

## Fourier Transform Infrared and Ultraviolet-Visible Spectroscopic Characterization of Biofield Treated Salicylic Acid and Sparfloxacin

Mahendra Kumar Trivedi<sup>1</sup>, Alice Branton<sup>1</sup>, Dahryn Trivedi<sup>1</sup>, Harish Shettigar<sup>1</sup>, Khemraj Bairwa<sup>2</sup> and Snehasis Jana<sup>2\*</sup>

<sup>1</sup>Trivedi Global Inc., 10624 S Eastern Avenue Suite A-969, Henderson, NV 89052, USA

<sup>2</sup>Trivedi Science Research Laboratory Pvt. Ltd., Hall-A, Chinar Mega Mall, Chinar Fortune City, Hoshangabad Rd., Bhopal, Madhya Pradesh, India

### Abstract

Salicylic acid is a naturally occurring derivative of benzoic acid, and widely used in organic synthesis and as a plant hormone. Sparfloxacin is fluorinated quinolone antibiotic having broad spectrum antimicrobial property. The present study was aimed to evaluate the impact of biofield treatment on spectral properties of salicylic acid and sparfloxacin using FT-IR and UV-Vis spectroscopic techniques. The study was carried out in two groups, one was set to control, and another was subjected to biofield treatment. FT-IR spectrum of treated salicylic acid showed the upstream shifting in wavenumber of C-H stretching from 2999 to 3004  $\text{cm}^{-1}$  and 2831 to 2837  $\text{cm}^{-1}$  and C=O asymmetric stretching vibration from 1670 to 1683  $\text{cm}^{-1}$  and 1652 to 1662  $\text{cm}^{-1}$ . The peak intensity in treated salicylic acid at 1558  $\text{cm}^{-1}$  (aromatic C=C stretching) and 1501  $\text{cm}^{-1}$  (C-C stretching) was increased as compared to control. FT-IR spectrum of treated sparfloxacin showed a downstream shifting in wavenumber of C-H stretching from 2961 to 2955  $\text{cm}^{-1}$  and 2848 to 2818  $\text{cm}^{-1}$ , and upstream shifting in wavenumber of C=O (pyridone) stretching from 1641 to 1648  $\text{cm}^{-1}$ . Besides, increased intensity of peaks in treated sparfloxacin was found at 1628  $\text{cm}^{-1}$  [C=C stretching (pyridone)] and 1507  $\text{cm}^{-1}$  (N-H bending) as compared to control. UV spectrum of biofield treated salicylic acid exhibited a shifting of wavelength ( $\lambda_{\text{max}}$ ) from 295.8 to 302.4 nm and 231.2 to 234.4 nm, with respect to control. Likewise, biofield treated sparfloxacin showed the shifting in UV wavelength ( $\lambda_{\text{max}}$ ) from 373.8 to 380.6 nm and 224.2 to 209.2 nm.

Over all, the results suggest that alteration in wavenumber of IR peaks in treated samples might be occurred due to biofield induced alteration in force constant and dipole moment of some bonds. The changes in UV wavelength ( $\lambda_{\text{max}}$ ) of treated sample also support the FT-IR results. Due to alteration in force constant and bond strength, the chemical stability of structure of treated drugs might also be increased, which could be beneficial for self-life of biofield treated drugs.

**Keywords:** Salicylic acid; Sparfloxacin; Biofield treatment; Fourier transform infrared spectroscopy; Ultraviolet-Visible spectroscopy

### Introduction

Salicylic acid is a mono-hydroxyl benzoic acid, and naturally occurring in the bark of willow tree (*Salix alba*) [1]. It is an important active metabolite of aspirin, which acts as a prodrug to salicylic acid. The salts and esters of salicylic acid are known as salicylates that are widely used as rubefacient and analgesic in several topical formulations. Salicylic acid alleviates peeling of intercellular cement and binds with scales in the stratum corneum, thereby loosening the keratin. This keratolytic effect also renders an antifungal effect as removal of the stratum corneum suppresses the fungal growth [1,2]. It exerts anti-inflammatory activity by suppressing the cyclooxygenase (COX) activity that caused to inhibition of pro-inflammatory mediators production. Therefore, it is widely used for the treatment of several skin diseases like acne, psoriasis, seborrhoeic dermatitis, calluses, keratosis pilaris, and warts due to its keratolytic, fungicidal, bacteriostatic, and photo-protective properties [3,4]. Salicylic acid is also a phytohormone and useful in growth and development of whole plant [5].

