

Gantenerumab: A Novel Human Anti-A β Antibody Demonstrates Sustained Cerebral Amyloid- β Binding and Elicits Cell-Mediated Removal of Human Amyloid- β

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Abstract. The amyloid- β lowering capacity of anti-A β antibodies has been demonstrated in transgenic models of Alzheimer's disease (AD) and in AD patients. While the mechanism of immunotherapeutic amyloid- β removal is controversial, antibody-mediated sequestration of peripheral A β versus microglial phagocytic activity and disassembly of cerebral amyloid (or a combination thereof) has been proposed. For successful A β immunotherapy, we hypothesized that high affinity antibody binding to amyloid- β plaques and recruitment of brain effector cells is required for most efficient amyloid clearance. Here we report the generation of a novel fully human anti-A β antibody, gantenerumab, optimized *in vitro* for binding with sub-nanomolar affinity to a conformational epitope expressed on amyloid- β fibrils using HuCAL[®] phage display technologies. In peptide maps, both N-terminal and central portions of A β were recognized by gantenerumab. Remarkably, a novel orientation of N-terminal A β bound to the complementarity determining regions was identified by x-ray analysis of a gantenerumab Fab-A β ₁₋₁₁ complex. In functional assays gantenerumab induced cellular phagocytosis of human amyloid- β deposits in AD brain slices when co-cultured with primary human macrophages and neutralized oligomeric A β ₄₂-mediated inhibitory effects on long-term potentiation in rat brain. In APP751_{swedish}XPS2_{N141I} transgenic mice, gantenerumab showed sustained binding to cerebral amyloid- β and, upon chronic treatment, significantly reduced small amyloid- β plaques by recruiting microglia and prevented new plaque formation. Unlike other A β antibodies, gantenerumab did not alter plasma A β suggesting undisturbed systemic clearance of soluble A β . These studies demonstrated that gantenerumab preferentially interacts with aggregated A β in the brain and lowers amyloid- β by eliciting effector cell-mediated clearance.

Keywords: Alzheimer's disease, A β , amyloid- β , antibody, HuCAL[®], immunotherapy

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INTRODUCTION

Alzheimer's disease (AD) is the world's leading cause of dementia and the most prevalent neurodegenerative disease. The majority, an estimated 50–75% of all dementia cases, are caused by AD. The disease is characterized by a progressive loss of cognitive function and because of the increasing prevalence, AD is expected to become a major challenge in increasingly aging societies. Currently available medications are based on symptomatic intervention but do not cure AD. The amyloid- β (A β) hypothesis suggests an age-dependent accumulation of the A β peptide as a critical step in the cascade of pathological events that cause deficits in synaptic functionality and lead to severe neurodegeneration in AD [1]. Thus, approaches to decrease accumulation of A β are actively explored as a therapeutic modality for treatment of AD.

Initial studies using active immunization with fibrillar A β in transgenic mouse models of AD demonstrated reduction of amyloid- β plaque pathology and ability to restore memory deficits [2, 3]. However, subsequent clinical evaluation of active immunization with fibrillar A β_{1-42} (AN-1792) revealed unwanted inflammatory reactions attributable to infiltrating T cells caused by the immunogen in combination with detergent and the used adjuvant QS-21 in 6% of patients suggestive for a deleterious Th1 response [4]. Several patients with anti-A β antibodies showed signs of plaque clearance at autopsy [5, 6] but subsequent clinical studies failed to show unequivocal improvement in patients with substantial levels of anti-A β antibodies [7, 8, 9]. As an alternative to active immunization, passive immunization demonstrated activity in transgenic animal models, e.g., [10, 11], through mechanisms that appear to include: (i) prevention of A β fibril formation [12], (ii) inhibition of neuro/synapto-toxic oligomers of A β [13], (iii) enhancement of cerebral A β efflux by a postulated equilibrium shift [11] or by receptor-mediated efflux of antibody-bound A β [14], and (iv) eliciting Fc receptor-mediated phagocytosis of antibody-bound A β [10].

An increasing number of anti-A β antibodies that were proposed to act by different, although not mutually exclusive mechanism(s), are currently explored as potential therapeutics for AD [15]. The molecular properties that determine successful therapeutic application of anti-A β antibodies are under debate and controlled clinical studies are needed to identify the major mechanism(s) of successful amyloid removal by antibodies. We identified specific anti-A β antibodies by screening a human phage display library

HuCAL[®]-Fab1 [16] and optimized selected candidates for specific and high avidity binding to A β plaques, both *in vitro* and *in vivo*. The finally selected antibody, gantenerumab, showed a unique molecular binding mode to the N-terminal portion of A β together with binding to central amino acids of the A β peptide, which is markedly different from other anti-A β antibodies previously described [17, 18, 19]. The amyloid- β lowering capacity was confirmed in an *ex vivo* phagocytosis assay using human effector cells and *in vivo* in PS2APP double transgenic mice. This is the first report of a fully human anti-A β monoclonal antibody characterized in preclinical models of AD focused on A β clearance mechanism(s). The activity of gantenerumab in these models indicates its potential for efficacy in humans.

MATERIALS AND METHODS

Monoclonal anti-A β antibodies

Gantenerumab was derived from a parental clone obtained from the MorphoSys HuCAL[®]-Fab1 phage display library [16] and engineered by *in vitro* maturation using CDR cassette exchanges [20]. Gantenerumab antibody material was obtained from a CHO production clone. Murine anti-A β antibodies BAP2 (specific for the A β peptide epitope comprising amino acids 2–8 [21]), BAP41 (specific for the A β peptide epitope comprising amino acids 5–10), and BAP44 (specific for the A β peptide epitope comprising amino acids 20–21) were generated and selected by using standard hybridoma technologies [22].

Affinity measurements by surface plasmon spectroscopy

Affinity was measured online with surface plasmon resonance using a Biacore3000 instrument. Purified human anti-A β antibody preparations with >99% monomer verified by size exclusion chromatography were analyzed.

Synthetic A β_{40} peptides were obtained from Bachem (Bachem, Inc., Switzerland). Solubilization and aggregate formation were adapted from published procedures [23]. Binding to monomeric A β peptides was studied with samples resolubilized in 1,1,1,3,3,3-hexafluoro-2-propanol (HFIP, Fluka 52512) at 1 mg/mL. A β monomers were immobilized at 5 response units (RU; equivalent to ~ 5 pg/mm²) on a Streptavidin sensor chip (SA BR-1000-32 from GE Healthcare).

A β oligomers were prepared by HFIP pre-treated synthetic A β ₄₀ and solubilized at 50 μ M in 10 mM Tris, pH 7. Care was taken to avoid extensive vortexing and to use pre-cooled buffer. Oligomer preparations were analyzed by analytical ultracentrifugation and revealed a mixture consisting predominantly of trimers, tetramers, and higher oligomeric states with residual minor fraction of monomers. This preparation of oligomers was stable for at least 2 weeks if stored at 4°C (data not shown). A β fibrils were obtained from synthetic A β ₄₀ peptides incubated at a concentration of 200 μ g/ml in 10 mM sodium-acetate buffer (pH 4.0) for three days at 37°C. Electron microscopic analysis confirmed either spheroidal (i.e., oligomeric) or fibrillar ultrastructure for the oligomers and fibril preparations, respectively (data not shown). A β ₄₀ fibrils and oligomers generated *in vitro* were coupled to a CM5 chip at 200 RU as described in instruction manual of the manufacturer. The equilibrium dissociation constant (K_D) values were obtained from equilibrium responses with a Scatchard type analysis by retrieving values for divalent binding from the maximum slope and values for monomeric binding from the minimum slope. The K_D values and the kinetic rate constants k_a and k_d were retrieved by a kinetic fit based on a monovalent binding model and plotting the equilibrium sensor responses versus the respective concentration and fitting the resulting curve with the mathematical equation of a Langmuir isotherm.

Epitope mapping

The amino acid sequence encompassing A β ₄₂ ISEVKM¹DAEF RHDSGYEVHH QKLVFFAEDV GSNKGAIIGL MVGGVVI⁴²ATV IV was divided into 43 overlapping decapeptides with a frameshift of 1 amino acid. 43 peptide sequences were used starting with ISEVKMDAEF and ending with GGVVIATVIV. The 43 decapeptides were synthesized with N-terminal acetylation and C-terminal covalent attachment to a cellulose sheet (“pepspot”) by a commercial supplier (Jerini BioTools). The cellulose sheet was incubated for 2 h on a rocking platform with gantenerumab at 1 μ g/ml in blocking buffer (50 mM Tris·HCl, 140 mM NaCl, 5 mM Na₂EDTA, 0.05% NP40 (Fluka), 0.25% gelatine (Sigma), 1% bovine serum albumin fraction V (Sigma), pH 7.4). The sheet was transferred to a semi-dry blotting stack with the peptide side facing a PVDF membrane (Biorad) of equal size. After blotting, reactive peptides were detected by goat anti-human IgG (H+L) labeled with fluorochrome IRdye800 (Rockland code# 609-132-123) added at 1 : 10,000 dilution

in Odyssey blocking buffer and further diluted 1 : 1 with PBS, 0.05% Tween20 and scanned at 800 nm by a fluorescence scanner (Odyssey).

X-ray crystallography

Fab fragments were purified after papain digestion of gantenerumab and the Fc and Fab fragments were separated on a Resource S column by ion exchange chromatography. Fab fragment containing fractions were concentrated to 15 mg/mL and crystallized by vapor diffusion in sitting drops. Gantenerumab Fab were crystallized out of 25% PEG 3350, 0.1 M Bis-Tris 6.5, and 0.2 M ammonium sulfate. The Fab/A β complex was obtained by soaking the crystals with a 10 mM peptide solution of A β ₁₋₁₁ or A β ₃₋₁₁ for 24 h. For data collection, crystals were cryoprotected by addition of 15% glycerol and flash frozen in liquid nitrogen. Data have been collected to a resolution of 1.8 Å at a wavelength of 0.9999 Å at the beamline X10SA at the Swiss Light Source (Villigen, Switzerland). Crystals belong to the monoclinic space group P2₁ with cell axes of $a = 71.14$ Å, $b = 66.24$ Å, $c = 105.16$ Å, $\beta = 99.1^\circ$. Programs [24] were used to process and subsequently refine the data by rigid body and positional refinement against an existing in house apo structure of the gantenerumab Fab fragment. Difference electron density was used to place the peptide by real space refinement. Manual rebuilding of protein and peptide was done with COOT [25]. In the final structure, the heavy chains excluding part of CDR H3, the complete light chains and the bound A β peptides (residues 1–11) were traced. Two sulfate ions, 731 water molecules and, at residue Asn52 of the heavy chains, one NAG sugar unit was modeled into the electron density.

