



Fungal Planet description sheets: 128–153

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Key words

ITS DNA barcodes
LSU
novel fungal species
systematics

Abstract Novel species of microfungi described in the present study include the following from Australia: *Catenulostroma corymbiae* from *Corymbia*, *Devriesia stirlingiae* from *Stirlingia*, *Penidiella carpentariae* from *Carpentaria*, *Phaeococcomyces eucalypti* from *Eucalyptus*, *Phialophora livistonae* from *Livistona*, *Phyllosticta aristolochiicola* from *Aristolochia*, *Clitopilus austroprunulus* on sclerophyll forest litter of *Eucalyptus regnans* and *Toxicocladosporium posoqueriae* from *Posoqueria*. Several species are also described from South Africa, namely: *Ceramothyrium podocarp* from *Podocarpus*, *Cercospora chrysanthemoides* from *Chrysanthemoides*, *Devriesia shakazulii* from *Aloe*, *Penidiella drakensbergensis* from *Protea*, *Strelitziana cliviae* from *Clivia* and *Zasmidium syzygii* from *Syzygium*. Other species include *Bipolaris microstegii* from *Microstegium* and *Synchaetomella acerina* from *Acer* (USA), *Brunneiapiospora austropalmicola* from *Rhopalostylis* (New Zealand), *Calonectria pentaseptata* from *Eucalyptus* and *Macadamia* (Vietnam), *Ceramothyrium melastoma* from *Melastoma* (Indonesia), *Collembolispora aristata* from stream foam (Czech Republic), *Devriesia imbrexigena* from glazed decorative tiles (Portugal), *Microcyclospora rhoicola* from *Rhus* (Canada), *Seiridium phyllicae* from *Phyllica* (Tristan de Cunha, Inaccessible Island), *Passalora lobeliaefistulosis* from *Lobelia* (Brazil) and *Zymoseptoria verkleyi* from *Poa* (The Netherlands). *Valsalinicola* represents a new ascomycete genus from *Alnus* (Austria) and *Parapenidiella* a new hyphomycete genus from *Eucalyptus* (Australia). Morphological and culture characteristics along with ITS DNA barcodes are also provided.

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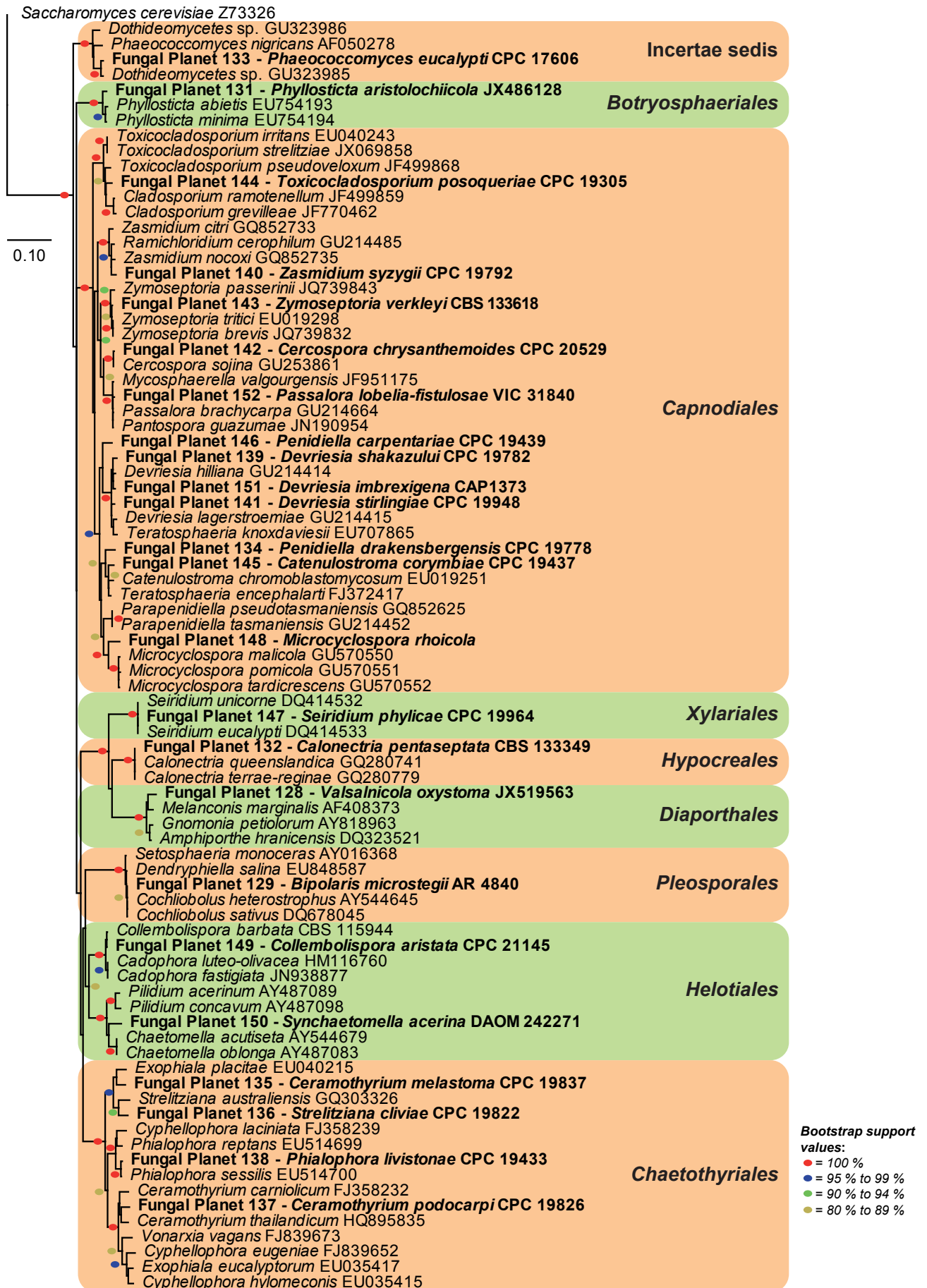
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Neighbour-joining tree obtained using a distance analysis with a general time reversible (GTR) substitution model on the partial 28S nrRNA gene alignment (817 nucleotides including alignment gaps) as implemented in PAUP v. 4.0b10 (Swofford 2003). Novel species are indicated in a **bold** font and the orders are indicated on the right-hand side of the figure. The scale bar indicates the number of substitutions per site and the bootstrap support values (based on 1 000 replicates) are shown by colour-coded dots for values > 79 % (see legend on figure). The tree was rooted to a sequence of *Saccharomyces cerevisiae* (GenBank Z73326.)