Sparfloxacin is a difluorinated quinolone antibiotic with broad spectrum antibacterial activity. Additionally, it also possesses a good *in vitro* activity against several unusual pathogens such as legionellae, chlamydia, rickettsiae, mycoplasmas, mycobacteria, etc. [6,7]. Sparfloxacin showed good topical absorption and had excellent penetration into upper and lower respiratory tissues; therefore it is extensively used in respiratory infections. It inhibits DNA replication and transcription in bacteria by inhibiting the DNA gyrase or topoisomerase IV enzyme [8]. The gastrointestinal discomfort and CNS effects are the most common adverse effects associated with sparfloxacin [9]. Chemical stability of pharmaceutical drugs or active ingredients is a matter of great concern as it affects the safety, efficacy, and shelf life of drugs or drug products [10]. Therefore, it is important

to find out an alternate approach, which could enhance the stability of drugs by changing the structural (bond strength, bond length, dipole moment etc.) properties of these compounds.

Recently, biofield treatment has been reported to alter the physical and structural properties of various living and non-living things [11,12]. Biofield is the electromagnetic field that permeates and surrounds the living organisms. It is the scientific term, used for biologically created electromagnetic energy, essential for regulation and communications within the organism [13]. As per Planck M, electrical current exists inside the human body in the form of vibratory energy particles like ions, protons, and electrons. These moving particles generates a magnetic field in the human body [14,15]. Mr. Trivedi has the ability to harness the energy from environment or universe and can transmit into any living or nonliving object around this Globe. The object(s) always receive the energy and responding into useful way, this process is known as biofield treatment. The National Center for Complementary and Alternative Medicine considered this biofield treatment (therapy) in subcategory of energy therapies [16].

Mr. Trivedi's biofield treatment has substantially changed the antimicrobial susceptibility, biochemical reactions pattern and

**\*Corresponding author:** Snehasis Jana, Trivedi Science Research Laboratory Pvt. Ltd., Hall-A, Chinar Mega Mall, Chinar Fortune City, Hoshangabad Rd., Bhopal-462026, Madhya Pradesh, India, Tel: +91-755-6660006; E-mail: [publication@trivedisrl.com](mailto:publication@trivedisrl.com)

Received July 14, 2015; Accepted August 18, 2015; Published August 21, 2015

**Citation:** Trivedi MK, Branton A, Trivedi D, Shettigar H, Bairwa K, et al. (2015) Fourier Transform Infrared and Ultraviolet-Visible Spectroscopic Characterization of Biofield Treated Salicylic Acid and Sparfloxacin. Nat Prod Chem Res 3: 186. doi:10.4172/2329-6836.1000186

**Copyright:** © 2015 Trivedi MK, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

biotype number of different human pathogens [13,17]. It also showed a significant impact in the field of agriculture and biotechnology, with respect to yield, nutrient value, and quality of products [18-20]. Mr. Trivedi's biofield treatment has also changed the various physicochemical and structural properties of metals and ceramics [12,21-23].

By conceiving the impact of biofield treatment on structural property of metals and ceramics, the present study was aimed to further explore the biofield treatment on two pharmaceutical drugs i.e., salicylic acid and sparfloxacin with respect to effects on their structural property. The effects were analyzed using Fourier transform infrared (FT-IR) and Ultraviolet-Visible (UV-Vis) spectroscopic techniques.

## Materials and Methods

### Study design

Salicylic acid was procured from Qualigens Fine Chemicals (Mumbai, India), and sparfloxacin was procured from Sigma-Aldrich (MA, USA). Each drug sample was divided into two parts i.e., control and treatment. The control groups were remained as untreated and treatment groups were handed over in sealed pack to Mr. Trivedi for biofield treatment under standard laboratory condition. Mr. Trivedi provided this treatment through his energy transmission process to the treatment groups without touching the samples. After that, both the control and treated samples of salicylic acid and sparfloxacin (Figure 1) were analyzed using FT-IR spectroscopy and UV-vis spectroscopy.

### FT-IR spectroscopic characterization

FT-IR spectra of salicylic acid and sparfloxacin (control and treated) were recorded on Shimadzu's Fourier transform infrared spectrometer (Japan) with the frequency range of 4000-500  $\text{cm}^{-1}$ . The FT-IR spectroscopic analysis was carried out to evaluate the impact of biofield treatment at atomic level like bond strength, and stability of the structure of both drugs [24].

### UV-Vis spectroscopic analysis

UV spectroscopic analysis of salicylic acid and sparfloxacin were acquired on a Shimadzu UV-2400 PC series spectrophotometer with 1 cm quartz cell and a slit width of 2.0 nm. The analysis was performed using wavelength range of 200-400 nm. This study was performed to evaluate the effect of biofield treatment on the structural property of tested drugs with respect to functional groups and their position.

