Genetic studies of leptin concentrations implicate leptin in the regulation of early

adiposity

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Abstract

Leptin influences food intake by informing the brain about the status of body fat stores. Rare *LEP* mutations associated with congenital leptin deficiency cause severe earlyonset obesity that can be mitigated by administering leptin. However, the role of genetic regulation of leptin in polygenic obesity remains poorly understood. We performed an exome-based analysis in up to 57,232 individuals of diverse ancestries to identify genetic variants that influence adiposity-adjusted leptin concentrations. We identify five novel variants, including four missense variants, in *LEP, ZNF800, KLHL31*, and *ACTL9*, and one intergenic variant near *KLF14*. The missense variant Val94Met (rs17151919) in *LEP* was common in individuals of African ancestry only and its association with lower leptin concentrations was specific to this ancestry (P=2x10⁻¹⁶, n=3,901). Using *in vitro* analyses, we show that the Met94 allele decreases leptin secretion. We also show that the Met94 allele is associated with higher BMI in young African-ancestry children but not in adults, suggesting leptin regulates early adiposity.

Introduction

Leptin is an adipocyte-derived hormone that helps maintain homeostatic control of fat tissue mass by signaling the status of body energy stores to the appetite-regulating circuits of the brain [1]. Rare homozygous mutations in the leptin (*LEP*) gene can cause complete leptin deficiency that results in hyperphagia and severe early-onset obesity, which can be treated effectively by exogenous leptin administration [2, 3]. Mice and patients heterozygous for these mutations show partial leptin deficiency and increased body weight [4-6].

In the general population, leptin concentrations correlate closely with body fat mass. However, there is wide inter-individual variability; about 10-20% of obese individuals have leptin concentrations that are similar to those observed in non-obese individuals, which is in part due to genetic differences [7, 8]. Twin and family studies suggest that 30-50% of variation in leptin at any given level of adiposity and across different ethnic groups is explained by genetic differences [8]. The implications of this variability for body weight regulation remain poorly understood.

Identification of genetic variants associated with circulating leptin may shed new light on the role of variability in leptin levels in the general population. In a recent genome-wide association study (GWAS) of leptin concentrations, we identified four loci associated with leptin concentrations independent of body mass index (BMI) [9]. The variant most strongly associated with leptin concentrations was rs10487505, located 21 kb upstream from *LEP*, in a region shown to harbor a long non-coding RNA (EST EL947753) that influences the transcriptional control of leptin expression [10]. The leptin-decreasing allele of

rs10487505 was nominally associated with ~0.03 kg/m² higher BMI in adults and 1.05fold increased risk of early-onset obesity [9]. More recently, the association of the leptindecreasing allele of rs10487505 with higher adult BMI, body fat percentage, and risk of extreme obesity was replicated in the UK Biobank [10]. The most pronounced association, however, was observed for body size at 10 years of age; carriers of the leptin-decreasing allele reported being "plumper" at age 10 compared to peers" more frequently than carriers of the allele associated with higher leptin concentration. The association between rs10487505 and childhood body size was recently replicated in 14,521 Norwegian children, and the peak effect of rs10487505 on BMI was observed in 1.5-year-old children [11].

In the present study, we sought to elucidate the genetic basis of leptin concentrations through screening genetic variants with an exome-targeted array in up to 57,232 individuals of European, African, East Asian or Hispanic ancestry. We confirm five previously established and identify five novel variants associated with leptin concentrations, including four missense variants in *LEP*, *ZNF800*, *KLHL31*, and *ACTL9*, and one intergenic variant near *KLF14*. The novel *LEP* variant, Val94Met (rs17151919), is associated with leptin concentrations in adults of African ancestry only. The leptin-lowering Met94 allele of the rs17151919 variant is associated with higher BMI in young children, but shows a weak or no association with BMI in adulthood, suggesting leptin regulates early adiposity.

Research Design and Methods

Study design

We performed an exome-based association study using data from 35 cohorts comprising up to 57,232 adults (≥18 years) of whom 50,321 were of European descent, 4,387 of African descent, 2,036 of East Asian descent, and 488 of Hispanic descent. We carried out additional analyses in men and women separately. All analyses were performed in models combining studies of all ancestries and in European ancestry cohorts only, for both additive and recessive genetic models. All participating institutions and coordinating centers approved the project. Informed consent was obtained from all study participants. We have reported the study-specific design, sample quality control, and descriptive statistics in **Tables S1-S2**.

Outcome traits

The participating studies acquired residuals for leptin concentrations (in ng/mL) using linear regression, adjusting for age, genome-wide principal components, and any study-specific covariates (e.g. study center). The residuals were calculated with and without adjustment for BMI. Studies with unrelated individuals acquired the residuals in men and women separately, whereas family-based studies additionally acquired sex-combined residuals adjusting for sex as a covariate. Case-control studies acquired the residuals in cases and controls separately. Finally, we rank-transformed the residuals using inverse normal transformation to follow a distribution with a mean of 0 and a standard deviation of 1.

Genotyping

All participating studies performed genotyping using the Illumina HumanExome BeadChip. The genotype calling was performed using the designated manufacturer's software, followed by zCall. Study-specific quality control measures were implemented before the association analyses to remove poorly genotyped variants (**Table S3**).

Study-level association analyses

Associations of the exome-wide variants with the residuals of leptin concentrations were examined using linear mixed models implemented in either RAREMETALWORKER [12] or RVTEST [13] (**Table S3**). The model accounted for potential cryptic relatedness by incorporating a kinship matrix. We performed the single variant association analyses using both additive and recessive genotypic models. We also calculated covariance matrices capturing LD relationships between markers within 1 Mb for use in gene-level meta-analyses.

Quality control of study-level association results

We applied the EasyQC package in R to association summary statistics from each participating study to identify cohort-specific QC issues. This included (i) identifying issues with calculation of leptin residuals and transformation of the residuals, (ii) identifying strand issues by comparing allele frequencies against reference alleles from the 1000 Genomes Project phase 1, and (iii) identifying issues arising from population stratification.

Single variant meta-analyses

The meta-analyses of summary statistics from the participating studies were carried out using RAREMETAL [14] by two different analysts in parallel. We excluded all variants with a call rate <98%, Hardy Weinberg equilibrium *P*-value <1x10⁻⁶, or an allele frequency that strongly deviated from the 1000 Genomes reference frequency (>0.60 for all-ancestry analyses and >0.30 for ancestry-specific analyses). To identify the leptin-associated variants, we used the array-wide Bonferroni-corrected threshold of *P*<2x10⁻⁷ for ~250,000 variants in the single variant analyses.

Gene-based meta-analyses

We performed gene-based analyses using the sequence kernel association test [15] (SKAT) and variable threshold [16] (VT) methods in RAREMETAL. The analyses were performed with two different sets of criteria (broad and strict) to select predicted damaging rare and low-frequency variants with MAF<5% annotated using five prediction algorithms: PolyPhen-2, HumDiv, HumVar, LRT, MutationTaster, and SIFT. The broad gene-based tests included nonsense, stop-loss, splice-site, and missense variants that were annotated as damaging by at least one of the five algorithms whereas the strict tests only included variants predicted as damaging by all of the five algorithms. The statistical significance for the gene-based tests was set at a Bonferroni-corrected threshold of P<2.5x10⁻⁶ for 20,000 genes.

Age-stratified BMI analyses of variants in and near LEP

To study the influence of age on the association of the Val94Met variant in *LEP* and the rs10487505 variant near *LEP* with childhood BMI, we performed age-stratified analyses

in children with African and European ancestry from the Center for Applied Genomics at Children's Hospital of Philadelphia (CHOP) cohort recruited from 2006 to present [17]. The participants had multiple BMI measurements at different ages and analyses were performed with measurements in 1-year age bins. The number of BMI measurements in each age bin is shown in **Tables S9** and **S10**. Statistical significance was defined as P<0.05. The Val94Met and rs10487505 variants were genotyped using the Illumina Infinium II HumanHap550 and Human610 BeadChip and imputed to the HRC r1.1 reference panel using the Sanger Imputation Server. All participants were biologically unrelated, aged between 2 and 18 years, and between -3 and +3 standard deviations of CDC-corrected BMI. The study was approved by the Institutional Review Board of the Children's Hospital of Philadelphia. Parental informed consent was given for each study participant.

Additionally, we used information on comparative body size at age 10 (data field 1687) for 452,264 individuals of European ancestry and 8,154 individuals of African ancestry from the UK Biobank. The participants were asked to choose one of the three categories of "about average", "thinner", or "plumper" to describe their body size compared to average when they were 10 years old.

Pathway enrichment analyses

We utilized the EC-DEPICT [18, 19] gene set enrichment analysis method to evaluate nonsynonymous index variants (strongest nonsynonymous variant within ± 1 Mb boundary) with *P*<5x10⁻⁴ for association with either i) leptin unadjusted for BMI, or ii) leptin adjusted for BMI. EC-DEPICT's primary innovation is the use of "reconstituted" gene sets,

which consist of gene sets downloaded from several databases that have been extended based on publicly available large-scale co-expression data [18]. Two analyses were performed: (i) all coding variants (N=93 loci for leptin unadjusted for BMI and N=91 loci for leptin adjusted for BMI) and (ii) coding variants with MAF<5% only (N=77 loci for leptin unadjusted for BMI and N=65 loci for leptin adjusted for BMI).

We also utilized PASCAL [20] to study the enrichment of exome-wide association results in gene sets and pathways using two estimation approaches: MAX and SUM. The MAX estimation is more powerful for single variant-driven associations whereas the SUM estimation is more powerful when multiple variants are driving the signal [20]. We used reconstituted gene sets from DEPICT and the reference data from UK10K [TwinsUK [21] and ALSPAC [22]] to estimate LD. The PASCAL analyses were performed for all exomechip variants (N_{all} =265,780 for leptin adjusted for BMI, N_{all} =265,780 for leptin unadjusted). and for coding variants only (N_{coding} =176,035 for leptin adjusted for BMI, N_{coding} =180,864 for leptin unadjusted). No allele frequency or *P*-value thresholds were used to select variants for the PASCAL analyses. The pathway scoring method used by PASCAL combines individual gene scores without the need for a tuneable threshold parameter to determine inclusion of genes in the enrichment analysis [20].

Leptin adjusted for BMI is correlated with body fat free mass (correlation with fat-free mass index in the Fenland cohort = -0.39). The initial pathway analyses for leptin adjusted for BMI using EC-DEPICT and PASCAL suggested enrichment of skeletal-muscle related pathways. To make sure that the gene set enrichment results were not due to correlation between leptin adjusted for BMI and fat-free mass index, we corrected the effect sizes

using the following equation [23]: $Beta_{corrected} = beta_{leptin} - (beta_{FFMI} \times r_{FFMIvs.LEPTIN})$, where $r_{FFMIvs.LEPTIN} = -0.39$ (Pearson correlation coefficient in the Fenland Study). The beta_{FFMI-} coefficients were extracted from an ongoing exome-wide association study of fat-free mass index in ~500,000 individuals.

Collider bias

Given that we adjusted leptin concentrations for BMI in our exome-based analyses and leptin and BMI are strongly correlated (r~0.5-0.8) [9], we tested all exome-based significant loci for evidence of collider bias [23-25]. For each index we extracted the association results from our BMI-unadjusted leptin analyses and from the largest published exome-wide analysis for BMI [19]. We corrected BMI-adjusted associations for potential bias due to phenotypic correlation between leptin concentrations and BMI, and compared the strength and significance of association with leptin concentrations unadjusted for BMI, leptin adjusted for BMI, and association with BMI (**Table S6**).

eQTL colocalization analyses

The *cis*-expression quantitative trait locus (*cis*-eQTL) analyses were carried out by using abdominal subcutaneous adipose tissue from 770 participants of the METSIM (Metabolic Syndrome in Men) study who all were Finnish men from Kuopio, Finland [26]. The eQTL mapping in 770 METSIM individuals was performed by EPACTS implementing a linear mixed model to account for the population structure among the samples. The eQTLs were defined as *cis* (local) if the peak association was within 1 Mb on either side of the exon boundaries of the gene. We also identified variants most strongly associated with

genes/transcripts from the index variant ("eSNP"). We used METSIM LD (based on n=770, HRC imputation) to assess LD r^2 between the index variant and the lead eSNP. If the pairwise LD was $r^2>0.80$, we performed a reciprocal conditional analysis. We tested association between the lead SNP and transcript level when the lead eSNP was included in the model, and *vice versa*.

Expression of the potential causal genes in preadipocytes and mature adipocytes

We compared the expression of the candidate causal genes in the novel leptin-associated loci, including *ZNF800*, *KLF14*, *KLHL31*, *ACTL9*, *CNTD1* and *DNAJC18* in preadipocytes and mature adipocytes, two major constituent cell types of adipose tissue. Human preadipocytes isolated from adipose tissue were induced to undergo adipocyte differentiation *in vitro* [27]. RNA samples were obtained from preadipocytes and lipid-laden mature adipocytes at post-differentiation day 12.

Impact of Val94Met variant in *LEP* on leptin protein stability

We used UCSF Chimera 1.13.1 to model the 3D protein structure and valine-tomethionine substitution in the leptin protein [28]. The Rotamers tool and the Dunbrack Rotamer Libary were used to view and evaluate amino acid sidechain rotamers. The displayed orientation of methionine was chosen based on the clashes and contacts observed in the protein and hydrogen bonds [29]. To predict protein stability, we used SDM [30], iStable [31], Cupsat [32] and iMutant 2.0 [33]. All analyses applied the 3D structure for leptin (ID 1AX8) from the RSCCP Protein Data Bank as the reference data set.

Effect of the Val94Met variant in LEP on leptin protein stability and secretion rate

We tested the effect of the Val94Met variant on leptin protein stability and secretion rate in HEK293 cells in vitro. Human leptin cDNA clone was obtained from Open Biosystems Inc (Huntsville, AL) and subcloned into pcDNA3.1 vector. The original cDNA clone encodes the Val94 variant. The 94Met variant was created using Quikchange II sitedirected Mutagenesis kit (Agilent, Santa Clara, CA), with the Val94 plasmid as template and the following mutagenesis primers (forward: 5'-atgccttccagaaacatgatccaaa tatccaac-3', reverse: 5'-attagatatttagatcatatttctagaaagacat-3'). Plasmids carrying Val94 or 94Met cDNA (0.05 µg) were introduced into HEK293 cells (0.65 million cells/well in 12-well plate) using Lipofectamine 2000 as previously described [34]. To measure intracellular leptin protein turnover and secretion rates, cells were treated with protein synthesis inhibitor cycloheximide (CHX, 20 µg/ml) in fresh media for 0.5 and 1 hr at 72 hr post-transfection. Cells incubated with fresh media for 1 hr without CHX were used as untreated controls. Conditioned media were saved for leptin assay, and cell lysate were prepared using NP-40 lysis buffer (50 mM Tris-HCl, pH 8.0, 150 mM NaCl, 1 mM DTT, 1 mM EDTA, 0.5% NP-40, 10% glycerol, and 1x Roche protein inhibitor mixture). Leptin concentrations in cell lysates and the amount of leptin in conditioned media were determined using a human leptin ELISA kit (R&D Systems, Minneapolis, MN). Little or no cell debris was observed in the conditioned media after centrifugation, suggesting little or no cell breakage during the incubation. The experiments were carried in duplicates or triplicates and repeated four times.

Results

Five novel genetic variants show association with leptin concentrations independent of adiposity

To identify genetic variants associated with leptin concentrations, we tested the associations of 246,328 single nucleotide variants (SNVs), genotyped on an exometargeted genotyping array, with leptin concentrations in up to 57,232 individuals of European (n=50,321), African (n=4,387), East Asian (n=2,036) or Hispanic ancestry (n=488) from 35 studies (**Tables S1-S3**). The exome-array provides a detailed coverage of gene-coding regions and includes tags for variants identified in previously published GWASs for human complex traits. Given the strong correlation between leptin and BMI (r~0.5-0.8) [9], we examined associations with leptin concentrations with and without adjustment for BMI. Additional analyses were performed in men (n=23,862) and women (n=32,940) separately. All the analyses were performed in all ancestries combined and in European-ancestry individuals only.

We confirmed five previously established [9] and identified five novel variants associated with leptin concentrations. The novel associations include four missense variants, in *LEP*, *ZNF800*, *KLHL31*, and *ACTL9*, and one intergenic variant near *KLF14* (**Table 1, Table S4**). The associations at already established loci include intergenic variants near *LEP* and *CCNL1*, a missense variant in *GCKR*, and intronic variants in *COBLL1* and *FTO* (**Table 1, Table S4**). To detect additional independent signals at the 10 leptin-associated loci, we performed conditional analyses, but no further signals were identified.

