Genotype—Phenotype Analysis of the Branchio-Oculo-Facial Syndrome

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Branchio-oculo-facial syndrome (BOFS; OMIM#113620) is a rare autosomal dominant craniofacial disorder with variable expression. Major features include cutaneous and ocular abnormalities, characteristic facies, renal, ectodermal, and temporal bone anomalies. Having determined that mutations involving TFAP2A result in BOFS, we studied a total of 30 families (41 affected individuals); 26/30 (87%) fulfilled our cardinal diagnostic criteria. The original family with the 3.2 Mb deletion including the TFAP2A gene remains the only BOFS family without the typical CL/P and the only family with a deletion. We have identified a hotspot region in the highly conserved exons 4 and 5 of TFAP2A that harbors missense mutations in 27/ 30 (90%) families. Several of these mutations are recurrent. Mosaicism was detected in one family. To date, genetic heterogeneity has not been observed. Although the cardinal criteria for BOFS have been based on the presence of each of the core defects, an affected family member or thymic remnant, we documented TFAP2A mutations in three (10%) probands in our series without a classic cervical cutaneous defect or ectopic thymus. Temporal bone anomalies were identified in 3/5 patients investigated. The occurrence of CL/P, premature graying, coloboma, heterochromia irides, and ectopic thymus, are evidence for BOFS as a neurocristopathy. Intrafamilial clinical variability can be marked. Although there does not appear to be mutation-specific genotype-phenotype correlations at this time, more patients need to be studied. Clinical testing for TFAP2A mutations is now available and will assist geneticists in confirming the typical cases or excluding the diagnosis in atypical cases. © 2010 Wiley-Liss, Inc.

Key words: branchio-oculo-facial syndrome; cleft lip/palate; mutation analysis; neurocristopathy; *TFAP2A*

INTRODUCTION

Branchio-oculo-facial syndrome (BOFS; OMIM#113620) is a rare, distinctive, autosomal dominant developmental disorder with variable manifestations [Lin et al., 1995]. The name of the condition reflects involvement of the three major systems. The classic features are thinned erythematous cutaneous defects in the cervical or infra- and/or supra-auricular region, ocular anomalies (microphthalmia or anophthalmia, coloboma, strabismus, cataract, ptosis), and nasolacrimal duct obstruction. The characteristic craniofacial features include dolichocephaly, malformed pinnae, thick nasal tip, upslanted eyes, and cleft lip (CL) (including lesser forms, such as microform, "pseudocleft," or abnormal philtrum [Lin et al., 2009]) with or without cleft palate (CP). Additional findings include conductive/mixed/sensorineural hearing loss, ectodermal anomalies (small teeth, dysplastic nails, sparse and prematurely gray hair), ectopic dermal thymus, and scalp cysts. Growth restriction, renal anomalies (dysplastic, multicystic or absent kidneys, vesicoureteral reflux), upper-lip pits and mild mental retardation are reportedly infrequent findings. More recently, inner ear malformations (incomplete partition type II, enlarged vestibule, and enlarged vestibular aqueduct) and temporal

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bone anomalies (stenosis of the round and oval windows, malformations of the stapes, and hypoplastic long process of the incus) [Stoetzel et al., 2009; Tekin et al., 2009] are of diagnostic and clinical importance.

We originally described five families with BOFS that had heterozygous mutations or a deletion of the *TFAP2A* gene [Milunsky et al., 2008]. Four research groups have confirmed our original findings [Gestri et al., 2009; Stoetzel et al., 2009; Tekin et al., 2009; Reiber et al., 2010a]. This article extends ongoing clinical and molecular research to include 30 BOFS families (41 affected individuals), the largest series involving this syndrome.

MATERIALS AND METHODS Patients

Families were enrolled under a protocol at the Boston University School of Medicine, or submitted as individual patients for clinical genetic testing. Permission for publication of those photos that appear was signed by each participant and/or parent. Pictures of selected patients appear in Figure 1. The diagnostic criteria used for BOFS are summarized in Table I. We also reviewed previously described patients [Gestri et al., 2009; Stoetzel et al., 2009; Tekin et al., 2009; Reiber et al., 2010a].

Methods

In the original family we identified a 3.2 Mb deletion at chromosome 6p24.3, utilizing a 500K Microarray (Affymetrix, Santa Clara, CA). We then employed Multiplex ligation-dependant probe amplification (MLPA) to confirm that the deletion included the *TFAP2A* gene. Details of both of these methods are previously published [Milunsky et al., 2008, Patients 1a and b]. For *TFAP2A* sequencing analysis, genomic DNA was processed with the Autopure automated DNA extractor according to manufacturer's instructions (Gentra Systems, Minneapolis, MN). The seven coding exons and intron/exon boundaries of the *TFAP2A* gene were amplified with PCR using appropriate primers. Primer sequences for the generation of amplicons were derived from the NCBI gene weebsite. The method used to sequence the gene utilizes the ABI