## Results and Discussion

### FT-IR spectroscopic analysis

The FT-IR spectra of salicylic acid (control and treated) are shown in Figure 2. FT-IR spectrum of control sample (Figure 2a) showed characteristic vibrational peaks at wavenumber 3233  $\text{cm}^{-1}$  and 2999-2831  $\text{cm}^{-1}$  that were assigned to OH and C-H stretching, respectively. The C=O (COO<sup>-</sup>) asymmetric and symmetric stretching were assigned to IR peaks observed at 1652-1670  $\text{cm}^{-1}$  and 1386  $\text{cm}^{-1}$ , respectively. Further, IR peaks appeared at 1558-1610  $\text{cm}^{-1}$  were attributed to C=C (phenolic) multiple peaks. The C-C stretching peaks were observed at 1444-1503  $\text{cm}^{-1}$ . The O-H (phenolic) bending was assigned to IR peak appeared at 1324  $\text{cm}^{-1}$ . The COO<sup>-</sup> (C-O) stretching and C-OH (phenolic) stretching were assigned to IR peaks appeared at 1296  $\text{cm}^{-1}$  and 1156-1248  $\text{cm}^{-1}$ , respectively. The vibrational peaks appeared at 759-669  $\text{cm}^{-1}$  were attributed to =C-H bending. The observed FT-IR data of control salicylic acid was well supported by the literature data [25].

The FT-IR spectrum of biofield treated salicylic acid (Figure 2b) showed the IR absorption peaks at 3233 and 2837-3004  $\text{cm}^{-1}$  that were assigned to O-H and C-H stretching, respectively. Vibration peaks observed at 1662-1683  $\text{cm}^{-1}$  and 1558-1612  $\text{cm}^{-1}$  were attributed to C=O asymmetric stretching and C=C (phenolic) stretching peaks, respectively. In addition, C-C stretching peak was appeared at 1445-1501  $\text{cm}^{-1}$  and C=O (COO<sup>-</sup>) symmetric stretching peak was observed at 1387  $\text{cm}^{-1}$ . The O-H (Ph-OH) and =C-H bending peaks were appeared at 1324  $\text{cm}^{-1}$  and 760-669  $\text{cm}^{-1}$ , respectively. Vibrational peaks observed at 1296  $\text{cm}^{-1}$  and 1157-1249  $\text{cm}^{-1}$  were assigned to COO<sup>-</sup> (C-O) stretching and phenolic (C-OH) stretching, respectively.

Overall, the FT-IR data indicates a significant impact of biofield treatment at atomic level of salicylic acid as compared to control. The FT-IR data of treated salicylic acid exhibited the shifting in wavenumber of some bonds with respect to control sample. For instance, an upstream shifting in wavenumber of C-H stretching from 2999 (control) to 3004  $\text{cm}^{-1}$  (treated) and 2831 to 2837  $\text{cm}^{-1}$ ; and C=O asymmetric stretching vibrations from 1670 to 1683  $\text{cm}^{-1}$  and 1652 to 1662  $\text{cm}^{-1}$ . The frequency of vibrational peak ( $\nu$ ) depends on two factors i.e., force constant and reduced mass, which can be explained by following equation [26].

$$\nu = 1/2\pi c \sqrt{k/\mu}$$

here,  $c$  is speed of light,  $k$  is force constant and  $\mu$  is reduced mass.

If reduced mass is constant, then the frequency is directly proportional to the force constant; therefore, increase in frequency of any bond suggested a possible enhancement in force constant of respective bond [24]. Based on this it is speculated that the force constant and bond strength of C-H and C=O bond might increased after biofield treatment as compared to control. Additionally, the intensity of peaks at 1558  $\text{cm}^{-1}$  (aromatic C=C stretching) and 1501  $\text{cm}^{-1}$  (C-C stretching) was significantly increased in biofield treated sample, as compared to control. This might be due to alteration in ratio of change in dipole moment ( $\partial\mu$ ) to change in bond distance ( $\partial r$ ) [27].

The FT-IR spectrum of control sparfloxacin is shown in (Figure 3a), which showed the characteristic vibrational peaks at 3093-3462  $\text{cm}^{-1}$  that were collectively assigned to O-H and N-H stretching. Vibrational peaks appeared at 2848-2961  $\text{cm}^{-1}$  were assigned to C-H (CH<sub>3</sub>) stretching. Further, the vibrational peaks observed at 1715  $\text{cm}^{-1}$  and 1641  $\text{cm}^{-1}$  were attributed to C=O (COO<sup>-</sup>) asymmetric stretching and C=O (pyridone) stretching, respectively. The IR peaks appeared at 1586, and 1533  $\text{cm}^{-1}$  were assigned to C=C (benzene ring) stretching and N-H bending, respectively. The C-H asymmetrical bending and C=O (COO<sup>-</sup>) symmetric stretching peaks were observed at 1437 and 1334  $\text{cm}^{-1}$  respectively. The C-N (aryl) stretching and C-F stretching peaks were assigned to IR peaks observed at 1186-1293  $\text{cm}^{-1}$  and 1151  $\text{cm}^{-1}$ , respectively. Further, the IR peaks appeared at 1084, 1029, and 668  $\text{cm}^{-1}$  were assigned to C-O (COO<sup>-</sup>) stretching, C-N (alkyl) stretching, and =C-H bending, respectively. The FT-IR data of control sparfloxacin was well supported by the literature data [28,29].