The association between rs1121980 near *FTO* and leptin concentrations became nonsignificant after adjustment for BMI (P_{unadj} =8x10⁻¹⁷; P_{adjBMI} =0.45). The effects of all other known and novel loci were independent of BMI (**Table S5**). We tested whether the adjustment for BMI, a strongly correlated covariate [23], may have introduced collider bias, but found no evidence of such bias (**Table S6**).

The strongest variant associated with leptin concentrations was rs791600, an intergenic variant near the *LEP* gene. The rs791600 variant is in linkage disequilibrium (LD) (EUR r^2 =0.70) with the rs10487505 variant identified in our previously published GWAS [9], which is not included in the exome array and was therefore not available for analyses in the present study. In the prior GWAS study, the rs10487505 variant showed a more significant association with BMI-adjusted leptin concentrations (beta=0.034 per allele, P=2.7x10⁻¹¹, n=29,252) than rs791600 (beta=0.029 per allele, P=3.0x10⁻⁹, n=31,800) and thus is still considered the lead variant at this locus (**Figure S1**).

Nine of the 10 identified loci showed an association with leptin concentrations in all ancestries combined and in European ancestry only analyses. However, the novel *LEP* variant Val94Met (rs17151919) only showed a significant association in all ancestries combined (P=2x10⁻¹⁶) and not in European-ancestry individuals alone (P=0.47, **Table S7**). In further ancestry-stratified analyses, we observed that the Met94-coding allele is common in populations of African ancestry (MAF=8%), less common in those with Hispanic ancestry (MAF=2%), very rare in those with European ancestry (MAF=0.02%) and monomorphic in people with East Asian ancestry [35]. In individuals of African descent, each Met94-coding allele was associated with 0.34 standard deviations (SD)

lower leptin concentrations ($P=2x10^{-16}$, n=3,901) (**Figure S2**). The direction of effect was consistent in individuals with Hispanic (-0.21 SD effect per allele, P=0.29, n=488) and European ancestry (-0.19 SD effect per allele, P=0.47, n=44,401), but did not reach statistical significance, most likely because very few carriers were available (N_{HIS}=24, N_{EUR}=15) (**Table S7**).

Gene-based analysis identifies two novel genes with sex-specific effect on leptin

In addition to single variant-based association tests, we performed gene-based tests using rare and low-frequency coding variants in aggregate [15, 16] (**Methods**). We identified two genes associated with leptin concentrations. *CNTD1* showed association with leptin concentrations unadjusted for BMI in men ($P=1\times10^{-7}$) but not in women (P=0.27) (**Table 2, Table S8**). The association in men was driven by five coding variants and was strongly attenuated by adjusting for BMI (P=0.007), suggesting that the association of *CNTD1* with leptin concentrations may be due to a link between *CNTD1* and adiposity, although no such connection has been previously reported. The *CNTD1* gene encodes cyclin N-terminal domain-containing 1, which is critical for meiotic crossover maturation and deselection of excess pre-crossover sites.

Another gene, *DNAJC18*, showed association with BMI-adjusted leptin concentrations in women ($P=6\times10^{-8}$), but not men (P=0.02). The association in women was driven by two coding variants (**Table 2, Table S8**). *DNAJC18* is part of the Dnaj heat shock protein family. However, no function has yet been described to C18 subfamily.

LEP Val94Met regulates leptin secretion and early adiposity

The Val94Met (rs17151919) LEP variant was associated with BMI-adjusted leptin concentrations in individuals of African ancestry. A previous study in 2,129 African Americans in the CARDIA study (not included in the present meta-analyses) reported a significant association between the leptin-decreasing Met94 allele of the Var94Met (rs17151919) variant in LEP and up to 1.12 kg/m² higher BMI in adulthood (P=0.018) [36]. However, results from two larger studies of BMI by the African Ancestry Anthropometry Genetics Consortium (n=42,752; P=0.88) [37] and the African ancestry population of the UK Biobank study (N=7,820, P=0.17), did not replicate the association. Nevertheless, among the African ancestry population in the UK Biobank, carriers of the leptindecreasing Met94 allele reported more often that, at age 10, they were "plumper" (compared to peers) (OR=1.11, P=0.04), suggesting that the effect of this variant may be age-dependent. To study the influence of age, we performed age-stratified analyses in up to 2,726 children with African-ancestry from the Center for Applied Genomics at Children's Hospital of Philadelphia (CHOP) cohort [17]. Comparing the effect sizes across different age points revealed that each leptin-decreasing Met94 allele was associated with 0.12-0.20 units higher BMI z-score between the ages 3 and 7 (P<0.05). The most pronounced effect was reached at age 6 years (Figure 1, Table S9), and no association with BMI was observed after age 8 years (betas -0.04 to 0.05) (Figure 1, Table S9), suggesting that the BMI-increasing effect of the Met94 allele wanes shortly before puberty. The rs10487575 variant near LEP showed a similar trajectory of association with childhood BMI as the Val94Met variant but the effect sizes were much more modest (Figure 1, Table S10), consistent with the five-fold smaller effect of rs10487505 on leptin concentrations compared to Val94Met in adults (Table 1).

The Val94Met variant is located at position 94 in the 167 amino acid leptin precursor protein and results in a valine to methionine change at position 73 of the mature protein (**Figure 2A**). Position 73 is situated at the leptin protein surface and is not believed to be involved in binding of leptin to its receptor. Nevertheless, structural prediction tools [30-33] suggested that the substitution of valine with methionine at this position is likely to lead to reduced stability of the mature leptin protein (**Figure 2A**, **Figures S3-4**, **Table S11**). This is consistent with our observation that the methionine-coding allele is associated with lower leptin concentrations.

To study the impact of the Val94Met variant on the intracellular turnover of the leptin protein and its secretion rate, we performed *in vitro* experiments in HEK293 cells. Leptin secretion rate – calculated as the amount of leptin secreted in 1 hour normalized to the respective cellular leptin content – was 20.4% lower in Met94 than in Val94 cells (*P*=0.0007 by repeated measures 1-way ANOVA) 72 hours post-transfection (**Figure 2B**). Leptin secretion rates between 48-72 hours post-transfection and during a 1-hour treatment with cycloheximide were 11.8% (*P*=0.0005) and 17.9% (*P*=0.0002) lower, respectively, in Met94 compared to Val94 (**Figure S5**). Notably, no difference was found in the intracellular turnover rate of leptin between Val94 and Met94 cells during a 0.5 or 1-hour incubation with cycloheximide to impair protein synthesis (**Figure 2C**). The unchanged turnover rate incorporates protein secretion and degradation, suggesting that decreased leptin secretion rate was likely associated with increased intracellular leptin degradation in Met94 cells. Overall, these *in vitro* experiments suggest that methionine substitution in position 73 of the mature leptin protein decreases the rate of leptin

secretion from the cells, which may contribute to the association of the Met94 allele with lower leptin concentrations.

ZNF800 locus regulates adipose gene expression and body composition

The Pro103Ser (rs62621812) variant in *ZNF800* changes the amino acid sequence of CH2 zinc finger protein, a putative transcription factor [38]. We found that the Ser103 allele (frequency=2.8%) is associated with lower BMI-adjusted leptin concentrations (P=2.0x10⁻¹²). As shown before [26], Pro103Ser is the lead variant associated with expression of *ZNF800* in subcutaneous adipose tissue in the Finnish METSIM Study (P=2.4x10⁻¹⁶); the Ser103 allele is associated with higher *ZNF800* expression levels (**Table S12**). *ZNF800* is a master regulator in subcutaneous adipose tissue expression of nine other genes [26]. In the eQTL data, the leptin-decreasing Ser103 allele was not significantly associated with the expression of *LEP* (P=0.20), located 866 kb downstream, and the observed direction of the effect on *LEP* expression was opposite to that observed for leptin concentrations (beta=0.14 SD/allele vs. beta= -0.13 SD/allele, respectively), suggesting that the leptin-lowering effect of the Ser103 allele on leptin concentrations is unlikely to be mediated by direct transcriptional regulation of *LEP*.

In the UK Biobank study, we found that each leptin-decreasing Ser103 allele is associated with 0.14 kg/m² higher BMI (P=8.1x10⁻⁶). However, there was no association between Ser103 allele and body fat percentage (-0.045% per allele, P=0.25), indicating that the variant impacts BMI primarily by increasing fat free body mass. Indeed, the leptin-decreasing Ser103 allele was associated with a 0.33 kg higher fat free mass (P=4.6x10⁻⁶)

²⁰) and only 0.13 kg higher fat mass (P=0.023). The Ser103 allele is associated with higher expression of the *ZNF800* gene in the tibial nerve (GTEx v8, P=1.4x10⁻⁶, n=532) that innervates the muscles of the leg, and has been previously identified for association with increased appendicular lean mass [39]. There was no association between the Ser103 allele and self-reported body size at age 10 (P=0.75).

The KLF14 locus regulates adipogenesis and fat distribution

The rs972283 variant (MAF_{EUR}=48%), associated with leptin concentrations, is located 51 kb upstream from KLF14 and 2.5 Mb downstream from LEP, and is in near-perfect LD with previously reported GWAS variants for type 2 diabetes [40], insulin-resistance [41], HDL cholesterol [42], and body fat distribution [43]. As reported earlier [26], rs972283 is associated with KLF14 expression in subcutaneous adipose tissue (Table S12). As KLF14 is a master regulator in adipose tissue, rs972283 is also associated with the expression of multiple other genes in trans [44]. No significant association was observed between rs972283 and LEP expression in the METSIM eQTL study [44], suggesting that *KLF14* may not regulate leptin production at the transcriptional level, at least not in men. Lower expression of KLF14 has been implicated in impaired adipogenesis due to defective adipocyte glucose uptake in women, characterized by the presence of fewer but larger adipocytes and a shift in fat distribution from gynoid stores to abdominal tissues [44]. However, while the effects of KLF14 on adipogenesis and adipose redistribution have been found to be specific to women, there was no difference in the association of rs972283 with leptin levels between men and women (Table S4).

Interestingly, the carriers of the rs972283-G allele reported more frequently being plumper (P=2.8x10⁻⁵) and shorter (P=0.014) than average at age 10 in the UK Biobank than noncarriers, whereas the same allele was associated with a lower BMI (P=6.8x10⁻⁹) and increased height (P=0.010) in adults, suggesting that the effect of the rs972283 variant on body size may change during life course. In previous GWAS of adults, the rs972283-G allele has been identified to be associated with higher risk of type 2 diabetes [40] and insulin resistance [41], and lower hip circumference (adjusted for BMI) [43] and HDL cholesterol [42]. In the UK Biobank study, the rs972283-G allele was associated with lower body fat percentage in adults (P=5.9x10⁻²²).

The KLHL31 locus is implicated in adipogenesis in adult females

The Val156IIe (rs3799260) variant (MAF_{EUR}=18%) in *KLHL31*, associated with leptin concentrations in female-only analyses, changes the amino acid sequence of the kelchlike family member 31 protein. *KLHL31* suppresses Wnt- β -catenin signaling that is involved in promoting adipocyte differentiation and suppressing oxidative metabolism in adipocytes. The Val156IIe variant is predicted to be benign/tolerated by SIFT/Polyphen [45, 46]. Previous genetic associations have identified a variant in low LD (rs7739232; EUR r²=0.27) to be associated with BMI-adjusted hip circumference, also specific to women [43]. The rs7739232 variant was not included in the exome-array and was thus not analyzed in the present study. Our *in vitro* experiments showed that *KLHL31* is only expressed in mature adipocytes, but not in preadipocytes (**Figure S6**), suggesting that the gene is developmentally regulated.

In the UK Biobank, similar to the variants in and near *LEP*, the carriers of the leptindecreasing lle156 allele reported more often being plumper than average at age 10 (P=5.6x10⁻⁶), but there was a weaker association with higher BMI (P=0.045) in adulthood.

In men, the ACTL9 locus may regulate leptin concentrations in a cell nonautonomous fashion

Homozygosity for the minor allele of the Ser37Phe (rs2340550) variant in *ACTL9* was associated with leptin concentrations in men only in a recessive genetic model. While the Ser37Phe variant is predicted to be benign/tolerated by SIFT/Polyphen, another missense variant, Ala51Val (rs10410943), in high LD (EUR r^2 =0.99) is predicted to be deleterious/probably damaging and could be the causal variant at the locus. The Ser37Phe variant is also in high LD (r^2 >0.8) with several nearby non-coding variants (**Figure S7**). However, none of these overlaps with regulatory elements in adipocytes. The expression of *ACTL9* is restricted to the testis and it is therefore likely to act in an adipocyte non-autonomous fashion to influence leptin concentrations. Actin proteins have cytoskeletal functions and have also been implicated in signaling and nuclear activities.

Gene-set analyses implicate adipocyte-related pathways

We performed gene-set enrichment analyses using EC-DEPICT [18, 19, 47] and PASCAL [20] to identify biological processes and candidate pathways enriched for loci associated with leptin unadjusted or adjusted for BMI. Among coding variants associated with BMIunadjusted leptin concentrations, PASCAL identified significant enrichment of the geneset for "positive regulation of reproductive success" ($P_{empirical}$ =1.6x10⁻⁵) (**Table S13**),

consistent with the crucial permissive role of leptin in the integrity of the gonadal axis [48]. Among coding variants associated with leptin adjusted for BMI, we found enrichment of the immune-related TRIM39 protein-protein interaction subnetwork [49, 50] ($P_{empirical}$ =8.4x10⁻⁶) (**Table S14**). No gene sets were found to be significantly enriched in PASCAL analyses where all exome-wide variants (coding and non-coding) for leptin adjusted for BMI were included, nor in the EC-DEPICT analyses.

Discussion

We identified 10 genetic variants associated with leptin concentrations and two genebased associations using an exome-based genotyping array in up to 57,232 individuals with varying ancestries. The two independent variants most strongly associated with leptin concentrations were located in and near the *LEP* gene. The African ancestry-driven variant within *LEP*, Val94Met (rs17151919), was found to decrease leptin secretion in HEK293 cells whereas rs10487505 located near *LEP* overlaps a lncRNA that regulates *LEP* expression [10]. Both variants showed significant association with increased adiposity in children, whereas only a nominal or no association was observed in adults.

Previous analyses have shown that the leptin-lowering allele of rs10487505 is only weakly associated with higher BMI in adulthood but shows a pronounced association with BMI in early childhood [10, 11]. Similarly, we showed that the *LEP* Met94 allele, associated with lower leptin concentration, is associated with early childhood BMI. Our results suggest that leptin has an impact specifically on early adiposity, encouraging further studies to uncover the molecular mechanisms that underlie this age-dependent relationship between leptin and BMI.

The Val94Met and rs10487505 variants in and near *LEP* are likely to influence leptin concentrations by different molecular mechanisms. The novel African ancestry-driven variant Val94Met may affect circulating levels of leptin by reducing leptin secretion. The rs10487505 variant is associated with leptin mRNA levels in adipose tissue. Located upstream of *LEP* within a lncRNA (EL947753), we hypothesize that this variant interacts with enhancer regions to regulate the expression of *LEP*. Defects in *LEP* regulation in

mice lead to a relative hypoleptinemic form of obesity that is responsive to leptin administration [10].

We identified four new loci associated with leptin concentrations, located in or near the *ZNF800*, *KLF14*, *KLHL31* and *ACTL9* genes. Two additional genes, *CNTD1* and *DNAJC18* were identified in gene-based analyses. The *ZNF800* and *KLF14* genes are master *trans*-regulators of adipose tissue gene expression [26] and located in the proximity of the *LEP* gene (866 kb and 2.5 Mb away, respectively). The variants in *ZNF800* and near *KLF14* were not associated with *LEP* mRNA levels, however, suggesting that they may be involved in translational or post-translational rather than transcriptional regulation of leptin production. *KLHL31* has been shown to promote adipocyte differentiation and suppress oxidative metabolism in adipocytes, whereas *ACTL9* is not expressed in adipocytes and could affect circulating levels by a non-cell autonomous mechanism. The *KLHL31* and *ACTL9* loci, and the *CNTD1* and *DNAJC18* genes, were only identified in sex-specific models and narrowly passed the array-wide significance threshold. Further validation of the association of these loci with leptin concentrations is warranted.

In summary, we identified a new genetic association of an African ancestry-specific missense variant rs17151919 in *LEP* with leptin concentrations and replicated the association of the rs10487505 variant near *LEP*. The pronounced association of these variants with BMI in early childhood implicates genetic regulation of *LEP* in early growth and suggests that young children may be particularly sensitive to the metabolic/behavioral effects of leptin. We also identified novel loci at *ZNF800*, *KLF14*, *KLHL31*, *ACTL9*, *CNTD1*

and *DNAJC18* associated with leptin concentrations, providing additional insights into leptin physiology.

