

Genetic Change Within Populations of *Phytophthora infestans* in the United States and Canada During 1994 to 1996: Role of Migration and Recombination

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ABSTRACT

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Dramatic changes occurred within populations of *Phytophthora infestans* in the United States and Canada from 1994 through 1996. Occurrence of the US-8 genotype, detected rarely during 1992 and 1993, increased rapidly and predominated in most regions during 1994 through 1996. US-7, which infected both potato and tomato and made up almost 50% of the sample during 1993, was detected only rarely among 330 isolates from the United States analyzed during 1994. It was not detected at all in more limited samples from 1996. Thus, ability to infect both potato and tomato apparently did not increase the fitness of this genotype relative to US-8, as predicted previously. US-1, the previously dominant genotype throughout the United States and Canada, made up 8% or less of the samples analyzed during 1994 through 1996. A few additional genotypes were detected, which could indicate the beginnings of sexual reproduction of *P. infestans* within the United States and Canada. However, clonal reproduction still predominated in all locations sampled; opportunities for sexual reproduc-

tion probably were limited, because the A1 and A2 mating types usually were separated geographically. The high sensitivity of the US-1 genotype to the fungicide metalaxyl also could have reduced opportunities for contact between the mating types in fields where this compound was applied. The previous correlation between metalaxyl sensitivity and genotype was confirmed and extended to a new genotype, US-17: all US-1 isolates tested were sensitive; all isolates of the US-7, US-8, and US-17 genotypes tested to date have been resistant. Isolates of *P. capsici* and *P. erythroseptica*, two other species often found on tomato and potato, could be easily distinguished from each other and from *P. infestans* using a simple allozyme assay for the enzyme *glucose-6-phosphate isomerase*. This technique could be useful for rapid identification of species, in addition to genotype of *P. infestans*. It generally was not possible to predict which genotypes would be present in a location from 1 year to the next. Long-distance movement of US-8 in seed tubers was documented, and this was probably the primary means for the rapid spread of this genotype from 1993 through 1996.

Additional keywords: DNA fingerprinting, fungicide resistance, late blight, migration, population genetics.

Late blight of potato and tomato, caused by *Phytophthora infestans* (Mont.) de Bary, reemerged as a serious disease in the United States and Canada during the early 1990s (15). The disease had not been particularly problematic during the 1970s and 1980s. Disease-control failures caused by strains resistant to the fungicide metalaxyl were first noted in the western United States during 1990 (4). Problems worsened during 1992 and, by the end of 1993, epidemics had been reported throughout the United States and western Canada (5,8,25), usually caused by metalaxyl-resistant strains (8,25). Severe epidemics continued in the eastern United States during 1994 and shifted to the western states during 1995 (15). The economic cost of late blight to U.S. potato and tomato growers during this period was estimated at more than \$230 million (14,26).

The increased disease severity coincided with major genetic changes in populations of *P. infestans* throughout the United States and western Canada (6,17,23). Allozyme and DNA fingerprint analyses revealed that most of the problems during 1992 and 1993

were caused primarily by two new genotypes (US-7 and US-8) that probably were introduced from northwestern Mexico during or shortly before 1992 (15,16,23). These genotypes were A2 mating type, resistant to metalaxyl (25), and highly pathogenic (24,27,29,30), which undoubtedly exacerbated the severity of the 1990s' epidemics (15,24).

US-7 was the predominant genotype during 1993 (23), both numerically (55% of all isolates analyzed) and geographically. It was found first in New York, Tennessee, and North Carolina during 1992, but was identified in 10 states during 1993 (23). This genotype is highly pathogenic to tomato in addition to potato (24,30), which may have given it a selective advantage. By contrast, during 1993, US-8 was found only in one county in Maine (23).

The rapid spread of US-7 and the persistence of the A1 US-1 genotype meant that both mating types were widely distributed during 1993 (5,23). However, opportunities for sexual reproduction probably were limited. Among hundreds of fields sampled, both mating types were found together only in a small number of fields in Florida (5,23), Texas (5), and British Columbia, Canada (23). More intensive sampling revealed that A1 and A2 coexisted in an additional 23 fields in British Columbia during 1993, in seven fields in New Brunswick during 1994 (2), and in one of six intensively sampled fields in Oregon and Washington during 1995 (P. B. Hamm, *personal communication*). However, probable sex-

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ual recombinants were found only in British Columbia during 1992 (23) and in the Columbia Basin of Oregon and Washington during 1993 (33). In Florida, Texas, and New Brunswick, coexistence of both mating types probably was too recent for the products of sexual reproduction to be detectable. Thus, as of 1993, although both mating types were present, there was only limited evidence of sexual reproduction in the Pacific Northwest and none in other areas of the United States.

The purpose of this study was to analyze populations of *P. infestans* during its recent resurgence in the United States to (i) identify which genotypes caused the devastating epidemics during 1994 through 1996; (ii) determine whether any additional migrations from Mexico or Europe had occurred since the last survey during 1993; and (iii) search for isolates with recombinant genotypes to determine whether sexual recombination was occurring more frequently in the United States. A secondary goal was to test whether genotype frequencies remained stable from year to year and thus could be used to predict the genetic composition of future populations. A final goal was to determine whether other species of *Phytophthora* that infect potato or tomato could be identified using the rapid cellulose-acetate electrophoresis system (21) used for identifying genotypes of *P. infestans*.

MATERIALS AND METHODS

Sources of isolates. Isolates of *P. infestans* were obtained from potato, tomato, or hairy nightshade (*Solanum sarrachoides* Sendtner) plants infected during calendar years 1994 through 1996. In addition, during 1993, 109 isolates from potato also were analyzed. Most isolates were obtained from infected leaf, stem, fruit, or tuber tissue sent to Cornell University (Ithaca, NY) or the Vegetable Laboratory (Beltsville, MD) from cooperators throughout the United States. Some tissue was collected by laboratory personnel directly, and other isolates were received as axenic cultures. In total, 556 isolates were obtained from at least 220 fields including 455 isolates from potato (185 fields), 95 from tomato (32 fields), and six

from hairy nightshade (3 fields). Fields were from commercial growers (some organic), home gardens, volunteer potatoes, or research plots and were separated spatially from other fields. Isolates were obtained by transferring fresh sporangia to agar media (usually rye B [1]) using a small agar block on the tip of a spatula as described previously (5,23). Tissue that was not sporulating was placed in a petri dish with a small amount of moistened filter paper at 18°C for 1 to 4 days to induce sufficient sporulation for isolation. For tissue that was beginning to decay, *P. infestans* was rescued by placing a fresh tuber slice (potato cv. Norchip, approximately 0.75 cm thick) on top of the infected tissue in a 9-cm petri plate. Usually, *P. infestans* would grow through the tuber slice and sporulate within 2 to 6 days. Clean-up medium (22), pea (17), and 10% V-8 juice (34) agars were occasionally used to make isolations. After isolation, cultures were grown on rye A agar (1). A subset of the isolates was stored cryogenically at -135°C.

In addition to *P. infestans*, isolates of two other *Phytophthora* species were analyzed during 1994. Four isolates of *P. capsici* were obtained from infected tomato fruits from a metalaxyl (Ridomil)-sprayed field near Sacramento, CA, and 21 isolates of *P. erythro-septica* were obtained from potato tubers in New York and Maine (20). The total number of isolates of all species analyzed was 581.

Isolate characterization. All isolates collected during 1993 and 1994 and some of those from 1996 were analyzed for mating type and for genotype at the two allozyme loci *glucose-6-phosphate isomerase (Gpi)* and *peptidase (Pep)*. Isolates from 1995 and some of those from 1996 were analyzed for mating type and *Gpi* genotype; others were analyzed for *Gpi* genotype alone at the Cornell University Diagnostic Laboratory. Mating type was determined by pairing each isolate with known A1 and A2 testers (17). Each tester was placed in the center of a 9-cm petri plate, and three unknowns were placed equidistant around the edges. Thus, only two plates were required to test three isolates. Ambiguous reactions were repeated as necessary to obtain clear results.