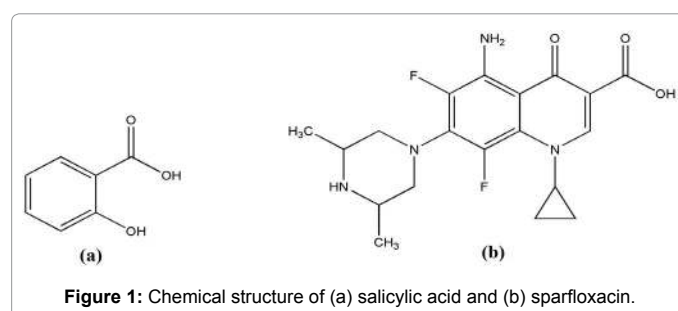


Figure 1: Chemical structure of (a) salicylic acid and (b) sparfloxacin.

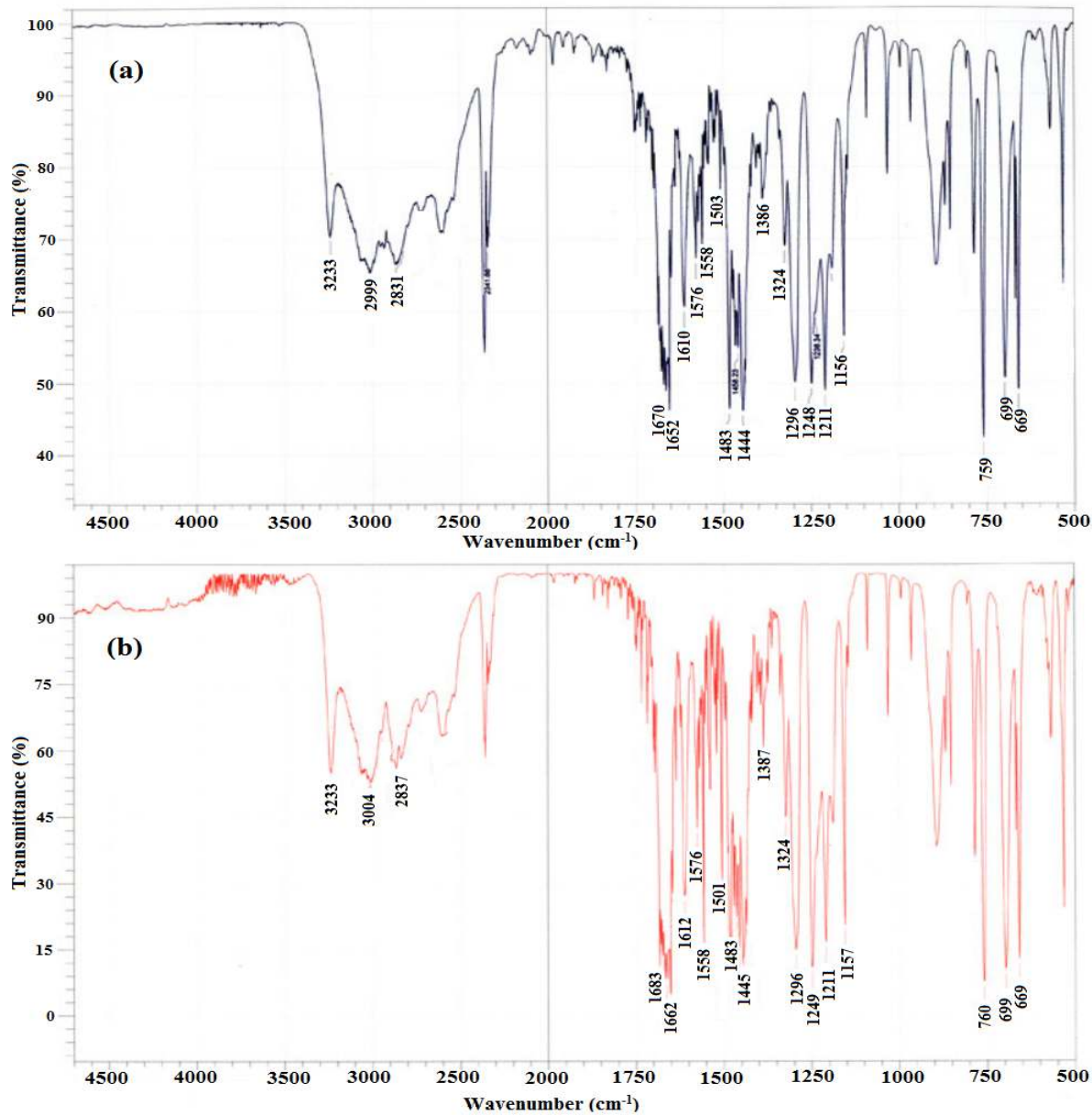


Figure 2: FT-IR spectra of salicylic acid (a) control and (b) treated.