Transgenic mice

Male and female APP751^{swedish} × PS2^{N141I} (PS2APP) [26] in-house strain Bl6.152 H on a homozygous C57BL/6 background were used at plaque-bearing ages between 4 and 10 months. Mice were intravenously (iv) injected with different doses of anti-A β antibodies and 20 mM histidine buffer with 140 mM NaCl and 0.01% Tween-20, pH 5.5 as vehicle control. For plaque binding analysis a single injection of gantenerumab was administered to approximately 7-month-old PS2APP mice and analyzed after 3 days or up to 9 weeks after gantenerumab administration. Antibody-injected mice were anaesthetized (~2 min in 5% v/v Forene™) and animals perfused with 20 mL PBS (~10 mL/min). Brains were removed

and immediately frozen on powdered dry ice. All procedures were conducted in strict adherence to the Swiss federal regulations on animal protection, to the rules of the Association for Assessment and Accreditation of Laboratory Animal Care and with the explicit approval of the local veterinary authority.

For chronic treatment, gantenerumab was administered to 5–6 month old male PS2A β PP mice by weekly iv bolus injections, 20 mg/kg body weight, via the tail vein for 5 months. The baseline level of A β plaque load at the beginning of the study was determined in untreated PS2APP mice. To prevent an antibody response to gantenerumab in chronically treated transgenic mice such mice received a non-lytic anti-CD4 monoclonal antibody acting on regulatory T cells [27]. A single iv injection of 0.5 mg of an anti-CD4 antibody (ATCC hybridoma clone GK 1.5) was given to all mice, including the wildtype C57BL/6 mice and PS2APP vehicle treated mice, to transiently deplete peripheral CD4+ T-helper cells one day before the start of gantenerumab administration. Presence of anti-drug antibodies against gantenerumab was monitored by surface plasmon resonance spectroscopy at monthly intervals by retroorbital blood sampling during the course of the study. Behavioral testing of locomotor activity and memory in the Morris water maze was done as described earlier [26]. After 5 month treatment, all groups learned to find the visible platform (data not shown). Unexpectedly, all groups had a spatial learning impairment, as shown by the lack of preference for the platform quadrant during the probe trial (data not shown). It is important to note that also in the vehicle-treated wildtype group animals did not learn the task. All animal experimentation was compliant with the guide of the institution in which the experiments were done and protocol approval by the Institutional Animal Care and Use Committee.

Quantification of A β peptides in plasma

Four month old PS2APP mice received a single iv injection of anti-A β antibodies gantenerumab, BAP41, and BAP44 at a dose of 20 mg/kg body weight and plasma concentrations of A β ₄₀ and A β ₄₂ peptides were determined after 3 days using Liquid Phase Electrochemiluminescence (LPECL) assay technology. The immunoassay was done according to manufacturer's instructions (Meso Scale Discovery, Gaithersburg, Maryland). Streptavidin pre-coated 96-well plates were incubated with blocking buffer (5% BSA, 20 mM Na-phosphate, 120 mM NaCl, 0.05% CHAPS, pH 7.4) for 2 h at room temperature. After

discarding blocking solution, plates were filled with 50 μ L/well of capture antibody 4G8-bio (Signet) diluted at 1 μ g/ml in assay buffer (20 mM Na-phosphate, 120 mM NaCl, 0.05% CHAPS, 0.5% BSA, 0.4% milk powder, 2 mM EDTA, 0.05% NaN₃, pH 7.4) and incubated for 2 h at room temperature. Plates were then washed three times with wash buffer (20 mM Na-phosphate, 120 mM NaCl, 0.05% CHAPS, pH 7.4) followed by addition of 50 μ L/well of either A β peptide standards, blanks or samples diluted 1 : 3 in assay buffer and incubated overnight at 4°C. After washing, either TAG-labeled antibody BAP24 (for A β ₄₀) or TAG-labeled antibody BAP15 (for A β ₄₂ [28]) was added to the plates and incubated for 3 h at room temperature. After washing, plates were filled with Mesoscale Read Buffer and read in a Sector Imager reader 6000 according to manufacturer's instructions. Data were statistically analyzed by one-way ANOVA with Tukey's multiple comparison test.

Quantification of A β peptides

A β ₄₀ and A β ₄₂ were extracted from PS2APP mice treated over 5 months with gantenerumab using a two-step protocol. Cerebral hemispheres were homogenized in 1 mL of DEA buffer (2% diethanolamine, 50 mM NaCl) using the MagNAlyser (F. Hoffmann-La Roche). The supernatants containing DEA soluble A β were collected after centrifugation and neutralized to pH 8 with 1M Tris-HCl. To extract insoluble A β , pellets were resuspended in 1 mL 9M urea, incubated overnight at 4°C and supernatants collected after centrifugation. Both supernatants (collected after either DEA or urea extraction) were tested in ELISA. The antibody combination used for the detection of A β peptides in brain extracts comprised biotinylated monoclonal antibody 4G8 (Senetek) for the capture and horseradish peroxidase (HRP) conjugated Roche in-house monoclonal antibodies HRP-BAP24 and HRP-BAP15 for the detection of A β ₄₀ and A β ₄₂, respectively. Control experiments addressing the influence of human anti-A β antibody were done by spiking brain homogenate from the vehicle treated group with gantenerumab up to 1 μ g/mL. No interference of gantenerumab in the A β measurements was observed. The ELISA was performed as previously described [26].

Long-term potentiation in vivo

Sprague Dawley rats (300–350 g) were anesthetized with an i.p., injection of urethane (ethyl carbamate, 1.5 g/kg) and supplementary injections were given

when necessary. Body temperature was maintained at 36–37°C and constantly monitored. Field excitatory postsynaptic potentials (fEPSPs) were recorded in the stratum radiatum of the hippocampus by stimulating the Schaffer collateral commissural pathway. For this, a recording electrode (TM33B10KT, WPI, Sarasota, FL) was positioned 3.4 mm posterior and 2.5 mm lateral to the midline and a stimulation electrode (PTN3CC02INS, WPI) 4.2 mm posterior and 3.8 mm lateral to the midline. For intracerebroventricular (icv) injection, a cannula (Introcan 18 G, B. Braun Melsungen AG) was placed in the lateral ventricle, 0.8 mm posterior, 1.5 mm lateral to the midline and 3.7 mm ventral to bregma. Test stimuli lasting for 100 μ s were delivered every 30 s at a current intensity evoking 50% of maximum fEPSP. Long-term potentiation (LTP) was induced by a high frequency stimulation (HFS) protocol consisting of 10 trains of 20 stimuli (200 μ s) at 200 Hz, 2 s intertrain interval at an intensity evoking a 75% maximum fEPSPs. The fEPSP slopes (30–70%) were normalized to the 10-min period before icv injection and represented as mean \pm SEM. A β_{42} (stored at –80°C) was solubilized to 50 μ M in 10 mM Tris, pH 7.4 and incubated overnight at 4°C (oligomer forming conditions). Immediately before use, the A β_{42} solution was diluted in Tris to a concentration of 100 nM. Vehicle or A β_{42} solutions (5 μ L) with and without gantenerumab or the N-terminal specific anti-A β antibody 6E10 (Senetek) were injected over a 5-min period 1 h before high frequency stimulation. LTP was induced with A β_{42} in absence and presence of gantenerumab. Statistical significance was assessed using one way ANOVA with Dunnett's post-hoc analysis for comparison to vehicle and Tukey's post-hoc analysis for pair-wise comparisons ($p < 0.05$ was considered significant).

Immunohistochemistry

Cryosections of mouse brains and postmortem human frontal cortices from AD cases at Braak stage VI [29] were prepared at 10 and 20 μ m nominal thickness, treated with ice-cold acetone and washed in PBS. Blocking of unspecific binding sites was done with 2% BSA or sequential incubation in Ultra V block (LabVision) for 15 min followed by a PBS wash and incubation in power block solution (BioGenex) with 10% normal sheep serum for 20 min. Gantenerumab was detected by an affinity-purified goat anti-human IgG (H+L) conjugated to Alexa555 or Cy3 (#A-21433, lot 54699A, Molecular Probes, or #109-165-003, lot 49353, Jackson Immuno Research)

at a concentration of 15 μ g/mL in 2% normal goat serum in PBS, pH 7.4 for 1 h at room temperature. Counterstaining for A β plaques was done by incubation with BAP2, a mouse monoclonal antibody against A β conjugated to Alexa488 at a concentration of 0.5 μ g/mL for 1 h at room temperature. For deconvolution image processing, 40 μ m thick slices were used after postfixation with 2% formaldehyde followed by double staining as described above. Triple staining against microglia was done after perfusion fixation with 4% formaldehyde and sequential of 40 μ m thick floating sections with a rat anti-mouse antibody that reacts against I-A/I-E MHC class II alloantigens present on activated microglia (clone 2G9, BD Biosciences Pharmingen, San Jose, CA) at 10 μ g/mL incubated overnight and detected by goat anti-rat IgG conjugated to Alexa 488 (Molecular Probes) followed by secondary antibodies against gantenerumab and BAP2 conjugated to Alexa647 at concentrations as described above. AD cases included in this study were obtained from the departments of neuropathology of the Sun Health Research Institute (University of Arizona) with informed consent obtained for the use of human brain tissue and for access to medical records for research purposes. Tissue was obtained and used in a manner compliant with the ethical standards of the Committee on Human Experimentation of the institution in which the experiments were done and in accord with the Helsinki Declaration of 1975 and was approved by the local Institutional Review Board.

Microscopy and image processing

Confocal images were recorded using a HCX PL APO 40 \times , NA 1.25 oil immersion or HCX PL APO 63 \times , NA 1.4 oil immersion objectives on a Leica SP2 confocal microscope and processed with IMARIS software (Bitplane, Switzerland). Relative fluorescence intensity of plaque bound gantenerumab was determined from 5 images per animal and three animals per dose group using the IMARIS MeasurementPro module. Images shown represent maximum intensity projections of the z-series images. Deconvolution was done with HUYGENS software (Scientific Volume Imaging, Netherland) and colocalization was analyzed using the IMARIS Coloc module. Plaque load in the phagocytosis assay was determined by the average surface area obtained from 5 images in grey matter regions and calculated using the IMARIS SurpassPro software module. Surface visualization of triple stainings was also created with IMARIS SurpassPro.

