JAMA Neurology | Original Investigation

Genome-wide Pleiotropy Between Parkinson Disease and Autoimmune Diseases

Aree Witoelar, PhD; Iris E. Jansen, PhD; Yunpeng Wang, PhD; Rahul S. Desikan, MD, PhD; J. Raphael Gibbs, MS; Cornelis Blauwendraat, MSc; Wesley K. Thompson, PhD; Dena G. Hernandez, MS; Srdjan Djurovic, PhD; Andrew J. Schork, MSc; Francesco Bettella, PhD; David Ellinghaus, PhD; Andre Franke, PhD; Benedicte A. Lie, PhD; Linda K. McEvoy, PhD; Tom H. Karlsen, MD, PhD; Suzanne Lesage, PhD; Huw R. Morris, PhD; Alexis Brice, MD; Nicholas W. Wood, PhD, FRCP, FMedSci; Peter Heutink, PhD; John Hardy, PhD; Andrew B. Singleton, PhD; Anders M. Dale, PhD; Thomas Gasser, MD, PhD; Ole A. Andreassen, MD, PhD; Manu Sharma, PhD; for the International Parkinson's Disease Genomics Consortium (IPDGC), North American Brain Expression Consortium (NABEC), and United Kingdom Brain Expression Consortium (UKBEC) Investigators

IMPORTANCE Recent genome-wide association studies (GWAS) and pathway analyses supported long-standing observations of an association between immune-mediated diseases and Parkinson disease (PD). The post-GWAS era provides an opportunity for cross-phenotype analyses between different complex phenotypes.

OBJECTIVES To test the hypothesis that there are common genetic risk variants conveying risk of both PD and autoimmune diseases (ie, pleiotropy) and to identify new shared genetic variants and their pathways by applying a novel statistical framework in a genome-wide approach.

DESIGN, SETTING, AND PARTICIPANTS Using the conjunction false discovery rate method, this study analyzed GWAS data from a selection of archetypal autoimmune diseases among 138 511 individuals of European ancestry and systemically investigated pleiotropy between PD and type 1 diabetes, Crohn disease, ulcerative colitis, rheumatoid arthritis, celiac disease, psoriasis, and multiple sclerosis. NeuroX data (6927 PD cases and 6108 controls) were used for replication. The study investigated the biological correlation between the top loci through protein-protein interaction and changes in the gene expression and methylation levels. The dates of the analysis were June 10, 2015, to March 4, 2017.

MAIN OUTCOMES AND MEASURES The primary outcome was a list of novel loci and their pathways involved in PD and autoimmune diseases.

RESULTS Genome-wide conjunctional analysis identified 17 novel loci at false discovery rate less than 0.05 with overlap between PD and autoimmune diseases, including known PD loci adjacent to *GAK*, *HLA-DRB5*, *LRRK2*, and *MAPT* for rheumatoid arthritis, ulcerative colitis and Crohn disease. Replication confirmed the involvement of *HLA*, *LRRK2*, *MAPT*, *TRIM10*, and SE*TD1A* in PD. Among the novel genes discovered, *WNT3*, *KANSL1*, *CRHR1*, *BOLA2*, and *GUCY1A3* are within a protein-protein interaction network with known PD genes. A subset of novel loci was significantly associated with changes in methylation or expression levels of adjacent genes.

CONCLUSIONS AND RELEVANCE The study findings provide novel mechanistic insights into PD and autoimmune diseases and identify a common genetic pathway between these phenotypes. The results may have implications for future therapeutic trials involving anti-inflammatory agents.

JAMA Neurol. 2017;74(7):780-792. doi:10.1001/jamaneurol.2017.0469 Published online June 5. 2017. Editorial page 769

Supplemental content

Author Affiliations: Author affiliations are listed at the end of this article.

Group Information: The International Parkinson's Disease Genomics Consortium (IPDGC), North American Brain Expression Consortium (NABEC), and United Kingdom Brain Expression Consortium (UKBEC) investigators are listed at the end of this article.

Corresponding Author: Manu Sharma, PhD, Centre for Genetic Epidemiology, Institute for Clinical Epidemiology and Applied Biometry, University of Tübingen, Silcherstrasse 5, 72076 Tübingen, Germany (manu.sharma@uni-tuebingen.de).

jamaneurology.com

merging evidence suggests a substantial genetic component underlying Parkinson disease (PD).^{1,2} Linkage analysis and genome-wide association studies (GWAS) confirmed the role of genes involved in familial and sporadic forms of PD.^{3,4} Genome-wide association studies are able to identify variants with strong genetic effects; however, true polygenic risk alleles with weaker evidence for association may be overlooked.^{5,6} The estimated heritability in PD GWAS substantially increases when weak effect loci are also considered,² further emphasizing the involvement of a large proportion of genetic risk variants below standard genome-wide significance thresholds. Moreover, these studies^{2,5} nominate novel loci that have not been implicated in disease pathogenesis.

The association between inflammation and neurodegenerative diseases has long been observed in Alzheimer disease (AD),^{7,8} amyotrophic lateral sclerosis, and, highlighted in this work, PD.⁹⁻¹¹ In an epidemiological study¹² in Sweden, 6 of 33 studied types of autoimmune disorders were identified with an increased risk of developing PD, including amyotrophic lateral sclerosis, hyperthyroidism, hypothyroidism, multiple sclerosis, pernicious anemia, and polymyalgia rheumatica, although this finding was not observed in a population-based casecontrol study¹³ from Denmark. The association between PD and MS has been confirmed in other studies. 14-16 Furthermore, in clinical studies, 17,18 regular users of nonsteroidal antiinflammatory drugs were found to have lowered risk of PD. It is still not clear whether immune dysfunction has an important role in early stages of PD or is simply the end product of a neuronal degeneration process.¹⁹

The occurrence of PD in patients with autoimmune diseases, or vice versa, could reflect genetically determined factors influencing both lipid metabolism and immune disorders that cannot be elucidated by epidemiological and clinical studies alone. 19 Genome-wide-based pathway analyses in PD supported the association between PD and autoimmune diseases.⁵ Early independent studies showed that at least one gene, LRRK2, is statistically significant in both PD²⁰ and Crohn disease.²¹ The results of a recently published study²² suggested that, along with known PD loci USP25, HLA-DRA, and LRRK2, additional genetic factors are present that contribute to genetic comorbidity shared by PD and CD. A systematic study is needed to decipher whether shared polygenetic risk variants (ie, genetic pleiotropy) exist between PD and autoimmune diseases and whether particular molecular biological pathways are involved.

An approach combining GWAS data from 2 disorders with shared pathways can significantly increase the power to discover novel loci and partly reveal the missing heritability in GWAS. Our group recently developed a novel statistical framework to identify single-nucleotide polymorphisms (SNPs) exhibiting genetic pleiotropy between multiple phenotypes and applied it to identify pleiotropy between AD and autoimmune diseases. ²³ This approach also identified novel loci between schizophrenia and cardiovascular diseases, ²⁴ psychiatric disorders, ²⁵ and neurological diseases. ²⁶

Herein, we applied this approach to investigate the potentially shared genetic basis for PD and autoimmune diseases. Autoimmune diseases were selected based on available large

Key Points

Question Are there genome-wide genetic risk factors for Parkinson disease that are shared with pathways of autoimmune diseases?

Findings In this study of combined genome-wide association data with control replication, we identified 17 novel genetic loci shared between Parkinson disease and type 1 diabetes, Crohn disease, ulcerative colitis, rheumatoid arthritis, celiac disease, psoriasis, and multiple sclerosis.

Meaning Our findings identify a common genetic pathway between Parkinson disease and autoimmune diseases and suggest that the immune system influences Parkinson disease pathogenesis.

GWAS, including PD, type 1 diabetes, CD, ulcerative colitis, rheumatoid arthritis celiac disease, psoriasis, and multiple sclerosis. ²⁷⁻³³ We used conditional and conjunction false discovery rate analyses to define SNPs associated with both groups of phenotypes (pleiotropic SNPs).

Methods

Participant Samples

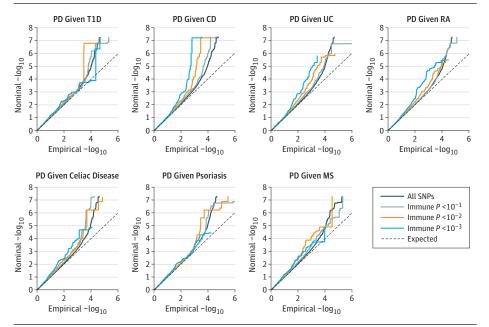
Using the conjunction false discovery rate method, this study analyzed GWAS data from a selection of archetypal autoimmune diseases among 138 511 individuals of European ancestry and systemically investigated pleiotropy between PD and type 1 diabetes, Crohn disease, ulcerative colitis, RA, celiac disease, psoriasis, and multiple sclerosis. Genome-wide association studies summary statistic P values and z scores were obtained from the studies of PD, 4 CD, 27 ulcerative colitis, 28 RA, 29 type 1 diabetes,³⁰ celiac disease,³¹ psoriasis,³² and multiple sclerosis³³ (eTable 1 in the Supplement). Details of the inclusion criteria and phenotype characteristics of the GWAS are described in the original publications. The relevant institutional review boards or ethics committees approved the research protocol of the individual GWAS used in the present analysis, and all participants gave written informed consent. The dates of the analysis were June 10, 2015, to March 4, 2017. All P values were corrected for inflation using a genomic control procedure.24,25

Statistical Analysis

Conditional Quantile-Quantile Plots

The quantitative estimates of true associations and statistical enrichment were calculated from the distributions of summary statistics. 34,35 We plotted conditional quantile-quantile (Q-Q) plots for a primary phenotype by filtering SNPs based on their level of association with a secondary phenotype. Pleiotropic enrichment between PD and an autoimmune disorder was evident if the degree of deflection of PD P values from the expected null line produced successive leftward deflection when conditioned on an autoimmune disease. 24,25,36 To control for linkage disequilibrium (LD), we performed a random pruning procedure. 37

 $\label{eq:policy} Figure \ 1. \ Pleiotropic Enrichment of Parkinson \ Disease \ (PD) \ Conditioned \ on \ Association \ P \ Values \ of \ Autoimmune \ Diseases$



Conditional quantile-quantile plots (nominal vs empirical -log₁₀ P values) are calculated from single-nucleotide polymorphism (SNP) populations of varying degrees of association with autoimmune diseases. Each population is composed of SNPs that pass certain significance of association (type 1 diabetes [T1D], Crohn disease [CD], ulcerative colitis [UC], rheumatoid arthritis [RA], celiac disease, psoriasis, and multiple sclerosis [MS]) at $P \le 1$ (All SNPs), $P < 10^{-1}$, $P < 10^{-2}$, and $P < 10^{-3}$. All Pvalues have been corrected for genomic inflation. Dotted lines indicate the expected line under the null hypothesis, and leftward deflection shows increasing degrees of enrichment

Conditional and Conjunction False Discovery Rate

We defined conditional false discovery rate, denoted by $FDR_{trait1|trait2}$, as the posterior probability that a given SNP is null for the first trait given that the P values in both traits are smaller than their observed P values. 24,25 We defined conjunction false discovery rate, denoted by $FDR_{trait1&trait2}$, as the posterior probability that a given SNP is null for both phenotypes simultaneously given that the P values for both traits are as small or smaller than the observed P values. We obtained a conservative estimate of conjunction false discovery rate by taking the minimum of $FDR_{trait1|trait2}$ and $FDR_{trait2|trait1}$. To control for LD, we applied a random pruning procedure. 37 Detailed information on the methods can be found in prior studies. 24,25

NeuroX Data

We replicated the top conjunction false discovery rate loci, highly associated with both PD and autoimmune disorders, in a second independent PD data set. The data set was generated with the NeuroX exome array, $^{3.38}$ including 6927 PD cases and 6108 controls. Variants passing standard quality control (Hardy-Weinberg equilibrium $P > 1 \times 10^{-6}$ and maximum missingness rate of 5%) were tested for association with PD with a logistic model correcting for the first 4 multidimensional scaling components and sex.