The FT-IR spectrum (Figure 3b) of treated sparfloxacin showed the characteristic vibrational peaks at 3084-3462  $\text{cm}^{-1}$ , were attributed to overlapped peaks of O-H and N-H stretching. The vibrational peak appeared at 2818-2955  $\text{cm}^{-1}$  were assigned to C-H ( $\text{CH}_2$ ) stretching. Further, the vibrational peaks appeared at 1715  $\text{cm}^{-1}$  and 1648  $\text{cm}^{-1}$  were attributed to C=O ( $\text{COO}^-$ ) asymmetric stretching and C=O (pyridone) stretching, respectively. The IR peaks observed at 1628, 1586, and 1533-1507  $\text{cm}^{-1}$  were attributed to C=C (pyridone) stretching, C=C (benzene ring) stretching, and N-H bending, respectively. The IR peaks observed at 1437 and 1335  $\text{cm}^{-1}$  were attributed to asymmetrical bending of C-H group and symmetric stretching of C=O ( $\text{COO}^-$ ) group, respectively. Likewise, the C-N (aryl) stretching and C-F stretching were assigned to peaks observed at 1289-1186  $\text{cm}^{-1}$  and 1135  $\text{cm}^{-1}$ , respectively. Further, the C-O ( $\text{COO}^-$ ) stretching, C-N (alkyl) stretching, and =C-H bending were assigned to absorption peak observed at 1084, 1030, and 652  $\text{cm}^{-1}$ , respectively.

Overall, the FT-IR results of biofield treated sparfloxacin exhibited the shifting in wavenumber of some bonds with respect to control sample. For instance, a downstream shifting in frequency of vibrational peaks like C-H stretching from 2961 to 2955  $\text{cm}^{-1}$  and 2848 to 2818  $\text{cm}^{-1}$ ; C-F stretching from 1151 to 1135  $\text{cm}^{-1}$ ; and upstream shifting in wavenumber of C=O (pyridone) stretching from 1641 to 1648  $\text{cm}^{-1}$ . This slight change in wavenumber of peaks referred to corresponding changes in the force constant of that bond as compared to control. Increase in the force constant of stretching peaks, suggests the enhancement of bond strength and *vice versa*; likely, increasing in the force constant of bending peaks referred to increase in rigidity of bond in a molecule [25]. Additionally, C=C stretching (pyridone) peaks at 1628  $\text{cm}^{-1}$  and N-H bending 1507  $\text{cm}^{-1}$  were not observed in the control sample of sparfloxacin. However, these were significantly observed in treated sample. This might be observed due to very low intensity to be