Quantification of plaque number in PS2APP mice was obtained from five sagittal sections per mouse within the hippocampal formation using every 10th section cut at 10 μm nominal thickness. After staining of sections with gantenerumab at 5 $\mu\text{g}/\text{mL}$ for 1 h at room temperature and detection by affinity-purified goat anti-human IgG (H+L) conjugated to Alexa555 at 15 $\mu\text{g}/\text{mL}$ for 1 h at room temperature, images were scanned with a GenePix Personal 4100A microarray scanner (Axon Instruments, now Molecular Devices, Sunnyvale CA, USA) and analyzed with MCID M7 Elite software (Interfocus Ltd., Haverhill, England). Calculations were made with common spreadsheet software (Microsoft Excel, Redmond, WA, USA). Statistical evaluation was done using a two-tailed, heteroscedastic *t*-test. Immunogold labeling and electron microscopy was done with gantenerumab at 10 $\mu\text{g}/\text{mL}$ and detected with goat anti-human detection antibody coupled to 10 nanometer colloidal gold. Immunogold stainings were done with synthetic A β_{42} fibrils and visualized by negative staining with uranyl acetate. Immunogold labeling of brain tissue sections was done after freeze substitution and low temperature resin embedding as previously described [26]. Images were recorded with a JEOL 1210 transmission electron microscope.

Isolation of primary human cells and phagocytosis assay

Monocytes were obtained from human peripheral blood mononuclear cells (PBMCs) from a buffy coat (obtained from a local blood bank) by Ficoll density centrifugation. Monocytes were isolated from PBMCs by magnetic labeling using MACS[®] separation (Miltenyi Biotec, Germany #130-091-153) that consists of the Monocyte Isolation Kit II for isolation of human monocytes through depletion of non-monocytes (negative selection). Monocytes were differentiated to macrophages by adding 0.3 $\mu\text{g}/\text{mL}$ human macrophage colony stimulating factor (GenScript Z02001). We used differentiated human macrophages cultured in RPMI 1640 (Gibco #61870-044) medium with 100 U/mL penicillin and 100 $\mu\text{g}/\text{mL}$ streptomycin (Gibco #15140-122).

Differentiated macrophages were incubated in an antibody-dependent cellular phagocytosis assay employing cryosectioned postmortem human AD brain sections as substrate. Human AD brain tissue sections from cortical regions (Braak stage VI) were prepared at a nominal thickness of 20 μm and placed onto removable poly-D-lysine coated 2-well culture

dishes (Biocoat[™] #40629). Brain sections were preincubated with different concentrations of gantenerumab for 1 h, washed with PBS before human primary cells were seeded at 0.8 to 1.5 $\times 10^6$ cells/mL and cultured at 37°C with 5% carbon dioxide for 2 to 3 days. An unrelated human IgG1 (Serotec, PHP010) antibody was used as an additional control. Detection of amyloid- β plaques was done after fixation with 2% formaldehyde for 10 min, washing and staining with BAP2 conjugated to AlexaFluor488 at 10 $\mu\text{g}/\text{ml}$ for 1 h at room temperature. Double-labeling of macrophages was done with antibodies against A β and gantenerumab as described above and lysosomal marker antibody against LAMP2 (RDI Division of Fitzgerald Industries Intl).

Pharmacokinetics

Each mouse received a single iv injection of gantenerumab at various doses. Plasma samples were analyzed for gantenerumab by a validated gantenerumab specific ELISA. Pharmacokinetic analysis was performed by standard non-compartmental analysis with the software package ToxKin 3 (Entimo AG, Berlin) using composite plasma concentration-time profiles for each dose group.

RESULTS

Generation and in vitro A β binding profile of gantenerumab

Gantenerumab represents a new type of a fully human monoclonal anti-A β antibody derived from a synthetic human combinatorial antibody library (HuCAL[®]) based on phage display [16]. The desired profile of the antibody was engineered successively by *in vitro* maturation on fibrillar A β using reiterative cycles of CDR optimization [30] and applying targeted CDR-diversification [20]. This allowed affinity optimization for A β binding at sub-nanomolar K_D values. The finally selected format of gantenerumab was an IgG1 molecule of Gm(17)(z) and Gm(1)(a) allotypes [31].

Equilibrium binding of gantenerumab to different aggregation states of synthetic A β_{40} , as measured by surface plasmon resonance under conditions that monitor the bivalent binding, revealed K_D values of 0.6 nM, 1.2 nM, and 17 nM for A β fibrils, A β oligomers, and A β monomers, respectively. The kinetically most stable complex formation was observed for A β fibrils (k_d

2.8×10^{-4} 1/s) and A β oligomers (k_d 4.9×10^{-4} 1/s) whereas for monomeric A β a higher dissociation rate was observed (k_d 1.2×10^{-2} 1/s) suggesting rapid exchange of antibody-bound monomeric A β .

Epitope mapping by pepspot analysis using overlapping decameric A β peptides revealed that gantenerumab recognizes two regions within the A β peptide (Fig. 1a). Reactivity was observed with peptides containing the N-terminal sequence EFR with the decapeptide EFRHDSGYEV showing the strongest signal. A second reactivity was observed with a central region of A β that was most pronounced with the decapeptide VFFAEDVGSN and both flanking decapeptides.

The binding of gantenerumab to the N-terminal part of A β was investigated in detail by x-ray diffraction and structural analyses. Crystallized Fab fragments of gantenerumab were soaked with peptide solutions of A β_{1-11} or A β_{3-11} . Structures of both complexes were determined with 1.8 Å and 2.0 Å resolution, respectively. Only minor differences between the two complex structures were noted. The average root mean square deviation over all atoms in a superimposition of the peptides was 0.36 Å. Comparison of gantenerumab Fab-A β structure with published A β complex structures available in the Protein Data Bank

(pdb) demonstrated a unique A β binding mode of gantenerumab [17–19] (Fig. 1b and c). For the heavy chain, when all Fab fragments were superimposed, gantenerumab-bound A β peptide was flipped by 180°. In contrast, conformations of bound A β peptide of all published anti-A β antibodies are markedly different. The conformational difference might be explained by the sequence differences in the CDRs between gantenerumab and the other anti-A β antibodies published so far.

In complex with gantenerumab A β_{1-11} is bound in an extended conformation in the groove defined by residues from the CDRs H1, H2, H3, and L3. All amino acids of the peptide are visible in the electron density map and participate in the interaction pattern with the heavy or light chain, at least by hydrogen bonding of the main chain nitrogen and carbonyl atoms, with the exception of Ser8 and Gly9. Both amino acids are located in a short γ -turn pointing away from the Fab fragment. The conformation of the turn is fixed by a main chain hydrogen bond between the backbone carbonyl atom of Asp7 and the backbone nitrogen of Tyr10. The first 4 residues of A β_{1-11} are predominantly bound to the heavy chain of gantenerumab whereas residues 5 to 11 contact both heavy and light chains. Residues Asp1 and Ala2 interact with the anti-

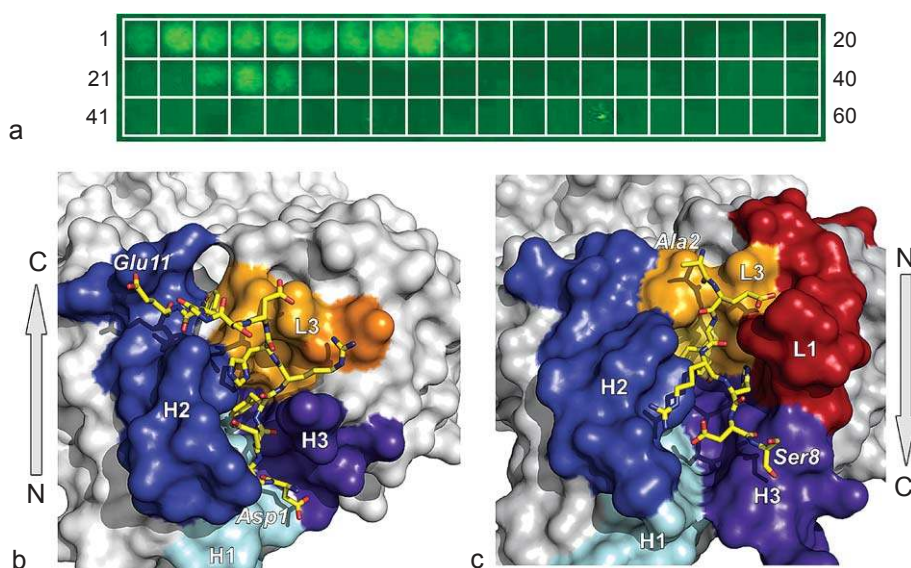


Fig. 1. Binding of gantenerumab to A β . Epitope mapping of gantenerumab by pepspot analysis with overlapping A β decapeptides (a). Pepspot signals of numbered overlapping decapeptide spots indicate recognition of multiple amino acids at the N-terminal and central part of A β . Strongest reactivity is seen with peptides 8 (AEFRHDSGYE), 9 (EFRHDSGYEV) at the N-terminus and peptide 24 (VFFAEDVGSN) within the central part of A β . Comparison of the atomic binding mode of A β_{1-11} peptide bound to gantenerumab (b) and WO2 (pdb entry 3bkj) (c). The surfaces of the Fab fragments are colored in grey except the CDRs which contact the A β peptides. H1 is shown in cyan, H2 in blue, H3 in purple, L1 in red, and L3 in orange. The orientation of the bound peptide with respect to the heavy chain is indicated by the arrow. The figure was generated with PyMOL (<http://pymol.sourceforge.net/>).

body mainly by hydrogen bonds formed by main chain atoms. An anchor point for the peptide is the side chain of Phe4 that is deeply buried in a hydrophobic pocket formed by amino acids Ile92L, M95L, Ile97L, Ala33H, Met34H, Ser35H, Ala50H, Val110H, and Phe113H. Further tight interactions were visible for Arg5 and His6. A more detailed description of the structural analysis of gantenerumab/A β complexes will be published elsewhere. Taken together, the crystal structure obtained from gantenerumab Fab with A β ₁₋₁₁ or A β ₃₋₁₁ peptides revealed a unique binding motif with frequent contacts of the A β peptides to CDRs 1, 2, and 3 of the heavy chain and CDR3 of the light chain, all likely contributing to the observed high avidity for A β plaques.

Immunoelectron microscopy of brain sections labeled with gantenerumab showed specific ultrastructural localization of gold label in close proximity to fibrillar structures present in A β plaques of AD patients and transgenic PS2APP mice. (Fig. 2a and b). Specific binding to the A β fibril surface was confirmed using synthetic A β ₄₂ fibrils immunolabeled *in vitro* (Fig. 2c). Immunofluorescence staining revealed specific binding of gantenerumab to all types of A β deposits. Typical dense core and diffuse A β plaques in human AD and murine PS2APP brain tissues were specifically labeled (Fig. 2d and e). The used immunohistochemical assay employed cryosections without chemical fixatives to preserve native antigen structures and more closely mimic *in vivo* tissue. The minimal effective concentration of gantenerumab for consistent staining of human A β plaques was 0.07 nM equivalent to 10 ng/mL (Fig. 2f). Our data suggest that comparable low concentrations of gantenerumab when present in brain interstitial fluid will result in effective cerebral A β binding *in vivo*.

