Opinion

H5N1 Viruses and Vaccines

Kanta Subbarao*, Catherine Luke

he establishment and spread of highly pathogenic avian influenza (HPAI) viruses of the H5N1 subtype in birds and coincident infections in humans since 2003 have raised concerns that we may be facing an influenza pandemic caused by an H5N1 influenza virus. In this brief Opinion piece, we consider the pandemic threat posed by H5N1 viruses and review the published data on the evaluation of H5N1 vaccines in preclinical and clinical studies.

HPAI H5N1 viruses have been isolated from avian species in more than 50 countries. As of 29 January 2007, 270 laboratory-confirmed cases of H5N1 infection in humans had been reported by the World Health Organization, 164 of which were fatal [1], resulting in a case fatality rate of approximately 60%.

In order to cause a pandemic, H5N1 viruses will have to acquire the ability to transmit efficiently from person to person. The H5 hemagglutinin (HA) is found in influenza viruses that typically infect avian species, so efficient personto-person spread could happen if the H5N1 virus reassorts, or exchanges genes, with circulating human influenza viruses giving rise to a virus with the H5 HA (to which the population is not immune) in a gene constellation that confers the property of transmissibility. Alternatively, efficient personto-person spread could occur if the H5N1 virus evolves and adapts to more efficient replication and transmissibility in the human population.

Two observations have led to questions about the likelihood of a reassortant H5N1 virus causing a pandemic. First, reassortant viruses have not been isolated despite ongoing H5N1 outbreaks in birds and infections in humans, even with concurrent circulation of human influenza viruses since 2003. Second, laboratory studies have found that reassortant viruses that derived the surface glycoprotein genes from an H5N1 virus and internal protein genes from an H3N2 influenza A virus were not efficiently transmitted and were somewhat less infectious to ferrets (an animal model for human influenza) than the wild-type H5N1 viruses [2]. The concern that an H5N1 virus could adapt to the human host and acquire mutations that confer transmissibility prompts very careful analysis of each cluster of human H5N1 infections that is reported ([1,3-5]). At present, the data suggest that human-to-human transmission is inefficient and very limited. Nevertheless, from the standpoint of public health preparedness, it is important to move forward in developing approaches for dealing with H5N1 in humans.

Vaccination is the preferred strategy for prevention and control of influenza. The most expeditious way to generate an H5N1 vaccine is to use licensed technology, such as inactivated or live attenuated vaccines. However, several practical and scientific challenges to the development of H5N1 vaccines exist. These include high pathogenicity of wild-type H5N1 influenza viruses, reduced yield of candidate vaccine viruses in embryonated hens' eggs compared to that of human influenza viruses, limited manufacturing capacity, and poor immunogenicity of the H5 HA. Despite these obstacles, several approaches have been used to generate candidate vaccines and a few have advanced to clinical trials (Table 1). Table 1 also includes data published on vaccines that are being developed for veterinary use.

Perhaps the most significant scientific challenge for the development and licensure of pandemic vaccines for humans is that assessment of vaccine efficacy for humans will have to be inferred from preclinical studies in experimental animals and immunogenicity studies in humans, as it will not be possible to assess the efficacy of a pandemic vaccine in a clinical trial before a pandemic begins. Table 2 summarizes the preclinical and clinical findings from inactivated H5N1 vaccines evaluated in humans to date. Preclinical studies of influenza vaccines are generally conducted in mice or ferrets. In most cases, the 1997 and 2003 H5N1 vaccine candidates were promising in terms of immunogenicity and efficacy, with complete protection of animals from lethal H5N1 infection, and significant, if not complete, reduction of pulmonary viral replication following challenge. Preclinical data in ferrets have not been published on the 2004 H5N1 vaccines that were evaluated in clinical trials, so data are not available to directly assess how accurately preclinical studies would have predicted the outcome of evaluation of these vaccines in humans.

In clinical trials, inactivated virus vaccines based on H5N1 viruses isolated in 2004 [6,7], a recombinant H5 HA subunit vaccine based on an H5N1 virus isolated in 1997 expressed in a baculovirus vector [8], and an inactivated virus vaccine based on a surrogate low pathogenicity avian H5N3 virus [9-11], were poorly immunogenic when administered to volunteers without adjuvant. Clinical trials of H1N1 influenza vaccines in 1977 established that whole virion vaccines are more immunogenic than split-virion vaccines (in which the virus particles are disrupted by detergent treatment to obtain a preparation enriched for the surface antigens) [12,13]; however, the former are also more reactogenic than the latter. Consistent with this observation, in recent trials in humans of an alum-adjuvanted inactivated H5N1 virus vaccine, much lower doses of a whole virion vaccine elicited higher levels of antibody compared to a split-virion vaccine

Editor: Marianne Manchester, The Scripps Research Institute, United States of America

Citation: Subbarao K, Luke C (2007) H5N1 viruses and vaccines. PLoS Pathog 3(3): e40. doi:10.1371/journal.ppat.0030040

This is an open-access article distributed under the terms of the Creative Commons Public Domain declaration which stipulates that, once placed in the public domain, this work may be freely reproduced, distributed, transmitted, modified, built upon, or otherwise used by anyone for any lawful purpose.

