

FOOD CHEMICAL CONTAMINANTS

Fumonisin Levels in Uruguayan Corn Products¹

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A survey was conducted to evaluate fumonisins FB₁ and FB₂ in Uruguayan corn products. Sixty-four samples of different local brands were purchased from retail stores during a 15-month period and analyzed for FB₁ and FB₂ by methanol-water extraction, cleanup with a 1 mL strong-anion-exchange solid-phase extraction column, and liquid chromatography with *o*-phthaldialdehyde-2-mercaptoethanol derivatization and fluorescence detection. Contamination levels for FB₁ varied from 50 ng/g (detection limit) to 6342 ng/g. Values were highest in feed samples (up to 6342 ng/g), unprocessed corn kernel (up to 3688 ng/g), and milled products, which included polenta (up to 427 ng/g). They were lowest in processed corn kernel (up to 155 ng/g) and snacks (up to 314 ng/g). FB₂ was determined in one-fourth of the total samples and detected at trace levels in only one feed sample. The data demonstrated the natural occurrence of fumonisins in corn products in Uruguay. Feed and polenta that contain fumonisins could be of concern because they are consumed in large amounts and are often the main nutrient source in Uruguay.

Fumonisin is a family of mycotoxins produced by *Fusarium moniliforme*, *F. proliferatum*, and other related species (1, 2) that colonize corn worldwide. Fumonisin has been reported in corn-based human food (3, 4). The toxins produced can accumulate to levels known to be toxic to animals and potentially harmful to humans. The fumonisins cause leukoencephalomalacia (ELEM) in horses (5) and porcine pulmonary edema (PPE) in swine (6). It has been proposed that one of the mechanisms of toxicity of fumonisins is inhibition of the enzyme sphinganine *N*-acyltransferase, which results in a decrease in sphingosine and accumulation of free sphinganine, an

intermediate in the biosynthetic pathway for complex sphingolipids (7).

Esophageal cancer has been linked to one of these toxins, fumonisin B₁ (FB₁; 8, 9). In humans, ingestion of corn infected with *F. moniliforme* has been implicated in the high incidence of esophageal cancer in northeastern Italy (10), the Transkei region in South Africa, and the Linxian region of China (11). Uruguay has a high rate of esophageal cancer that has been associated with drinking "mate," a local herbal infusion sipped very hot through a metal straw. However, this association has never been demonstrated. Because the natural occurrence of fumonisins in Uruguayan corn foods had not been studied, a survey was conducted to evaluate FB₁ and FB₂ in priority corn products.

Experimental

Sample Collection

Sixty-four different local brands of corn products were purchased from retail stores during a 15-month period (1/1995 to 4/1996). Samples included 22 unprocessed corn kernel, 8 polentas, 2 popcorn, 5 snacks, 4 corn starch, 3 breakfast cereals, 4 canned corn, 2 frozen corn, 1 mazamorra (grits), and 13 corn-based feeds. These products were grouped into 5 categories: milled products (I), snacks and breakfast cereals (II), corn-based feeds (III), processed corn kernel (IV), and unprocessed corn kernel (V). The samples were a random selection of the most popular national brands taken from store shelves. A subsample of ca 1 kg of each was ground to uniform consistency and mixed well before analysis.

Reagents and Apparatus

(a) *Solvents and reagents.*—Liquid chromatographic (LC) grade acetonitrile and methanol; American Chemical Society (ACS) grade acetic acid, sodium bicarbonate, and sodium tetraborate; *o*-phthaldialdehyde (OPA), Sigma Chemical Co., St. Louis, MO; LC quality water.

(b) *Solid-phase extraction (SPE) tubes.*—Strong-anion-exchange (SAX) columns; 1 mL Supelclean LC-SAX, Supelco (Belfonte, PA).