Gantenerumab elicits clearance of human A β plaques ex vivo

To test whether gantenerumab reduces A β amyloid *ex vivo*, we developed a phagocytosis assay employing postmortem AD brain tissue slices cultured with primary human effector cells. We used differentiated primary human macrophages or primary human microglia prepared from human brain tissue after brain surgery. AD brain sections were preincubated with different concentrations of gantenerumab followed by incubation with effector cells in culture medium for 40–72 h. A β plaque load was assessed by immunohistochemistry and quantitative image analysis. A concentration-dependent decrease of A β plaque load

was seen (Fig. 3a–c). Notably, gantenerumab preincubated at 0.07 nM (10 ng/mL) was the minimal effective concentration ($p \leq 0.05$) able to lower A β amyloid in this assay (Fig. 3b and e) with amyloid removal highly significant ($p \leq 0.01$ to $p \leq 0.001$) and almost complete after preincubation with gantenerumab at ≥ 7 nM (Fig. 3c and e). The observed minimal effective concentration was consistent with the lowest effective concentration of gantenerumab to stain human A β plaques (Fig. 2f). EC₅₀ of gantenerumab in the phagocytosis assay was around 0.7 nM as determined in three independent experiments with macrophages from different human donors (Fig. 3f). Preincubation with gantenerumab up to 5 μ g/ml followed by incubation in culture medium in the absence of macrophages did not influence immunoreactivity with detection antibody BAP2 excluding a competitive effect of gantenerumab in the quantification of the amyloid load (Fig. 3d). Another control experiment with unspecific human IgG1 in the presence of macrophages showed unchanged A β load confirming that induction of phagocytic activity required gantenerumab (Fig. 3e). A similar gantenerumab dependent reduction of amyloid- β was obtained employing primary human microglia cells [32]. Double labeling of macrophages against A β and gantenerumab showed intracellular co-localization, demonstrating cellular uptake of gantenerumab-associated A β (Fig. 3g–j). Double labeling of A β and lysosomal marker LAMP2 revealed co-localization at vesicular structures indicative of lysosomal degradation of internalized A β amyloid (Fig. 3k–n).

Sustained binding of gantenerumab to A β plaques in vivo leads to reduction of small A β plaques

To examine the binding of gantenerumab to A β plaques *in vivo*, we administered the antibody systemically to PS2APP mice. Mice were sacrificed at various times after receiving different doses of gantenerumab and brain sections analyzed after staining against A β by confocal microscopy. Three days after dosing, the earliest time point investigated, gantenerumab was found associated with cerebral A β plaques. Specific binding to A β plaques was confirmed by co-localization stainings for A β . A typical image showing the reactivity of gantenerumab in PS2APP mice is shown for an immunodecorated A β plaque at high resolution obtained after deconvolution image processing (Fig. 4a–c). The entire plaque surface formed by filamentous structures in the depicted A β plaque has reacted with gantenerumab. Binding of

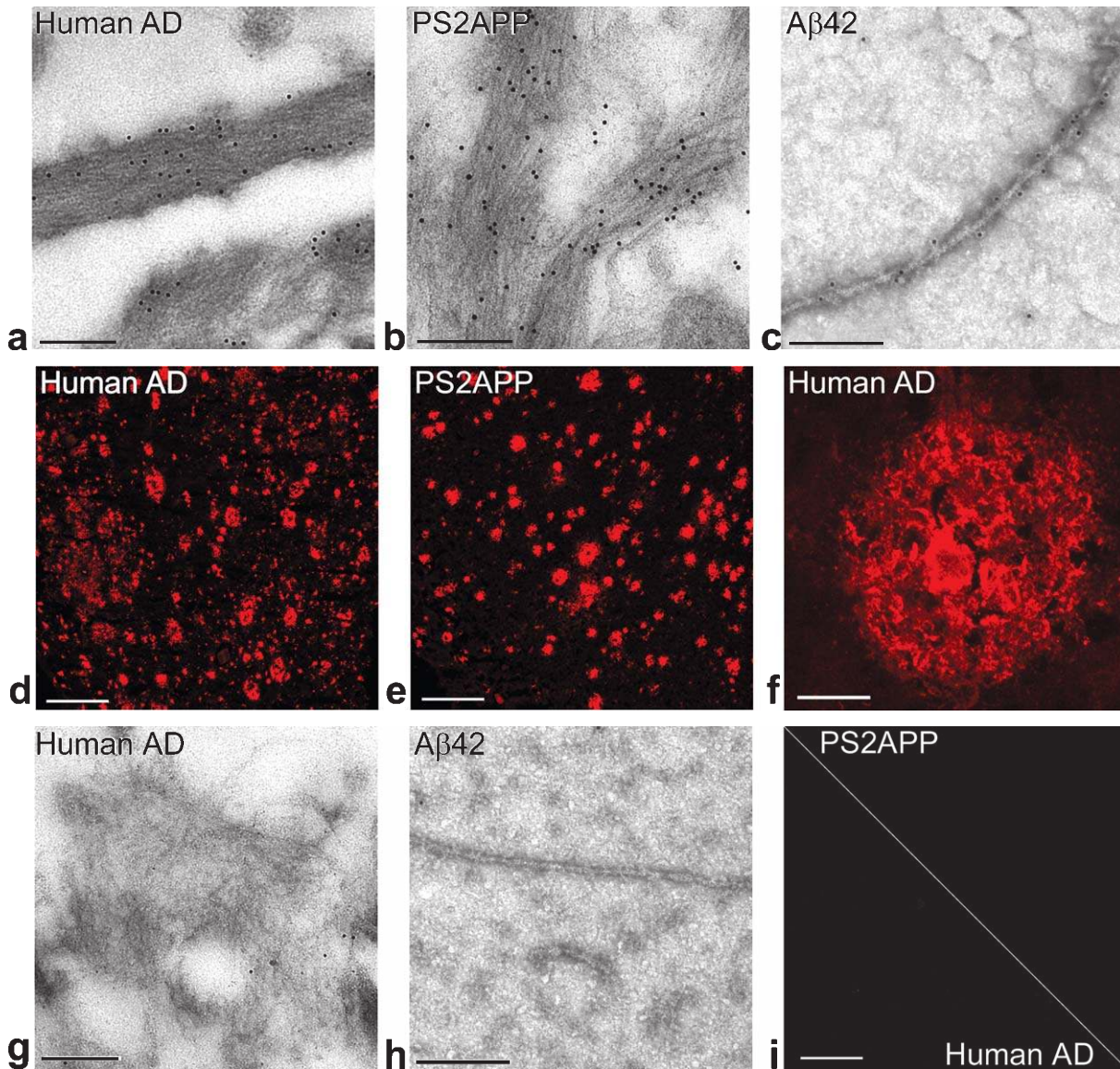


Fig. 2. Immunohistochemical localization of gantenerumab. Electron micrographs demonstrate binding of gantenerumab to A β fibrils within amyloid- β plaques from an AD patient (a) a PS2APP mouse (b) and synthetic A β_{1-42} fibrils (c). Gantenerumab was detected by a secondary goat anti-human antibody conjugated to 10 nm colloidal gold. Immunofluorescence staining of gantenerumab at 7 nM (1 μ g/mL) revealed specifically diffuse and dense-cored amyloid- β plaques in the frontal cortex of an AD patient (d) and PS2APP mouse (e). Staining of a human AD brain section with gantenerumab at 0.07 nM (10 ng/mL) demonstrated sensitive reactivity with a dense-core A β plaque (f). Controls with an unrelated human IgG1 antibody were negative for human amyloid- β plaques (g), A β_{1-42} fibrils (h) and immunofluorescence staining of human (left) and PS2APP (right) brain tissue (i). Bars on electron micrographs in a, b, c, g and h represent 200 nm. Bars in fluorescence micrographs, d, e and i represent 200 μ m except 20 μ m in f.

gantenerumab was dose-dependent with increasing fluorescence intensity at higher doses (Fig. 4d–f). Binding was observed with all A β plaque types and in all brain regions with plaque formation, similar to the examples obtained within the frontal cortex as shown in

Fig. 4. The reactivity of gantenerumab was consistently observed in all animals investigated demonstrating effective penetration into the brain.

Microglia cells are hypothesized to be mediators of A β clearance. We therefore applied a triple label-

