

1 Genomic quantitative genetics to study evolution in the wild

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19 **Keywords**

20 adaptation, high-throughput genotyping, quantitative genetics, relatedness, natural populations

21

22 **Abstract**

23 Quantitative genetic theory provides a means of estimating the evolutionary potential of natural
24 populations. However, this approach was previously only feasible in systems where the genetic
25 relatedness between individuals could be inferred from pedigrees or experimental crosses. The
26 genomic revolution opened up the possibility of obtaining the realized proportion of genome shared
27 among individuals in natural populations of virtually any species, which could promise (more)
28 accurate estimates of quantitative genetic parameters in virtually any species. Such a ‘genomic’
29 quantitative genetics approach relies on fewer assumptions, offers a greater methodological
30 flexibility, and is thus expected to greatly enhance our understanding of evolution in natural
31 populations, for example, in the context of adaptation to environmental change, eco-evolutionary
32 dynamics, and biodiversity conservation.

33

34

35 **Understanding trait evolution in natural populations using quantitative genetics**

36 Reliably predicting responses to selection is essential in animal and plant breeding, for studying
37 evolution in natural populations, and understanding population and species response to rapid
38 anthropogenic environmental change [e.g. 1]. Building on the seminal work of RA Fisher [2], who
39 first proposed to decompose the variation in a continuous trait into genetic and environmental
40 components, breeders and evolutionary biologists have long used quantitative genetic (QG) theory
41 to estimate the heritable proportion of trait variation and co-variation with other traits to predict
42 response to natural and artificial selection [3]. Partitioning the variance in a trait has been possible
43 by assuming a simple model of genetic determinism: each trait is affected by many loci of
44 individually (infinitesimally) small effects, known as the infinitesimal model (see Glossary).
45 Despite this strong, and not necessarily realistic assumption underlying QG [4], it has been very
46 efficient and successful in predicting selection responses in animal and plant breeding over the last
47 century [5].

48 The key requirement for any QG analyses is information about the relatedness among individuals.
49 The resemblance between relatives is the basis for separating the heritable proportion of the trait
50 variance (the additive genetic variance) that determines the response to selection and potential for
51 evolution. The generalization of QG theory to include any kind of relatives in a single statistical
52 framework called the ‘animal model’ has become the major tool in animal breeding to predict the
53 ‘genetic merit’ or breeding value of individuals [6]. The animal model also opened up the
54 possibility to study evolution in the wild, because it was no longer necessary to perform crosses to
55 obtain many pairs of individuals of the same relationship, instead, any degree of relatedness was
56 regarded as informative. Nevertheless, the application of QG in wild populations remained a tool of
57 privilege, limited to long-term studies, where pedigrees were possible to establish from individual
58 observations, biasing the approach towards a handful of mammal and bird species [7].

59 With the genomic revolution, QG moved away from the spotlight, and the focus shifted to searching
60 for the genetic determinants of phenotypic variation, i.e. mapping quantitative trait loci (QTL).
61 Thanks to lower genotyping costs, mapping at the finest possible level was enabled by genome wide
62 association studies (GWAS) that aim to find markers, most often single nucleotide polymorphisms
63 (SNPs), associated with phenotypic variation [e.g. 8]. However, GWAS so far did not meet the high
64 expectations: in many cases, a large part of the phenotypic variance remained unexplained by SNPs
65 that reached genome-wide significance (the so-called missing heritability problem); a problem that
66 is now partly resolved by increased sample sizes [8]. Furthermore, even the largest GWAS studies
67 in humans have not yet reached a plateau of the number of 'risk factors' (significant SNPs)
68 associated with trait variation [8]. Not so surprisingly, GWAS conducted in natural plant and animal
69 populations had so far limited power and often failed to identify SNPs associated with traits
70 [reviewed in 9]; yet the lack of any genome-wide significant loci indicates the absence of major
71 effect loci. Some of these studies instead found that the proportion of variance explained by a
72 chromosome correlated with its size [10, 11], as expected if these traits are under the influence of
73 many loci of small effects. All these findings support that the infinitesimal model of classical QG

74 theory is a reasonable approximation for most quantitative traits, including life-history, behavior, or
75 morphology [12]. The QG approach also has the advantage of not relying on unknown underlying
76 molecular or functional details of the genetic basis of quantitative traits and directly provides
77 estimates of the evolutionary potential of such traits [13]. In this Opinion paper, we promote a so-
78 called ‘genomic QG’, which makes use of genomic data almost exclusively for estimating the
79 relatedness between individuals, thus opening the door to many applications of QG to decipher the
80 architecture of quantitative traits and their response to selection; questions often off-limits in many
81 natural systems. Genomic QG has the potential to improve our ability to mitigate species’ responses
82 to environmental change [e.g. 1], and contribute to the field of conservation genetics, such as
83 identifying ‘pre-adapted’ individuals in programs of assisted migration to ‘help’ species keep up
84 with climate change by estimating genetic merit of translocated individuals [e.g. 14].

