

FLUVIRUCINS A<sub>1</sub>, A<sub>2</sub>, B<sub>1</sub>, B<sub>2</sub>, B<sub>3</sub>, B<sub>4</sub> AND B<sub>5</sub>, NEW ANTIBIOTICS  
ACTIVE AGAINST INFLUENZA A VIRUSIII. THE STEREOCHEMISTRY AND ABSOLUTE  
CONFIGURATION OF FLUVIRUCIN A<sub>1</sub>

NOBUAKI NARUSE, MASATAKA KONISHI and TOSHIKAZU OKI

Bristol-Myers Squibb Research Institute,  
2-9-3 Shimo-meguro, Meguro-ku, Tokyo 153, Japan

YOSHINOBU INOUE and HIROSHI KAKISAWA\*

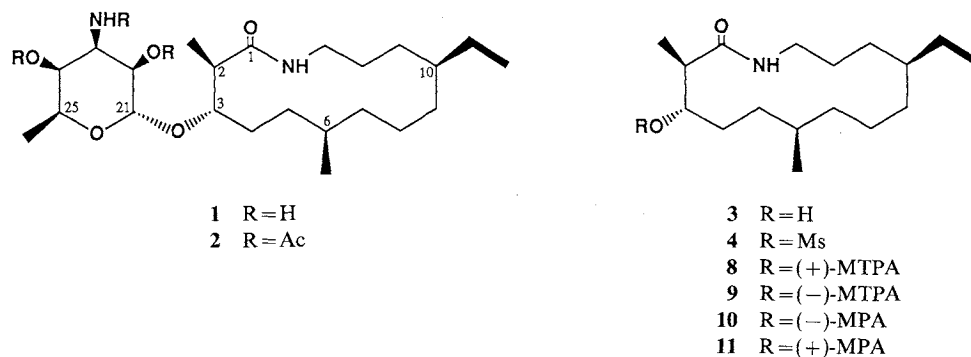
Department of Chemistry, University of Tsukuba,  
Tennodai, Tsukuba, Ibaraki 305, Japan

(Received for publication January 14, 1991)

Fluvirucin A<sub>1</sub> was established as (2*R*,3*S*,6*R*,10*S*)-3-[(3-amino-3,6-dideoxy- $\alpha$ -L-talopyranosyl)-oxy]-2,6-dimethyl-10-ethyl-13-tridecanolactam by chemical, spectroscopic, and X-ray crystallographic analyses.

Fluvirucins are new antibiotics isolated from the fermentation broth of unidentified actinomycete species and show potent inhibitory activity against influenza A virus.<sup>1,2)</sup> The structure **1** was proposed for the major component, fluvirucin A<sub>1</sub>, based on various spectroscopic and chemical properties. The 14-membered macrocyclic lactam was the first such aglycone from a natural source but there remained a question that no direct connectivity between C-11 and C-12 was observed in the 2D-incredible natural abundance double quantum transfer experiment (INADEQUATE) spectrum. The stereochemistry of the sugar part was established as 3-amino-3,6-dideoxy-L-talose but that of the aglycone remained uncertain. We report here several spectroscopic and chemical properties of **1** and a single crystal X-ray analysis of triacetyl derivative **2** of **1** to confirm the structure and the absolute configuration of the 14-membered macrocycle. Hydrolysis of fluvirucin A<sub>1</sub> (**1**) with methanolic hydrogen chloride afforded an aglycone, named fluvirucinine A<sub>1</sub> (**3**) together with two anomeric sugars.<sup>2)</sup> When a mesylate **4** derived from **3** was treated with 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) in *N,N*-dimethylformamide at 100°C for 4 hours, three dehydration-products **5**, **6**, and **7** were isolated in 36, 13, and 22%, respectively (Scheme 1). Both **5** and **6** have an olefinic double bond and the structures were easily elucidated by <sup>1</sup>H NMR spectroscopy. The remaining **7** contained a  $\beta$ -lactam skeleton (IR 1740 cm<sup>-1</sup>) and the small coupling constant ( $J=2.2$  Hz) between the adjacent protons on the 4-membered ring requires these two hydrogens to be *trans*. As the  $\beta$ -lactam ring can be considered to derive through an SN<sub>2</sub>-displacement mechanism, the stereochemistry of the original C-2 and C-3 substituents is estimated as *trans*. The large coupling constant between C2-H and C3-H in **4** ( $J=10.3$  Hz) or an acetate of **3** ( $J=9.4$  Hz)<sup>2)</sup> supports this assignment.

