# nature genetics

# Genetic loci influencing kidney function and chronic kidney disease

John C Chambers<sup>1,2,66\*</sup>, Weihua Zhang<sup>1,2,66</sup>, Graham M Lord<sup>3–5</sup>, Pim van der Harst<sup>6</sup>, Debbie A Lawlor<sup>7</sup>, Joban S Sehmi<sup>2,8</sup>, Daniel P Gale<sup>9</sup>, Mark N Wass<sup>10</sup>, Kourosh R Ahmadi<sup>11</sup>, Stephan J L Bakker<sup>6</sup>, Jacqui Beckmann<sup>12</sup>, Henk J G Bilo<sup>6</sup>, Murielle Bochud<sup>13</sup>, Morris J Brown<sup>14</sup>, Mark J Caulfield<sup>15</sup>, John M C Connell<sup>16</sup>, H Terence Cook<sup>17</sup>, Ioana Cotlarciuc<sup>11</sup>, George Davey Smith<sup>7</sup>, Ranil de Silva<sup>2,8</sup>, Guohong Deng<sup>18</sup>, Olivier Devuyst<sup>19</sup>, Lambert D Dikkeschei<sup>20</sup>, Nada Dimkovic<sup>21</sup>, Mark Dockrell<sup>22</sup>, Anna Dominiczak<sup>16</sup>, Shah Ebrahim<sup>23</sup>, Thomas Eggermann<sup>24</sup>, Martin Farrall<sup>25</sup>, Luigi Ferrucci<sup>26</sup>, Jurgen Floege<sup>27</sup>, Nita G Forouhi<sup>28</sup>, Ron T Gansevoort<sup>29</sup>, Xijin Han<sup>30</sup>, Bo Hedblad<sup>31</sup>, Jaap J Homan van der Heide<sup>32</sup>, Bouke G Hepkema<sup>33</sup>, Maria Hernandez-Fuentes<sup>3–5</sup>, Elina Hypponen<sup>34</sup>, Toby Johnson<sup>35</sup>, Paul E de Jong<sup>36</sup>, Nanne Kleefstra<sup>36</sup>, Vasiliki Lagou<sup>37</sup>, Marta Lapsley<sup>22</sup>, Yun Li<sup>30</sup>, Ruth J F Loos<sup>28</sup>, Jian'an Luan<sup>28</sup>, Karin Luttropp<sup>38</sup>, Céline Maréchal<sup>19</sup>, Olle Melander<sup>31</sup>, Patricia B Munroe<sup>15</sup>, Louise Nordfors<sup>38</sup>, Afshin Parsa<sup>39</sup>, Leena Peltonen<sup>40–42</sup>, Brenda W Penninx<sup>43–45</sup>, Esperanza Perucha<sup>3–5</sup>, Anneli Pouta<sup>42,46</sup>, Inga Prokopenko<sup>47–49</sup>, Paul J Roderick<sup>50</sup>, Aimo Ruokonen<sup>51</sup>, Nilesh J Samani<sup>52</sup>, Serena Sanna<sup>53</sup>, Martin Schalling<sup>54</sup>, David Schlessinger<sup>55</sup>, Georg Schlieper<sup>27</sup>, Marc A J Seelen<sup>29</sup>, Alan R Shuldiner<sup>56</sup>, Marketa Sjögren<sup>31</sup>, Johannes H Smit<sup>43–45</sup>, Harold Snieder<sup>37</sup>, Nicole Soranzo<sup>11</sup>, Timothy D Spector<sup>11</sup>, Peter Stenvinkel<sup>57</sup>, Michael J E Sternberg<sup>10</sup>, Ramasamyiyer Swaminathan<sup>58</sup>, Toshiko Tanaka<sup>26</sup>, Lielith J Ubink-Veltmaat<sup>59</sup>, Manuela Uda<sup>53</sup>, Peter Vollenweider<sup>60</sup>, Chris Wallace<sup>61</sup>, Dawn Waterworth<sup>62</sup>, Klaus Zerres<sup>24</sup>, Gerard Waeber<sup>60</sup>, Nicholas J Wareham<sup>28</sup>, Patrick H Maxwell<sup>9</sup>, Mark I McCarthy<sup>47,48,49</sup>, Marjo-Riitta Jarvelin<sup>1,42,46,63</sup>, Vincent Mooser<sup>62</sup>, Goncalo R Abecasis<sup>30</sup>, Liz Lightstone<sup>64,66</sup>, James Scott<sup>8,66</sup>, Gerjan Navis<sup>6,66</sup>, Paul Elliott<sup>1,65,66</sup> & Jaspal S Kooner<sup>2,8,66</sup>