85

86 **From pedigrees to genome sharing**

87 Relatedness is traditionally based on pedigrees at the base of which individuals are assumed to be
88 unrelated (founders) [15]. Given a pedigree, it can be determined whether two alleles at a particular
89 locus that are identical (by state or IBS) are also identical by descent (IBD) because they were
90 inherited from the same ancestor. Due to Mendelian segregation, there is inherent variation in IBD
91 across loci. For example, full-sibs carry, on average, 50% of their alleles IBD, but the exact
92 percentage of genome sharing and hence their realized relatedness is subject to variation (see
93 Fig.1A). The amount of variation around the expectations depends upon the pedigree-relationship
94 class, and is zero only for parent and their offspring and mono-zygotic twins. Pedigree-based
95 relatedness has well known shortcomings, such as its arbitrariness with respect to a founder
96 population and its imprecision regarding IBD allele sharing [e.g. 16]. Nevertheless, it has been
97 standard in the vast majority of past QG studies.

98 Even though different relatedness estimators that approximate IBD probabilities have been
99 suggested in the microsatellite era, they had too large sampling errors [e.g. 17], thus it was widely

100 accepted that markers should only be used to check, correct, and complete pedigrees rather than
101 replacing them [18]. With the advent of genomic data (Box 1), it has become possible to estimate
102 the realized proportion of the genome that two individuals share IBD. The proportion of genome
103 shared between all pairs of individuals in a population can be summarized with the genomic
104 relationship matrix (GRM) (see Box 1 for methods).

105 The number of markers necessary to accurately estimate the GRM ultimately depends on the
106 effective number of independently segregating chromosome segments that depend on the effective
107 population size, the recombination rate and the genome size [19] and are also influenced by the
108 mating system and population structure and history, which concertedly determine linkage
109 disequilibrium decay. Generally speaking, species with small genomes need fewer markers than
110 species with big genomes, where the difference between expected and realized relatedness can be
111 high [19, 20]. A greater number of markers is also expected to be necessary in outcrossing
112 populations in comparison to selfing, partially selfing or inbred populations because outcrossing
113 populations generally have large effective population sizes and high recombination rates [21, 20]. It
114 is difficult to provide a quantitative guidance for the required number of markers for a specific
115 study. Thus, performing simulations using characteristics of the studied species and populations,
116 and exploring multiple GRM estimation methods (see Box 1) might be the best approach.

117

118 **The ‘dawn’ of genomic quantitative genetics**

119 For humans and species of economic importance, genomic data became available much earlier than
120 for other species, thus the first genomic QG tools appeared in these fields. Many of these methods
121 have not reached evolutionary and global change biology yet. First, we review these tools, then
122 discuss the initial applications in natural populations.

123

124 *Animal and plant breeding*

125 Animal breeders traditionally estimated breeding values based on phenotypic scores, such as milk
126 yield, of a focal individual and its relatives, using an animal model (Box 2). With high density SNP
127 chips (Table 1) becoming available for many domesticated species, breeders were the first to
128 suggest the use of the GRM in an animal model to predict genomic breeding values (GEBVs) [22]
129 and also developed other approaches for genomic selection (GS) that even bypass the animal model
130 (see Box 2 for details). GS revolutionized animal breeding and the field has experienced a surge of
131 method development [23]. It was also demonstrated that an animal model coupled with a GRM
132 instead of a pedigree leads more accurate estimates of the variance components than pedigree-based
133 equivalents [24, 23]. Further, most pre-genomic breeding applications ignored estimating non-
134 additive genetic effects because they generally require complex breeding designs or pedigree
135 structures, and also because it was believed that their effects are negligible [e.g. 5]. A handful of
136 recent empirical studies from tree breeders demonstrated that, in comparison to classical pedigree
137 based approaches, the use of a GRM in an animal model can yield substantially more accurate
138 separation of the additive and non-additive components of genetic variance [25, 26].