Calonectria pentaseptata



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Calonectria pentaseptata L. Lombard, M.J. Wingf., P.Q. Thu & Crous, *sp. nov.*

Etymology. Name refers to the 5-septate macroconidia produced by this fungus.

Sexual morph unknown. *Conidiophores* consisting of a stipe bearing a suit of penicillate fertile branches, a stipe extension, and terminal vesicle; stipe septate, hyaline, smooth 47–133 × 6–10 µm; stipe extension septate, straight to flexuous, 168–350 µm long, 3–6 µm wide at the apical septum, terminating in a narrowly clavate vesicle, 2–6 µm diam. *Conidiogenous apparatus* 70–99 µm long, 23–90 µm wide; primary branches 0–1-septate, 19–31 × 4–7 µm; secondary branches aseptate, 16–34 × 4–7 µm; tertiary branches aseptate, 14–22 × 4–6 µm, each terminal branch producing 1–3 phialides; phialides cylindrical to allantoid, obpyriform when carried singly, hyaline, aseptate, 15–24 × 4–6 µm; apex with minute periclinal thickening and inconspicuous collarete. *Macroconidia* cylindrical, rounded at both ends, straight, (75–)87–109(–115) × (5–)6–8(–10) µm (av. = 98 × 7 µm), 5(–8)-septate, lacking a visible abscission scar, held in parallel cylindrical clusters by colourless slime. *Megaconidia* and *microconidia* not seen.

Culture characteristics — (in the dark, 24 °C after 1 wk): Colonies fast growing, with optimum growth at 24 °C on MEA; surface sienna to dark brick, reverse sepia-brown; abundant aerial mycelium and sporulation; chlamydospores extensive throughout the medium, forming microsclerotia.