As the C3-OH may be equatorially oriented, application of the MOSHER<sup>3)</sup> or TROST method<sup>4)</sup> to determine the absolute stereochemistry was investigated. The results, shown in Fig. 2, are in accord with the prediction that the absolute stereochemistry of C-3 is *S*, though most of the protons on the 14-membered ring could not be assigned due to the overlap of the signals. The configuration of remaining C-6 and C-10

Fig. 1. Absolute structures of fluvirucin A<sub>1</sub> (1) and fluvirucinine A<sub>1</sub> (3), and their derivatives.

Scheme 1.

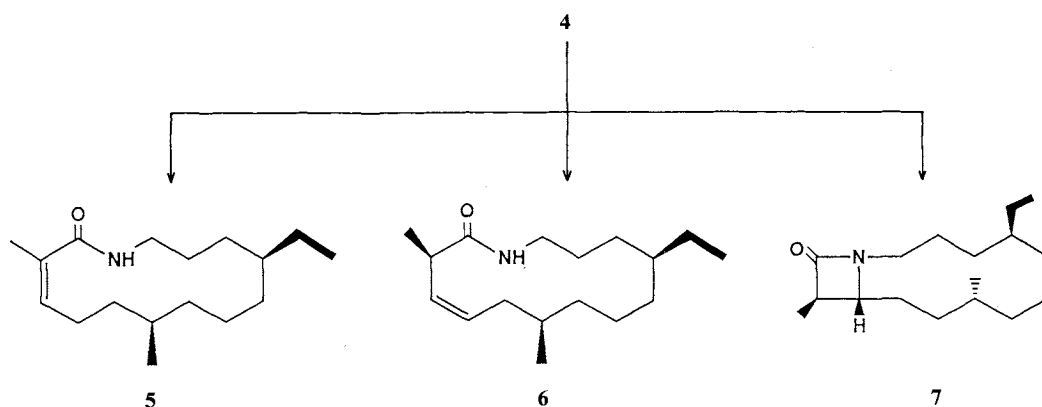
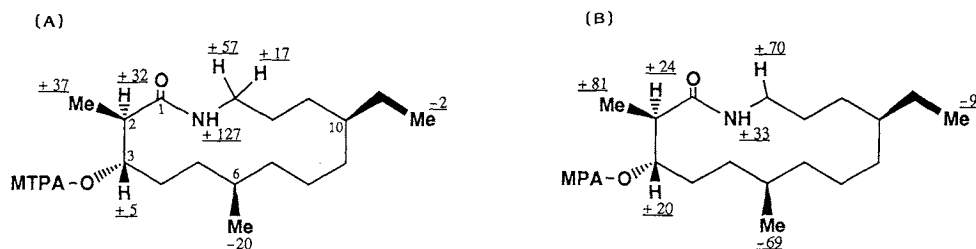


Fig. 2. Chemical shifts differences.

*R*-(+)- and *S*-(-)-MTPA esters:  $\Delta\delta$  in Hz (400 MHz) =  $\delta(S) - \delta(R)$  (A), *R*-(-) and *S*-(+)-MPA esters:  $\Delta\delta$  in Hz (400 MHz) =  $\delta(R) - \delta(S)$  (B).