139

140 *Human genetics*

141 Human populations cannot be manipulated experimentally, so different 'non-invasive' genomic QG
142 methods have been developed in this field. Most notably, it was suggested to exploit variation in
143 realized genome sharing within a pedigree relationship class to estimate the heritability: Visscher *et al.*
144 *al.* [27] proposed this idea within full-sib families, and Yang *et al.* [28] among unrelated
145 individuals. Most importantly, these methods led to more accurate estimates of the variance
146 components. Nevertheless, population parameters such as linkage disequilibrium between marker
147 and quantitative trait loci can lead to different performance of this method in different species or
148 populations. To explore this further, Visscher and Goddard [29] investigated the sampling error in
149 the heritability estimates as a function of the sampling scheme, the sample size and the effective
150 population size, and concluded that overall, more individuals are needed to achieve the same

151 precision with random population samples in comparison to pedigrees, and higher sample size is
152 required when there is little variance in relatedness such with large effective population size or in
153 expanding populations.

154

155 *The first 'wild' applications*

156 Applications of the genomic QG approach in wild populations have been lagging behind breeding
157 and human genetics for obvious genomic resource reasons, even though the idea to use molecular
158 markers to estimate heritability in natural populations had been proposed back in 1996 by Ritland
159 [31]. Most early applications, based on a few microsatellites, yielded estimates too imprecise and
160 often out of range [30, 31]. Recently, with dense marker panels becoming accessible in some wild
161 pedigreed populations, it has been shown that marker-based relatedness estimates agreed with those
162 from pedigrees (see Fig.1A and B) and also that heritability estimates based on high-density
163 markers and pedigrees agree (Soay sheep (*Ovis aries*) [32], great tits (*Parus major*) [10]). Some
164 empirical studies also investigated how many markers are required to estimate the heritability by
165 comparing estimates from a decreasing proportion of the available loci. For example, in the selfing
166 plant species, *Medicago truncatula*, heritability estimates with 25k SNPs (i.e. one marker every 10
167 Kb, which approximates the distance of linkage disequilibrium decay) were nearly unbiased in
168 comparison to those obtained with >5 million SNPs (Fig.1C). Similarly, for a small island
169 population of a wild ungulate, Soay sheep, heritability with 10k SNPs yielded unbiased estimates of
170 the heritability obtained with 33k SNPs across several traits (Fig.1D).

171

172 **The genomic QG toolbox for ecology and evolution**

173 Bringing QG to the wild by using the pedigree-based animal model has greatly advanced our
174 understanding of evolution [7]. Today, the acquisition of high-density marker data is possible in
175 virtually any species (Table 1), which opens up the possibility to apply the QG research program in

176 the absence of a pedigree or controlled cross in any population of any species. Genomic QG also
177 offers a greater methodological flexibility than classical QG and many evolutionary and global
178 change research questions could benefit from it.

179

180 *Genomic estimated breeding values*

181 GEBVs could have many potential uses outside animal and plant breeding, such as in the context of
182 conservation biology to identify suitable individuals for assisted migration or release programs. The
183 objective of assisted migration is to ‘help’ species migrating to keep up with the pace of climate
184 change, and it is becoming common practice for forest trees [14]. It has been suggested that GEBVs
185 can be combined with gene-environment association analysis to predict phenotypes and identify
186 changes in performance along environmental gradients [33]; Box 3 provides a proof-of-concept
187 example of this approach in *Arabidopsis thaliana*. Since GS is based on a statistical model built on
188 data from a ‘training population’ (Box 2); in order to obtain reliable predictions of the breeding
189 values; it seems necessary to include individuals across the full environmental range over that
190 predictions are desired. A further potential use of GEBVs is in breeding programs of endangered
191 species, where genomic data could help to optimize breeding of remaining individuals to minimize
192 inbreeding or outbreeding depression [34]. Releasing individuals bred in captivity into the wild is
193 often used to restore extinct populations or help threatened populations to survive, but programs
194 might fail if individuals are maladapted to the new environment [e.g. 35]. Identifying potentially
195 well-adapted individuals based on their GEBVs, as described in Box 2, could be an option, or
196 combined with allele specific genetic rescue, e.g. when disease resistance genes are known [see
197 examples in 36]. As pointed out above, it remains unclear whether estimating GEBVs will be
198 generally feasible in natural populations, but in an outbreeding tree species GEBVs estimated with
199 reasonable accuracies [37]. The fact that QG estimates based on genomic markers agreed with
200 ‘standard’ QG estimates in the studies on mammals, birds and plants [10, 38, 32] also indicates that
201 it should be possible.