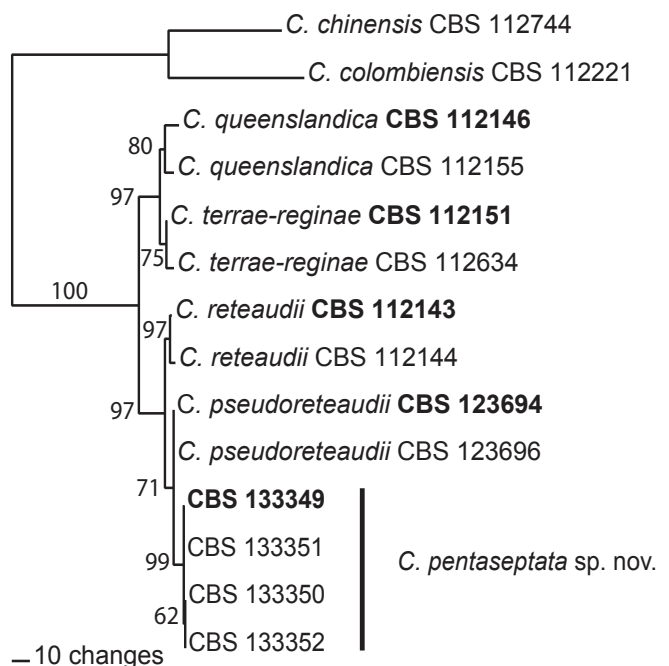
Typus. VIETNAM, Bavi, Hanoi, *Eucalyptus* hybrid, Sept. 2011, P.Q. Thu, holotype CBS H-21062, culture ex-type CBS 133349, β-tubulin (TUB) sequence GenBank JX855942, Histone H3 (HIS3) sequence GenBank JX855946, ITS sequence GenBank JX855950, LSU sequence GenBank JX855954 and translations elongation factor 1-alpha (TEF1-α) sequence GenBank JX855958, MycoBank MB801468.

Colour illustrations. *Eucalyptus* plantation in Vietnam; conidiophore; clavate vesicles; conidiogenous apparatus; conidia. Scale bars = 10 µm.

Other specimens examined. VIETNAM, Bavi, Hanoi, *Macadamia* sp., Sept. 2011, P.Q. Thu, CBS 133351, TUB sequence GenBank JX855944, HIS3 sequence GenBank JX855948, ITS sequence GenBank JX855952, LSU sequence GenBank JX855956 and TEF1-α sequence GenBank JX855960; *ibid.*, *E. urophylla*, Sept. 2011, P.Q. Thu, CBS 133350, TUB sequence GenBank JX855943, HIS3 sequence GenBank JX855947, ITS sequence GenBank JX855951 and TEF1-α sequence GenBank JX855959; *ibid.*, *Eucalyptus* hybrid, Sept. 2011, P.Q. Thu, CBS 133352, TUB sequence GenBank JX855945, HIS3 sequence GenBank JX855949, ITS sequence GenBank JX855953 and TEF1-α sequence GenBank JX855961.

Notes — *Calonectria pentaseptata* resides in the *C. reteaudii* species complex (Kang et al. 2001, Lombard et al. 2010a, b, c) based on morphological characteristics supported by phylogenetic inference. The macroconidia of *C. pentaseptata* (av. = 98 × 7 µm) are smaller than those of *C. pseudoreteaudii* (av. = 104 × 8 µm), and larger than those of *C. queenslandica* (av. = 69 × 6 µm), *C. reteaudii* (av. = 84 × 6.5 µm) and *C. terrae-reginae* (av. = 76 × 6 µm) (Lombard et al. 2010c). As with *C. queenslandica* and *C. terrae-reginae*, *C. pentaseptata* failed to produce microconidiophores and microconidia, distinguishing this fungus from *C. pseudoreteaudii* and *C. reteaudii*, which readily form these structures in culture (Lombard et al. 2010a, b, c).

One of two equally most parsimonious trees (TI = 380, CI = 0.942, RI = 0.921, RC = 0.868) obtained from a heuristic search with 1 000 random taxon additions of the combined sequences of TUB, HIS3 and TEF1-α sequence alignments of the *C. reteaudii* complex using PAUP v. 4.0b10. The bootstrap support values from 1 000 replicates are shown at the nodes. The tree was rooted to *C. chinensis* (CBS 112744) and *C. colombiensis* (CBS 112221). The ex-type strains are printed in **bold**.



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Ceramothyrium melastoma Crous & M.J. Wingf., *sp. nov.**Etymology.* Named reflects the host genus, *Melastoma*.

Description of colonies sporulating on synthetic nutrient-poor agar (SNA). *Mycelium* consisting of pale brown, septate, branched, finely verruculose, 2–3 µm diam hyphae. *Conidiophores* reduced to conidiogenous cells. *Conidiogenous cells* integrated, lateral on hyphae, phialidic with small collarete (flaring or not), 2 µm wide, 1–1.5 µm high. *Conidia* pale brown to subhyaline, subcylindrical to obclavate, apex subobtuse, base tapering, truncate, 1–12-septate, but commonly forming lateral branches as in *Stanhughesia* morphs of *Ceratothyrium* (especially on potato-dextrose agar (PDA) and malt extract agar (MEA), but less so on SNA), conidial body (25–)40–60(–90) × (2.5–)3 µm, lateral branches 7–25 × 2.5–3 µm. *Tripodosporium* morph on PDA and MEA: *central conidial body* 15–30 µm long, 3–4 µm wide at clavate apex, giving rise to two apical, lateral branches that angle upwards, of unequal length, lateral arms 15–35 × 2.5–3 µm; constricted at septa where lateral arms join the conidial body.