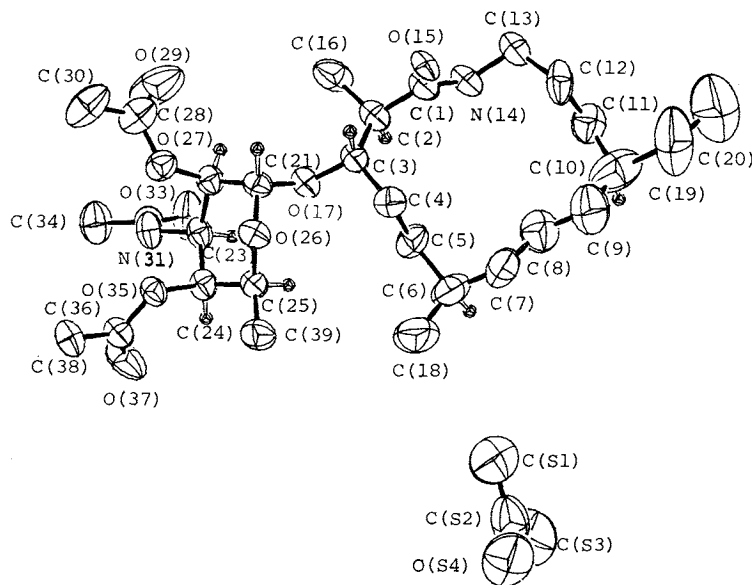
could not yet be clarified by a chemical or spectroscopic method.

We finally tried a single crystal X-ray analysis of **2** to confirm unambiguously the structure and the absolute configuration of the 14-membered macrolactam. **2** was recrystallized from acetone to give colorless rods with one equivalent of acetone as solvent of crystallization. The crystallographic data are summarized as follows: monoclinic,  $P2_1$ ;  $a = 8.317(1)$ ,  $b = 28.472(3)$ ,  $c = 8.132(1)\text{\AA}$ ,  $\beta = 110.43(1)^\circ$ ,  $V = 1805(5)\text{\AA}^3$ ,  $Z = 2$ ;  $D_m = 1.14(1)$ ,  $D_{calc} = 1.13\text{ g/cm}^3$ .

Several direct attempts with various starting phase-sets estimated from a convergence map failed to

Fig. 3. ORTEP drawing<sup>10)</sup> of the major conformer of **2** with thermal ellipsoids scaled at the 50% probability level and numbering scheme.

H-Atoms are calculated and are represented by circles of radius 0.1 Å.



solve the structure. The RANTAN approach in MULTAN11/82<sup>5)</sup> finally succeeded in location of 20 atoms (the sugar part) on the *E*-map. Successive weighted Fourier syntheses revealed the position of the remained non-hydrogen atoms except an ethyl group. The structure was refined by the full-matrix least-squares method (SHELX76).<sup>6)</sup> The atomic scattering factors were taken from International Tables for X-Ray Crystallography.<sup>7)</sup> All numerical calculations were carried out on a Facom M780/20 computer at the Science Information Processing Center, University of Tsukuba.

After isotropic refinement on 37 atoms, weighted *D*-Fourier syntheses found the ethyl group and solvent (acetone). Isotropic refinement on all non-hydrogen atoms converged the *R* factor to 0.154; at this stage a disorder was found around the 14-membered ring. Two conformations were estimated with an occupancy of 0.7 and 0.3. Since several bond-lengths in the disordered part, however, had not been converged to the normal values, the bond lengths in the minor part were fixed to 1.540 Å with e.s.d.=0.001 in the final stage. One reflection (020) considered to suffer to a secondary extinction was omitted. Final blocked full-matrix least squares with anisotropic for non-hydrogen atoms and isotropic for those in the minor disordered part converged the *R* factor to 0.098 ( $wR=0.100$ ,  $S=3.72$ ). Final weighting scheme was  $\omega=2.9/[\sigma^2(F_o)+0.0012F^2]$ . The ratio of maximum least-squares shift to error was less than 0.1.

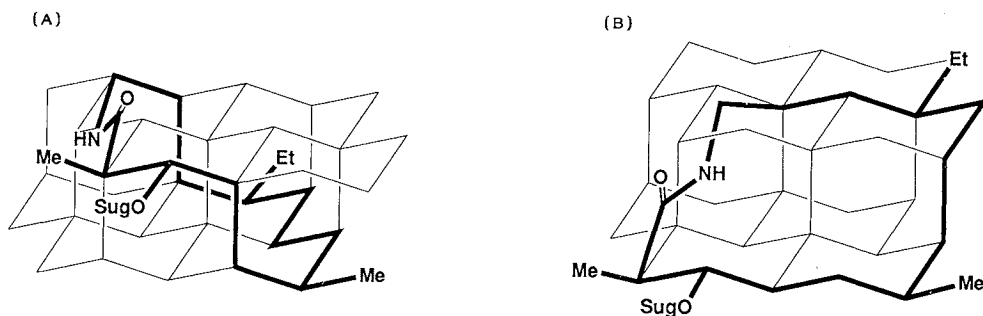
Molecular structure of the major conformer with atomic numbering scheme is shown in Fig. 3.<sup>†</sup>

