>	25.01.91	CB3m	10	CB2f	9	0	Captivity
123	US2	<i>Chrysocyon brachyurus</i>	19.12.90	CB4m	6	CB3f	8	0	Captivity
124	US2	<i>Chrysocyon brachyurus</i>	01.01.93	CB4m	9	CB4f	11	0	Captivity
125	US2	<i>Chrysocyon brachyurus</i>	29.12.91	CB4m	7	CB4f	9	0	Captivity
126	US3	<i>Canis lupus</i>	16.04.90	CL1m	11	CL1f	12	0	
127	US3	<i>Canis lupus</i>	17.04.89	CL1m		CL1f	11	0	
128	US3	<i>Ursus maritimus</i>	02.11.95	UM1m	10	UM1f	11	0	
129	US3	<i>Ursus maritimus</i>	25.11.96	UM1m	11	UM1f	12	0	
130	US4	<i>Ailurus fulgens</i>	04.07.88	AF4m	10	AF4f	10	0	Captivity
131	US4	<i>Ailurus fulgens</i>	17.07.89	AF4m	11	AF4f	11	0	Captivity
132	US4	<i>Ailurus fulgens</i>	30.06.88	AF5m	2	AF5f	3	193	Captivity
133	US4	<i>Ailurus fulgens</i>	04.07.91	AF5m	5	AF6f	3	0	Captivity
134	US4	<i>Ailurus fulgens</i>	07.07.90	AF5m	4	AF6f	2	0	Captivity
135	US4	<i>Ailurus fulgens</i>	20.6.93	AF6m	2	AF7f	2	0	Captivity
136	US4	<i>Canis lupus</i>	17.04.94	CL4m		CL4f	5	0	
137	US4	<i>Canis lupus</i>	21.04.96	CL4m		CL4f	7	0	
138	WE1	<i>Suricata suricatta</i>	09.11.94	SS9m		SS12f		200	
139	WE1	<i>Suricata suricatta</i>	13.01.94	SS9m		SS12f		200	
140	WE1	<i>Suricata suricatta</i>	22.01.95	SS9m		SS12f		200	
141	WE1	<i>Suricata suricatta</i>	22.08.94	SS9m		SS12f		200	
142	WE2	<i>Suricata suricatta</i>	10.09.93	SS13m		SS19f	5	200	
143	WE2	<i>Suricata suricatta</i>	15.10.95	SS13m		SS19f	7	200	
144	WE2	<i>Suricata suricatta</i>	18.08.90	SS13m		SS19f	2	200	

	J	K	L	M	N	O	P	Q	R
	Rearing	Litter size	no of dead	Cause of death	age of death	Cage size	ind/cage	no of dens	no of keepers
109	Parental	1	0						
110	Parental	5	0						
111	Parental	2	0						
112	Parental	1	1		3 days				
113	Parental	1	0						
114	Parental	1	1			0			
115	Unknown	3	3		0-1 day				
116	Unknown	4	2		2-3 days				
117	Unknown	4	0						
118	Parental	5	5		1-2 days				
119	Parental	3	0						
120	Parental	1	1			0			
121	Unknown	1	1	Unknown		0			
122	Unknown	2	1	Unknown		0			
123	Unknown	2	0						
124	Unknown	3	0						
125	Unknown	3	0						
126	Hand	3	0						
127	Hand	2	1		3 months				
128	Parental	2	2		0-10 days				
129	Parental	2	1		2 days				
130	Unknown	2	1	Unknown		0			
131	Unknown	1	1	Unknown	2 days				
132	Unknown	3	0						
133	Unknown	2	0						
134	Unknown	2							
135	Unknown	3	3	Unknown		0			
136	Parental	1	1		7 days				
137	Parental	1	1			0			
138	Parental	2	2	Vanished	8 days	40	5	1	4
139	Parental	4	4	Vanished	5 days	40	5	1	4
140	Parental	4	0			40	5	1	4
141	Parental	2	2	Vanished	6 days	40	5	1	4
142	Parental	1	1	Vanished	20 days	18	9	1	4
143	Parental	2	2	Vanished	5 days	18	9	1	4
144	Parental	3	0			18	9	1	4

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	A	B	C	D	E	F	G	H	I
	Zoo	Species	Date of birth	Sire	Age	Dam	Age	Travel (km)	Origin
145	WE2	<i>Suricata suricatta</i>	22.03.91	SS13m		SS19f	3	200	
147	WE2	<i>Suricata suricatta</i>	22.11.89	SS13m		SS19f	1	200	
148	WE2	<i>Suricata suricatta</i>	24.06.93	SS13m		SS19f	5	200	
149	WE2	<i>Suricata suricatta</i>	29.03.92	SS13m		SS19f	4	200	
150	WE2	<i>Suricata suricatta</i>	29.08.89	SS13m		SS19f	0.5	200	
151	WE3	<i>Suricata suricatta</i>	09.07.93	SS4m	9	SS4f	9	0	
152	WE3	<i>Suricata suricatta</i>	07.11.90	SS5m	4	SS5f	7	200	
153	WE3	<i>Suricata suricatta</i>	30.01.91	SS5m	5	SS5f	8	200	

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	J	K	L	M	N	O	P	Q	R
	Rearing	Litter size	no of dead	Cause of death	age of death	Cage size	ind/cage	no of dens	no of keepers
145	Parental	5	5	Vanished	11 days	18	9	1	4
147	Parental	4	0			18	9	1	4
148	Parental	1	1	Unknown	0	18	9	1	4
149	Parental	4	0			18	9	1	4
150	Parental	2	0			18	9	1	4
151	Parental	2	2		0-1 day				
152	Unknown	2	2		1 day				
153	Unknown	6	5		1 day				

