## DISPATCHES

## Global Distribution of Mycobacterium tuberculosis Spoligotypes

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We present a short summary of recent observations on the global distribution of the major clades of the *Mycobacterium tuberculosis* complex, the causative agent of tuberculosis. This global distribution was defined by data-mining of an international spoligotyping database, SpoIDB3. This database contains 11,708 patterns from as many clinical isolates originating from more than 90 countries. The 11,708 spoligotypes were clustered into 813 shared types. A total of 1,300 orphan patterns (clinical isolates showing a unique spoligotype) were also detected.

**S** ince the publication of the second version of our spoligotypes database on *Mycobacterium tuberculosis* (1), the causative agent of tuberculosis (TB), the proportion of clustered isolates (shared types [STs]) increased from 84% (2,779/ 3,319) to 90% (11,708/13,008). Fifty percent of the clustered isolates were found in only 20 STs. Three of these isolates are *M. bovis*, including *M. bovis* BCG (ST 481, 482, and 683). The addition of the next 30 most frequent STs increased the total proportion of clustered isolates (65% instead of 50% initially).

A total of 36 potential subfamilies or subclades of M. tuberculosis complex have been tentatively identified, leading to the definition of major and minor visual recognition rules (Table). The ancestral East-African Indian family (EAI) is made up of at least five main subclades, whereas at least three major spoligotyping patterns are found within the Haarlem family (2). Two families found in central and Middle Eastern Asia (CAS1 and CAS2) are newly defined. The X family (3) is also currently split into at least three well-defined subclades. However, the subdivision of family T (T1-T4, likely to represent relatively old genotypes), which differs from the classic ST 53 (all spacers present except 33-36), remains poorly defined. Similarly, the Latino-American and Mediterranean family (LAM) is tentatively split into subclades LAM1-LAM10 (4). Spoligotyping used alone is not well suited for studying the phylogeny of these two clades (T and LAM). Such study will require results from other genotyping methods such as IS6110-restriction fragment length polymorphism (5) or mycobacterial interspersed repetitive units-variable number of DNA tandem repeats (6). Among well-characterized major clades of tubercle bacilli, four families represent 35% of 11,708 clustered isolates (Beijing 11%, LAM 9.3%, Haarlem 7.5%, and the X clade 7%).

The global distribution of the most frequently observed spoligotypes by continent in SpolDB3 is as follows. Among the patterns originating in North America (n= 4,276, 32% of the total number of isolates in the database), 16% of the strains are of the Beijing type, 14% belong to ST 137 or ST 119 (X family), and 8% are unique (results not shown). In Central America (n=587, 4.5%), 8% of the strains belong to the ubiquitous ST 53, 7% are ST 50, and 6% are ST 2; the last two STs are part of the Haarlem family. In South America (n=861, 6.6%), the distribution of ST 53 and ST 50 accounts for 10% and 9%, respectively, of the spoligotypes, whereas ST 42 accounts for as much as 9% of the total isolates. The origin of ST 42 remains to be established. In Africa (n=1,432, 11%), ST 59 and ST 53 account for 9% of all isolates studied thus far; however, the values obtained for ST 59 are biased because strains from Zimbabwe are overrepresented. We also observed that M. africanum ST 181 accounts for as much as 6% of all spoligotypes from Africa in our sample.

In Europe (n=4,360, 33.5%), ST 53 represents as much as 9% of the spoligotypes, ST 50 and 47 (Haarlem family) repre-

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Rk	ST	Class <sup>b</sup>	Total (n) <sup>c</sup>	Rules <sup>d</sup>	Binary description	Octal
1	1	Beijing	1282	Δ1–34		00000000003771
2	53	T1	864	F		777777777760771
11	52	T2	163	$\Delta 40$ and F		77777777760731
30	37	T3	71	$\Delta 13$ and F		777737777760771
64	40	T4	26	$\Delta 19$ and F		777777377760771
7	47	Haarlem1	246	$\Delta 26$ –30 and E		777777774020771
20	2	Haarlem2	104	$\Delta$ 1–24, $\Delta$ 26–30 and E		00000004020771
3	50	Haarlem3	519	Е		777777777720771
6	119	X1	310	С		777776777760771
4	137	X2	427	C and Δ39–42		777776777760601
31	92	X3	70	$\Delta$ 4–12 and C		700036777760731
15	48	EAI1	118	A and $\Delta 40$		77777777413731
13	19	EAI2	130	$\Delta 3$ , $\Delta 20$ –21 and A		677777477413771
16	11	EAI3	121	$\Delta$ 2-3, A and $\Delta$ 37–39		477777777413071
8	139	EAI4	234	$\Delta 26$ –27 and A		77777774413771
46	236	EAI5	41	А		77777777413771
24	181	Afri1	91	$\Delta79$ and $\Delta39$		77077777777671
ND	331	Afri2	9	$\Delta 8$ –12, $\Delta 21$ –24 and $\Delta 37$ –39		774077607777071
ND	438	Afri3	3	$\Delta 8$ –12 and $\Delta 37$ –39		774077777777071
17	482	M. bovis-BCG	26	$\Delta 3$ , $\Delta 9$ , $\Delta 16$ and D		67677377777600
ND	641	M. microti	8	4-7, 23–24, 37–38		074000030000600
ND	592	M. canetti	6	30 and 36		00000000101000
21	26	CAS1	102	Δ4–7, Δ23–34		703777740003771
ND	288	CAS2	6	Δ4–10, Δ23–34		700377740003771
12	20	LAM1	152	$\Delta 3$ and B		677777607760771
22	17	LAM2	92	$\Delta 3$ , $\Delta 13$ and B		677737607760771
19	33	LAM3	108	$\Delta 9$ –11 and B		776177607760771
49	60	LAM4	37	$\Delta 40$ and B		777777607760731
42	93	LAM5	44	$\Delta 13$ and B		777737607760771
37	64	LAM6	47	$\Delta 29$ and B		777777607560771
36	41	LAM7	48	$\Delta 20,\Delta 26\text{-}27$ and B		777777404760771
NA	290	LAM8	9	$\Delta 27$ and B		777777606760771
5	42	LAM9 <sup>e</sup>	344	В		777777607760771
9	61	LAM10	202	$\Delta 23$ –25 and F		777777743760771
26	34	$\mathbf{S}^{\mathbf{f}}$	82	$\Delta 9$ –10 and F		776377777760771
28	451	H37Rv	78	$\Delta 20$ –21 and F		777777477760771