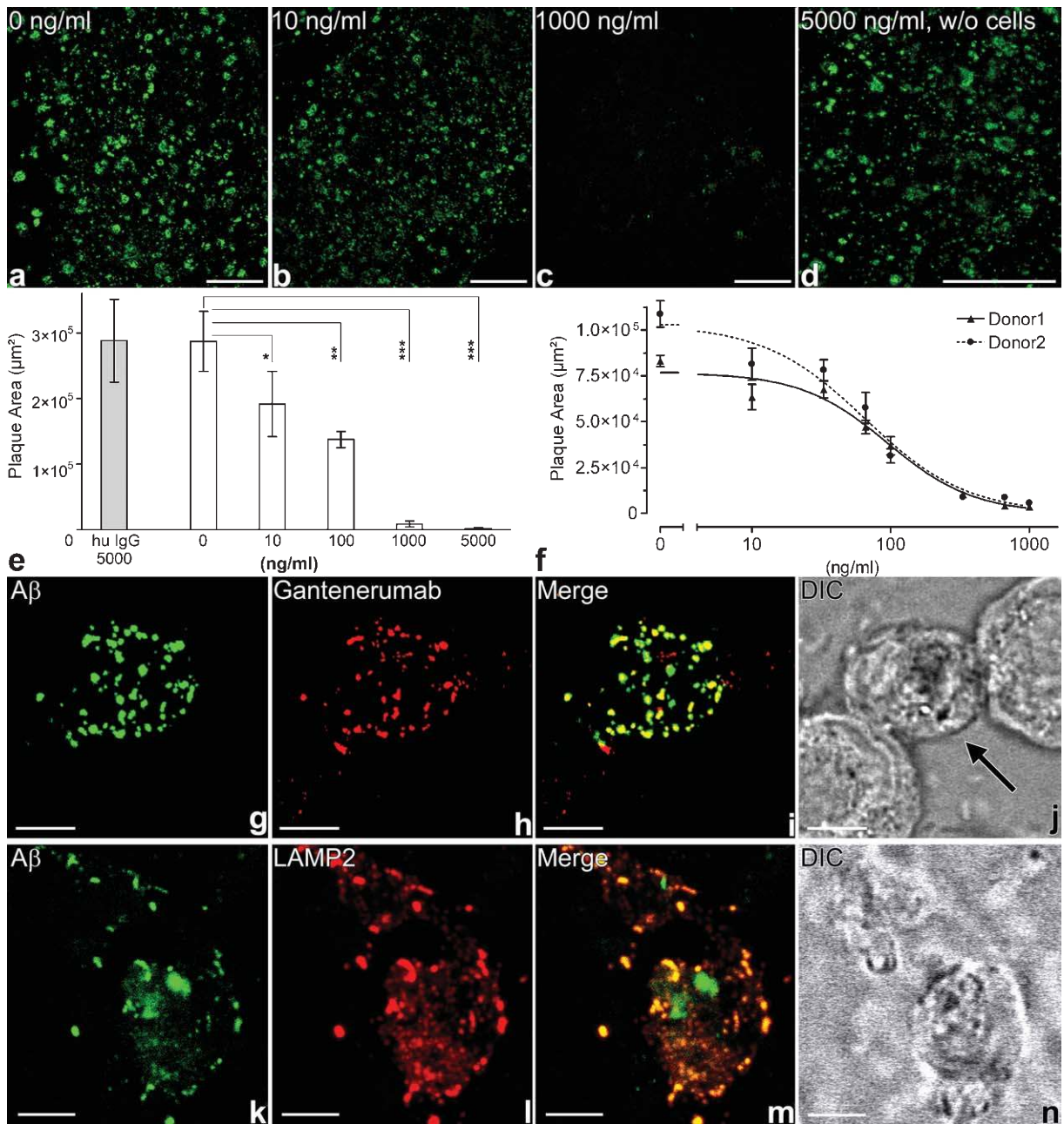


Fig. 3. Cellular phagocytosis of human amyloid- β plaques. Human AD brain sections were preincubated with gantenerumab followed by cell culturing in presence of primary human macrophages as effector cells. Plaques were labeled afterwards with an anti-A β antibody (BAP2) conjugated to Alexa488. Brain section without gantenerumab shows the distribution of numerous amyloid- β plaques in the grey matter of the frontal cortical lobe after culturing with human macrophages (a) and a concentration-dependent decrease of amyloid- β plaques when gantenerumab was added (b and c). Plaques were slightly reduced after preincubation with gantenerumab at 0.07 nM (10 ng/mL) (b) and substantially at 7 nM (1,000 ng/mL) (c). Control section preincubated with gantenerumab 35 nM (5,000 ng/mL) in the absence of cells indicated ability of BAP2 to label amyloid- β plaques (d). Quantitative analysis of the experiment shown in (a–c) revealed a significant concentration-dependent reduction of amyloid- β plaques, while control with unrelated human IgG1 antibody is inactive (e). Determination of EC50 showed a consistent plaque clearance by macrophages from two different human donors in additional independent experiments (f). Double-staining of human macrophages for A β and gantenerumab (g–j) and A β together with the lysosomal marker LAMP2 (k–n) indicated internalization of AD brain derived amyloid- β into vesicles. Differential interference contrast (DIC) images are shown for comparison to depict individual cells. The arrow in j points to a macrophage cell with numerous internalized complexes of colocalized A β and gantenerumab. Bars represent 300 μm in a–d, 5 μm in g–j and 3 μm in k–n. * $p \leq 0.05$, ** $p \leq 0.01$, *** $p \leq 0.001$.

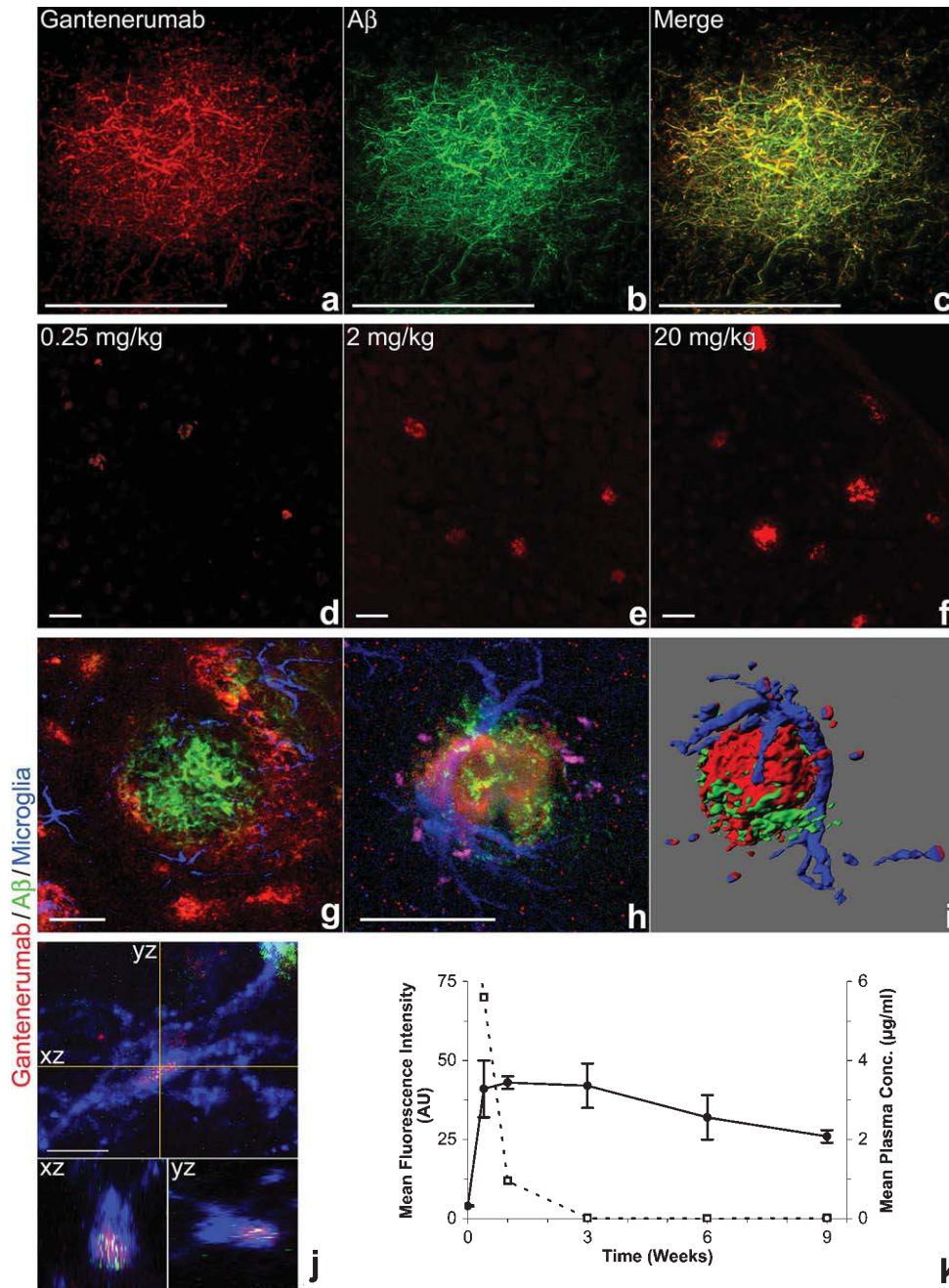


Fig. 4. Cerebral amyloid- β plaque binding of gantenerumab in PS2APP mice. High resolution confocal microscopy image of gantenerumab binding at amyloid- β plaque after a single iv injection of 20 mg/kg and imaging 3 days postdose (a-c). Gantenerumab was detected as described in Fig. 2. Structural details are shown after image deconvolution with strong reactivity of gantenerumab bound to an amyloid- β plaque (a) counterstained with anti-A β antibody BAP2 (b) demonstrating the amyloid- β specificity by the high degree of colocalization shown in the merged image (c). Gantenerumab binds dose-dependent to plaques as shown after one week iv bolus at indicated doses (d-f). Triple labeling of gantenerumab, A β and anti- I-A/I-E-antibody revealed microglia cells adjacent to gantenerumab bound to A β deposits (g). At higher magnification the close proximity of a microglia cell to a gantenerumab-reactive plaque (h) can be appreciated and is depicted in a volumetric visualization (i) of the microglia cell shown in h. Evidence for internalized gantenerumab that colocalized with A β in the cytoplasm of a microglia cell is shown in three different planes of view (j). The pharmacokinetic profile of gantenerumab bound to cerebral plaques showed sustained binding in PS2APP mice (k). Values represent mean fluorescence intensity at plaques in the occipital cortex over 9 weeks after a single iv bolus dose at 20 mg/kg. Error bars are \pm SD ($n=3$ animals per time point). The sustained binding of gantenerumab to plaques is markedly different from its peripheral mean plasma levels (dotted line). Bars represent 25 μ m in a-i and 5 μ m in j.

ing procedure to visualize gantenerumab, A β , and microglia. Microglia cells and microglia ramifications were frequently detected in the vicinity of plaques that reacted with gantenerumab (Fig. 4g–i). Occasionally, labeled gantenerumab and A β was detected intracellularly in the cytoplasm of microglia cells (Fig. 4j). These findings indicate recruitment of murine microglia by gantenerumab-reactive A β plaques and are consistent with a phagocytic plaque clearance mechanism.

To establish the kinetic profile of gantenerumab binding *in vivo* in the PS2APP model, we measured its reactivity with A β plaques for up to nine weeks. There was maximal binding of gantenerumab after two to three weeks that was consistently seen with different doses in all brain regions containing plaques, i.e., cortex, hippocampus, and thalamus. The binding of gantenerumab to A β plaques was sustained and detectable for over 2 months as shown for plaques in the frontal cortex (Fig. 4k). The minimal effective dose for detectable reactivity was 0.1 mg/kg after a single iv injection (data not shown). The pharmacokinetic profile of A β -bound gantenerumab in PS2APP mice was markedly different from the peripheral presence of gantenerumab in plasma where its half-life was 6.5 days and no longer detectable after three weeks (Fig. 4k; dotted line). Our findings demonstrate a consistent and long-lasting binding of gantenerumab to A β plaques in the brain of PS2APP mice.

To test whether gantenerumab elicited a lowering of the cerebral A β amyloid burden, we evaluated a five month treatment study in PS2APP transgenic mice. Since gantenerumab is a human antibody, mice were made tolerant by administering a single anti-CD4 injection one day before receiving gantenerumab. This treatment has been described to induce a long-lasting unresponsiveness in mice against human γ -globulin [33]. Potential anti-gantenerumab antibody responses were monitored monthly. No evidence for anti-drug antibodies was observed over the 5 months treatment period and residual plaques at the end of the study were consistently immunodecorated with gantenerumab in all animals.