Gene Expression and Methylation Changes

We determined the regional methylation and expression patterns within ± 1 megabases of 103 SNPs of interest. We investigated frontal cortex and cerebellum microarray data from the North American Brain Expression Consortium (NABEC)³⁹ and the United Kingdom Brain Expression Consortium (UKBEC)⁴⁰ of 396 European samples without neuropathological evidence of disease. We also accessed a second expression quan-

titative trait loci (eQTL) data set based on cap analysis gene expression profiling technique of the frontal cortex of 119 NABEC samples. ⁴¹ A total of 98 variants (3 not testable) and 83 variants (20 not testable) were studied for the microarray-based and cap analysis gene expression-based data sets, respectively. For details of these procedures, see the eMethods in the Supplement.

Genetic Correlations Among Implicated Loci

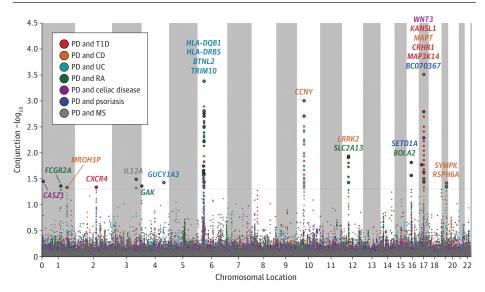
To investigate the genetic relatedness among implicated SNPs, we performed protein-protein interaction analyses using STRING version 10.⁴² The input consisted of novel loci as identified in pleiotropic analyses. We considered total scores above 0.400 (medium confidence) that correspond to the combination of the following 4 different scores: coexpression, experimental, knowledge, and text mining.

Results

Significant Genetic Overlap Between PD and Autoimmune Diseases

Conditional Q-Q plots for PD conditioned on association P values of autoimmune diseases showed strong enrichment for Crohn disease (**Figure 1**). Successive leftward shifts for decreasing nominal PD P values indicated that the proportion of non-null SNPs in PD increased considerably with higher levels of association with an autoimmune phenotype. For example, when conditioned on CD, the proportion of SNPs reaching a significance of PD $P < 10^{-5}$ in the category Crohn disease $P < 10^{-3}$ is 20 times greater than when all SNPs were examined (Figure 1 and eFigure 1 in the Supplement). Similar enrichment was found with ulcerative colitis and RA, and weaker

Figure 2. Conjunctional False Discovery Rate Manhattan Plot of – \log_{10} Values for the Associated Autoimmune Phenotypes



All single-nucleotide polymorphisms without pruning are plotted: enlarged points represent significant single-nucleotide polymorphisms with conjunction false discovery rate less than 0.05, and small points represent the nonsignificant single-nucleotide polymorphisms. The most significant single-nucleotide polymorphism in each linkage disequilibrium block is marked with black circles and annotated with its closest gene, showing the localization of 17 common loci (some loci may have multiple genes) between Parkinson disease and autoimmune diseases listed in Table 1. CD indicates Crohn disease; MS, multiple sclerosis; RA, rheumatoid arthritis: T1D, type 1 diabetes; and UC, ulcerative colitis.

enrichment was found with celiac disease and multiple sclerosis. The enrichment remained after removing the major histocompatibility complex and *MAPT* regions (eFigure 2 in the Supplement).

Shared Susceptibility Loci for PD and Autoimmune Disorders

We performed a conjunction false discovery rate analysis and visualized the pleiotropic loci between PD and autoimmune diseases in a Manhattan plot (Figure 2). Based on conjunction false discovery rate less than 0.05, we detected 17 independent pleiotropic loci for the 7 autoimmune diseases (Table 1). Nine loci remained after excluding the major histocompatibility complex and MAPT regions (eTable 2 in the Supplement). Of the 17 loci, the directions of PD effect given by z scores were mostly the same with Crohn disease, ulcerative colitis, and celiac disease and opposite with rheumatoid arthritisand psoriasis (Table 1 and eTable 3 in the Supplement). The conjunction false discovery rate analyses over multiple autoimmune phenotypes showed overlapping susceptibility loci between Crohn disease and ulcerative colitis and demonstrated some overlap between ulcerative colitis, RA, celiac disease, and multiple sclerosis (eFigure 3 in the Supplement). We were able to replicate 5 loci in our in-house independent NeuroX data at P < .05 (Table 1). In addition to the previously published HLA, LRRK2, and MAPT associations, 4 we also identified 2 new loci adjacently located to TRIM10 and SETD1A.

Functional Interpretation of Shared Susceptibility Loci

A total of 103 associated variants resulting from conditional false discovery rate less than 0.01 and conjunction false discovery rate less than 0.05 were tested for being a methylation QTL (methQTL) or an eQTL. **Table 2** summarizes the significant methQTL and eQTL in the brain in which the affected gene is implied by the literature (see the Discussion section) to have

a function in the immune system. As expected, most hits are for variants located in the *HLA* locus and *MAPT* locus, both of which have been implicated in PD. ^{3,37,43-45} Within the NABEC data set, 31 of the 103 variants were shown to have a significant effect on the methylation status of 16 genes (eTable 4 in the Supplement). Likewise, 29 variants were significantly associated with changes in expression of 14 genes in the NABEC, UKBEC, or in-house eQTL data set (eTable 5 in the Supplement).

In addition to the exploration for methQTL and eQTL within the described data sets, we compared a recent elaborate eQTL study 46 of multiple immune cell types (B cells, CD4 T cells, CD8 T cells, monocytes, and neutrophils) in patients with inflammatory bowel disease and healthy controls. Our 103 candidate SNPs intersected with those authors' significant (false discovery rate < 0.05) eQTL results. eTable 6 in the Supplement lists significant eQTL for 10 variants influencing the expression of 8 genes, 5 of which (DGKQ, IDUA, BST1, CD38, and SNCA) have previously been discussed in the context of PD.4 These SNPs could contribute to PD risk through immune mechanisms by regulating the gene expression of these PD-related genes in these immune-specific cells. Six immune eQTL that regulate the expression of 2 genes (DGKQ and DMPK) were also observed in the brain eQTL and methQTL data, affirming the immune-related involvement of these genes in PD.

Shared Biological Pathways Between Significant Risk Loci

Using functional gene networks and protein interaction networks, the connectivity among the loci in the combined network increased considerably compared with the networks represented by pleiotropic and PD loci (**Figure 3**). The network analyses revealed interaction between the 17 loci identified in our study with nodes defined by PD loci (eg, *GUCY1A3*, *KANSL1*, *CRHR1*, *WNT3*, and *BOLA2*). This finding

Table 1. Seventeen Independent Pleiotropic Loci in Parkinson Disease (PD) and Autoimmune Diseases With Conjunction False Discovery Rate (FDR) Less Than 0.05

