

Genome Sequence of the Butanol Hyperproducer *Clostridium saccharoperbutylacetonicum* N1-4

Carlos del Cerro,^a Carmen Felpeto-Santero,^a Antonia Rojas,^b Marta Tortajada,^b Daniel Ramón,^{b,c} José L. García^a

Environmental Biology, Centro de Investigaciones Biológicas, Consejo Superior de Investigaciones Científicas, Madrid, Spain^a; Biopolis SL, Parc Científic Universitat de Valencia, Paterna, Spain^b; Lifesequencing SL, Parc Científic Universitat de Valencia, Paterna, Spain^c

C.D.C. and C.F.-S. contributed equally to this work

***Clostridium saccharoperbutylacetonicum* is one of the most important acetone-butanol-ethanol (ABE)-generating industrial microorganisms and one of the few bacteria containing choline in its cell wall. Here, we report the draft genome sequence of *C. saccharoperbutylacetonicum* strain N1-4 (6.6 Mbp; G+C content, 29.4%) and the findings obtained from the annotation of the genome.**

Received 30 January 2013 Accepted 6 February 2013 Published 7 March 2013

Citation del Cerro C, Felpeto-Santero C, Rojas A, Tortajada M, Ramón D, García JL. 2013. Genome sequence of the butanol hyperproducer *Clostridium saccharoperbutylacetonicum* N1-4. *Genome Announc.* 1(2):e00070-13. doi:10.1128/genomeA.00070-13.

Copyright © 2013 del Cerro et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](http://creativecommons.org/licenses/by/3.0/).

Address correspondence to José L. García, jlgarcia@cib.csic.es.

Clostridium is one of the largest bacterial genera, ranking second in size after *Streptomyces*, and members of the genus are classified as Gram-positive endospore-forming obligate anaerobes (1). Many species of *Clostridium* are of biotechnological importance, such as *Clostridium acetobutylicum*, which was used for acetone-butanol-ethanol (ABE) production during the first half of the last century before being replaced by petrochemical synthesis in the industrial production of chemicals (2). However, there has been a revival of interest in ABE fermentation, since renewable resources have become possible alternative substrates for the production of biofuels at a low cost (3). Despite the fact that *Clostridium saccharoperbutylacetonicum* has been considered a reference microorganism for ABE fermentation (4–8), it was not genetically characterized until very recently (9), and its genome remained unknown.

The genome of *C. saccharoperbutylacetonicum* N1-4 (ATCC 27021) has been sequenced using the Titanium kit and the GS-FLX pyrosequencing equipment from Roche. Preliminary assembly of raw reads was performed using Newbler software from Roche. This assembly was manually revised and improved to obtain a quality draft of 210 contigs. The genome was structurally and functionally annotated using Rapid Annotations using Subsystems Technology (RAST) (10), an automated genome annotation system, and the functions, names, and general properties of the gene products were predicted using this method. *C. saccharoperbutylacetonicum* N1-4 has one of the largest clostridial genomes (6.6 Mbp); it has a G+C content of 29.4%, encodes 20 RNAs, and contains 5,987 coding sequences.

Remarkably, *C. saccharoperbutylacetonicum* is one of the few bacteria that contain choline in the teichoic acids of their cell walls (11, 12). This property usually correlates with the expression of different modular proteins, named choline-binding proteins (CBPs), which have evolved from the fusion of a typical choline-binding domain (13) with a variety of functional protein modules that play important physiological roles (14, 15). We have anno-

tated 66 CBPs encoded by the genome of *C. saccharoperbutylacetonicum*. At least nine of these CBPs contained functional modules showing high similarity with cell wall lytic enzymes (16).

JSpecies (17) comparison of *C. saccharoperbutylacetonicum* N1-4 and *Clostridium beijerinckii* NCIMB 8052 gives an average nucleotide identity based on BLAST (ANIb) of 78.86% (ANIb aligned 36.85%) and an average nucleotide identity based on MUMmer (ANIm) of 85.69% (ANIm aligned 20.12%). These results confirmed that although the two species share a very large number of genes, they can be taxonomically classified as different species.

Nucleotide sequence accession number. The *C. saccharoperbutylacetonicum* N1-4 (ATCC 27021) genome sequence has been submitted to GenBank under the accession no. [AOIF00000000](https://www.ncbi.nlm.nih.gov/nuclink/AOIF00000000).