202

203 *Realized genome sharing to quantify fitness*

204 The GRM is just the first step in genomic QG, but it can also be used in another innovative way in
205 evolutionary biology: The average relatedness of an individual with the population, estimated from
206 a GRM, could be used a measure of inclusive fitness integrated across the current population as
207 individuals whose ancestors produced many offspring should have a higher than average relatedness
208 with the population. Such a fitness measure could also be interesting for behavioral ecology, where
209 estimating relatedness among interacting individuals is necessary to understand social behaviors.
210 Here, the average genomic relatedness of an individual could better estimate its inclusive fitness.
211 Particularly, relatedness estimates based on coalescence could be adapted here [see also 39 and Box
212 1]. Estimating reproductive success is a challenging endeavor in wild populations for several
213 reasons [40] but could potentially be tackled by estimating GRMs.

214

215 *Evolutionary potential in the wild*

216 Evolutionary potential is a scaled measure of the additive genetic variance (V_A); it is the heritability
217 when it is scaled with the total phenotypic variance and evolvability when it is scaled by the trait
218 mean [41]. Bringing genomic QG to the wild will allow assessing the evolutionary potential of a
219 population *in situ*, which is a considerable advancement because it estimates the additive genetic
220 variance and covariance of phenotypic traits under the conditions in which natural selection is
221 acting, which is not the case when using common garden experiments. Such common garden
222 estimates are not guaranteed to apply under natural conditions as V_A might vary across
223 environments and strongly depends on population size and inbreeding levels [reviewed in 42].
224 Estimates of trait heritability in natural populations, however, are also prone to biases due to plastic
225 responses to environmental variables [see 43 for a recent review]. Consequently, a good
226 understanding of environmental but also individual-level variables, such age or body condition, that

227 affect the trait is necessary to account for this ‘environmental’ variation by including the relevant
228 variables in the model. Alternatively, if the exact variables responsible for plastic trait variation are
229 unknown or difficult to estimate but are known to vary spatially or temporally, appropriately
230 modelling this spatial or temporal variation is a suitable approach. For example, in the case of an
231 environmental variable that varies in discrete ‘units’, such as the effect of spring climate on timing
232 of flowering, breeding or migration, fitting the temporal or spatial ‘unit’ as a (fixed or random)
233 factor would be appropriate. If variation is more gradual, fitting an appropriate autocorrelation in
234 the model can remove this plastic variation in traits [e.g. 44]. Another approach to remove
235 confounding effects between genetic and environmental similarities is to estimate QG parameters
236 from variation in relatedness within families, such as full-sibs [27], which could be possible in
237 clutches of birds, fish or insects or half-sibs in plants [e.g. 45]. These *in situ* estimates of the
238 evolutionary potential can be useful in many fields of ecology, evolution and conservation biology.
239 Climate change induced new selective pressures such as increased drought, heat or seasonal shifts,
240 which have been reported in many ecosystems but for most species we lack *in situ* or even any
241 estimates of V_A at ecologically key traits including phenology or drought tolerance [e.g. 46, 47].
242 Climate change is also expected to drive changes in community assemblages due to different
243 dispersal abilities at different species, as has been shown for alpine plants [48]. *In situ* evaluation of
244 the evolutionary potential of key life-history traits is necessary to assess whether these species will
245 be able to adapt to the changed competition regime.

246 Estimates of V_A could also be particularly useful for models of eco-evolutionary dynamics (Hendry
247 2016). For example, Ellner [49] suggested a framework to partition change in a population into
248 contributions of evolution, non-heritable trait change and environmental. To date, this theoretical
249 framework has been limited to experimental systems, but with genomic QG it could be applied in
250 natural settings.

251 With the decreasing costs of genotyping, measuring phenotypes of individuals can become the
252 limiting factor for QG studies in natural populations. Although the variance in (genomic) breeding

253 values is not V_A [see e.g. 50], it could be used as a proxy for it, which would allow prediction of
254 evolutionary potential in new populations without the need to phenotype individuals. However,
255 genotype-by-environment interactions might limit the application of this approach to species where
256 reaction norms are known across environmental gradients.