Culture characteristics — (in the dark, 25 °C after 2 wk): Colonies on MEA, PDA and oatmeal agar erumpent, spreading, with smooth, even margin and sparse aerial mycelium. Surface pale olivaceous-grey, reverse olivaceous-grey, reaching 5 mm diam.

Typus. INDONESIA, North Sumatra, Lake Toba, on leaves of *Melastoma* sp. (*Melastomataceae*), 20 Aug. 2011, M.J. Wingfield, holotype CBSH-21077, culture ex-type CPC 19837 = CBS 133576, ITS sequence GenBank KC005771, LSU sequence GenBank KC005793, MycoBank MB801771.

Notes — Based on a megablast search of NCBI's GenBank nucleotide database, the closest hits using the LSU sequence are *Phaeococcomyces catenatus* (GenBank AF050277; Identities = 847/875 (97 %), Gaps = 0/875 (0 %)), *Exophiala placitae* (GenBank EU040215; Identities = 841/871 (97 %), Gaps = 0/871 (0 %)), and *Sarcinomyces petricola* (GenBank FJ358249; Identities = 835/865 (97 %), Gaps = 0/865 (0 %)). Closest hits using the ITS sequence had highest similarity to *Trichomerium deniquatum* (GenBank JX313654; Identities = 559/664 (84 %), Gaps = 38/664 (6 %)), *Phaeococcomyces chersonesos* (GenBank AJ507323; Identities = 534/641 (83 %), Gaps = 43/641 (7 %)), and *Trichomerium gleosporum* (GenBank JX313656; Identities = 417/480 (87 %), Gaps = 18/480 (4 %)). *Ceramothyrium melastoma* clusters in a basal lineage to the *Chaetothyriales*, and renders *Ceramothyrium* paraphyletic. For a discussion on *Ceramothyrium*, see Fungal Planet 137.

Colour illustrations. Flower and leaves of *Melastoma* sp.; colonies growing on synthetic nutrient-poor agar; conidiogenous cells giving rise to conidia, which become star-shaped with age. Scale bars = 10 µm.

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Seiridium phylicae* Crous & M.J. Wingf., sp. nov.Etymology.* Name refers to the host genus, *Phyllica*.

Caulicolous. *Conidiomata* stromatic, pycnidia, scattered to aggregated, erumpent, conical, up to 350 µm diam, uniloculate, dark brown to black, opening by irregular rupture; basal stroma of dark brown *textura angularis*. *Conidiophores* lining cavity, filamentous, creating impression of paraphyses, septate, branched, hyaline, smooth, up to 80 µm long, and 3.5 µm wide. *Conidiogenous cells* subcylindrical, terminal and lateral, integrated, smooth, hyaline, 10–20 × 1.5–3 µm; proliferating percurrently. *Conidia* fusoid to ellipsoid, dark to golden brown, granular, 5-septate, not constricted at septa, with visible central septal pore, (23–)28–30(–35) × (9–)10(–11) µm; basal cell conical with truncate hilum, pale brown to hyaline, 3–5 µm long; 4 median cells doliform to subcylindrical, brown, with wall and septa being darker, cells together 17–23 µm long; apical cell broadly conical, apex rounded, hyaline, 2–4 µm long. Apical appendages tubular, unbranched, eccentric, 6–8 µm long; basal appendages unbranched, centric, 2–5 µm long.

Culture characteristics — (in the dark, 24 °C after 2 wk): Colonies erumpent, spreading, with moderate aerial mycelium and even, lobate margins. On malt extract agar surface pale olivaceous-grey, with patches of dirty white; reverse cinnamon. On potato-dextrose agar surface dirty white with patches of black sporulation; reverse dirty white. On oatmeal agar surface pale grey-olivaceous with patches of dirty white, reaching 30 mm diam.