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Guarantor Statement

HY and TOK are the guarantor of this work and, as such, had full access to all the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis.

Conflict of Interest statement

This work was conducted prior to M.E.G's current affiliation with the National Heart, Lung, and Blood Institute, and, as such, the views expressed in this article do not represent the views of the NHLBI, NIH, or other government entity. D.M.-K. is a part-time clinical research consultant for Metabolon, Inc. M.A.N's participation is supported by a consulting contract between Data Tecnica International and the National Institute on Aging, National Institutes of Health. V.S. has served in advisory boards for Novo Nordisk and Sanofi and received honoraria from these companies. He also has ongoing research collaboration with Bayer Ltd (all unrelated to the present study). B.M.P. serves on the Steering Committee of the Yale Open Data Access Project funded by Johnson & Johnson. J.R.K. reports stock ownership in Bristol Myers Squibb, Johnson & Johnson, Merck, and Pfizer.

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Tables

Table 1. Leptin-associated loci identified in exome-based association analyses

SNP	Chr	Position	Nearest gene	Trait	Most significant model	Annotation	EA	OA	EAF	Beta	SE	P value	Ν
Novel variant	s												
rs3799260	6	53,519,605	KLHL31	LeptinAdjBMI	Additive / All ancestries / Women	missense	С	Т	0.175	0.055	0.010	1.0E-07	32,886
rs62621812	7	127,015,083	ZNF800	LeptinAdjBMI	Additive / All ancestries	missense	А	G	0.028	-0.127	0.018	2.0E-12	56,708
rs17151919	7	127,894,592	LEP	LeptinAdjBMI	Additive / All ancestries	missense	А	G	0.007	-0.333	0.040	1.5E-16	49,034
rs972283	7	130,466,854	KLF14	LeptinAdjBMI	Additive / European	intergenic	А	G	0.479	0.056	0.006	3.8E-18	49,830
rs2340550	19	8,808,942	ACTL9	LeptinAdjBMI	Recessive / European / Men	missense	А	G	0.316	0.071	0.014	2.0E-07	21,883
Previously identified variants													
rs1260326	2	27,730,940	GCKR	LeptinAdjBMI	Additive / All ancestries	missense	Т	С	0.375	-0.050	0.006	2.7E-15	56,708
rs13389219	2	165,528,876	COBLL1	LeptinAdjBMI	Additive / All ancestries	intronic	Т	С	0.410	0.053	0.007	3.0E-15	50,297
rs900399	3	156,798,732	CCNL1	LeptinAdjBMI	Additive / All ancestries / Women	intergenic	G	А	0.391	-0.054	0.008	1.2E-10	29,510
rs791600	7	127,865,816	LEP	LeptinAdjBMI	Additive / All ancestries	intergenic	А	G	0.422	-0.066	0.007	1.1E-23	49,282
rs1121980	16	53,809,247	FTO	Leptin	Additive / European	intronic	А	G	0.432	0.055	0.007	7.7E-17	49,909

The chromosomal positions are based on hg19.

Chr, chromosome; EA, Effect allele; OA, Other allele; EAF, Effect allele frequency; LeptinAdjBMI, leptin adjusted for body mass index
Table 2. Leptin-associated genes identified by gene-based exome-wide association analyses

Gene	Chr	Position	Trait	Most significant model	Method	Ν	P value	Beta	SE	N variants
CNTD1	17	40,950,810-40,963,605	Leptin	Additive / European / Men	SKAT broad	18,882	1.3E-07	0.898	0.165	5
DNAJC18	5	138,743,559-198,780,898	LeptinAdjBMI	Additive / All ancestries / Women	SKAT strict	29,510	5.5E-08	0.757	0.169	2

The chromosomal positions are based on hg19.

Chr, chromosome; LeptinAdjBMI, leptin adjusted for body mass index

Figure 1. Association of the leptin-decreasing alleles of the *LEP* Val94Met (rs17151919) variant (on the left) and the rs10487505 variant near *LEP* (on the right) with BMI standard deviation score (SDS) in the CHOP cohort. The analyses for the Val94Met variant were performed in up to 2,726 African ancestry participants and the analyses for the rs10487505 variant in up to 3,681 African and European ancestry participants of the CHOP cohort. The y-axis reports the effect of each leptin-decreasing allele on BMI at each age year. The error bars indicate 1 standard error of the mean (SEM).

Figure 2. Impact of Val94Met transversion at LEP rs17151919 on leptin secretion rate in HEK293 cells. The rs17151919 variant changes valine to methionine in position 73 of the mature leptin protein. A) The 3D illustration of leptin structure derived from RSCCB Protein Data Bank and modified with UCSF Chimera1.13.1. The prediction of protein stability is derived from the SDM2 server [30]. B) Leptin secretion rates for Val94 and Met94 expressed as the amount of leptin secreted in ng during a 1 hr incubation (72-73 hr post-transfection) (LEPs/hr) normalized by the respective cellular leptin content (LEPc) in untreated control cells at the end of incubation. Individual data points from four separate experiments (each with 2-3 technical replicates) are plotted. The normality of data distribution was examined using D'Agostino & Pearson normality test (p=0.65 and 0.54 for LEPV94 and LEPM94, respectively) and repeated measures oneway ANOVA was performed to assess the difference in secretion rate between the genotypes. Mean ± SD and AVOVA results (F and p values) are reported in the table below the graph. C) Intracellular leptin turnover rates for Val94 and Met94 alleles, obtained by measuring the relative cellular leptin contents in the untreated control cells

1

(defined as 1 for the respective *LEP* variant) and in samples treated with the protein synthesis inhibitor cycloheximide (CHX, 20 μ g/ml) for 0.5 and 1.0 hour. Mean ± SD at each time point from four separate experiments (each with 2-3 technical replicates) are plotted. Paired t-test was used to assess the genotype effect on the fractions of cellular LEP remained after 0.5 hr and 1 hr of CHX treatment (p values are reported in the table below the graph). The average hourly turnover rates for Val94 and Met94 were 61±2%, and 60±3%, respectively, calculated by subtracting the percent cellular LEP remained after one hour of CHX treatment from those of the respective untreated controls (defined as 100%).

Figure 1



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LEPV94

LEPM94

F (1,8)

р

1.62±0.18

1.29±0.09

28.82

0.0007



CHX treatment time (hr)

0.5

● LEPV94

-O- LEPM94

	0.5 hr	1 hr
LEPV94	0.56±0.06	0.39±0.02
LEPM94	0.54±0.03	0.40±0.03
P (V94 vs M94)	0.316	0.366

Table S1. Study design, number of individuals and sample quality control for ExomeChip study cohorts

Study		Study design	Ethnicity	Total	Sample C	QC	Samples	BMI	References
Short name	Full name			sample size (N)	Call rate*	Other exclusions	in analyses (N)	assessment method	
ARIC	Atherosclerosis Risk in Communities Study	Population- based	European American (EA) African American (AA)	462	≥ 95%	 call rate <95%, 2) PCA outliers, 3)sex mismatch , 4) inbreeding coefficient +/-6SD from mean of ancestry distribution, 5) first degree relatedness; comparison with GWAS data, exclude if >40% mismatch, 7) (p10GC) genotype quality score, representing the 10th percentile of the distribution of GenCall scores across all SNPs, 8) missing leptin, adiponectin, or BMI measures (only exclude from analyses missing respective phentoype trait) 	340	Measured	PMID: 2646917 PMID: 2387450 PMCID: PMC3709915 PMID: 12829649
CHS EA and AA	Cardiovascular Health Study	Population- based	European American (EA) African- American (AA)	5088	≥ 95%	Following the central QC and joint variant calling, additional QC steps were applied to the CHS data using PLINK. SNPs with a missingness rate of >95% were removed and individuals meeting the following criteria were excluded from analysis. We further excluded individuals with low P10GC call, a missing genotype rate of > 97%, gender mis-matches identified by X chromosome homozygosity rates. The sample was limited to those of self-described European-ancestry (EA) and African-American (AA) participants. Principal components analysis was performed using a subset of common LD-pruned variants from the Exome Chip both for the full sample as well as in EA and AA strata. Individuals whose full-sample first principal component suggested a different ancestry from their self-reported ancestry were excluded as were individuals who were outliers for the first 10 ancestry-specific principal components. Pairwise IBD measures were calculated and outliers with high levels of IBD were removed.	5044	Measured	PMID: 23874508 PMID: 1669507
CLHNS	Cebu Longitudinal Health and Nutrition Survey	Population Based Longitudinal	Filipino	1799	≥98%	1) Missing study specific covariates (household assets or household income)	1,792	Measured	PMID: 20507864
Ely	Ely study	Longitudinal cohort study	European ancestry	1592	> 98%	 Heterozygosity check, 2) Ethnic outliers, 3) Duplicate individuals, Sex discrepancy, 5) Unusually high number of singleton genotypes, 6) impossible IBD values, 7) phenotype missing 	1,432	Measured	PMID:17257284
ERF study	Erasmus Rucphen Family study	Family-based	White European	2963	≥ 95%		1146	Measured	http://www.eras musmc.nl/klinisch e_genetica/resear ch/intro/genepi/
FAMHS	Family Heart Study	Family-based	White European		≥ 98%	 1) Variants with missing rate > 5% (based on aggregate data) 2) pHWE<1e-6 3) Mendelian errors 4) minor allele count (MAC)<5 for variant-wise tests 	1505	Measured	PMID:8651220
Fenland-CE	Fenland Study	Population- based	European ancestry	1077	> 98%	Heterozygosity check; Ethnic outliers; sex discrepancy; unusually high number of singleton genotypes; impossible IBD values; phenotype missing; excluding overlap exomechip samples	368	Measured	PMID: 20519560

Fenland- Exomechip	Fenland Study	Population- based	White European	1650	> 98%	1) heterozygosity outliers (>3.5 SDs), 2) ethnic outliers, 3) sex discrepancy, 4) unusually high number of singleton genotypes, 5) related (IBD > 0.1875)	1342	Measured	PMID: 20519560
FHS	Framingham Heart Study	Family-based	White European	8153	≥ 97%	1) Missing GWAS PCs, 2) Ethnic outlier, 3) Missing trait or covariate	7458	Measured	PMID: 23874508
FINRISK 1997	Finland National FINRISK Health Survey 1997	Population- based	White European	8325 (4006)	≥ 95%	1) Missing leptin or adiponectine levels, 2) Missing BMI, 3)Pregnancy	3917	Measured	PMID: 29165699
FINRISK 2007	Finland National FINRISK Health Survey 2007	Population- based	White European	6086 (3465)	≥ 95%	1) Missing leptin or adiponectin levels, 2) Missing BMI, height or weight, 3) Missing fat free mass or fat mass, 4) Pregnancy	2945	Measured	PMID: 29158543
HABC AA	Health, aging and body composition study	Population- based	African American ancestry	1139	> 95%	1) missing data, 2) relatedness, 3) acestry outliers, 4) heterozygosity outliers	1060	Measured	
HABC EA	Health, aging and body composition study	Population- based	European ancestry	1663	> 95%	1) missing data, 2) relatedness , 3) acestry outliers, 4) heterozygosity outliers	1572	Measured	
Inter99	Inter99	Population- based	European	6141	≥ 98%	 Missing body weight and height. 2) Heterozygosity were calculated separately for maf < 1% and maf > 1% and samples were dropped judged by plots, 3) Cryptic relatedness (related to 20 or more individuals), 3) Technical duplicates, 4) Non-European population outliers from PCA plot (based on AIM SNPs), 5) Sex discrepancy 	5594	Measured	PMID: 14663300
JHS	Jackson Heart Study	Population- based cohort with subset of families	African American	2803	≥ 95%	1) Missing outcome or covariate, 2) Heterozygosity, 3) PC outlier 4) Half of overlap with ARIC African Americans (coordinated with ARIC)	2312	Measured	PMID: 16320381
KORA	Kooperative Gesundheitsforschung in der Region Augsburg (Cooperative Health Research in the Region of Augsburg)	Population- based	White European	2921	≥98%	 excess heterozygosity [i.e. het_rate > mean+/-5sd]], 2) seccheck based on y-chromosome (remove men with <50% and women with >50% calls on y-chromosome), 3) remove of HAPMAP-samples remove duplicates (keep sample with higher callrate), 5) remove samples with genetic inconsistencies with other genotyping / indication for contamination / population outliers 	2916	Measured	
Leipzig- adults	Leipzig Adults Study	Population- based	White European	902	≥ 99%	1) Missing phenotype, 2) Heterozygosity, 3) Non-European population outliers, 4) Technical duplicates with lower call rate 5) Sex discrepancy	902	Measured	PMID: 20935630
MESA CAU, CHN, AFA and HIS	Multi-Ethnic Study of Atherosclerosis (MESA) Cohort	Population- based	Caucasia n;Chines e;Hispani c;African- American were recruited from six field centers	6375	≥ 95%	1) Ethnic outliers, 2) duplicates, 3) gender mismatch, 4) Phenoty outliers	CAU 2497 AFA 1655 CHN 769 HIS 1435	Measured	
NEO Study	The Netherlands Epidemiology of Obesity Study	Population- based	European ancestry	6.604	≥ 98%	1) remove duplicate/swap samples, 2) remove samples with gender mismatch, 3) remove outliers in PCA	6.127	Measured	PMID: 23576214]

OMICS- Fenland	Fenland Study	Population- based	White European	8994	> 97%	1) Heterozygosity check, 2) Ethnic outliers, 3) sex discrepancy, 4) unusually high number of singleton genotypes, 5) impossible IBD values, 6) phenotype missing, 7) excluding overlap exomechip samples	7845	Measured	PMID: 20519560
PIVUS	Prospective Investigation of the Vasculature in Uppsala Seniors	Population- based	White European	961	≥ 99%	1) Missing phenotype, 2) Heterozygosity, 3) Non-European population outliers, 4) technical duplicates with lower call rate 5) Sex discrepancy	961	Measured	PMID: 16141402
RAINE Study	Western Australian Pregnancy Cohort (RAINE) Study	Population- based	White European	1527	>=95%	 Samepl disconcordance with GWAS data, 2) Heterozygosity Missing body weight and height, 4) Did not participant in DEXA scan 	1006	Measured	
RISC	Relationship between Insulin Sensitivity and Cardiovascular disease	Population- based	White European	313	0.99	1) heterozygosity, 2) duplicates, 3) relatedness	313	Measured	PMID:14968294
RSI	Rotterdam Study	Population- based	White European	3163	≥ 98%	1) Heterozygosity, 2) gender-check	554	Measured	PMID: 29064009
SHIP- TREND	Study of Health in Pomerania - TREND	Population- based	White European	4270	≥ 98%	1) missing data, 2) duplicate samples (by estimated IBD), 3) reported and genotyped sex mismatch, 4) Heterozygosity	4149	Measured	PMID: 20167617
TwinsUK	TwinsUK	twin study	White European	4081	≥ 95%	1) missing phenotype, 2) sample call rate	1864	Measured	
WGHS	Women's Genome Health Study	population based trial	European	22618	>98%	1) Heterozygosity, 2) Batch effects, 3) see also Grove et al. (PLoS One (2013) doi: 10.1371/journal.pone.0068095)	789	Self- reported	PMID: 18070814
WHI	Women's Health Initiative	Cohort	European	21,857	≥ 95%	1) Unexpected Duplicates, 2) PC ancestry outliers, 3) Missing body weight and height	5886	Measured	PMID: 9492970
WHI	Women's Health Initiative	Cohort	African American	3,516	≥ 95%	1) Unexpected Duplicates, 2) PC ancestry outliers, 3) Missing body weight and height	884	Measured	PMID: 9492970
YFS	The Cardiovascular Risk in Young Finns Study	Population- based	White European	1998	≥ 95%	1) Pregnancy, 2) Heterozygosity, 3) Gender discrepancy, 4) MDS outliers	1681	Measured	PMID: 18263651

* Call rate to exclude individuals for whom genotyping success rate is less than a certain percentage (to exclude 'bad' samples/DNA)

**Exome-chip samples from this study

Table S2. Study-specific descriptive statistics of ExomeChip cohorts.