257

258 *Predicting response to selection*

259 The evolutionary potential informs about the capacity of a population to respond to selection.
260 Knowledge of V_A for a trait or for fitness directly applies to predictive QG models of eco-
261 evolutionary dynamics. This is especially relevant in the context of predicting shifts of species'
262 ranges caused by climate change, and many voices have called for the development of eco-
263 evolutionary forecasting models in that context [e.g. 51]. Quantitative genetics theory provides a
264 well formed framework for predicting eco-evolutionary dynamics in single or among competitively
265 interacting species [e.g. 52]. Current research efforts provide practical examples of implementation
266 of this framework, together with modeling of species ecological niche distributions, to predict
267 changes of species' distribution in *Drosophilid* flies in Australia [53] or endemic alpine plants in
268 Austria [54]. However, this approach is data hungry and needs estimates of V_A for climate-
269 adaptation traits and the selection gradients acting on them. There is hope that by using genomic
270 QG tools such as environment-related GEBVs in natural populations (Box 3) we will be able to
271 alleviate the dearth of estimates of QG parameters necessary to correctly forecast the fate of natural
272 species facing a warming climate.

273

274 **Conclusions**

275 Genomic QG has the potential to advance our understanding of evolutionary processes in natural
276 populations in several ways. Its main advantage is that we no longer need pedigrees or breeding
277 experiments for QG analysis, which means that we can study additive genetic (co)variances, but

278 also selection, in virtually any species in a natural setting. Genomic QG parameters estimates can be
279 more precise than pedigree-based estimates, because they have fewer assumptions and offer a
280 greater methodological flexibility. This ability to conduct quantitative genetic analyses in natural
281 populations could allow more accurate predictions of species' persistence under climate change and
282 enable informed conservation strategies. As we pointed out, the feasibility of this approach will
283 partly depend on the specific study system. The relative ease with which high-density markers can
284 be obtained make us cautiously optimistic about the potential of marker-based quantitative genetics
285 to considerably contribute to our understanding of evolutionary processes in natural populations and
286 mitigating climate change.

287

288 **Acknowledgements**

289 We thank Thomas Mitchell-Olds, Jean-Luc Jannink, Marcel Visser and two anonymous reviewers
290 for constructive comments on the manuscript. The idea for this manuscript developed at Monte
291 Verità Conference in 2016 on “The genomic basis of eco-evolutionary change” organized by the
292 ETH Zurich's center for Adaptation to a changing environment (ACE). We would like to
293 acknowledge the organizers for providing such a fruitful, scientific atmosphere. Camillo Béréños,
294 Josephine Pemberton, John Stanton-Geddes and Peter Tiffin kindly allowed us to re-use their
295 figures and provided the data to do so. SF was partly supported by the Swiss National Science
296 Foundation (project 31003A_160123). FG is supported by grant PP00P3_144846 from the Swiss
297 National Science Foundation. KC was supported by an ACE Fellowship while working on the
298 manuscript.

299

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Table 1 Overview over high-throughput genotyping approaches and sequencing approaches that are potentially suitable to estimate relatedness.

Data type	Genomic resources needed	Pros	Cons	Cost	Examples of studies applying this approach to calculate the GRM
SNP chip	Reference genome	“Ready to use” - No bioinformatics work	Availability mostly limited to model and breeding systems, and possibly close relatives. Ascertainment bias: new populations can have different polymorphisms [55]	Low, if chip available	animal and plant breeding [24, 23], human genetics [28], wild species [32]
RRL- or RAD-sequencing	None, but improved if there is a reference genome	Flexibility of experimental design assisted by software predictions on number of markers. Established bioinformatics for data	Bias on relatedness estimates due to allele dropout and high proportion of missing data [56]	Medium	[56, 57]

		analysis with easy-to-use pipelines			
Whole-genome sequencing	Reference genome	High-confidence genome-wide information and use of established bioinformatics tools	High costs for large genomes, and can yield excessive amount of information [38]	Very high	[58, 38]
Low-coverage whole-genome sequencing	Reference genome	Dense genome representation including causal variants that can be suitable for parallel association studies.	Requires bioinformatics knowledge to use software building on probabilistic framework.	High	[59, 60]
Sequence or Exome capture	Reference genome of closely related species or	Targeted sequencing of selected loci, which can include causal variants	Requires high-quality genomic resources for bait design and to minimize allele bias [61]	High	[62]

	transcriptome	suitable for parallel association studies.		
Genome skimming	Reference genome	Minimal sequencing effort to survey high-copy regions of the genome, typically organellar and ribosomal DNA	No studies yet built a GRM. Complex evolution of high-copy nuclear loci can limit applicability in natural populations.	Low

451 **BOX 1 Methods to estimate the genomic relationship matrix (GRM)**

452 *1. Approximate identity-by-descent (IBD) probabilities at single locus.* This approach includes a
453 whole class of maximum-likelihood [e.g. 63, 60] and method of moments estimators [see 17 for an
454 overview].