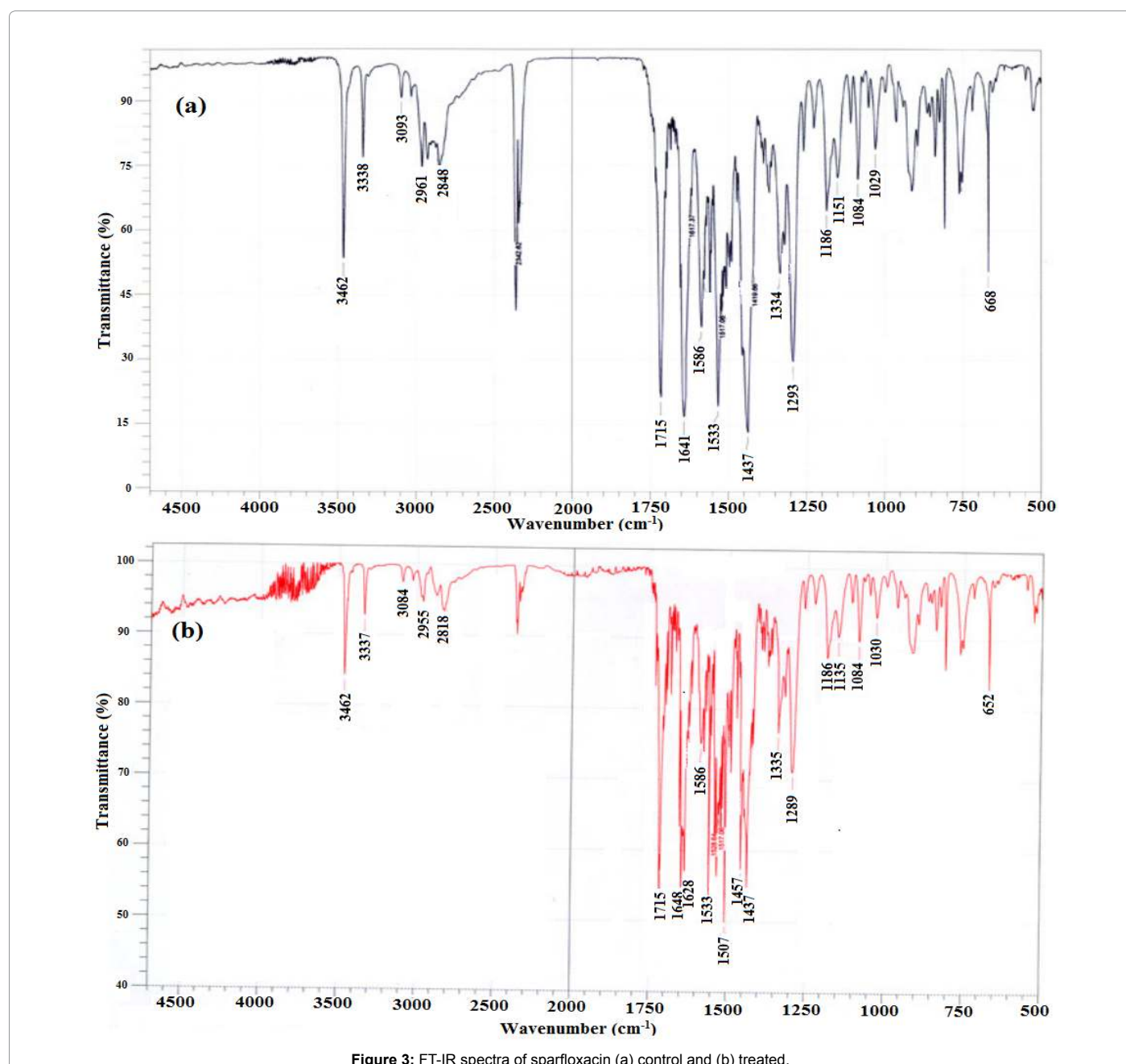


Figure 3: FT-IR spectra of sparfloxacin (a) control and (b) treated.

detected in control sample. It indicates a possible change in the  $\partial\mu/\partial r$  of these peaks [27], which might be occurred due to impact of biofield treatment at the atomic level (dipole moment) of treated sparfloxacin as compared to control.

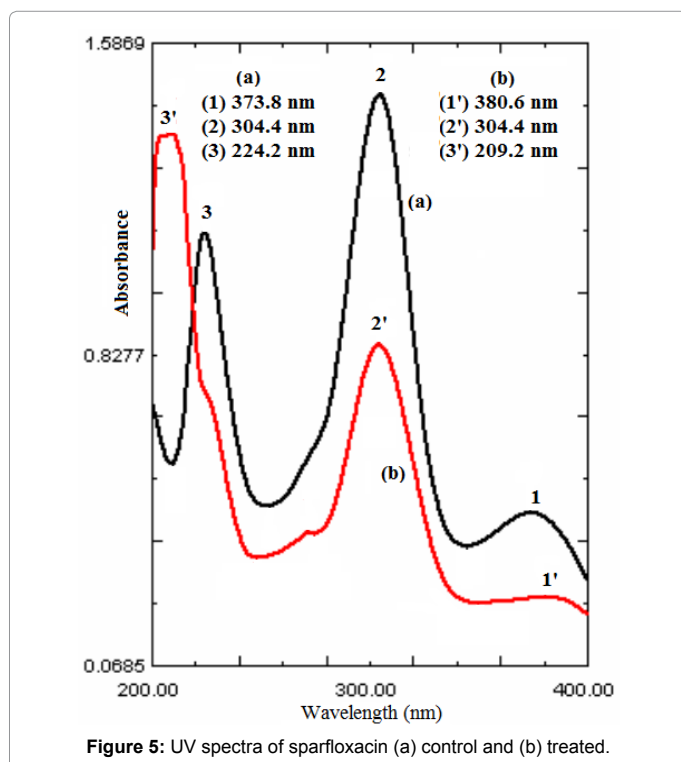
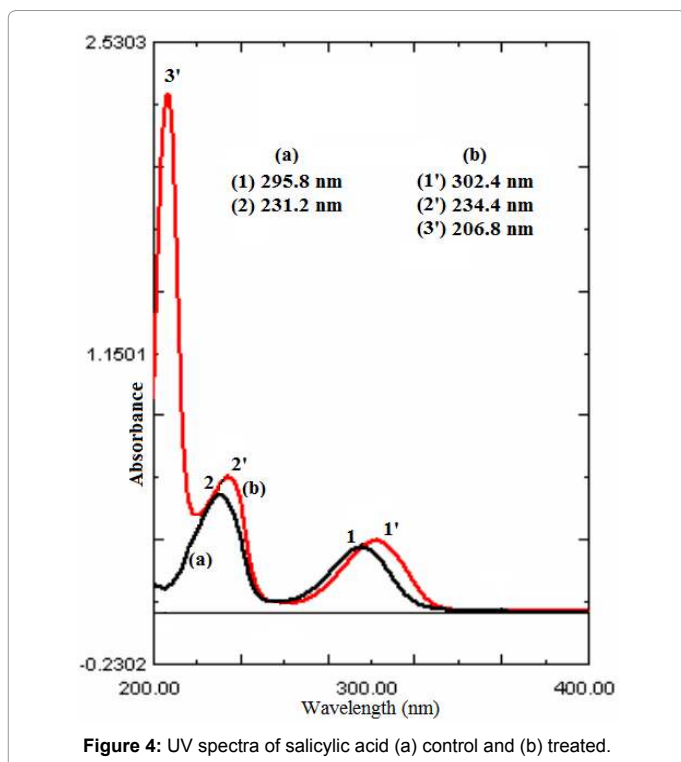
### UV-Vis spectroscopy

