Short Communication

First Report of Microcystin-LR in the Cyanobacterium Gloeotrichia echinulata

Cayelan C. Carey, 1,* James F. Haney, 2 Kathryn L. Cottingham 1

Received 9 October 2006; accepted 6 December 2006

ABSTRACT: Gloeotrichia echinulata is a bloom-forming cyanobacterium that is common in eutrophic lakes, and less prevalent but increasing in oligotrophic lakes. We used an enzyme-linked immunosorbent assay analysis to test for the presence of the hepatotoxin microcystin-LR (MC-LR) in G. echinulata collected from an oligotrophic lake in central New Hampshire, USA. We found that G. echinulata contained MC-LR at mean concentrations of 97.07 ± 7.78 (1 s.e.) ng MC-LR g^{-1} dry wt colonies. This suggests that recent outbreaks of G. echinulata in oligotrophic lakes used as water sources throughout New England (USA) may pose a health concern. The toxicity of G. echinulata reported here suggests the need for future monitoring of microcystins in oligotrophic lakes. © 2007 Wiley Periodicals, Inc. Environ Toxicol 22: 337–339, 2007.

Keywords: Gloeotrichia echinulata; cyanobacteria; microcystin-LR; ELISA; toxicity; oligotrophic lakes; drinking water

INTRODUCTION

Gloeotrichia echinulata is a nitrogen-fixing cyanobacterium that is well-known for blooming in eutrophic lakes in northern Europe (Pierson et al., 1992; Istvanovics et al., 1993; Pettersson et al., 1993; Jacobsen, 1994; Forsell and

Correspondence to: C. C. Carey; e-mail: cayelan.carey@ebc.uu.se

*Current address: Department of Limnology, Uppsala University, 751-23 Uppsala, Sweden.

Contract grant sponsor: Andrew W. Mellon Foundation.

Contract grant sponsor: Paul K. Richter and Evelyn E. Cook Richter Memorial Fund.

Contract grant sponsor: Lake Sunapee Protective Association.

Contract grant sponsor: Northeast Aquatic Plant Management Society.
Contract grant sponsor: Dartmouth College Department of Biological Sciences.

Contract grant sponsor: UNH Agricultural Experiment Station.

Contract grant number: H469.

Published online in Wiley InterScience (www.interscience.wiley.com). DOI 10.1002/tox.20245

© 2007 Wiley Periodicals, Inc.

Pettersson, 1995; Pettersson, 1998; Hyenstrand et al., 2001; Karlsson, 2003; Karlsson-Elfgren et al., 2003, 2005). Since 2002, *G. echinulata* blooms in oligotrophic lakes have occurred across northern New England (Maine Department of Environmental Protection, New Hampshire Department of Environmental Services). Because of their historically high water quality, many of these lakes are used for drinking water and recreation. Therefore, determining whether *G. echinulata* contains toxins is relevant for human health.

Previous research indicates that *G. echinulata* may contain toxins. Cronberg (1999) and Cronberg et al. (1999) established that recreational human exposure to *G. echinulata* blooms in Lake Ringsjön, Sweden can create skin irritation, and Gromov et al. (1996) found that injections of *G. echinulata* isolated from Lake Ladoga, Russia, were fatal to mice. *G. echinulata* is in the order Nostocales and family Rivulariaceae (Prescott, 1973), and toxicity surveys of phylogentically-related cyanobacteria such as *Aphanizomenon* and *Nostoc* (Dos S Vieira et al., 2005) suggest that



¹Department of Biological Sciences, Dartmouth College, Hanover, New Hampshire 03755, USA

²Department of Zoology, University of New Hampshire, Rudman Hall, Durham, New Hampshire 03824, USA

G. echinulata may contain the hepatotoxin microcystin. However, the toxicity of G. echinulata has never been definitively identified or measured. Here, we test whether G. echinulata contains the hepatotoxin microcystin-LR (MC-LR).

METHODS

We tested for MC-LR in G. echinulata colonies collected from Lake Sunapee, a large, deep, oligotrophic lake in central New Hampshire, USA, at $43^{\circ}24'N$, $72^{\circ}2'W$. The lake has an area of 16.55 km^2 , a volume of $1.88 \times 10^8 \text{ m}^3$, and a mean depth of 10 m. G. echinulata colonies were first identified in Lake Sunapee in July 2004 (Lake Sunapee Protective Association, 2006, personal communication). The lake is a water source for many local residents.