Tissue for allozyme and DNA fingerprint analyses was grown as described previously (17,21). Allozyme genotypes were deter-

TABLE 1. Characteristics and frequencies of genotypes of *Phytophthora infestans* detected among 556 isolates analyzed during 1993 to 1996 from the United States and Canada and a few isolates each of *P. capsici* and *P. erythro-septica* analyzed during 1994

Genotype	Mating type	Allozyme genotype		Metalaxyl response ^c	Frequency of detection during			
		<i>Gpi</i> ^a	<i>Pep</i> ^b		1993 ^d	1994 ^e	1995 ^f	1996 ^g
<i>P. infestans</i>								
US-1	A1	86/100	92/100	S	7.4	7.6	3.1	7.7
US-6	A1	100/100	92/100	S	11.9	1.2	0.0	0.0
US-6.5	A1	100/100	92/100	R	0.9	0.0	0.0	0.0
US-7	A2	100/111	100/100	R	37.6	5.8	43.0	0.0
US-8	A2	100/111/122	100/100	R	5.5	77.2	47.7	69.2
US-11	A1	100/100/111	100/100	R	0.0	6.7	6.2	0.0
US-12	A1	100/111	92/100	R	0.0	0.3	0.0	0.0
US-13	A2	100/100	100/100	R	0.0	0.9	0.0	0.0
US-14	A2	100/122	100/100	R	0.0	0.3	0.0	0.0
US-16	A1	100/111	100/100	R	0.0	0.0	0.0	7.7
US-17	A1	100/122	100/100	R	0.0	0.0	0.0	15.4
BC-1/US-7 ^h	A2	100/111	100/100	I/R	22.9	0.0	0.0	0.0
BC-1	A2	100/111	100/100	I	11.9	0.0	0.0	0.0
CA-8	A1	100/111/111	100/100	... ⁱ	1.9	0.0	0.0	0.0
<i>P. capsici</i>								
CAP-1	...	106/106	94/94	S	...	1.0
<i>P. erythro-septica</i>								
ERY-1	SF ^j	91/91	...	S, I, R ^k	...	1.0

^a *Glucose-6-phosphate isomerase.*

^b *Peptidase.*

^c S = sensitive, R = resistant, and I = intermediate. No intermediate isolates were tested in this study, but BC-1 isolates were scored intermediate previously (25).

^d N = 109.

^e N = 330.

^f N = 65.

^g N = 52.

^h These isolates probably were either BC-1 or US-7.

ⁱ Not determined.

^j Self fertile.

^k Reported previously (20).

mined by starch gel analysis (17) or with cellulose acetate (21). During 1996, *Gpi* genotype often was determined by analyzing mycelia or sporangia plucked or washed directly from infected tissue (21). Procedures for DNA fingerprinting were as described previously (19). Because of the predominance of the US-8 genotype (discussed in Results) and the high ability of allozyme analysis to identify recombinant genotypes in these populations, only a low number of isolates was scored for DNA fingerprint.

Metalaxyl sensitivity was determined for a small subset of the isolates using amended-agar or floating leaf-disc assays as described previously (25,32). Isolates were grown on rye B medium containing 0, 5, or 100 µg of metalaxyl per ml, and colony diameters were measured after 6 to 9 days. Some isolates were tested only on 0 and 100 µg of metalaxyl per ml. For the leaf-disc assays, only 0 and 5 µg of metalaxyl per ml were used, and the assays were scored after 5 to 8 days. Isolates of *P. capsici* were scored on amended-agar assays after 3 days. Isolates that grew more than 50% of the control on 5 µg of metalaxyl per ml were considered resistant; all others were scored sensitive.

RESULTS

Thirteen genotypes of *P. infestans* were found in the total sample of 556 isolates analyzed during 1993 through 1996 (Table 1). Genotype frequencies varied widely from year to year, and only two genotypes (US-1 and US-8) were detected during all 4 years (Table 1).

Genotype identification: 1993. Analyses of 181 isolates from 1993 were reported previously (23). The 109 additional isolates analyzed here were mostly from Texas and British Columbia and were of particular interest because these were among only five locations where both mating types have been found in the same field (5,23). Analysis of 38 isolates from Texas revealed that 12 were US-6 (A1 mating type) and 26 were US-7 (A2 mating type) (Table 2). Among 58 isolates from Canada, one (from Alberta) was US-1, one (from British Columbia) was US-6, one was US-6.5 (confirmed by DNA fingerprint analysis), and two had a new genotype, CA-8 (Table 2). The remaining 54 isolates were A2 mating type and *Gpi* 100/111, *Pep* 100/100, which corresponds to the US-7 and BC-1 genotypes. DNA fingerprint analysis of 14 of these isolates revealed that 4 were BC-1 and 10 were US-7. Identities of five additional US-7 and nine BC-1 isolates were confirmed by random amplified polymorphic DNA analysis (Z. K. Punja, unpublished data). The remaining isolates were not analyzed for DNA fingerprint, but were assumed to represent either US-7 or BC-1.

The remaining 1993 isolates were from seed tubers grown during 1993 that were collected and analyzed during early 1994. Two isolates, collected from Maine seed shipped to Florida during late 1993, had the US-8 genotype, as did four other isolates from Maine seed sent to New York during April 1994. Seven isolates, from North Dakota seed shipped into New York during April 1994, were US-1 (Table 2).

Genotype identification: 1994. Among the 330 isolates of *P. infestans* analyzed during 1994, 52 (16%) were A1 and the rest were A2. No self-fertile isolates were identified. The A1 and A2 mating types were found together only in 1 among the 134 fields sampled (Table 3); one A1 isolate was found in the same field as a US-8 genotype. Unfortunately, the A1 isolate from that field was lost before its genotype could be determined. Among the remaining isolates, eight genotypes were identified, only four of which were detected commonly (frequency >5%) (Table 1). By far the most commonly detected genotype was US-8, found on potato, tomato, and hairy nightshade (Table 3) throughout the United States and in New Brunswick, Canada. Two US-8 isolates were obtained from hairy nightshade growing in a cabbage field four miles from the nearest field of potato. US-8 was detected in 12 counties in New York during 1994 (Table 3), even though it was not detected at all in New York during 1993 (23).

TABLE 2. Sampling information and genotype for 109 isolates of *Phytophthora infestans* from potato during the 1993 growing season in the United States and Canada

State or province	County or city	Host cultivar	Month collected	No. of fields	No. of isolates	Genotype(s)
Alberta	Brooks	Russet Burbank	August	1	1	US-1
British Columbia	Chilliwack	... ^a	July	...	3	BC-1/US-7 ^b US-6.5 US-7
	Cloverdale	Eramosa	June	1	1	BC-1/US-7 ^b
		Norchip	July	1	1	BC-1/US-7 ^b
		Norkotah	July	1	1	BC-1/US-7 ^b
		Norland	July	1	4	BC-1/US-7 ^b
		Russet Burbank	June	1	1	BC-1
			July	3	1	US-7
			August	1	1	BC-1
			September	1	1	US-7
		Russet Norkotah	June	2	2	BC-1
					1	US-6
					2	US-7
			July	2	1	BC-1/US-7 ^b
					2	US-7
		Warba	June	2	1	BC-1/US-7 ^b
					2	US-7
		Yukon Gold	July	1	1	BC-1/US-7 ^b
		...	July	1	1	BC-1/US-7 ^b
	Delta	Norchip	June	1	1	US-7
		Red LaSoda	June	2	1	BC-1/US-7 ^b
					3	BC-1
		Russet Norkotah	June	1	2	BC-1/US-7 ^b
					1	BC-1
					1	CA-8
			August	1	1	BC-1/US-7 ^b
		Sunrise	July	1	1	BC-1
		Warba	July	1	1	BC-1/US-7 ^b
	Ladner	Warba	July	1	2	BC-1/US-7 ^b
	Okanagan	Nooksack	September	1	1	BC-1/US-7 ^b
		Russet Burbank	September	1	1	BC-1
					1	US-7
		Russet Norkotah	August	1	1	BC-1/US-7 ^b
	Vancouver Island	Russet Burbank	September	1	1	BC-1
		Russet Norkotah	August	1	1	CA-8
		Yukon Gold	August	1	2	BC-1
		...	August	1	1	BC-1/US-7 ^b
Maine	Aroostook	...	January 1994 ^c	1	2	US-8
		Norwis	April 1994 ^d	1	1	US-8
		Snowden	April 1994 ^d	1	3	US-8
North Dakota	...	FL 1533	April 1994 ^d	...	7	US-1
Texas	Cameron	FL 1533	Jan.–March	...	13	US-7
	Hidalgo	FL 1533	Jan.–March	...	12	US-6
			Jan.–March	...	13	US-7

^a Not known.