UV spectra of control and treated samples of salicylic acid are shown in Figure 4. It revealed that UV absorption peaks of treated sample were shifted to higher wavelength ( $\lambda_{\max}$ ) from 295.8  $\rightarrow$  302.4 nm and 231.2  $\rightarrow$  234.4 nm. The existing literature on principle of UV spectroscopy suggests that a compound can absorb UV light due to presence of either or both conjugated pi ( $\pi$ )-bonding systems ( $\pi - \pi^*$  transition) and nonbonding electron system ( $n - \pi^*$  transition) in the compound. The UV absorption phenomenon occurred when electrons travelled from low energy orbital (i.e.,  $\sigma$ ,  $n$  and  $\pi$ ) to high energy orbital

(i.e.,  $\sigma^*$  and  $\pi^*$ ). There is certain energy gap between  $\sigma - \sigma^*$ ,  $\sigma - \pi^*$ ,  $\pi - \pi^*$  and  $n - \pi^*$  orbitals. When this energy gap altered, the wavelength ( $\lambda_{\max}$ ) was also altered respectively [24]. Based on this, it is speculated that, due to influence of biofield treatment, the energy gap between  $\pi - \pi^*$  and  $n - \pi^*$  transition in salicylic acid might be altered, which causes shifting of wavelength ( $\lambda_{\max}$ ) in treated salicylic acid as compared to control.

UV spectra of sparfloxacin (control and treated) are shown in Figure 5. It showed the shifting of lambda max ( $\lambda_{\max}$ ) of UV absorption peaks in treated sample from 373.8  $\rightarrow$  380.6 nm and 224.2  $\rightarrow$  209.2 nm; and absorption peak at 304.4 were appeared at the same wavelength in both samples. This suggests a possible alteration in the energy gap between  $\pi - \pi^*$  and  $n - \pi^*$  transition in treated sparfloxacin as compared to control.





## Conclusion

The FT-IR data of both drugs showed an alteration in the wavenumber of some functional groups like C-H and C=O (COO<sup>-</sup>) in treated salicylic acid, and C-H, C=O (pyridine), and C-F in treated sparfloxacin as compared to their respective control. The increased intensity of peaks at 1558 and 1501 cm<sup>-1</sup> was found in treated salicylic

acid. While the increased intensity of peaks at 1628 and 1507 cm<sup>-1</sup> was found in treated sparfloxacin, as compared to respective control. The UV spectra of both drugs showed a biofield induce shifting of wavelength ( $\lambda_{max}$ ) as compared to control. This might be occurred due to some possible alteration in structural properties like force constant, bond strength, dipole moments etc. of treated drugs through the biofield treatment. Due to possible alterations in structural properties of treated drugs, the chemical stability might also be altered, which could be beneficial for the shelf life of pharmaceutical drugs.

## Acknowledgement

The authors would like to acknowledge the whole team of MGV Pharmacy College, Nashik for providing the instrumental facility.