Gantenerumab was administered by weekly iv bolus injections at 20 mg/kg and A β plaque load was determined by immunostaining and morphometrical analysis in the brain regions exhibiting substantial pathology. A significant reduction of A β amyloid plaques was evident in gantenerumab-treated PS2APP mice compared with vehicle controls (Fig. 5a and b). A detailed morphometric analysis revealed a significant ($p \leq 0.001$) reduction of A β plaque numbers in hippocampus (36%), neocortex (40%), and thalamus

(70%), respectively (Fig. 5c–e). Notably, the activity of gantenerumab correlated with plaque dimensions. The number of smaller, less than 400 μm^2 , A β plaques was most efficiently reduced when compared with vehicle treated animals (Fig. 5f). This reduction reflected not only a significant ($p \leq 0.001$) prevention of *de novo* plaque formation as revealed by comparison to the vehicle treated group, but also a significant ($p \leq 0.05$ to $p \leq 0.01$) clearance of pre-existing small plaques up to a size range of $\leq 300 \mu\text{m}^2$ when compared to the baseline group of PS2APP mice. The build-up of larger plaques with surfaces of $\geq 400 \mu\text{m}^2$ was similar for both the vehicle- and gantenerumab-treated groups after the 5 month treatment period (Fig. 5f) suggesting that larger plaques were more resistant to clearance in the PS2APP mouse model used in our study. By measuring the surface fraction of stained plaques, a significant decrease ($p \leq 0.001$) was seen in thalamus and a trend in cortex ($p = 0.07$) and hippocampus ($p = 0.23$) compared to vehicle treated animals (data not shown). Taken together these data suggested that the antibody-mediated clearing rate of large plaques is too slow to compete with the rate of A β accumulation in this aggressive transgenic mouse model of amyloidosis.

We have also analyzed the levels of both soluble and insoluble A β_{40} and A β_{42} in whole brain homogenates of PS2APP mice after 5 month treatment by ELISA. A small reduction of the mean levels of soluble A β_{42} (measured per mg wet brain) was detected in the gantenerumab-treated PS2APP mice with 1.07 ± 0.28 ng/mg (range 0.72 to 1.48 ng/mg) versus 1.28 ± 0.32 ng/mg (range 0.76 to 1.79 ng/mg) in the vehicle-treated group. However, the levels for soluble A β_{40} and insoluble A β after urea dissociation were not significantly different between treatment and control groups. A relatively high variability of insoluble A β levels among individual animals was noted in whole brain homogenates which could have masked the pronounced effect of gantenerumab on small plaques as determined by immunohistochemistry. In addition, as larger plaques ($\geq 400 \mu\text{m}^2$) were rather resistant to treatment, the biochemical analysis of total A β in whole brain extracts may not be sensitive enough to reveal the observed significant reduction of smaller plaques.

A similar finding was recently reported showing that the bulk of formic acid extractable A β was derived from dense core amyloid plaques and was not significantly affected by anti-A β antibody treatment. However, histological analysis based on silver staining revealed a marked reduction of small and diffuse amyloid plaques [34].

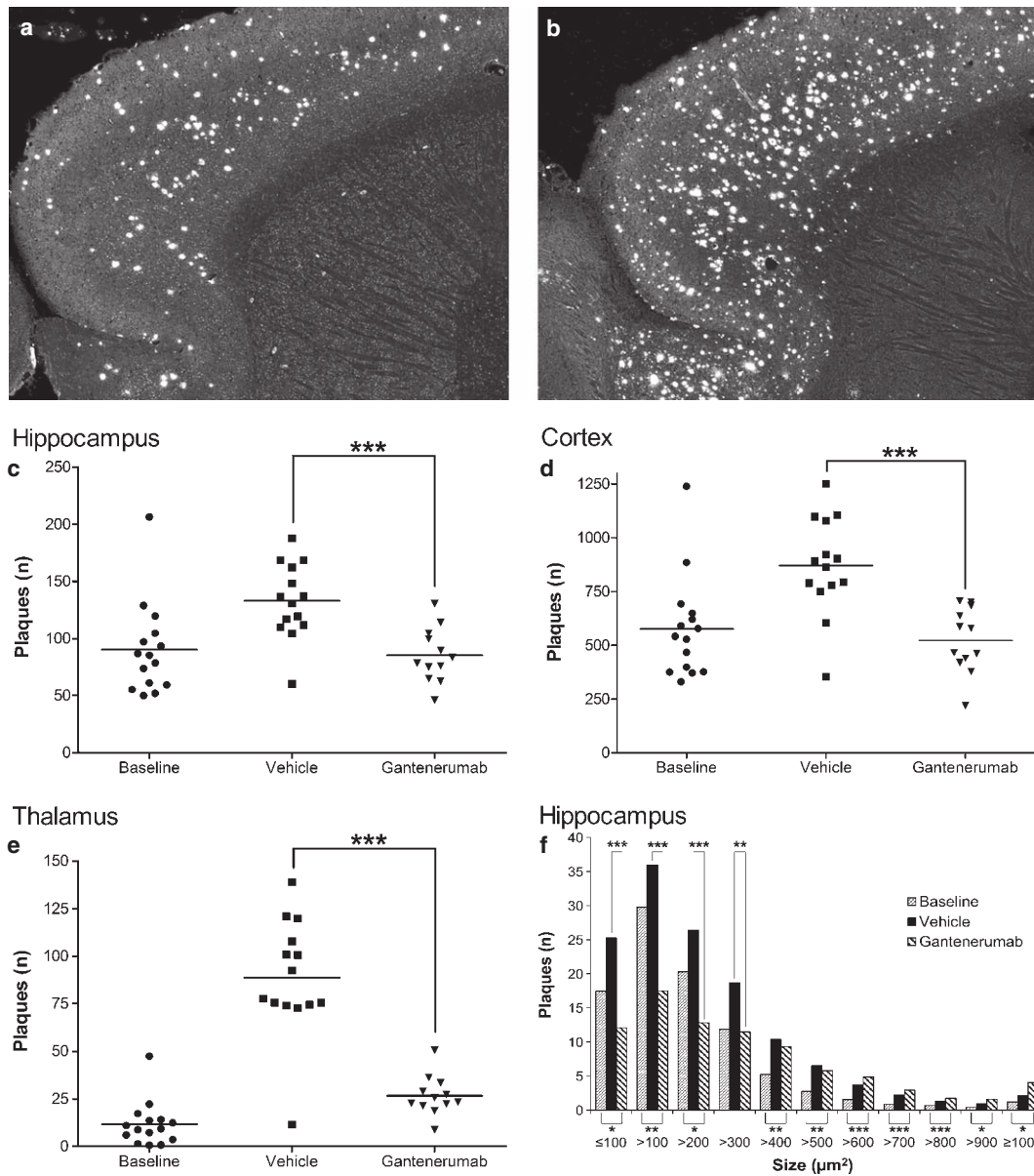


Fig. 5. PS2APP mice treated with gantenerumab for 5 months. Plaque bearing PS2APP mice were treated by weekly iv injections of gantenerumab at 20 mg/kg. Reduction of Aβ plaques was seen after treatment with gantenerumab (a) compared with vehicle (b). Quantitative morphometry after immunohistochemical staining of Aβ plaques is shown for the hippocampus (c), neocortex (d) and thalamus (e). Plaque load of untreated animals sacrificed at an age of 5 months is shown for comparison as baseline level of amyloidosis at study begin (day 0, $n = 15$). A significant reduction in plaque number is evident after treatment with gantenerumab ($n = 12$), compared to the progressive plaque formation seen in vehicle treated animals ($n = 14$). A significant clearance activity of gantenerumab was seen specifically for smaller amyloid deposits up to a size of $400 \mu\text{m}^2$ as shown for the hippocampal region (f). * $p \leq 0.05$, ** $p \leq 0.01$, *** $p \leq 0.001$.

Following 5 months of treatment with gantenerumab, there was no treatment-induced neurological or locomotor deficit and all groups learned to find the visible platform. Although no general behavioral deterioration was observed, cognitive assessment in the Morris water maze test was compromised by lack

of learning in wildtype and transgenic PS2APP mice injected with vehicle (control groups) as well as in gantenerumab treated PS2APP mice. Since learning was unexpectedly impaired in the wildtype animals, a stress-induced effect caused by weekly iv injections and monthly retro-orbital blood samplings are a likely

explanation for the observed learning deficit in the Morris water maze test.

Potential adverse effects in relation to the presence and reduction of amyloid plaques were also assessed in the five months study. No evidence of inflammatory reactions in general or other adverse effects were observed in this study. Gantenerumab was well tolerated and did not induce reactive inflammation following the degradation of AD plaques or exacerbate the manifestation of CAA or CAA-related microhemorrhage. Anti-gantenerumab antibodies were monitored once every month and were not detected up to the end of the study.

Gantenerumab treatment does not affect peripheral A β levels

To study whether gantenerumab affected peripheral A β levels, we analyzed plasma A β_{40} and A β_{42} after a single gantenerumab administration to PS2APP transgenic mice. For comparison we administered two murine anti-A β antibodies which recognize either an N-terminal epitope (BAP41) or a central epitope of A β

(BAP44) [22]. A β_{40} and A β_{42} plasma levels were measured 3 days after administration of the antibodies to PS2APP mice. No changes of total A β_{40} and A β_{42} were observed between vehicle and gantenerumab treated animals (Fig. 6). In contrast, treatment with BAP41 significantly ($p \leq 0.001$) elevated A β_{40} and A β_{42} concentrations by ≥ 20 fold. Similarly, BAP44 antibody significantly ($p \leq 0.001$) increased total A β_{40} and A β_{42} by 13 and 17-fold, respectively. These data demonstrated that systemic levels of soluble A β were not altered by gantenerumab. Presumably, due to the relatively fast off-rate of gantenerumab-bound monomeric A β the normal degradation pathway of circulating A β is not disturbed. A number of A β antibodies have been described which increase total peripheral A β levels upon administration to A β PP transgenic animal models or AD patients. This effect is thought to be due to stabilization of circulating antibody-bound A β interfering with the physiological systemic clearance of peripheral A β [35].

To establish a comprehensive pharmacokinetic profile of gantenerumab in mice, exposure levels and plasma concentration-time curves were determined.