| 1 | SNP | Gene | OMIM Accession No. | Chromosome | Human Genome Build 19 Position | PD P Value | PD FDR | Minimum Conjunction FDR | Phenotype | Direction | NeuroX P Value | Meta-analysis P Value |
|----|------------|----------|-----------------------|------------|-----------------------------------|------------|----------|----------------------------|----------------|---------------|-------------------|--------------------------|
| | rs616519 | CASZ1 | 609895 | 1 | 10,768,002 | 2.03E-04 | 2.03E-01 | 3.59E-02 | Celiac disease | Same | NA | <.05 |
| 7 | rs7515174 | FCGR2A | 146790 | 1 | 161,476,949 | 2.68E-04 | 2.28E-01 | 4.38E-02 | RA | Not available | NA | <.05 |
| 3 | rs1572789 | MROH3P | NA | 1 | 200,932,305 | 4.79E-04 | 3.24E-01 | 4.68E-02 | CD | Same | NA | <.05 |
| 4 | rs2011946 | CXCR4 | 162643 | 2 | 136,817,616 | 2.44E-04 | 2.28E-01 | 4.64E-02 | T1D | Opposite | NA | 6.20E-07 |
| 2 | rs2243123 | IL12A | 161560 | 6 | 159,709,651 | 2.98E-04 | 2.57E-01 | 3.30E-02 | MS | Opposite | NA | <.05 |
| 9 | rs3755963 | GAK | 602052 | 4 | 894,255 | 2.36E-05 | 5.77E-02 | 4.38E-02 | RA | Opposite | NA | 1.41E-10 |
| 7 | rs2625276 | GUCY1A3 | 139396 | 4 | 156,600,197 | 6.47E-05 | 1.06E-01 | 3.76E-02 | nc | Same | NA | <.05 |
| ∞ | rs9261531 | TRIM10 | 605701 | 9 | 30,120,268 | 1.11E-04 | 1.38E-01 | 1.81E-02 | OUC | Opposite | NA | <.05 |
| 8 | rs2022065 | TRIM10 | 605701 | 9 | 30,121,460 | 2.25E-04 | 2.28E-01 | 2.71E-02 | MS | Same | NA | <.05 |
| 8 | rs9261535 | TRIM10 | 605701 | 9 | 30,127,323 | 1.36E-04 | 1.58E-01 | 1.82E-02 | Celiac disease | Same | .047 | <.05 |
| 6 | rs9268480 | BTNL2 | 000909 | 9 | 32,363,844 | 5.60E-06 | 1.66E-02 | 1.29E-03 | OC | Same | NA | <1.00E-04 |
| 6 | rs3763312 | BTNL2 | 000909 | 9 | 32,376,348 | 1.25E-05 | 3.51E-02 | 2.00E-03 | Celiac disease | Same | NA | 1.20E-11 |
| 6 | rs28366337 | HLA-DRB5 | 604776 | 9 | 32,564,699 | 8.53E-05 | 1.21E-01 | 1.51E-02 | OUC | Same | NA | NA |
| 6 | rs17425622 | HLA-DRB5 | 604776 | 9 | 32,571,961 | 1.43E-06 | 6.07E-03 | 4.26E-04 | OUC | Same | 1.37E-05 | 8.07E-05 |
| 6 | rs4642516 | HLA-DQB1 | 604305 | 9 | 32,657,543 | 2.69E-04 | 2.28E-01 | 3.64E-02 | OU | Same | NA | <.05 |
| 6 | rs9275328 | HLA-DQB1 | 604305 | 9 | 32,666,822 | 2.18E-05 | 4.91E-02 | 3.17E-03 | Celiac disease | Same | NA | 1.65E-07 |
| 6 | rs9275356 | HLA-DQB1 | 604305 | 9 | 32,667,850 | 1.19E-05 | 3.51E-02 | 1.59E-03 | CD | Not available | NA | 2.52E-12 |
| 10 | rs12242110 | CCNY | 612786 | 10 | 35,535,695 | 8.71E-06 | 2.44E-02 | 1.01E-03 | 00 | Same | .339 | <1.00E-04 |
| 11 | rs7960662 | SLC2A13 | 611036 | 12 | 40,479,067 | 2.33E-05 | 5.77E-02 | 1.24E-02 | RA | Same | NA | <.05 |
| 12 | rs17467164 | LRRK2 | 200609 | 12 | 40,814,197 | 1.06E-04 | 1.38E-01 | 1.17E-02 | CD | Same | .030 | 1.07E-07 |
| 13 | rs4787495 | BOLA2 | 613182 | 16 | 30,165,725 | 3.50E-06 | 1.12E-02 | 1.54E-02 | RA | Opposite | NA | <.05 |
| 14 | rs11640961 | SETD1A | 611052 | 16 | 30,979,818 | 3.99E-05 | 7.89E-02 | 2.75E-02 | Psoriasis | Opposite | 2.81E-06 | 3.07E-07 |
| 15 | rs1975974 | BC070367 | 601519 | 17 | 21,707,060 | 5.15E-05 | 9.16E-02 | 1.72E-02 | Psoriasis | Same | NA | >.05 |
| 16 | rs2867316 | MAP3K14 | 604655 | 17 | 43,376,447 | 1.83E-04 | 2.03E-01 | 3.66E-02 | T1D | Same | 796. | 8.29E-10 |
| 16 | rs393152 | CRHR1 | 122561 | 17 | 43,719,143 | 1.31E-18 | 6.84E-07 | 9.18E-03 | T1D | Same | .074 | <5.00E-08 |
| 16 | rs1467967 | MAPT | 157140 | 17 | 43,986,179 | 6.07E-08 | 3.38E-04 | 3.15E-04 | 0 | Same | NA | <5.00E-08 |
| 16 | rs17652121 | MAPT | 157140 | 17 | 44,073,973 | 3.41E-18 | 6.84E-07 | 3.18E-02 | CD | Same | .034 | <5.00E-08 |
| 16 | rs17661428 | KANSL1 | 612452 | 17 | 44,208,144 | 3.60E-15 | 6.84E-07 | 1.64E-03 | T1D | Same | NA | <5.00E-08 |
| 16 | rs2074404 | WNT3 | 165330 | 17 | 44,865,439 | 5.27E-11 | 6.84E-07 | 5.22E-03 | Celiac disease | Same | .599 | <5.00E-08 |
| 17 | rs12463359 | RSPH6A | 607548 | 19 | 46,304,585 | 9.31E-06 | 2.94E-02 | 3.86E-02 | 00 | Same | NA | <.05 |
| 17 | rs10500292 | SYMPK | 602388 | 19 | 46,327,933 | 8.82E-06 | 2.44E-02 | 3.86E-02 | CD | Same | NA | <.05 |

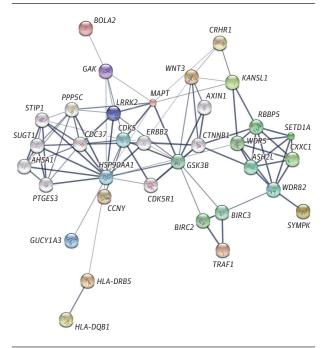
Abbreviations: CD, Crohn disease; MS, multiple sclerosis; NA, not applicable; RA, rheumatoid arthritis; SNP, single-nucleotide polymorphism; TID, type I diabetes; and UC, ulcerative colitis.

phenotypes, as well as the minimum conjunction FDR across all phenotypes. One SNP is listed for the major histocompatibility complex region on chromosome 6. Nine of the top loci were available for association testing within the NeuroX data set. Meta-analysis association P values in the study by Nalls et al³ were obtained from publicly available PDGene (pdgene.org).

 $^{^{3}}$ Listed are independent gene loci with SNPs with conjunction FDR less than 0.05 in both PD and the associated autoimmune disease represented by the SNP with the minimum conjunction FDR in each linkage disequilibrium block (ℓ^{2} <0.200). For comparison, the conjunction FDR values for each identified SNP are listed for all

| Table 2. Overv | iew of Expressic | Table 2. Overview of Expression Quantitative Trait Loci (eQTL) and | | lethylation | Methylation Effects Related to the Immune System | to the Im | nune System | | | | |
|--------------------------------|---|--|--|--|--|-----------------|--|---|-----------------------|-----------------------------------|------------------------|
| SNP | Chromosome | Human Genome Build 19 Position | Major Allele, Minor Allele | maf Data Set ^a | European maf 1000G | Assay | Tissue | Trait | Gene | Effect Size of Minor Allele | P Value |
| rs7515174 | 1 | 161,476,949 | C, G | 0.15 | 0.14 | CpG | Frontal cortex | cg24422489 | FCGR2A | -0.603 | 8.58×10^{-7} |
| rs9261531 | 9 | 30,120,268 | C, T | 0.23 | 0.25 | CpG | Cerebellum | cg00679556 | TRIM31 | 0.467 | 1.29×10^{-6} |
| | | | | 0.23 | 0.25 | CpG | Frontal cortex | cg20879959 | HLA-A | -0.458 | 2.08×10^{-6} |
| rs9261535 | 9 | 30,127,323 | G, A | 0.23 | 0.23 | CpG | Cerebellum | cg00679556 | TRIM31 | 0.472 | 1.27×10^{-6} |
| | | | | 0.23 | 0.23 | CpG | Frontal cortex | cg20879959 | HLA-A | -0.453 | 3.20×10^{-6} |
| rs9268480 | 9 | 32,363,844 | C, T | 0.27 | 0.27 | eQTL C | Frontal cortex | L2 None chr6 - 32557582 | HLA-DRB1 | -0.485 | 1.44×10^{-5} |
| rs4642516 | 9 | 32,657,543 | 7, 6 | 0.52 | 0.48 | CpG | Cerebellum | cg25764570 | HLA-DRA | -0.352 | 3.85×10^{-5} |
| rs1059504 | 17 | 43,472,321 | G, A | 0.43 | 0.49 | eQTL C | Frontal cortex | L2 None chr17 + 44270964 | CRHR1 | -0.300 | 5.67×10^{-5} |
| rs11012 | 17 | 43,513,441 | C, T | 0.23 | 0.19 | eQTL C | Frontal cortex | L2 None chr17 + 44270964 | CRHR1 | 0.599 | 1.18×10^{-16} |
| rs393152 | 17 | 43,719,143 | A, G | 0.27 | 0.24 | eQTL C | Frontal cortex | L2 None chr17 + 44270964 | CRHR1 | 0.610 | 1.85×10^{-22} |
| rs8072451 | 17 | 43,893,716 | C, T | 0.26 | 0.24 | eQTL M | Cerebellum | ILMN 1709549 | PLEKHM1 | -0.507 | 8.62×10^{-10} |
| rs2301689 | 17 | 43,935,838 | C, T | 0.33 | 0.32 | CpG | Frontal cortex | cg07321605 | NSF | -0.442 | 4.65×10^{-6} |
| rs17652121 | 17 | 44,073,973 | T, C | 0.25 | 0.24 | eQTL M | Cerebellum | ILMN 1709549 | PLEKHM1 | -0.499 | 9.54×10^{-10} |
| | | | | 0.25 | 0.24 | eQTL C | Frontal cortex | L2 None chr17 + 44270964 | CRHR1 | 0.569 | 1.27×20^{-19} |
| rs4792827 | 17 | 44,131,305 | C, T | 0.44 | 0.41 | CpG | Cerebellum | cg07321605 | NSF | -0.347 | 8.82×10^{-5} |
| rs17661428 | 17 | 44,208,144 | C, G | 0.25 | 0.24 | eQTL M | Cerebellum | ILMN 1709549 | PLEKHM1 | -0.759 | 2.50×10^{-5} |
| rs183211 | 17 | 44,788,310 | G, A | 0.27 | 0.26 | eQTL C | Frontal cortex | L2 None chr17 + 44270964 | CRHR1 | 0.566 | 1.32×10^{-21} |
| | | | | 0.27 | 0.26 | eQTL C | Frontal cortex | L2 None chr17 + 43861684 | CRHR1 | 0.169 | 6.21×10^{-5} |
| rs169201 | 17 | 44,790,203 | A, G | 0.24 | 0.22 | eQTL C | Frontal cortex | L2 None chr17 + 44270964 | CRHR1 | 0.615 | 6.57×10^{-5} |
| | | | | 0.24 | 0.22 | eQTL C | Frontal cortex | L2 None chr17 + 43861684 | CRHR1 | 0.192 | 1.11×10^{-5} |
| rs142167 | 17 | 44,795,234 | A, G | 0.27 | 0.26 | eQTL C | Frontal cortex | L2 None chr17 + 44270964 | CRHR1 | 0.566 | 1.32×10^{-5} |
| | | | | 0.27 | 0.26 | eQTL C | Frontal cortex | L2 None chr17 + 43861684 | CRHR1 | 0.169 | 6.21×10^{-5} |
| rs199533 | 17 | 44,828,931 | G, A | 0.24 | 0.22 | eQTL C | Frontal cortex | L2 None chr17 + 44270964 | CRHR1 | 0.603 | 1.95×10^{-23} |
| | | | | 0.24 | 0.22 | eQTL C | Frontal cortex | L2 None chr17 + 43861684 | CRHR1 | 0.197 | 5.61×10^{-6} |
| rs199515 | 17 | 44,856,641 | C, G | 0.23 | 0.22 | eQTL C | Frontal cortex | L2 None chr17 + 44270964 | CRHR1 | 0.616 | 8.35×10^{-23} |
| rs2074404 | 17 | 44,865,439 | T, G | 0.26 | 0.27 | eQTL C | Frontal cortex | L2 None chr17 + 44270964 | CRHR1 | 0.562 | 3.08×10^{-20} |
| Abbreviations: expression data | CAGE, cap analysi ; eQTL M, eQTL a le polymorphism; | Abbreviations: CAGE, cap analysis gene expression; CpG, methylation assay; eQTL C, eQTL assay based on CAGE expression data; eQTL M, eQTL assay based on microarray expression data; maf, minor allele frequency; SNP, single-nucleotide polymorphism; Trait, probe name of methylation or expression assay. | nethylation assay expression data; hylation or expre | ; eQTL C, eQ maf, minor a ssion assay. | ΣTL assay based α | on CAGE SNP, | ^a In North Amer (UKBEC), or C, | ^a In North American Brain Expression Consortium (NABEC), United Kingdom Brain Expression Consortium (UKBEC), or CAGE data set. | ABEC), United Kingdor | n Brain Expressior | . Consortium |