Using genome-wide association, we identify common variants at 2p12-p13, 6q26, 17q23 and 19q13 associated with serum creatinine, a marker of kidney function ( $P = 10^{-10}$  to 10<sup>-15</sup>). Of these, rs10206899 (near NAT8, 2p12-p13) and rs4805834 (near SLC7A9, 19q13) were also associated with chronic kidney disease ( $P = 5.0 \times 10^{-5}$  and  $P = 3.6 \times 10^{-4}$ , respectively). Our findings provide insight into metabolic, solute and drug-transport pathways underlying susceptibility to chronic kidney disease.

In North America and Europe, chronic kidney disease (CKD) affects ~11% of adults. CKD is associated with high morbidity and, in the advanced

stage, requires life-support treatment by renal dialysis or transplantation<sup>1</sup>. CKD is also a major risk factor for myocardial infarction and stroke.

CKD is a multifactorial disorder with an important genetic component<sup>2</sup>. A number of monogenic disorders underlying CKD have been identified, though these account for only a small proportion of the total burden of kidney disease. Recent studies have identified common variants at the UMOD, SHROOM3, GATM and MYH9 loci that are associated with kidney function in European and African-American populations<sup>2,3</sup>. We carried out both a genome-wide association study and a replication study to identify loci associated with serum creatinine levels. Although creatinine levels may be partially influenced by nonrenal factors, including diet and generation from muscle metabolism, serum creatinine is a validated measure of glomerular filtration rate<sup>4</sup>.

Genome-wide association was done in 23,812 participants of European descent from nine studies; characteristics of participants and genotyping arrays used are summarized in the Supplementary Methods and in Supplementary Table 1. Creatinine levels were transformed by log<sub>10</sub> to achieve approximate normality, and SNP associations were tested by linear regression using an additive genetic model adjusted for age and sex. Principal component scores were included as ancestry covariates in regression analyses, and test statistics were corrected for the genomic control inflation factor to adjust for population substructure (Supplementary Methods)<sup>5</sup>. Analyses were performed separately in each cohort and were followed by meta-analysis using z scores weighted by the square root of the sample size. Quantilequantile plots showed good adherence to null expectations ( $\lambda = 1.024$ ; Supplementary Fig. 1). Our genome-wide association study had 80% power to detect SNPs associated with 0.14% of population variation in creatinine levels at  $P < 5 \times 10^{-7}$ .

There were 109 SNPs associated with creatinine at  $P < 5 \times 10^{-7}$ , which were distributed over five loci (2p12-p13, 4q21, 6q26, 17q23 and 19q13; Fig. 1 and Supplementary Fig. 2). At four of these loci (2p12-p13, 6q26, 17q23 and 19q13), common variants have not previously been reported to be associated with kidney function or CKD; at each locus, we selected the most strongly associated SNP for replication testing with creatinine in a further sample of 16,626 Europeans (Supplementary Methods and Supplementary Table 2). All four SNPs showed strong replication with creatinine ( $P = 2.4 \times$  $10^{-3}$  to 7.0 ×  $10^{-9}$ ; Table 1 and Supplementary Table 3). At 4q21, the most strongly associated SNP was rs9992101 ( $P = 5.9 \times 10^{-9}$ ), which is located in SHROOM3 and is in high linkage disequilibrium (LD) with rs17319721 ( $r^2 = 0.78$ , HapMap CEU population), a SNP previously reported to be associated with glomerular filtration rate<sup>2</sup>.