^b These isolates were A2 mating type and *glucose-6-phosphate isomerase* 100/111, *peptidase* 100/100. They were not tested for DNA fingerprint, but were assumed to be either BC-1 or US-7.

^c These were assayed from seed tubers sent to Florida. Even though they were collected during 1994, they reflected genotypes present in the field during the 1993 growing season. The remainder of the seed was planted.

^d These were assayed from seed tubers sent to New York. Even though they were collected during 1994, they reflected genotypes present in the field during the 1993 growing season.

The second most commonly detected genotype (8%) during 1994 was US-1 (Table 1). This genotype was found only in seed tubers from Nebraska and in potato fields in single counties in six states (Michigan, North Carolina, North Dakota, Oregon, Washington, and Wisconsin). Many New York potato growers planted infected seed after we determined that it contained US-1, yet US-1 was not recovered from the field in New York during 1994.

US-11 was the third most commonly detected genotype, making up 7% of the total sample (Table 1). The US-11 genotype was found in the Mt. Vernon variety trials in Skagit County, WA. This genotype had an unbalanced heterozygote pattern for the *Gpi* locus, with three bands in an approximately 4:4:1 ratio of intensities (Fig. 1). Thus, it probably has two copies of the 100 allele and was scored *Gpi* 100/100/111 (Table 1).

The only other widespread genotype was US-7 (6% of the total sample). This genotype was found only on tomato in five states, most of which were in the south (Table 3).

US-6, common during the early 1990s (17), was found only in two fields in Umatilla County, OR, during 1994 (Table 3) and made up only 1% of the sample that year (Table 1).

The remaining three genotypes were rare (less than 1% each) (Table 1) and were found only in our more extensive samples from New York (Table 3). One US-12 isolate was obtained from a tomato fruit from Maclean, NY. This genotype was A1 mating type and had a *Gpi* genotype like US-7, but a *Pep* genotype like US-1 or US-6. The US-13 genotype was found in two isolates from a commercial tomato field in Dryden, NY. These isolates were A2 mating type and 100/100 for *Gpi* and *Pep*. A third isolate from the same field also was A2 mating type, but was lost before its genotype could be determined. The final genotype, US-14, was found in a potato field that also had US-8. It differed from US-8 only in *Gpi* genotype: 100/122 for US-14 versus 100/111/122 for US-8.

Four isolates of *P. capsici* were tested during 1994, and all had the same genotype: *Gpi* 106/106, *Pep* 94/94. All 21 isolates of *P. erythroseptica* were *Gpi* 91/91 (Table 3). We could not obtain useful resolution of *Pep* genotypes of *P. erythroseptica*. Because the isozyme genotypes of *P. capsici* and *P. erythroseptica* were different from each other and from all of the *P. infestans* genotypes, it was easy to distinguish all three species on the basis of *Gpi* genotype alone.

TABLE 3. Sampling information and genotype for isolates of *Phytophthora infestans* (N = 330), *P. capsici* (N = 4), and *P. erythroseptica* (N = 21) collected in the United States and Canada during 1994

State or province	County	Host cultivar	Month collected	No. of fields	No. of isolates	Genotype	State or province	County	Host cultivar	Month collected	No. of fields	No. of isolates	Genotype
Isolates from potato						116	277						
Florida	Dade	... ^a	January	8	12	US-8	N. Dakota	Walsh	FL 1533	August	1	1	US-1
	St. Johns	...	July	4	5	US-8	Oregon	Umatilla	Russet Burbank	July	2	4	US-6
Georgia	Miller	Atlantic	May	2	6	US-8				August	1	1	US-1
Maine	Aroostook	FL 1533	July	2	2	US-8			Shepody	July	1	1	US-8
		Gold Rush	August	1	1	US-8			Washington ...	September	1	4	US-8
		Russet Burbank	August	1	1	US-8	Pennsylvania	Centre	Katahdin	September	1	2	US-8
		St. John	August	1	2	US-8			Erie	September	1	3	US-8
Michigan	Isabella	...	August	1	3	US-8			Potter	August	1	1	US-8
	Kent	...	August	1	3	US-8				August	1	3	US-8
	Mecosta	...	August	2	7	US-8				August	1	3	US-8
	Montcalm	...	August	13	49	US-8	Washington	Franklin	Ranger Russet	June	1	2	US-1
		...	October	4	5	US-8				July	2	4	US-1
	Presque Isle	...	October	3	7	US-1			Russet Burbank	August	1	1	US-1
		Onaway	October	1	2	US-1		Skagit	Halite	August	1	1	US-11
	Saginaw	...	November	1	4	US-8			Kennebec	August	1	1	US-11
Nebraska	Dawes	Red LaSoda ^b	October	1	3	US-1			Norkota	August	1	1	US-11
NB ^c	Carleton	Gold Rush	August	1	3	US-8			Ranger Russet	August	1	1	US-11
		Snowden	August	1	4	US-8			White Rose	August	2	10	US-11
New York	Cayuga	...	September	1	2	US-8			...	August	2	7	US-11
	Chautauqua	Kennebec	September	1	2	US-8			...	August	1	1	US-11
	Erie	...	September	1	1	US-8	W. Virginia	Preston	Elba	August	1	1	US-8
	Niagara	Chippewa	November	1	2	US-8			Katahdin	August	1	1	US-8
	Oneida	...	August	3	9	US-8	Wisconsin	August	...	2	US-1
	Ontario	Snowden	September	1	6	US-8	Isolates from tomato						
		...	October	1	2	US-8	Florida	Manatee	...	April	2	8	US-7
	Steuben	657	August	2	2	US-8	New York	Chautauqua	...	September	3	14	US-8
		Atlantic	August	1	3	US-8			Genesee	September	1	1	US-8
		FL 1533	September	1	1	US-8			Ontario	Sept., Oct.	2	6	US-8
		Monona	Sept., Oct.	2	3	US-8			Steuben	August	1	3	US-8
		...	Aug.-Oct.	14	34	US-8,			Tompkins	September	1	3	US-7
					1	US-7 ^d			...	September	1	1	US-12
					1	US-14 ^e			...	September	1	3	US-13
	Tioga	...	August	1	5	US-8	S. Carolina	Charleston	...	June	1	4	US-7
	Tompkins	...	September	1	4	US-8	Tennessee	Rhea	...	September	1	3	US-7
	Unknown	...	September	1	1	US-8	Virginia	Giles	...	August	1	1	US-7
	Washington	...	September	1	3	US-8	Isolates from hairy nightshade (<i>Solanum sarrachoides</i>)						
		Katahdin	October	1	1	US-8	New York	Ontario	...	September	2	4	US-8
	Wyoming	...	August	1	3	US-8			Orleans	September	1	2	US-8
		Kanona	September	1	1	US-8	Isolates of <i>Phytophthora capsici</i>						
		Monona	September	1	3	US-8	California	Sacramento	...	September	1	4	CAP-1
N. Carolina	Camden	...	May	1	2	US-1	Isolates of <i>Phytophthora erythroseptica</i>						
	Pamlico	...	June	1	1	US-8	Maine	Aroostook	1	1	ERY-1
	Washington	...	June	1	4	US-8	New York	Orleans	...	October	1	3	ERY-1
N. Dakota	Pembina	FL 1533	August	1	3	US-8		Suffolk	Norwis	November	6	17	ERY-1

^a Not known.