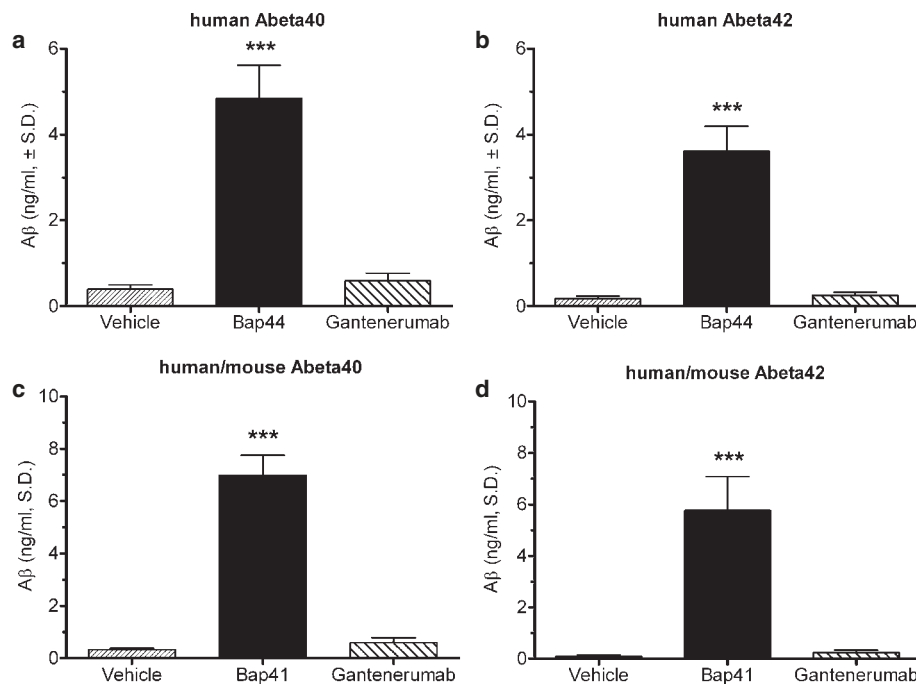


Fig. 6. Effect of gantenerumab on peripheral A β_{40} and A β_{42} . A β levels were determined in plasma of 4 month old PS2APP mice after a single iv injection of gantenerumab, BAP44, or BAP41 antibodies at 20 mg/kg and measured after three days. A β_{40} and A β_{42} were discriminated by C-terminal specific detection antibodies. BAP44 significantly increased peripheral A β_{40} (a) and A β_{42} (b). Likewise BAP41 also significantly elevated peripheral A β_{40} (c) and A β_{42} (d). Note that, in mice treated with BAP44 (recognizes A β at position 20-21) the antibody used to capture A β_{40} and A β_{42} , 6E10, recognized the N-terminal residues of human A β while in mice treated with BAP41 (recognizes A β at position 5-10), the antibody used to capture A β_{40} and A β_{42} , 4G8, reacts with both human and mouse A β . Remarkably, treatment with gantenerumab did not increase peripheral A β_{40} and A β_{42} levels. *** $p \leq 0.001$.

Gantenerumab exhibited a biphasic appearance in both C57BL/6 wild type and PS2APP transgenic mice with approximately dose-proportional increases in C_{max} and AUC parameters. Gantenerumab was cleared slowly, with clearance values of 0.0769 mL/min/kg and 0.0667 mL/min/kg in PS2APP and wildtype C57BL/6 mice, respectively. The initial volume of distribution (V_c) was about 38 and 50 mL/kg in PS2APP and C57BL/6 mice, respectively. The apparent volumes of distribution at steady state (V_{ss}) were markedly higher (around 300 mL/kg). During the β -elimination phase, plasma concentrations decreased in a log-linear manner with a half-life of 6.5 days in PS2APP mice. Thus, similar linear pharmacokinetics profiles of gantenerumab were obtained in both PS2APP mice and C57BL/6 wild type mice, consistent with the absence of a relevant target-mediated decrease of gantenerumab in PS2APP mice expressing human A β .

Gantenerumab neutralizes A β_{42} oligomers *in vivo*

To evaluate the ability of gantenerumab to neutralize neurotoxic A β_{42} oligomers that can interfere with synaptic function, we performed LTP measurements *in vivo* (Fig. 7). High frequency stimulation (HFS) of the Schaffer collaterals produced post-tetanic potentiation in the CA1 region of the hippocampus of anaesthetized, adult male Sprague Dawley rats. Stable LTP was obtained after icv injection of vehicle, while injection of A β_{42} oligomers lowered the induction of LTP after 5 min (Fig. 7a) and significantly ($p \leq 0.05$) blocked LTP after 150 min after HFS (Fig. 7b). Co-injection of A β_{42} oligomers with

gantenerumab or 6E10, attenuated the inhibition of LTP induction towards control levels after vehicle injection (Fig. 7a and b). The observed capacity of gantenerumab to prevent A β_{42} -oligomer induced inhibition of LTP in rat brain suggested that gantenerumab can effectively neutralize soluble A β_{42} aggregates and by consequence protect synaptic functionality.

DISCUSSION

Here we present the molecular and pharmacological profiles of a novel, human monoclonal IgG1 antibody, termed gantenerumab, which has been selected and optimized *in vitro* by phage display technologies. A number of murine anti-A β antibodies with remarkable amyloid lowering capacities in transgenic models of AD have been described in the past. Some of them have been humanized and are in clinical development as potential disease-modifying therapies based on the amyloid- β hypothesis of AD. Passive immunization holds promise for treatment of AD patients, and human rather than xenogenic antibodies are preferential choice for chronic administration.

While all of the A β antibodies described so far, including those in clinical testing, typically bind to one region of the A β sequence [15], gantenerumab binds to both N-terminal and central regions of A β as determined by solid phase peptide mapping analysis. The highest binding affinity of gantenerumab was observed for aggregated A β states suggesting recognition of a conformational epitope constituted by N-terminal and central A β residues in a configuration not present in

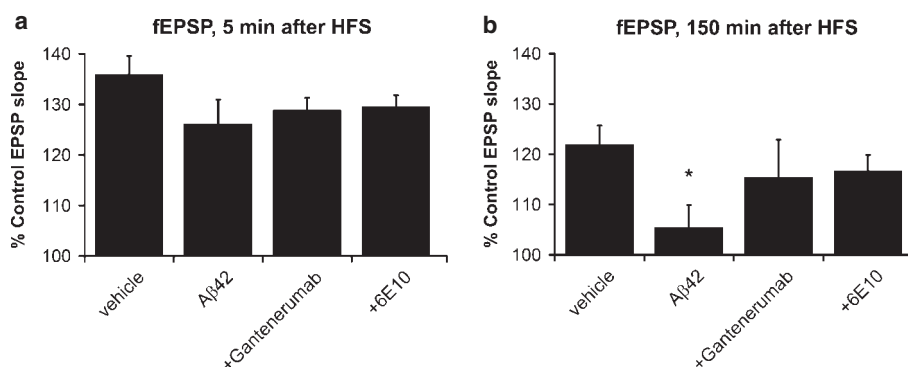


Fig. 7. Gantenerumab attenuates the inhibition of long-term potentiation (LTP) by A β_{42} in rat CA1 *in vivo*. After stable baseline recordings were obtained, rats were injected icv with 5 μ L of vehicle or 10 nM A β_{42} oligomers in the absence or presence of gantenerumab or control anti-A β antibody 6E10 at 1 μ g/ μ L. One hour later, high frequency stimulation (HFS) was issued that induced LTP in vehicle treated animals and in animals injected with soluble A β_{42} oligomers co-injected with gantenerumab or 6E10. Histograms represent normalized, mean fEPSP slopes \pm SEM of 5-6 animals normalized to the 10-min period before the HFS. For comparison, mean fEPSP values are shown 5 min (a) and 150 min (b) after HFS with a significant blocking of LTP induction seen after injecting A β_{42} oligomers after 150 min. Co-injection of gantenerumab or 6E10, effectively attenuated the A β_{42} oligomers induced inhibition of LTP induction. * $p \leq 0.05$.

the structure of monomeric A β . Since A β fibrils with well-defined spatial arrangement of A β segments were used for antibody selection and maturation *in vitro*, the identified A β binding properties of gantenerumab are unique and different from other antibodies raised by standard *in vivo* immunization procedures. These latter A β antibodies usually recognize linear epitopes of the most prominent immunogenic sequence present in the immunogen. Effective binding of gantenerumab to the supramolecular conformation of synthetic A β fibrils was also confirmed by immuno-electron microscopy. To further elucidate the A β -binding mode of gantenerumab the structure of a gantenerumab Fab-A β ₁₋₁₁ complex was determined by x-ray diffraction. Interestingly, the orientation of the N-terminal A β segment in the antigen binding cleft was opposite to what has been described for other Fab-A β complexes [17–19]. A high resolution structure of A β fibrils has been determined previously by NMR analysis demonstrating that the N-terminal and central portions of A β are exposed in close juxtaposition at the surface of the fibril [36]. Based on these data and the dual epitope recognition as measured in the dot blot assay we propose a binding mode of gantenerumab to fibrillar A β that involves both N-terminal and spatially adjacent central A β sequences (Fig. 8). According to this model the flexible N-termini of A β are the initial contact points of gantenerumab binding followed by interaction with the adjacent central A β part which confers increased binding avidity. Such a model is compatible with current structural information of A β fibrils and the gantenerumab Fab-A β ₁₋₁₁ complex but additional biostructural investigations are required to elucidate the exact atomic binding mode of gantenerumab to fibrillar A β .

The specificity of gantenerumab predicted strong binding to native A β plaques. Indeed, in brain sections from AD patients and PS2APP transgenic mice, highly efficient immunostaining of fibrillar A β was observed with gantenerumab at concentrations as low as 0.07 nM. In addition to combining specificity and high affinity binding, the therapeutic activity of anti-A β antibodies may require Fc functionality. Here we showed that gantenerumab has the capacity to clear human A β plaques from AD brain slices in the presence of human primary macrophages as effector cells but not in their absence, strongly suggesting the involvement of Fc receptor-mediated processes. Co-staining of A β , gantenerumab, and LAMP2 confirmed cellular uptake and lysosomal localization of antibody-coated A β . Notably, the minimal gantenerumab concentration to initiate A β clearance was

0.07 nM, i.e., the same concentration that was effective to yield significant A β plaque binding as measured by immunostaining. Similar results were obtained with primary human microglia cells [32] further supporting the hypothesis of Fc receptor/microglia-mediated phagocytosis and lysosomal degradation. Cellular phagocytosis was previously reported with murine anti-A β antibodies applied to transgenic mouse brain sections at 10 μ g/mL (\sim 70 nM) and cultured with primary murine microglia cells [10]. Differences in the assay conditions, higher binding affinity and/or mode of conformational epitope recognition might account for the several orders of magnitude higher potency of gantenerumab in the human *ex vivo* assay compared to the published murine assay results.