Next we tested the four top-ranking SNPs for association with estimated glomerular filtration rate (eGFR) and cystatin c (two additional measures of kidney function<sup>4</sup>) and with CKD among the participants from the replication sample (Supplementary Methods). rs10206899 (2p12-p13) and rs4805834 (19q13) were associated with eGFR, cystatin c

Received 10 September 2009; accepted 15 March 2010; published online 11 April 2010; doi:10.1038/ng.566

<sup>\*</sup>A full list of author affiliations appears at the end of the paper.

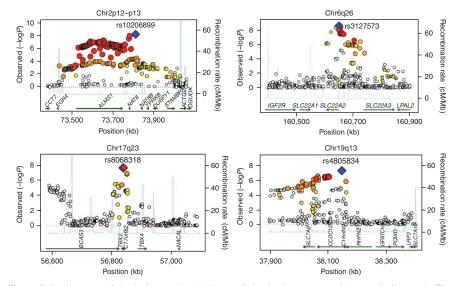


Figure 1 Architecture of the loci associated with creatinine in the genome-wide association study. The most significant SNP in each region is plotted in blue. LD is based on the HapMap CEU sample and is color-coded as red ( $r^2$  to top SNP, 0.8–1.0), orange (0.5–0.8), yellow (0.2–0.5) and white (<0.2).

and CKD (**Table 1** and **Supplementary Table 4**). In contrast, rs3127573 (6q26) and rs8068318 (17q23) were associated with eGFR but not with cystatin c or CKD. None of the four SNPs were associated with weight, hypertension, diabetes or other clinical parameters known to influence creatinine levels (**Supplementary Table 5**), and the relationships of these SNPs with creatinine were similar among people with and without diabetes or hypertension (**Supplementary Table 6**).

rs10206899 (2p12–p13), which is associated with creatinine, eGFR, cystatin c and CKD, is near several genes, including *NAT8*, *NAT8B*, *ALMS1*, *DUSP11* and *TPRKB* (**Fig. 1**). *NAT8* is a biologically compelling candidate for the observed association. NAT8 is a member of the GCN5-related N-acetyltransferase (GNAT) superfamily, a group of enzymes that catalyze transfer of an acetyl group from acetyl-coenzyme A to a wide range of acceptor substrates<sup>6</sup>. *NAT8* is strongly and almost exclusively expressed in kidney (**Supplementary Fig. 3**), in particular by tubular cells of the renal cortex (**Supplementary Fig. 4** and **5**). Acetylation is a key metabolic pathway for the detoxification of nephrotoxic substances such as aminoglycosides, inhalational anesthetics and environmental toxins, including industrial solvents such as trichloroethylene<sup>7,8</sup>. rs10206899 is in high LD ( $r^2 = 1.0$ ) with the only common nonsynonymous SNP in *NAT8*, rs15358 (causing a A595G change). rs15358 produces a nonconservative amino acid change (F143S) within the acetyl-coenzyme

*TPRKB* encodes the p53-related proteinkinase–binding protein and is of unknown function. Neither is strongly or preferentially expressed in kidney.

A binding site, an effect predicted to influ-

ence acetylation by NAT8 (Supplementary

Fig. 6). rs15358 was also closely associated with

creatinine levels in the genome-wide study

 $(P = 1.8 \times 10^{-8})$ . Our findings raise the possi-

bility that common genetic variation in NAT8

may influence acetylation pathways, disturbances of which are known to be associated

with drug- and toxin-induced kidney injury.