The 14-membered lactam skeleton **1** was unequivocally confirmed including the substituents correctly positioned. Since the absolute configuration of the sugar had been elucidated as 3-amino-3,6-dideoxy-L-talose,<sup>2)</sup> those on the aglycone were established as 2*R*, 3*S*, 6*R*, 10*S*. The definition of 2*R*, 3*S* is accord with the result obtained by chemical and spectroscopic methods as mentioned above.

<sup>†</sup> Atomic parameters, selected bond lengths, and torsion angles of the 14-membered lactam and the sugar are deposited in Cambridge Crystallographic Data Centre.

Fig. 4. Diamond-lattice representations.

Major (A) and minor (B) conformations.



The characteristic feature of the crystal structure is that the 14-membered lactam ring is not in a single conformation. The large thermal parameters and abnormal bond lengths of several atoms<sup>†</sup> show that other conformational isomers are still possible together with disorder of the ethyl group. Two conformations obtained are shown schematically on the diamond lattice in Fig. 4. The C13-N14-C1-C2-C3 regions of both conformers are essentially same with C3-O-sugar in an equatorial position. The difference is detected in three -CH<sub>2</sub>-CH<sub>2</sub>- units (-C4-C5-, -C7-C8-, and -C11-C12-). The conformations are not in accord with those of the 14-membered lactone antibiotics, erythromycin A<sup>8)</sup> or oleandomycin derivatives.<sup>9)</sup> The difference may arise by the presence of many substituents in the latter cases, in which the conformation should be fixed by the steric environment of the substituents.

### Experimental

The mp's are uncorrected. IR spectra were determined on a Jasco IR-810 spectrometer. <sup>1</sup>H NMR spectra were recorded on a Jeol JNM-GX 400 (400 MHz) and chemical shifts were obtained relative to CDCl<sub>3</sub> as an internal standard.

#### Mesylate (4)

A mixture of 200 mg of **3** and 1 ml of methanesulfonyl chloride in 10 ml of dry pyridine was stirred at room temperature for 16 hours. After the whole had been poured onto ice-water, products were extracted with EtOAc (70 ml × 2). The organic layer was washed with 1 M HCl, a satd NaHCO<sub>3</sub> soln, and water (each 50 ml × 2) and was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>. Evaporating the solvent gave 220 mg of **4** as solids. Crystallization from CHCl<sub>3</sub> - MeOH afforded a pure **4**. MP 216~217°C; IR  $\nu_{\max}$  (KBr) cm<sup>-1</sup> 1320, 1160; <sup>1</sup>H NMR (CDCl<sub>3</sub> - CD<sub>3</sub>OD)  $\delta$  4.91 (1H, dt,  $J$  = 10.3 and 4.3 Hz, 3-H), 3.71 (1H, m, 13-H), 3.08 (3H, s, 3-OMs), 2.69 (1H, dq,  $J$  = 10.3 and 6.8 Hz, 2-H), 2.64 (1H, m, 13-H), 1.27 (3H, d,  $J$  = 6.8 Hz, 2-CH<sub>3</sub>), 0.91 (3H, d,  $J$  = 6.8 Hz, 6-CH<sub>3</sub>), 0.87 (3H, t,  $J$  = 7.3 Hz, 10-CH<sub>2</sub>CH<sub>3</sub>).

Anal Calcd for C<sub>18</sub>H<sub>35</sub>NO<sub>4</sub>S: C 59.80, H 9.76, N 3.87, S 8.87.

Found: C 59.55, H 9.77, N 3.74, S 9.21.