Table. Excerpt from SpoIDB3 database showing prototype spoligotypes, visual recognition rules, and binary and octal description<sup>a</sup>

<sup>a</sup>Rk, ranking no.; ND, not done; ST, arbitrary designation; *M., Mycobacterium*.

 <sup>RK</sup>, ranking no.; ND, not done; S1, arbitrary designation; *M., Mycobacterium*.
<sup>b</sup>Class: family definition. See text for the definition of the family acronyms.
<sup>c</sup>Total (n), size of the class; binary and octal, description.
<sup>d</sup>Rule A, absence of spacers 29–32, presence of spacer 33 and absence of spacer 34; rule B, absence of spacers 21–24 and spacers 33–36; rule C : absence of spacer 18 and spacers 33– 36; rule D, absence of spacers 39-43; rule E, absence of spacer 31 and spacers 33-36; rule F, absence of spacers 33-36. Clades defined with low sample size, such as Afri2, Afri3, CAS2, and LAM8 are subject to change.

eFormerly LAM1.

<sup>f</sup>Formerly LAM2

sent 8% of the cases, and the Beijing family accounts for 4% of the spoligotypes. In the Middle Eastern and central Asian region, where the number of samples obtained is still very low (n=351, 2.7%), a high diversity of strains within the EAI and CAS families has been observed, and no single pattern currently exceeds 5%. Further studies of isolates from these regions are needed, e.g., in India, where our sampling is still anecdotal (n=44 isolates). Notwithstanding the scarcity of available data from this region, the observed diversity suggests that this region might be of great interest for further study of the genetic variation of tubercle bacilli. Contrary to what we observed for the Middle East and central Asia, the Far East Asian region (n=801, 6.1%) is characterized by the prevalence of a single genotype, the Beijing type family, a family linked to emerging multiresistance (7). One out of two strains in the Far East is a Beijing type. In Oceania (n=340, 2.6%), ST 19 and Beijing account for 15% and 13%, respectively, of clustered isolates. Thus, this preliminary analysis of the spoligotype distribution of SpolDB3 clearly shows major differences in the population structure of tubercle bacilli within the eight subcontinents studied (Africa; Europe; North America; Central America; South America; Middle East and Central Asia; Far East Asia; and Oceania).

At present, SpoIDB 3 is an experimental tool that has yet to prove its usefulness in tracking epidemics. Nevertheless, the facility with which matches between spoligotypes can be detected suggests that this tool may be a good screening mechanism for population-based studies on recent TB transmission. Indeed, the detection of a rarely found ST in SpoIDB3 may be a catalyst that signals researchers to look for the clonality of the isolates and to study their epidemiologic relatedness.

Data-exchange protocols through inter-networking will also be implemented in the near future. Working groups such as the European Network for Exchange of Molecular Typing Information (available from: URL: www.rivm.nl/enemti) are coordinating such initiatives. The expanded use of the Bionumerics software (third upgrade; Applied Maths, St. Martens-Latem, Belgium) may also foster this research field. SpolDB3 will also be instrumental in facilitating better understanding of the driving forces that shape tubercle bacilli evolution. Further research should now emphasize the use of data-mining methods, in combination with experts' knowledge, to tackle the complex dynamics of the population's genetics of tubercle bacilli and TB transmission (3). Our sample represents the compilation of many national studies and, as such, should be considered as an ongoing population-based project aimed at studying global TB genetic diversity. Nevertheless, obtaining a more precise and representative snapshot of the genetic variability of *M. tuberculosis* complex will require a larger sampling. Although only partially representative of worldwide spoligotypes of *M. tuberculosis* complex, Spo1DB3 contains a reservoir of genetic information that has already proved useful for defining the phylogenetic links that exist within the TB

genomes and for constructing theoretical models of genome evolution. Much remains to be done to evaluate the potential of global genetic databases to better characterize casual contacts (that could lead to identification of sporadic cases) in TB epidemiology. An improved version of our database, which will focus on areas with a high prevalence of TB, is currently in development; as of August 26, 2002, it had 20,000 isolates and 3,000 alleles. Ongoing population-based genotyping projects will likely help shed light on contemporary and ancient tubercle bacilli's evolutionary history.

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