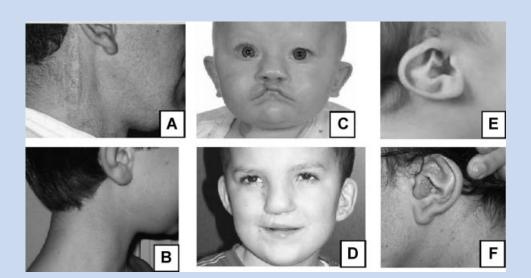


FIG. 1. A: Illustrates the typical "B," the cutaneous defect scar in an adult. In striking contrast, (B) (Patient 27) lacked a typical cutaneous defect, but had a very fine hair extension in the characteristic distribution. Though difficult to illustrate in a photograph, this feature was documented over the years by his pediatrician, mother and barber. C (Patient 12): Typical mildly up-slanted palpebral fissures, full nasal tip, low set protruding ears and lesser form cleft lip prior to surgical repair. The tears pooling in the right eye illustrate nasolacrimal duct atresia. D (Patient 15c): Mildly low-set pinnae, hypertelorism, high nasal bridge, thick nasal tip, following repair of cleft lip and palate. E,F: Variation of the characteristic pinnae anomalies.

VariantSEQr Resequencing System (Applied Biosystems, Foster City, CA). Mutation analysis was performed with the Mutation Surveyor program (SoftGenetics, State College, PA). Several of the mutations were further confirmed using restriction digestion with the appropriate enzyme (New England Biolabs, Beverly, MA), specific to each mutation. In addition, more than 300 normal control individuals were sequenced (see Supplementary Online Version for detailed methodology).

RESULTS

The current BOFS diagnostic criteria (Table I) were derived from Lin et al. [1995] and Milunsky et al. [2008], with the addition of an independently verified first-degree relative or distinctive defect (ectopic thymus). The molecular and clinical features of 41 patients,

TABLE I. Diagnostic Criteria for the Branchio-Oculo-Facial Syndrome (BOFS)*

- (1) All three of the main features present: Branchial (cutaneous) defect Ocular anomaly
- Facial (characteristic facial anomalies)
- (2) Two of the three main features plus one of the following Affected first-degree relative, independently diagnosed Ectopic thymus (dermal)

*Adapted from Lin [2009].

20 males and 21 females (6 months–69 years, mean 18 years) are summarized in Table II. Twenty families were enrolled using our Institutional Review Board research informed consent process. An additional 10 families/individuals are reported after consent was obtained following clinical genetic testing. We also reviewed 16 cases of BOFS reported in the literature (Table II).

Molecular Analysis

The molecular data presented in Table II include the exon where the mutation was found, the protein consequence of the mutation, and whether the mutation was de novo. The original family with the 3.2 Mb deletion including the TFAP2A gene is the only BOFS family without the typical CL/P and our only family with a gene deletion. One proband (Family 29) had a single base pair insertion (c. 376_377 Ins G [p.Asp126GlyfsX43]) in exon 2 that resulted in a stop codon in exon 3, 43 amino acids downstream from the insertion. This was the only frameshift mutation in our series. All other mutations were missense; the majority clustered in exon 4 (23/28, 82%). There were six recurrent mutations (five in exon 4 and one in exon 5; Table III). All missense mutations involved highly conserved amino acids (Fig. 2). Family 22 was the only one with a missense mutation outside of this hotspot region (H384Y) in exon 7. Family 7 was interesting in that there was classic BOFS phenotype in the 11-year-old daughter (R254G mutation in exon 4) and a milder phenotype in her 45-year-old father, who had premature gray hair, preauricular pits, and a supernumerary nipple. On sequencing and restriction digestion (Fig. 3), the father appeared to be a mosaic for the mutation in blood. Further studies from other tissues have been requested.

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Protein consequence	Whole	Whole	Whole	R2556	L249P R3	R254W	G262E	R254W	Mos 22546	R2546	A2 56V	A2 56V	3254W	R236P	G251E	R254W	A256V
	del	del	del														
De novo	No	No	Π	Yes	Yes	Yes	Yes	Π	Π	No		No	Π		Yes	Yes	Yes
Sex	ш	Σ	ш	ш	Σ	Σ	ш	Σ	Σ	ш	Σ	ш	Σ	ш	Σ	ш	ш
Age (years)	25	1	46	18	2	14	17	11	45	11	52	13	6	~	22	2	പ
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anomaly																	
Ectopic thymus	Π		Π	+	I	I	I	Π				Π	Π		+	+	⊐
Ocular anomalies																	
Coloboma	Ι	٣	Ι	٣	I	٣	в	I	I	I	I	в	в	в	I	I	ш
An/microphthalmia	I	٣	I	Ж	I	٣	в	I	I	в	I	I	I	в	8	I	I
NLDS/A		I		в	в	в		I	I	в	I	I	в	в	+	+	в
Cataract	I	I	I	I	I	I	I	I	I	I	I	I			I	I	⊃
Facial anomalies																	
Auricular anomaly	в	в	Ι	в	в	в	в	Ι	I	в	Ι	Ι	в	в	В	в	в
Characteristic face	I	+	I	+	+	+	+	+	I	+	Ι	+	+	+	+	+	+
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Snarse hair	I	I	I	I	I	I	I	I	I	I	I	I	=	=	eyebrow -	I	+
Additional features))			B PAP
Temporal bone	Π	Π		Π	Π			Π	Π	+		Π		Π	Π		Π
anomalies																	
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Supernumerary nipples	Ι	Ι		Ι	Ι	I	·	I	+	+	Ι	I		. –	D	Ι	I
CHD	Π		Π	I	I		I	Π	- 1		I	Π	Π				
Psychiatric symptoms	I	I	I	+	I	I	+	I	I	I	I	I		+	I	I	I
Cancer	I	I	I	I	I	I	+	I	Ι	I	I	I			I	I	I