Typus. UK, British Overseas Territory of Saint Helena, Ascension and Tristan da Cunha, Inaccessible Island, Blenden Hall, S37°17'41" W12°42'08", stems of *Phyllica arborea* (*Rhamnaceae*), Sept. 2011, P.G. Ryan, holotype CBS H-21089, cultures ex-type CPC 19962–19965 (CPC 19964 = CBS 133587), β-tubulin (TUB) sequence GenBank KC005819–KC005821, TEF1-α sequences GenBank KC005815–KC005817, ITS sequences GenBank KC005785–KC005785, LSU sequences GenBank KC005807–KC005810, MycoBank MB801788.

Notes — Conidia of *Seiridium cardinale* are 21–30 × 8–10 µm, with basal appendage being 1 µm long when present, and apical appendage 0.5–1.5 µm (Sutton 1980), which clearly distinguishes it from *Seiridium phylicae*. Based on a megablast search of NCBI's GenBank nucleotide database, the closest hits using the LSU sequence are *Seiridium eucalypti* (GenBank DQ414533; Identities = 833/833 (100 %), Gaps = 0/833 (0 %)), *Seiridium unicorn* (GenBank DQ414532; Identities = 833/833 (100 %), Gaps = 0/833 (0 %)) and *Lepteutypa cupressi* (GenBank AF382379; Identities = 872/875 (99 %), Gaps = 3/875 (0 %)). Closest hits using the ITS sequence had highest similarity to *Seiridium cardinale* (GenBank AF409995; Identities = 552/558 (99 %), Gaps = 2/558 (0 %)), *Seiridium cupressi* (GenBank FJ430600; Identities = 558/567 (98 %), Gaps = 4/567 (1 %)) and *Seiridium unicorn* (GenBank AF377299; Identities = 567/578 (98 %), Gaps = 2/578 (0 %)). Closest hits using the TUB sequence had highest similarity to *Seiridium cardinale* (GenBank DQ926973; Identities = 353/366 (96 %), Gaps = 3/366 (1 %)) and *Seiridium cupressi* (GenBank AF320495; Identities = 385/401 (96 %), Gaps = 2/401 (0 %)). Only distant hits (e.g. Identities = 218/249 (88 %), Gaps = 12/249 (5 %)) with *Pestalotiopsis* spp. were obtained when the TEF sequences were used in a megablast search.

Colour illustrations. *Phyllica arborea* growing on Inaccessible Island; colony on synthetic nutrient-poor agar; conidiophores, conidiogenous cells and conidia. Scale bars = 10 µm.