Figure 3. Functional Gene Network for Novel Pleiotropic Loci From the Present Analysis and Previously Confirmed Parkinson Disease



The protein-coding genes closest to the most associated single-nucleotide polymorphism in the pleiotropic loci and the previously confirmed Parkinson disease loci were used to construct a functional similarity network of genes (see the Methods section). Nodes are colored to show association. The thickness of lines connecting nodes indicates the strength of the association between nodes.

demonstrates biological relatedness between the pleiotropic loci from the present analysis and PD loci from previous reports. 47

Discussion

Genetic comorbidities between PD and immune-related genes have only been explored for high-risk PD loci. 5,6,22 Our study using a genome-wide unbiased statistical approach identified 17 shared loci between PD and 7 autoimmune diseases. This finding strengthens the hypothesis that known PD risk genes might contribute to PD through immune system defects (eg, LRRK2, GAK, HLA, and MAPT), ^{21,48} while suggesting that additional loci with weaker associations also contribute to pleiotropy (Table 1). TRIM10 is closely positioned to the HLA region, and, although not extensively described, variants in SETD1A have also previously been associated with PD. 49,50 While we found an overlap between genetic risk factors for autoimmune diseases and PD, the specific loci involved for each trait differed. This result is consistent with the findings that some susceptibility loci are associated with risk in multiple immune-mediated diseases.51

Because our method considers joint *P* values, some SNPs with strong association with PD might not pass the significance threshold if they only had marginal association

with the autoimmune disease, and vice versa. For instance, a locus in SNCA has one of the strongest associations in PD⁴ (PD $P = 3.67 \times 10^{-26}$), but it is not associated with autoimmune diseases (RA P = .6184). Taking the 2 P values together, its conjunction false discovery rate is not significant (PD and rheumatoid arthritisconjunction false discovery rate of 0.9856). Among the 13 significant genes in our PD data set, we did not find significant conjunction false discovery rate loci among SYT11, ACMSD, STK39, MCCC1/ LAMP3, BST1, SNCA, and CCDC62/HIP1R owing to weak association with autoimmune disorders (eTable 7 in the Supplement). Likewise, some known risk loci for immune diseases were not found in our results: one locus near CARD15/NOD2 was significant in Crohn disease $(P = 1.21 \times 10^{-58})$ but not in PD (P = .2163), and its conjunction false discovery rate of 0.8496 did not pass our threshold. MC1R has been reported to be associated with PD and immune-mediated diseases,52,53 but it had not been reported in our data sets of PD (P = .2936) or Crohn disease (P = .2936).

Our brain-based QTL analyses suggest immune function-related genes for which the expression or methylation level is changed by of one of our identified susceptibility loci. Most of these genes are located in the HLA locus or MAPT locus, and owing to the complex underlying LD structures, it is difficult to define the true causal genes. However, our analyses implicate that, in addition to the PD-associated HLA genes and MAPT in these loci, TRIM31, CRHR1, PLEKHM1, and NSF might also be related to PD through defective components of the immune system. 54-56 For example, methylation levels of TRIM31 seem to be affected in the cerebellum by 2 susceptibility loci (rs9261531 and rs9261535) in TRIM10. It is hypothesized that TRIM family proteins are active in the innate immune response to intracellular infectious agents. 57,58 In addition to known PD loci, there is one novel susceptibility locus that has an effect on methylation levels. This variant (rs7515174) is located in the third intron and affects the frontal cortex methylation state of FCGR2A. This gene encodes a protein belonging to the IgGFc receptor gene family in which the encoded proteins are located on the surface of many immune response cells and take part in clearing of immune complexes and phagocytosis. 59-61 Variants in FCGR2A have been associated with inflammatory bowel disease, Crohn disease, and ulcerative colitis, 62 and variants in other genes from the same family were associated with RA.⁶³ Of further interest are the 8 identified genes in which the expression is regulated by 10 pleiotropic SNPs (from 5 loci) in several specific immune cell types. 46 Five of the 8 genes are located in 3 loci previously associated with PD.3 For example, SNCA expression is regulated by a pleiotropic variant (rs2736990) in intron 2 of SNCA in monocytes. This finding seems in line with a previous study⁶⁴ describing an increase of monocytes in peripheral blood of patients with PD, implying an immune-related manifestation of PD through monocytes. RNPS1, DMPK, and DMWD (with the latter 2 genes involved in myotonic dystrophy⁶⁵) are 3 of the 8 immune-based eQTL that are newly associated with PD in the present study. The biological functions of these genes involve messenger RNA modification and intracellular trafficking or are unknown. ^{66,67}

We used pathway analyses to discern the underlying relevant pathways; however, functional studies are pertinent to provide biological insight. Performing downstream pathogenetic analyses using cell-based models is beyond the scope of the present work. Nevertheless, we anticipate that the genetic loci highlighted in our study will motivate the scientific community to pursue this line of research.

The strong pleiotropic enrichment observed between PD and Crohn disease suggests a common pathogenetic link between these 2 phenotypes. Previous studies^{21,48} highlighted LRRK2 as a significant risk factor for both of these phenotypes. LRRK2 has been identified as having 2 independent Crohn disease risk loci (rs11564258 and rs3761863)⁶⁸: only one of them is in high LD to our shared susceptibility locus rs17467164 ($r^2 = 0.992$ and $r^2 = 0.075$, respectively). The observed association between PD and Crohn disease indicates that defects in cargo transport mechanism might underline the disease pathogenesis in both phenotypes.⁶⁹ It is known that gastrointestinal tract dysfunction is associated with PD, perhaps even preceding the onset of central motor symptoms.⁷⁰ Several of the identified overlapping genes (CCNY, LRRK2, MAPT, RSPH6A, and SYMPK) are involved in basic cellular functions that may be related to alterations in enteric neurotransmission or intestinal motility disturbances. 71 Furthermore, the shared gene HLA-DQB1 has a central role in the immune system by introducing peptides derived from extracellular proteins, which implicate overlapping factors related to the immune system (CD4 T cells).

We found moderate polygenic pleiotropic enrichment between PD and ulcerative colitis or RA, whereas genetic enrichment with type 1 diabetes, celiac disease, psoriasis, and multiple sclerosis was weak. In comparison, in a populationbased study, 13 the risk for PD was observed to increase in a subset of the cohort with autoimmune diseases and ulcerative colitis and to decrease in those with a previous diagnosis of RA. The epidemiological data in that investigation are in agreement with a recently published study 22 in which genetic comorbidities with PD were explored using top loci from diverse phenotypes. The authors observed a decreased risk for rheumatoid arthritisand psoriasis, but the findings were not statistically significant because of the small sample size. It is unlikely that patients with unrecognized immune-related disorders were included in the PD study¹³ population in large enough numbers to affect the results; however, some participants in the autoimmune disorders population¹³ could develop PD over time.

Inflammation of microglia, the major resident immune cells in the brain, ⁷² has been involved in degeneration of dopaminergic neurons affecting PD. ^{19,73} The extent of genetic pleiotropy observed between PD and autoimmune diseases will help us to understand novel pathogenetic aspects of neurodegeneration in PD, a chronic immune activation of microglia, which may cause or contribute to degeneration of neurons. For example, it has been shown that aggregated

 α -synuclein protein (by overexpression of *SNCA*) activates the microglia, which increases nitration of α -synuclein and maintains the proinflammatory innate immune response in PD. ⁷⁴ In this context, the present findings of a polygenic link between PD and inflammatory biological function are likely to be relevant. Furthermore, recent evidence suggests that immune factors are also involved in other neurodegenerative diseases, such as AD. ²³

Limitations and Strengths

Our pleiotropy-based statistical framework was limited to GWAS with a high coverage of SNPs (>500 000); therefore, autoimmune disorders selected for this study were based on available GWAS data that fit these criteria. With more extensive GWAS data, it would be worthwhile to study a larger set of immune disorders associated with PD from epidemiological or clinical studies (eg, thyroid disease). Our study is also limited in distinguishing between immune-mediated and autoimmune disease. It has been hypothesized that PD itself is an autoimmune disease. Although we have shown herein using GWAS of autoimmune disorders that PD has a strong immune component, the conclusion of the hypothesis that PD is an autoimmune disease should be investigated through further cell-based functional studies.

This work has clinical implications. Our data suggest more extensive clinical studies of patients with immunemediated disorders for PD signs to develop possible screening schemes for PD, and vice versa, for monitoring immune and inflammatory status among individuals with an increased risk for PD.⁷⁶ According to our study, apparently healthy individuals with a high load of these shared risk genotypes, predisposing them to inflammation disturbances, could be at particularly increased risk for developing PD. Further prospective studies in these individuals may clarify these issues. Our findings also suggest the need for further investigation of the role of immune-modulating agents in the treatment of PD. There is some evidence indicating that anti-immune drugs could be a viable option for therapeutic interventions in PD. A 2004 study⁷⁶ showed that candesartan cilexetil, a drug used for hypertension, reduces the α-synuclein-induced microglia phenotype. Therefore, data generated from our study may facilitate novel treatment strategies by increasing our understanding of the pathogenetic mechanisms influenced by pleiotropic disease loci.

Conclusions

In summary, our study highlights the usefulness of crossphenotype analyses to identify genetic overlap (ie, pleiotropic loci) between PD and a selection of autoimmune disorders. Our results suggest that these PD-associated loci contribute to PD through immune defects and that immune dysfunction is not simply the end product of the neurodegeneration process. The findings strongly support the presence of interaction between the immune system and neurodegeneration in PD.