Studya	Trait	Men						Women						
Study	Indit	n	mean	SD	median	min	max	n	mean	SD	median	min	max	
	Age (yrs)	249	53.8	5.7	53	45	65	342	53.2	5.6	53	44	65	
ARIC	BMI (kg/m²)	249	28.7	4.04	28.2	20.4	44.9	342	28.1	5.6	26.8	18.1	49.5	
	Leptin levels (ng/ml)	249	8.4	9.6	5.9	0.5	105.3	342	25.6	22.1	18.7	0.7	147.3	
	Age (yrs)	484	72.9	5.4	72	65	91	533	72.6	4.9	72	65	92	
CHS-EA	BMI (kg/m²)	482	26.4	3.6	26	16.9	39.4	531	26.3	4.9	25.5	15.6	47.7	
	Leptin levels (ng/ml)	484	9.5	10.4	7.2	1.3	100	533	27.2	22.6	19.2	1.4	100	
	Age (yrs)	88	73.6	5.7	73	65	89	121	73.7	5.4	73	66	90	
CHS-AA	BMI (kg/m²)	88	26.4	3.8	26.1	18.2	37.7	121	29.6	5.3	29.3	18.3	44.5	
	Leptin levels (ng/ml)	88	9.6	8.8	7.1	1.3	46.8	121	41.7	26.6	36.1	1.4	100	
	Age (yrs)	-	-	-	-	-	-	1792	48.5	6.1	47.7	35.7	69.3	
CLHNS	BMI (kg/m²)	-	-	-	-	-	-	1780	21.3	4.4	24.1	12.3	42.1	
	Leptin levels (ng/ml)	-	-	-	-	-	-	1792	25.5	19.4	21.3	0	154.2	
	Age (yrs)	742	61.5	9.1	61.6	35.7	77.4	849	60.8	9.3	60.2	36.3	78.9	
Ely	BMI (kg/m²)	742	27.4	3.9	26.8	16	45.8	849	27.3	5.4	26.3	16.9	59.3	
	Leptin levels (ng/ml)	658	9.2	8.1	7.1	0.1	63.1	769	33	26.7	25.7	0.7	198	
	Age (yrs)	262	49.4	14.2	49.6	17.6	81.8	316	50.0	15.4	51.0	18.6	81.4	
ERF study	BMI (kg/m²)	262	27.5	5.0	26.9	17.4	50.8	316	27.1	5.2	26.5	17.7	61.8	
	Leptin levels (ng/ml)	262	27.7	43.8	16.8	0.6	535.9	316	91.3	89.1	60.0	0.0	599.3	
	Age (yrs)	737	52.5	13.9	53.9	25.2	91.0	768	52.8	13.1	53.9	25.2	88.7	
FAMHS EA	BMI (kg/m²)	737	27.8	5.0	27.0	16.0	49.6	768	28.1	6.9	26.4	16.1	55.1	
	Leptin levels (ng/ml)	737	8.5	7.0	6.6	1.1	77.1	768	23.4	17.9	18.4	2.2	123.6	
	Age (yrs)	164	49.6	7.0	50.4	36.1	61.6	204	49.3	7.6	50.1	30.7	62.3	
Fenland-CE	BMI (kg/m²)	164	27.4	4.1	26.9	18.2	42.3	204	26.6	4.8	25.4	19.1	45.5	
	Leptin levels (ng/ml)	164	7.46	7.26	5.70	0.50	57.90	204	23.69	19.80	16.75	2.20	112.00	
Fenland-	Age (yrs)	621	48.5	7.2	48.5	31.3	61.5	713	48.6	7.2	49.0	33.7	61.1	
Exomechip	BMI (kg/m²)	621	27.5	4.0	27.1	18.0	46.6	713	26.6	5.5	25.2	16.6	59.9	

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	1		1	1	1	1	1	1	1		1	1	
	Leptin levels (ng/ml)	621	7.7	7.5	5.9	0.1	74.5	713	24.2	21.3	17.7	0.5	169.0
	Age (yrs)	3035	48.3	7.4	48.6	30.9	62.3	3376	48.4	7.2	48.7	30.5	62.8
Fenland-	BMI (kg/m²)	3035	27.3	4.2	26.8	15.3	50.6	3376	26.4	5.2	25.4	14.5	58.7
OMICS	Leptin levels (ng/ml)	3035	7.7	7.3	5.6	0.1	72.1	3376	23.3	20.3	17.3	0.1	199.0
	Age (yrs)	1800	40.3	8.9	40.0	19.0	72.0	2034	40.0	8.8	40.0	19.0	70.0
FHS	BMI (kg/m²)	1800	27.9	4.7	27.3	16.4	56.5	2030	26.0	6.1	24.4	15.6	60.6
	Leptin levels (ng/ml)	1800	6.1	6.2	4.3	0.2	64.2	2034	18.2	17.1	12.3	0.7	110.3
	Age (yrs)	1786	46.1	13.1	45.3	24.2	74.1	2134.0	44.8	12.4	44.2	24.2	73.8
FINRISK97	BMI (kg/m²)	1783	26.6	3.9	26.1	14.7	47.1	2133.0	26.0	4.9	25.1	16.6	51.6
	Leptin levels (ng/ml)	1761	6.2	6.2	4.3	1.6	76.2	2111.0	18.0	14.0	14.0	1.6	100.0
	Age (yrs)	1298	52.2	13.6	53.0	25.0	74.0	1647.0	51.0	15.5	51.0	25.0	74.0
FINRISK07	BMI (kg/m²)	1284	26.9	4.1	26.3	15.7	62.8	1635.0	26.6	5.4	25.4	15.9	52.7
	Leptin levels (ng/ml)	1284	7.8	8.2	5.3	0.1	89.1	1602.0	19.1	15.8	14.9	0.5	100.0
	Age (yrs)	457	73.5	2.8	73.0	69.0	79.0	603	73.3	2.9	73.0	68.0	80.0
НАВС АА	BMI (kg/m²)	457	27.1	4.2	26.8	14.9	43.2	603	29.4	5.6	29.0	14.6	47.5
	Leptin levels (ng/ml)	457	8.1	7.2	6.4	0.0	60.3	603	24.8	15.0	22.3	0.3	99.3
	Age (yrs)	825	73.9	2.9	74	69	80	747	73.6	2.8	73	69	80
HABC EA	BMI (kg/m²)	825	27	3.7	26.6	17.6	44.2	747	26.1	4.5	25.6	15.6	44.7
	Leptin levels (ng/ml)	825	7.7	6.8	6	0.2	59.1	747	18.9	14	14.8	0.3	86.9
	Age (yrs)	2675	46.6	7.8	45.2	29.9	61.1	2828	45.8	8.0	45.1	29.7	61.3
Inter99	BMI (kg/m²)	2674	26.8	4.0	26.3	17.1	56.9	2825	25.8	5.0	24.7	15.2	55.7
	Leptin levels (ng/ml)	2675	4.6	5.1	3.2	0.2	70.7	2828	15.1	16.1	10.3	0.4	260.6
	Age (yrs)	861	51.9	12.8	51.0	21.0	81.0	1434	53.8	12.6	53.0	21.0	91.0
JHS	BMI (kg/m²)	861	30.4	30.4	29.2	16.4	66.1	1434	31.9	6.2	31.5	16.0	91.8
	Leptin levels (ng/ml)	861	12.0	11.5	8.8	0.8	106.9	1434	36.1	21.5	32.7	1.4	291.0
	Age (yrs)	1415	49.6	13.4	50.0	25.0	74.0	1506	48.4	13.2	48.0	25.0	74.0
KORA	BMI (kg/m²)	1411	27.4	3.8	26.9	16.3	55.1	1491	26.8	5.1	25.9	15.8	51.2
	Leptin levels (ng/ml)	1410	9.4	10.3	6.3	0.0	140.0	1506	27.9	23.6	20.5	0.3	212.0
	Age (yrs)	223	42.3	17.1	40.5	18.0	99.0	276	41.5	16.6	38.0	18.0	89.0
Leipzig-adults	BMI (kg/m²)	223	35.4	12.6	32.6	18.8	120.4	276	36.1	12.6	33.6	14.7	70.0
	Leptin levels (ng/ml)	223	14.3	13.8	10.1	0.2	62.1	276	35.1	23.5	34.1	0.2	142.9

	Age (yrs)	395	62.6	10.2	63.0	45.0	84.0	360	62.9	9.2	62.5	45.0	84.0
MESA CALL	BMI (kg/m²)	395	28.2	4.0	27.6	19.9	41.1	360	27.5	5.7	26.5	16.9	45.7
	Leptin levels (ng/ml)	395	10.4	10.6	7.1	0.2	79.9	360	27.2	23.0	20.7	1.1	156.5
	Age (yrs)	129	62.6	10.7	63.0	45.0	82.0	115	62.3	9.7	61.0	44.0	84.0
MESA CHN	BMI (kg/m²)	129	24.3	2.8	23.9	16.8	32.3	115	24.4	3.2	24.6	17.8	33.0
	Leptin levels (ng/ml)	129	5.8	5.8	3.7	0.4	36.5	115	18.7	16.4	13.2	1.2	113.9
	Age (yrs)	158	61.7	9.7	62	45	83	180	63.6	9.6	64	46	84
MESA AFA	BMI (kg/m²)	158	28.6	4.5	28.3	19	46.9	180	30.3	5.7	29.4	19.7	47.3
	Leptin levels (ng/ml)	158	15.3	17.3	9.3	0.2	150	180	41.6	29.4	37.3	2.8	190.9
	Age (yrs)	246	60.0	9.9	59.0	44.0	82.0	242	62.3	9.2	63.0	45.0	82.0
MESA HIS	BMI (kg/m²)	246	29.0	4.5	28.7	19.4	45.8	242	30.1	5.5	29.6	18.3	52.5
	Leptin levels (ng/ml)	246	11.0	11.0	7.1	0.0	66.8	242	33.8	25.9	27.8	0.9	224.9
	Age (yrs)	2941	56.2	6.0	57.0	44.0	66.0	3186	55.8	5.9	56.0	44.0	66.0
NEO study	BMI (kg/m²)	2941	29.8	3.9	29.3	19.3	54.4	3186	30.3	5.5	29.8	17.2	61.2
	Leptin levels (ng/ml)	2929	12.9	9.2	10.5	0.5	98.6	3172	36.0	23.1	31.9	0.5	262.0
	Age (yrs)	479	70.1	0.2	70.1	69.8	72.3	466	70.3	0.1	70.3	69.9	70.8
PIVUS	BMI (kg/m²)	479	27.0	3.7	26.8	17.7	43.4	466	27.1	4.9	26.5	16.6	49.8
	Leptin levels (ng/ml)	479	8.0	5.6	6.5	1.1	41.8	466	19.4	11.9	17.0	1.7	90.0
	Age (yrs)	467	20.1	0.4	20.0	19.4	22.1	412	20.0	0.4	19.9	18.3	21.9
RAINE Study	BMI (kg/m²)	467	24.5	4.3	23.8	16.9	48.9	412	24.2	5.0	23.0	15.4	46.5
	Leptin levels (ng/ml)	467	6.1	9.9	3.4	0.1	162.1	412	26.2	18.7	21.5	2.2	98.2
	Age (yrs)	156	44.7	8.3	-	-	-	157	45.8	7.9	-	-	-
RISC	BMI (kg/m²)	156	26.0	3.5	26.0	17.9	39.3	157	25.2	4.5	24.3	16.9	42.9
	Leptin levels (ng/ml)	156	5.5	5.6	4.1	0.0	35.7	157	20.9	16.6	16.1	0.9	110.0
	Age (yrs)	273	66.7	7.1	66.1	55.2	88.7	279	69.2	7.6	69.4	55.1	90.8
RSI	BMI (kg/m²)	268	25.7	2.8	25.8	18.4	35.3	272	26.8	4.6	26.0	18.2	59.5
	Leptin levels (ng/ml)	273	5.6	4.5	4.0	0.4	25.2	281	17.9	13.1	15.0	0.7	61.4
	Age (yrs)	410	50.5	14.1	51.0	22.0	80.0	545	50.0	13.3	50.0	20.0	81.0
SHIP-TREND	BMI (kg/m²)	410	28.1	3.7	28.0	19.2	43.9	545	27.0	5.1	26.3	18.5	53.7
	Leptin levels (ng/ml)	410	7.4	5.6	6.2	1.0	43.1	545	21.8	15.7	18.0	1.9	165.0
TwinsUK	Age (yrs)	-	-	-	-	-	-	1015	48.8	11.2	49.1	18.4	73.5

	BMI (kg/m²)	-	-	-	-	-	-	1015	25.2	4.5	24.3	15.1	46.0
	Leptin levels (ng/ml)	-	-	-	-	-	-	1015	16.9	12.0	13.6	1.1	79.4
	Age (yrs)	-	-	-	-	-	-	789	58.8	8.5	58.0	45.0	87.0
WGHS	BMI (kg/m²)	-	-	-	-	-	-	789	25.9	4.7	25.0	14.6	49.9
	Leptin levels (ng/ml)	-	-	-	-	-	-	789	22.8	16.9	19.1	1.4	145.0
	Age (yrs)	-	-	-	-	-	-	1901	68.3	6.4	69.0	50.0	79.0
WHI EA	BMI (kg/m²)	-	-	-	-	-	-	1901	27.7	6.6	26.5	15.7	159.8
	Leptin levels (ng/ml)	-	-	-	-	-	-	1901	20.9	18.6	16.2	0.2	148.8
	Age (yrs)	-	-	-	-	-	-	468	65.5	6.8	66.0	50.0	79.0
WHIAA	BMI (kg/m²)	-	-	-	-	-	-	468	30.2	7.7	29.1	17.2	141.0
	Leptin levels (ng/ml)	-	-	-	-	-	-	468	33.0	20.4	29.1	2.1	117.2
	Age (yrs)	759	32	5	33	24	39	922	32.1	5	33	24	39
YFS	BMI (kg/m²)	755	25.7	4	25.1	15.7	47.8	919	24.4	4.6	23.5	15.7	47.2
	Leptin levels (ng/ml)	759	5.4	4.2	4.3	0.8	32.1	922	15.2	9.7	13	1.5	63.3

* only report descrptives for the individuals included in each of the analyses

CHS NOTE: For age and BMI, I included all individuals who are included in one or more of the analyses

CHS NOTE: For leptin and adiponectin, I included all individuals in the biggest analysis (not adjusted for fat percentage or BMI)

Table S3. Information on genotyping methods, quality control of SNPs, imputation, and statistical analysis for ExomeChip study cohorts

			Principal co	mponents	Inclusio	on criteria			Association analyses		
Cohort	Genotyping Array	Genotype calling algorithm	Software	SNPs used from GWAS/ExomeCHIP/AI MS/Other	MAF	Call rate*	<i>P</i> -value for HWE	SNPs that met QC criteria	Polymorphic SNPs in meta- analysis	Analyses software	
ARIC	Illumina ExomeChip V1.0	GenTrain 2.0 clustering algorithm	Eigensoft v3.0	Exomchip (MAF>5%)	≥ 0%	≥ 95%	> 10 ⁻⁶	237898 **	163,162 (EA)	rvtests	
CHS EA and AA	Illumina ExomeChip V1.0	-	R	ExomeChip	> 0%	≥ 97%	No filter	227061		raremetalworker	
CLHNS	Affymetrix 500K	Birdseed v2	MACH	GWAS/ExomeCHIP	≥ 0%	≥95%	> 10 ⁻⁶	2304702	28,560,246	mach2QTL	
Ely	Illumina HumanCoreExom e	GenCall	PLINK	GWAS	>0%	>95%	> 5x10 ⁻⁶	231349	231349	RAREMETALWORKER	
ERF study	Illumina HumanExome chip v1.1	GenomeStudio v1.9. and zCall			>5%	>95%	> 10 ⁻⁵		240017	rvtests	
FAMHS	Illumina Human Exome 12v1.0 BeadChip	Genome Studio via central CHARGE-S genotyping	EIGENSTR AT	GWAS	≥0%	≥ 98%	> 10 ⁻⁶	237373 **		raremetalworker	
Fenland- CE	Illumina HumanCoreExom e	GenCall	PLINK v1.9beta	GWAS	>0%	>95%	> 10 ⁻⁶	1508325 9	234201	RAREMETALWORKER	
Fenland- Exomechi P	Illumina ExomeChip v1.0	Gencall + zcall	PLINK v1.07	ExomeChip	≥ 0%	>=97%	> 10 ⁻⁶	241979	240859	RAREMETALWORKER	
FHS	Illumina Infinium HumanExome BeadChip v1.0	Illumina issued cluster file HumanExome-12v1.egt + zCall + CHARGE best practices and joint calling	EIGENSOF T	GWAS	≥ 0%	≥ 97%	No filter	237767		raremetalworker	