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14 4 14 7es F 69	B D L	heterochromia irides B B H - I B H - I B	B CL	+	+ Scalp B Mixed	+ ⊃
Patient Exon Protein consequence De novo Sex Age (years) FH	Cervical anomanes Cervical cutaneous anomaly Ectopic thymus Ocular anomalies	Loloboma An/microphthalmia NLDS/A Cataract Facial anomalies Auricular anomaly Characteristic face	Chin dimple/cleft CL/P	Lip pits, upper Facial nerve weak/palsy Preauricular pits Ectodermal anomalies Dental anomalies Nail anomalies Premature gray hair	Cysts Sparse hair Additional features Temporal bone anomalies Hearing loss	Growth restriction Kidney anomalies Supernumerary nipples CHD Psychiatric symptoms Cancer

TABLE II. (Continued)

Literature ⁵⁻⁸ # pts (%), 16 (100) See text See text	8/16 (50) F 8/16 (50) Range 1–57 years, mean 17.2	8/16 (50) 10/16 (63)	0/16 [0] 5/6 [83] 10/14 [71],	1 heterochromia iris 11/15 (73) 9/14 (64) 1/7 (14) 16/16 (100) 15/16 (94) 5/6 (83) 0/16 (0) Any 5/16 (13), CL 3/16 (19), CL/P 2/16 (13), pseudo 1/16 (6)	0/3 (0) 1/11 (9) 3/11 (27) 6/13 (46) 0/5 (0) 6/9 (67) 1/16 (6) 1/16 (6)
New # pts [%], Lit 41 [100]	12/23 (52) F 21/41 (51) Range 0.5–69 years, mean 18 years y	18/41 [44] 33/41 [81]	6/19 (32) 32/41 (78) 20/38 (53)		not stated 1/41 (∠) 4/37 (11) 7/40 (18) 14/40 (35) 14/40 (35) 8/39 (20) 14/37 (38) 8/37 (21) 5/38 (13)
New # probands [%], 30 (100)	11/18 (61) M 17/30 (57) Range 0.5-69 years,	mean 11.b years 10/30 (33) 27/30 (90)	6/14 [43] 27/30 [90] 17/28 [61]	12/30 (40) 24/30 (80) 2/25 (8) 30/30 (100) 23/30 (77) 29/30 (97) 5/28 (18) Any 29/30 (97), CL 8/30 (27), CL 8/30 (27), CL/P 6/30 (20), pseudo 15/30 (50);	1 abn phirtum 4/27 (15) 7/29 (24) 9/29 (31) 13/26 (46) 7/26 (27) 7/26 (27) 5/27 (18)
30 4 R254P	$\supset \Sigma \stackrel{1}{_{0}}$	m	+ m	C m + + m + + +	$ \supset ++++ $
29 2 p.Asp1266ly <i>f</i> .cv13	Yes M 13	🗠		CL Bseudo Pseudo Pseudo	+ ⊃
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27 4 S247L	⊃ ∑ ത	- + Faint hair	only U B	ссedo suedo	+ +
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23 4 E242K	л т 2 29	<u>م</u> ا	⊃		+
Patient Exon Protein consequence	De novo Sex Age (years)	FH Branchial anomalies Cervical cutaneous anomalu	Ectopic thymus Ocular anomalies Coloboma	An/microphthalmia NLDS/A Cataract Facial anomalies Auricular anomaly Characteristic face Chin dimple/cleft CL/P	Lip pits, upper Facial nerve weak/palsy Preauricular pits Ectodermal anomalies Dental anomalies Nail anomalies Nail anomalies Premature gray hair Cysts Sparse hair Additional features

(Continued)