NAT8B is highly homologous to NAT8

and also contains an acetyltransferase

domain but is only expressed at low levels in

kidney (Supplementary Fig. 3). Mutations

in ALMS1 are responsible for Alström

Syndrome, a rare autosomal-recessive multi-

system disorder characterized by progressive kidney and hepatic failure, obesity and

insulin resistance, and blindness and hear-

ing loss<sup>9</sup>. Although DUSP11 and TPRKB

are also near rs10206899, neither has been

implicated in kidney function. DUSP11

is a dual-specificity protein phosphatase;

rs4805834 (19q13) is near *SLC7A9*, a cationic amino acid transporter highly expressed in kidney tubular cells (**Supplementary Fig. 3**)<sup>10</sup>. *SLC7A9* is a strong candidate for the association of rs4805834 with creatinine, eGFR, cystatin c and CKD; mutations in *SLC7A9* cause cystinuria and nephrolithiasis and are associated with increased risk of CKD<sup>10</sup>. rs4805834 is also near *CCDC123* and *C19orf40*. The latter (also known as *FAAP24*) has been identified as a component of the Fanconi anemia core complex, which plays a crucial role in DNA damage response<sup>11</sup> but has no reported relationship to kidney function. The function of *CCDC123* is not known.

rs3127573 (6q26) and rs8068318 (17q23) were associated with creatinine and eGFR. rs3127573 is near *SLC22A2*, an organic cation transporter strongly and preferentially expressed in kidney (**Supplementary Fig. 3**) that contributes to secretion of creatinine and other substrates by renal tubular epithelial cells<sup>12</sup>. Common variants at this locus are reported to influence kidney injury caused by nephrotoxic drugs such as cisplatin<sup>13</sup>. rs8068318 is located in *TBX2*, which encodes a member of the highly conserved T-box family of transcription factors<sup>14</sup>. Mouse *Tbx2<sup>-/-</sup>* mutants have a range of morphological defects, including limb deformities and cardiac anomalies, but a renal phenotype has not previously been described for them<sup>14</sup>. *Tbx2* is widely expressed in many tissues, including developing and adult

## Table 1 Association results for top-ranking SNPs in the genome-wide association and replication study

SNP	Locus	Alleles	MAF	Creatinine								Chronic kidney disease (replication sample)	
				GWA sample			Replication sample			Combined			
				п	Effect size (95% CI)	Р	п	Effect size (95% CI)	Р	Effect size (95% CI)	Р	Odds ratio (95% CI)	Р
rs10206899	2p12-p13	A/G	0.22	23,812	-0.9 (-1.2 to -0.6)	5.9 × 10 <sup>-9</sup>	16,167	-1.0 (-1.4 to -0.7)	7.0 × 10 <sup>-9</sup>	-1.0 (-1.2 to -0.7)	$1.2 \times 10^{-15}$	0.85 (0.79 to 0.92)	$5.0 \times 10^{-5}$
rs3127573	6q26	A/G	0.13	21,857	1.4 (1.0 to 1.8)	$5.0 \times 10^{-9}$	16,427	0.7 (0.2 to 1.1)	$2.4 \times 10^{-3}$	1.1 (0.8 to 1.4)	$6.5 \times 10^{-10}$	1.07 (0.97 to 1.17)	0.17
rs8068318	17q23	A/G	0.27	23,812	0.9 (0.6 to 1.2)	2.2 × 10 <sup>-8</sup>	16,350	0.6 (0.2 to 0.9)	$6.1 \times 10^{-4}$	0.8 (0.6 to 1.0)	$3.4 \times 10^{-10}$	1.05 (0.98 to 1.13)	0.16
rs4805834	19q13	G/A	0.13	23,812	-1.1 (-1.5 to -0.7)	$5.3 \times 10^{-8}$	16,241	–0.9 (–1.3 to –0.5)	$4.7 \times 10^{-5}$	-1.0 (-1.3 to -0.7)	$4.5 \times 10^{-11}$	0.84 (0.76 to 0.92)	$3.6 \times 10^{-4}$

Alleles, reference allele/minor allele; MAF, minor allele frequency; GWA, genome-wide association; effect size, % change in serum creatinine (95% Cl) or odds ratio for CKD (95% Cl) per copy of minor allele under an additive genetic model and adjusted for (i) age, gender and principal component scores in the genome-wide association study and (ii) age and gender in the replication study. Effect sizes were estimated by meta-analysis of cohort-specific beta estimates using the inverse variance method and a fixed effects model.

kidneys<sup>15</sup>, but the function of Tbx2 in the kidney is not known. rs8068318 is also near BCAS3 and hypothetical gene C17orf82. BCAS3 may be involved in angiogenesis, but is not known to be involved in kidney function.