FINRISK 1997	Illumina HumanHap 610k		PLINK	ExomeCHIP	> 0%	≥ 90%	> 10 ⁻⁶	509376	495420	rvtests
FINRISK 2007	Illumina HumanHap 610k		PLINK	ExomeCHIP	> 0%	≥ 90%	> 10 ⁻⁶	509376	495420	rvtests
НАВС АА	Illumina ExomeChip V1.0	CHARGE protocol	eignestrat	AIMs	≥ 0%	>95%	> 10 ⁻⁶	228554	228554	rvtests
HABC EA	Illumina ExomeChip V1.0	CHARGE protocol	eignestrat	AIMs	≥ 0%	>95%	> 10 ⁻⁶	228565	228565	rvtests
Inter99	Illumina HumanExome- 12v1	GenCall + Zcall	PLINK	AIM SNPs for outlier detection, ExomeCHIP fo adjustment	> 0%	≥ 98%	> 10 ⁻⁴	137187	137187	RMW
JHS	Illumina ExomeChip V1.0	CHARGE joint calling (Illumina GenomeStudio v2011.1 software was utilized with the GenTrain 2.0 clustering algorithm)	Eigenstrat smartpca	Bi-allelic ExomeChip SNPs with MAF > 0.05, HWE p > 0.00001, callrate > 99%, pruned to be pairwise independent with r = 0.3 in plink.	> 0%	≥ 95%	No filter	137716		rvtests
KORA	Illumina ExomeChip V1.0	GeneCall + Zcall (CHARGE Protocol)	genomest udio	ExomeCHIP	> 0%	≥98%	≥10 ⁻⁸	1409	247868	rvtests
Leipzig- adults	Illumina HumanExome- 12v1_A	GeneCall + Zcall (Oxford Protocol)	PLINK	ExomeCHIP MAF>1%	> 0%	≥ 99%	> 10 ⁻⁴	231460		RareMetalWorker
MESA CAU, CHN, AFA, and HIS	Illumina Exome Chip v1.0	Illumina GenomeStudio2011.1	EIGENSTR AT	ExomeCHIP	> 0%	≥ 90%	> 10 ⁻⁶	238876	238876	rvtests
NEO Study	Illumina HumanCoreExom eChip-24V1.0	GeneCall (SOP v5)	PLINK	Based on LD prune	> 0%	≥ 98%	> 10 ⁻⁶	209874	209874	rvtests
OMICS- Fenland	Affymetrix Axiom UKBiobank	Axiom GT1	PLINK v1.9beta	GWAS	> 0%	≥ 95%	> 10 ⁻⁶	719871	58240	RAREMETALWORKE R

PIVUS	Illumina HumanExome- 12v1_A	GeneCall + Zcall (Oxford Protocol)	plink/MDS	AIMS	> 0%	≥ 99%	> 10 ⁻⁴	233149		raremetalworker
RAINE Study	Illuminia HumanExome- 12v1_A	Illumina GenomeStudio GenTrain Clustering algorithm + zCall	EIGENSOF T - smartpca	AIMS	>0%	>=95%	> 10 ⁻⁴	240806	240062	rvtests
RISC	Illumina Human Exome Beadchip v1	GenCall followed by zCall	PLINK	ExomeCHIP	≥ 0%	0.99	> 10 ⁻⁴	236875	236871	RMW
RSI	Illumina ExomeChip V1.1	GeneCall + Zcall (CHARGE Protocol)	PLINK	GWAS	>0%	≥ 90%	> 10 ⁻⁶	237766	109402	rvtests
SHIP- TREND	Illumina ExomeChip V1.0	GeneCall (CHARGE JointCalling Clusterfile)	Illumina GenomeSt udio v2011.1	AIMs	> 0%	≥ 98% (together with SHIP)	> 10 ⁻⁶	238205		raremetalworker
TwinsUK	Illumina12v1-1_A	GeneCall	Plink	GWAS	> 0%	≥ 90%	> 10 ⁻⁶	222804		raremetalworker 4.13.6
WGHS	Illumina HumanExome Beadchip v.1.1A	genomeStudio + zCall	EIGENSTR AT	GWAS	≥ 0%	>95%	> 10 ⁻⁶	235667	234710	raremetal
WHI	Illumina Human Exome BeadChip v1.0	GenomeStudio v2010.3	SNPRelate	ExomeCHIP	> 0%	≥ 90%	> 10 ⁻⁶	246470	246,303	rvtests
YFS	Illumina CoreExome v1.0b	GenCall	PLINK	ExomeCHIP	> 0%	≥ 95%	> 10 ⁻⁶	238194	237,852	rvtest

* Call rate to exclude SNPs for which less than a certain percentage of individuals were successfully genotyped (i.e. to exclude 'bad' SNPs)

** Includes monomorphic SNPs

Table S4. Single-variant results in all statistical models for the leptin-associated loci

SNP	Nearest Gene	EA	OA	EAF	Beta	SE	P value	N
Leptin / Additive / All	ancestries							
rs1121980	FTO	A	G	0.424	0.050	0.006	9.4E-16	56,802
rs2340550	ACTL9	G	A	0.696	-0.005	0.007	4.6E-01	54,433
rs13389219	COBLL1	т	с	0.410	0.048	0.007	1.0E-12	50,386
rs1260326	GCKR	с	т	0.624	0.035	0.006	4.9E-08	56,802
rs900399	CCNL1	G	A	0.389	-0.036	0.007	2.5E-08	50,386
rs3799260	KLHL31	т	с	0.822	-0.023	0.008	3.7E-03	56,802
rs62621812	ZNF800	А	G	0.028	-0.097	0.018	8.0E-08	56,802
rs791600	LEP	A	G	0.422	-0.048	0.007	2.7E-13	49,371
rs17151919	LEP	А	G	0.007	-0.259	0.040	1.3E-10	49,111
rs972283	KLF14	G	A	0.551	-0.038	0.006	6.0E-10	56,802
Leptin / Additive / Eu	ropean							
rs1121980	FTO	А	G	0.432	0.055	0.007	7.7E-17	49,909
rs2340550	ACTL9	G	A	0.685	-0.008	0.007	2.8E-01	48,008
rs13389219	COBLL1	т	с	0.394	0.046	0.007	7.3E-11	43,493
rs1260326	GCKR	С	Т	0.607	0.032	0.007	1.7E-06	49,909
rs900399	CCNL1	G	A	0.396	-0.033	0.007	2.4E-06	43,493
rs3799260	KLHL31	т	с	0.818	-0.024	0.008	3.8E-03	49,909
rs62621812	ZNF800	А	G	0.031	-0.098	0.018	8.2E-08	49,909
rs791600	LEP	А	G	0.411	-0.043	0.007	1.4E-09	42,478
rs17151919	LEP	А	G	0.000	0.134	0.261	6.1E-01	44,474
rs972283	KLF14	G	A	0.521	-0.041	0.006	1.1E-10	49,909
Leptin / Additive / All	ancestries / Men							
rs1121980	FTO	A	G	0.433	0.075	0.009	9.7E-16	23,861
rs2340550	ACTL9	G	A	0.693	-0.027	0.010	7.5E-03	23,861
rs13389219	COBLL1	т	с	0.417	0.059	0.010	9.6E-09	20,822
rs1260326	GCKR	с	т	0.625	0.028	0.010	4.1E-03	23,861
rs900399	CCNL1	G	A	0.387	-0.030	0.010	2.8E-03	20,822
rs3799260	KLHL31	т	с	0.819	-0.007	0.012	5.7E-01	23,861
rs62621812	ZNF800	А	G	0.029	-0.100	0.027	2.6E-04	23,861
rs791600	LEP	А	G	0.406	-0.035	0.010	4.4E-04	20,822

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rs17151919	LEP	A	G	0.006	-0.310	0.066	3.1E-06	22,153
rs972283	KLF14	G	A	0.544	-0.036	0.009	1.3E-04	23,861
Leptin / Additive / Euro	pean / Men							
rs1121980	FTO	A	G	0.433	0.077	0.010	1.8E-15	21,921
rs2340550	ACTL9	G	A	0.684	-0.029	0.010	5.9E-03	21,921
rs13389219	COBLL1	т	с	0.395	0.057	0.011	7.0E-08	18,882
rs1260326	GCKR	с	Т	0.608	0.026	0.010	8.4E-03	21,921
rs900399	CCNL1	G	A	0.395	-0.026	0.011	1.5E-02	18,882
rs3799260	KLHL31	т	с	0.819	-0.007	0.012	5.6E-01	21,921
rs62621812	ZNF800	A	G	0.031	-0.100	0.027	2.7E-04	21,921
rs791600	LEP	A	G	0.410	-0.032	0.010	2.0E-03	18,882
rs17151919	LEP	A	G	0.000	0.155	0.349	6.6E-01	20,213
rs972283	KLF14	G	A	0.522	-0.037	0.010	1.1E-04	21,921
Leptin / Additive / All a	ncestries / Women							
rs1121980	FTO	A	G	0.417	0.035	0.008	1.5E-05	32,940
rs2340550	ACTL9	G	A	0.697	0.010	0.009	2.5E-01	30,571
rs13389219	COBLL1	т	с	0.405	0.040	0.009	5.6E-06	29,563
rs1260326	GCKR	с	Т	0.624	0.043	0.008	2.1E-07	32,940
rs900399	CCNL1	G	A	0.391	-0.049	0.008	6.2E-09	29,563
rs3799260	KLHL31	т	с	0.825	-0.041	0.010	5.6E-05	32,940
rs62621812	ZNF800	A	G	0.027	-0.102	0.024	2.2E-05	32,940
rs791600	LEP	A	G	0.434	-0.060	0.009	2.9E-12	28,548
rs17151919	LEP	A	G	0.007	-0.233	0.049	1.8E-06	26,957
rs972283	KLF14	G	A	0.555	-0.043	0.008	6.5E-08	32,940
Leptin / Additive / Euro	pean / Women							
rs1121980	FTO	A	G	0.431	0.044	0.009	6.2E-07	27,987
rs2340550	ACTL9	G	A	0.685	0.007	0.010	4.6E-01	26,086
rs13389219	COBLL1	т	с	0.392	0.038	0.009	6.1E-05	24,610
rs1260326	GCKR	с	т	0.606	0.040	0.009	6.0E-06	27,987
rs900399	CCNL1	G	A	0.397	-0.048	0.009	2.5E-07	24,610
rs3799260	KLHL31	т	с	0.818	-0.043	0.011	8.9E-05	27,987
rs62621812	ZNF800	A	G	0.032	-0.103	0.024	2.1E-05	27,987
rs791600	LEP	A	G	0.413	-0.054	0.009	8.3E-09	23,595
rs17151919	LEP	A	G	0.000	-0.039	0.380	9.2E-01	24,260

rs972283	KLF14	G	A	0.521	-0.048	0.009	1.6E-08	27,987
Leptin / Recessive / .	All ancestries							
rs1121980	FTO	A	G	0.424	0.071	0.011	1.1E-10	56,802
rs2340550	ACTL9	G	A	0.696	-0.006	0.009	4.6E-01	54,433
rs13389219	COBLL1	т	с	0.410	0.062	0.012	3.2E-07	50,386
rs1260326	GCKR	с	т	0.624	0.046	0.009	2.3E-07	56,802
rs900399	CCNL1	G	A	0.389	-0.042	0.012	6.4E-04	50,386
rs3799260	KLHL31	т	с	0.822	-0.029	0.009	1.5E-03	56,802
rs62621812	ZNF800	A	G	0.021	-0.037	0.124	7.7E-01	56,802
rs791600	LEP	A	G	0.422	-0.074	0.012	6.9E-10	49,371
rs17151919	LEP	A	G	0.007	-0.527	0.190	5.6E-03	49,111
rs972283	KLF14	G	A	0.551	-0.049	0.009	2.5E-07	56,802
Leptin / Recessive /	European							
rs1121980	FTO	A	G	0.432	0.078	0.012	3.2E-11	49,909
rs2340550	ACTL9	G	A	0.685	-0.011	0.009	2.5E-01	48,008
rs13389219	COBLL1	т	с	0.394	0.061	0.013	4.5E-06	43,493
rs1260326	GCKR	с	т	0.607	0.041	0.009	1.2E-05	49,909
rs900399	CCNL1	G	A	0.396	-0.036	0.013	6.6E-03	43,493
rs3799260	KLHL31	т	с	0.818	-0.031	0.010	1.3E-03	49,909
rs62621812	ZNF800	A	G	0.023	-0.037	0.124	7.7E-01	49,909
rs791600	LEP	A	G	0.411	-0.070	0.013	5.8E-08	42,478
rs17151919	LEP	A	G	0.000	NA	NA	NA	44,474
rs972283	KLF14	G	A	0.521	-0.054	0.010	9.5E-08	49,909
Leptin / Recessive / .	All ancestries / Men							
rs1121980	FTO	A	G	0.433	0.096	0.017	8.5E-09	23,861
rs2340550	ACTL9	G	Α	0.693	-0.031	0.013	1.7E-02	23,861
rs13389219	COBLL1	т	с	0.417	0.085	0.019	7.0E-06	20,822
rs1260326	GCKR	с	т	0.625	0.031	0.014	2.4E-02	23,861
rs900399	CCNL1	G	A	0.387	-0.029	0.019	1.3E-01	20,822
rs3799260	KLHL31	т	с	0.819	-0.014	0.014	3.3E-01	23,861
rs62621812	ZNF800	А	G	0.021	-0.067	0.192	7.3E-01	23,861
rs791600	LEP	А	G	0.406	-0.065	0.019	4.6E-04	20,822
rs17151919	LEP	A	G	0.005	-0.725	0.260	5.3E-03	22,153
rs972283	KLF14	G	A	0.544	-0.050	0.015	6.0E-04	23,861

Leptin / Recessive / Eu	uropean / Men							
rs1121980	FTO	A	G	0.433	0.100	0.017	7.6E-09	21,921
rs2340550	ACTL9	G	A	0.684	-0.035	0.014	1.1E-02	21,921
rs13389219	COBLL1	т	с	0.395	0.080	0.020	7.1E-05	18,882
rs1260326	GCKR	с	т	0.608	0.027	0.014	6.0E-02	21,921
rs900399	CCNL1	G	А	0.395	-0.023	0.020	2.6E-01	18,882
rs3799260	KLHL31	Т	с	0.819	-0.015	0.014	2.9E-01	21,921
rs62621812	ZNF800	A	G	0.023	-0.067	0.192	7.3E-01	21,921
rs791600	LEP	A	G	0.410	-0.065	0.019	8.7E-04	18,882
rs17151919	LEP	A	G	0.000	NA	Inf	NA	20,213
rs972283	KLF14	G	А	0.522	-0.052	0.015	5.6E-04	21,921
Leptin / Recessive / A	ll ancestries / Women							
rs1121980	FTO	A	G	0.417	0.058	0.015	6.8E-05	32,940
rs2340550	ACTL9	G	A	0.697	0.011	0.012	3.6E-01	30,571
rs13389219	COBLL1	т	с	0.405	0.052	0.016	9.4E-04	29,563
rs1260326	GCKR	с	т	0.624	0.059	0.012	3.9E-07	32,940
rs900399	CCNL1	G	A	0.391	-0.061	0.016	1.5E-04	29,563
rs3799260	KLHL31	т	с	0.825	-0.047	0.012	8.4E-05	32,940
rs62621812	ZNF800	A	G	0.019	-0.149	0.162	3.6E-01	32,940
rs791600	LEP	A	G	0.434	-0.088	0.016	1.9E-08	28,548
rs17151919	LEP	А	G	0.007	-0.195	0.280	4.9E-01	26,957
rs972283	KLF14	G	A	0.555	-0.055	0.012	8.5E-06	32,940
Leptin / Recessive / Eu	uropean / Women							
rs1121980	FTO	A	G	0.431	0.068	0.016	1.4E-05	27,987
rs2340550	ACTL9	G	A	0.685	0.006	0.013	6.5E-01	26,086
rs13389219	COBLL1	т	с	0.392	0.055	0.018	2.1E-03	24,610
rs1260326	GCKR	с	т	0.606	0.055	0.013	1.3E-05	27,987
rs900399	CCNL1	G	A	0.397	-0.057	0.017	9.9E-04	24,610
rs3799260	KLHL31	Т	с	0.818	-0.049	0.013	1.5E-04	27,987
rs62621812	ZNF800	A	G	0.022	-0.149	0.162	3.6E-01	27,987
rs791600	LEP	A	G	0.413	-0.083	0.017	1.8E-06	23,595
rs17151919	LEP	А	G	0.000	NA	NA	NA	24,260
rs972283	KLF14	G	А	0.521	-0.062	0.014	5.6E-06	27,987
LeptinAdjBMI / Additi	ve / All ancestries							

