

# Forests trapped in nitrogen limitation – an ecological market perspective on ectomycorrhizal symbiosis

Oskar Franklin<sup>1</sup>, Torgny Näsholm<sup>2</sup>, Peter Högberg<sup>2</sup> and Mona N. Högberg<sup>2</sup>

<sup>1</sup>IIASA- International Institute for Applied Systems Analysis, A-2361 Laxenburg, Austria; <sup>2</sup>Department of Forest Ecology and Management, Swedish University of Agricultural Sciences, SE-901 83 Umeå, Sweden

## Summary

Author for correspondence:

Oskar Franklin

Tel: +43 2236 807251

Email: franklin@iiasa.ac.at

Received: 31 January 2014

Accepted: 2 April 2014

*New Phytologist* (2014) **203**: 657–666

doi: 10.1111/nph.12840

**Key words:** below-ground, elevated carbon dioxide, game theory, mutualism, optimization, pine, trade, tragedy of the commons.

- Ectomycorrhizal symbiosis is omnipresent in boreal forests, where it is assumed to benefit plant growth. However, experiments show inconsistent benefits for plants and volatility of individual partnerships, which calls for a re-evaluation of the presumed role of this symbiosis.
- We reconcile these inconsistencies by developing a model that demonstrates how mycorrhizal networking and market mechanisms shape the strategies of individual plants and fungi to promote symbiotic stability at the ecosystem level.
- The model predicts that plants switch abruptly from a mixed strategy with both mycorrhizal and nonmycorrhizal roots to a purely mycorrhizal strategy as soil nitrogen availability declines, in agreement with the frequency distribution of ectomycorrhizal colonization intensity across a wide-ranging data set. In line with observations in field-scale isotope labeling experiments, the model explains why ectomycorrhizal symbiosis does not alleviate plant nitrogen limitation. Instead, market mechanisms may generate self-stabilization of the mycorrhizal strategy via nitrogen depletion feedback, even if plant growth is ultimately reduced.
- We suggest that this feedback mechanism maintains the strong nitrogen limitation ubiquitous in boreal forests. The mechanism may also have the capacity to eliminate or even reverse the expected positive effect of rising CO<sub>2</sub> on tree growth in strongly nitrogen-limited boreal forests.

## Introduction

The combination of vast carbon (C) stores and a particularly strong projected temperature rise makes boreal forests a critical component of the future climate system (Foley *et al.*, 1994). While low nitrogen (N) supply typically constrains C input by plant growth in this biome (Jarvis & Linder, 2000), large amounts of C are stored in the soils (Post *et al.*, 1982). Although well-known factors such as net primary production, temperature, and precipitation all influence C storage in forest soils, there is an even stronger predictor of high soil C – the presence of ectomycorrhiza (Averill *et al.*, 2014). This fundamental component of forest ecosystems is highly sensitive to ongoing global changes such as rising atmospheric CO<sub>2</sub> (Fransson *et al.*, 2005; Garcia *et al.*, 2008) and N deposition (Cox *et al.*, 2010; Högberg *et al.*, 2010; Bahr *et al.*, 2013). Not only ectomycorrhizal fungi (EMF) but also arbuscular mycorrhizal fungi (AMF) have the potential to significantly influence forest N dynamics (Hodge & Fitter, 2010). Consequently, the stability and behavior of mycorrhizal symbiosis in forests may play a significant role in the progression of global change. However, evaluation of mycorrhizal responses to global changes is hampered by our limited mechanistic understanding of the role of mycorrhizal symbiosis in ecosystems (Johnson *et al.*, 2013). Here we aim to establish principles for the

stability and function of ectomycorrhizal symbiosis in a forest ecosystem context, focusing on boreal forests.

Evolution of symbiosis between N-limited plants and C-limited mycorrhizal fungi should require mutualistic C–N exchange (Hoeksema & Schwartz, 2003); that is, both parties benefit by participating in the exchange. However, plants do not always benefit (Egger & Hobbie, 2004; Jones & Smith, 2004; Corrêa *et al.*, 2012; Walder *et al.*, 2012). Even in strongly N-limited boreal forest, a recent study suggests that EMF sustain rather than alleviate plant N limitation by reducing the fraction of fungal N uptake transferred to trees as soil N availability declines (Näsholm *et al.*, 2013). Conversely, experimental N additions increased the proportion of N transferred to the trees and the N : C exchange ratio between fungi and trees, implying a greater symbiotic benefit for the trees at high than at low soil N availability (Näsholm *et al.*, 2013). These findings appear inconsistent with the common view of ectomycorrhizal symbiosis and its predominance in nutrient-poor forests.

Recent theoretical progress on mycorrhizal symbiosis has elucidated the conditions for evolutionary stability of pair-wise partnerships in the presence of nonsymbiotic alternative strategies and potential cheaters, that is, individuals taking benefits without providing something in return to their partners (de Mazancourt & Schwartz, 2010; Grman *et al.*, 2012). However, these models

are not obviously applicable to the multiple-partner structure of ectomycorrhizal symbiosis and its dynamic nature (Kennedy, 2010; Pickles *et al.*, 2010). More specifically, in addition to the inconsistent benefits for plants discussed above, lack of partner specificity and fidelity in ectomycorrhizal symbiosis (Kennedy, 2010) challenge mycorrhizal theories based on evolved species-specific pair-wise partnerships. An ecological market perspective (Noë & Hammerstein, 1994; Jones *et al.*, 2012; Werner *et al.*, 2014) opens a way forward by allowing mutualistic symbiosis to be maintained by market mechanisms, such as optimal selection of trading partners (a host distributes trading among its partners to minimize overall 'cost' per unit resource (Kummel & Salant, 2006)) and reciprocal rewards, that is, beneficial behavior toward a partner is reciprocated (Kiers *et al.*, 2011; Fellbaum *et al.*, 2012). However, observations of strongly nonreciprocal behavior (Walder *et al.*, 2012) appear to call for another explanation. More generally, the ecologically critical questions remain: why does mycorrhizal symbiosis often persist even when it appears not to benefit plants, and what are the consequences of this behavior for ecosystem function? We hypothesize that the interactions among multiple partners of both plants and fungi (mycorrhizal networks) shape the strategies of individuals to promote symbiotic stability even if forest productivity is reduced. The hypothesis is implemented as a tractable market model of C–N exchange in boreal forest, which goes beyond earlier market models (e.g. Kummel & Salant, 2006) by accounting for multiple simultaneous partners of both plants and fungi and adaptation of individuals' strategies. We evaluate the model against published empirical observations, and discuss its implications for the role of mycorrhizal symbiosis in the boreal forest and more generally.

## Description

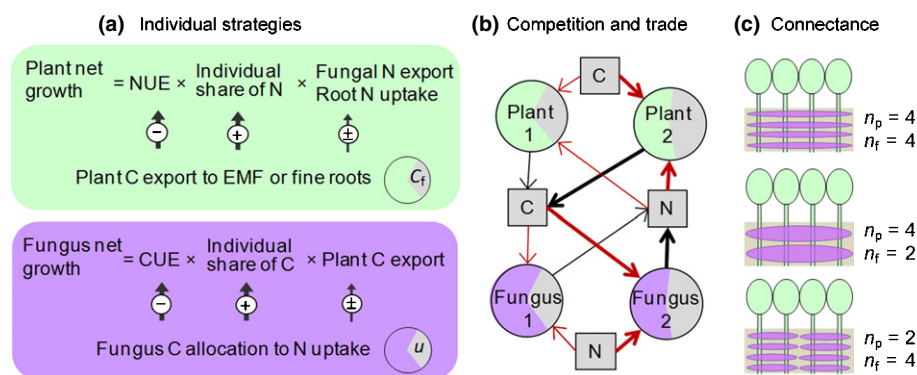
We developed a mycorrhizal symbiosis model (Fig. 1, Supporting Information Methods S1, Table S1, Fig. S1) based on established equations for N-limited plant growth (Franklin *et al.*, 2012) and

C-limited (or C- and N-co-limited) fungal growth (Näsholm *et al.*, 2013). However, in contrast to previous stand-based applications of these physiological models, here each fungus and plant is allowed to individually adjust its strategy in terms of internal C–N allocation and interaction (trading) with its (multiple) symbiotic partners. The interaction among multiple individuals is modeled as a market, where each plant supplies C in exchange for N and each fungus supplies N in exchange for C. The resulting N : C exchange rate is analogous to the price in a financial market and will depend on the supply and demand of C and N, the intensity of market competition (e.g. competition for plant C among multiple N-supplying fungi), and an individual's influence on supply (e.g. one fungus' N export influence on plant C supply). A mycorrhizal network provides the physical structure facilitating these market interactions.

## Individual physiology

Net plant growth is modeled as photosynthesis minus C costs of respiration, litter production, and C export to EMF, where both photosynthesis and respiration are functions of plant N content. Photosynthesis is also a function of light absorption and atmospheric CO<sub>2</sub>, where elevated CO<sub>2</sub> enhances photosynthetic N use efficiency (Franklin, 2007). Plants regulate C export to EMF depending on their N demand and the fungal N return. Gross N uptake from soil is a function of soil N availability and the N uptake component of EMF biomass, or fine-root biomass for nonmycorrhizal roots, limited by its soil exploration efficiency at low soil N, and by uptake capacity at high N. Nitrogen uptake by EMF that is in excess of their N demand for growth (determined by C import times a fixed biomass N : C ratio) is exported to host plants (Näsholm *et al.*, 2013).

Physiological parameter values were estimated based on our recent dual stable C and N isotope labeling experiments in boreal forest (Högberg *et al.*, 2010; Näsholm *et al.*, 2013) and other published data typical of boreal pine forests; for example, the

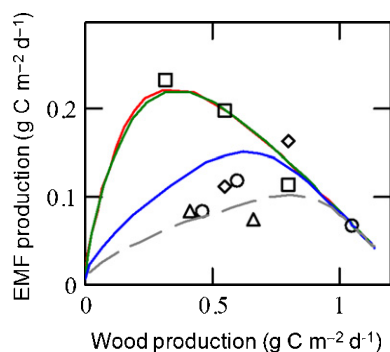


**Fig. 1** (a) An individual's strategy, that is, plant carbon (C) export ( $C_f$ ) to ectomycorrhizal fungi (EMF) and fungal allocation to nitrogen (N) uptake ( $u$ ), is optimized to maximize net growth (including reproductive production) via three factors: its hosts' total resource export (and/or nonmycorrhizal N uptake by plants), resource share obtained in intra-party competition, and resource use efficiency (N use efficiency (NUE) or C use efficiency (CUE)). (b) Competition is illustrated by uptake (red arrows) and export (black arrows) of C and N. Arrow thickness indicates flux rates: plant 2 exports more C to EMF, which results in a higher N uptake than in plant 1. Fungus 2 delivers more N and a higher N : C exchange rate, and thus receives more C than fungus 1. (c) Each plant hosts multiple competing fungi (the number of fungal partners per plant host ( $n_f$ )) and each fungus hosts multiple competing plants (the number of plant partners per fungal host ( $n_p$ )). See Supporting Information Methods S1 for the full mathematical description of the model.

biomass N : C ratio was estimated to be 0.03 for foliage and fine roots (Luoma, 1997) and 0.085 for EMF (Mikusinska *et al.*, 2013), the mean life-span of foliage was estimated to be 4 yr, that of fine roots 2 yr (Keel *et al.*, 2012), and that of EMF 17 d (Högberg *et al.*, 2008, 2010), and light-saturated photosynthetic N use efficiency was estimated to be  $0.14 \text{ g C g N}^{-1} \text{ h}^{-1}$  (Luoma, 1997). Unknown parameters were adjusted to match the range of measured EMF and tree productivity in soil N gradients in boreal forests (Fig. 2; Methods S1).