In addition to SHROOM3, we also replicated previously reported associations<sup>2</sup> of rs12917707 in UMOD ( $P = 1.7 \times 10^{-5}$ ) and rs2467853 in GATM ( $P = 6.0 \times 10^{-6}$ ) with creatinine in the genome-wide association study. Although we did not find a relationship of the MYH9 locus with creatinine, this may simply reflect the low prevalence (~4%) of the MYH9 risk haplotype in Europeans (Supplementary Methods)<sup>3</sup>.

Our findings of common genetic variants associated with creatinine, cystatin c and CKD provide insight into the metabolic, solute and drugtransport mechanisms underlying kidney function and CKD. Further evaluation of these pathways may enable biomarker discovery and the development of new strategies to protect kidney function and prevent CKD.

Note: Supplementary information is available on the Nature Genetics website.

#### ACKNOWLEDGMENTS

Study-specific acknowledgments are provided in the Supplementary Note. We thank S. Asquith and J. Collier at K-bioscience for their help with the replication genotyping.

# AUTHOR CONTRIBUTIONS

J.C.C., P.E., L.L., J.S., G.N. and J.S.K. designed the study. J.C.C., W.Z., D.A.L. and P.v.d.H. led the data analysis. J.C.C., P.E., L.L., J.S., G.N. and J.S.K. wrote the manuscript, with contributions from all the authors.

### COMPETING FINANCIAL INTERESTS

The authors declare no competing financial interests.

Published online at http://www.nature.com/naturegenetics/.

Reprints and permissions information is available online at http://npg.nature.com/ reprintsandpermissions/.

- 1. Levey, A.S. et al. Kidney Int. 72, 247-259 (2007).
- 2. Köttgen, A. et al. Nat. Genet. 41, 712-717 (2009).
- 3. Kopp, J.B. et al. Nat. Genet. 40, 1175-1184 (2008).
- Stevens, L.A., Coresh, J., Greene, T. & Levey, A.S. N. Engl. J. Med. 354, 4. 2473-2483 (2006).
- Price, A.L. et al. Nat. Genet. 38, 904-909 (2006).
- Dyda, F., Klein, D.C. & Hickman, A.B. Annu. Rev. Biophys. Biomol. Struct. 29, 6. 81-103 (2000).
- Kharasch, E.D. Clin. Pharmacol. Ther. 84, 158-162 (2008).
- Lash, L.H., Fisher, J.W., Lipscomb, J.C. & Parker, J.C. Environ. Health Perspect. 8. 108 Suppl 2, 177-200 (2000).
- Marshall, J.D., Beck, S., Maffei, P. & Naggert, J.K. Eur. J. Hum. Genet. 15, 9. 1193-1202 (2007)
- 10. Mattoo, A. & Goldfarb, D.S. Semin. Nephrol. 28, 181-191 (2008).
- 11. Ciccia, A. et al. Mol. Cell 25, 331-343 (2007).
- 12. Fujita, T., Urban, T.J., Leabman, M.K., Fujita, K. & Giacomini, K.M. J. Pharm. Sci. 95, 25-36 (2006).
- 13. Filipski, K.K., Mathijssen, R.H., Mikkelsen, T.S., Schinkel, A.H. & Sparreboom, A. Clin. Pharmacol. Ther. 86, 396-402 (2009).
- 14. Naiche, L.A., Harrelson, Z., Kelly, R.G. & Papaioannou, V.E. Annu. Rev. Genet. 39, 219-239 (2005).
- 15. Chapman, D.L. et al. Dev. Dyn. 206, 379-390 (1996).