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rs1121980	FTO	A	G	0.424	0.003	0.006	5.7E-01	56,708
rs2340550	ACTL9	G	A	0.695	-0.014	0.007	3.2E-02	54,339
rs13389219	COBLL1	т	с	0.410	0.053	0.007	3.0E-15	50,297
rs1260326	GCKR	с	т	0.624	0.050	0.006	2.7E-15	56,708
rs900399	CCNL1	G	A	0.389	-0.041	0.007	5.2E-10	50,297
rs3799260	KLHL31	т	с	0.822	-0.036	0.008	4.0E-06	56,708
rs62621812	ZNF800	А	G	0.028	-0.127	0.018	2.0E-12	56,708
rs791600	LEP	А	G	0.422	-0.066	0.007	1.1E-23	49,282
rs17151919	LEP	А	G	0.007	-0.333	0.040	1.5E-16	49,034
rs972283	KLF14	G	A	0.550	-0.053	0.006	6.3E-18	56,708
LeptinAdjBMI / Additive ,	/ European							
rs1121980	FTO	A	G	0.432	0.005	0.007	4.5E-01	49,830
rs2340550	ACTL9	G	A	0.685	-0.016	0.007	2.6E-02	47,929
rs13389219	COBLL1	т	с	0.394	0.053	0.007	1.1E-13	43,419
rs1260326	GCKR	с	т	0.607	0.048	0.007	4.3E-13	49,830
rs900399	CCNL1	G	A	0.396	-0.040	0.007	9.2E-09	43,419
rs3799260	KLHL31	т	с	0.818	-0.038	0.008	3.8E-06	49,830
rs62621812	ZNF800	A	G	0.031	-0.127	0.018	2.8E-12	49,830
rs791600	LEP	A	G	0.411	-0.063	0.007	5.4E-19	42,404
rs17151919	LEP	A	G	0.000	-0.187	0.261	4.7E-01	44,401
rs972283	KLF14	G	A	0.521	-0.056	0.006	3.8E-18	49,830
LeptinAdjBMI / Additive ,	/ All ancestries / Men							
rs1121980	FTO	A	G	0.433	0.028	0.009	2.6E-03	23,822
rs2340550	ACTL9	G	A	0.693	-0.050	0.010	8.5E-07	23,822
rs13389219	COBLL1	т	с	0.417	0.052	0.010	3.8E-07	20,787
rs1260326	GCKR	с	т	0.624	0.043	0.010	8.4E-06	23,822
rs900399	CCNL1	G	A	0.387	-0.036	0.010	4.3E-04	20,787
rs3799260	KLHL31	т	с	0.819	-0.023	0.012	6.0E-02	23,822
rs62621812	ZNF800	A	G	0.029	-0.148	0.027	7.0E-08	23,822
rs791600	LEP	A	G	0.406	-0.054	0.010	6.8E-08	20,787
rs17151919	LEP	A	G	0.006	-0.399	0.066	1.2E-09	22,119
rs972283	KLF14	G	A	0.544	-0.045	0.009	1.8E-06	23,822
LeptinAdjBMI / Additive ,	/ European / Men							
rs1121980	FTO	A	G	0.433	0.026	0.010	6.4E-03	21,883

rs2340550	ACTL9	G	A	0.684	-0.053	0.010	4.1E-07	21,883
rs13389219	COBLL1	т	с	0.395	0.048	0.011	5.2E-06	18,848
rs1260326	GCKR	с	т	0.608	0.042	0.010	2.1E-05	21,883
rs900399	CCNL1	G	A	0.395	-0.033	0.011	2.1E-03	18,848
rs3799260	KLHL31	т	с	0.819	-0.025	0.012	4.3E-02	21,883
rs62621812	ZNF800	A	G	0.031	-0.146	0.028	1.1E-07	21,883
rs791600	LEP	A	G	0.410	-0.049	0.010	2.5E-06	18,848
rs17151919	LEP	A	G	0.000	-0.225	0.352	5.2E-01	20,180
rs972283	KLF14	G	A	0.522	-0.048	0.010	4.8E-07	21,883
LeptinAdjBMI / Additive	/ All ancestries / Women							
rs1121980	FTO	A	G	0.417	-0.013	0.008	1.2E-01	32,886
rs2340550	ACTL9	G	A	0.697	0.007	0.009	4.2E-01	30,517
rs13389219	COBLL1	т	с	0.405	0.052	0.009	2.0E-09	29,510
rs1260326	GCKR	с	т	0.624	0.059	0.008	6.2E-13	32,886
rs900399	CCNL1	G	A	0.391	-0.054	0.008	1.2E-10	29,510
rs3799260	KLHL31	т	с	0.825	-0.055	0.010	1.0E-07	32,886
rs62621812	ZNF800	A	G	0.027	-0.125	0.024	2.2E-07	32,886
rs791600	LEP	A	G	0.434	-0.079	0.009	4.1E-20	28,495
rs17151919	LEP	A	G	0.007	-0.291	0.050	5.7E-09	26,915
rs972283	KLF14	G	A	0.555	-0.063	0.008	3.7E-15	32,886
LeptinAdjBMI / Additive	/ European / Women							
rs1121980	FTO	A	G	0.431	-0.009	0.009	3.2E-01	27,947
rs2340550	ACTL9	G	A	0.685	0.008	0.009	4.1E-01	26,046
rs13389219	COBLL1	т	с	0.392	0.055	0.009	3.8E-09	24,571
rs1260326	GCKR	с	т	0.606	0.057	0.009	9.4E-11	27,947
rs900399	CCNL1	G	A	0.398	-0.058	0.009	3.4E-10	24,571
rs3799260	KLHL31	т	с	0.818	-0.057	0.011	2.2E-07	27,947
rs62621812	ZNF800	A	G	0.032	-0.126	0.024	1.9E-07	27,947
rs791600	LEP	A	G	0.413	-0.080	0.009	2.9E-17	23,556
rs17151919	LEP	A	G	0.000	-0.310	0.375	4.1E-01	24,221
rs972283	KLF14	G	А	0.521	-0.066	0.009	1.3E-14	27,947
LeptinAdjBMI / Recessiv	e / All ancestries	1						
rs1121980	FTO	A	G	0.424	0.000	0.011	9.8E-01	56,708
rs2340550	ACTL9	G	A	0.695	-0.014	0.009	9.8E-02	54,339

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rs13389219	COBLL1	т	с	0.410	0.080	0.012	7.0E-11	50,297
rs1260326	GCKR	с	т	0.624	0.057	0.009	1.8E-10	56,708
rs900399	CCNL1	G	A	0.389	-0.057	0.012	4.3E-06	50,297
rs3799260	KLHL31	т	с	0.822	-0.044	0.009	1.7E-06	56,708
rs62621812	ZNF800	A	G	0.021	-0.145	0.124	2.4E-01	56,708
rs791600	LEP	A	G	0.422	-0.099	0.012	2.4E-16	49,282
rs17151919	LEP	А	G	0.007	-0.795	0.190	2.9E-05	49,034
rs972283	KLF14	G	A	0.550	-0.071	0.009	6.1E-14	56,708
LeptinAdjBMI / Recessive	e / European							
rs1121980	FTO	A	G	0.432	-0.005	0.012	6.8E-01	49,830
rs2340550	ACTL9	G	A	0.685	-0.018	0.009	5.7E-02	47,929
rs13389219	COBLL1	т	с	0.394	0.081	0.013	1.4E-09	43,419
rs1260326	GCKR	с	т	0.607	0.053	0.009	1.9E-08	49,830
rs900399	CCNL1	G	A	0.396	-0.054	0.013	3.8E-05	43,419
rs3799260	KLHL31	т	с	0.818	-0.047	0.010	1.3E-06	49,830
rs62621812	ZNF800	A	G	0.023	-0.145	0.124	2.4E-01	49,830
rs791600	LEP	A	G	0.411	-0.099	0.013	2.7E-14	42,404
rs17151919	LEP	A	G	0.000	NA	Inf	NA	44,401
rs972283	KLF14	G	A	0.521	-0.079	0.010	8.0E-15	49,830
LeptinAdjBMI / Recessive	e / All ancestries / Men							
rs1121980	FTO	A	G	0.433	0.024	0.017	1.6E-01	23,822
rs2340550	ACTL9	G	A	0.693	-0.065	0.013	6.5E-07	23,822
rs13389219	COBLL1	т	с	0.417	0.082	0.019	1.4E-05	20,787
rs1260326	GCKR	с	т	0.624	0.046	0.014	7.9E-04	23,822
rs900399	CCNL1	G	A	0.387	-0.036	0.019	6.2E-02	20,787
rs3799260	KLHL31	т	с	0.819	-0.030	0.014	2.9E-02	23,822
rs62621812	ZNF800	A	G	0.021	-0.293	0.192	1.3E-01	23,822
rs791600	LEP	A	G	0.406	-0.095	0.019	4.2E-07	20,787
rs17151919	LEP	А	G	0.005	-0.942	0.258	2.5E-04	22,119
rs972283	KLF14	G	A	0.544	-0.058	0.015	6.2E-05	23,822
LeptinAdjBMI / Recessive	e / European / Men							
rs1121980	FTO	A	G	0.433	0.021	0.017	2.2E-01	21,883
rs2340550	ACTL9	G	A	0.684	-0.071	0.014	2.0E-07	21,883
rs13389219	COBLL1	т	С	0.395	0.072	0.020	3.3E-04	18,848

rs1260326	GCKR	с	т	0.608	0.045	0.014	1.6E-03	21,883
rs900399	CCNL1	G	A	0.395	-0.032	0.020	1.1E-01	18,848
rs3799260	KLHL31	т	с	0.819	-0.034	0.015	2.0E-02	21,883
rs62621812	ZNF800	A	G	0.023	-0.293	0.192	1.3E-01	21,883
rs791600	LEP	A	G	0.410	-0.092	0.019	2.1E-06	18,848
rs17151919	LEP	A	G	0.000	NA	NA	NA	20,180
rs972283	KLF14	G	A	0.522	-0.065	0.015	2.0E-05	21,883
LeptinAdjBMI / Recessiv	e / All ancestries / Women							
rs1121980	FTO	A	G	0.417	-0.016	0.015	2.6E-01	32,886
rs2340550	ACTL9	G	A	0.697	0.016	0.012	1.8E-01	30,517
rs13389219	COBLL1	т	с	0.405	0.080	0.016	4.3E-07	29,510
rs1260326	GCKR	с	т	0.624	0.068	0.012	4.4E-09	32,886
rs900399	CCNL1	G	A	0.391	-0.083	0.016	2.3E-07	29,510
rs3799260	KLHL31	т	с	0.825	-0.063	0.012	1.3E-07	32,886
rs62621812	ZNF800	A	G	0.019	-0.241	0.162	1.4E-01	32,886
rs791600	LEP	A	G	0.434	-0.112	0.016	5.7E-13	28,495
rs17151919	LEP	A	G	0.007	-0.570	0.284	4.5E-02	26,915
rs972283	KLF14	G	A	0.555	-0.086	0.012	2.5E-12	32,886
LeptinAdjBMI / Recessiv	e / All ancestries / Women							
rs1121980	FTO	A	G	0.431	-0.022	0.015	1.5E-01	27,947
rs2340550	ACTL9	G	A	0.685	0.015	0.012	2.3E-01	26,046
rs13389219	COBLL1	т	с	0.392	0.093	0.018	1.7E-07	24,571
rs1260326	GCKR	с	т	0.606	0.062	0.013	7.0E-07	27,947
rs900399	CCNL1	G	A	0.398	-0.085	0.017	9.2E-07	24,571
rs3799260	KLHL31	т	с	0.818	-0.066	0.013	2.2E-07	27,947
rs62621812	ZNF800	A	G	0.022	-0.241	0.162	1.4E-01	27,947
rs791600	LEP	A	G	0.413	-0.118	0.017	1.4E-11	23,556
rs17151919	LEP	A	G	0.000	NA	NA	NA	24,221
rs972283	KLF14	G	А	0.521	-0.096	0.014	1.4E-12	27,947

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SNP	Chr	Position	Gene	Meta-analysis	Annotation	EA	OA	Beta	Beta	SE	SE	Р	Р	N	N
								AdjBMI		AdjBMI		AdjBMI		AdjBMI	
Novel loci															
rs3799260	6	53519605	KLHL31	Additive / All ancestries /Women	missense	C	Т	0.055	0.041	0.010	0.010	1.0E-07	5.6E-05	32,886	32,940
rs62621812	7	127015083	ZNF800	Additive / All ancestries	missense	G	A	0.127	0.097	0.018	0.018	2.0E-12	8.0E-08	56,708	56,802
rs17151919	7	127894592	LEP	Additive / All ancestries	missense	G	A	0.333	0.259	0.040	0.040	1.5E-16	1.1E-10	49,034	49,111
rs972283	7	130466854	KLF14	Additive / European	intergenic	A	G	0.056	0.041	0.006	0.006	3.8E-18	1.1E-10	49,830	49,909
rs2340550	19	8808942	ACTL9	Recessive / European / Men	missense	A	G	0.071	0.035	0.014	0.014	2.0E-07	1.1E-02	21,883	21,921
Previously ide	ntified lo	ci													
rs1260326	2	27730940	GCKR	Additive / All ancestries	missense	C	Т	0.050	0.035	0.006	0.006	2.7E-15	4.9E-08	56,708	56,802
rs13389219	2	165528876	COBLL1	Additive / All ancestries	intronic	Т	С	0.053	0.048	0.007	0.007	3.0E-15	1.0E-12	50,297	50,386
rs900399	3	156798732	CCNL1	Additive / All ancestries /Women	intergenic	A	G	0.054	0.049	0.008	0.008	1.2E-10	6.2E-09	29,510	29,563
rs791600	7	127865816	LEP	Additive / All ancestries	intergenic	G	A	0.066	0.048	0.007	0.007	1.1E-23	2.7E-13	49,282	49,371
rs1121980	16	53809247	FTO	Additive / European	intronic	A	G	0.005	0.055	0.007	0.007	4.5E-01	7.7E-17	49,830	49,909

The chromosomal positions are based on hg19.

Chr, chromosome; EA, Effect allele; OA, Other allele; EAF, Effect allele frequency; LeptinAdjBMI, leptin adjusted for body mass index

Locus	MarkerName	EA	EAF	xL	pL	xLadjB	pLadjB	xLadjBa	хВ	рВ
FTO	rs1121980	A	0.4316428	0.05486291	7.71E-17	0.004952214	4.47E-01	0.04153	0.07481	6.70E-225
ACTL9*	rs2340550	G	0.6846579	-0.007649348	2.79E-01	-0.01562951	2.62E-02	-0.01394	0.00345	1.51E-01
COBLL1	rs13389219	Т	0.3938886	0.04618875	7.30E-11	0.05254237	1.13E-13	0.05871	0.01261	8.16E-08
GCKR	rs1260326	C	0.6070448	0.03182518	1.71E-06	0.04773787	4.32E-13	0.04993	0.00449	5.24E-02
CCNL1	rs900399	G	0.3961604	-0.03304733	2.43E-06	-0.04024503	9.25E-09	-0.04198	-0.00355	1.24E-01
KLHL31*	rs3799260	Т	0.8183081	-0.02397769	3.79E-03	-0.03820487	3.83E-06	-0.03499	0.00657	1.71E-02
ZNF800	rs62621812	A	0.03142557	-0.09769461	8.18E-08	-0.1273454	2.80E-12	-0.11685	0.02147	1.24E-03
LEP	rs791600	A	0.4110841	-0.04264698	1.36E-09	-0.06262022	5.35E-19	-0.06034	0.00466	4.54E-02
LEP*	rs17151919	A	0.000166917	0.1342779	6.07E-01	-0.1868478	4.73E-01	-0.18299	0.00789	9.20E-01
KLF14	rs972283	G	0.5211696	-0.04137304	1.12E-10	-0.05554037	3.84E-18	-0.05942	-0.00793	2.68E-04

Table S6. Examination of collider bias with BMI among the exome-array significant loci associated with leptin adjusted for BMI

xL, Effect size for leptin

pL, P value for leptin

xLadjB, Effect size for leptin adjusted for BMI

pLadjB, P value for leptin adjusted for BMI

xLadjBa, Corrected effect size for leptin adjusted for BMI

xB, Effect size for BMI

pB, P value for BMI

* The ACTL9, KLHL31, and LEP rs17151919 loci reached array-wide significance (P<2x10⁻⁷) in meta-analyses of European-ancestry men (recessive model), all-ancestry women, and Africanancestry men and women combined, respectively. The results shown are from meta-analyses of European ancestry individuals only.