455 *2. Cumulative excess of recent coalescence.* A coalescent tree is used to trace the origin of alleles in
456 the current population back to a most recent common ancestor (MRCA). Rousset [39], suggested to
457 define kinship coefficients as the degree of excess coalescent time to their MRCA at each
458 independently segregating segments of the genome, however, no estimator or software have been
459 proposed yet.

460 *3. Average allelic correlation (a.k.a. unified additive relationship).* This is the most standard class
461 of approaches to estimate the GRM in animal and plant breeding and human genetics (see Table I
462 for software). In breeding, the method appeared with the uprise of genomic selection (GS), and
463 calculated as the correlation between genotypes of two individuals across all loci [64]. In human
464 genetics, the approach appeared in Yang et al. [28], and was explained in Powell et al. [65]. The
465 approach circumvents the problem of the arbitrarily reference population by defining it as the
466 current population. As a result, IBD is no longer a probability, but a correlation coefficient that can
467 be used to predict if a gamete carries the same allele as another irrespective of its origin (i.e. IBS).
468 Adjustment for linkage disequilibrium (LD) been proposed by [16], which can lead to improved
469 estimates, especially with sparse SNP data [66]. When using dense SNP data, LD-adjustment
470 attributes too much weight to the SNPs with low minor allele frequencies.

471 *4. Haplotype sharing.* Haplotype sharing approaches require both marker data and a genetic map
472 [e.g. 67]. A common approach in GS is to use dense SNP chip on a few key individuals to identify
473 haplotypes, and a sparse chip on the rest of the individuals [see review by 23].

474 Table I. Examples of software to estimate the GRM

Software	PLINK (v1.07)	GCTA	LDAK	BEAGLE
Meth	1	3	3	4
Reference (software)	[68]	[69]	[16]	[67]
Websites	http://zzz.bwh.harvard.edu/plink/	http://cnsgenomics.com/software/gcta/	http://dougspeed.com/ldak/	http://faculty.washington.edu/browning/beagle/beagle.html

475

476

477 **BOX 2 Two key statistical models to estimate QG parameters using genomic data**

478 The two main types of statistical models dominate the QG literature and both can make use of
 479 genomic data. In the animal model, the pedigree can be replaced with the GRM, and in genome
 480 regressions, genomic data is a necessary ingredient.

481

	Animal model	Genome regression
Key reference	Henderson (1975)	Meuwissen, Hayes, Goddard (2001)
The model	$y_i = \mu + a_i + e_i$ where a_i is the breeding value of individual i. Since a_i 's are unknown, they are estimated from the covariance between relatives in additive genetic effects given by $2 \cdot \text{kinship} \cdot V_A$, thus using a pedigree or a GRM	$y_i = \mu + \sum (SNP_{i,j} * g_j) + e_i$ where $SNP_{i,j}$ is the genotype of individual i at loci j, and g_j is the effect of the SNP on the phenotype
explanation of common model terms	y_i is a trait value of individual i, μ is the population mean, and e_i is the residual term	
Breeding value (BV) estimation	Using the best linear unbiased predictor (BLUP) in a mixed effects model	BLUPs assuming equal variances across SNPs is possible, but Bayesian methods generally work better

482

483 **About the animal model**

484 Animal model is most often used to estimate variance components, such as the additive genetic
485 variance, which serves to estimate the heritability of traits or genetic correlations between traits.
486 Breeding values are the estimated random effects and can be used to characterize individuals,
487 however, several authors warn not to use them in further statistical analysis without incorporating
488 their estimated standard errors [e.g. 50], e.g. in a hierarchical Bayesian framework. Generally large
489 pedigrees are required but it can also be used with a common garden experimental data.

490

491 **About genome regressions**

492 Genome regressions are principally used in a prediction context to help breeders identify superior
493 individuals. Thus, the model is fitted in a training population to estimate breeding values of
494 individuals of unknown trait value in a candidate population (such as phenotyping milk yield at an
495 early age or at a male cow). GEBVs are generally much more accurate than EBVs. Particularly high
496 accuracy is expected with a large number of individuals in the training population, for a highly
497 heritable trait, and when the number of segregating chromosome segments is small (i.e. small
498 genome and small effective population size). GS does not work well across breeds. Instead, the best
499 performance is achieved when individuals in the candidate population are related to individuals in
500 the training population.