© 2010 Nature America, Inc. All rights reserved. <sup>1</sup>Department of Epidemiology and Biostatistics, School of Public Health, Imperial College of London, London, UK. <sup>2</sup>Ealing Hospital National Health Service (NHS) Trust,

Middlesex, UK. <sup>3</sup>Department of Nephrology and Transplantation and <sup>4</sup>Medical Research Council (MRC) Centre for Transplantation, King's College London, UK. <sup>5</sup>National Institute for Health Research (NIHR) Comprehensive Biomedical Research Centre, Guy's and St. Thomas' NHS Foundation Trust and King's College London, London, UK. <sup>6</sup>University Medical Center Groningen, University of Groningen, Groningen, The Netherlands. <sup>7</sup>MRC Centre for Causal Analyses in Translational Epidemiology, University of Bristol, Bristol, UK. <sup>8</sup>National Heart and Lung Institute, Hammersmith Hospital Campus, Imperial College London, London, UK. <sup>9</sup>Division of Medicine, University College London, London, UK. <sup>10</sup>Structural Bioinformatics Group, Imperial College London, London, UK. <sup>11</sup>Twin Research and Genetic Epidemiology Department, King's College London, St. Thomas' Hospital Campus, London, UK. <sup>12</sup>Department of Medical Genetics, University of Lausanne, Lausanne, Switzerland. <sup>13</sup>University Institute of Social and Preventive Medicine, Centre Hospitalier Universitaire Vaudois (CHUV), Lausanne, Switzerland. <sup>14</sup>Clinical Pharmacology Unit, University of Cambridge, Addenbrookes Hospital, Cambridge, UK. 15Clinical Pharmacology and The Genome Centre, William Harvey Research Institute, Barts and The London School of Medicine and Dentistry, London, UK. <sup>16</sup>Glasgow Cardiovascular Research Centre, University of Glasgow, Glasgow, Glasgow, UK. <sup>17</sup>Department of Histopathology, Imperial College, Hammersmith Hospital, London, UK. <sup>18</sup>Department of Gastroenterology and Hepatology, Imperial College London, London, UK. <sup>19</sup>Division of Nephrology, Université Catholique de Louvain Medical School, Brussels, Belgium. <sup>20</sup>Isala Clinics, Zwolle, The Netherlands. <sup>21</sup>Center for Renal Diseases, Zvezdara University Medical Center, Belgrade, Serbia. <sup>22</sup>South West Thames Institute for Renal Research, Epsom and St. Helier University Hospitals NHS Trust, Carshalton, UK. 23 London School of Hygiene and Tropical Medicine, London, UK. <sup>24</sup>Department of Human Genetics, Rheinisch-Westfaelische Technische Hochschule University Hospital Aachen, Aachen, Germany. <sup>25</sup>Cardiovascular Medicine, University of Oxford, Wellcome Trust Centre for Human Genetics, Oxford, UK. <sup>26</sup>National Institute of Aging, Clinical Research Branch–Longitudinal Studies Section, Baltimore, Maryland, USA. <sup>27</sup>Department of Nephrology and Clinical Immunology, RWTH University Hospital Aachen, Aachen, Germany. <sup>28</sup>MRC Epidemiology Unit, Institute of Metabolic Science, Addenbrooke's Hospital, Cambridge, UK. <sup>29</sup>Department of Internal Medicine, University Medical Center Groningen, University of Groningen, Groningen, The Netherlands. <sup>30</sup>Center for Statistical Genetics, University of Michigan, Ann Arbor, Michigan, USA. <sup>31</sup>Department of Clinical Sciences, Lund University, Malmö, Sweden. <sup>32</sup>Department of Nephrology, University Medical Center Groningen, Groningen, The Netherlands. <sup>33</sup>Department of Transplantation Immunology, University Medical Center Groningen, Groningen, The Netherlands. <sup>34</sup>Centre of Epidemiology for Child Health, University College London