### Strategies, trade, and competition

Importantly, the model captures two critical features of ectomycorrhizal symbiosis not addressed in existing models: (1) each plant interacts simultaneously with multiple EMF and each fungus interacts with multiple plants (i.e. a mycorrhizal network; e.g. Southworth *et al.*, 2005; Beiler *et al.*, 2010), and (2) the strategies of individuals adapt to the environment (biotic and abiotic) by maximizing fitness, approximated by net growth (including reproduction) for plants (Franklin *et al.*, 2012) and reproductive growth for fungi (Pringle & Taylor, 2002). A plant's strategy in the model is defined by its C allocation to EMF ( $C_f$ ), which in effect regulates the trade-off between C investments in N uptake (via EMF) and C use for growth. A fungus strategy is defined by its ratio ( $u$ ) of C allocation to reproductive growth versus N uptake components, in line with the observed trade-off between intrinsic growth rate and potential N uptake capacity (measured as efficiency of protein mineralization) among EMF species (Eaton & Ayres, 2002). Changes in  $u$  may reflect both phenotypic plasticity and changes in active EMF community (species) composition. Thus, we do not differentiate between acclimation of  $u$  of the existing EMF and replacement of these EMF with new individuals of a different (or the same) species that have a different  $u$ . The responses of other characteristics of the fungal community to soil N availability (e.g. uptake capacity for different forms of N and spatial distribution) are implicitly subsumed in the response of N uptake to N availability (Methods S1).



**Fig. 2** Modeled (lines) versus measured (symbols) production (biomass growth) of ectomycorrhizal fungi and wood along soil nitrogen (N) gradients in boreal forests (Nilsson *et al.*, 2005; Ekblad *et al.*, 2013; Methods S1), where the sites are Varjisaan (squares), Flakastugan (diamonds), Betsela (circles), and Kryddgrovan (triangles). Line colors correspond to connectance scenarios, that is, numbers of fungal partners per plant and vice versa ( $n_f$  and  $n_p$ , respectively): green ( $n_f = 8$ ;  $n_p = 8$ ), red ( $n_f = 2$ ;  $n_p = 8$ ) blue ( $n_f = 8$ ;  $n_p = 2$ ), and gray dashed ( $n_f = 100$ ;  $n_p = 1$ ).

As a result of the resource trading, the strategy of an individual ( $u$  or  $C_f$ ) affects not only the individual's own nutrient status and fitness but also those of its trading partners. For example, if a fungus increases N export (via increased  $u$ ) this may reduce or increase the C export by its plant partners, that is, change the plants' strategy, which implies that the strategies of plants and EMF are always linked. However, if each fungus has more than one plant partner, its influence on the plant strategies, that is, C export, is 'diluted' in proportion to how many fungal partners each plant has, and vice versa for a plant's influence on its fungal partners. Thus, in the presence of multiple partners, a fungus' potential to influence its host plants' C export declines; instead, competition for this C with the other fungal partners of the same plants becomes important. If one fungus returns more N per additional plant C received than the other fungal partners, the plant can increase its fitness by trading more with this particular partner. Thus, we assume that fungal partners of the same host plants compete via their N : C exchange rate, that is, N exported to the plants per C received (and vice versa for a plant competing for fungal N export). The relative fraction of plant-derived C obtained by an individual fungus ( $F_i$ ) is a function of its N : C exchange ratio ( $x$ ) with its host plants relative to its competitors,  $F_i = F[(x_i/\bar{x})^z]$ , where  $\bar{x}$  is mean  $x$  among all competitors, and  $z$  determines how strongly  $F_i$  is affected by  $x_i$  (partner discrimination). In the absence of an empirical basis for estimating  $z$  we used a value of 1 and tested the effect of higher and lower values. In contrast to the proximity of different fungal partners on plant roots, the spatial separation between different plant partners of a fungus should enable strong partner discrimination, that is, a high  $z$ , in line with empirical observations of AMF (Lekberg *et al.*, 2010). Strong partner discrimination will force equal N : C exchange ratios among competing plants so that they effectively compete for fungal N via how much fungal biomass they support. Thus, the fraction of fungal N export a plant receives in competition with other plant partners of the same EMF is a function of its C export ( $C_f$ ) relative to its competitors.

An optimal, or equilibrium, strategy (evolutionarily stable strategy) means that no individual can gain fitness by changing its strategy, which corresponds to resource exchange with equal marginal return from all trading partners. Under our assumption of identical individuals in each party, this principle corresponds to the reciprocal rewards concept (Kiers *et al.*, 2011) but it is more general because it does not exclude nonreciprocity if partners of the same host differ in their C or N supply versus demand (price) response, for example for different plant species linked to the same mycorrhizal fungus (Walder *et al.*, 2012).

An additional constraint on the N : C exchange ratio is set by the possibility for plants to switch to a nonmycorrhizal strategy. If the market mechanisms described above result in an N : C exchange ratio that renders the nonmycorrhizal roots more profitable for a plant than mycorrhizal roots, to persist, EMF must increase their N : C exchange ratio via increased allocation to N uptake ( $u$ ) so that their utility for plants matches that of nonmycorrhizal roots. EMF with lower  $u$  will not receive C from the plants.

## Symbiotic stability

The ecological stability of the mycorrhizal plant strategy depends on its competitiveness against a nonmycorrhizal strategy, which in turn depends on soil N availability. EMF have a much higher (by a factor of 300 in the model) specific N uptake efficiency than nonmycorrhizal roots at low soil N availability (Jennings, 1995; Smith & Read, 2008), whereas the efficiencies converge for saturating N availability (Methods S1). Thus, under increasing soil N availability, the mycorrhizal advantage in gross N uptake rate declines and is eventually outweighed by the C costs and fungal N immobilization it incurs. As a consequence of the differences in N uptake and resulting soil N depletion between mycorrhizal and nonmycorrhizal roots, the relative benefits of each root strategy also depend on the strategy of the surrounding competitors of the plant (Methods S1). In essence, the nonmycorrhizal strategy can invade if a plant can gain fitness by growing nonmycorrhizal roots in a population of mycorrhizal plants and vice versa. If both strategies can invade each other they will coexist.

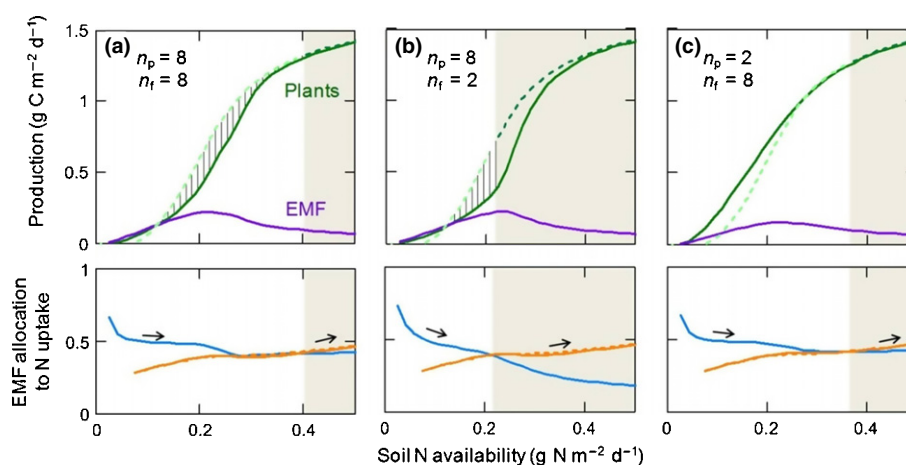
## Results

### Results and evaluation against empirical observations

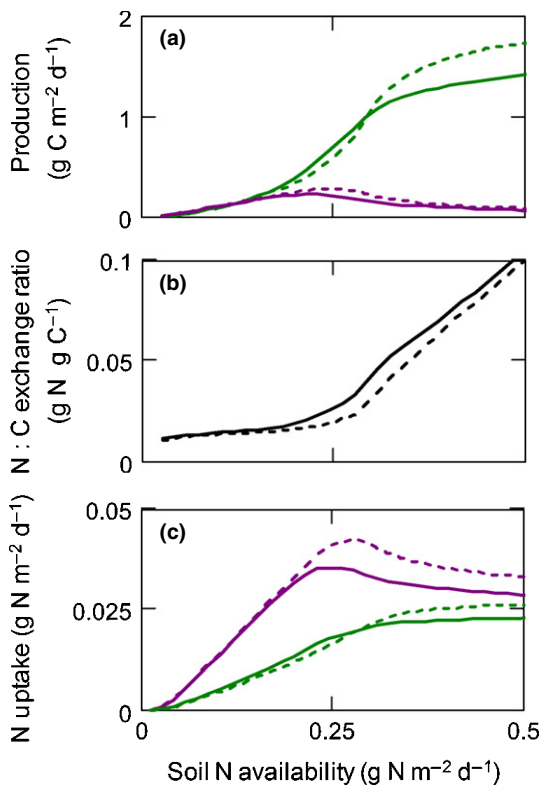
We evaluated the model's predictions in comparison to published empirical results on effects of soil N availability and atmospheric CO<sub>2</sub> in boreal or temperate ectomycorrhizal forests. The modeled responses of plants and EMF to soil N are modulated by the number of fungal partners per plant host ( $n_f$ ) and the number of plant partners per fungal host ( $n_p$ ; Fig. 3), which determines the importance of intra-party competition for an individual's strategy (Fig. 1). Competition induces tragedy-of-the-commons effects in each party that prevent either the plants or the EMF from collectively optimizing the N:C exchange rate to maximize their

benefits. A hypothetical absence of inter-fungal competition (a single fungus individual monopoly) would quickly make plants abandon the fully mycorrhizal strategy as N availability increases or, without a nonmycorrhizal option for the plants, would severely hamper plant productivity (Fig. S2). However, as long as plants have a nonmycorrhizal option or there is inter-fungal competition for plant C, C and N fluxes between plants and EMF will largely follow the basic principles of supply and demand versus cost (here the N:C exchange rate). As the N demand of fungi is constrained by their access to C from the plants, increasing soil N availability leads to higher N export to plants, resulting in increased plant growth (Fig. 3) and higher N:C exchange rate (Fig. 4), in line with results from field experiments (Näsholm *et al.*, 2013). At the same time, plant N demand drives C allocation to EMF to first increase and then decline as other factors (e.g. light) constrain plant N demand (Fig. 3), explaining observed patterns of EMF growth (Nilsson *et al.*, 2005; Hasselquist *et al.*, 2012; Kjoller *et al.*, 2012) analogously to fine-root allocation in nonmycorrhizal plants (Franklin *et al.*, 2012). Although often only the declining phase of mycorrhizal response to increasing N availability is observed, it must be preceded by an increasing phase as productivity of both plants and hosted EMF must converge to zero at zero N availability. In response to elevated atmospheric CO<sub>2</sub>, the plant-fungal N:C exchange rate is reduced (Fig. 4b) in line with observations (Alberton & Kuyper, 2009). N transfer to plants and corresponding plant net growth (mainly stem growth in trees) are enhanced by elevated CO<sub>2</sub> at high soil N availability but not at low soil N availability (Fig. 4b,c) in agreement with stand-scale observations (Oren *et al.*, 2001; Dieleman *et al.*, 2010).