501

502 **Box 3 Using genomic relatedness and home environments to identify locally adapted**
503 **genotypes**

504 A common challenge in conservation genetics is to identify locally-adapted genotypes for a given
505 site. Here, we demonstrate that GRMs for wild accessions collected across diverse climates
506 ('ecotypes') can be applied to predict which genotypes are adapted to cold sites and which are
507 adapted to warm sites. Our approach relies on a three-way correlation between genomic similarity
508 (GRMs), similarity in home environments (winter cold), and similarity in locally adaptive traits
509 (unobserved and predicted). As such, correlation between population structure and environmental
510 gradients causing local adaptation is not a problem for our approach, rather this correlation is
511 leveraged to predict adaptive trait variation. By contrast, population structure-environment
512 correlation is problematic for genome-environment association studies that seek to identify
513 individual causal loci involved in local adaptation. We reanalyzed published data on *Arabidopsis*
514 *thaliana* [70-72] and global climate [73]. We used >200k SNPs [72] to estimate a GRM based on
515 average allelic correlation (Box 1). We fitted an animal model where an ecotype's native winter
516 cold environment (minimum winter temperature) was used as 'phenotype' and modeled as a
517 function of the GRM with 10-fold cross validation to predict native temperatures as GEBVs [74].
518 Based on the assumption that populations are locally adapted along temperature gradients, these
519 GEBVs can also be interpreted as predictions of a 'latent cold-adapted phenotype', *i.e.* an
520 unobserved phenotype assumed to confer high fitness in particular temperatures. We next show that
521 these predictions closely correspond to which genotypes are locally adapted along temperature
522 gradients. Although here we validated our predictions with experimental data, note that no
523 experiments were used to generate our predictions; predictions are based solely on the relationship
524 between GRM and native environments. Individual relative fitness of 157 ecotypes was measured in
525 four common gardens by Fournier-Level et al. [71]. In each garden, we regressed relative fitness
526 against predicted latent cold-adapted phenotypes to identify the expected most fit latent phenotype
527 at each site (Fig. I, data from two sites shown in insets, curve shows fitted quadratic regression).

528 The fittest latent cold-adapted phenotype at each site varied closely with differences in winter cold
529 among sites (central panel, $R^2 = 0.92$). This relationship indicates that the GRM association with
530 native environments can be used to predict which genotypes are adapted to cold sites and which are
531 adapted to warm sites, information that might guide conservation and restoration actions.

532

533 **Glossary**

534

535 **Infinitesimal model:** The infinitesimal model assumes that the genetic component of a trait is
536 determined by an infinite number of unlinked genes that each has an infinitesimally small, additive
537 effect. This will lead to continuous genetic variation in the trait and this model is hence generally
538 used to model so-called ‘quantitative’ or ‘complex’ traits. It is, however, also applicable to discrete
539 traits, as e.g. litter size, and even binary traits if trait expression depends on one or more thresholds
540 that the summed allelic effects need to pass.

541 **Heritability:** The narrow-sense heritability (h^2) is the ratio of the additive genetic variance (V_A) to
542 the total phenotypic variance (V_P) of a trait: $h^2 = \frac{V_A}{V_P}$. The narrow-sense heritability is used to
543 predict the response to selection of a trait when using the breeders’ equation. When only the whole
544 genetic variance (V_G) is known, which also includes non-additive genetic effects (dominance and
545 epistasis), broad-sense heritability (H^2) is estimated.

546 **Evolvability** can be defined in the broader and narrower sense. In the broader sense it simply is the
547 ability of a trait to evolve (including from novel mutations). In the narrower sense it is the additive
548 genetic variance scaled by the trait mean: $volvability = \frac{V_A}{\text{trait mean}^2}$, or the additive genetic
549 coefficient of variation (CV_A): $CV_A = \frac{V_A}{\text{trait mean}}$.

550 **Quantitative trait locus:** A quantitative trait locus (QTL) is the genomic section, i.e. a locus, that
551 contribute to the variation (together with other QTL) of a quantitative trait, i.e. a trait that shows
552 continuous phenotypic variation.

553 **Genome-wide association study:** A genome-wide association study (GWAS) aims to identify loci
554 determining a trait by testing for associations between molecular markers and trait values. In the
555 common case when SNP chips are used, the underlying assumption is that the markers are in tight
556 linkage with or within the loci responsible for trait variation.