^b Isolated from seed tubers sent to Florida.

^c New Brunswick.

^d One A1 isolate was found in a field with a US-8 isolate, but was lost before its genotype could be determined.

^e One US-14 isolate was found in a field with a US-8 isolate.

DNA fingerprint analysis of 15 isolates confirmed that 10 with mating type and allozyme genotype like US-8 (from Pennsylvania, Maine, and four counties in New York) plus the US-14 isolate had the characteristic US-8 fingerprint pattern. Representative isolates of the US-1, US-11 (two isolates), and US-12 genotypes also were tested for DNA fingerprint (Fig. 2).

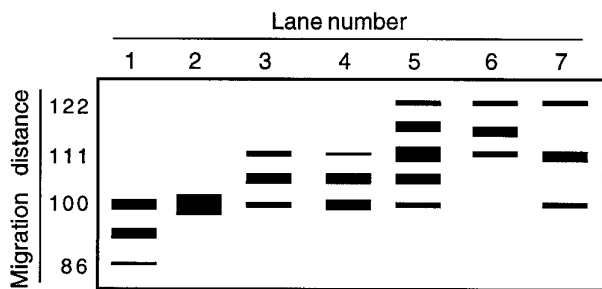


Fig. 1. Schematic gel-banding patterns at the *glucose-6-phosphate isomerase* (*Gpi*) locus for the most commonly detected genotypes of *Phytophthora infestans* in the United States and Canada. The *Gpi* genotypes and associated clonal lineages of *P. infestans* are as follows: lane 1, *Gpi* 86/100/100, characteristic of the US-1 clonal lineage; lane 2, *Gpi* 100/100, US-6; lane 3, *Gpi* 100/111, US-7 and BC-1; lane 4, *Gpi* 100/100/111, US-11; lane 5, *Gpi* 100/111/122, US-8; lane 6, *Gpi* 111/122, US-10; and lane 7, *Gpi* 100/122, US-17. Note the unbalanced heterozygote patterns in lanes 1 and 4, most likely caused by two copies of the *Gpi* 100 allele. Unbalanced heterozygotes are characteristic of the US-1 and US-11 genotypes and have three bands in an approximately 1:4:4 ratio of intensities (4:4:1 for US-11). Compare these with the balanced heterozygote patterns in lanes 3, 6, and 7, in which the three bands are in an approximately 1:2:1 ratio of intensities.

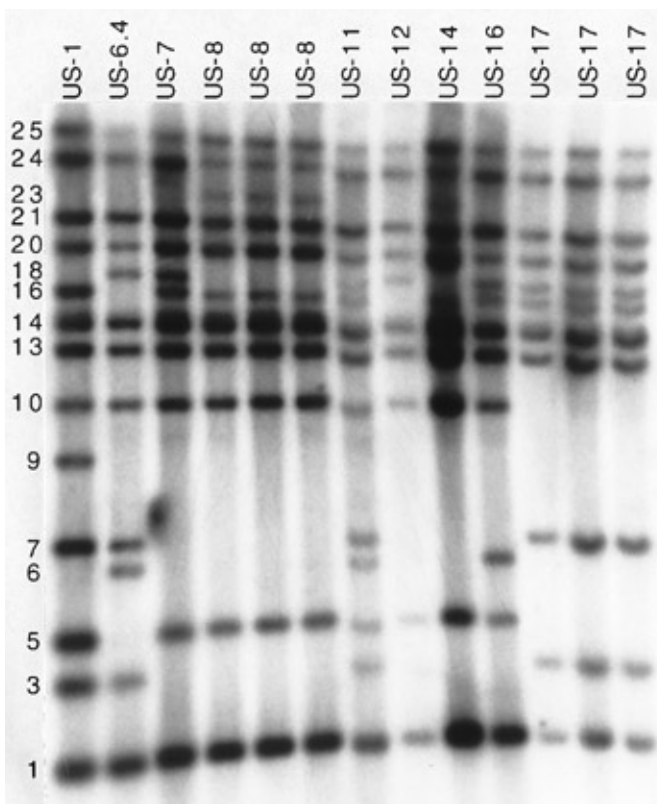


Fig. 2. RG57 fingerprint patterns of nine genotypes of *Phytophthora infestans* detected in the United States or Canada during 1994 through 1996. Genotype names are indicated above each lane. RG57 fingerprint band numbers are indicated on the left. US-14 is identical to US-8 and is probably a somatic variant. US-11, US-12, US-16, and US-17 are probably recombinants between US-1 or US-6 (A1 mating type) and US-7 or US-8 (A2 mating type). Locations represented by each genotype are as follows: US-1, Nebraska; US-6.4, Washington; US-8, US-12, and US-14, New York; US-16, Oregon; and US-17, Alabama, New Jersey, and New York.

Genotype identification: 1995. A similar pattern occurred during 1995 and 1996. Most isolates collected during 1995 were US-8, but US-7 isolates were found in Alaska, California, Florida, and New Jersey (Table 4). The US-11 genotype was isolated from tomato in California and New York (Table 4). US-1 was found in Michigan and Texas. During 1995 and 1996, multilocus genotype often was inferred from *Gpi* genotype alone, so the numbers of genotypes reported here may be underestimates. The genotypes of 12 isolates were confirmed by DNA fingerprinting including seven that were US-7 (from Alaska and California), one that was US-8 (from New York), and four that were US-11 (from California and New York).

Genotype identification: 1996. The vast majority of the isolates tested during 1996 were US-8, including isolates from Alabama, Maine, and 14 counties in New York (Table 5). US-1 was isolated from potato and tomato in Schoharie and Schenectady counties, NY, respectively (adjacent counties in eastern New York). Two unusual genotypes were found on tomato: US-16 in California and US-17 in Alabama, Florida, New Jersey, and New York (Table 5). DNA fingerprint analysis confirmed the genotype of one US-16 and seven US-17 isolates (representing all four states where this genotype was found).

Changes in genotype frequencies over time. To test whether the US-1 genotype was being replaced, frequencies of the four most commonly detected genotypes in the United States and Canada during 1979 to 1996 were calculated from data published previously (17,23) and this study. Analysis of the combined data set of 1,112 isolates revealed a decline in detection frequency of the US-1 genotype (Table 6). US-1 made up more than 60% of the isolates collected during 1979 to 1987, but declined to less than 10% during 1994 through 1996. US-6 showed a rapid increase in frequency during 1987 to 1991, followed by a rapid decrease to less than 2% of the isolates collected during 1994. It was not detected in our more limited samples during 1995 and 1996. US-7 peaked during 1993, declined during 1994, rebounded slightly during 1995, but was not detected during 1996. Most of the US-7

TABLE 4. Sampling information and genotype for 65 isolates of *Phytophthora infestans* collected in the United States during 1995

State	County	Month collected	No. of fields	No. of isolates	Genotype(s) ^a	
Isolates from potato			26	33		
Alaska	Matanuska-Susitna	October	1	1	US-7	
Arizona	Maricopa	March	1	1	US-8	
Florida	Hendry	January	1	1	US-8	
Idaho	Canyon	July	1	1	US-8	
Michigan	1	1	US-1	
New York	Ontario	September	1	1	US-8	
		October	1	1	US-8	
	Steuben	July	2	2	US-8	
		August	2	2	US-8	
	Wayne	October	1	1	US-8	
		September	1	1	US-8	
	Wyoming	July	1	3	US-8	
		August	2	2	US-8	
	North Carolina	Camden	June	6	6	US-8
			June	1	2	US-8
Pamlico		June	1	2	US-8	
Washington		June	1	3	US-8	
Texas		Frio	March	1	2	US-1, US-8
Isolates from tomato			7	32		
California	San Luis Obispo	October	1	4	US-7 (1), US-11 (3)	
		November	1	23	US-7	
Florida	Hillsborough	April	1	1	US-8	
		April	2	2	US-7	
New Jersey	Cumberland	July	1	1	US-7	
New York	Erie	October	1	1	US-11	

^a Determined from analysis of mating type, *glucose-6-phosphate isomerase* (*Gpi*) allozyme genotype, and RG57 fingerprint; for some isolates, from *Gpi* genotype alone.
^b Not known.