Institute of Child Health, London, UK. <sup>35</sup>Department of Medical Genetics, University of Lausanne and the Swiss Institute of Bioinformatics, Lausanne, Switzerland. <sup>36</sup>Langerhans Medical Research Group, Zwolle, The Netherlands. <sup>37</sup>Unit of Genetic Epidemiology and Bioinformatics, Department of Epidemiology, University Medical Center Groningen, University of Groningen, Groningen, The Netherlands. <sup>38</sup>Department of Molecular Medicine and Surgery, Karolinska University Hospital, Karolinska Institutet, Stockholm, Sweden. <sup>39</sup>Division of Nephrology, University of Maryland School of Medicine, Baltimore, Maryland, USA. <sup>40</sup>Wellcome Trust Sanger Institute, Wellcome Trust Genome Campus, Hinxton, Cambridge, UK. 41 Institute for Molecular Medicine Finland (FIMM), Nordic European Molecular Biology Laboratory (EMBL) Partnership for Molecular Medicine, Biomedicum Helsinki 2U and University of Helsinki, Helsinki, Finland. <sup>42</sup>National Institute for Health and Welfare, Helsinki, Finland. <sup>43</sup>Department of Psychiatry, Institute for Research in Extramural Medicine (EMGO), Neuroscience Campus, Vrije Universiteit (VU) Medical Center, Amsterdam, The Netherlands. <sup>44</sup>Department of Psychiatry, Leiden University Medical Center, Leiden, The Netherlands. <sup>45</sup>Department of Psychiatry, University Medical Center Groningen, University of Groningen, Groningen, The Netherlands. <sup>46</sup>Institute of Health Sciences, University of Oulu, Oulu, Finland. <sup>47</sup>Wellcome Trust Centre for Human Genetics, University of Oxford, Oxford, UK. <sup>48</sup>Oxford Centre for Diabetes, Endocrinology and Metabolism and <sup>49</sup>Oxford NIHR Biomedical Research Centre, Oxford, UK. <sup>50</sup>University of Southampton, Southampton, UK. <sup>51</sup>Institute of Clinical Medicine, Department of Clinical Chemistry, University of Oulu, Oulu, Finland. <sup>52</sup>Cardiovascular Sciences, University of Leicester, Glenfield Hospital, Leicester, UK. <sup>53</sup>Istituto di Neurogenetica e Neurofarmacologia, Consiglio Nazionale delle Ricerche, Cagliari, Italy. <sup>54</sup>Department of Molecular Medicine and Surgery, Karolinska University Hospital, Karolinska Institutet, Stockholm, Sweden. 55 Laboratory of Genetics, US National Institutes of Health Biomedical Research Center, National Institute on Aging, Baltimore, Maryland, USA. <sup>56</sup>Division of Endocrinology, Diabetes and Nutrition, University of Maryland School of Medicine, Baltimore, Maryland, USA. <sup>57</sup>Division of Renal Medicine K56, Karolinska University Hospital at Huddinge, Stockholm, Sweden. <sup>58</sup>Chemical Pathology, Guy's and St Thomas' Hospitals Trust, London, UK. <sup>59</sup>General Practice 't Veen, Hattem, The Netherlands. <sup>60</sup>Department of Internal Medicine, University Hospital Center, University of Lausanne, Lausanne, Switzerland. <sup>61</sup>Cambridge Institute for Medical Research, Addenbrooke's Hospital, Hills Road, Cambridge, UK. <sup>62</sup>Medical Genetics, Clinical Pharmacology and Discovery Medicine, GlaxoSmithKline, King of Prussia, Pennsylvania, USA. 63Biocenter Oulu, University of Oulu, Oulu, Finland. 64Renal Section, Division of Medicine, Hammersmith Hospital Campus, Imperial College London, London, UK. 65 MRC-HPA Centre for Environment and Health, Department of Epidemiology and Biostatistics, School of Public Health, Imperial College of London, London, UK. 66 These authors contributed equally to the work. Correspondence should be addressed to J.C.C. (john.chambers@ic.ac.uk) or J.S.K. (j.kooner@imperial.ac.uk).