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REFERENCES

- Aa HA van der, Vanev S. 2002. A revision of the species described in *Phyllosticta*. Centraalbureau voor Schimmelcultures, Utrecht, The Netherlands.
- Azranlou M, Bakhshi M. 2011. *Microcyclospora rumicis*, a new species on *Rumex crispus* from Iran. *Mycotaxon* 118: 181–186.
- Azranlou M, Crous PW. 2006. *Strelitziana africana*. *Fungal Planet* No. 8. Centraalbureau voor Schimmelcultures, Utrecht, The Netherlands.
- Bensch K, Braun U, Groenewald JZ, Crous PW. 2012. The genus *Cladospodium*. *Studies in Mycology* 72: 1–401.
- Berbee ML, Pirseyedi M, Hubbard S. 1999. *Cochliobolus* phylogenetics and the origin of known, highly virulent pathogens, inferred from ITS and glyceraldehyde-3-phosphate dehydrogenase sequences. *Mycologia* 91: 964–977.
- Braun U. 1995. A monograph of *Cercosporiella*, *Ramularia* and allied genera (Phytopathogenic Hyphomycetes). IHW Verlag, München, Germany.
- Cannon PF, Hawksworth DL, Sherwood-Pike MA. 1985. The British Ascomycotina. An annotated checklist. Commonwealth Mycological Institute, Kew, Surrey, England.
- Cheewangkoon R, Groenewald JZ, Summerell BA, Hyde KD, To-anun C, Crous PW. 2009. Myrtaceae, a cache of fungal biodiversity. *Persoonia* 23: 55–85.
- Chomnunti P, Ko Ko TW, Chukeatirote E, Hyde KD, Cai L, et al. 2012. Phylogeny of the Chaetothyriaceae in northern Thailand including three new species. *Mycologia* 103: 382–395.
- Co-David D, Langeveld D, Noordeloos M. 2009. Molecular phylogeny and spore evolution of Entolomataceae. *Persoonia* 23: 147–176.
- Constantinescu O, Holm K, Holm L. 1989. Teleomorph-anamorph connections in Ascomycetes. 1–3. *Stanhughesia* (Hyphomycetes) new genus, the anamorph of *Ceramothyrium*. *Studies in Mycology* 31: 69–84.
- Coutinho ML, Miller AZ, Gutierrez-Patricio S, Hernandez-Marine M, Gomez-Bolea A, et al. 2012. Microbial communities on deteriorated artistic tiles from Pena National Palace (Sintra, Portugal). *International Biodeterioration & Biodegradation*. doi.org/10.1016/j.ibiod.2012.05.028.
- Crous PW. 1999. Species of *Mycosphaerella* and related anamorphs occurring on Myrtaceae (excluding *Eucalyptus*). *Mycological Research* 103: 607–621.
- Crous PW, Braun U. 2003. *Mycosphaerella* and its anamorphs. 1. Names published in *Cercospora* and *Passalora*. *CBS Biodiversity Series* 1: 1–571. Utrecht, The Netherlands.
- Crous PW, Braun U, Groenewald JZ. 2007. *Mycosphaerella* is polyphyletic. *Studies in Mycology* 58: 1–32.
- Crous PW, Braun U, Hunter GC, Wingfield MJ, Verkley GJM, et al. In press. Phylogenetic lineages in *Pseudocercospora*. *Studies in Mycology* 75: 37–114.
- Crous PW, Groenewald JZ. 2011. Why everlasting don't last. *Persoonia* 26: 70–84.
- Crous PW, Groenewald JZ, Shivas RG. 2010a. *Devriesia fraseriae*. *Fungal Planet* 65. *Persoonia* 25: 150–151.
- Crous PW, Groenewald JZ, Shivas RG. 2010b. *Strelitziana eucalypti*. *Fungal Planet* 62. *Persoonia* 25: 144–145.
- Crous PW, Groenewald JZ, Shivas RG, Edwards J, Seifert KA, et al. 2011. *Fungal Planet* description sheets: 69–91. *Persoonia* 26: 108–156.
- Crous PW, Schoch CL, Hyde KD, Wood AR, Gueidan C, et al. 2009a. Phylogenetic lineages in the Capnodiales. *Studies in Mycology* 64: 17–47.
- Crous PW, Summerell BA, Carnegie AJ, Wingfield MJ, Groenewald JZ. 2009b. Novel species of *Mycosphaerellaceae* and *Teratosphaeriaceae*. *Persoonia* 23: 119–146.
- Crous PW, Summerell BA, Carnegie AJ, Wingfield MJ, Hunter GC, et al. 2009c. Unravelling *Mycosphaerella*: do you believe in genera? *Persoonia* 23: 99–118.
- Crous PW, Summerell BA, Shivas RG, Burgess TI, Decock CA, et al. 2012. *Fungal Planet* description sheets: 107–127. *Persoonia* 28: 138–182.
- Crous PW, Wingfield MJ, Groenewald JZ. 2009d. Niche sharing reflects a poorly understood biodiversity phenomenon. *Persoonia* 22: 83–94.
- Crous PW, Wood AR, Okada G, Groenewald JZ. 2008. Foliicolous microfungi occurring on *Encephalartos*. *Persoonia* 21: 135–146.
- Decock C, Delgado Rodriguez G, Seifert KA. 2005. Phylogeny of *Synchaetomella lunatospora*, a new genus and species of synnematosus fungi from Southeast Asia. *Antonie van Leeuwenhoek* 88: 231–240.
- Ellis MB. 1971. *Dematiaceae* hyphomycetes. CAB International, Kew, UK.
- Farr DF, Bills GF, Chamuris GP, Rossman AY. 1989. *Fungi on plants and plant products in the United States*. APS Press, USA.
- Fernando AA, Currah RS. 1995. *Leptodontidium orchidicola* (*Mycelium radicans atrovirens* complex): Aspects of its conidiogenesis and ecology. *Mycotaxon* 54: 287–294.