As a consequence of the intra-party competition effect, a fungus' N export to its host plant (and plant growth) increases with the effective number of competing fungal partners ( $n_f$ ) while plant C export to a fungus increases with the number of plant partners per fungus ( $n_p$ ) (Fig. 3). Supporting this result, plants



**Fig. 3** Modeled net growth of plants with and without mycorrhiza (solid and dashed green lines, respectively) and ectomycorrhizal fungi (EMF; purple lines), where in (a) each plant has eight fungal partners ( $n_f = 8$ ) and each fungus has eight plant partners ( $n_p = 8$ ), in (b) each plant has fewer fungal partners ( $n_f = 2$ ;  $n_p = 8$ ), and in (c) each fungus has fewer plant partners ( $n_f = 8$ ;  $n_p = 2$ ). Fungal strategy, that is, fractional carbon (C) allocation to nitrogen (N) uptake components ( $u$ ; lower panels), is determined either by inter-fungal competition (nonshaded area, blue line) or by competition with nonmycorrhizal roots (shaded area), which determines the minimum  $u$  at which a mycorrhizal plant strategy can invade a nonmycorrhizal strategy (dashed orange line) and avoid invasion of a nonmycorrhizal strategy (solid orange line). The mycorrhizal strategy may persist although it reduces plant growth (vertically dashed area) compared with a nonmycorrhizal strategy (light-green dashed line).



**Fig. 4** Model results under ambient CO<sub>2</sub> (solid lines) and elevated CO<sub>2</sub> (dashed lines). (a) Plant net growth (green line) and growth of ectomycorrhizal fungi (EMF; purple line). (b) Plant–fungal nitrogen : carbon (N : C) exchange ratio. (c) N uptake of EMF (purple lines) and N transfer to plants (green lines). Plant–fungi connectance corresponds to Fig. 3(a), i.e. each plant had eight fungal partners ( $n_f = 8$ ) and vice versa ( $n_p = 8$ )

with several fungal partner species increased N uptake and growth at elevated CO<sub>2</sub> more than plants with only one fungal partner (Alberton *et al.*, 2005). Although not modeled, an additional partner may also have a competitive advantage that drives improved N : C exchange rate (for the host) in competition with existing partners. Thus, both fungi and plants should strive to increase the number of partners, which means that connectance should increase concurrently for plants and EMF, until other factors (e.g. grazing; Högberg *et al.*, 2010) limit further network expansion. Increased connectance enhances plant C allocation to EMF growth and shifts the strategy of the EMF toward increased N uptake (Fig. 3). Both these effects increase gross N uptake by EMF, which depletes marginal soil N availability and competitively hampers a nonmycorrhizal strategy. Importantly, this can stabilize the mycorrhizal strategy even when it reduces plant growth compared with a nonmycorrhizal strategy. Thus, while the mycorrhizal strategy allows plants to survive at lower soil N than without mycorrhiza, the symbiosis may reduce plant growth but still persist at higher soil N availability (Fig. 3).

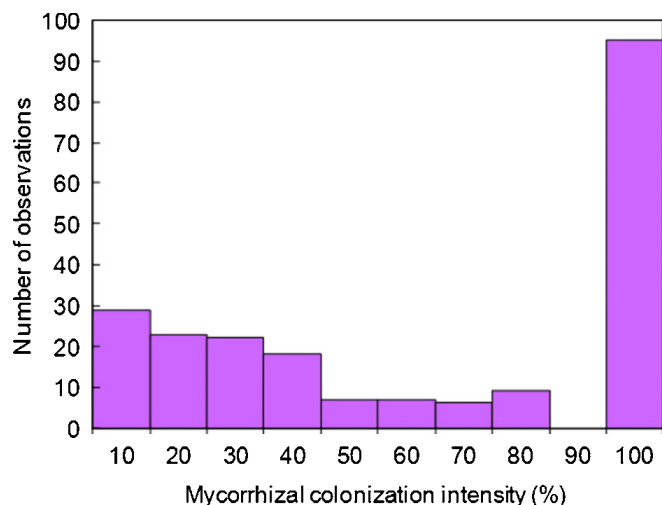
When soil N availability increases to a level where a nonmycorrhizal strategy can invade, EMF only persist if their utility for a plant matches that of nonmycorrhizal roots, which requires EMF to invest more in N uptake (increase  $u$ ) than in the absence of a nonmycorrhizal strategy (Fig. 3 versus Fig. S2). However, EMF

will not increase  $u$  more than necessary as this would reduce fitness. For a plant this means a switch from a clear advantage of the mycorrhizal strategy to equal utility of mycorrhizal and nonmycorrhizal roots. At the same time, there exists a  $u$  that allows the mycorrhizal strategy to invade a resident nonmycorrhizal strategy. Thus, both root strategies may coexist under further increasing soil N availability until EMF are forced to allocate all resources to N uptake ( $u \rightarrow 1$ , leaving nothing for reproductive growth), making the symbiosis unviable. Although mutual invasibility and convergence of the utilities of the alternative root strategies do not imply strictly equal frequency, they suggest that, on average, mycorrhizal and nonmycorrhizal roots coexist in similar proportions. These results agree with the patterns typically observed in gradients of increasing soil N availability: while plant growth gradually increases there is almost complete mycorrhizal root colonization under a range of low soil N availability, followed by coexistence of nonmycorrhizal and mycorrhizal roots, and finally disappearance of EMF (Taylor *et al.*, 2000; Högberg *et al.*, 2003; Nilsson *et al.*, 2005; Kjoller *et al.*, 2012). A corresponding negative trend was also found for soil mycelia where ectomycorrhizal species declined and disappeared (Högberg *et al.*, 2014). Furthermore, because the increasing fungal allocation to N uptake components occurs at the expense of reproductive allocation (increased  $u$ ), it agrees with empirically observed reductions in fruiting-body (reproductive structures) production at high N additions (Peter *et al.*, 2001; Hasselquist *et al.*, 2012). Although many factors influence at which point the plants switch between a pure mycorrhizal strategy and a mixed strategy with both mycorrhizal and nonmycorrhizal roots, the model implies that along environmental gradients there should always be abrupt shifts between very high ( $\approx 100\%$ ) mycorrhizal colonization and much lower colonization. Although not supported by all studies (e.g. Børja & Nilsen, 2009) this threshold behavior agrees with observations of ectomycorrhizal colonization intensity across a wide-ranging data set comprising 125 species and 44 sites (Akhmetzhanova *et al.*, 2012), which were clustered at very high or relatively low values (Fig. 5).

## Discussion

### Scope and limitations

We have shown that market mechanisms can shape the strategies of plants and mycorrhizal fungi to stabilize the mycorrhizal symbiosis under a wide range of soil N availability. Whereas the model formulation is general enough to encompass ectomycorrhizal symbiosis as well as arbuscular mycorrhizal (AM) symbiosis, we have chosen to focus the model evaluation on N-limited ectomycorrhizal forest. The main reason for this focus is the dominant role that the symbiosis plays in these ecosystems in terms of N and C cycling (e.g. Näsholm *et al.*, 2013). Hypothetically, many of our results should be qualitatively valid also for AM symbiosis under nutrient (N or phosphorus)-limited conditions. However, the more open N cycle (less N limitation) in forests with predominantly AM symbiosis (Phillips *et al.*, 2013) suggests that the stabilization of symbiosis via nutrient depletion feedback (see 'Implication 2: mutual stabilization of



**Fig. 5** Field measurements of ectomycorrhizal colonization intensity, for 125 species and 44 sites in the former Soviet Union (Akhmetzhanova *et al.*, 2012), show high frequency of observations at very high or relatively low colonization intensity and fewer observations at intermediate colonization intensity, as predicted by our model. Statistical testing by *k*-means cluster analysis separated two clusters centered at high (95%) and low (21%) values with  $P < 0.0001$ . The colonization intensity is measured in per cent of maximal colonization intensity, based on the mean number of mycorrhizal root tips per root length for each species at each site, relative to a reference scale (Akhmetzhanova *et al.*, 2012).

nitrogen limitation and ectomycorrhizal symbiosis') is less relevant in AM-dominated ecosystems.

Another difference between mycorrhizal types is that significant amounts of organic N are taken up by EMF in ectomycorrhizal forests whereas inorganic N uptake dominates in AMF (Phillips *et al.*, 2013). A change from an organic to an inorganic N source would increase the N:C ratio of fungal uptake and metabolic pools and would therefore lead to a higher N:C exchange ratio with the plants in our model. This would lead to quantitative shifts in our results and slightly different values of the fitted parameters (uptake capacities) but no qualitative changes in the conclusions.

#### Implication 1: two stability regimes of ectomycorrhizal symbiosis

The model suggests a previously not recognized dichotomy in how the strategies of EMF respond to soil N availability: (1) partner discrimination by plants induces competition for C among their fungal partners that enhances fungal C allocation to N uptake, stabilizing a fungal strategy beneficial for host plants at low N availability, whereas (2) competition (or choice) between mycorrhizal and nonmycorrhizal plant strategies constrains fungal strategy at higher N availability. However, although empirical evidence supports the general importance of competition in structuring EMF communities (Kennedy, 2010), the specific mechanism (1) has been empirically confirmed only for AM symbiosis (Kiers *et al.*, 2011; Fellbaum *et al.*, 2012). Without mechanism (1), that is, plant discrimination among partner fungi, there would be no stabilizing effect of inter-fungal competition at low N availability

and the stability of the symbiosis would rely solely on mechanism (2) ( $z=0$  in Fig. S3). The model would (with some parameter adjustments) still be able to reproduce many observed phenomena, including the effects of soil N availability and elevated CO<sub>2</sub> on N:C exchange rates (Alberton & Kuyper, 2009; Näsholm *et al.*, 2013) and the stronger decline in fruiting-body production than in mycelia of EMF under high N availability (Wallenda & Kottke, 1998; Peter *et al.*, 2001; Hasselquist *et al.*, 2012). However, the necessity of both mechanisms (1) and (2) is supported by the high colonization rates of EMF at low soil N, and the threshold behavior of ectomycorrhizal colonization intensity (Fig. 5), which indicates switching between the two controlling mechanisms. Mechanism (2) alone, that is, absence of intra-party fungal competition, would lead to a continuously declining fungal allocation to N uptake components at decreasing soil N availability, maintaining similar utility of mycorrhizal and nonmycorrhizal plant strategies and thus lower ectomycorrhizal colonization at low N availability than the near 100% commonly observed (Taylor *et al.*, 2000; Fig. 5).