ARTICLE INFORMATION

Accepted for Publication: March 8, 2017. **Published Online:** June 5, 2017.

doi:10.1001/jamaneurol.2017.0469 Author Affiliations: Norwegian Centre for Mental Disorders Research (NORMENT), K. G. Jebsen Centre for Psychosis Research, Institute of Clinical Medicine, University of Oslo, Oslo (Witoelar, Wang, Djurovic, Bettella, Andreassen); Division of Mental Health and Addiction, Oslo University Hospital, Oslo, Norway (Witoelar); Department of Clinical Genetics, Vrije Universiteit (VU) University Medical Center, Amsterdam, the Netherlands (Jansen, Heutink); German Center for Neurodegenerative Diseases (DZNE), Tübingen (Jansen, Blauwendraat, Heutink, Gasser); Multimodal Imaging Laboratory, University of California at San Diego, La Jolla (Wang, Desikan, Schork, McEvoy, Dale); Department of Radiology and Biomedical Imaging, University of California, San Francisco (Desikan); Laboratory of Neurogenetics, National Institute on Aging, Bethesda, Maryland (Gibbs, Hernandez, Singleton); Department of Psychiatry, University of California at San Diego, La Jolla (Thompson, Dale); Department of Psychiatry, University of Copenhagen, Copenhagen, Denmark (Thompson); Department of Medical Genetics, University of Oslo, Oslo, Norway (Djurovic, Lie); Department of Medical Genetics, Oslo University Hospital, Oslo, Norway (Djurovic, Lie); Sciences Graduate Program, University of California at San Diego, La Jolla (Schork); Department of Neurosciences, University of California at San Diego, La Jolla (Schork, Dale): Institute of Clinical Molecular Biology, Christian Albrechts University of Kiel, Kiel, Germany (Ellinghaus, Franke); K. G. Jebsen Inflammation Research Centre, Research Institute of Internal Medicine, Oslo, Norway (Lie, McEvoy, Karlsen): Division of Cancer Medicine, Surgery and Transplantation, Oslo University Hospital Rikshospitalet, Oslo, Norway (Lie, Karlsen); Section of Clinical Immunology and Infectious Diseases, Oslo University Hospital Rikshospitalet, Oslo. Norway (Lie); Division of Gastroenterology, Institute of Medicine, University of Bergen, Bergen, Norway (Karlsen); Norwegian Primary Sclerosing Cholangitis (PSC) Research Center, Department of Transplantation Medicine, Oslo (Karlsen): Sorbonne Universités, Université Pierre-et-Marie Curie (UPMC) Paris O6, UM 1127, Institut du Cerveau et de la Moelle Epinière (ICM), Paris, France (Lesage, Brice); Institut National de la Santé et de la Récherche Médicale (INSERM), Unité 1127, Institut du Cerveau et de la Moelle Epinière (ICM), Paris,

Lila Weston Institute, University College London, London, England (Hardy); Department of Radiology, University of California at San Diego, La Jolla (Dale); Centre for Genetic Epidemiology, Institute for Clinical Epidemiology and Applied Biometry, University of Tübingen, Tübingen, Germany (Sharma).

Author Contributions: Dr Witoelar had full access to all of the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

Study concept and design: Witoelar, Desikan, Thompson, Wood, Dale, Gasser, Andreassen, Sharma.

Acquisition, analysis, or interpretation of data: Witoelar, Jansen, Wang, Gibbs, Blauwendraat, Hernandez, Djurovic, Schork, Bettella, Ellinghaus, Franke, Lie, McEvoy, Karlsen, Lesage, Morris, Brice, Heutink, Hardy, Singleton, Dale, Andreassen, Sharma.

Drafting of the manuscript: Witoelar, Jansen, Desikan, Andreassen, Sharma.

Critical revision of the manuscript for important intellectual content: Witoelar, Wang, Desikan, Gibbs, Blauwendraat, Thompson, Hernandez, Djurovic, Schork, Bettella, Ellinghaus, Franke, Lie, McEvoy, Karlsen, Lesage, Morris, Brice, Wood, Heutink, Hardy, Singleton, Dale, Gasser, Andreassen, Sharma.

Statistical analysis: Witoelar, Jansen, Wang, Thompson, Ellinghaus, Lie, Dale. Obtained funding: Heutink, Hardy, Dale, Gasser, Andreassen. Sharma.

Administrative, technical, or material support: Wang, Desikan, Gibbs, Hernandez, Djurovic, Franke, Singleton, Dale, Andreassen.

Study supervision: Wang, Desikan, Wood, Heutink, Hardy, Dale, Gasser, Andreassen, Sharma.

Conflict of Interest Disclosures: None reported.

Funding/Support: This work was supported by the German Federal Ministry of Education and Research (BMBF) within the framework of the e:Med research and funding concept (SysINFLAME grant OIZX1306A). This project received infrastructure support from the Deutsche

Forschungsgemeinschaft (DFG) Excellence Cluster 306 "Inflammation at Interfaces." Dr Jansen receives funding from Prinses Beatrix Fonds. Dr Franke receives an endowment professorship by the Foundation for Experimental Medicine (Zurich, Switzerland). Dr Andreassen and his team are supported by The Research Council of Norway (213837, 225989, 223273, and 475 237250/EU Joint Programme-Neurodegenerative Disease Research [EU-JPND]), the South East Norway Health Authority (2013-123), the Norwegian Health Association, and the K. G. Jebsen Foundation. Dr Sharma receives funding from The Michael J Fox Foundation for Parkinson's Research and the EU-JPND program (Comprehensive Unbiased Risk Factor Assessment for Genetics and Environment in Parkinson's Disease [COURAGE-PD]).

Role of the Funder/Sponsor: The funding sources had no role in the design and conduct of the study; collection, management, analysis, or interpretation of the data; preparation, review, or approval of the manuscript; and decision to submit the manuscript for publication.

Group Information: The International Parkinson's Disease Genomics Consortium (IPDGC), North

American Brain Expression Consortium (NABEC), and United Kingdom Brain Expression Consortium (UKBEC) investigators are as follows:

IPDGC: Mike A. Nalls (Laboratory of Neurogenetics, National Institute on Aging, National Institutes of Health, Bethesda, Maryland), Vincent Plagnol (UCL Genetics Institute, London, England), Dena G. Hernandez (Laboratory of Neurogenetics, National Institute on Aging, Bethesda, Maryland; and Department of Molecular Neuroscience, UCL Institute of Neurology, London, England), Manu Sharma (Department for Neurodegenerative Diseases, Hertie Institute for Clinical Brain Research, University of Tübingen, Tübingen, Germany; and DZNE, German Center for Neurodegenerative Diseases, Tübingen), Una-Marie Sheerin (Department of Molecular Neuroscience, UCL Institute of Neurology, London, England), Mohamad Saad (INSERM U563, CPTP, Toulouse, France; and Paul Sabatier University, Toulouse, France). Javier Simón-Sánchez (Department for Neurodegenerative Diseases, Hertie Institute for Clinical Brain Research, University of Tübingen, Tübingen, Germany; and DZNE, German Center for Neurodegenerative Diseases, Tübingen, Germany), Iris E. Jansen (Department of Clinical Genetics, VU University Medical Center, Amsterdam, the Netherlands), Claudia Schulte (Department for Neurodegenerative Diseases, Hertie Institute for Clinical Brain Research, University of Tübingen, Tübingen, Germany), Suzanne Lesage (INSERM, UMR_S975 [formerly UMR_S679], Paris, France; Université Pierre et Marie Curie-Paris, Centre de Recherche de l'Institut du Cerveau et de la Moelle Épinière, Paris, France; and CNRS, Paris, France), Sigurlaug Sveinbiörnsdóttir (Department of Neurology, Landspítali University Hospital, Reykjavík, Iceland; Department of Neurology, MEHT Broomfield Hospital, Chelmsford, Essex. England; and Queen Mary College, University of London, London, England), Sampath Arepalli (Laboratory of Neurogenetics, National Institute on Aging, Bethesda, Maryland), Roger Barker (Department of Neurology, Addenbrooke's Hospital, University of Cambridge, Cambridge, England), Yoav Ben-Shlomo (School of Social and Community Medicine, University of Bristol, Bristol, England), Henk W. Berendse (Department of Neurology and Alzheimer Center, VU University Medical Center, Amsterdam, the Netherlands), Daniela Berg (Department for Neurodegenerative Diseases, Hertie Institute for Clinical Brain Research, University of Tübingen, Tübingen, Germany; and DZNE, German Center for Neurodegenerative Diseases, Tübingen), Kailash Bhatia (Department of Motor Neuroscience, UCL Institute of Neurology, London, England), Rob M. A. de Bie (Department of Neurology, Academic Medical Center, University of Amsterdam, Amsterdam, the Netherlands), Alessandro Biffi (Center for Human Genetic Research and Department of Neurology, Massachusetts General Hospital, Boston; and Program in Medical and Population Genetics, Broad Institute, Cambridge, Massachusetts), Bas Bloem (Department of Neurology, Radboud University Nijmegen Medical Centre, Nijmegen, the Netherlands), Zoltan Bochdanovits (Department of Clinical Genetics, Section of Medical Genomics, VU University Medical Centre, Amsterdam, the Netherlands), Michael Bonin (Department of Medical Genetics,

Tübingen, Germany (Heutink, Gasser, Sharma); Rita

France (Lesage, Brice); Centre National de la

Recherche Scientifique (CNRS) UMR 7225, Institut

France (Lesage, Brice); Institut du Cerveau et de la

Moelle Epinière (ICM), Paris, France (Lesage, Brice);

Assistance Publique-Hôpitaux de Paris, Hôpital de

Hospital for Neurology and Neurosurgery (NHNN),

(Morris); Department of Molecular Neurosciences,

Institute of Neurology, University College London,

Neurodegenerative Diseases, Hertie Institute for

Clinical Brain Research, University of Tübingen,

la Salpêtrière, Département de Génétique et

Cytogénétique, Paris, France (Lesage, Brice);

Department of Clinical Neuroscience, National

University College London, London, England

London, England (Wood); Department of

du Cerveau et de la Moelle Epinière (ICM), Paris,

Institute of Human Genetics, University of Tübingen, Tübingen, Germany), Jose M. Bras (Department of Molecular Neuroscience, UCL Institute of Neurology, London, England), Kathrin Brockmann (Department for Neurodegenerative Diseases, Hertie Institute for Clinical Brain Research, University of Tübingen, Tübingen, Germany: and DZNE. German Center for Neurodegenerative Diseases, Tübingen, Germany), Janet Brooks (Laboratory of Neurogenetics, National Institute on Aging, Bethesda, Maryland), David J. Burn (Newcastle University Clinical Ageing Research Unit, Campus for Ageing and Vitality, Newcastle Upon Tyne, England), Elisa Majounie (Laboratory of Neurogenetics, National Institute on Aging, Bethesda, Maryland), Gavin Charlesworth (Department of Molecular Neuroscience, UCL Institute of Neurology, London, England), Codrin Lungu (National Institutes of Health Parkinson Clinic, NINDS, National Institutes of Health, Bethesda, Maryland), Honglei Chen (Epidemiology Branch, National Institute of Environmental Health Sciences, National Institutes of Health, Research Triangle Park, North Carolina), Patrick F. Chinnery (Neurology M4104, The Medical School, Framlington Place, Newcastle Upon Tyne, England), Sean Chong (Laboratory of Neurogenetics, National Institute on Aging, Bethesda, Maryland), Carl E. Clarke (School of Clinical and Experimental Medicine, University of Birmingham, Birmingham, England; and Department of Neurology, City Hospital, Sandwell and West Birmingham Hospitals NHS Trust, Birmingham, England), Mark R. Cookson (Laboratory of Neurogenetics, National Institute on Aging, Bethesda, Maryland), J. Mark Cooper (Department of Clinical Neurosciences, UCL Institute of Neurology, London, England), Jean Christophe Corvol (INSERM LIMR S975 Paris France; Université Pierre et Marie Curie-Paris, Paris, France: CNRS. Paris. France: and INSERM CIC-9503. Hôpital Pitié-Salpêtrière, Paris, France), Carl Counsell (University of Aberdeen, Division of Applied Health Sciences, Population Health Section, Aberdeen, Scotland), Philippe Damier (CHU Nantes, CICOOO4, Service de Neurologie, Nantes, France), Jean-François Dartigues (INSERM U897, Université Victor Segalen, Bordeaux, France), Panos Deloukas (Wellcome Trust Sanger Institute, Wellcome Trust Genome Campus, Cambridge, England). Günther Deuschl (Klinik für Neurologie. Universitätsklinikum Schleswig-Holstein, Campus Kiel, Christian Albrechts University of Kiel, Kiel, Germany), David T. Dexter (Parkinson's Disease Research Group, Faculty of Medicine, Imperial College London, London, England), Karin D. van Dijk (Department of Neurology and Alzheimer Center, VU University Medical Center, Amsterdam, the Netherlands), Allissa Dillman (Laboratory of Neurogenetics, National Institute on Aging, Bethesda, Maryland), Frank Durif (Service de Neurologie, Hôpital Gabriel Montpied, Clermont-Ferrand, France), Alexandra Dürr (INSERM, UMR S975, Paris, France; Université Pierre et Marie Curie-Paris; CNRS, Paris, France; and AP-HP, Pitié Salpêtrière Hospital, Paris, France), Sarah Edkins (Wellcome Trust Sanger Institute, Cambridge, England), Jonathan R. Evans (Cambridge Centre for Brain Repair, Cambridge, England), Thomas Foltynie (UCL Institute of Neurology, London, England), Jing Dong (Epidemiology Branch, National Institute of Environmental Health Sciences, Research Triangle Park, North Carolina), Michelle Gardner (Department of Molecular Neuroscience, UCL