- Flory SL, Kleczewski N, Clay K. 2011. Ecological consequences of pathogen accumulation on an invasive grass. *Ecosphere* 2: 1–12.
- Frank J, Crous PW, Groenewald JZ, Oertel B, Hyde KD, et al. 2010. *Microcyclospora* and *Microcyclosporella*: novel genera accommodating epiphytic fungi causing sooty blotch on apple. *Persoonia* 24: 93–105.
- Glienke C, Pereira OL, Stringari D, Fabris J, Kava-Cordeiro V, et al. 2011. Endophytic and pathogenic *Phyllosticta* species, with reference to those associated with Citrus Black Spot. *Persoonia* 26: 47–56.
- Groenewald JZ, Nakashima C, Nishikawa J, Shin H-D, Park JH, et al. In press. Species concepts in *Cercospora*: spotting the weeds among the roses. *Studies in Mycology* 75: 115–170.
- Hartley AJ, Mattos-Shipleay K, Collins CM, Kilaru S, Foster GD, Bailey AM. 2009. Investigating pleuromutilin-producing *Clitopilus* species and related basidiomycetes. *FEMS Microbiology Letters* 297: 24–30.
- Hausknecht A, Noordeloos ME. 1998. Neue oder seltene arten der Entolomataceae (Agaricales) aus Mittel- und Südeuropa. *Österreichische Zeitschrift für Pilzkunde* 8: 199–221.
- Hoog GS de. 1977. *Rhinocladiella* and allied genera. *Studies in Mycology* 15: 1–140.
- Hoog GS de, Weenink XO, Gerrits van den Ende AHG. 1999. Taxonomy of the *Phialophora verrucosa* complex with the description of two new species. *Studies in Mycology* 43: 107–121.
- Hyde KD, Frölich J, Taylor JE. 1998. Fungi from palms XXXVI. *Sydowia* 50: 21–79.
- Kang JC, Crous PW, Old KM, Dudzinski MJ. 2001. Non-conspecificity of *Cylindrocladium quinquesepatum* and *Calonectria quinquesepata* based on beta-tubulin gene phylogeny and morphology. *Canadian Journal of Botany* 79: 1241–1247.
- Kang JC, Hyde KD, Kong RYC. 1999. Studies on the Amphisphaeriales 1. The *Clypeosphaeriaceae*. *Mycoscience* 40: 151–164.
- Kang JC, Kong RYC, Hyde KD. 1998. Studies on the Amphisphaeriales 1. Amphisphaeriaceae (sensu stricto) and its phylogenetic relationships inferred from 5.8S rDNA and ITS2 sequences. *Fungal Diversity* 1: 147–157.
- Kleczewski NM, Flory SL. 2010. Leaf blight disease of the invasive grass *Microstegium vimineum* caused by a *Bipolaris* sp. *Plant Disease* 94: 807–811.
- Kleczewski NM, Flory SL, Clay K. 2012. Variation in pathogenicity and host range of *Bipolaris* sp. causing leaf blight disease on the invasive grass *Microstegium vimineum*. *Weed Science* 60: 486–493.
- Kobayashi T. 2007. Index of fungi inhabiting woody plants in Japan. Host, distribution and literature. *Zenkoku-Noson-Kyoiku Kyokai Publishing Co., Ltd., Japan*.
- Kornerup A, Wanscher JH. 1978. *Methuen handbook of colour*. Eyre Methuen Ltd., UK.
- Lombard L, Crous PW, Wingfield BD, Wingfield MJ. 2010a. Species concepts in *Calonectria* (*Cylindrocladium*). *Studies in Mycology* 66: 1–14.
- Lombard L, Crous PW, Wingfield BD, Wingfield MJ. 2010b. Phylogeny and systematics of the genus *Calonectria*. *Studies in Mycology* 66: 31–69.
- Lombard L, Zhou XD, Crous PW, Wingfield BD, Wingfield MJ. 2010c. *Calonectria* species associated with cutting rot of *Eucalyptus*. *Persoonia* 24: 1–11.
- Manamgoda DS, Cai L, McKenzie EHC, Crous PW, Madrid H, et al. 2012. A phylogenetic and taxonomic re-evaluation of the *Bipolaris* – *Cochliobolus* – *Curvularia* complex. *Fungal Diversity* 56: 131–144.
- Marvanová L, Pascoal C, Cássio F. 2003. New and rare hyphomycetes from streams of Northwest Portugal. Part I. *Cryptogamie Mycologie* 24: 339–358.
- Noordeloos ME, Gates GM. 2012. The Entolomataceae of Tasmania. *Fungal Diversity Research Series*, vol. 22.
- Overholts L. 1943. *Mycological notes for 1939–40*. *Mycologia* 35: 243–254.
- Park D. 1972. On the ecology of heterotrophic micro-organisms in freshwater. *Transactions of the British Mycological Society* 58: 291–299.
- Park RF, Keane PJ, Wingfield MJ, Crous PW. 2000. Fungal diseases of eucalypt foliage. In: Keane PJ, Kile GA, Podger FD, Brown BN (eds), *Diseases and pathogens of eucalypts*: 153–239. CSIRO publishing, Australia.
- Pereira JM, Barreto RW, Ellison AC, Maffia LA. 2003. *Corynespora casicola* f. sp. *lantanae*: a potential biocontrol agent from Brazil for *Lantana camara*. *Biological Control* 26: 21–31.
- Pisetta M, Montecchio L, Longa CMO, Salvadori C, Zottele F, Maresi G. 2012. Green alder decline in the Italian Alps. *Forest Ecology and Management* 281: 75–83.
- Quaedvlieg W, Kema GHJ, Groenewald JZ, Verkley GJM, Seifbarghi S, et al. 2011. *Zymoseptoria* gen. nov.: a new genus to accommodate *Septoria*-like species occurring on graminicolous hosts. *Persoonia* 26: 57–69.
- Rayner RW. 1970. *A mycological colour chart*. Commonwealth Mycological Institute, Kew, Surrey, England.