Under the ongoing global rise in N deposition, switching between the two mechanisms delineated above may influence not only the colonization rate and productivity of EMF, but also forest N cycling. While forests growing at high and very low N availabilities should experience gradual change in response to N deposition, trees growing at intermediate N availability may suddenly reduce mycorrhizal colonization (as a result of switching of stabilizing mechanisms from (1) to (2)), which reduces N retention and may induce ecosystem N leaching (Bahr *et al.*, 2013; Högborg *et al.*, 2013).

#### Implication 2: mutual stabilization of nitrogen limitation and ectomycorrhizal symbiosis

In addition to the primary mechanisms (1) and (2), the model postulates that mechanism (1) gives rise to a secondary stabilizing mechanism: stabilization of the mycorrhizal strategy via reduction of marginal N availability (N uptake gain per additional root mass), that is, environmental resource feedback (Fig. 6). Thus, the mycorrhizal symbiosis creates its own environment where it outcompetes nonmycorrhizal plant strategies. Its potential to stabilize mycorrhizal symbiosis even when it reduces plant growth suggests that it may be responsible for the large fraction of negative effects among observations of mycorrhizal effects on plant growth (Corrêa *et al.*, 2008, 2012). Importantly, the feedback stabilization of N limitation and mycorrhizal symbiosis under low N availability suggests that it underpins the syndrome of low productivity and mycorrhizal dominance in boreal forests (Högborg *et al.*, 2011; Näsholm *et al.*, 2013). This feedback effect would be exacerbated if EMF contribute to continuing soil N immobilization, as suggested by the increase in natural abundance of <sup>15</sup>N with increasing soil depth observed in ectomycorrhizal forests (Högborg *et al.*, 1996; Billings & Richter, 2006; Hobbie & Ouimet, 2009). Although the full extent of the feedback stabilization hypothesis is not easily tested experimentally, the link between high N retention and EMF has been established in plots recovering from long-term N addition (Högborg *et al.*, 2011, 2014).

### Implication 3: ectomycorrhizal symbiosis hampers CO<sub>2</sub> fertilization of tree growth

The model suggest that N transfer to plants and resulting plant net growth (growth minus litter production) is enhanced by elevated CO<sub>2</sub> at high soil N availability but not at low N availability, where it may even decrease (Fig. 4). Thus, only at sufficiently high N availability will trees in ectomycorrhizal forests increase stem growth under rising CO<sub>2</sub>. This suggests that mycorrhizal symbiosis may make the strongly N-limited boreal forests particularly susceptible to progressive N limitation postulated as a consequence of rising atmospheric CO<sub>2</sub> (Luo *et al.*, 2004; Alberton *et al.*, 2007). The underlying reason for this is that, at low soil N availability, the increase in tree C export to EMF in response to elevated CO<sub>2</sub> enhances N immobilization in EMF biomass more than N export to trees, which is reflected in a reduced N : C exchange rate (Fig. 4b). Thus, the trees generate a self-imposed N limitation by excessive C export to EMF in response to elevated CO<sub>2</sub>. Although this may seem highly maladaptive, from an individual tree's perspective it is an adaptive response (increasing fitness) to competition for N among trees. Just like in the stabilization of a nonbeneficial mycorrhizal plant strategy discussed above (Implication 2: mutual stabilization of nitrogen limitation and ectomycorrhizal symbiosis), individual fitness maximization leads to a collective loss, that is, a tragedy-of-the-commons effect.

### Theoretical implications in a wider context: intra-party competition stabilizes symbiosis for better or worse

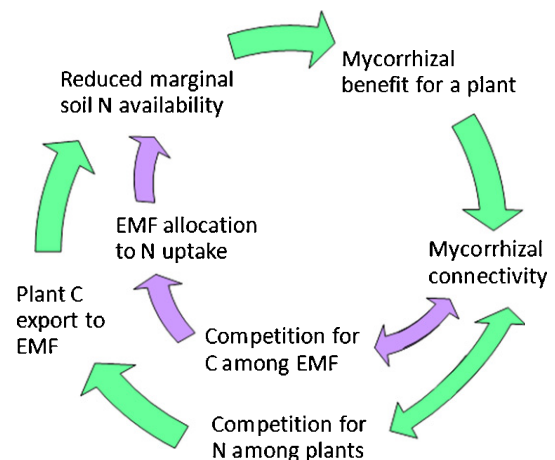
The importance of intra-party competition in a market context (out-bidding) for the stability of mutualism has long been recognized (Noë & Hammerstein, 1994; Jones *et al.*, 2012; Werner *et al.*, 2014). This principle has been applied to mycorrhizal symbiosis, showing how a single plant should optimize the use of multiple fungal partners (Kummel & Salant, 2006). Our model extends this theory by accounting for adaptive strategies in both parties of the symbiosis, quantifying the effects of partner numbers, and exploring its potential to stabilize nonmutualistic symbiosis. Because the beneficial effect for a host of intra-party competition among its partners increases with the number of partners (Fig. 3), this mechanism may drive the observed positive relationship between connectance and stability in mutualistic networks; that is, a species' chance of survival increases with the number of partners (James *et al.*, 2012). Similarly, the mechanism may contribute to the observed increase in productivity with diversity of mycorrhizal fungi (van der Heijden *et al.*, 1998), because species diversity should correlate with the number of individuals and the range of fungal strategies that compete for plant C. However, while mycorrhizal networks have been viewed as a facilitating 'socialism' in terms of nutrient sharing among plants (Van Der Heijden & Horton, 2009), in our theory they rather promote 'capitalism', by enhancing competition among profit-maximizing trading individuals. For example, when different plants share a fungal partner delivering N, the allocation of N to each plant is driven by the fungus' strategy to maximize C

return rather than by a plant behavior to share N with other plants. Thus, perhaps expectedly, the resulting increased trading is not always beneficial for both parties of the trade. Whereas partner competition enhances resource transfer to a host, our model shows that the resulting increase in resource depletion can stabilize mycorrhizal symbiosis even if a nonmycorrhizal strategy would have been more productive for the plants. More generally, this implies that resource feedback can drive inherently mutualistic (trading) entities into nonmutualistic cooperation (Figs 3a,b, 6). This resource feedback stabilization of a suboptimal strategy has interesting parallels in other market contexts, such as lock-in of inferior energy technologies (Unruh, 2000), highlighting the cross-disciplinary relevance of such mechanisms.

### Conclusions and way forward

While recent theoretical progress on mycorrhizal symbiosis mainly has focused on the evolution and stability of mutualism in pair-wise interactions, empirical research has revealed a picture of multiple simultaneous and highly dynamic interactions. Here we have taken a step to extend theory in this direction, based on a model that links the strategy (governed by evolutionary principles) of each individual to its multiple exchangeable trading partners and competitors in a network.

The model is able to reproduce a wide range of observed responses to N and atmospheric CO<sub>2</sub> in ectomycorrhizal forest, indicating that it captures relevant mechanisms despite its highly simplified representations of ecophysiology and functional diversity of EMF. However, more empirical research is needed to allow further evaluation of the hypotheses generated and some of the model's assumptions. A central assumption is that a host is able to discriminate and induce competition among its partners, which



**Fig. 6** Feedback loop stabilizing the mycorrhizal strategy, showing the interrelation between effects of ectomycorrhizal fungi (EMF; purple arrows) and plants (green arrows). Increased fungal-plant (mycorrhizal) connectivity enhances competition among both EMF and plants, which drives plants to increase carbon (C) export to EMF and EMF to increase nitrogen (N) uptake and N export to plants. These effects interact in depleting available soil N, which favors the mycorrhizal strategy compared with a nonmycorrhizal plant strategy, which in turn promotes mycorrhizal connectivity.

benefits the host and promotes symbiotic stability. While such partner discrimination has been confirmed for AM symbiosis, empirical quantification and investigation of its role in ectomycorrhizal symbiosis are urgently needed. Furthermore, while plant allocation responses to resource availability are relatively well known, the importance of corresponding acclimation/adaptation of fungal strategies indicated by our results is much less explored. Progress in understanding and modeling of mycorrhizal ecosystems would strongly benefit from exploration of additional traits and trade-offs delineating functional/strategic diversity in EMF, including, for example, life-span, biomass N : C ratio, and enzyme production.

The results suggest that persistence of ectomycorrhizal symbiosis in boreal forests emerges at the ecosystem level, via positive feedback between low nutrient availability and stability of the mycorrhizal plant strategy. This mechanism may also have the capacity to eliminate the expected positive effect of rising CO<sub>2</sub> on tree growth or even reverse it to a negative effect in strongly N-limited boreal forests. The potential significance of these results for the global C cycle underlines the need for an ecosystem perspective in understanding mycorrhizal symbiosis and the importance of including the effects of mycorrhizal symbiosis in applied forest and vegetation models.

## Acknowledgements

This study was financed by grants to T.N., M.N.H. and P.H. from the Kempe foundations, Swedish University of Agricultural Sciences (TC4F) and the research councils: The Swedish Research Council for Environment, Agricultural Sciences and Spatial Planning, the Swedish Governmental Agency for Innovation Systems, and The Swedish Research Council. Funding was received from the European Research Council under the European Union's Seventh Framework Programme (FP/2007–2013/ERC Grant Agreement no. 610028). We thank Jan Parsby, Thomas Hörnlund, Mark Blackburn and Stephan Schaffner for technical assistance, and Henrik Holmgren for access to his land. We thank Daniel Metcalfe, Sonja G. Keel, Catherine Campbell, Vaughan Hurry, and Sune Linder for scientific contributions. We thank Thomas W. Kuyper and two anonymous reviewers for important contributions.