DISCUSSION

isolates from 1995 came from a single field in California. Thus, rapid changes in genotype frequencies were detected over time, even from year to year.

The locations in which each genotype was detected also varied from year to year. It was not always possible to use information from 1 year to predict where a genotype would occur the next year. This was particularly evident in New York (Fig. 3). US-8 was not detected in New York during 1993, but was common and widespread during 1994. US-7 was the most common genotype in New York during 1993, but was only detected in one county during 1994. After 1994, US-8 predominated everywhere.

Metalaxyl sensitivities. Metalaxyl tests confirmed the associations between sensitivity and genotype reported previously (25) for the US-1, US-7, and US-8 genotypes (Table 7). All US-1 isolates tested were sensitive, whereas all US-7 and US-8 isolates were resistant. In addition, eight US-17 isolates were tested, and all were highly resistant (Table 7). Two US-6 isolates from Texas were highly sensitive, as reported previously for that population (5). Floating leaf-disc assays gave better discrimination between resistance and sensitivity and had a smaller variance than amended-agar assays. None of the US-1 isolates tested grew or sporulated on leaf discs floating on 5- μ g/ml metalaxyl solution, whereas one US-1 isolate grew up to 12% of the control on 5- μ g/ml metalaxyl-amended agar (Table 7). Similarly, US-7 and US-8 isolates grew from 64 to 106% of the control on 5- μ g/ml metalaxyl-amended agar, but from 95 to 103% on floating leaf discs. One isolate, FL 318-2, was tested with both methods and grew at 71% on 5- μ g/ml metalaxyl-amended agar, but at 95% on the floating leaf-disc test.

Both isolates of *P. capsici* tested were highly sensitive to metalaxyl (Table 7), even though they came from a field that had been sprayed with Ridomil and the grower believed they probably were resistant.

TABLE 5. Sampling information and genotype for 52 isolates of *Phytophthora infestans* collected in the United States during 1996

State	County	Month collected	No. of fields	No. of isolates	Genotype ^a
Isolates from potato			28	36	
Alabama	Baldwin	April	1	2	US-8
Maine	Aroostook	July	1	1	US-8
New York	Clinton	August	4	4	US-8
	Delaware	October	1	6	US-8
	Franklin	August	1	1	US-8
	Genesee	October	1	1	US-8
	Monroe	August	1	1	US-8
		September	1	1	US-8
	Ontario	September	1	1	US-8
	Oswego	September	1	1	US-8
	Schoharie	July	1	3	US-1
	Seneca	August	1	1	US-8
	St. Lawrence	September	1	1	US-8
	Suffolk	July	2	2	US-8
	Tioga	August	4	4	US-8
	Tompkins	October	1	1	US-8
Wayne	August	2	2	US-8	
Wyoming	August	1	1	US-8	
	September	1	1	US-8	
	October	1	1	US-8	
Isolates from tomato			10	16	
Alabama	Blount	October	1	1	US-17
California	San Luis Obispo	September	1	4	US-16
Florida	Lee	December	1	4	US-17
New Jersey	Gloucester	September	1	1	US-17
New York	Chautauqua	October	1	1	US-17
	Clinton	September	1	1	US-8
	Ontario	September	1	1	US-8
	Orange	August	1	1	US-17
	Schenectady	August	1	1	US-1
	Tioga	September	1	1	US-8

^a For 36 isolates, multilocus genotype was predicted based on *glucose-6-phosphate isomerase* genotype alone.

The devastating late blight epidemics in the United States during 1994 through 1996 were caused primarily by the US-8 genotype of *P. infestans*. Based on our experiences through 1993, this was unexpected. US-7 was the most commonly detected genotype during 1993 and occurred throughout the country (23). It infected tomato in addition to potato (24), which we thought might give it a selective advantage. However, although US-7 caused some epidemics on tomato, primarily in the southeastern states during 1994, it was a relatively minor component of the population that year. This continued during 1995, and, by 1996, US-7 was not detected at all. In its place, a new genotype, US-17, began to appear on tomato.

By 1994, the old US-1 clonal lineage of *P. infestans* was almost totally replaced in the United States by new genotypes from Mexico. Although US-1 still occurs at a low frequency, it is no longer the major component of the U.S. population. The almost total replacement of US-1 occurred within 3 years after the new genotypes (US-7 and US-8) were first detected. Rapid replacement of US-1 by US-8 also has been documented locally in Wisconsin (31), the Columbia Basin of Oregon and Washington (33), and in central and eastern Canada (Z. K. Punja, *unpublished data*). Similar rapid replacements of US-1 have been documented in Europe (3, 9, 10, 13, 36–38), Asia (28), and South America (11). However, the genotypes causing the replacements were different in each location. So far, the *Gpi III* allele (characteristic of the US-7 and US-8 genotypes) has not been found outside North America (16, 18). Conversely, genotypes characteristic of the European, South American, and Asian migrations have not been detected within North America. Thus, there is still potential for additional migrations that could spread damaging genotypes to new locations.

A rather disturbing result is that it was not always possible to predict which genotypes would be present from 1 year to the next. US-8 was a relatively minor component of the 1993 collections, found in less than 10% of the isolates analyzed and only in those from Maine. However, it predominated during 1994 and made up 77% of all isolates tested. After 1994, US-8 occurred throughout the United States, and no more major changes were noticed.

Inability to predict genotypes from year to year was particularly evident in New York during 1993 and 1994. US-7 was the only genotype detected in New York during 1993 and was distributed throughout the state (23). However, during 1994, US-7 was found only in one home garden, and instead, US-8 was found throughout the state. It is probable that in situ survival is low, so seed tuber infections are much more important. In addition, US-8 appears to have much greater pathogenic fitness than US-7 on potato (27, 29, 30), which may have played a role in its rapid increase. We know that seed tubers planted in New York during 1994 were infected with US-8 (Table 2), and these must have provided the inoculum,

TABLE 6. Changes in frequency of the four most common genotypes of *Phytophthora infestans* detected in samples analyzed from the United States and Canada during 1979 to 1996^a

Genotype	Frequency of detection (%) during						
	1979–1987 ^b	1987–1991 ^c	1992 ^d	1993 ^e	1994 ^f	1995 ^g	1996 ^h
US-1	65	19	30	21	8	3	8
US-6	8	79	12	8	1
US-7	34	48	6	43	...
US-8	21	8	77	48	69

^a Based on a total sample of 1,112 isolates.

^b *N* = 26. Data for 1979 through the first half of 1987 are from Goodwin et al. (17).

^c *N* = 146. Data for the second half of 1987 through 1991 are from Goodwin et al. (17).

^d *N* = 203. Data from Goodwin et al. (23).

^e *N* = 290. Data from Goodwin et al. (23) and this study.

^f *N* = 330. This study.

^g *N* = 65. This study.

^h *N* = 52. This study.

because there was no evidence for US-8 from the previous year. This again underscores the importance of planting disease-free seed.

Distribution of *P. infestans* in seed tubers also was demonstrated in Florida. Seed tubers shipped into Florida from Maine and North Dakota during January 1993 were infected with US-1, and US-1 was the only genotype isolated from the field later that year (23). Seed shipped into Florida from Maine during late 1993 contained US-8, which was the only genotype identified in southern Florida during 1994. Thus, genotypes in seed gave a much better prediction of genotypes in the field from 1993 to 1994 than did the geno-

types that were present in the field the previous year. The only exception was US-1, which evidently was prevented from causing epidemics from seed tubers by early applications of metalaxyl.