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Table 1. Occurrence of fumonisins B₁ and B₂ in Uruguayan corn-based products

Category	Product	FB ₁ , ng/g	FB ₂ , ng/g	Range, positives/ category FB ₁ , ng/g	Average FB ₁ , ng/g	Incidence/category FB ₁ pos./total, %			
I	Polenta	277	— ^a	100–427	105	3/12 (25)			
	Polenta	ND ^b	—						
	Polenta	100	—						
	Polenta	ND	—						
	Polenta	ND	ND						
	Polenta	ND	ND						
	Polenta	ND	—						
	Polenta	427	—						
	Corn starch	ND	ND						
	Corn starch	ND	ND						
	Corn starch	ND	ND						
	Corn starch	ND	ND						
	II	Pop corn	199				—	152–314	118
Snacks		314	ND						
Corn flakes		218	—						
Snacks		152	ND						
Corn flakes		ND	ND	152–314	118	4/10 (40)			
Pop corn		ND	ND						
Snacks		ND	ND						
Snacks		ND	—						
Snacks		ND	—						
Corn flakes		ND	—						
III	Feed	3733	—				256–6342	2573	13/13 (100)
	Feed	2800	—						
	Feed	2450	—						
	Feed	2700	—						
	Feed	2390	—						
	Feed	3480	Trace						
	Feed	1848	—						
	Feed	1200	—						
	Feed	5021	—						
	Feed	690	—						
	Feed	540	—						
	Feed	6342	—						
	Feed	256	—						
IV	Canned cream corn	ND	—	155	65	1/7 (14)			
	Canned corn	ND	ND						
	Frozen corn	155	—						
	Mazamorra	ND	—						
	Canned corn	ND	ND						
	Canned corn	ND	—						
	Frozen corn	ND	—						
V	Corn	3153	—	165–3688	963	11/22 (50)			
	Corn	ND	—						
	Corn	ND	—						
	Corn	1672	—						
	Corn	513	—						
	Corn	676	—						
	Corn	1877	—						
	Corn	ND	—						
	Corn	ND	—						
	Corn	3688	—						
	Corn	ND	—						
	Corn	5787	—						
	Corn	212	ND						

Table 1. (continued)

Category	Product	FB ₁ , ng/g	FB ₂ , ng/g	Range, positives/ category FB ₁ , ng/g	Average FB ₁ , ng/g	Incidence/category FB ₁ pos./total, %
V (continued)	Corn	1313	—			
	Corn	ND	—			
	Corn	ND	—			
	Corn	ND	—			
	Corn	1582	—			
	Corn	ND	ND			
	Corn	ND	—			
	Corn	ND	—			
	Corn	165	—			

^a Not determined.

^b ND (not detected); equated to detection limit (50 ng/g).

(c) *o*-Phthaldialdehyde (OPA) solution.—Dissolve 40 mg in 1 mL methanol in amber vial; add 5 mL 0.1M sodium tetraborate and 50 μ L 2-mercaptoethanol (MCE); prepare daily.

(d) *Fumonisin standards*.—Plant Research Centre, Agriculture and Agri-Food, Ottawa, ON, Canada.

(e) *FB₁ and FB₂ standard solutions*.—Prepare stock solution of 100 μ g/mL of each fumonisin in acetonitrile–water (1 + 1, v/v). Dilute 100 μ L of each stock solution to 1 mL in acetonitrile–water (1 + 1, v/v) to obtain working solution of 10 μ g/mL. Store all standard solutions in freezer.

(f) *Mobile phase*.—Methanol–0.1M NaH₂PO₄ (77 + 23, v/v; pH 3.3) and H₂O.

(g) *LC system*.—Hewlett Packard Model 1050 isocratic system with 20 μ L sample loop, 100 \times 4.6 mm C₁₈ ODS Hypersil column with 5 μ m particle size and precolumn; Hewlett Packard fluorescence detector Model 1046A with excitation wavelength of 232 nm and emission wavelength of 425 nm; and Hewlett Packard Model 1050 integrator.