## References

- Akhmetzhanova AA, Soudzilovskaia NA, Onipchenko VG, Cornwell WK, Agafonov VA, Selivanov IA, Cornelissen JH. 2012. A rediscovered treasure: mycorrhizal intensity database for 3000 vascular plant species across the former Soviet Union: Ecological Archives E093-059. *Ecology* 93: 689–690.
- Alberton O, Kuyper TW. 2009. Ectomycorrhizal fungi associated with *Pinus sylvestris* seedlings respond differently to increased carbon and nitrogen availability: implications for ecosystem responses to global change. *Global Change Biology* 15: 166–175.
- Alberton O, Kuyper TW, Gorissen A. 2005. Taking myco-centrism seriously: mycorrhizal fungal and plant responses to elevated CO<sub>2</sub>. *New Phytologist* 167: 859–868.
- Alberton O, Kuyper TW, Gorissen A. 2007. Competition for nitrogen between *Pinus sylvestris* and ectomycorrhizal fungi generates potential for negative feedback under elevated CO<sub>2</sub>. *Plant and Soil* 296: 159–172.
- Averill C, Turner BL, Finzi AC. 2014. Mycorrhiza-mediated competition between plants and decomposers drives soil carbon storage. *Nature*. doi: 10.1038/nature12901.
- Bahr A, Ellström M, Akselsson C, Ekblad A, Mikusinska A, Wallander H. 2013. Growth of ectomycorrhizal fungal mycelium along a Norway spruce forest nitrogen deposition gradient and its effect on nitrogen leakage. *Soil Biology and Biochemistry* 59: 38–48.
- Beiler KJ, Durall DM, Simard SW, Maxwell SA, Kretzer AM. 2010. Architecture of the wood-wide web: *Rhizopogon* spp. genets link multiple Douglas-fir cohorts. *New Phytologist* 185: 543–553.
- Billings SA, Richter DD. 2006. Changes in stable isotopic signatures of soil nitrogen and carbon during 40 years of forest development. *Oecologia* 148: 325–333.
- Børja I, Nilsen P. 2009. Long term effect of liming and fertilization on ectomycorrhizal colonization and tree growth in old Scots pine (*Pinus sylvestris* L.) stands. *Plant and Soil* 314: 109–119.
- Corrêa A, Gurevitch J, Martins-Loução MA, Cruz C. 2012. C allocation to the fungus is not a cost to the plant in ectomycorrhizae. *Oikos* 121: 449–463.
- Corrêa A, Strasser RJ, Martins-Loução MA. 2008. Response of plants to ectomycorrhizae in N-limited conditions: which factors determine its variation? *Mycorrhiza* 18: 413–427.
- Cox F, Barsoum N, Lilleskov EA, Bidartondo MI. 2010. Nitrogen availability is a primary determinant of conifer mycorrhizas across complex environmental gradients. *Ecology Letters* 13: 1103–1113.
- Dieleman WIJ, Luyssaert S, Rey A, De Angelis P, Barton CVM, Broadmeadow MSJ, Broadmeadow SB, Chigwerewe KS, Crookshanks M, Dufrene E *et al.* 2010. Soil [N] modulates soil C cycling in CO<sub>2</sub>-fumigated tree stands: a meta-analysis. *Plant, Cell & Environment* 33: 2001–2011.
- Eaton GK, Ayres MP. 2002. Plasticity and constraint in growth and protein mineralization of ectomycorrhizal fungi under simulated nitrogen deposition. *Mycologia* 94: 921–932.
- egger KN, Hibbett DS. 2004. The evolutionary implications of exploitation in mycorrhizas 1. *Canadian Journal of Botany* 82: 1110–1121.
- Ekblad A, Wallander H, Godbold DL, Cruz C, Johnson D, Baldrian P, Björk RG, Epron D, Kieliszewska-Rokicka B, Kjoller R *et al.* 2013. The production and turnover of extramatrical mycelium of ectomycorrhizal fungi in forest soils: role in carbon cycling. *Plant and Soil* 366: 1–27.
- Fellbaum CR, Gachomo EW, Beesetty Y, Choudhari S, Strahan GD, Pfeiffer PE, Kiers ET, Bücking H. 2012. Carbon availability triggers fungal nitrogen uptake and transport in arbuscular mycorrhizal symbiosis. *Proceedings of the National Academy of Sciences, USA* 109: 2666–2671.
- Foley JA, Kutzbach JE, Coe MT, Levis S. 1994. Feedbacks between climate and boreal forests during the Holocene epoch. *Nature* 371: 52–54.
- Franklin O. 2007. Optimal nitrogen allocation controls tree responses to elevated CO<sub>2</sub>. *New Phytologist* 174: 811–822.
- Franklin O, Johansson J, Dewar RC, Dieckmann U, McMurtrie RE, Brännström Å, Dybzinski R. 2012. Modeling carbon allocation in trees: a search for principles. *Tree Physiology* 32: 648–666.
- Fransson PMA, Taylor AFS, Finlay RD. 2005. Mycelial production, spread and root colonisation by the ectomycorrhizal fungi *Hebeloma crustuliniforme* and *Paxillus involutus* under elevated atmospheric CO<sub>2</sub>. *Mycorrhiza* 15: 25–31.
- Garcia MO, Ovasapyan T, Greas M, Treseder KK. 2008. Mycorrhizal dynamics under elevated CO<sub>2</sub> and nitrogen fertilization in a warm temperate forest. *Plant and Soil* 303: 301–310.
- Grman E, Robinson TMP, Klausmeier CA. 2012. Ecological specialization and trade affect the outcome of negotiations in mutualism. *American Naturalist* 179: 567–581.
- Hasselquist NJ, Metcalfe DB, Högberg P. 2012. Contrasting effects of low and high nitrogen additions on soil CO<sub>2</sub> flux components and ectomycorrhizal fungal sporocarp production in a boreal forest. *Global Change Biology* 18: 3596–3605.
- van der Heijden MGA, Klironomos JN, Ursic M, Moutoglis P, Streitwolf-Engel R, Boller T, Wiemken A, Sanders IR. 1998. Mycorrhizal fungal diversity determines plant biodiversity, ecosystem variability and productivity. *Nature* 396: 69–72.



- Hobbie EA, Ouimette AP. 2009. Controls of nitrogen isotope patterns in soil profiles. *Biogeochemistry* 95: 355–371.
- Hodge A, Fitter AH. 2010. Substantial nitrogen acquisition by arbuscular mycorrhizal fungi from organic material has implications for N cycling. *Proceedings of the National Academy of Sciences, USA* 107: 13754–13759.
- Hoeksema JD, Schwartz MW. 2003. Expanding comparative-advantage biological market models: contingency of mutualism on partners' resource requirements and acquisition trade-offs. *Proceedings of the Royal Society B: Biological Sciences* 270: 913–919.
- Högberg P, Högberg MN, Göttlicher SG, Betson NR, Keel SG, Metcalfe DB, Campbell C, Schindlbacher A, Hurry V, Lundmark T *et al.* 2008. High temporal resolution tracing of photosynthate carbon from the tree canopy to forest soil microorganisms. *New Phytologist* 177: 220–228.
- Högberg P, Högberg L, Schinkel H, Högberg M, Johannisson C, Wallmark H. 1996. <sup>15</sup>N abundance of surface soils, roots and mycorrhizas in profiles of European forest soils. *Oecologia* 108: 207–214.
- Högberg P, Johannisson C, Yarwood S, Callesen I, Näsholm T, Myrold DD, Högberg MN. 2011. Recovery of ectomycorrhiza after 'nitrogen saturation' of a conifer forest. *New Phytologist* 189: 515–525.
- Högberg MN, Bååth E, Nordgren A, Arnebrant K, Högberg P. 2003. Contrasting effects of nitrogen availability on plant carbon supply to mycorrhizal fungi and saprotrophs – a hypothesis based on field observations in boreal forest. *New Phytologist* 160: 225–238.
- Högberg MN, Briones MJI, Keel SG, Metcalfe DB, Campbell C, Midwood AJ, Thornton B, Hurry V, Linder S, Näsholm T *et al.* 2010. Quantification of effects of season and nitrogen supply on tree below-ground carbon transfer to ectomycorrhizal fungi and other soil organisms in a boreal pine forest. *New Phytologist* 187: 485–493.
- Högberg MN, Högberg L, Kleja DB. 2013. Soil microbial community indices as predictors of soil solution chemistry and N leaching in *Picea abies* (L.) Karst. forests in S. Sweden. *Plant and Soil* 372: 507–522.
- Högberg MN, Yarwood SA, Myrold DD. 2014. Fungal but not bacterial communities recover after termination of decadal nitrogen additions to boreal forest. *Soil Biology and Biochemistry* 72: 35–43.
- James A, Pitchford JW, Plank MJ. 2012. Disentangling nestedness from models of ecological complexity. *Nature* 487: 227–230.
- Jarvis P, Linder S. 2000. Constraints to growth of boreal forests. *Nature* 405: 904–905.
- Jennings D. 1995. *The physiology of fungal nutrition*. Cambridge, UK: Cambridge University Press.
- Johnson NC, Angelard C, Sanders IR, Kiers ET. 2013. Predicting community and ecosystem outcomes of mycorrhizal responses to global change. *Ecology Letters* 16(Suppl. 1): 140–153.
- Jones EI, Bronstein JL, Ferrière R. 2012. The fundamental role of competition in the ecology and evolution of mutualisms. *Annals of the New York Academy of Sciences* 1256: 66–88.
- Jones MD, Smith SE. 2004. Exploring functional definitions of mycorrhizas: are mycorrhizas always mutualisms? *Canadian Journal of Botany* 82: 1089–1109.
- Keel SG, Campbell CD, Högberg MN, Richter A, Wild B, Zhou X, Hurry V, Linder S, Näsholm T, Högberg P. 2012. Allocation of carbon to fine root compounds and their residence times in a boreal forest depend on root size class and season. *New Phytologist* 194: 972–981.
- Kennedy P. 2010. Ectomycorrhizal fungi and interspecific competition: species interactions, community structure, coexistence mechanisms, and future research directions. *New Phytologist* 187: 895–910.
- Kiers ET, Duhamel M, Beesetty Y, Mensah JA, Franken O, Verbruggen E, Fellbaum CR, Kowalchuk GA, Hart MM, Bago A *et al.* 2011. Reciprocal rewards stabilize cooperation in the mycorrhizal symbiosis. *Science* 333: 880–882.
- Kjøller R, Nilsson LO, Hansen K, Schmidt IK, Vesterdal L, Gundersen P. 2012. Dramatic changes in ectomycorrhizal community composition, root tip abundance and mycelial production along a stand-scale nitrogen deposition gradient. *New Phytologist* 194: 278–286.
- Kummel M, Salant SW. 2006. The economics of mutualisms: optimal utilization of mycorrhizal mutualistic partners by plants. *Ecology* 87: 892–902.
- Lekberg Y, Hammer EC, Olsson PA. 2010. Plants as resource islands and storage units – adopting the mycocentric view of arbuscular mycorrhizal networks. *FEMS Microbiology Ecology* 74: 336–345.
- Luo Y, Su B, Currie WS, Dukes JS, Finzi A, Hartwig U, Hungate B, McMurtrie RE, Oren R, Parton WJ *et al.* 2004. Progressive nitrogen limitation of ecosystem responses to rising atmospheric carbon dioxide. *BioScience* 54: 731–739.
- Luoma S. 1997. Geographical pattern in photosynthetic light response of *Pinus sylvestris* in Europe. *Functional Ecology* 11: 273–281.
- de Mazancourt C, Schwartz MW. 2010. A resource ratio theory of cooperation. *Ecology Letters* 13: 349–359.
- Mikusinska A, Persson T, Taylor AFS, Ekblad A. 2013. Response of ectomycorrhizal extramatrical mycelium production and isotopic composition to in-growth bag size and soil fauna. *Soil Biology and Biochemistry* 66: 154–162.
- Näsholm T, Högberg P, Franklin O, Metcalfe D, Keel SG, Campbell C, Hurry V, Linder S, Högberg MN. 2013. Are ectomycorrhizal fungi alleviating or aggravating nitrogen limitation of tree growth in boreal forests? *New Phytologist* 198: 214–221.
- Nilsson LO, Giesler R, Bååth E, Wallander H. 2005. Growth and biomass of mycorrhizal mycelia in coniferous forests along short natural nutrient gradients. *New Phytologist* 165: 613–622.
- Noë R, Hammerstein P. 1994. Biological markets: supply and demand determine the effect of partner choice in cooperation, mutualism and mating. *Behavioral Ecology and Sociobiology* 35: 1–11.
- Oren R, Ellsworth DS, Johnsen KH, Phillips N, Ewers BE, Maier C, Schafer KVR, McCarthy H, Hendrey G, McNulty SG *et al.* 2001. Soil fertility limits carbon sequestration by forest ecosystems in a CO<sub>2</sub>-enriched atmosphere. *Nature* 411: 469–472.
- Peter M, Ayer F, Egli S. 2001. Nitrogen addition in a Norway spruce stand altered macromycete sporocarp production and below-ground ectomycorrhizal species composition. *New Phytologist* 149: 311–325.
- Phillips RP, Brzostek E, Midgley MG. 2013. The mycorrhizal-associated nutrient economy: a new framework for predicting carbon-nutrient couplings in temperate forests. *New Phytologist* 199: 41–51.
- Pickles BJ, Genney DR, Potts JM, Lennon JJ, Anderson IC, Alexander IJ. 2010. Spatial and temporal ecology of Scots pine ectomycorrhizas. *New Phytologist* 186: 755–768.
- Post WM, Emanuel WR, Zinke PJ, Stangenberger AG. 1982. Soil carbon pools and world life zones. *Nature* 298: 156–159.
- Pringle A, Taylor JW. 2002. The fitness of filamentous fungi. *Trends in Microbiology* 10: 474–481.
- Smith SE, Read DJ. 2008. *Mycorrhizal symbiosis*. Cambridge, UK: Academic Press.
- Southworth D, He XH, Swenson W, Bledsoe CS, Horwath WR. 2005. Application of network theory to potential mycorrhizal networks. *Mycorrhiza* 15: 589–595.
- Taylor AFS, Martin F, Read DJ. 2000. Fungal diversity in ectomycorrhizal communities of Norway spruce [*Picea abies* (L.) Karst.] and beech (*Fagus sylvatica* L.) along north-south transects in Europe. *Ecological Studies* 142: 343–365.
- Unruh GC. 2000. Understanding carbon lock-in. *Energy Policy* 28: 817–830.
- Van Der Heijden MGA, Horton TR. 2009. Socialism in soil? The importance of mycorrhizal fungal networks for facilitation in natural ecosystems. *Journal of Ecology* 97: 1139–1150.
- Walder F, Niemann H, Natarajan M, Lehmann MF, Boller T, Wiemken A. 2012. Mycorrhizal networks: common goods of plants shared under unequal terms of trade. *Plant Physiology* 159: 789–797.
- Wallenda T, Kottke I. 1998. Nitrogen deposition and ectomycorrhizas. *New Phytologist* 139: 161–168.
- Werner GD, Strassmann JE, Ivens AB, Engelmoer DJ, Verbruggen E, Queller DC, Noë R, Johnson NC, Hammerstein P, Kiers ET. 2014. Evolution of microbial markets. *Proceedings of the National Academy of Sciences, USA* 111: 1237–1244.