Immunoglobulins reach CSF/plasma ratios of 0.1 to 0.2% [37, 38] predicting that efficacious brain interstitial fluid concentrations of gantenerumab, i.e., in the subnanomolar range based on the *ex vivo* phagocytosis experiments, can be achieved at attainable plasma levels. While the exact mechanism of amyloid- β clearance is still a matter of debate, our data and earlier studies in transgenic mice [10] and AD patients [5, 6] suggest that microglia-mediated phagocytosis of antibody-coated A β plaques is a major mechanism of A β plaque clearance in the brain. In addition it is possible that other innate immune mechanisms such as complement activation also contribute to the clearance of amyloid- β plaques *in vivo*. Stimulation of microglia by A β -bound antibodies may be particularly efficient in AD patients who are compromised by dysfunctional macrophages and pathologically altered microglia with poor capacity to degrade A β [39, 40]. A vaccination study in vervet monkeys also supported the role of brain penetrating anti-A β antibodies in A β clearance since therapeutically active antibodies raised in these animals showed significant reactivity against fibrillar A β [41, 42].

Only partial and/or transient A β plaque reactivity, most prominent 1 h after injection, has been reported for murine anti-A β antibodies investigated in transgenic mouse models [43, 44]. In contrast, gantenerumab showed sensitive, dose-dependent and sustained binding to A β plaques in the brains of PS2APP mice that lasted more than 9 weeks after single administration indicating an extraordinary thermodynamic binding stability towards amyloid- β plaques *in vivo*. It is interesting to note that despite this significant “decoration” of A β plaques by gantenerumab large plaques ($>400 \mu\text{m}^2$) were relatively resistant to clearance as evident from the 5-months treatment study. Whether attenuated interactions of human Fc with

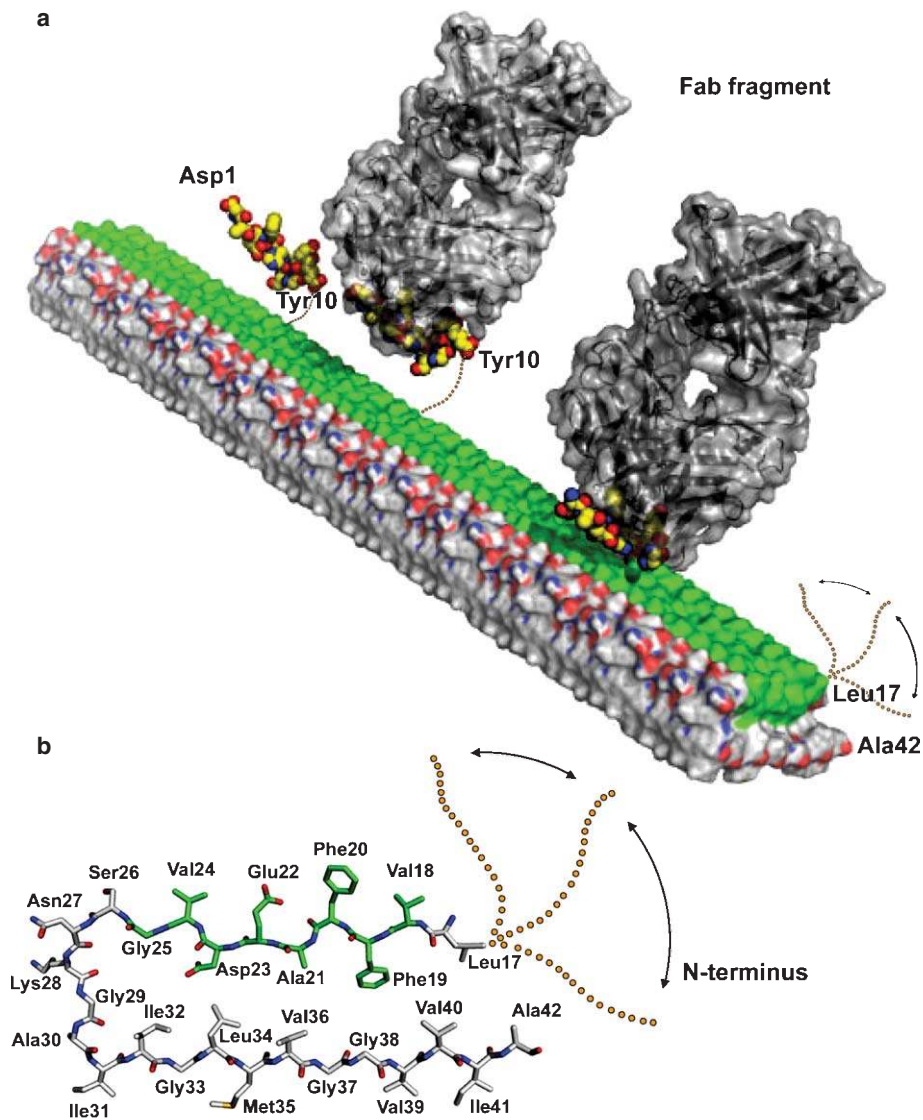


Fig. 8. Model of gantenerumab binding to the A β fibril surface. The binding of the gantenerumab Fab fragments to the A β fibril is schematically shown in a stepwise manner from left to right (a). The N-terminal part of A β is shown according to the atomic binding data obtained with gantenerumab shown for A β ₁₋₁₀ (DAEFRHDSGY). Initial binding to this free N-terminal part is followed in a second step involving the central A β epitope comprising amino acids 19–26 (VFFAEDVG) with their position highlighted in green on the fibril surface; the remaining surface is colored according to atom type with nitrogen and oxygen shown in blue and red. This dual mode of interaction is considered feasible by the spatial vicinity of both epitopes at the fibril surface and the flexibility of the N-terminus of A β . By consequence, an increased avidity of gantenerumab to fibrillar A β is suggested even in the presence of N-terminal truncated A β . The presented fibril structure is derived from data obtained by the NMR study by Lühns et al. [36]. A cross-sectional view perpendicular to the fibril axis showing one β -strand ordered stretch of A β ₁₇₋₄₂ with amino acids at position 19–26 shown in green (b). The approximate position of the flexible N-terminus at Leu17 is schematically illustrated with dotted lines in a and b.

murine microglia or other mechanisms contribute to the apparent persistence of these larger amyloid- β plaques needs further investigation. However, gantenerumab clearly elicited clearance of small-sized A β plaques and inhibited formation of new plaques after chronic treatment of PS2APP mice. A similar long-lasting binding of a murine anti-A β antibody

to amyloid plaques and resistance of thioflavine-S-positive dense cored plaques to antibody-mediated clearance was recently reported [34].

It is known that human IgG1 Fc binds less efficiently to murine Fc γ receptors [45] and activation of phagocytosis by microglia might not be optimally supported by a human IgG1 antibody like gantenerumab in a

murine animal model. While murine anti-A β antibodies have repeatedly been shown to lower amyloid- β load in a variety of transgenic mouse models likely involving activated microglia [46, 47], our study is the first to confirm the amyloid-lowering capacity of a fully human antibody in a transgenic mouse model. Gantenerumab is a human IgG1 which functionally corresponds to the murine IgG2a subtype. Murine IgG2a anti-A β antibody subtypes were shown to induce cellular phagocytosis and lower thioflavin-S positive amyloid plaques [48, 49]. In this context, a recent comparative study between murine 3D6 (IgG2a) and murine m266 (IgG1) anti-A β antibodies did not confirm removal of brain amyloid by m266 whereas the A β plaque binding 3D6 antibody was effective [50]. Besides the differences of the two antibodies in amyloid recognition (plaque-binding 3D6 versus A β peptide-capturing m266), the murine IgG1 isotype of m266 may also have contributed to the lack of efficacy in this model. In conclusion, the observed strong binding capacity of gantenerumab to amyloid plaques holds promise for effective treatment of human AD patients, in line with the notion that therapeutic efficacy likely requires the use of an antibody that can optimally cross the blood-brain-barrier [51].

Remarkably, gantenerumab did not increase peripheral A β_{40} or A β_{42} levels in PS2APP mice whereas massive increases of plasma A β have been reported for murine m266 and 3D6 antibodies in transgenic models [11, 50]. Both 3D6 and m266 when converted into humanized antibodies showed marked elevation of peripheral A β in AD patients [15, 52]. The similarity in the pharmacokinetic profiles of investigated anti-A β antibodies and A β indicates a stabilization of soluble A β by binding to the circulating anti-A β antibodies [35, 53]. In contrast to these antibodies, gantenerumab did not stabilize soluble A β in the periphery or in the brain, where binding to aggregated A β is thermodynamically preferred. Therefore, soluble monomeric A β does not compete with gantenerumab-binding to A β plaques *in vivo* but may interfere with 3D6 or m266 binding to aggregated A β once stable complexes with soluble monomeric A β are formed. Impaired receptor-mediated efflux of antibody-bound A β by FcRn receptors in the brain [14] appears less likely to account for the observed lack of peripheral A β increase in presence of gantnerumab since recent studies comparing wt and FcRn knockout mice suggest that FcRn does not significantly contribute to transport of IgG across the blood-brain-barrier in mice [54]. Our findings support the concept that amyloid- β lowering requires brain uptake of antibody

and binding to cerebral amyloid. However, in addition to microglia-mediated A β clearance through activation of Fc γ receptors it cannot be excluded that either direct dissociation of aggregated A β [12, 55] or other Fc independent pathways also contribute to some extent to A β lowering *in vivo* [56–58].

Similar to the N-terminal specific anti-A β antibody 6E10, gantenerumab was also shown to neutralize A β_{42} oligomer-induced inhibition of LTP in rat brain confirming previous reports [13, 59]. Thus, consistent with the high avidity binding to aggregated A β_{42} , gantenerumab can functionally neutralize oligomeric A β_{42} . The ability of gantenerumab to react not only with the A β fibrils and plaques but also with oligomeric A β might contribute to its therapeutic use by restoring synaptic deficits that are known to occur in AD.

In our studies in PS2APP mice after chronic treatment with gantenerumab, we did not observe an increased incidence of cerebral A β angiopathy (CAA) or microhemorrhages. Previous reports attributed these vascular side effects either to the epitope specificity of the anti-A β antibody, exemplified by the N-terminal specific 3D6 antibody [60] or the Fc glycosylation pattern [57]. However, the relevance of these observations for immunotherapy in AD patients is not established. Intriguingly, histopathological evaluations of post-mortem AD brains in follow-up studies of the AN1792 trial suggested that increased CAA pathology observed in immunized patients may be due to a transient perturbation of the perivascular drainage pathway [61].

In summary, our findings provide evidence that the specificity and avidity of gantenerumab facilitates sustained binding to cerebral A β *in vivo* leading to reduction of the amyloid plaque burden by triggering effector cell-mediated clearance mechanisms. Thus gantenerumab represents a potent human anti-A β antibody with a preference for aggregated A β and is considered as a promising agent for disease-modifying interventions in AD.

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