Extraction and Cleanup

A 25 g ground subsample was shaken for 30 min with 250 mL methanol–water (3 + 1, v/v) and then filtered through Whatman No. 50 filter paper. The pH of the extract was adjusted if necessary to pH 5.8–6.5 with 1M NaOH (12). A 1 mL portion containing 0.1 g sample was applied to a 1 mL SAX SPE column previously conditioned with 2 mL methanol followed by 1 mL water (13). The column was washed successively with 500 μ L water and 500 μ L methanol. All washes were discarded. Fumonisin B₁ and B₂ were eluted with 1 mL methanol–acetic acid (99 + 1, v/v). Washing and elution were accomplished by gravity flow. The eluate was evaporated to dryness under a stream of nitrogen in a water bath (60°C). The residue was redissolved in 200 μ L methanol, and portions of this solution were used for derivatization.

Derivatization and LC Analysis

Residues were derivatized immediately prior to injection by addition of 250 μ L OPA reagent to 25 μ L sample solution. A 20 μ L portion of the derivatized samples was analyzed by a reversed-phase, isocratic system with fluorescence detection.

Quantitation was by peak height. The flow rate was 1 mL/min. FB₁ and FB₂ standards (10 μ each) were derivatized in the same way. Method recoveries from 6 samples spiked with 100 or 1000 ng FB₁/g ranged from 60 to 110%. Detection limit was 50 ng/g. Results below the level of detection were taken as 50 ng/g for calculations of average.

Results and Discussion

Naturally occurring levels of FB₁ in Uruguayan corn and corn-based products varied from 50 ng/g (detection limit) to 6342 ng/g (Table 1). Values were highest in feed samples (average, 2573; range, 256–6342 ng/g); unprocessed corn kernel (average, 963; range, ND–3688 ng/g); and milled products, including polenta (average, 105; range, ND–427 ng/g). They were lowest in snacks and breakfast cereals (average, 118; range, ND–314 ng/g) and processed corn kernel (average, 65; range, ND–155 ng/g).

FB₁ incidence was 100% in feed samples and a minimum of 14% in processed corn kernel. The distribution of FB₁ levels for all 4 categories as percent of total samples is shown in Figure 1. Feeds also gave the highest frequency of positive samples. The feed sample with the highest FB₁ value (6342 ng/g) originated from an incident involving a horse with fatal ELEM symptoms.

FB₂ was determined at random in one-fourth of the total samples and detected at trace levels in only one feed sample (Table 1).

The data presented are the first report of the natural occurrence of fumonisins in corn products in Uruguay. Feed and polenta that contain fumonisins could be of concern because they are consumed in high amounts and are often the main nutrient source for animals and humans, respectively.