## Supporting Information

Additional supporting information may be found in the online version of this article.

**Fig. S1** Comparison of nitrogen uptake for roots and mycorrhizal fungi.

**Fig. S2** Modeled effects of mycorrhizal symbiosis on trees and mycorrhizal fungi when trees lack the option of a nonmycorrhizal strategy.

**Fig. S3** Modeled effects of variation in the fungal ability to discriminate among plant partners (parameter  $z$ ) on productivity and symbiotic stability in the forest.

**Table S1** Model parameters and variables

**Methods S1** Full model description.

Please note: Wiley Blackwell are not responsible for the content or functionality of any supporting information supplied by the authors. Any queries (other than missing material) should be directed to the *New Phytologist* Central Office.



## About *New Phytologist*

- *New Phytologist* is an electronic (online-only) journal owned by the New Phytologist Trust, a **not-for-profit organization** dedicated to the promotion of plant science, facilitating projects from symposia to free access for our Tansley reviews.
- Regular papers, Letters, Research reviews, Rapid reports and both Modelling/Theory and Methods papers are encouraged. We are committed to rapid processing, from online submission through to publication 'as ready' via *Early View* – our average time to decision is <25 days. There are **no page or colour charges** and a PDF version will be provided for each article.
- The journal is available online at Wiley Online Library. Visit **www.newphytologist.com** to search the articles and register for table of contents email alerts.
- If you have any questions, do get in touch with Central Office (np-centraloffice@lancaster.ac.uk) or, if it is more convenient, our USA Office (np-usaoffice@ornl.gov)
- For submission instructions, subscription and all the latest information visit **www.newphytologist.com**

## Supporting Information Methods S1, Table S1, Figs S1–S3

## Methods S1: Mathematical model description

Table S1. Model variables and parameters.

Symbol	Value	Unit	Description
<i>dependent variables</i>			
$B_r, B_f, B_{fu}$		$\text{gC m}^{-2}$	Biomass of fine-roots, EMF, and EMF N uptake components
$C_r, C_f$		$\text{gC m}^{-2} \text{d}^{-1}$	C use for fine-root growth, and C export to EMF
$F$		-	Fraction resource obtained by one individual among competitors
$G$		$\text{gC m}^{-2} \text{d}^{-1}$	Net plant growth (GPP – respiration – litter production)
$G_f$		$\text{gC m}^{-2} \text{d}^{-1}$	Net EMF reproductive production
$N_c, N_r$		$\text{gN m}^{-2} \text{d}^{-1}$	N in canopy and fine-roots
$N_p$		$\text{gN m}^{-2}$	Plant N uptake
$P$		$\text{gC m}^{-2} \text{d}^{-1}$	Net canopy C uptake (photosynthesis)
$P_f$		$\text{gC m}^{-2} \text{d}^{-1}$	EMF biomass production
$U$		$\text{gN m}^{-2} \text{d}^{-1}$	N uptake by roots or hyphae
$U_c$		$\text{gN m}^{-2} \text{d}^{-1}$	N uptake capacity of fine-roots or EMF
$u$		-	EMF biomass fraction contributing to N uptake
$W$		$\text{gC m}^{-2} \text{d}^{-1}$	Plant C costs of respiration and litter production
$x$		$\text{gN gC}^{-1}$	EMF-Plant N:C exchange rate
<i>independent variables and parameters</i>			
$a$	0.14	$\text{gC gN}^{-1} \text{h}^{-1}$	Light saturated photosynthesis per canopy N (Luoma, 1997)
$c_c, c_r, c_G, c_f$	0.03, 0.03, 0.01, 0.085	$\text{gN gC}^{-1}$	N:C ratio of foliage (Luoma, 1997), fine-roots, plant net growth, and EMF (Mikusinska <i>et al.</i> , 2013)
$d$	0.25	m	Effective soil depth
$e_u$	0.5, 0.0017	$\text{m}^3 \text{gC}^{-1}$	Soil volume explored per EMF uptake components and fine-root biomass, respectively <sup>#</sup>
$f_r$	1	-	Minimum fine-root production per growth of EMF (Hasselquist <i>et al.</i> , 2012)
$N_{av}$		$\text{gN m}^{-3} \text{d}^{-1}$	Soil N availability = maximal N uptake per soil volume
$h$	12	h	day length in hours
$n_p, n_f$		-	Number of competitors for plants and EMF, respectively. Examples of observed values range from 1 -20 (Southworth <i>et al.</i> , 2005; Beiler <i>et al.</i> , 2010)
$Q$	1.5	$\text{gC h}^{-1}$	N saturated photosynthetic capacity = photosynthetic quantum efficiency (Wong <i>et al.</i> , 1979) times photosynthetically active radiation.
$t_o, t_r, t_f$	800, 400, 17	d	Lifespan of foliage <sup>*</sup> , fine-roots (Keel <i>et al.</i> , 2012), and EMF (Högberg <i>et al.</i> , 2008; Högberg <i>et al.</i> , 2010), respectively
$u_{cB}$	0.1	$\text{gN gC}^{-1} \text{d}^{-1}$	N saturated N uptake capacity per root or EMF biomass <sup>#</sup>
$w$	0.25	$\text{gC gN}^{-1} \text{d}^{-1}$	Plant litter and respiration C costs per canopy N <sup>#</sup>
$y_f$	0.3	$\text{gC gC}^{-1}$	Fungal net C use efficiency (Sinsabaugh <i>et al.</i> , 2013)
$Y_p$	0.7	$\text{gC gC}^{-1}$	Plant growth C conversion efficiency (Choudhury, 2001)
$z$			Exponent of the partner discrimination relationship

<sup>#</sup> calibrated to match measured data (see below- Model parameterization, and Fig. 2)

<sup>\*</sup> typical values in boreal pine forest were used. All rate values refer to the growing season (e.g.  $d$  means growing season days) spanning 150 days.

## Nitrogen uptake

Gross N uptake (both for fine-roots and ectomycorrhizal fungi (EMF);  $U$ ; eq. 1) is determined by maximal (N saturated) uptake capacity of the whole root system or EMF population ( $U_c$ , eq. 2), soil N availability ( $N_{av}$ ) and soil depth ( $d$ ).

$$U = \frac{U_c N_{av} d}{U_c + N_{av} d} \quad (1)$$

$U_c$  in turn depends on the biomass ( $B$ ) of fine-roots or fungal uptake components, the specific maximal N uptake capacities ( $u_{cB}$ ) and the specific maximum N uptake efficiency at low soil N availability ( $e_u$ ), which is equivalent to the soil volume accessed per  $B$ .

$$U_c = B \frac{u_{cB} N_{av} e_u}{u_{cB} + N_{av} e_u} \quad (2)$$

In terms of N uptake, fine-roots and hyphae of EMF differ only in their specific N uptake efficiency ( $e_u$ , eq. 2), which is much higher for hyphae than fine-roots. This difference combined with the higher turnover (shorter life-span) of EMF compared to fine-roots mean that mycorrhizal N uptake is more cost efficient at low than at high soil N availability (Fig. S1).

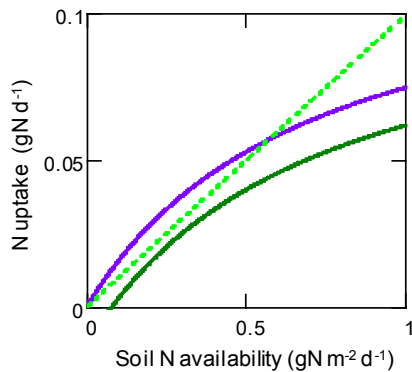


Figure. S1. Modeled nitrogen (N) uptake by ectomycorrhizal fungi (EMF, solid purple line), N transfer to mycorrhizal plants (solid green line), and N uptake of a non-mycorrhizal plant (dashed green line) for an equal fixed amount of C ( $= 0.5 \text{ g m}^{-2} \text{ d}^{-1}$ ) invested in the root system.