These analyses confirmed that metalaxyl sensitivity and genotype still are correlated at least for the US-1, US-7, and US-8 genotypes. All US-1 isolates tested so far from the United States have been sensitive (25,31,33), and this continued during 1994. Similarly, all US-7 and US-8 isolates still are resistant. The two US-6 isolates tested from Texas were highly sensitive. This confirms the polymorphism for metalaxyl sensitivity within the US-6

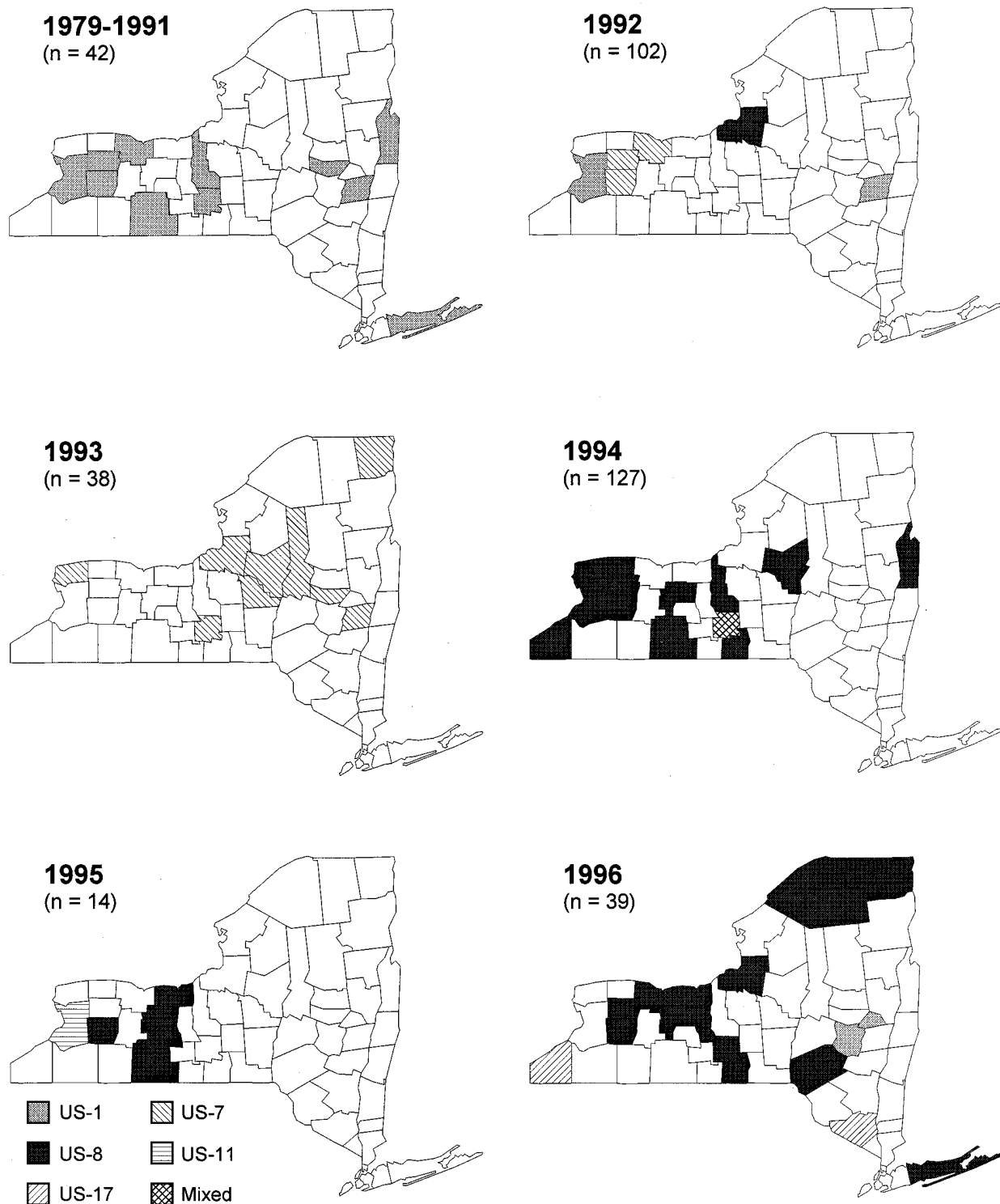


Fig. 3. Changes in collection locations of genotypes of *Phytophthora infestans* in New York by county among 362 isolates analyzed during 1979 through 1996. Sample sizes are in parentheses. Genotypes are as indicated in Table 1. "Mixed" refers to more than one genotype in the same county during 1 year. This only occurred during 1994, when the US-7, US-8, US-12, and US-13 genotypes were found in Tompkins County.

clonal lineage noted previously (25). US-17 appears to be a new resistant genotype, because all eight isolates tested from four different states were highly resistant.

The floating leaf-disc assay eliminated some of the variance seen in the amended-agar assay and is recommended for all future studies. The floating leaf-disc results also may reflect results in the field more accurately. US-8 grows significantly less than US-7 on metalaxyl-amended agar (25). However, both US-7 and US-8 grow at virtually 100% of the control on floating leaf-disc assays and apparently are unaffected by metalaxyl in the field (25). Therefore, the floating leaf-disc results may provide a better prediction of response to metalaxyl in growers' fields.

During 1994 to 1996 in the United States, there still was only limited evidence for sexual recombination of *P. infestans*, even though both mating types have been widely distributed since 1992 (5,23). This probably is because both mating types seldom were

found together in the same field. Goodwin et al. (23) found both mating types together only in one field in Florida among more than 100 fields from which multiple isolates were obtained during 1992 and 1993. Deahl et al. (5) analyzed 184 U.S. isolates collected during 1991 through 1993 and found both mating types together only occasionally in Florida (probably from the same field sampled by Goodwin et al. [23]) and Texas. The Texas isolates were analyzed in the current study and were all US-6 or US-7. Thus, in Florida and Texas, both mating types probably were introduced too recently for the products of sexual recombination to become detectable. Both mating types have been found together more often in British Columbia (2), where the first probable evidence for sexual recombination was found during 1992 (23). Both mating types also were found together in seven fields in New Brunswick during 1994 (2), but that is probably when A2 was first introduced, so opportunities for sexual recombination were minimal. Increased sampling in Florida, Texas, and New Brunswick is needed to determine whether coexistence of both mating types led to additional instances of sexual recombination.

Opportunities for sexual recombination remained limited during 1994 through 1996. In our samples, both mating types were found together only in one field in New York, one county in California, and one field in Texas among 94 U.S. fields with multiple samples. Furthermore, the mating types usually were separated geographically, even when they occurred in the same state. For example, during 1994 in Michigan, A1 isolates were found only in Presque Isle County at the northern tip of the southern peninsula, while A2 isolates were limited to the central part of the state. Similarly, during 1994 in North Carolina, A1 isolates were found only in one eastern county from which no A2 isolates were obtained. During 1996 in New York, A1 isolates were found only in one western and three eastern counties, while US-8 (A2) was found in 14 other counties throughout the state. Although the limited sampling obviously underestimates the true range of both mating types, it does indicate that, in the United States, opportunities for sexual recombination of *P. infestans* remain limited 4 years after A2 became widespread.

Opportunities for sexual reproduction also probably were limited by the high sensitivity of the US-1 clonal lineage to metalaxyl. US-1 was detected in shipments of seed tubers from North Dakota that were planted subsequently in New York (and presumably other states). Many growers who planted infected seed sprayed with metalaxyl in an attempt to eliminate US-1. Evidently, this strategy was successful. All US-1 isolates tested were highly sensitive, and this probably explains the absence of US-1 from metalaxyl-treated fields. Many of these fields did contain US-8, probably introduced by wind-borne inoculum from nearby fields, so sexual reproduction could have occurred had US-1 been present. Thus, use of metalaxyl may have provided an unseen benefit even when it did not halt an epidemic.