G. echinulata colonies were collected from the upper 1 m of the water column with an 80 μm plankton net on August 22 and September 14, 2005. On each collection day we isolated two samples of 100 colonies each (four total samples) with an Olympus SZH10 dissecting microscope and estimated each colony's biovolume from its radius. Samples were frozen and transported to the Center for Freshwater Biology Analytical Laboratory at the University of New Hampshire for analysis by enzyme-linked immunosorbent assay (ELISA). Suspensions of G. echinulata were subjected to three freeze-thaw cycles followed by sonification to disrupt the cells and release any microcystins. We passed these samples through a 13 mm, 0.2 μ m Whatman PTFE syringe filter to remove particulates immediately before analysis (Sasner et al., 2001). ELISA analyses were performed using following the instructions for the Microcystin 96-Well-Plate Kits (EnviroLogix, Portland, ME).

RESULTS

G. echinulata contained low levels of the hepatotoxin MC-LR equivalence. On August 22, 2005, the mean toxicity of *G. echinulata* was 94.91 ± 4.66 (1 s.e.) ng MC-LR g⁻¹ dry wt colonies, and on September 14, 2005, the mean toxicity of *G. echinulata* was 99.24 ± 18.23 ng MC-LR g⁻¹ dry wt colonies. Although expressed as MC-LR, it is not possible to state which of the 47 or more microcystins (Rinehart et al., 1994) were present in *G. echinulata* as the ELISA is sensitive to a broad range of microcystin analogs.

DISCUSSION

This study demonstrates that *G. echinulata* can produce the toxin MC-LR. MC-LR is a slow-acting cyclic heptapeptide hepatotoxin that has wide-ranging effects on vertebrates that drink or are in contact with contaminated water (Bischoff,

2001; Sasner et al., 2001). The concentration of MC-LR in G. echinulata is lower than microcystin concentrations measured in other cyanobacteria such as Microcystis aeruginosa (concentrations ranging from 0.01 to 1.73 mg g⁻¹ freezedried cells, reviewed in Vezie et al., 1997) or Radiocystis fernandoi (2.47 mg g⁻¹ d.w., Dos S Vieira et al., 2005).

In oligotrophic Lake Sunapee, G. echinulata blooms do not appear to cause an immediate threat to human health because colony density is low (maximum density in the upper 1 m was 2.96 colonies L⁻¹ in 2005, Carey et al., in preparation), contributing only $1.16 \times 10^{-5} \mu g$ MC-LR L^{-1} . This is well below the 1 μ g MC-LR L^{-1} toxicity threshold for drinking water set by the World Health Organization (Sasner et al., 2001). However, G. echinulata could have negative effects on humans or aquatic ecosystems when colony densities are sufficiently large to create high total MC-LR concentrations. For example, in Lake Erken, Sweden, G. echinulata abundance can be as high as 5000 colonies L^{-1} (Eiler et al., 2006). If each G. echinulata colony has a biovolume of 5.46×10^{-4} mL (the mean 2005 colonial biovolume in Lake Sunapee), then we would expect that MC-LR concentrations in this lake could be as high as 0.027 μ g MC-LR L⁻¹, within two orders of magnitude of the WHO drinking water guideline.

MC-LR has the potential to negatively affect aquatic food webs, humans, and livestock that are in contact with cyanobacterial blooms (Carmichael et al., 2001; Dittman and Wiegand, 2006). Traditionally, microcystins are only measured in eutrophic systems where the frequency and intensity of cyanobacterial blooms are high (reviewed in Sasner et al., 2001). *G. echinulata*'s toxicity in samples from Lake Sunapee, albeit low, indicates that microcystin toxicity in oligotrophic systems deserves more study, especially as *G. echinulata* outbreaks occur in oligotrophic lakes throughout New England. As *G. echinulata* has the potential for exponential growth (Carey et al., in preparation), long-term monitoring of *G. echinulata* recruitment and toxicity in Lake Sunapee and other oligotrophic lakes is necessary to evaluate potential microcystin health concerns.

Authors thank R. T. Wood, Lake Sunapee Steward, for his advice throughout this project and sampling assistance, K. C. Weathers for helpful comments, and M. and T. Eliassen for the use of their property on Lake Sunapee and sampling assistance.

REFERENCES

Bischoff K. 2001. The toxicology of microcystin-LR: Occurrence, toxicokinetics, toxicodynamics, diagnosis and treatment. Vet Hum Toxicol 43:294–297.