#### DBU Treatment on 4

To a suspension of 184 mg of **4** in 10 ml of DMF, there was added 1.5 ml of DBU and the mixture was heated at 100°C for 4 hours. After the whole had been cooled to room temperature, EtOAc was added up to 150 ml and the solution was washed with 1 M HCl, a satd NaHCO<sub>3</sub> soln, and water (each 100 ml × 2). The organic layer was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and evaporated to dryness to give a pale yellow solids. Chromatography on silica gel (toluene - MeOH, 19:1) afforded **5** (48.5 mg), **6** (17.7 mg), and **7**

<sup>†</sup> See p. 758.

(29.5 mg) as white solids.

5: MP 163~165°C; IR  $\nu_{\max}$  (KBr)  $\text{cm}^{-1}$  1660;  $^1\text{H NMR}$  ( $\text{CDCl}_3$ )  $\delta$  5.62 (1H, br s, NH), 5.40 (1H, ddq,  $J=9.0, 6.4$  and  $1.3$  Hz, 3-H), 3.44 (1H, ddt,  $J=13.3, 2.1$  and  $6.8$  Hz, 13-H), 3.26 (1H, m, 13-H), 2.20 (1H, ddt,  $J=13.3, 4.7$  and  $9.0$  Hz, 4-H), 1.94 (1H, m, 4-H), 1.89 (3H, s, 2- $\text{CH}_3$ ), 0.88 (3H, d,  $J=6.8$  Hz, 6- $\text{CH}_3$ ), 0.86 (3H, t,  $J=7.3$  Hz, 10- $\text{CH}_2\text{CH}_3$ ). NOESY spectrum showed a cross peak between C2- $\text{CH}_3$  and C3-H.

Anal Calcd for  $\text{C}_{17}\text{H}_{31}\text{NO}$ : C 76.92, H 11.77, N 5.28.

Found: C 76.97, H 11.95, N 5.25.

6: MP 172~174°C; IR  $\nu_{\max}$  (KBr)  $\text{cm}^{-1}$  1660;  $^1\text{H NMR}$  ( $\text{CDCl}_3$ )  $\delta$  5.61 (1H, br s, NH), 5.53 (1H, ddd,  $J=11.7, 10.3$  and  $3.8$  Hz, 4-H), 5.42 (1H, ddd,  $J=11.7, 9.0$  and  $1.9$  Hz, 3-H), 3.53 (1H, m, 13-H), 2.97 (1H, m, 13-H), 2.91 (1H, dq,  $J=9.0$  and  $6.8$  Hz, 2-H), 2.16 (1H, ddd,  $J=13.7, 4.4$  and  $3.8$  Hz, 5-H), 1.83 (1H, dt,  $J=13.7$  and  $10.3$  Hz, 5-H), 1.22 (3H, d,  $J=6.8$  Hz, 2- $\text{CH}_3$ ), 0.93 (3H, d,  $J=6.8$  Hz, 6- $\text{CH}_3$ ), 0.84 (3H, t,  $J=7.3$  Hz, 10- $\text{CH}_2\text{CH}_3$ ).

Anal Calcd for  $\text{C}_{17}\text{H}_{31}\text{NO}$ : C 76.92, H 11.77, N 5.28.

Found: C 76.82, H 11.88, N 5.17.

7: MP 77~78°C; IR  $\nu_{\max}$  (KBr)  $\text{cm}^{-1}$  1740;  $^1\text{H NMR}$  ( $\text{CDCl}_3$ )  $\delta$  3.75 (1H, dt,  $J=14.3$  and  $8.2$  Hz, 13-H), 3.10 (1H, dt,  $J=8.8$  and  $2.2$  Hz, 3-H), 2.70 (1H, dq,  $J=2.2$  and  $7.3$  Hz, 2-H), 2.66 (1H, ddd,  $J=14.3, 7.8$  and  $3.3$  Hz, 13-H), 1.24 (3H, d,  $J=7.3$  Hz, 2- $\text{CH}_3$ ), 0.89 (3H, d,  $J=6.6$  Hz, 6- $\text{CH}_3$ ), 0.86 (3H, t,  $J=7.3$  Hz, 10- $\text{CH}_2\text{CH}_3$ ).