50) 3/7 (42)		1	1	[11] 5/35 [14]		[20] 5/34 [14]	(4) 1/39 (5)	Abn, abnormal; ASD, atrial septal defect, B, bilateral; B PAP, bilateral post-axial polydactyly; CL/P, cleft lip/palate; Cond, conductive; CHD, congenital heart defect (determined by echocardiography); ded, deletion; FH, family history; L, left, mos, mosaic; LDs, learning disabilities; NLDSA, nasolacrimal duct stenosis/atresia; pts, patients; Pseudo CL, pseudocleft (includes mini-microform); R, right; Sev, severe; SN, sensorineural; U, unknown. ¹ Milunsky et al. [2008]; ² Mégarbané et al. [1998]; ³ Lin et al. [2009]; ⁵ Gestri et al. [2009]; ⁶ Stoetzel et al. [2009]; ⁷ Tekin et al. [2009]; ⁸ Reiber et al. [2009]; ⁹ Reiber et al. [2010]; ⁹ tekin et al. [2009]; ⁹ tekin at uthors (J.M.; A.L.) determined the status of certain features from published photos if not specifically stated in text.
3/6 [50]	23/29 [79]	12/30 [40]	10/26 [39]	3/27 (1/11	5/25 (1/28	. defect (determined udoclett (includes mi udoclet); ⁹ Reiber et
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 +]	B Mixed	Ι	⊃	+		Ι	I	bn, abnormal; ASD, atrial septal defect, B, bilateral; I el, deletion; EH, family history; L, left; mos, mosaic; , right; Sev, severe; SN, sensorineural; U, unknown. Milunsky et al. [2008]; ² Mégarbané et al. [1998]; he senior authors [J,M.; A.L.] determined the status

TABLE III. Recurrent *TFAP2A* Mutations Reveal Hotspot of Mutable Amino Acid Residues in BOFS

	Mutation of amino acid residue
Exon	(# individuals or families)
4	R254G/W/P (6)
4	R237G/P (3)
4	E242K (3)
5	A256V (3)
4	G251E (2)
4	R255G (2)

Clinical Features

The frequency and pattern of defects in our patients and in the literature is similar to that reported in both clinical [Lin et al., 1995] and molecular series [Milunsky et al., 2008], but includes several new findings. Importantly, the typical cervical or supra-auricular ("branchial") cutaneous defects were not ubiquitous (90% and 62% in our series and the literature, respectively). The most minimal expression was a patch of faint hair located in the same linear cervical distribution [Patient 27; Stoetzel et al., 2009, Patient 1.2]. Two patients had heterochromic irides [Patient 14; Stoetzel et al., 2009, Patient 1.2]. Congenital heart defects were rare (~8% among total patients), including two atrial septal defects [Patient 24; Gestri et al., 2009 Patient 2], and one tetralogy of Fallot [Reiber et al., 2010a, Patient SP2, previously reported by Bennaceur et al., 1998]. Two probands in our series had bilateral post-axial polydactyly that was not present in their families. This finding may be related to BOFS (rarely), as preaxial polydactyly, type A has been reported [Fujimoto et al., 1987, Patient 1].

Psychomotor development is usually normal, despite frequent visual and hearing handicaps, but, there are two patients reported with autism spectrum disorder and a patient with severe mental retardation [Reiber et al., 2010a, Patient SP2; Gestri et al., 2009; Reiber et al., 2010b].

DISCUSSION

The research families presented had a clinical diagnosis of BOFS based on probands demonstrating all three BOFS features (cervical cutaneous defects, ocular anomalies, facial anomalies) or two features and a first-degree affected relative. Aims of the study included determining genetic heterogeneity, degree of clinical variability, and whether there are any mutation-specific genotype–phenotype correlations. We do not have complete clinical data on all affected individuals, as most have not had temporal bone CT scans or echocardiograms, and some have not had audiology evaluations, or renal ultrasonography. The details of each patient's clinical findings are summarized in Table II. One of our original patients (Patient 5) remains the only affected individual with cancer (medulloblastoma) [see further discussion in Milunsky et al., 2008].

Both Tekin et al. [2009] and Stoetzel et al. [2009] highlighted the inner ear and temporal bone anomalies that had been reported previously in BOFS [Raveh et al., 2000] as well as in other disorders, such as CHARGE syndrome [Amiel et al., 2001] and branchio-

Homo sapiens	RLsllsstskykvtvaevq RR lspp E cln AS l L g G v $LRRA$ kskng G Rslre	267
Equus caballus	RLsllsstskykvtvaevorrlsppEclnASlLgGvLRRAkskngGrslre	269
Sus scrofa	RLsllsstskykvtvaevorrlsppEclnASlLgGvLRRAkskngGrslre	267
Pan troglodytes	$\mathbf{RL} \texttt{sllsstskykvtvaev} \\ \mathbf{RR} \\ \texttt{lspp} \\ \mathbf{E} \\ \texttt{cln} \\ \mathbf{AS} \\ \texttt{l} \\ \mathbf{L} \\ \texttt{G} \\ \mathbf{V} \\ \mathbf{LRRA} \\ \texttt{kskng} \\ \mathbf{G} \\ \texttt{rslre} \\ rs$	269
Rattus norvegicus	RLsllsstskykvtvaevQRRlsppEclnASlLgGvLRRAkskngGrslre	261
Mouse	RLsllsstskykvtvaevq RR lspp E cln AS l L g G v $LRRA$ kskng G Rslre	267
Monodelphis domestica	RLsllsstskykvtvaevq RR lspp E cln AS l L g G v $LRRA$ kskng G rslre	267
Ornithorhynchus	RLsllsstskykvtvaevq RR lspp E cln AS l L g G v $LRRA$ kskng G rslre	267
Canis familiaris	\mathbf{RL} sllsstskykvtvaevq \mathbf{RR} lspp \mathbf{E} cln \mathbf{AS} l \mathbf{L} g \mathbf{G} v \mathbf{LRRA} kskng \mathbf{G} rslre	264
Bos taurus	\mathbf{RL} sllsstskykvtvaevq \mathbf{RR} lspp \mathbf{E} cln \mathbf{AS} l \mathbf{L} g \mathbf{G} v \mathbf{LRRA} kskng \mathbf{G} rslre	267
Ovis aries	\mathbf{RL} sllsstskykvtvaevq \mathbf{RR} lspp \mathbf{E} cln \mathbf{AS} l \mathbf{L} g \mathbf{G} v \mathbf{LRRA} kskng \mathbf{G} rslre	321
Gallus gallus	RLsllsstskykvtvaevq RR lspp E cln AS l L g G v $LRRA$ kskng G rslre	267
Xenopus laevis	\mathbf{RL} sllsstskykvtvaevq \mathbf{RR} lspp \mathbf{E} cln \mathbf{AS} l \mathbf{L} g \mathbf{G} v \mathbf{LRRA} kskng \mathbf{G} rslre	241
Danio rerio	RLsllsstskykvtvaevq RR lspp E cln AS l L g G v $LRRA$ kskng G rslre	261
Ciona intestinalis	RLsllsstskykvtvaeiq RR lspp E cln AS l L g G v $LRRA$ kskdg G krlrd	400