ACKNOWLEDGMENTS

We acknowledge the financial support provided by the Ministry of Economy and Competitiveness Project BIOSOS (CENIT-E 2009) and by project Consolider CSD2007-00005.

REFERENCES

1. Garrity GM, Brenner DJ, Krieg NR, Staley JT (ed). 2005. *Bergey's manual of systematic bacteriology*, vol 2. Springer Verlag, New York, NY.
2. Dürre P. 2008. Fermentative butanol production: bulk chemical and biofuel. *Ann. N. Y. Acad. Sci.* 1125:353–362.
3. Gu Y, Jiang Y, Wu H, Liu X, Li Z, Li J, Xiao H, Shen Z, Dong H, Yang Y, Li Y, Jiang W, Yang S. 2011. Economical challenges to microbial producers of butanol: feedstock, butanol ratio and titer. *Biotechnol. J.* 6:1348–1357.
4. Ellis JT, Hengge NN, Sims RC, Miller CD. 2012. Acetone, butanol, and ethanol production from wastewater algae. *Bioresour. Technol.* 111: 491–495.
5. Kobayashi G, Eto K, Tashiro Y, Okubo K, Sonomoto K, Ishizaki A. 2005. Utilization of excess sludge by acetone-butanol-ethanol fermentation employing *Clostridium saccharoperbutylacetonicum* N1-4 (ATCC 13564). *J. Biosci. Bioengin.* 99:517–519.
6. Nakayama S, Kiyoshi K, Kadokura T, Nakazato A. 2011. Butanol production from crystalline cellulose by cocultured *Clostridium thermocellum*

- and *Clostridium saccharoperbutylacetonicum* N1-4. Appl. Environ. Microbiol. 77:6470–6475.
7. Oshiro M, Hanada K, Tashiro Y, Sonomoto K. 2010. Efficient conversion of lactic acid to butanol with pH-stat continuous lactic acid and glucose feeding method by *Clostridium saccharoperbutylacetonicum*. Appl. Microbiol. Biotechnol. 87:1177–1185.
 8. Thang VH, Kanda K, Kobayashi G. 2010. Production of acetone-butanol-ethanol (ABE) in direct fermentation of cassava by *Clostridium saccharoperbutylacetonicum* N1-4. Appl. Biochem. Biotechnol. 161: 157–170.
 9. Keis S, Shaheen R, Jones DT. 2001. Emended descriptions of *Clostridium acetobutylicum* and *Clostridium beijerinckii*, and descriptions of *Clostridium saccharoperbutylacetonicum* sp. nov. and *Clostridium saccharobutylicum* sp. nov. Int. J. Syst. Evol. Microbiol. 51:2095–2103.
 10. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. BMC Genomics 9:75.
 11. García JL, García E, Sánchez-Puelles JM, López R. 1988. Identification of a lytic enzyme of *Clostridium acetobutylicum* that degrades choline-containing pneumococcal cell walls. FEMS Microbiol. Lett. 52:133–138.
 12. Podvin L, Reysset G, Hubert J, Sebald M. 1988. Presence of choline in teichoic acid of *Clostridium acetobutylicum* NI-4 and choline inhibition of autolytic function. J. Gen. Microbiol. 134:1603–1609.
 13. Fernández-Tornero C, López R, García E, Giménez-Gallego G, Romero A. 2001. A novel solenoid fold in the cell wall anchoring domain of the pneumococcal virulence factor LytA. Nat. Struct. Biol. 8:1020–1024.
 14. García JL, Sánchez-Beato AR, Medrano FJ, López R. 1998. Versatility of choline-binding domain. Microb. Drug Resist. 4:25–36.
 15. Hakenbeck R, Madhour A, Denapaite D, Brückner R. 2009. Versatility of choline metabolism and choline-binding proteins in *Streptococcus pneumoniae* and commensal streptococci. FEMS Microbiol. Rev. 33: 572–586.
 16. López R, García E, García P, García JL. 1997. The pneumococcal cell wall degrading enzymes: a modular design to create new lysins? Microb. Drug Resist. 3:199–211.
 17. Richter M, Rosselló-Móra R. 2009. Shifting the genomic gold standard for the prokaryotic species definition. Proc. Natl. Acad. Sci. U. S. A. 106: 19126–19131.