## References

1. Miner J, Hoffhines A (2007) The discovery of aspirin's antithrombotic effects. *Tex Heart Inst J* 34: 179-186.
2. Davies M, Marks R (1976) Studies on the effect of salicylic acid on normal skin. *Br J Dermatol* 95: 187-192.
3. Vane JR (1971) Inhibition of prostaglandin synthesis as a mechanism of action for aspirin-like drugs. *Nat New Biol* 231: 232-235.
4. Madan RK, Levitt J (2014) A review of toxicity from topical salicylic acid preparations. *J Am Acad Dermatol* 70: 788-792.
5. Raskin I (1992) Salicylate, A new plant hormone. *Plant Physiol* 99: 799-803.
6. Johnson JH, Cooper MA, Andrews JM, Wise R (1992) Pharmacokinetics and inflammatory fluid penetration of sparfloxacin. *Antimicrob Agents Chemother* 36: 2444-2446.
7. Cooper MA, Andrews JM, Ashby JP, Matthews RS, Wise R (1990) In-vitro activity of sparfloxacin, a new quinolone antimicrobial agent. *J Antimicrob Chemother* 26: 667-676.
8. Hooper DC (2000) Mechanisms of action and resistance of older and newer fluoroquinolones. *Clin Infect Dis* 31: S24-S28.
9. Stein GE, Havlicek DH (1997) Sparfloxacin: Potential clinical and economic impact in the treatment of respiratory infections. *Pharmacotherapy* 17: 1139-1147.
10. Blessy M, Patel RD, Prajapati PN, Agrawal YK (2014) Development of forced degradation and stability indicating studies of drugs-A review. *J Pharm Anal* 4: 159-165.
11. Trivedi MK, Patil S (2008) Impact of an external energy on *Yersinia enterocolitica* [ATCC-23715] in relation to antibiotic susceptibility and biochemical reactions: an experimental study. *Internet J Alternat Med* 6-13.
12. Trivedi MK, Patil S, Tallapragada RM (2013) Effect of biofield treatment on the physical and thermal characteristics of vanadium pentoxide powders. *J Material Sci Eng S11*, 001.
13. Garland SN, Valentine D, Desai K, Langer C, Evans T et al. (2013) Complementary and alternative medicine use and benefit finding among cancer patients. *J Altern Complement Med* 19: 876-881.
14. Planck M (1903) *Treatise on Thermodynamics*, (3rd edn) translated by Alexander OGG, Longmans, Green, London (UK).
15. Einstein A (1905) Does the inertia of a body depend upon its energy-content. *Ann Phys* 18: 639-641.
16. Hok J, Tishelman C, Ploner A, Forss A, Falkenberg T (2008) Mapping patterns of complementary and alternative medicine use in cancer: an explorative cross-sectional study of individuals with reported positive "exceptional" experiences. *BMC Complement Altern Med* 8: 48.
17. Trivedi MK, Bhardwaj Y, Patil S, Shettigar H, Bulbule, A (2009) Impact of an external energy on *Enterococcus faecalis* [ATCC-51299] in relation to antibiotic susceptibility and biochemical reactions—an experimental study. *J Accord Integr Med* 5: 119-130.
18. Lenssen AW (2013) Biofield and fungicide seed treatment influences on soybean productivity, seed quality and weed community. *Agricultural Journal* 8: 138-143.
19. Sances F, Flora E, Patil S, Spence A, Shinde V (2013) Impact of biofield treatment on ginseng and organic blueberry yield. *Agrivita J Agric Sci* 35.

20. Patil SA, Nayak GB, Barve SS, Tembe RP, Khan RR (2012) Impact of biofield treatment on growth and anatomical characteristics of *Pogostemon cablin* (Benth.). *Biotechnology* 11: 154-162.
21. Trivedi MK, Patil S, Tallapragada RM (2013) Effect of bio field treatment on the physical and thermal characteristics of silicon, tin and lead powders. *J Material Sci Eng* 2: 125.
22. Trivedi MK, Patil S, Tallapragada RMR (2015) Effect of biofield treatment on the physical and thermal characteristics of aluminium powders. *Ind Eng Manage* 4: 151.
23. Trivedi MK, Nayak G, Patil S, Tallapragada RM, Latiyal O (2015) Studies of the atomic and crystalline characteristics of ceramic oxide nano powders after biofield treatment. *Ind Eng Manage* 4: 1000161.
24. Pavia DL, Lampman GM, Kriz GS (2001). *Introduction to spectroscopy*. (3rd edn), Thomson learning, Singapore.
25. Guan XH, Chen GH, Shang C (2007) ATR-FTIR and XPS study on the structure of complexes formed upon the adsorption of simple organic acids on aluminum hydroxide. *J Environ Sci (China)* 19: 438-443.
26. Stuart BH (2004) *Infrared Spectroscopy: Fundamentals and applications (analytical techniques in the sciences (AnTs))*. John Wiley & Sons Ltd; Chichester, UK.
27. Smith BC (1999) *Infrared Spectral Interpretation: A systematic approach*. CRC Press 1-288.
28. Gupta H, Aqil M, Khar RK, Ali A, Bhatnagar A, et al. (2010) Sparfloxacin-loaded PLGA nanoparticles for sustained ocular drug delivery. *Nanomedicine* 6: 324-333.
29. Salgado HRN, Moreno, PRH, Braga AL, Schapoval EES (2005) Photodegradation of sparfloxacin and isolation of its degradation products by preparative HPLC. *J Basic Appl Pharm Sci* 26: 47-54.