557 **Single nucleotide polymorphism:** A single nucleotide polymorphism (SNP) is a variation at a
558 specific genomic position. For example, at a specific position in a genome, the base C appears in
559 most individuals, but in a minority the base A appears. This specific SNP would have the two
560 nucleotide variations – C or A – as its alleles.

561 **Identity-by-descent:** A genomic section is identical by descent (IBD) in two individuals if they
562 have inherited it from a common ancestor.

563 **Identity-by-state:** A genomic section is identical by state (IBS) in two individuals if it has the same
564 nucleotide sequence in these two individuals. IBS can arise by chance (mutation) or because the
565 individuals inherited it from the same ancestor, in which case it would also be identical by descent.

566 **Realized relatedness:** The relatedness of two individuals calculated from a pedigree (kinship) is
567 their expected genetic similarity (or proportion of genome sharing). There is, however, variation
568 around this expectation (except for parent-offspring pairs and monozygotic twins) because of the
569 segregation of independent genome segments during meiosis (mendelian noise). This variation is
570 lowered by more independently segregating segments[e.g. 27, 19]. An interesting consequence of
571 this is that for species with ‘small’ genomes, i.e. with fewer independently segregating segments,
572 QG estimates based on realized relatedness will be more precise than estimates based on expected
573 relatedness.

574 **Genomic relationship matrix:** The pedigree-based relatedness matrix, often denoted A , contains
575 all pairwise relatedness estimates (which are two time the kinship coefficient θ) in the off-diagonals
576 and the individuals inbreeding coefficient in the diagonal. The genomic relatedness matrix (GRM),
577 often also denoted G , is the same but with the relatedness estimates derived from genomic markers.
578 Various approaches to estimate relatedness from markers exists (see Box 1) and GRMs can be
579 constructed with any of them.

580 **Linkage disequilibrium** is the non-random association of alleles at different loci. Loci are in
581 linkage disequilibrium, or linked, if certain combinations of alleles occur more often than expected

582 by chance. Linkage disequilibrium can be caused by a multitude of factors, including selection,
583 recombination rate, mating system, population structure or physical proximity in the genome.

584 **Non-additive effects** include dominance and epistasis. Dominance means that the effect of one
585 allele masks (is 'dominant' over) the effect of the other (recessive) allele. Incomplete and co-
586 dominance are cases when the dominant allele is not completely masking the recessive allele and
587 the heterozygous phenotype is somewhat intermediate between the homozygous dominant and
588 homozygous recessive phenotype. Epistasis, or gene-gene interactions, occurs when the allelic
589 effect at one locus depend on the alleles of one or more other loci, the genetic background.

590 **Assisted migration:** Many species have limited dispersal abilities or their dispersal is impaired by
591 habitat fragmentation, which means that they cannot reach other, isolated populations or currently
592 unoccupied but suitable habitat. In assisted migration, or assisted gene flow, individuals are
593 artificially moved from one population to improve another population's probability to persist or to
594 colonize novel but suitable habitat, e.g., in the context of climate change.

595

596 **Figure caption**

597 Fig.1 Genomic relatedness and heritability estimates. (A) Relationship between marker-based and
598 pedigree relatedness in one of the wild study populations with the deepest pedigree, the St. Kilda
599 soay sheep population. Realized or ‘genomic’ relatedness, estimated from ca. 37000 SNP markers,
600 is plotted against IBD relatedness, estimated from the pedigree. The variation of realized
601 relatedness estimates with the same IBD relatedness class, seen as vertical spread, is due to
602 Mendelian segregation (and sampling error). (B) shows, for the same population, that the
603 correlation between pedigree and marker-based relatedness estimates increases with the number of
604 SNPs used to estimate the GRM. (C) and (D) show the effect of number of markers on heritability
605 estimates in *Medicago truncatula* and soay sheep, respectively. Boxes and whiskers indicate the
606 variation in estimates from the repeated samples of the reduced data sets. When few markers are
607 used to estimate the GRM heritability, estimates are down-ward biased. Increasing the number of
608 markers reduces this bias, but, at some point, adding more markers has little effect on the estimate
609 anymore, only on its variation. In (B) and (D) the x-axis shows the proportion of SNPs used in
610 relation to the complete data set and the absolute numbers are 926, 1852, 3704, 11111, 18518,
611 25926, respectively. (A), (B) and (D) re-drawn after [32]; (C) re-drawn after [38].