Amino Acids		



oto-renal syndrome [Ceruti et al., 2002]. In our cohort, 3/6 probands investigated had temporal bone anomalies, whereas CHARGE syndrome is associated with semicircular canal aplasia/hypoplasia, which is nearly diagnostic [Glueckert et al., 2010; Zentner et al., 2010]. In contrast, BOR is associated with mostly cochlear anomalies [Ceruti et al., 2002]. This finding is likely to be under-diagnosed; further large clinical studies are needed to determine the frequency of temporal bone anomalies in BOFS.

The probands in every family had some form of CL, with or without CP. The one exception, without an overt cleft, was our deletion Family 1, who had a short-tented prominent philtrum. Gestri et al. [2009] reported a BOFS family with a paternally inherited whole gene deletion. The brother and sister with "unaffected" parents had an unusual phenotype with orbital cysts who were diagnosed as having BOFS [Fielding and Fryer, 1992], but viewed as "possibly affected" and "atypical" in a large series [Lin et al., 1995]. Recent molecular analysis confirmed the diagnosis and examination of their father showed premature graying and abnormal philtrum [Gestri et al., 2009]. None of the family members had an overt labial cleft, but rather were characterized by having a distinctly abnormal philtrum or a "pseudocleft." Further data come from Davies et al. [1999] and Misceo et al. [2008], who both reported chromosome 6 interstitial deletions, including *TFAP2A* that do not result in overt clefting, but rather a

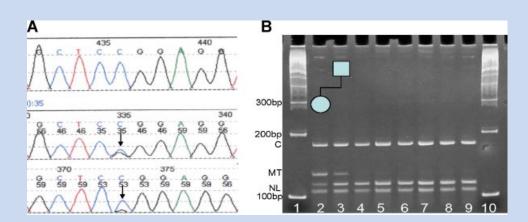


FIG. 3. A: Chromatogram of R254G mosaic mutation from Family 7 indicated by arrow. Top panel is the reference sequence. Middle panel is the female proband. Lower panel is the father's mosaic mutation. B: Restriction digest of the R254G mutation. The mutation destroys a Msp1 site. C = constant restriction site; MT = mutant band; NL = normal band. Lanes 1 and 10: 100 bp marker; lane 2: affected daughter's sample; lane 3: father's sample; lanes 4–9 normal control samples.

prominent philtrum. Lin et al. [2009] discussed that the abnormally short philtrum and bilateral notched vermilion-mucosa border are on the spectrum of microform CL and noted the absence of isolated CP in BOFS.

"Branchial" or pharyngeal arch involvement, typically manifesting as the cervical cutaneous anomaly is a classic finding in patients with BOFS [Lin et al., 1995]. Nevertheless, three described probands from Families 21, 22, and 28 did not have this anomaly (examined by geneticists A.R, [London], A.D., and M.W.). Two additional patients were viewed as having a very minor expression in the form of faint hair on the neck [Patient 27; Stoetzel et al., 2009, Patient 1.2]. Interestingly, Family 21 and 28 have the same mutation (E242K); however, the mutation was also found in another classic BOFS patient (Family 23) with bilateral cervical cutaneous anomalies. The third family (Family 22) had a missense mutation in the helix-span-helix region of the DNA binding and dimerization domain. Further families without this classic cervical anomaly should be genotyped in order to establish a more precise genotype-phenotype correlation, if one exists. Nevertheless, the lack of the cervical cutaneous anomaly further broadens the variability of the BOF diagnosis, as has recently been reported by Reiber et al. [2010a].

Twenty-eight of 40 (70%) patients investigated had hearing loss, mostly bilateral and categorized as sensorineural, conductive, or mixed. Additional details are not readily available from the majority of patients regarding hearing frequency, progression and effectiveness of amplification. Further studies to address these questions would be helpful. Echocardiography is uncommonly performed in BOFS, and the occurrence of atrial septal defect (one ach in our series and the literature) may not reflect true occurrence. Nevertheless, tetralogy of Fallot [Reiber et al., 2010a] may have some importance as a conotruncal defect in the analysis of the developmental mechanism (see below).