However, some potential recombinant genotypes were rarely detected during 1994 through 1996. The most common of these was the US-11 genotype, which was A1 mating type, *Gpi 100/100/111*, *Pep 100/100*, and was found in California, New York, and Washington. This genotype also was reported in the Columbia Basin of Oregon and Washington during 1993 (33). It could have originated by recombination between US-6 (A1) and US-7 or US-8 (A2). The presence of DNA fingerprint band 6 in US-11 excludes US-1 as a likely A1 parent, because this band also is absent from both US-7 and US-8. Both US-8 and US-11 apparently are trisomic (have three copies) for the chromosome containing the *Gpi* locus, and crosses involving US-8 yield a high frequency of trisomic progeny in the laboratory (22). Thus, recombination between US-6 and US-8 may be the most likely origin of the *Gpi 100/100/111* genotype of US-11.

Four other potential recombinant genotypes were found in fields of tomato. US-12 was found in Maclean, NY; US-13 in Dryden, NY; US-16 in California; and US-17 in Alabama, Florida, New Jersey, and New York (Table 8). US-13 could have been generated by hybridization between US-1 or US-6 (A1) and US-7

TABLE 7. Metalaxyl sensitivities of isolates of *Phytophthora infestans* and *P. capsici* by genotype

Genotype	Isolate	Percent of control ^a		Result	
		5 µg/ml	100 µg/ml		
Amended-agar assays					
US-1	151 MI	1	0	Sensitive	
	NC-364	12	9	Sensitive	
	NC-367	11	8	Sensitive	
US-6	TX 4-1 ^b	3	3	Sensitive	
	TX 4-3 ^b	3	3	Sensitive	
US-7	TX 3-4 ^b	104	49	Resistant	
	TX 3-8 ^b	94	65	Resistant	
	FL 318-2	71	61	Resistant	
	FL 319-5	91	54	Resistant	
	SC-1	106	96	Resistant	
	SC-4	103	105	Resistant	
US-8	GA-320	83	56	Resistant	
	NC-P-4	105	60	Resistant	
	NC-P-5	83	41	Resistant	
	NC-P-6	75	58	Resistant	
	NC-Pamlico-1	83	38	Resistant	
	146 Erie PA	64	33	Resistant	
US-11	US94-0478	... ^c	63	Resistant	
US-12	US94-0494	...	60	Resistant	
US-13	Fleming, fruit 1	...	77	Resistant	
US-14	US94-0502	...	53	Resistant	
US-16	US94-0298	...	75	Resistant	
US-17	96-444	...	100	Resistant	
	96-505	...	107	Resistant	
	96-543	...	103	Resistant	
	96-550	...	105	Resistant	
	US97-0001	...	105	Resistant	
	US97-0002	...	105	Resistant	
	US97-0003	...	105	Resistant	
	US97-0004	...	105	Resistant	
	Floating leaf-disc assays (5 µg/ml only)				
	US-1	#309 ND seed	0	...	Sensitive
#310 ND seed		0	...	Sensitive	
#313 ND seed		0	...	Sensitive	
#314 ND seed		0	...	Sensitive	
#315 ND seed		0	...	Sensitive	
#317 ND seed		0	...	Sensitive	
US-7	FL 318-2 ^d	95	...	Resistant	
US-8	14263-2 ME	101	...	Resistant	
	14263-3 ME	98	...	Resistant	
	14263-4 ME	103	...	Resistant	
	#311 Maine seed	99	...	Resistant	
<i>Phytophthora capsici</i> agar assays					
CAP-1	CA 2	3	1	Sensitive	
CAP-1	CA 3	1	0	Sensitive	

^a Growth of an isolate on media containing 5 or 100 µg of metalaxyl per ml relative to the unamended control. The percent leaf disc area covered by sporulation relative to the unamended control was measured for the floating leaf-disc assay.

^b Texas isolates collected during 1993, but analyzed during 1994.

^c Not tested.

^d Also tested with the amended-agar assay.

or US-8 (A2). This also could have been the origin of the CA-8 genotype from potato in British Columbia. For US-17, the parents were most likely US-6 and US-8, to have both the *Gpi 122* allele and DNA fingerprint band 18. US-16 could have originated by a cross between US-6 and US-7 or US-8 (not US-1). US-12 could not

have originated by hybridization between US-1 and US-8, as neither has DNA fingerprint band 18. However, it could have originated by a cross between US-1 and US-7, or US-6 by US-7 or US-8. In fact, all of these genotypes could have arisen by hybridization between US-6 and US-8. Isolates with the same mating type and

TABLE 8. Summary of genotypes of *Phytophthora infestans* identified in the United States and Canada during 1979 to 1996

Genotype ^a	Mating type	Allozyme genotype		RG57 fingerprint ^d	Metalaxyl sensitivity ^e	Comments
		<i>Gpi</i> ^b	<i>Pep</i> ^c			
US-1	A1	86/100	92/100	1011101011001101000110011	S	Old established genotype possibly present since the 1840s.
US-1.1	A1	86/100	100/100	1011101011001101000110011	...	One isolate from New York during 1979, and one from New Brunswick during 1980.
US-1.2	A1	86/100	92/100	1011101010001101000110011	...	Two isolates from Wisconsin during 1982, and one isolate during 1980 (location unknown).
US-1.3	A1	86/100	92/100	1011101001001101000110011	...	One isolate from Wisconsin during 1982.
US-1.4	A1	86/100	100/100	1011101010001101000110011	...	One isolate from North Carolina during 1985.
US-1.5	A1	86/100	92/100	1011101011001101010110011	...	One isolate from Prince Edward Island during 1992.
US-1.6	A1	86/100	92/100	1011101011001101000111011	...	One isolate from Peru during 1984.
US-1.7	A1	100/100	92/100	1011101011001101000110011	...	One isolate from Brazil during 1987.
US-1.8	A1	86/100	92/100	1011100011001101000110011	...	One isolate from Poland during 1988.
US-2	A1	86/100	92/100	1011101001001101011110011	...	One isolate, year and location of isolation not known.
US-3	A1	86/100	92/100	101110000001101000110011	...	One isolate from 1983, location unknown.
US-4	A1	100/100	92/92	1011101001001101100110011	...	One isolate from California during 1980.
US-5	A1	100/100	92/100	1011101001001101011110011	S	Two isolates from Maine during 1987, and one isolate during 1987, location unknown. Probably the same clonal lineage as US-2.
US-6	A1	100/100	92/100	1011111001001100010110011	R or S	Possibly introduced into California during 1979; since found throughout the United States and western Canada. Mostly on tomato, potato occasionally.
US-6.1	A1	100/100	92/92	1011111001001100010110011	...	One isolate from California during 1987.
US-6.2	A1	100/100	92/100	1011101001001100010110011	I	One isolate from western Washington during 1990.
US-6.3	A1	100/100	92/100	1011111001011100010110011	R	One isolate from British Columbia during 1991.
US-6.4	A1	100/100	100/100	1011011001001100010110011	R	One isolate from British Columbia during 1991.
US-6.5	A1	100/100	92/100	1011111001001100010010011	R	Single isolates from eastern Washington and British Columbia during 1991 and 1993, respectively.
US-7	A2	100/111	100/100	1001100001001101010110011	R	Recent immigrant from northwestern Mexico found throughout the United States and western Canada. Mostly on tomato, potato occasionally.
US-8	A2	100/111/122	100/100	1001100001001101000110111	R	Recent immigrant from northwestern Mexico now found throughout the United States and western Canada. Mostly on potato, tomato occasionally.
US-9	A1	100/100	83/100	...	R	One isolate from Idaho during 1993, on greenhouse tomato. Possibly introduced from Europe.
US-10	A2	111/122	100/100	...	S	One isolate from Wisconsin during 1993. Probably a sexual recombinant or recent immigrant from northwestern Mexico.
US-11	A1	100/100/111	100/100	1010111001001101010110011	R	Probable sexual recombinant; has two copies of <i>Gpi 100</i> allele. Found in western Washington during 1994, and California and New York during 1995.
US-12	A1	100/111	92/100	1000100001001100010110011	R	Probable sexual recombinant. Found in one field of tomato in Maclean, NY, during 1994.
US-13	A2	100/100	100/100	...	R	Possible sexual recombinant; also could have been imported from northwestern Mexico. Found in one field of tomato in Dryden, NY, during 1994.
US-14	A2	100/122	100/100	1000100001001101000110111	R	Probable clonal derivative of US-8 (by loss of <i>Gpi 111</i> allele). Found in one potato field in Steuben County, NY, during 1994. Also could be called US-8.1.
US-15	A2	100/100	92/100	Three isolates from tomato in Pennsylvania during 1994 (S. Kim, <i>personal communication</i>).
US-16	A1	100/111	100/100	1000110001001101010110011	R	Probable sexual recombinant between US-6 and US-7 or US-8. Isolated in Oregon during 1994, and from tomato in California during 1996.
US-17	A1	100/122	100/100	1010001000001101010110011	R	Probable sexual recombinant between US-6 and US-8. Isolated from tomato in Alabama, Florida, New Jersey, and New York during 1996.
CDA-1 ^f	A1	86/100	92/100	1111101011001101001110011	...	One isolate from New Brunswick during 1980. The fingerprint for this isolate also contains the rare band 11a.
CDA-2 ^f	A1	100/100	100/100	1011001000001100001110011	S	One isolate from British Columbia during 1991.
CA-2.1	A1	100/100	100/100	1011001000001100001111011	...	One isolate from British Columbia during 1991. Probably the same clonal lineage as CDA-2.
CDA-3 ^f	A2	86/100	100/100	1111101001001001100110011	...	Two isolates from British Columbia during 1991.
BC-1 ^f	A2	100/111	100/100	1000000001001101000110011	I	Possible sexual recombinant or recent immigrant. Isolated in British Columbia during 1992 and 1993.
BC-2 ^f	A2	100/100	100/100	1000110000001101000110011	S	Probable sexual recombinant. One isolate from British Columbia during 1992.
BC-3 ^f	A2	100/100	100/100	1010001001001100010110011	S	Probable sexual recombinant. One isolate from British Columbia during 1992.
BC-4 ^f	A2	100/100	100/100	1001000000001100010110011	S	Probable sexual recombinant. One isolate from British Columbia during 1992.
CA-8	A1	100/111/111	100/100	Probable sexual recombinant. Two isolates from British Columbia during 1993.