Anal Calcd for  $\text{C}_{17}\text{H}_{31}\text{NO}$ : C 76.92, H 11.77, N 5.28.

Found: C 76.77, H 11.88, N 5.28.

#### *R*-(+)- and *S*-(-)- $\alpha$ -Methoxy- $\alpha$ -(trifluoromethyl)phenylacetyl Esters 8 and 9

A mixture of 13 mg of **3**, 150  $\mu\text{l}$  of  $\alpha$ -methoxy- $\alpha$ -(trifluoromethyl)phenylacetyl chloride (*R*-(+)-MTPAC1), 1 ml of  $\text{Et}_3\text{N}$ , and 10 mg of 4-dimethylaminopyridine (DMAP) in 2 ml of dichloromethane was stirred at room temperature for 16 hours. EtOAc and ice-water were added to the solution and the whole was stirred for 1 hour. After had been separated, the organic layer was washed with 1 M HCl, satd  $\text{NaHCO}_3$  soln, and water. After concentrating under reduced pressure, the crude products were purified by preparative TLC (toluene-MeOH (17:3) followed with hexane-acetone, 9:1) and column-chromatography on Sephadex LH-20 (MeOH) to afford 15 mg of **8** as white solids. The *S*-(-)-MTPA ester **9** was prepared as similar procedure.

8: MP 185~187°C;  $^1\text{H NMR}$  ( $\text{CDCl}_3$ )  $\delta$  7.57 (2H, m), 7.42 (3H, m), 5.45 (1H, dd,  $J=8.1$  and  $4.3$  Hz, NH), 5.26 (1H, dt,  $J=3.4$  and  $8.1$  Hz, 3-H), 3.75 (1H, ddt,  $J=13.7, 1.7$  and  $8.1$  Hz, 13-H), 3.59 (3H, s), 2.50 (1H, m, 13-H), 2.44 (1H, dq,  $J=8.1$  and  $7.3$  Hz, 2-H), 1.72 (1H, tt,  $J=13.9$  and  $3.4$  Hz, 4-H), 1.06 (3H, d,  $J=7.3$  Hz, 2- $\text{CH}_3$ ), 0.85 (3H, t,  $J=7.3$  Hz, 10- $\text{CH}_2\text{CH}_3$ ), 0.84 (3H, d,  $J=7.3$  Hz, 6- $\text{CH}_3$ ).

9: MP 184~186°C;  $^1\text{H NMR}$  ( $\text{CDCl}_3$ )  $\delta$  7.54 (2H, m), 7.41 (3H, m), 5.76 (1H, dd,  $J=8.1$  and  $4.3$  Hz, NH), 5.27 (1H, dt,  $J=3.4$  and  $8.6$  Hz, 3-H), 3.79 (1H, ddt,  $J=13.7, 1.7$  and  $8.1$  Hz, 13-H), 3.54 (3H, s), 2.64 (1H, m, 13-H), 2.52 (1H, dq,  $J=8.6$  and  $7.3$  Hz, 2-H), 1.73 (1H, tt,  $J=13.9$  and  $3.4$  Hz, 4-H), 1.15 (3H, d,  $J=7.3$  Hz, 2- $\text{CH}_3$ ), 0.84 (3H, t,  $J=7.3$  Hz, 10- $\text{CH}_2\text{CH}_3$ ), 0.79 (3H, d,  $J=6.8$  Hz, 6- $\text{CH}_3$ ).

#### *R*-(-)- and *S*-(+)- $\alpha$ -Methoxyphenylacetic Acid (MPA) Esters 10 and 11

A mixture of 14 mg of **3**, 53 mg of *R*-(-)-MPA, 10 mg of DMAP, and 1-ethyl-3-(3-diethylaminopropyl)carbodiimide hydrochloride in 5 ml of dichloromethane was stirred at room temperature for 16 hours. After concentrating under reduced pressure, 40 ml of EtOAc was added and the whole was washed with a 5% citric acid soln, water, a 5%  $\text{NaHCO}_3$  soln, and water (each 30 ml  $\times$  2). After evaporating the solvent, the residue was purified by preparative TLC (toluene-MeOH, 17:3) and column-chromatography on Sephadex LH-20 (MeOH) to afford 11.5 mg of **10** as white solids. *S*-(+)-MPA ester **11** was prepared by a similar procedure.