A large percentage of patients (12/34; 35%) that were investigated had various renal anomalies, including dysplasia, agenesis, multicystic kidneys, and vesicoureteral reflux. These data support obtaining renal ultrasonography in the diagnostic evaluation of possible BOFS patients.

Premature graying of hair or poliosis is likely another underreported frequent finding in BOFS families. In our series, 14/37 (38%) individuals had this feature; it was especially prevalent in the more mildly affected older generations who harbored the familial deleterious mutation. Mégarbané et al. [1998] initially reported poliosis in a daughter and her father with BOFS (Family 8). This underscores that clinical screening of first-degree relatives of BOFS patients should include some of the less obvious phenotypes (as some individuals dye their hair). Interestingly, Patient 14, in addition to her premature graving and characteristic BOFS features, also had heterochromia of the irides, another feature of neurocristopathies. This is not surprising as TFAP2A is known to be expressed in premigratory and migratory neural crest cells [Hilger-Eversheim et al., 2000; Li and Cornell, 2007]. Thus, premature graving hair, sensorineural deafness, inner ear/temporal bone anomalies, coloboma, heterochromia, facial nerve weakness, tetralogy of Fallot, and ectopic thymus provide clinical support for neural crest involvement in BOFS. We had originally reported that Patients 2 and 5 had anxiety and depression [Milunsky et al., 2008]. In surveying the rest of the BOFS patients, we found another three who had psychiatric symptoms. It is difficult to determine if the psychiatric symptomatology is related to the psychosocial context of having a craniofacial disorder or related to their genotype. As previously reported, the AP-2 family may be involved in the regulation of the monoaminergic systems in the adult brain, resulting in neuropsychiatric disorders [Damberg, 2005]. It would seem prudent to inquire about such symptoms in the diagnostic and follow-up evaluations of BOFS patients.

The molecular spectrum in 30 families with 41 affected individuals with BOFS includes heterozygous missense mutations, a frameshift mutation, and a complete deletion of the *TFAP2A* gene. The frequency of partial or whole gene *TFAP2A* deletions remains unknown, but appears to be low. More deletion cases need to be identified to determine their frequency and whether lesser forms of CL are present in these patients. The data from Gestri et al. [2009] clearly demonstrate that a deletion of the *TFAP2A* gene results in a variety of ophthalmologic anomalies that have been reported in BOFS, including coloboma (typically posterior segment, but also iris), microphthalmia, nasolacrimal duct stenosis, and cataract. These ocular anomalies have also been described in patients with larger deletions including the *TFAP2A* gene [Davies et al., 1999; Misceo et al., 2008].

The intrafamilial clinical variability was marked in Families 1, 7, 8, and 19 (see Table II). This variability could be explained in Family 7 by the apparent mosaicism detected on sequencing and restriction digestion. Additional tissues from the father in Family 7 have been requested to further demonstrate the likely mosaicism. Given this marked intrafamilial variability, testing of parents of a molecularly confirmed case of BOFS is recommended for more accurate recurrence risk counseling. In addition to mosaicism, modifier genes are likely playing a role in the clinical variability seen in families. Once a mutation has been established, genetic counseling and prenatal diagnosis would be available.

We have thus far not demonstrated any genetic heterogeneity for those cases fulfilling the clinical criteria previously discussed.

We have identified a hotspot region in the highly conserved exons 4 and 5 (basic region of the DNA binding and dimerization domain) of *TFAP2A* that harbors missense mutations in 27/30 (90%) (Fig. 2) families. Several of these mutations are recurrent and are listed in Table III. One family harbors a mutation in exon 7 of the *TFAP2A* gene (helix-span-helix region of the DNA binding and dimerization domain). The proband in this family did not have obviously different clinical findings than our other patients. Not surprisingly, the proband of Family 29 did not have patently different clinical findings than our other patients; harboring a frameshift mutation in exon 2 leading to disruption of the DNA binding and dimerization domain.

In addition to the whole gene deletion cases reported by Gestri et al. [2009], they also described an individual with classic BOFS with a deletion between amino acid residues 233 and 236. Tekin et al. [2009] discovered a complex *TFAP2A* allele (deletion of 18 and insertion of 6 nucleotides) between amino acids 276 and 281 that altered amino acids in the basic DNA binding and dimerization domains. That patient also had a phenotype characteristic of BOFS. Stoetzel et al. [2009] reported one family and two sporadic cases with missense mutations in exon 4 (S239P, L249P, and L218P). We had previously found the L249P in Patient 3 [Milunsky et al., 2008], and noted another change (L218R) of the same amino acid residue in this report. Reiber et al. [2010a] published five patients with BOFS (two familial and three sporadic) with mutations in exons 4–6. They noted the recurrent R255G mutation in a familial and sporadic case [mutation seen in Case 2 in Milunsky et al., 2008] and a mutation of amino acid residue 237 that was found in three patients in this report (see Tables II and III). They also found two novel missense mutations in exon 5 (L269P) and exon 6 (E296K).