Institute of Neurology, London, England), J. Raphael Gibbs (Laboratory of Neurogenetics, National Institute on Aging, Bethesda, Maryland: and Department of Molecular Neuroscience, UCL Institute of Neurology, London, England), Alison Goate (Departments of Psychiatry and Neurology, Washington University School of Medicine in St Louis, St Louis, Missouri), Emma Grav (Wellcome Trust Sanger Institute, Cambridge, England), Rita Guerreiro (Department of Molecular Neuroscience. UCL Institute of Neurology, London, England), Clare Harris (University of Aberdeen, Aberdeen, Scotland), Jacobus J. van Hilten (Department of Neurology, Leiden University Medical Center, Leiden, the Netherlands), Albert Hofman (Department of Epidemiology, Erasmus University Medical Center, Rotterdam, the Netherlands), Albert Hollenbeck (AARP, Washington, DC), Janice Holton (Queen Square Brain Bank for Neurological Disorders, UCL Institute of Neurology, London, England), Michele Hu (Department of Clinical Neurology, John Radcliffe Hospital, Oxford, England), Xuemei Huang (Departments of Neurology, Radiology, Neurosurgery, Pharmacology, Kinesiology, and Bioengineering, Pennsylvania State University-Milton S. Hershey Medical Center, Hershey), Isabel Wurster (Department for Neurodegenerative Diseases, Hertie Institute for Clinical Brain Research, University of Tübingen, Tübingen, Germany; and German Center for Neurodegenerative Diseases, Tübingen, Germany), Walter Mätzler (Department for Neurodegenerative Diseases. Hertie Institute for Clinical Brain Research, University of Tübingen, Tübingen, Germany; and German Center for Neurodegenerative Diseases Tübingen, Germany), Gavin Hudson (Neurology M4104, The Medical School, Newcastle Upon Tyne, England), Sarah E. Hunt (Wellcome Trust Sanger Institute, Cambridge, England), Johanna Huttenlocher (deCODE genetics, Reykjavík, Iceland), Thomas Illig (Institute of Epidemiology, Helmholtz Zentrum München, German Research Centre for Environmental Health, Neuherberg), Pálmi V. Jónsson (Department of Geriatrics, Landspítali University Hospital. Reykjavík, Iceland), Jean-Charles Lambert (INSERM U744, Lille, France; and Institut Pasteur de Lille, Université de Lille Nord, Lille, France), Cordelia Langford (Cambridge Centre for Brain Repair, Cambridge, England), Andrew Lees (Oueen Square Brain Bank for Neurological Disorders, UCL Institute of Neurology, London, England), Peter Lichtner (Institute of Human Genetics, Helmholtz Zentrum München, German Research Centre for Environmental Health, Neuherberg), Patricia Limousin (Institute of Neurology, Sobell Department, Unit of Functional Neurosurgery, London, England), Grisel Lopez (Section on Molecular Neurogenetics, Medical Genetics Branch, NHGRI, National Institutes of Health, Bethesda. Maryland), Delia Lorenz (Klinik für Neurologie, Universitätsklinikum Schleswig-Holstein, Kiel, Germany), Codrin Lungu (National Institutes of Health Parkinson Clinic, NINDS, National Institutes of Health, Bethesda, Maryland), Alisdair McNeill (Department of Clinical Neurosciences, UCL Institute of Neurology, London, England), Catriona Moorby (School of Clinical and Experimental Medicine, University of Birmingham, Birmingham, England). Matthew Moore (Laboratory of Neurogenetics, National Institute on Aging, Bethesda, Maryland), Huw R. Morris (National

University College London, London, England), Karen E. Morrison (School of Clinical and Experimental Medicine. University of Birmingham. Birmingham, England; and Neurosciences Department, Queen Elizabeth Hospital, University Hospitals Birmingham NHS Foundation Trust, Birmingham, England), Valentina Escott-Price (MRC Centre for Neuropsychiatric Genetics and Genomics, Cardiff University School of Medicine, Cardiff, England), Ese Mudanohwo (Neurogenetics Unit, UCL Institute of Neurology, London, England; and National Hospital for Neurology and Neurosurgery, University College London, London, England), Sean S. O'Sullivan (Queen Square Brain Bank for Neurological Disorders, UCL Institute of Neurology, London, England), Justin Pearson (MRC Centre for Neuropsychiatric Genetics and Genomics, Cardiff University School of Medicine, Cardiff, England), Joel S. Perlmutter (Departments of Neurology, Radiology, and Neurobiology at Washington University in St Louis, St Louis, Missouri), Hjörvar Pétursson (deCODE genetics, Reykjavík, Iceland; and Department of Medical Genetics, Institute of Human Genetics, University of Tübingen, Tübingen, Germany), Pierre Pollak (Service de Neurologie, CHU de Grenoble, Grenoble, France), Bart Post (Department of Neurology, Radboud University Nijmegen Medical Centre, Nijmegen, the Netherlands), Simon Potter (Wellcome Trust Sanger Institute, Cambridge, England), Bernard Ravina (Translational Neurology, Biogen Idec, Cambridge, Massachusetts), Tamas Revesz (Oueen Square Brain Bank for Neurological Disorders, UCL Institute of Neurology, London, England), Olaf Riess (Department of Medical Genetics, Institute of Human Genetics, University of Tübingen, Tübingen, Germany), Fernando Rivadeneira (Departments of Epidemiology and Internal Medicine, Erasmus University Medical Center, Rotterdam, the Netherlands), Patrizia Rizzu (Department of Clinical Genetics, Section of Medical Genomics, VU University Medical Centre, Amsterdam, the Netherlands), Mina Ryten (Department of Molecular Neuroscience, UCL Institute of Neurology, London, England), Stephen Sawcer (University of Cambridge, Department of Clinical Neurosciences, Addenbrooke's Hospital, Cambridge, England), Anthony Schapira (Department of Clinical Neurosciences, UCL Institute of Neurology, London, England), Hans Scheffer (Department of Human Genetics, Radboud University Nijmegen Medical Centre, Nijmegen, the Netherlands), Karen Shaw (Queen Square Brain Bank for Neurological Disorders, UCL Institute of Neurology, London, England), Ira Shoulson (Department of Neurology, University of Rochester, Rochester, New York), Joshua Shulman (Baylor College of Medicine, Houston, Texas), Ellen Sidransky (Section on Molecular Neurogenetics, Medical Genetics Branch, NHGRI, National Institutes of Health, Bethesda, Maryland), Colin Smith (Department of Pathology, University of Edinburgh, Edinburgh, Scotland), Chris C. A. Spencer (Wellcome Trust Centre for Human Genetics, Oxford, England), Hreinn Stefánsson (deCODE genetics, Reykjavík, Iceland), Francesco Bettella (deCODE genetics, Reykjavík, Iceland), Joanna D. Stockton (School of Clinical and Experimental Medicine, University of Birmingham, Birmingham, England), Amy Strange (Wellcome Trust Centre for Human Genetics, Oxford, England), Kevin Talbot (University of Oxford, Department of Clinical Neurology, John Radcliffe Hospital, Oxford,

Hospital for Neurology and Neurosurgery,

England), Carlie M. Tanner (Clinical Research Department, The Parkinson's Institute and Clinical Center, Sunnyvale, California), Avazeh Tashakkori-Ghanbaria (Wellcome Trust Sanger Institute, Cambridge, England), François Tison (Service de Neurologie, Hôpital Haut-Lévêque, Pessac, France), Daniah Trabzuni (Department of Molecular Neuroscience, UCL Institute of Neurology, London, England), Bryan J. Traynor (Laboratory of Neurogenetics, National Institute on Aging, Bethesda, Maryland), André G. Uitterlinden (Departments of Epidemiology and Internal Medicine. Erasmus University Medical Center. Rotterdam, the Netherlands), Daan Velseboer (Department of Neurology, Academic Medical Center, University of Amsterdam, Amsterdam, the Netherlands), Marie Vidailhet (INSERM, UMR_S975, Université Pierre et Marie Curie-Paris, CNRS, UMR 7225, Paris, France), Robert Walker (Department of Pathology, University of Edinburgh, Edinburgh, Scotland), Bart van de Warrenburg (Department of Neurology, Radboud University Nijmegen Medical Centre, Nijmegen, the Netherlands), Mirdhu Wickremaratchi (Department of Neurology, Cardiff University, Cardiff, England), Nigel Williams (MRC Centre for Neuropsychiatric Genetics and Genomics, Cardiff University School of Medicine, Cardiff, England), Caroline H Williams-Gray (Department of Neurology, Addenbrooke's Hospital, University of Cambridge, Cambridge, England), Sophie Winder-Rhodes (Department of Psychiatry and Medical Research Council and Wellcome Trust Behavioural and Clinical Neurosciences Institute, University of Cambridge, Cambridge, England), Kári Stefánsson (deCODE genetics, Reykjavík, Iceland), Maria Martinez (INSERM UMR 1043, Paul Sabatier University, Toulouse, France), Nicholas W. Wood (UCL Genetics Institute, London, England; and Department of Molecular Neuroscience, UCL Institute of Neurology, London, England), John Hardy (Department of Molecular Neuroscience, UCL Institute of Neurology, London, England), Peter Heutink (DZNE, German Center for Neurodegenerative Diseases. Tübingen: and Department for Neurodegenerative Diseases, Hertie Institute for Clinical Brain Research, University of Tübingen, Tübingen, Germany), Alexis Brice (INSERM, UMR_S975, Université Pierre et Marie Curie-Paris, CNRS, UMR 7225, AP-HP, Pitié-Salpêtrière Hospital, Paris, France), Thomas Gasser (Department for Neurodegenerative Diseases, Hertie Institute for Clinical Brain Research, and DZNE, German Center for Neurodegenerative Diseases, Tübingen, Germany), and Andrew B. Singleton (Laboratory of Neurogenetics, National Institute on Aging, Bethesda, Maryland).