	1	1	1	1	1	1	1	1	1	1		1	
Ancestry	Trait	Chr:Position	EA	OA	N	EAF	N _{GG}	N _{GA+AG}	N _{AA}	beta	se	Pvalue	12
All	LeptinAdjBMI	7:127894592	A (Met94)	G (Val94)	49034	0.0067	40075	609	28	-0.333	0.040	1.53E-16	76%
European	LeptinAdjBMI	7:127894592	A (Met94)	G (Val94)	44401	0.0002	36065	15	0	-0.187	0.261	4.73E-01	0%
African	LeptinAdjBMI	7:127894592	A (Met94)	G (Val94)	3901	0.0800	3302	571	27	-0.343	0.042	2.40E-16	94%
Hispanic	LeptinAdjBMI	7:127894592	A (Met94)	G (Val94)	488	0.0221	464	23	1	-0.209	NA	2.85E-01	NA
East Asian	LeptinAdjBMI	7:127894592	A (Met94)	G (Val94)	244	NA	NA	NA	NA	NA	NA	NA	NA

Table S7. Ancestry-specific results for the Val94Met (rs17151919) missense variant in LEP

EA, effect allele; OA, other allele; EAF, effect allele frequency

Gene	Method	N	P value	beta	se	N variants
Leptin / Additive / All ar	ncestries					
CNTD1	SKAT broad	49,597	9.1E-04	0.350	0.094	6
CNTD1	SKAT strict	48,582	7.0E-02	1.043	0.330	1
CNTD1	VT broad	49,597	3.9E-06	0.746	0.149	4
CNTD1	VT strict	48,582	7.0E-02	1.043	0.330	1
DNAJC18	SKAT broad	56,013	2.2E-02	0.062	0.057	7
DNAJC18	SKAT strict	49,597	5.1E-05	0.466	0.135	2
DNAJC18	VT broad	56,013	4.3E-03	0.323	0.096	5
DNAJC18	VT strict	49,597	1.1E-03	0.466	0.135	2
Leptin / Additive / Euro	pean					
CNTD1	SKAT broad	42,704	1.4E-05	0.580	0.126	5
CNTD1	SKAT strict	NA	NA	NA	NA	NA
CNTD1	VT broad	42,704	1.1E-05	0.720	0.153	4
CNTD1	VT strict	NA	NA	NA	NA	NA
DNAJC18	SKAT broad	49,120	3.1E-02	0.045	0.060	7
DNAJC18	SKAT strict	42,704	5.3E-05	0.478	0.140	2
DNAJC18	VT broad	49,120	8.4E-03	0.360	0.112	5
DNAJC18	VT strict	42,704	1.3E-03	0.478	0.140	2
Leptin / Additive / All ar	ncestries / Men					
CNTD1	SKAT broad	20,822	2.0E-05	0.580	0.137	5
CNTD1	SKAT strict	NA	NA	NA	NA	NA
CNTD1	VT broad	20,822	6.4E-06	1.026	0.209	3
CNTD1	VT strict	NA	NA	NA	NA	NA
DNAJC18	SKAT broad	23,861	5.3E-01	-0.061	0.086	6
DNAJC18	SKAT strict	20,822	2.0E-01	0.034	0.223	2
DNAJC18	VT broad	23,861	3.4E-01	-0.569	0.360	2
DNAJC18	VT strict	20,822	1.3E-01	-0.896	0.499	1
Leptin / Additive / Euro	pean / Men					
CNTD1	SKAT broad	18,882	1.3E-07	0.898	0.165	5
CNTD1	SKAT strict		NA NA		NA	NA
CNTD1	VT broad	18,882	1.4E-07	0.898	0.165	5
CNTD1	VT strict	NA	NA	NA	NA	NA

Table S8. Gene-based results in all statistical models for leptin-associated genes

DNAJC18	SKAT broad	21,921	5.3E-01	-0.059	0.087	6
DNAJC18	SKAT strict	18,882	2.0E-01	0.034	0.223	2
DNAJC18	VT broad	21,921	6.4E-01	-0.566	0.446	2
DNAJC18	VT strict	18,882	1.3E-01	-0.896	0.499	1
Leptin / Additive / All ances	stries / Women					
CNTD1	SKAT broad	29,563	5.5E-01	0.178	0.123	6
CNTD1	SKAT strict	28,548	7.3E-02	0.981	0.297	1
CNTD1	VT broad	29,563	3.2E-02	0.553	0.211	4
CNTD1	VT strict	28,548	7.3E-02	0.981	0.297	1
DNAJC18	SKAT broad	32,940	1.3E-02	0.151	0.075	7
DNAJC18	SKAT strict	29,563	1.9E-05	0.717	0.166	2
DNAJC18	VT broad	32,940	5.8E-04	0.452	0.117	5
DNAJC18	VT strict	29,563	3.3E-05	0.717	0.166	2
Leptin / Additive / Europea	n / Women					
CNTD1	SKAT broad	24,610	2.7E-01	0.233	0.188	5
CNTD1	SKAT strict	NA	NA	NA	NA	NA
CNTD1	VT broad	24,610	1.3E-01	0.478	0.240	4
CNTD1	VT strict	NA	NA	NA	NA	NA
DNAJC18	SKAT broad	27,987	1.9E-02	0.132	0.081	6
DNAJC18	SKAT strict	24,610	1.3E-05	0.767	0.177	2
DNAJC18	VT broad	27,987	8.1E-04	0.557	0.146	4
DNAJC18	VT strict	24,610	2.8E-05	0.767	0.177	2
LeptinAdjBMI / Additive / A	All ancestries					
CNTD1	SKAT broad	49,508	4.6E-02	0.242	0.093	6
CNTD1	SKAT strict	48,493	9.1E-02	0.969	0.330	1
CNTD1	VT broad	49,508	9.0E-04	0.560	0.149	4
CNTD1	VT strict	48,493	9.1E-02	0.969	0.330	1
DNAJC18	SKAT broad	55,919	4.3E-03	0.083	0.057	7
DNAJC18	SKAT strict	49,508	1.2E-07	0.485	0.136	2
DNAJC18	VT broad	55,919	1.8E-02	0.279	0.096	5
DNAJC18	VT strict	49,508	7.1E-04	0.485	0.136	2
LeptinAdjBMI / Additive / E	uropean					
CNTD1	SKAT broad	42,630	3.8E-03	0.430	0.126	5
CNTD1	SKAT strict	NA	NA	NA	NA	NA
CNTD1	VT broad	42,630	2.0E-03	0.525	0.153	4

CNTD1	VT strict	NA	NA	NA	NA	NA
DNAJC18	SKAT broad	49,041	8.4E-03	0.063	0.060	7
DNAJC18	SKAT strict	42,630	2.3E-07	0.474	0.141	2
DNAJC18	VT broad	49,041	6.4E-02	0.286	0.113	5
DNAJC18	VT strict	42,630	1.6E-03	0.474	0.141	2
LeptinAdjBMI / Additive /	All ancestries / Men					
CNTD1	SKAT broad	20,787	7.1E-02	0.313	0.138	5
CNTD1	SKAT strict	NA	NA	NA	NA	NA
CNTD1	VT broad	20,787	1.5E-02	0.606	0.210	3
CNTD1	VT strict	NA	NA	NA	NA	NA
DNAJC18	SKAT broad	23,822	2.6E-01	-0.124	0.086	6
DNAJC18	VT broad	23,822	1.6E-01	-0.713	0.359	2
DNAJC18	SKAT strict	20,787	9.6E-02	0.036	0.223	2
DNAJC18	VT strict	20,787	4.2E-02	-1.138	0.498	1
LeptinAdjBMI / Additive /	European / Men					
CNTD1	SKAT broad	18,848	7.4E-03	0.565	0.165	5
CNTD1	SKAT strict	NA	NA	NA	NA	NA
CNTD1	VT broad	18,848	2.6E-03	0.565	0.165	5
CNTD1	VT strict	NA	NA	NA	NA	NA
DNAJC18	SKAT broad	21,883	2.6E-01	-0.109	0.087	6
DNAJC18	SKAT strict	18,848	9.6E-02	0.036	0.223	2
DNAJC18	VT broad	21,883	2.5E-01	-0.838	0.446	2
DNAJC18	VT strict	18,848	4.2E-02	-1.138	0.498	1
LeptinAdjBMI / Additive /	All ancestries / Women					
CNTD1	SKAT broad	29,510	3.0E-01	0.238	0.124	6
CNTD1	SKAT strict	28,495	7.7E-02	0.972	0.305	1
CNTD1	VT broad	29,510	1.3E-02	0.620	0.212	4
CNTD1	VT strict	28,495	7.7E-02	0.972	0.305	1
DNAJC18	SKAT broad	32,886	7.6E-04	0.234	0.075	7
DNAJC18	SKAT strict	29,510	5.5E-08	0.757	0.169	2
DNAJC18	VT broad	32,886	4.4E-04	0.460	0.118	5
DNAJC18	VT strict	29,510	1.5E-05	0.757	0.169	2
LeptinAdjBMI / Additive /	European / Women					
CNTD1	SKAT broad	24,571	1.7E-01	0.373	0.186	5
CNTD1	SKAT strict	NA	NA	NA	NA	NA

CNTD1	VT broad	24,571	6.3E-02	0.554	0.239	4
CNTD1	VT strict	NA	NA	NA	NA	NA
DNAJC18	SKAT broad	27,947	2.4E-03	0.207	0.081	6
DNAJC18	SKAT strict	24,571	7.9E-08	0.774	0.179	2
DNAJC18	VT broad	27,947	4.1E-03	0.496	0.147	4
DNAJC18	VT strict	24,571	3.2E-05	0.774	0.179	2

Age Bin	N	Allele freq.	Beta	SE	Р
2	2726	0.089	0.079	0.055	0.153
3	2570	0.089	0.123	0.056	0.029
4	2572	0.093	0.160	0.054	0.003
5	2381	0.089	0.154	0.060	0.010
6	2030	0.091	0.204	0.066	0.002
7	1769	0.092	0.143	0.070	0.041
8	1583	0.092	0.029	0.074	0.694
9	1476	0.099	0.017	0.078	0.824
10	1446	0.095	0.017	0.080	0.832
11	1500	0.095	-0.004	0.079	0.964
12	1455	0.096	-0.036	0.075	0.631
13	1460	0.101	-0.007	0.075	0.928
14	1417	0.104	0.004	0.074	0.959
15	1355	0.099	0.048	0.077	0.537
16	1287	0.093	-0.006	0.081	0.937
17	1098	0.102	0.055	0.087	0.527
18	451	0.085	-0.009	0.135	0.946

Table S9. Association of the leptin-decreasing Met94 allele of *LEP* Val94Met (rs1715919) with BMI z-score in African-ancestry children from the CHOP cohort.

Age Bin	N	Allele freq.	Beta	SE	Р
2	3681	0.462	0.033	0.026	0.203
3	3618	0.467	0.026	0.026	0.334
4	3681	0.469	0.058	0.026	0.027
5	3557	0.471	-0.002	0.027	0.929
6	3166	0.473	-0.044	0.029	0.132
7	2869	0.469	-0.006	0.031	0.835
8	2711	0.465	-0.021	0.032	0.504
9	2571	0.465	-0.035	0.033	0.290
10	2608	0.468	-0.033	0.033	0.317
11	2705	0.462	-0.028	0.032	0.380
12	2685	0.454	-0.021	0.032	0.502
13	2697	0.459	0.004	0.031	0.898
14	2679	0.454	-0.027	0.032	0.389
15	2604	0.451	-0.009	0.031	0.777
16	2463	0.458	0.012	0.033	0.719
17	17 2130		-0.004	0.036	0.917
18	663	0.456	-0.003	0.062	0.959

 Table S10. Association of the leptin-decreasing C allele of rs10487505 near LEP with BMI z-score in a metaanalysis of African-ancestry and European ancestry children from the CHOP cohort.

ΤοοΙ	Protein (PDB-ID)	WT/MT	Chain	Overall stability	Predicted ΔΔG
CUPSAT	LEP (1AX8)	VAL/MET	A	Decreased	-0.22
I-Mutant v2.0	LEP (1AX8)	VAL/MET	A	Decreased	
SDM	LEP (1AX8)	VAL/MET	A	Decreased	-0.72

Table S11. Predicted change in leptin protein stability upon the Val94Met change (Val73Met in the mature leptin protein) in the amino acid sequence

Table S12. Colocalization of METSIM subcutaneous adipose tissue eQTLs at GWAS loci for leptin

						GWAS va level	GWAS variant association with expression level				sociation	with expre	ession level					
SNP	Chr	Position	MAF	Probeset	Allele 1 / EA	Allele 2	eQTL gene	Beta ^{initial}	P _{initial}	Beta conditional	P conditional	Lead eSNP	Allele 1/ Allele 2	Beta ^{initial}	P _{initial}	Beta conditional	P conditional	LD r ²
rs62621812	7	127,015,083	0.06	11736419_a_at	G	А	ZNF800	-0.871	2.40E-16	0.000	3.18E-01	rs62621812	A/G	0.871	2.40E-16	0.000	3.2E-01	1.00
rs972283	7	130,466,854	0.45	11737563_at	А	G	KLF14	0.233	4.14E-06	-0.322	4.46E-01	rs6467315	G/C	-0.238	2.26E-06	-0.552	1.9E-01	0.98
rs1260326	2	27,730,940	0.36	11729870_x_at	с	т	EMILIN1	-0.230	9.22E-06	0.166	5.23E-01	rs780094	C/T	-0.240	3.33E-06	-0.407	1.1E-01	0.96
rs900399	3	156,798,732	0.32	11717399_a_at	A	G	TIPARP	-0.905	2.99E-72	-0.213	1.57E-01	rs13322435	G/A	0.922	9.57E-77	0.715	2.0E-06	0.91

LD r2 calculated using 770 METSIM samples (Finnish males) included in eQTL data

A1 (column E) is the leptin raising allele from the Exome Chip analysis. A1 is also the effect allele for the effect sizes listed in columns H and J. Allele 1 in column O is the effect allele for the effect in columns O/Q. FDR<1% (P < 2.37 x 10⁻⁴)

Table S13. PASCAL gene set enrichment analysis results for leptin unadjusted for BMI using coding variants only.

(A) Leptin not adjusted for BMI, European, additive model, sex-combined analysis. Coding variants included. SUM method used (Bonferroni correction for 1000 gene sets and 2 traits: P<2.5E-05 for both chi2Pvalue and empPvalue)

Name	chi2Pvalue	empPvalue	Annotation
GO:2000243	1.30E-04	8.80E-05	positive regulation of reproductive process
MP:0005501	3.40E-04	0.000284	abnormal skin physiology
ENSG0000204713	4.31E-04	0.000389	TRIM27 PPI subnetwork
ENSG00000112448	4.31E-04	0.000397	ENSG0000112448 PPI subnetwork
ENSG00000215641	4.31E-04	0.000404	TRIM27 PPI subnetwork
GO:0032769	4.85E-04	0.000335	negative regulation of monooxygenase activity
MP:0002769	5.05E-04	0.000492	abnormal vas deferens morphology
ENSG0000008853	8.38E-04	0.000432	RHOBTB2 PPI subnetwork
ENSG0000081019	9.58E-04	0.00058	RSBN1 PPI subnetwork
GO:0072527	1.37E-03	9.70E-04	pyrimidine-containing compound metabolic process
ENSG00000143344	1.56E-03	0.00076	RGL1 PPI subnetwork

(B) Leptin not adjusted for BMI, European, additive model, sex-combined analysis. Coding variants included. MAX method used (Bonferroni correction for 1000 gene sets and 2 traits: P<2.5E-05 for both chi2Pvalue and empPvalue)

Name	chi2Pvalue	empPvalue	Annotation
GO:2000243	1.53E-05	1.59E-05	positive regulation of reproductive process
ENSG00000143344	4.85E-04	1.56E-04	RGL1 PPI subnetwork
ENSG0000215641	1.95E-04	1.69E-04	TRIM27 PPI subnetwork
ENSG0000204713	1.95E-04	1.94E-04	TRIM27 PPI subnetwork
ENSG00000112448	1.95E-04	1.96E-04	ENSG00000112448 PPI subnetwork
MP:0002769	2.23E-04	2.33E-04	abnormal vas deferens morphology
GO:0032769	2.57E-04	2.93E-04	negative regulation of monooxygenase activity
ENSG0000074211	3.57E-04	4.03E-04	PPP2R2C PPI subnetwork
ENSG0000008853	1.30E-03	4.40E-04	RHOBTB2 PPI subnetwork
ENSG00000169682	6.59E-04	5.50E-04	SPNS1 PPI subnetwork
ENSG0000081019	8.20E-04	5.60E-04	RSBN1 PPI subnetwork
GO:0004715	7.77E-04	5.70E-04	non-membrane spanning protein tyrosine kinase activity
GO:0010458	7.32E-04	6.00E-04	exit from mitosis
ENSG0000090054	2.10E-03	9.00E-04	SPTLC1 PPI subnetwork
ENSG00000113578	6.06E-04	9.50E-04	FGF1 PPI subnetwork
MP:0008347	1.09E-03	9.70E-04	decreased gamma-delta T cell number

Table S14. PASCAL gene set enrichment analysis for leptin adjusted for BMI using coding variants only.