Carmichael WW, Azevedo SMFO, Ji SA., Molica RJR, Jochimsen EM, Lau S, Rinehart KL, Shaw GR, Eaglesham GK. 2001. Human fatalities from cyanobacteria: Chemical and biological

- evidence for cyanotoxins. Environ Health Perspect 109:663-
- Cronberg G. 1999. Qualitative and quantitative investigations of phytoplankton in Lake Ringsjon, Scania, Sweden. Hydrobiologia 404:27-40.
- Cronberg G, Annadotter H, Lawton LA. 1999. The occurrence of toxic blue-green algae in Lake Ringsjon, southern Sweden, despite nutrient reduction and fish biomanipulation. Hydrobiologia 404:123-129.
- Dittman E, Wiegand C. 2006. Cyanobacterial toxins- occurrence, biosynthesis and impact on human affairs. Mol Nutr Food Res 50:7-17.
- Dos S, Vieira JM, de P Azevedo MT, de Oliveira Azevedo SM, Honda RY, Correa B. 2005. Toxic cyanobacteria and microcystin concentrations in a public water supply reservoir in the Brazilian Amazonia region. Toxicon 45:901-909.
- Eiler A, Olsson JA, Bertilsson S. 2006. Diurnal variations in the auto- and heterotrophic activity of cyanobacterial phycospheres (Gloeotrichia echinulata) and the identity of attached bacteria. Freshwater Biol 51:298-311.
- Forsell L, Pettersson K. 1995. On the seasonal migration of the cyanobacterium Gloeotrichia echinulata in Lake Erken, Sweden, and its influence on the pelagic population. Mar Freshwater Res 46:287-293.
- Gromov BV, Vepritsky AA, Mamkaeva KA, Voloshko LN. 1996. A survey of toxicity of cyanobacterial blooms in Lake Ladoga and adjacent water bodies. Hydrobiologia 322:149-151.
- Hyenstrand P, Rydin E, Gunnerhed M, Linder J, Blomqvist P. 2001. Response of the cyanobacterium Gloeotrichia echinulata to iron and boron additions- an experiment from Lake Erken. Freshwater Biol 46:735-741.
- Istvanovics V, Pettersson K, Rodrigo ,MA, Pierson D, Padisak J, Colom W. 1993. Gloeotrichia echinulata, a colonial cyanobacterium with a unique phosphorus uptake and life strategy. J Plankton Res 15:531-534.

- Jacobsen BA. 1994. Bloom formation of Gloeotrichia echinulata and Aphanizomenon flos-aquae in a shallow, eutrophic, Danish lake. Hydrobiologia 289:193-197.
- Karlsson I. 2003. Benthic growth of Gloeotrichia echinulata cyanobacteria. Hydrobiologia 506:189-193.
- Karlsson-Elfgren I, Rydin E, Hyenstrand P, Pettersson K. 2003. Recruitment and pelagic growth of Gloeotrichia echinulata (Cyanophyceae) in Lake Erken. J Phycol 39:1050-1056.
- Karlsson-Elfgren I, Hyenstrand P, Rydin E. 2005. Pelagic growth and colony division of Gloeotrichia echinulata in Lake Erken. J Plankton Res 27:145-151.
- Pettersson K. 1998. Mechanisms for internal loading of phosphorus in lakes. Hydrobiologia 374:21–25.
- Pettersson K, Herlitz E, Istvanovics V. 1993. The role of Gloetrichia echinulata in the transfer of phosphorus from sediments to water in Lake Erken. Hydrobiologia 253:123-129.
- Pierson DC, Pettersson K, Istvanovics V. 1992. Temporal changes in biomass specific photosynthesis during the summer: Regulation by environmental factors and the importance of phytoplankton succession. Hydrobiologia 243:119–135.
- Prescott GW. 1973. Algae of the Western Great Lakes Area. Koenigstein, West Germany: Otto Koeltz Science Publishers. 977 p.
- Rinehart KL, Namikoshi H, Choi BW. 1994. Structure and biosynthesis of toxins from blue-green algae (cyanobacteria). J Appl Phycol 6:159-176.
- Sasner JJ, Haney JF, Ikawa M, Schloss JA. 2001. Early signs and determinants of biotoxins (microcystins) in lakes. University of New Hampshire, Center for Freshwater Biology. EPA STAR Report 2001R827407. Available at http://cfpub.epa.gov/ncer_ abstracts/index.cfm/fuseaction/display.abstractDetail/abstract/ 639/report/F.
- Vezie C, Brient L, Sivonen K, Bertru G, Lefeuvre JC, Salkinoja-Salonen M. 1997. Occurrence of microcystin-containing cyanobacterial blooms in freshwaters of Brittany (France). Archiv Hydrobiol 139:401-413.