NABEC: Sampath Arepalli (Laboratory of Neurogenetics, National Institute on Aging, National Institutes of Health, Bethesda, Maryland), Mark R. Cookson (Laboratory of Neurogenetics, National Institute on Aging, National Institutes of Health, Bethesda, Maryland), Allissa Dillman (Laboratory of Neurogenetics, National Institute on Aging, National Institutes of Health, Bethesda, Maryland), Luigi Ferrucci (Clinical Research Branch, National Institute on Aging, Baltimore, Maryland), J. Raphael Gibbs (Laboratory of Neurogenetics, National Institute on Aging, National Institutes of Health, Bethesda, Maryland; and Department of Molecular Neuroscience, UCL Institute of Neurology, London, England), Dena G. Hernandez (Laboratory of Neurogenetics, National Institute on

Aging, National Institutes of Health, Bethesda, Maryland; and German Center for Neurodegenerative Diseases (DZNE)-Tübingen. Tübingen, Germany), Robert Johnson (NICHD Brain and Tissue Bank for Developmental Disorders, University of Maryland Medical School, Baltimore). Dan L. Longo (Lymphocyte Cell Biology Unit, Laboratory of Immunology, National Institute on Aging, National Institutes of Health, Baltimore, Maryland), Michael A. Nalls (Laboratory of Neurogenetics, National Institute on Aging, National Institutes of Health, Bethesda, Maryland), Richard O'Brien (Brain Resource Center, The Johns Hopkins University, Baltimore, Maryland), Andrew Singleton (Laboratory of Neurogenetics, National Institute on Aging, National Institutes of Health, Bethesda, Maryland), Bryan Traynor (Laboratory of Neurogenetics, National Institute on Aging, National Institutes of Health, Bethesda, Maryland), Juan Troncoso (Brain Resource Center, The Johns Hopkins University, Baltimore, Maryland), Marcel van der Brug (Laboratory of Neurogenetics, National Institute on Aging, National Institutes of Health, Bethesda, Maryland; and ITGR Biomarker Discovery Group, Genentech, South San Francisco, California), H. Ronald Zielke (NICHD Brain and Tissue Bank for Developmental Disorders, University of Maryland Medical School, Baltimore), and Alan Zonderman (Research Resources Branch, National Institute on Aging, National Institutes of Health, Bethesda, Maryland).

UKBEC: John A. Hardy (Department of Molecular Neuroscience, UCL Institute of Neurology, London, England), Mina Ryten (Department of Molecular Neuroscience, UCL Institute of Neurology, London, England), Colin Smith (Department of Pathology, The University of Edinburgh, Edinburgh, Scotland), Daniah Trabzuni (Department of Molecular Neuroscience, UCL Institute of Neurology, London, England), Robert Walker (Department of Pathology, The University of Edinburgh, Edinburgh, Scotland), and Mike Weale (Department of Medical & Molecular Genetics, King's College London, London, England).

Additional Contributions: We thank Cisca Wijmenga, PhD (Department of Genetics, University of Groningen), David van Heel, PhD (Centre for Genomics and Child Health, Queen Mary University of London), Annegret Fischer, PhD (Sarcoidosis Research, Christian Albrechts University of Kiel), Eva Ellinghaus, PhD (Genetics & Bioinformatics, Christian Albrechts University of Kiel), the International Inflammatory Bowel Disease Genetics Consortium (IIBDGC), and the Psoriasis Association Genetics Extension (PAGE) for access to summary results data. No compensation was received.

REFERENCES

- 1. Simón-Sánchez J, Schulte C, Bras JM, et al. Genome-wide association study reveals genetic risk underlying Parkinson's disease. *Nat Genet*. 2009; 41(12):1308-1312.
- Keller MF, Saad M, Bras J, et al; International Parkinson's Disease Genomics Consortium (IPDGC); Wellcome Trust Case Control Consortium 2 (WTCCC2). Using genome-wide complex trait analysis to quantify "missing heritability" in Parkinson's disease. *Hum Mol Genet*. 2012;21(22): 4996-5009.
- **3**. Nalls MA, Pankratz N, Lill CM, et al; International Parkinson's Disease Genomics Consortium (IPDGC);

- Parkinson's Study Group (PSG) Parkinson's
 Research: The Organized Genetics Initiative
 (PROGENI); 23andMe; GenePD; NeuroGenetics
 Research Consortium (NGRC); Hussman Institute of
 Human Genomics (HIHG); Ashkenazi Jewish
 Dataset Investigator; Cohorts for Health and Aging
 Research in Genetic Epidemiology (CHARGE);
 North American Brain Expression Consortium
 (NABEC); United Kingdom Brain Expression
 Consortium (UKBEC); Greek Parkinson's Disease
 Consortium; Alzheimer Genetic Analysis Group.
 Large-scale meta-analysis of genome-wide
 association data identifies six new risk loci for
 Parkinson's disease. Nat Genet. 2014;46(9):989-993.
- 4. Nalls MA, Plagnol V, Hernandez DG, et al; International Parkinson Disease Genomics Consortium. Imputation of sequence variants for identification of genetic risks for Parkinson's disease: a meta-analysis of genome-wide association studies. *Lancet*. 2011;377(9766):641-649.
- Holmans P, Moskvina V, Jones L, et al.
 A pathway-based analysis provides additional support for an immune-related genetic susceptibility to Parkinson's disease. Hum Mol Genet. 2012.
- **6**. McCarthy MI, Abecasis GR, Cardon LR, et al. Genome-wide association studies for complex traits: consensus, uncertainty and challenges. *Nat Rev Genet*. 2008:9(5):356-369.
- 7. Khandelwal PJ, Herman AM, Moussa CE. Inflammation in the early stages of neurodegenerative pathology. *J Neuroimmunol*. 2011;238(1-2):1-11.
- **8**. Akiyama H, Barger S, Barnum S, et al. Inflammation and Alzheimer's disease. *Neurobiol Aging*. 2000;21(3):383-421.
- **9.** Phani S, Loike JD, Przedborski S. Neurodegeneration and inflammation in Parkinson's disease. *Parkinsonism Relat Disord*. 2012;18(suppl 1):5207-5209.
- **10**. McGeer PL, McGeer EG. Inflammation and neurodegeneration in Parkinson's disease. *Parkinsonism Relat Disord*. 2004;10(suppl 1):S3-S7.
- 11. Arai H, Furuya T, Mizuno Y, Mochizuki H. Inflammation and infection in Parkinson's disease. *Histol Histopathol*. 2006;21(6):673-678.
- 12. Li X, Sundquist J, Sundquist K. Subsequent risks of Parkinson disease in patients with autoimmune and related disorders: a nationwide epidemiological study from Sweden. *Neurodegener Dis.* 2012;10(1-4):277-284
- **13**. Rugbjerg K, Friis S, Ritz B, Schernhammer ES, Korbo L, Olsen JH. Autoimmune disease and risk for Parkinson disease: a population-based case-control study. *Neurology*. 2009;73(18):1462-1468.
- **14.** Pedemonte E, Trabucco E, Cella M, Solaro C. Parkinsonism in multiple sclerosis patients: a casual or causal association? *Parkinsonism Relat Disord*. 2013:19(4):492-493.
- **15.** Vieregge P, Klostermann W, Brückmann H. Parkinsonism in multiple sclerosis. *Mov Disord*. 1992;7(4):380-382.
- **16.** Nielsen NM, Pasternak B, Stenager E, Koch-Henriksen N, Frisch M. Multiple sclerosis and risk of Parkinson's disease: a Danish nationwide cohort study. *Eur J Neurol*. 2014;21(1):107-111. Medline:24053187
- **17**. Chen H, Jacobs E, Schwarzschild MA, et al. Nonsteroidal antiinflammatory drug use and the

- risk for Parkinson's disease. *Ann Neurol*. 2005;58 (6):963-967.
- **18**. Chen H, Zhang SM, Hernán MA, et al. Nonsteroidal anti-inflammatory drugs and the risk of Parkinson disease. *Arch Neurol*. 2003;60(8): 1059-1064.
- **19.** Mosley RL, Hutter-Saunders JA, Stone DK, Gendelman HE. Inflammation and adaptive immunity in Parkinson's disease. *Cold Spring Harb Perspect Med*. 2012;2(1):a009381.
- **20**. Zimprich A, Biskup S, Leitner P, et al. Mutations in *LRRK2* cause autosomal-dominant parkinsonism with pleomorphic pathology. *Neuron*. 2004;44 (4):601-607.
- 21. Barrett JC, Hansoul S, Nicolae DL, et al; NIDDK IBD Genetics Consortium; Belgian-French IBD Consortium; Wellcome Trust Case Control Consortium. Genome-wide association defines more than 30 distinct susceptibility loci for Crohn's disease. *Nat Genet*. 2008;40(8):955-962.
- 22. Nalls MA, Saad M, Noyce AJ, et al; International Parkinson's Disease Genomics Consortium (IPDGC); Wellcome Trust Case Control Consortium 2 (WTCCC2); North American Brain Expression Consortium (NABEC); United Kingdom Brain Expression Consortium (UKBEC). Genetic comorbidities in Parkinson's disease. *Hum Mol Genet*. 2014;23(3):831-841.
- **23**. Yokoyama JS, Wang Y, Schork AJ, et al; Alzheimer's Disease Neuroimaging Initiative. Association between genetic traits for immune-mediated diseases and Alzheimer disease. *JAMA Neurol.* 2016;73(6):691-697.
- 24. Andreassen OA, Djurovic S, Thompson WK, et al; International Consortium for Blood Pressure GWAS; Diabetes Genetics Replication and Meta-analysis Consortium; Psychiatric Genomics Consortium Schizophrenia Working Group. Improved detection of common variants associated with schizophrenia by leveraging pleiotropy with cardiovascular-disease risk factors. *Am J Hum Genet*. 2013;92(2):197-209.
- 25. Andreassen OA, Thompson WK, Schork AJ, et al; Psychiatric Genomics Consortium (PGC); Bipolar Disorder and Schizophrenia Working Groups. Improved detection of common variants associated with schizophrenia and bipolar disorder using pleiotropy-informed conditional false discovery rate. *PLoS Genet*. 2013;9(4):e1003455.
- 26. Andreassen OA, Harbo HF, Wang Y, et al; Psychiatric Genomics Consortium (PGC) Bipolar Disorder and Schizophrenia Work Groups; International Multiple Sclerosis Genetics Consortium (IMSGC). Genetic pleiotropy between multiple sclerosis and schizophrenia but not bipolar disorder: differential involvement of immune-related gene loci. *Mol Psychiatry*. 2015;20 (2):207-214.
- **27**. Franke A, McGovern DP, Barrett JC, et al. Genome-wide meta-analysis increases to 71 the number of confirmed Crohn's disease susceptibility loci. *Nat Genet*. 2010;42(12):1118-1125.
- **28**. Anderson CA, Boucher G, Lees CW, et al. Meta-analysis identifies 29 additional ulcerative colitis risk loci, increasing the number of confirmed associations to 47. *Nat Genet*. 2011;43(3):246-252.
- **29**. Stahl EA, Raychaudhuri S, Remmers EF, et al; BIRAC Consortium; YEAR Consortium. Genome-wide association study meta-analysis