Implications for Management

BOFS is a multisystem disorder that requires multidisciplinary care, including genetic counseling for affected families. This article emphasizes that absence of the "B" feature (cutaneous defect) does not exclude the diagnosis, and thus, patients who may have been followed as "possible BOFS" should be reconsidered with the advantage of molecular analysis. Management recommendations are summarized in Table IV. Ophthalmologic evaluation has always been essential because of the various ocular anomalies and the potential for visual limitation. Craniofacial surgery should be performed by a plastic surgeon experienced in treating children with orofacial clefts and congenital facial anomalies, preferably in the setting of a CP or craniofacial center. Lesser forms of CL, formerly known as "pseudocleft" may need surgical repair although seemingly minor defects, and may represent a minimal expression of the BOFS gene [Lin et al., 2009]. The recent reports of temporal bone anomalies are compelling evidence that aggressive evaluation of hearing loss should be accompanied by CT imaging. Given the frequency of kidney anomalies, renal ultrasonography is recommended in the initial diagnostic evaluation of BOFS patients. Congenital heart defects are probably rare, but the detection of a murmur should prompt referral for echocardiography. Although intelligence is usually normal, challenges to learning mean that an educational interventional program and formal psychometric testing should be arranged. Given the potential risks for psychiatric problems and predisposition to cancer, although routine testing is not recommended, continued monitoring seems prudent. The intrafamilial variability of BOFS appears significant, and warrants molecular testing of parents for more accurate recurrence risk counseling. Finally, the results of this study indicate that although genotyping does not predict a specific phenotype, further investigation is needed.

CONCLUSIONS

This is the largest series of patients with BOFS with genotyping of *TFAP2A*. Identification of a hotspot region and recurrent mutations remain important to the molecular diagnostic strategy in clinical laboratories. The diagnosis of the BOFS remains clinical in most patients. Nevertheless, molecular testing for *TFA-P2A* mutations will assist geneticists in confirming apparently typical cases or excluding the diagnosis in atypical cases. Aside from the data presented on the patients with a deletion, there does

TABLE IV. Management of Individuals With BOFS **Clinical issue** Guideline Ideally, follow children in a multidisciplinary cleft palate or craniofacial clinic setting General Branchial (cutaneous) When superficial or small, these may heal spontaneously. Usually, require surgical excision; a sinus tract defects must be dissected. This should be done only by an experienced pediatric plastic surgeon. Exploration for a thymic remnant may be necessary, and if detected, this should be sent for histopathologic examination Ophthalmologic defects Complete ophthalmologic examination is needed because of visual limitation and strabismus, as well as an evaluation for nasolacrimal duct patency Facial anomalies: Surgical treatment should be done only by a pediatric plastic surgeon experienced in treating children with orofacial clefts cleft lip. Lesser forms of cleft lip, formerly known as "pseudocleft" may need surgical correction Nose In addition to the nasal tip flattening or asymmetry that may be associated with cleft lip, there may be a characteristic full, flat nasal tip which may need a corrective procedure Teeth Hypoplastic or absent teeth should be carefully monitored Ears Malformed protruding pinnae may require surgical correction. If diagnosed in early infancy, auricular molding may be indicated Hearing In addition to the newborn hearing screen, a referral to an audiologist is essential. The recent reports of temporal bone anomalies are compelling evidence that CT imaging should be done to anticipate optimal hearing correction Heart defect Echocardiogram indicated if there is a murmur or symptomatic Kidney defect Renal ultrasonography recommended at the time of diagnosis Development Speech therapy is likely and learning challenges are common if there are visual and hearing special needs Psychologic issues Monitor Cancer Monitor Genetic issues Confirmation of the TFAP2A mutation should be performed by a CLIA-approved laboratory. Parental targeted molecular analysis recommended given significant phenotypic variability. Genetic counseling to review the inheritance and reproductive risks should be provided

not appear to be mutation-specific genotype-phenotype correlations at this time, but more patients need to be studied.