10: MP 220~221°C;  $^1\text{H NMR}$  ( $\text{CDCl}_3$ )  $\delta$  7.44 (2H, br d,  $J=7$  Hz), 7.35 (3H, m), 5.70 (1H, br dd,  $J=7.3$  and  $4.7$  Hz, NH), 5.05 (1H, dt,  $J=2.7$  and  $8.8$  Hz, 3-H), 4.77 (1H, s), 3.63 (1H, m, 13-H), 3.43 (3H, s), 2.83 (1H, dddd,  $J=13.5, 7.3, 4.7$  and  $2.1$  Hz, 13-H), 2.39 (1H, dq,  $J=8.8$  and  $6.8$  Hz, 2-H), 1.09 (3H, d,  $J=6.8$  Hz, 2- $\text{CH}_3$ ), 0.83 (3H, t,  $J=7.3$  Hz, 10- $\text{CH}_2\text{CH}_3$ ), 0.62 (3H, d,  $J=6.8$  Hz, 6- $\text{CH}_3$ ).

11: MP 250~252°C;  $^1\text{H NMR}$  ( $\text{CDCl}_3$ )  $\delta$  7.45 (2H, br d,  $J=7$  Hz), 7.37 (3H, m), 5.62 (1H, br dd,

$J=7.3$  and  $4.7$  Hz, NH),  $5.00$  (1H, dt,  $J=2.6$  and  $6.8$  Hz, 3-H),  $4.78$  (1H, s),  $3.65$  (1H, ddt,  $J=13.3, 2.6$  and  $7.3$  Hz, 13-H),  $3.42$  (3H, s),  $2.66$  (1H, dddd,  $J=13.3, 8.6, 4.7$  and  $2.1$  Hz, 13-H),  $2.33$  (1H, quint,  $J=6.8$  Hz, 2-H),  $0.88$  (3H, d,  $J=6.8$  Hz, 2-CH<sub>3</sub>),  $0.86$  (3H, t,  $J=7.3$  Hz, 10-CH<sub>2</sub>CH<sub>3</sub>),  $0.79$  (3H, d,  $J=7.3$  Hz, 6-CH<sub>3</sub>).

#### Crystallographic and Diffraction Data Collection of 2

The crystals were grown from an acetone solution as colorless rods. MP  $246\sim 247^\circ\text{C}$  (changed to opaque above  $148^\circ\text{C}$  due to the solvent loss).

*Anal.* Calcd for C<sub>29</sub>H<sub>50</sub>N<sub>2</sub>O<sub>8</sub>·C<sub>3</sub>H<sub>6</sub>O: C 62.72, H 9.21, N 4.57.

Found: C 62.21, H 9.18, N 4.57.

The densities ( $D_m$ ) were measured by flotation in aqueous KI solution. The unit-cell dimensions were refined by least-squares refinement from 25 reflections with  $11^\circ < \theta < 13^\circ$  on an Enraf-Nonius CAD4 automated kappa-axis diffractometer with graphite monochromated Mo K $\alpha$  radiation (50 kV and 26 mA). The diffraction intensities were measured for a single crystal of about  $0.4 \times 0.3 \times 0.3 \text{ mm}^3$  on the diffractometer up to  $2\theta=60^\circ$ ;  $2\theta-\omega$  scan, the scan rate with  $4^\circ \text{ min}^{-1}$  in  $\theta$ , the  $\omega$  scan width =  $0.6 + 0.35 \tan \theta$ . Three standard reflections were measured after every 2 hours. The intensities were corrected for Lorentz, polarization, and decay ( $-1.2\%$  in intensity) but not for absorption. Systematic absence ( $k=2n$ ) indicated unambiguously the space group  $P2_1$ . Of 5,345 unique reflections, 3,453 of independent structure factors with  $|F_o| > 3\sigma(F_o)$  were used for structure determination.

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