- identifies seven new rheumatoid arthritis risk loci. *Nat Genet*. 2010;42(6):508-514.
- **30.** Barrett JC, Clayton DG, Concannon P, et al; Type 1 Diabetes Genetics Consortium. Genome-wide association study and meta-analysis find that over 40 loci affect risk of type 1 diabetes. *Nat Genet.* 2009;41(6):703-707.
- **31**. Dubois PC, Trynka G, Franke L, et al. Multiple common variants for celiac disease influencing immune gene expression. *Nat Genet*. 2010;42(4): 295-302.
- **32.** Ellinghaus D, Ellinghaus E, Nair RP, et al. Combined analysis of genome-wide association studies for Crohn disease and psoriasis identifies seven shared susceptibility loci. *Am J Hum Genet*. 2012;90(4):636-647.
- **33.** Sawcer S, Hellenthal G, Pirinen M, et al; International Multiple Sclerosis Genetics Consortium; Wellcome Trust Case Control Consortium 2. Genetic risk and a primary role for cell-mediated immune mechanisms in multiple sclerosis. *Nature*. 2011;476(7359):214-219.
- **34**. Efron B. Large-Scale Inference: Empirical Bayes Methods for Estimation, Testing and Prediction. New York, NY: Cambridge University Press; 2010.
- **35**. Schweder T, Spjotvoll E. Plots of P-values to evaluate many tests simultaneously. *Biometrika*. 1982:69(3):493-502.
- **36.** Liu JZ, Hov JR, Folseraas T, et al; UK-PSCSC Consortium; International PSC Study Group; International IBD Genetics Consortium. Dense genotyping of immune-related disease regions identifies nine new risk loci for primary sclerosing cholangitis. *Nat Genet*. 2013;45(6):670-675.
- **37**. Desikan RS, Schork AJ, Wang Y, et al; ADNI, ADGC, GERAD, CHARGE and IPDGC Investigators. Genetic overlap between Alzheimer's disease and Parkinson's disease at the *MAPT* locus. *Mol Psychiatry*. 2015;20(12):1588-1595.
- **38**. Nalls MA, Bras J, Hernandez DG, et al. NeuroX, a fast and efficient genotyping platform for investigation of neurodegenerative diseases. *Neurobiol Aging*. 2014.
- **39**. Gibbs JR, van der Brug MP, Hernandez DG, et al. Abundant quantitative trait loci exist for DNA methylation and gene expression in human brain. *PLoS Genet*. 2010;6(5):e1000952.
- **40**. Ramasamy A, Trabzuni D, Guelfi S, et al; UK Brain Expression Consortium; North American Brain Expression Consortium. Genetic variability in the regulation of gene expression in ten regions of the human brain. *Nat Neurosci.* 2014;17(10):1418-1428.
- **41**. Blauwendraat C, Francescatto M, Gibbs JR, et al. Comprehensive promoter level expression quantitative trait loci analysis of the human frontal lobe. *Genome Med*. 2016;8(1):65.
- **42**. Szklarczyk D, Franceschini A, Wyder S, et al. STRING v10: protein-protein interaction networks, integrated over the tree of life. *Nucleic Acids Res*. 2015;43(database issue):D447-D452.
- **43**. Wider C, Vilariño-Güell C, Jasinska-Myga B, et al. Association of the *MAPT* locus with Parkinson's disease. *Eur J Neurol*. 2010;17(3):483-486.
- **44.** Wissemann WT, Hill-Burns EM, Zabetian CP, et al. Association of Parkinson disease with structural and regulatory variants in the HLA region. *Am J Hum Genet*. 2013;93(5):984-993.

- **45**. Ahmed I, Tamouza R, Delord M, et al. Association between Parkinson's disease and the *HLA-DRB1* locus. *Mov Disord*. 2012;27(9):1104-1110.
- **46**. Peters JE, Lyons PA, Lee JC, et al. Insight into genotype-phenotype associations through eQTL mapping in multiple cell types in health and immune-mediated disease. *PLoS Genet*. 2016;12(3): e1005908.
- **47**. Teslovich TM, Musunuru K, Smith AV, et al. Biological, clinical and population relevance of 95 loci for blood lipids. *Nature*. 2010;466(7307):707-713.
- **48**. Liu Z, Lenardo MJ. The role of *LRRK2* in inflammatory bowel disease. *Cell Res*. 2012;22(7): 1092-1094
- **49**. International Parkinson's Disease Genomics Consortium (IPDGC); Wellcome Trust Case Control Consortium 2 (WTCCC2). A two-stage meta-analysis identifies several new loci for Parkinson's disease. *PLoS Genet*. 2011;7(6):e1002142.
- **50**. Wang JY, Gong MY, Ye YL, et al. The *RIT2* and *STX1B* polymorphisms are associated with Parkinson's disease. *Parkinsonism Relat Disord*. 2015;21(3):300-302.
- **51.** Cotsapas C, Voight BF, Rossin E, et al; FOCiS Network of Consortia. Pervasive sharing of genetic effects in autoimmune disease. *PLoS Genet*. 2011;7 (8):e1002254.
- **52.** Gao X, Simon KC, Han J, Schwarzschild MA, Ascherio A. Genetic determinants of hair color and Parkinson's disease risk. *Ann Neurol.* 2009;65(1): 76-82.
- **53.** Maaser C, Kannengiesser K, Specht C, et al. Crucial role of the melanocortin receptor MC1R in experimental colitis. *Gut.* 2006;55(10):1415-1422.
- **54.** Webster EL, Torpy DJ, Elenkov IJ, Chrousos GP. Corticotropin-releasing hormone and inflammation. *Ann N Y Acad Sci.* 1998;840:21-32.
- **55.** McEwan DG, Richter B, Claudi B, et al. PLEKHM1 regulates *Salmonella*-containing vacuole biogenesis and infection. *Cell Host Microbe*. 2015;17 (1):58-71.
- **56**. Stow JL, Manderson AP, Murray RZ. SNAREing immunity: the role of SNAREs in the immune system. *Nat Rev Immunol*. 2006;6(12):919-929.
- **57.** Nisole S, Stoye JP, Saïb A. TRIM family proteins: retroviral restriction and antiviral defence. *Nat Rev Microbiol.* 2005;3(10):799-808.
- **58**. Kawai T, Akira S. Regulation of innate immune signalling pathways by the tripartite motif (TRIM) family proteins. *EMBO Mol Med*. 2011;3(9):513-527.
- **59.** Indik ZK, Park JG, Hunter S, Schreiber AD. The molecular dissection of Fc gamma receptor mediated phagocytosis. *Blood*. 1995;86(12):4389-4399.
- **60**. Dijstelbloem HM, van de Winkel JG, Kallenberg CG. Inflammation in autoimmunity: receptors for IgG revisited. *Trends Immunol*. 2001;22(9):510-516.
- **61**. Gessner JE, Heiken H, Tamm A, Schmidt RE. The IgG Fc receptor family. *Ann Hematol*. 1998;76 (6):231-248
- **62.** Weersma RK, Crusius JB, Roberts RL, et al. Association of FcgR2a, but not FcgR3a, with inflammatory bowel diseases across three Caucasian populations. *Inflamm Bowel Dis.* 2010;16 (12):2080-2089.

- **63**. Franke L, el Bannoudi H, Jansen DT, et al. Association analysis of copy numbers of FC-gamma receptor genes for rheumatoid arthritis and other immune-mediated phenotypes. *Eur J Hum Genet*. 2016;24(2):263-270.
- **64**. Grozdanov V, Bliederhaeuser C, Ruf WP, et al. Inflammatory dysregulation of blood monocytes in Parkinson's disease patients. *Acta Neuropathol*. 2014;128(5):651-663.
- **65**. Cho DH, Tapscott SJ. Myotonic dystrophy: emerging mechanisms for DM1 and DM2. *Biochim Biophys Acta*. 2007;1772(2):195-204.
- **66.** Kaliman P, Llagostera E. Myotonic dystrophy protein kinase (DMPK) and its role in the pathogenesis of myotonic dystrophy 1. *Cell Signal*. 2008;20(11):1935-1941.
- **67**. Trembley JH, Tatsumi S, Sakashita E, et al. Activation of pre-mRNA splicing by human RNPS1 is regulated by CK2 phosphorylation. *Mol Cell Biol*. 2005;25(4):1446-1457.

- **68**. Trabzuni D, Ryten M, Emmett W, et al; International Parkinson Disease Genomics Consortium (IPDGC). Fine-mapping, gene expression and splicing analysis of the disease associated *LRRK2* locus. *PLoS One*. 2013;8(8):e70724.
- **69**. Zhang Q, Pan Y, Yan R, et al. Commensal bacteria direct selective cargo sorting to promote symbiosis. *Nat Immunol*. 2015;16(9):918-926.
- **70**. Rao M, Gershon MD. The bowel and beyond: the enteric nervous system in neurological disorders. *Nat Rev Gastroenterol Hepatol*. 2016;13 (9):517-528.
- 71. Pellegrini C, Colucci R, Antonioli L, et al. Intestinal dysfunction in Parkinson's disease: lessons learned from translational studies and experimental models. *Neurogastroenterol Motil.* 2016;28(12):1781-1791.
- **72**. Ransohoff RM, Perry VH. Microglial physiology: unique stimuli, specialized responses. *Annu Rev Immunol*. 2009;27:119-145.

- **73.** Stone DK, Reynolds AD, Mosley RL, Gendelman HE. Innate and adaptive immunity for the pathobiology of Parkinson's disease. *Antioxid Redox Signal*. 2009;11(9):2151-2166.
- **74**. Daniele SG, Béraud D, Davenport C, Cheng K, Yin H, Maguire-Zeiss KA. Activation of MyD88-dependent TLR1/2 signaling by misfolded a-synuclein, a protein linked to neurodegenerative disorders. *Sci Signal*. 2015;8(376):ra45.
- **75.** Monahan AJ, Warren M, Carvey PM. Neuroinflammation and peripheral immune infiltration in Parkinson's disease: an autoimmune hypothesis. *Cell Transplant*. 2008;17(4):363-372.
- **76.** American Diabetes Association; American Psychiatric Association; American Association of Clinical Endocrinologists; North American Association for the Study of Obesity. Consensus development conference on antipsychotic drugs and obesity and diabetes. *Diabetes Care*. 2004;27 (2):596-601.