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REFERENCES

- Amiel J, Attieé-Bitach T, Marianowski R, Cormier-Daire V, Abadie V, Bonnet D, Gonzales M, Chemouny S, Brunelle F, Munnich A, Manach Y, Lyonnet S. 2001. Temporal bone anomaly proposed as a major criteria for diagnosis of CHARGE syndrome. Am J Med Genet 99:124–127.
- Bennaceur S, Buisson T, Bertolus C, Couly G. 1998. Branchio-oculo-facial syndrome with cleft lip and bilateral dermal thymus. Cleft Palate Craniofac J 35:454–459.
- Ceruti S, Stinckens C, Cremers CW, Casselman JW. 2002. Temporal bone anomalies in the branchio-oto-renal syndrome: Detailed computed tomographic and magnetic resonance imaging findings. Otol Neurotol 23:200–207.
- Damberg M. 2005. Transcription factor AP-2 and monoaminergic functions in the central nervous system. J Neural Transm 112:1281–1296.
- Davies AF, Mirza G, Flinter F, Ragoussis J. 1999. An interstitial deletion of 6p24-p25 proximal to the FKHL7 locus and including AP-2 α that affects anterior eye chamber development. J Med Genet 36:708–710.
- Fielding DW, Fryer AE. 1992. Recurrence of orbital cysts in the branchiooculc-facial syndrome. J Med Genet 29:430–431.
- Fujimoto A, Lipson M, Lacro RV, Shinno RW, Boelter WD, Jones KL, Wilson MG. 1987. New autosomal dominant branchio-oculo-facial syndrome. Am J Med Genet 27:943–951.
- Gestri G, Osborne RJ, Wyatt AW, Gerrelli D, Gribble S, Stewart H, Fryer A, Bunyan DJ, Prescott K, Collin JR, Fitzgerald T, Robinson D, Carter NP, Wilson SW, Ragge NK. 2009. Reduced TFAP2A function causes variable optic fissure closure and retinal defects and sensitizes eye development to mutations in other morphogenetic regulators. Hum Genet 126:791–803.
- Glueckert R, Rask-Andersen H, Sergi C, Schmutzhard J, Mueller B, Beckmann F, Rittinger O, Hoefsloot LH, Schrott-Fischer A, Janecke AR. 2010. Histology and synchrotron radiation-based microtomography of the inner ear in a molecularly confirmed case of CHARGE syndrome. Am J Med Genet Part A 152A:665–673.
- Hilger-Eversheim K, Moser M, Schorle H, Buettner R. 2000. Regulatory roles of AP-2 transcription factors in vertebrate development, apoptosis and cell cycle-control. Gene 260:1–12.

- Li W, Cornell R. 2007. Redundant activities of Tfap2a and Tfap2c are required for neural crest induction and development of other non-neural ectoderm derivatives in zebrafish embryos. Dev Biol 304:338–354.
- Lin AE. 2009. Cardinal signs. Branchio-Oculo-Facial syndrome (BOFS Abstract). Am Coll Med Genet Annual Meeting, Tampa, Florida, March 28, 2009.
- Lin AE, Gorlin RJ, Lurie IW, Brunner HG, van der Burgt I, Naumchik IV, Rumyantseva NV, Stengel-Rutkowski S, Rosenbaum K, Meinecke P, et al. 1995. Further delineation of the branchio-oculo-facial syndrome. Am J Med Genet 56:42–59.
- Lin AE, Yuzuriha S, McLean S, Mulliken JB. 2009. Lesser forms of cleft lip associated with the branchio-oculo-facial syndrome. J Craniofac Surg Supplement 1:608–611.
- Mégarbané A, Hawat N, Chedid P, Bleik J, Délézoïde AL. 1998. Branchiooculo-facial syndrome associated with a white forelock. Clin Dysmorphol 7:221–223.
- Milunsky JM, Maher TA, Zhao G, Roberts AE, Stalker HJ, Zori RT, Burch MN, Clemens M, Mulliken JB, Smith R, Lin AE. 2008. TFAP2A mutations result in branchio-oculo-facial syndrome. Am J Hum Genet 82:1171–1177.
- Misceo D, Bjørgo K, Ormerod E, Ringen Ø, Rocchi M, Birger van der Hagen C, Frengen E. 2008. A de novo 6p interstitial deletion and a complex translocation involving chromosomes 2, 6, and 14 in a mildly developmentally delayed patient. Am J Med Genet Part A 146A: 3230–3233.
- Raveh E, Papsin BC, Forte V. 2000. Branchio-oculo-facial syndrome. Int J Pediatr Otorhinolaryngol 53:149–156.
- Reiber J, Sznajer Y, Guillén Posteguillo E, Müller D, Lyonnet S, Baumann C, Just W. 2010a. Additional clinical and molecular analyses in patients with branchio-oculo-facial syndrome. Am J Med Genet Part A 152A:994–999.
- Reiber J, Sznajer Y, Guillén Posteguillo E, Müller D, Lyonnet S, Baumann C, Just W. 2010b. Additional clinical and molecular analyses of TFAP2A in patients with the branchio-oculo-facial syndrome: Previously reported patient. Am J Med Genet Part A 152A:2143.
- Stoetzel C, Riehm S, Bennouna Greene V, Pelletier V, Vigneron J, Leheup B, Marion V, Hellé S, Danse JM, Thibault C, Moulinier L, Veillon F, Dollfus H. 2009. Confirmation of TFAP2A gene involvement in branchio-oculofacial syndrome (BOFS) and report of temporal bone anomalies. Am J Med Genet Part A 149A:2141–2146.
- Tekin M, Sirmaci A, Yüksel-Konuk B, Fitoz S, Sennaroğlu L. 2009. A complex TFAP2A allele is associated with branchio-oculo-facial syndrome and inner ear malformation in a deaf child. Am J Med Genet Part A 149A:427–430.
- Zentner GE, Layman WS, Martin DM, Scacheri PC. 2010. Molecular and phenotypic aspects of CHD7 mutation in CHARGE syndrome. Am J Med Genet Part A 152A:674–686.Review.