Name	chi2Pvalue	empPvalue	Pathway/Gene-set
ENSG0000175575	3.69E-05	7.90F-06	TRIM39PPI subnetwork
ENSG0000204599	3.69E-05	7.90E-06	PAAF1 PPI subnetwork
ENSG0000206495	3.69E-05	8.80F-06	FNSG00000206419 PPI subnetwork
ENSG0000206419	3.69E-05	1.03E-05	ENSG0000105972 PPI subnetwork
ENSG00000105972	7.14F-05	1.13E-05	mitochondrial large ribosomal subunit
GO:0005762	1.62E-04	2.66E-05	organellar large ribosomal subunit
GO:0000315	1.62E-04	2.76E-05	KLF1 PPI subnetwork
ENSG0000105610	1.96E-04	2.78E-05	negative regulation of monooxygenase activity
GO:0032769	1.58E-04	5.20E-05	BCI 10 PPI subnetwork
ENSG0000142867	6.84E-05	6.10E-05	CHD2 PPI subnetwork
ENSG00000173575	1.20E-04	6.20E-05	UBE38 PPI subnetwork
ENSG0000151148	6.91E-05	6.70E-05	abnormal skin physiology
MP:0005501	8.97E-05	6.70E-05	CCDC33 PPI subnetwork
ENSG0000140481	3.61E-04	9.70E-05	abnormal cell migration
ENSG0000198925	1.53E-04	2.13E-04	HSPA12A PPI subnetwork
MP:0003091	2.13E-04	1.59E-04	SV2A PPI subnetwork
ENSG0000159164	5.38E-04	1.61E-04	MTHFD1L PPI subnetwork
ENSG0000120254	4.09E-04	1.75E-04	REACTOME_REGULATION_OF_ACTIVATED_PAK:2P34_BY_PROTEASOME_ MEDIATED_DEGRADATION
REACTOME_REGULATION_OF_ACTIVATED_PAK:2P34_BY_PROTEASOME_ MEDIATED_DEGRADATION	3.32E-04	1.79E-04	ATG9A PPI subnetwork
ENSG0000165868	7.61E-04	2.16E-04	ENO2 PPI subnetwork
ENSG0000178363	2.21E-04	4.50E-04	EEF1A2 PPI subnetwork
ENSG0000111674	8.97E-04	2.31E-04	RHOBTB2 PPI subnetwork
ENSG0000008853	6.23E-04	2.49E-04	exit from mitosis
GO:0010458	5.32E-04	2.67E-04	ZNF462 PPI subnetwork
ENSG0000148143	8.18E-04	2.78E-04	TOP2B PPI subnetwork
ENSG0000077097	5.67E-04	2.82E-04	RSBN1 PPI subnetwork
ENSG0000081019	6.96E-04	3.13E-04	HLA-G PPI subnetwork
ENSG0000204632	1.21E-03	3.16E-04	ENSG0000206443 PPI subnetwork
ENSG0000206443	1.21E-03	3.21E-04	HLA-G PPI subnetwork
ENSG0000206506	1.21E-03	3.22E-04	acanthosis
MP:0001874	3.68E-04	3.41E-04	REACTOME_AUTODEGRADATION_OF_CDH1_BY_CDH1APCC
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REACTOME_AUTODEGRADATION_OF_CDH1_BY_CDH1APCC	5.20E-04	3.43E-04	SBF1 PPI subnetwork
ENSG0000100241	9.96E-04	3.67E-04	ENSG0000206413 PPI subnetwork
ENSG0000206413	1.46E-03	3.75E-04	NIPSNAP1 PPI subnetwork
ENSG0000184117	1.14E-03	3.78E-04	abnormal CD4-positive T cell differentiation
MP:0008076	7.62E-04	3.81E-04	HLA-E PPI subnetwork
ENSG0000206493	1.46E-03	3.85E-04	REACTOME_P53:INDEPENDENT_G1S_DNA_DAMAGE_CHECKPOINT
REACTOME_P53:INDEPENDENT_G1S_DNA_DAMAGE_CHECKPOINT	8.27E-04	4.13E-04	ZNF317 PPI subnetwork
ENSG0000130803	5.25E-04	4.35E-04	REACTOME_P53:INDEPENDENT_DNA_DAMAGE_RESPONSE
REACTOME_P53:INDEPENDENT_DNA_DAMAGE_RESPONSE	8.27E-04	4.40E-04	PDE1A PPI subnetwork
ENSG00000198838	4.45E-04	5.70E-04	TOMM34 PPI subnetwork
ENSG00000115252	1.34E-03	4.46E-04	FNBP1 PPI subnetwork
ENSG0000187239	1.01E-03	4.47E-04	CALML3 PPI subnetwork
ENSG0000101210	5.88E-04	4.50E-04	REACTOME_MYD88_DEPENDENT_CASCADE_INITIATED_ON_ENDOSOME
REACTOME_MYD88_DEPENDENT_CASCADE_INITIATED_ON_ENDOSOME	8.42E-04	4.55E-04	REACTOME_UBIQUITIN_MEDIATED_DEGRADATION_OF_PHOSPHORYLATE D_CDC25A
REACTOME_UBIQUITIN_MEDIATED_DEGRADATION_OF_PHOSPHORYLATE D_CDC25A	8.27E-04	4.62E-04	cellular defense response
GO:0006968	9.97E-04	4.76E-04	macrolide binding
GO:0005527	6.63E-04	4.91E-04	REACTOME_TOLL_LIKE_RECEPTOR_78_TLR78_CASCADE
REACTOME_TOLL_LIKE_RECEPTOR_78_TLR78_CASCADE	8.42E-04	5.10E-04	RPN1 PPI subnetwork
ENSG00000163902	1.62E-03	5.10E-04	FK506 binding
GO:0005528	6.63E-04	5.20E-04	ARID5B PPI subnetwork
ENSG0000150347	7.60E-04	5.20E-04	SEC31A PPI subnetwork
ENSG00000138674	1.23E-03	5.20E-04	SEPT3 PPI subnetwork
ENSG0000100167	1.37E-03	5.20E-04	ROGDI PPI subnetwork
ENSG0000067836	1.31E-03	5.30E-04	NAPB PPI subnetwork
MP:0004957	5.34E-04	8.00E-04	DCLK1 PPI subnetwork
ENSG00000125814	1.34E-03	5.50E-04	ADARB2 PPI subnetwork
ENSG00000185736	8.02E-04	5.60E-04	abnormal cardinal vein morphology
MP:0004783	1.81E-03	5.60E-04	RYR3 PPI subnetwork
ENSG0000025772	1.35E-03	5.80E-04	columnar/cuboidal epithelial cell differentiation
GO:0002065	1.43E-03	5.90E-04	RTN3 PPI subnetwork
ENSG00000133318	1.50E-03	6.00E-04	LIN7B PPI subnetwork
ENSG0000104863	1.80E-03	6.00E-04	AMOTL1 PPI subnetwork
ENSG0000166025	1.02E-03	6.10E-04	REACTOME_UBIQUITIN:DEPENDENT_DEGRADATION_OF_CYCLIN_D1
REACTOME_UBIQUITIN:DEPENDENT_DEGRADATION_OF_CYCLIN_D1	1.62E-03	6.10E-04	USP11 PPI subnetwork

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ENSG0000102226	1.28E-03	6.20E-04	ATP2B1 PPI subnetwork		
ENSG0000070961	9.08E-04	6.50E-04	ENSG00000186979 PPI subnetwork		
ENSG0000186979	9.97E-04	6.70E-04	ZNF174 PPI subnetwork		
ENSG0000103343	1.81E-03	6.90E-04	REACTOME_ANTIGEN_PRESENTATION_FOLDING_ASSEMBLY_AND_PEPTI DE_LOADING_OF_CLASS_I_MHC		
REACTOME_ANTIGEN_PRESENTATION_FOLDING_ASSEMBLY_AND_PEPTI DE_LOADING_OF_CLASS_I_MHC	1.25E-03	7.10E-04	abnormal vascular development		
MP:0000259	1.29E-03	7.10E-04	KIF21A PPI subnetwork		
ENSG0000139116	1.80E-03	7.70E-04	REACTOME_CDK:MEDIATED_PHOSPHORYLATION_AND_REMOVAL_OF_C DC6		
REACTOME_CDK:MEDIATED_PHOSPHORYLATION_AND_REMOVAL_OF_C DC6	1.59E-03	7.90E-04	testis tumor		
MP:0006262	1.72E-03	7.90E-04	abnormal blastocyst morphology		
ENSG0000133083	2.15E-03	8.40E-04	APPBP2 PPI subnetwork		
ENSG0000062725	1.08E-03	8.60E-04	PDE1B PPI subnetwork		
ENSG0000123360	1.95E-03	8.60E-04	abnormal body weight		
MP:0001259	1.22E-03	8.70E-04	REACTOME_REGULATION_OF_MRNA_STABILITY_BY_PROTEINS_THAT_BI ND_AU:RICH_ELEMENTS		
REACTOME_REGULATION_OF_MRNA_STABILITY_BY_PROTEINS_THAT_BI ND_AU:RICH_ELEMENTS	1.13E-03	8.90E-04	REACTOME_APCCCDC20_MEDIATED_DEGRADATION_OF_SECURIN		
REACTOME_APCCCDC20_MEDIATED_DEGRADATION_OF_SECURIN	1.33E-03	8.90E-04	STXBP5 PPI subnetwork		
ENSG0000164506	1.05E-03	9.10E-04	embryonic digestive tract morphogenesis		
GO:0048557	1.86E-03	9.10E-04	REACTOME_DESTABILIZATION_OF_MRNA_BY_AUF1_HNRNP_D0		
REACTOME_DESTABILIZATION_OF_MRNA_BY_AUF1_HNRNP_D0	1.94E-03	9.10E-04	HLA-F PPI subnetwork		
ENSG0000204642	2.31E-03	9.50E-04	REACTOME_ACTIVATED_TLR4_SIGNALLING		
REACTOME_ACTIVATED_TLR4_SIGNALLING	1.92E-03	9.60E-04	REACTOME_UBIQUITIN:DEPENDENT_DEGRADATION_OF_CYCLIN_D		
REACTOME_UBIQUITIN:DEPENDENT_DEGRADATION_OF_CYCLIN_D	1.62E-03	9.90E-04	REACTOME_ACTIVATION_OF_CHAPERONES_BY_IRE1ALPHA		
REACTOME_ACTIVATION_OF_CHAPERONES_BY_IRE1ALPHA	2.20E-03	9.90E-04	CAMK1 PPI subnetwork		
ENSG00000134072	3.14E-03	9.90E-04	COPE PPI subnetwork		
(B) Leptin adjusted for BMI, European, additive model, sex-combined analysis. Coding variants included. MAX method used (Bonferroni correction for 1000 gene sets and 2 traits: P<2.5E- 05 for both chi2Pvalue and empPvalue)					
Name	chi2Pvalue	empPvalue	Pathway/Gene-set		
GO:0032769	6.37E-05	1.93E-05	negative regulation of monooxygenase activity		
MP:0005501	3.04E-05	4.62E-05	abnormal skin physiology		
ENSG0000206495	1.68E-04	4.71E-05	TRIM39 PPI subnetwork		
ENSG0000204599	1.68E-04	5.40E-05	TRIM39 PPI subnetwork		
ENSG0000206419	1.68E-04	6.00E-05	ENSG0000206419 PPI subnetwork		
ENSG0000198925	7.30E-05	9.60E-05	ATG9A PPI subnetwork		
ENSG0000105972	3.12E-04	1.70E-04	ENSG0000105972 PPI subnetwork		
MP:0003091	3.86E-04	1.87E-04	abnormal cell migration		
ENSG00000142867	8.77E-04	3.36E-04	BCL10 PPI subnetwork		

GO:0033273	3.51E-04	5.01E-04	response to vitamin
ENSG0000163902	1.19E-03	4.91E-04	RPN1 PPI subnetwork
ENSG0000138674	1.06E-03	5.09E-04	SEC31A PPI subnetwork
GO:0002065	1.32E-03	6.10E-04	columnar/cuboidal epithelial cell differentiation
ENSG00000120254	9.48E-04	6.40E-04	MTHFD1L PPI subnetwork
REACTOME_TOLL_LIKE_RECEPTOR_78_TLR78_CASCADE	1.39E-03	6.40E-04	REACTOME_TOLL_LIKE_RECEPTOR_78_TLR78_CASCADE
REACTOME_MYD88_DEPENDENT_CASCADE_INITIATED_ON_ENDOSOME	1.39E-03	6.40E-04	REACTOME_MYD88_DEPENDENT_CASCADE_INITIATED_ON_ENDOSOME
ENSG00000165699	2.08E-03	7.60E-04	TSC1 PPI subnetwork
ENSG00000164506	1.09E-03	7.80E-04	STXBP5 PPI subnetwork
ENSG00000185825	1.38E-03	9.00E-04	BCAP31 PPI subnetwork
GO:0071299	2.22E-03	9.80E-04	cellular response to vitamin A

SUPPLEMENTARY INFORMATION

Yaghootkar H, Zhang Y, Spracklen CN, Karaderi T, Huang LO, Bradfield J, et al. Genetic studies of leptin concentrations implicate leptin in the regulation of early adiposity



Figure S1. Association of rs10487505 and rs791600 variants near *LEP* with leptin concentrations adjusted for BMI in a genome-wide association study of up to 32,161 individuals of European ancestry (Kilpeläinen et al., 2016).



Figure S2. Meta-analysis of the association of the Met94 allele of rs17151919 with leptin concentrations adjusted for BMI in cohorts of African ancestry.



Figure S3: Surface region of the leptin protein with the Val94Met position (Val73Met in the mature protein) highlighted.



Figure S4: Leptin structure and the predicted impact of mutagenesis in position 73 from valine to methionine. The Rotamer list on the left shows sidechain torsions (Chi 1 and 2), with the probability and number of interatomic clashes, i.e. unfavourable interactions where atoms are too close together. On the right, the lower picture shows all possibilities for sidechain torsions when methionine is substituted with valine, whereas the upper picture displays the substitution with the highest probability (marked with red square in the Rotamer list).



Fig S5. Impact of Val94Met transversion at *LEP* rs17151919 on leptin secretion rate in HEK293 cells in different conditions. A) Leptin secretion rates for Val94 and Met94 during a 24-hr incubation period (48-72 hr post-transfaction), expressed as the amount of leptin secreted in ng per hour over 24 hrs (LEPs/hr) normalized by the respective cellular leptin content (LEPc, ng) at the end of incubation. B) Leptin secretion rates for Val94 and Met94 during a 1-hour incubation (72-73 hr post-transfection) in the presence of cycloheximide (CHX, 20 µg/ml) expressed as the amount of leptin secreted in ng during the 1-hour incubation (LEPs/hr), normalized by the respective cellular leptin content (LEPc, ng). Individual data points from four separate experiments (each with 2-3 technical replicates) are plotted. All data passed D'Agostino & Pearson normality test and repeated measures one-way ANOVA was performed to assess the difference in secretion rate between the genotypes. Mean ± SD and AVOVA results (F and p values) are reported in the table below each graph. C-D. The amounts of leptin secreted (LEPs) during a 1 hr incubation (72-73 hr post-transfection) in untreated control cells (C), and the corresponding cellular leptin content (LEPc) at the end of the

incubation (D). Leptin secretion rates shown in Fig 2B were ratios of the amounts of leptin secreted (LEPs) over the corresponding cellular leptin contents (LEPc) shown here.



Figure S6. Expression of leptin modifiers in human preadipocytes and mature adipocytes. De-identified human subcutaneous adipose stromal cells were generously provided by the Boston NORC and were cultured and differentiated as previously described (Lee and Fried, 2014). Preadipocytes and *in vitro*-differentiated adipocytes from two females and two males were studied. Lipid-laden cells were assayed between 10-14 days after initial treatment with differentiation factors. Transcript levels were determined by RT-qPCR, normalized to the geometric mean of *RPLP0* and *PPIA*, and expressed relative to levels in preadipocytes. Two-way repeated measures ANOVA with posthoc Sidak's multiple comparison tests were performed *: p<0.05, **: p<0.01, ****: p<0.0001, ns (no statistical difference) are indicated, comparing the transcript levels between preadipocytes and mature adipocytes. There was an interaction

between sex and differentiation stage for ZNF800 (p<0.01). No *ACTL9* transcript was detected (nd: none detected). Genotypes of the individuals were marked as R-reference allele and A-alternative allele.



Figure S7. Linkage disequilibrium between the Ser37Phe (rs2340550) variant in *ACTL9* and variants within ±500 kb in the 1000 Genomes European ancestry reference panel. The numbering refers to Regulome DB score of the variants (www.regulomedb.org). Non-coding variants are marked in orange color and coding variants in red. The plot was produced using LDlink (https://ldlink.nci.nih.gov).

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