## Plant physiology

To model the interaction between plants and EMF, plant growth and allocation is expressed in terms of carbon export to EMF ( $C_f$ ) and N uptake ( $N_p$ ) via EMF or directly from soil.  $C_f$  is optimized to maximize net growth ( $G_p$ , including biomass increase and reproductive production), which is a reasonable fitness proxy (Franklin *et al.*, 2012). We calculate  $G_p$  based on a C flux balance equation (eq. 3) expressed in terms of photosynthesis ( $P$ , eq. 4) minus C costs due to maintenance respiration and litter production ( $W$ , eq. 5), and  $C_f$ . Both  $P$  and  $W$  are functions of N, which links the C balance equations to the N balance equation (eq. 6). This model corresponds to the forest stand model developed in (Franklin *et al.*, 2012) with the addition of C export to EMF.

$$G_p = y_p (P - W - C_f) \quad (3)$$

$$P = h \frac{a N_c Q}{a N_c + Q} \quad (4)$$

$$W = w (N_c + N_r) \quad (5)$$

$$N_p = \frac{N_c}{t_c} + \frac{N_r}{t_r} + c_G G_p \quad (6)$$

$$N_r = B_r c_r, \text{ where } B_r = B_{r0} + B_{rf} \quad (7)$$

$$B_{r0} = C_{r0} t_r \quad (8)$$

$$B_{rf} = f_r C_f y_f t_f \quad (9)$$

Eq. 4 means that  $P$  is limited by radiation ( $Q$ = photosynthetically active radiation  $\times$  photosynthetic quantum efficiency), canopy  $N$  ( $N_c$ ) and day-length ( $h$ ). The parameter  $a$  is the light saturated photosynthetic capacity per leaf  $N$ . In eq.5,  $w$  is respiration and litter production per  $N$  in foliage ( $N_c$ ) and fine-roots ( $N_r$ ). In eq. 6,  $N_p$  is plant  $N$  uptake,  $t_c$  and  $t_r$  are life-spans of foliage and fine-roots, and  $c_G$  is the  $N:C$  ratio of net growth. In eq. 7,  $B_r$  is the biomass of fine-roots, which includes non-mycorrhizal roots ( $B_{r0}$ , eq. 8) and mycorrhizal roots ( $B_{rf}$ , eq. 9). We assume that  $B_{rf}$  is a constant factor ( $f_r$ ) of the hosted fungal growth, which in turn is a function of  $C$  export to EMF and their growth  $C$  efficiency,  $y_f$ .

Using eqs. 3-9,  $G_p$  can be expressed analytically (expression too large to show here) in terms of fixed parameters and the variables nitrogen uptake ( $N_p$ ),  $C$  export to EMF ( $C_f$ ), and  $C$  export to non-mycorrhizal roots ( $C_{r0}$ ) (eq. 10).

$$G_p = f[C_f, C_{r0}, N_p] \quad (10)$$

### Fungal physiology

Fungal biomass production ( $P_f$ , eq. 11) is modeled as a function of plant  $C$  export ( $C_f$ ) as done in (Näsholm *et al.*, 2013), with the addition that fungal biomass ( $B_f$ ) is divided into components for  $N$  uptake ( $B_{fu}$ ) and reproductive growth, which is produced at a rate  $G_{fu}$  (eq. 11). The fraction of uptake components ( $u$ ) relative to reproductive growth ( $1-u$ ) defines the strategy of a fungus.

$$G_f = P_f (1-u), \text{ where } P_f = y_f C_f \quad (11)$$

$$B_{fu} = B_f u, \text{ where } B_f = P_f t_f \quad (12)$$

In eq. 12,  $t_f$  is the life-span of EMF.

EMF take up  $N$  as a function of  $B_{fu}$  ( $U[B_{fu}]$ , eq. 13) of which a part is used for their own growth and the rest ( $N_p$ ) is exported to the plants (Näsholm *et al.*, 2013).

$$N_p = U[B_{fu}] - P_f c_f \quad (13)$$

In eq. 13,  $c_f$  is fungal biomass  $N:C$  ratio.

The representation of fungal growth in terms of only two components was made to focus on the most relevant strategic component of fungal adaptation to  $N$  availability (Eaton & Ayres, 2002). Any other structures or functions responding to  $N$  and  $C$  availability, e.g. structural differences and capabilities to use organic  $N$  sources (Lilleskov *et al.*, 2011), are implicitly subsumed in the one of the two focal components to which they contribute the most and/or in the in the response of  $N$  uptake capacity to soil  $N$  availability ( $U_c$ , eq. S2).

Although fungal growth is assumed to be C limited it can also be simultaneously N limited indirectly, in the sense that N additions to the soil may (i) increase C export to EMF by plants and (ii) allow EMF to increase fitness by enhanced allocation to reproductive production at the expense of N uptake capacity.

### Fungal competition for C and N

Whereas the above equations define physiology of EMF, to derive fungal strategy it is necessary to define functions for a single fungus (genotype, denoted by index  $i$ ) in relation to its competitors (denoted by index  $o$ ). Each fungus sharing a host plant with other fungi compete for C by individually increasing its N:C exchange rate with the plants ( $x_i$ , eq. 15) by adjusting its relative allocation to N uptake components ( $u_i$ ) until its reproductive production (the fitness proxy,  $G_{fi}$ , eq. 14) peaks.  $G_{fi}$  is a function of the C use efficiency of growth ( $y_f$ ) and three variable factors: the total plant C supply ( $C_f$ ), the fraction of this C that is captured by the fungus ( $F_i$ , eq. 15) among its  $n_f$  competitors, and the fungus' allocation to reproductive production ( $1-u_i$ )

$$G_{fi} = y_f C_f F_i (1-u_i) \quad \text{where} \quad (14)$$

$$F_i = \frac{x_i^z}{x_i^z + x_o^z (n_f - 1)} \quad \text{where} \quad x_i = \frac{N_{pi}}{C_{fi}} \quad \text{and} \quad N_{pi} = U_i - P_{fi} c_f \quad (15)$$

The partner discrimination parameter ( $z$ ) determines how strongly the plants are able to prioritize fungi with higher  $x$ , which influences the N uptake benefits of EMF for the plants and thereby at what soil N availability plants switch between mycorrhizal and non-mycorrhizal strategies (Fig. S3). Although not addressed here, this trading strategy may have evolved by mechanisms of competition-based screening and partner fidelity feedback (Archetti *et al.*, 2011).

In addition to plant derived C, N uptake from soil ( $U_i$ ) is also subject to inter-fungal competition (eq. 16), which we assume to depend on the number of fungal partners per tree ( $n_f$ ) similarly to competition for plant derived C.

$$U_i = U[B_{fui}] n_f F_{ui} \quad \text{where} \quad F_{ui} = \frac{B_{fui}}{B_{fui} + B_{fuo} (n_f - 1)} \quad (16)$$

In eq. 16,  $U$  (eq. 1) is a function of the total biomass of EMF uptake components ( $B_{fui}$ , eq. 18), and  $F_{ui}$  is the fraction N uptake captured by fungus  $i$ , which depends on its individual uptake component biomass ( $B_{fui}$ ). Using eq. 14,  $B_{fui}$  can be expressed as a function of  $G_{fi}$  and  $u_i$  (eq. 17).  $B_{fui}$  (eq. 18) links fungal mean strategy ( $u$ ) to plant C export ( $C_f$ ).

$$B_{fui} = G_{fi} \frac{1-u_i}{u_i} t_f \quad (17)$$

$$B_{fui}[u_i] = y_f C_f u \quad \text{where} \quad u = \frac{u_i}{u_i + (n_f + 1)u_o} \quad (18)$$

After sequentially inserting eqs. 17-18 in eq. 16, eq.16 in eq. 15, and eq. 15 in eq. 14, eq. 14 can be solved (numerically) for  $G_{fi}$ . The resulting  $G_{fi}$  (eq. 19) is a function of  $u_i$  and  $C_f$ , where  $C_f$  is determined by optimal plant behavior (eq. 22), thus linking fungal and plant behavior.

$$G_{fi} = G_{fi}[C_f, u_i] \quad (19)$$

### Plant competition for N

While eq. 10 defines the fitness (net growth) of a plant in terms of its own properties, in order to define its fitness in the presence of competitors for soil N, we must account for an individual's uptake capacity ( $U_{ci}$ ) in relation to the capacity of its competitors ( $U_{co}$ ). Using eq.2, the uptake capacities are calculated from the plants' C export ( $C_f$ ) and the resulting biomass of EMF uptake components ( $B_{fu}$ , eq. 18). The resulting N acquisition of an individual plant ( $N_{pi}$ , eq. 20) can be expressed as: The average uptake per area ( $N_p$ , eqs. 13, 21) times the number of competitors with overlapping uptake areas or common partner EMF ( $n_p$ ), times the fraction N uptake captured by individual  $i$  ( $F_{pi}$ , eq. 21). Based on the clear-cut spatial separation and evidence of strong fungal discrimination among plant partners (Lekberg *et al.*, 2010) we assumed that competition is too efficient to allow any plant to pay less C per N received than other plants and thus will equalize N:C exchange rate ( $x$ ) among plants. In effect, plants will compete via the amount of EMF supported by their C export and its uptake capacity. Thus,  $F_{pi}$  is equal to the uptake capacity contributed by individual ( $U_{ci}$ ) relative to the total uptake capacity.

$$N_{pi} = N_p n_p F_{pi} \quad \text{where} \quad F_{pi} = \frac{U_{ci}[C_{fi}, u]}{U_{ci}[C_{fi}, u] + U_{co}[C_{fo}, u](n_p - 1)} \quad (20)$$

$$N_p = N_p [U_c] \quad \text{where} \quad U_c = \frac{U_{ci}[C_{fi}, u] + (n_p - 1)U_{co}[C_{fo}, u]}{n_p} \quad (21)$$

Inserting  $N_{pi}$  (eq. 20) in eq.10, net growth of an individual plant among competitors ( $G_{pi}$ , eq. 22) can be expressed as a function of its C allocation to EMF ( $C_{fi}$ ), and mean fungal allocation strategy ( $u$ ) of the EMF.

$$G_{pi} = f[C_{fi}, C_{r0}, u] \quad (22)$$

In eq. 22, C allocation to non-mycorrhizal roots ( $C_{r0}$ ) is zero except when evaluating a non-mycorrhizal strategy (see below).

### Optimal (ESS) strategies of interacting plants and EMF

The behavior of plants and EMF are linked via their allocation strategies defined by  $C_{fi}$  and  $u_i$ , which determine fitness of plants (eq. 22) and EMF (eq. 19). The simultaneous optimization of plant and EMF behavior is done as follows: For each  $u_i$  the optimal  $C_f$  is calculated by maximizing eq. 22 with respect to  $C_{fi}$ , i.e. each plant individually increases its C export to the EMF ( $C_{fi}$ ) to increase its N uptake ( $N_{pi}$ ) until its net growth ( $G_{pi}$ ) peaks, i.e.  $\partial G_{pi} / \partial C_{fi} = 0$ . In the calculation of  $G_p$  all individuals are identical ( $C_{fi} = C_{fo}$ ) while the *change* in  $G_p$ , i.e.  $\partial G_{pi} / \partial C_{fi}$ , is evaluated with respect to only  $C_{fi}$  (and not  $C_{fo}$ ). This resulting optimal  $C_f$  is a function of  $u_i$  and is inserted in the fungal fitness function (eq. 19). Optimal  $u$  is then calculated by maximizing fungal fitness ( $G_{fi}$ ) with respect to  $u_i$ . In the evaluation of  $G_{fi}$  all individuals are identical ( $u_i = u_o$ ) while the *change* in  $G_{fi}$ , i.e.  $\partial G_{fi} / \partial C_{fi}$ , is evaluated with respect to only  $u_i$  (and not  $u_o$ ). The optimal (ESS) values of  $C_f$  and  $u$  determines the system and all dependent variables.