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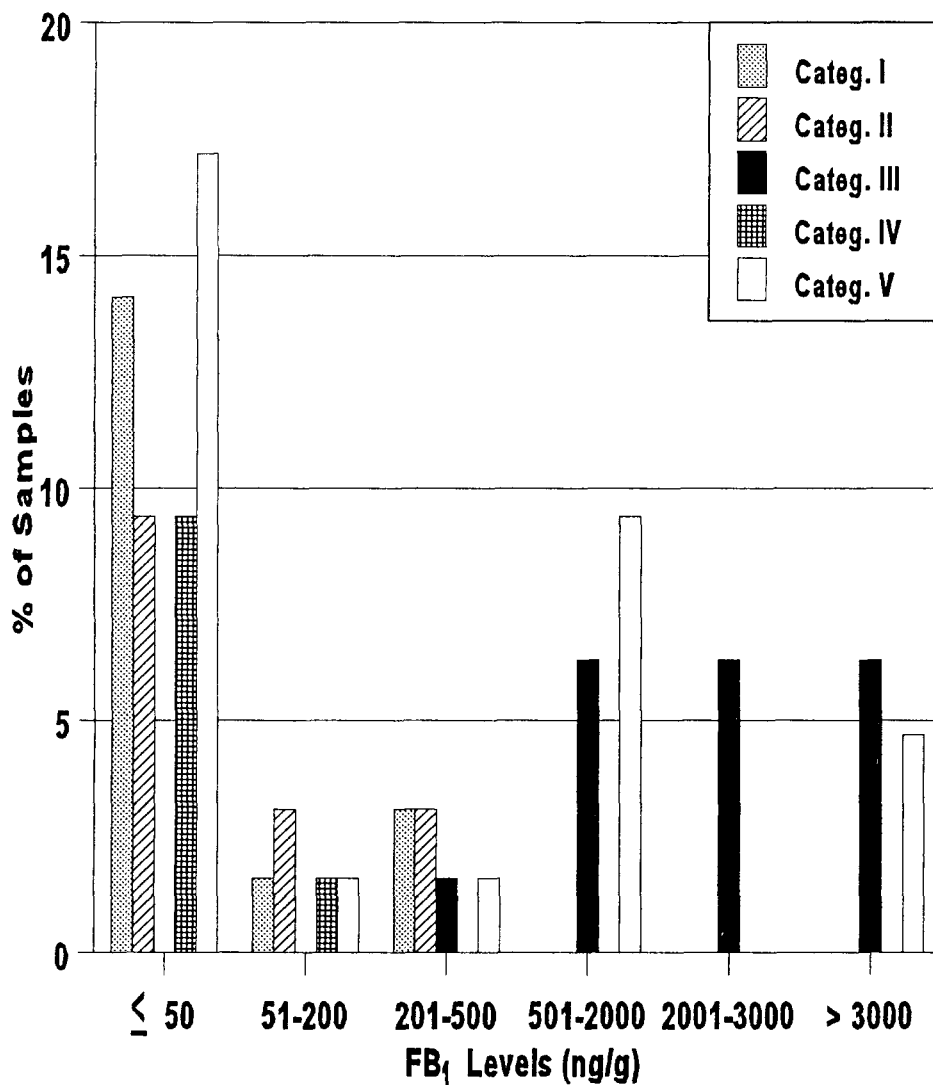


Figure 1. Frequency distribution of FB₁ levels.

References

- (1) Nelson, P.E., Desjardins, A.E., & Plattner, R.D. (1993) *Annu. Rev. Phytopathol.* **31**, 233-252
- (2) Norred, W.P. (1993) *J. Toxicol. Environ. Health* **38**, 309-328
- (3) Bullermann, L.B., & Tsai, W.J. (1994) *J. Food Prot.* **57**, 541-546
- (4) Shepard, G.S., Thiel, P.G., Stockenstrom, S., & Sydenham, E.W. (1996) *J. AOAC Int.* **79**, 671-687
- (5) Ross, P.F., Nelson, P.E., Richard, J.L., Osweiler, G.D., Rice, L.G., Plattner, R.D., & Wilson, T.M. (1990) *Appl. Environ. Microbiol.* **56**, 3225-3226
- (6) Harrison, L.R., Colvin, B.M., Green, J.J., Newmann, L.E., & Cole, J.R. (1990) *J. Vet. Diagn. Invest.* **2**, 217-221
- (7) Riley, R.T., Wang, E., & Merrill, A.H. (1994) *J. AOAC Int.* **77**, 533-540
- (8) Scott, P.M. (1993) *Int. J. Food. Microbiol.* **18**, 257-270
- (9) Sydenham, E.W., Gelderblom, W.C., Thiel, P.G., & Marasas, W.F. (1990) *J. Agric. Food Chem.* **38**, 285-290
- (10) Franceschi, S., Bidoli, E., Baron, A.E., & La Vecchia, C. (1990) *J. Nat. Cancer Inst.* **82**, 1407-1411
- (11) Sydenham, E.W., Shepard, G.S., Thiel, P.G., Marasas, W.F.O., & Stockenstrom, S. (1991) *J. Agric. Food Chem.* **25**, 767-771
- (12) *Official Methods of Analysis* (1996) AOAC INTERNATIONAL, Gaithersburg, MD, sec. **49.10.03**
- (13) Scott, P.M., & Lawrence, G.A. (1996) *Food Addit. Contam.* **13**, 823-832