- Roldán A, Puig MA. 1992. Hifomycetos acuáticos en la cuenca del río Esva (Asturias, norte de España). *Anales del Real Jardín Botánico de Madrid* 17: 3–11.
- Ronquist F, Teslenko M, Mark P, Ayres D, Darling A, Hohna S, Larget B, Liu L, Suchard M, Huelsenbeck J. 2012. MrBayes 3.2: Efficient Bayesian phylogenetic inference and model choice across a large model space. *Systematic Biology* 61: 539–542.
- Rossmann AY, Aime MC, Farr DF, Castlebury LA, Peterson KR, Leahy R. 2004. The coelomycetous genera *Chaetomella* and *Pilidium* represent a newly discovered lineage of inoperculate discomycetes. *Mycological Progress* 4: 275–290.
- Saccardo PA. 1882. *Sylloge fungorum omnium hucusque cognitorum* I. Patavii, Italy.
- Seifert KA. 1985. A monograph of *Stilbella* and some allied hyphomycetes. *Studies in Mycology* 27: 1–235.
- Seifert KA, Nickerson NL, Corlett M, Jackson ED, Lois-Seize G, Davies RJ. 2004. *Devriesia*, a new hyphomycete genus to accommodate heat-resistant, cladosporium-like fungi. *Canadian Journal of Botany* 82: 914–926.
- Shimizu K, Tanaka C, Peng Y-L, Tsuda M. 1998. Phylogeny of *Bipolaris* inferred from nucleotide sequences of Brn1, a reductase gene involved in melanin biosynthesis. *Journal of General and Applied Microbiology* 44: 251–258.
- Shivas RG, Alcorn JL. 1996. A checklist of plant pathogenic and other microfungi in the rainforests of the wet tropics of northern Queensland. *Australasian Plant Pathology* 25: 158–173.
- Sivanesan A. 1987. Graminicolous species of *Bipolaris*, *Curvularia*, *Drechslera*, *Exserohilum* and their teleomorphs. *Mycological Papers* 158: 1–261.
- Sprague R. 1950. *Diseases of cereals and grasses in North America*. Ronald Press Co., New York, USA.
- Stukenbrock EH, Quaerndt W, Javan-Nikhah M, Zala M, Crous PW, McDonald BA. 2012. *Zymoseptoria ardabiliae* and *Z. pseudotritici*, two progenitor species of the septoria tritici leaf blotch fungus *Z. tritici* (synonym: *Mycosphaerella graminicola*). *Mycologia* 104: 1397–1407.
- Sutton BC. 1980. *The Coelomycetes. Fungi imperfecti with pycnidia, acervuli and stromata*. Commonwealth Mycological Institute, Kew, Surrey, England.
- Swofford DL. 2003. PAUP* 4.0b10. *Phylogenetic Analysis Using Parsimony (*and other methods)*. Version 4. Sinauer Associates, Sunderland, MA, USA.
- Tomlinson PB. 2006. The uniqueness of palms. *Botanical Journal of the Linnean Society* 151: 5–14.
- Vizzini A, Musumeci E, Ercole E, Contu M. 2011. *Clitopilus chrischonensis* sp. nov. (Agaricales, Entolomataceae), a striking new fungal species from Switzerland. *Nova Hedwigia* 92: 425–434.
- Yang ZL. 2007. *Clitopilus amygdaliformis*, a new species from tropical China. *Mycotaxon* 100: 241–246.
- Zhang R, Yang HL, Sun GY, Li HY, Zhuang JL, Zhai XR, Gleason ML. 2009. *Strelitziana mali*, a new species causing sooty blotch on apple fruit. *Mycotaxon* 110: 477–485.
- Zwickl D. 2006. *Genetic algorithm approaches for the phylogenetic analysis of large biological sequence datasets under the maximum likelihood criterion*. PhD thesis, The University of Texas at Austin.