### Non-mycorrhizal plants

The physiological model for non-mycorrhizal plants differ from the mycorrhizal plant model only by the replacement of uptake via EMF with direct uptake by fine-roots ( $C_f = 0$ ,  $C_{r0} \neq 0$  in eq. 22) and the corresponding change in specific N uptake efficiency ( $e_u$ , Table S1). Competition for soil N among fine-roots of different plants is modeled in the same way as competition between different plants for the fungal derived N, but  $N_p$ ,  $B_{fu}$  and  $C_f$  are replaced by  $U$ ,  $B_{r0}$ , and  $C_r$ , respectively. We assumed that on average two plants compete for the same N ( $n_p = 2$ ) in a non-mycorrhizal plant population.

### Non-mycorrhizal versus mycorrhizal root strategies

A non-mycorrhizal strategy can invade a population of resident mycorrhizal plants if the increase in net growth per C invested is higher for a non-mycorrhizal root than for a mycorrhizal-root (including C allocation to EMF and supporting root structure), i.e.  $\partial G_{pi} / \partial C_{ri} > \partial G_{pi} / \partial C_{fi} > 0$ , where the last inequality follows from the definition of the ESS strategy of the resident mycorrhizal strategy. To calculate the N uptake due to the addition of a non-mycorrhizal root among existing mycorrhizal roots, the N uptake function (eq. 1) is extended by adding the uptake capacity of the non-mycorrhizal root to eqs. 20-21, in which  $U_c$  and  $F_{pi}$  are replaced by:

$$U_c = \frac{U_{ci}[C_{fi}, u] + U_{ci}[C_{ri}] + (n_p - 1)U_{co}[C_{fo}, u]}{n_p} \quad (23)$$

$$F_{pi} = \frac{U_{ci}[C_{fi}, u] + U_{ci}[C_{ri}]}{U_{ci}[C_{fi}, u] + U_{ci}[C_{ri}] + U_{co}[C_{fo}, u](n_p - 1)} \quad (24)$$

The invasion potential  $\partial G_{pi} / \partial C_{ri}$  is then evaluated as done in the calculation of optimal  $C_f$  of mycorrhizal plants based on eq. 22, where, after differentiation,  $C_{r0}$  is set to zero, which implies that the non-mycorrhizal root start growing at zero biomass.

The evaluation of the potential for a mycorrhizal strategy to invade a resident non-mycorrhizal strategy is analogous to the above, switching the positions of non-mycorrhizal and mycorrhizal N uptake in the equations.

### Model parameterization

Physiological parameters typical of boreal pine forest were used (Table S1), with unknown parameters adjusted to match the range of measured productivities of EMF and wood in boreal forest soil N gradients (Fig. 2). Measured productivity of EMF represented only the upper 10 cm soil layer, while the total productivity is at least twice as high (Ekblad *et al.*, 2013), and was therefore multiplied by 2.5 to match our modeled effective soil depth of 25 cm. Modeled wood production was assumed to contribute 75% of total net growth. Because the specific N uptake efficiency ( $e_u$ ) of EMF and fine-roots are unknown, a plausibly higher  $e_u$  for EMF than for plant roots was used (a factor of 300; Jennings, 1995; Smith & Read, 2008), which also enables the model to reproduce the commonly observed shift from mycorrhizal forest to non-mycorrhizal forest when going from low to high soil N availability (Taylor *et al.*, 2000; Högberg *et al.*, 2003; Kjoller *et al.*, 2012). Importantly, this parameterization does not build into the model any of the emergent results, such as for coexistence of strategies, symbiosis stabilization – soil N feedback, mycorrhizal colonization rates, effects of plant-fungal connectance, or effects of elevated CO<sub>2</sub>. The (unknown) parameter  $z$  was set to  $z=1$  and tested for a range of higher and lower values (Fig. S3). The ranges of partner numbers used (connectance) are within observed ranges (Table S1). The effect of elevated atmospheric [CO<sub>2</sub>] was modeled by increasing light saturated photosynthetic N use efficiency ( $a$ ) by 30%, which lies within the range of observed effects in FACE experiments (Franklin, 2007).



Figure S2

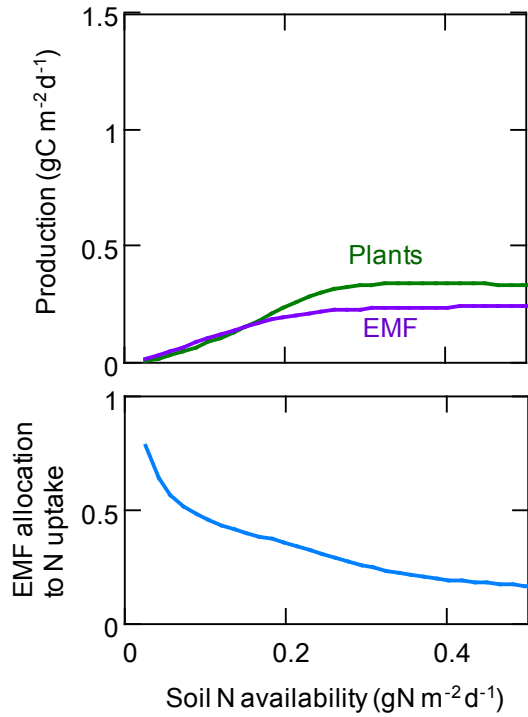


Figure S2. Modelled growth of plants (green lines) and ectomycorrhizal fungi (EMF, purple lines) and corresponding fungal strategy (fractional C allocation to N uptake ( $u$ ), blue line lower panel) as functions of soil N availability, for connectance  $n_p = 8$ , and  $n_f = 1$ , i.e. no inter-fungal competition for C. In contrast to fig. 3, here the plants do not have the option to switch to a non-mycorrhizal strategy.

Figure S3

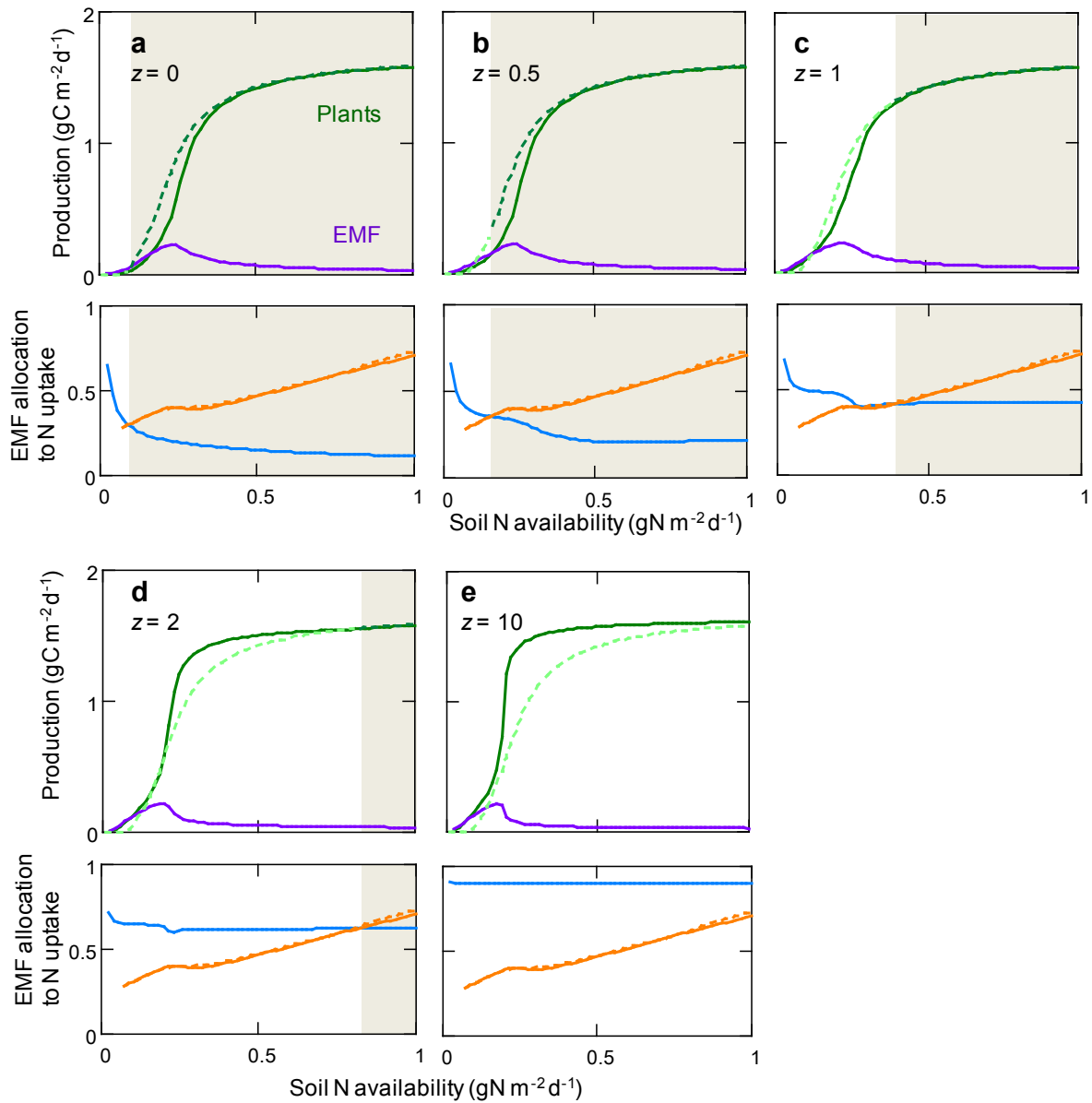


Figure S3. Modelled growth of plants (green lines) and ectomycorrhizal fungi (EMF, purple lines) and corresponding fungal strategy (fractional C allocation to N uptake ( $u$ ), lower figures) as functions of soil N availability as shown in Fig. 2, but for different values of the partner discrimination parameter ( $z$ , Table E1, eq. 15). As soil N availability increases, fungal strategy ( $u$ ) is first mainly determined by competition among EMF for plant C (blue line) and then (in shaded area) by competition with a non-mycorrhizal root strategy (solid orange line). The intensity of inter-fungal competition for plant C increases with  $z$ , resulting in higher fungal allocation to N uptake ( $u$ ) and higher plant N uptake, which delays the shift from a purely mycorrhizal to a mixed mycorrhizal and non-mycorrhizal plant strategy (shaded area) as soil N availability increases. Fungal-plant connectance were  $n_f = n_p = 8$  in all panels.

## References (that are not included in the main text)

- Archetti M, Scheuring I, Hoffman M, Frederickson ME, Pierce NE, Yu DW. 2011.** Economic game theory for mutualism and cooperation. *Ecology Letters* **14**(12): 1300-1312.
- Choudhury BJ. 2001.** Implementing a nitrogen-based model for autotrophic respiration using satellite and field observations. *Tropical Ecology* **2**: 141-174.
- Lilleskov EA, Hobbie EA, Horton TR. 2011.** Conservation of ectomycorrhizal fungi: Exploring the linkages between functional and taxonomic responses to anthropogenic N deposition. *Fungal Ecology* **4**(2): 174-183.
- Sinsabaugh RL, Manzoni S, Moorhead DL, Richter A. 2013.** Carbon use efficiency of microbial communities: stoichiometry, methodology and modelling. *Ecology Letters* **16**(7): 930-939.
- Wong SC, Cowan IR, Farquhar DG. 1979.** Stomatal conductance correlates with photosynthetic capacity. *Nature* **282**: 424-426.