^a Includes variants of the common clonal lineages detected outside the United States or Canada. Variants within a lineage are indicated by appending a period and a number after the genotype name and are numbered sequentially (17). Thus, US-1.3 is the third variant identified within the US-1 clonal lineage. These variants are identical to the "parent" clone except for one or two changes at allozyme or DNA fingerprint loci. They are assumed to have arisen from asexual (somatic) variation within lineages.

^b *Glucose-6-phosphate isomerase*.

^c *Peptidase*.

^d Presence (1) or absence (0) of RG57 fingerprint bands 1 to 25 (19) are indicated from left to right. DNA fingerprint band 4 is not reproducible and should not be used for genotype identification. ... = Not determined.

^e S = sensitive, R = resistant, I = intermediate, and ... = not determined.

^f Genotypes CDA-1, CDA-2, CDA-3, BC-1, BC-2, BC-3, and BC-4 were renamed recently according to a standard nomenclature (12). The new names are CA-1 through CA-7, respectively.

Gpi genotype as US-11 through US-14 also were reported in the Pacific Northwest during 1993 (33), along with two additional genotypes not detected in our samples. Additional sampling probably will reveal more genotypes. Even so, potential recombinant genotypes made up only a small proportion of the total sample, compared with some newly sexual populations in Europe in which nearly 100% of the isolates tested were recombinant (10,38).

One other new genotype, US-14, was from a potato field in southern New York that also contained US-8. US-14 had the same DNA fingerprint as US-8 and almost certainly arose as a somatic variant of US-8 by loss of the *Gpi III* allele.

In addition to sexual recombination, another potential source of new genotypes is continued migration from Mexico or other areas. Isolates identical to US-10 (from Wisconsin during 1993 [23]), US-13, and US-14 for mating type and dilocus allozyme genotype were found in northwestern Mexico during 1993 (S. B. Goodwin and W. E. Fry, unpublished data), and together accounted for almost 57% of the 109 isolates sampled that year. However, US-11 and US-12 so far have not been found in Mexico (22; S. B. Goodwin and W. E. Fry, unpublished data). Thus, although migration remains a possibility for some of the new genotypes, it seems remote for others. The most likely explanation is that at least some of the rare, new genotypes originated by sexual recombination within the United States. This also could be the origin of the US-15 genotype identified by S. H. Kim (personal communication) in Pennsylvania during 1994 (Table 8) and of the two additional genotypes identified recently in the Pacific Northwest (33). Other genotypes that probably originated by in situ sexual recombination were identified in British Columbia during 1992 (23) and 1993 (Table 8). Future monitoring is required to determine whether sexually reproducing populations of *P. infestans* now have become established in the United States and Canada, and how they might affect late blight epidemiology.

Isolates of two other species of *Phytophthora* that are found commonly on potato or tomato were easy to distinguish from *P. infestans* and from each other on the basis of *Gpi* genotype alone. All four isolates of *P. capsici* tested had the same genotype at the *Gpi* (106/106) and *Pep* (94/94) loci. This genotype probably is the same as genotype CAP-1 of Oudemans and Coffey (35), and we have retained their appellation. All 21 isolates of *P. erythroseptica* were *Gpi* 91/91; we could not resolve *Pep* bands for this species. Because each species had a distinct genotype, cellulose-acetate electrophoresis provides a fast, easy method for identification of *P. erythroseptica* and *P. capsici*, as well as genotypes of *P. infestans* (21). It is interesting that the two isolates of *P. capsici* tested were highly sensitive to metalaxyl, because they came from a field that had been sprayed and the grower thought they might be resistant. Thus, in *P. capsici*, loss of effectiveness of metalaxyl may be due to other factors besides resistance.

The mechanisms for the rapid spread of the US-8 genotype of *P. infestans* in the United States and Canada during 1994 are partially known. US-8 evidently became established in seed-producing areas of Maine and possibly other states during 1993 (23) and probably was distributed in infected seed during 1994. When both genotypes were in infected seed, sometimes in adjacent fields, the resistance of US-8 to metalaxyl probably explains its advantage compared with US-1. However, the most likely explanation may be that US-8 simply has higher pathogenic fitness on potato (both foliage and tubers) than the other common genotypes (27,29). US-8 also infected hairy nightshade (as does US-1 [23]), which may provide a bridge from field to field. Isolates of *P. infestans* now have been obtained from hairy nightshade (*S. sarrachoides*) in California (39), Wisconsin (23), Washington (7), and New York (this study). Clearly, this weed should be eliminated as much as possible from potato and tomato production areas.

The new genotypes obviously are much more difficult to control; a recent study showed that more frequent applications of fungicide will be required to achieve the same level of disease suppres-

sion for US-8 compared with US-1 (27). Models for predicting late blight epidemics (e.g., Blitecast) should be modified, or at least verified, against the new genotypes. The role and effect of oospore inoculum on late blight epidemics is not known. Epidemics could occur more often, earlier in the season, and with greater severity. Continued monitoring of the pathogen may be warranted for advance notice of future changes. The next few years could be very difficult until improved methods of disease management are developed to combat the new genotypes.

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