Abbreviation: HA, hemagglutinin

Kanta Subbarao and Catherine Luke are with the Laboratory of Infectious Diseases, National Institute of Allergy and Infectious Diseases, Bethesda, Maryland, United States of America.

* To whom correspondence should be addressed. E-mail: ksubbarao@niaid.nih.gov

Table 1. Vaccine Strategies against H5N1 Influenza that Have Been Evaluated in Preclinical and Clinical Studies

Type of Vaccine	Published Studies in which Indicated Vaccine Has Been Evaluated		Intended Use	References
	Preclinical Studies	Clinical Trials		
				[4,4,00]
Inactivated whole virus			Human	[14-20]
Inactivated subvirion	×		Human	[6,7]
Inactivated, surrogate low	1		Human	[9–11,21–23]
pathogenicity avian H5 virus				
Live attenuated virus	Lan .	×	Human	[24–27]
Subunit (surface glycoprotein	1	∠ ^a	Human	[8,28]
preparation or recombinant H5 HA)				
Adenovirus vectored H5		×	Human and veterinary	[29,30]
Fowlpox vectored H5		×	Veterinary	[31,32]
Newcastle disease virus vectored H5		×	Veterinary	[33,34]
DNA (H5 or NP/M)		×	Human	[35,36]

^aRecombinant H5 HA [8].

doi:10.1371/journal.ppat.0030040.t001

Table 2. Summary of Preclinical and Clinical Findings for Inactivated H5N1 Virus Vaccines Evaluated in Humans

Vaccine	Virus	Published Preclinical Findings	Published Clinical Trial Results	References
Inactivated subvirion	A/duck/Singapore/ 97 (H5N3)	Two doses ± alum required to elicit HI Ab in 65% of mice; detected Ab cross-reactive with H5N1 viruses. High degree of protection from lethality, pulmonary and extrapulmonary infection following challenge.	Two doses of 7.5, 15, or 30 μg, 3 wk apart, with or without MF59 adjuvant. Vaccine was well-tolerated at all doses.	[9–11,21,23]
	A/VN/1203/2004 PR8 reassortant	None published	Poorly immunogenic without adjuvant. Two doses of 90, 45, 15, or 7.5 μg 4 wk apart. Vaccine was well tolerated	[6]
			Two doses of 90 µg elicited NAb in 54% of individuals and HI Ab titers ≥1:40 in 58% of individuals.	
	A/VN/1194/2004 PR8 reassortant	None published	7.5, 15, or 30 μg, 2 doses, 3 wk apart, with or without alum adjuvant.	[7]
			Highest Ab responses were seen after 30 µg with adjuvant.	
Inactivated whole virion	A/HK/213/2003 PR8 reassortant	In mice, a single dose of 7 or 15 μ g with incomplete Freund's adjuvant elicited high levels of HI Ab and NAb and provided protection from pulmonary virus replication and lethal challenge. In ferrets, one dose (7 μ g or 15 μ g) with alum adjuvant or two doses without adjuvant (7 μ g) induced a protective Ab response and complete protection from lethal challenge with homologous wild-type virus, with significantly reduced lung virus titers. All ferrets were protected from lethal challenge with the heterologous A/VN/1203/04 wild-type virus.	Not done	[16,17,19]
	A/VN/1194/2004 PR8 reassortant	None published	Two doses of 1.25, 2.5, 5, or 10 μ g HA with aluminum hydroxide 4 wk apart.	[14]
			Vaccines were well tolerated. Ab was detected after one dose, and two doses of 10 µg resulted in seropositivity in 78% of individuals. Two doses of all doses met EMEA requirements for seasonal influenza vaccine licensing.	

Ab, antibody; EMEA, European Agency for the Evaluation of Medicinal Products; HI, hemagglutination inhibiting; NAb, neutralizing antibody; PR8, Influenza A/Puerto Rico/8/34 (H1N1). doi:10.1371/journal.ppat.0030040.t002

Clinical trials have demonstrated that the immunogenicity of H5 vaccines can be enhanced by an increased dose of the HA, the use of adjuvants, use of multiple doses, or use of a whole virion vaccine. More studies are needed to directly compare findings from preclinical and clinical evaluation of pandemic influenza vaccines to establish whether animal models can be used to guide decisions on which vaccine candidates to take forward for evaluation in humans. Although there is no evidence that H5N1 viruses have yet acquired pandemic potential, the consequences of such an event are serious enough that preparation for a possible pandemic is essential.

Acknowledgments

Author contributions. KS and CL wrote the paper.

Funding. This research was supported in part by the Intramural Research Program of the US National Institutes of Health, National Institute of Allergy and Infectious Diseases.

Competing interests. Our laboratory has a cooperative research and development agreement with MedImmune Vaccines to develop vaccines against potential pandemic strains of influenza.

References

- World Health Organization (2007) Disease outbreak news. Available: http://www.who.int/csr/don/en. Accessed 2 February 2007.
- Maines TR, Chen LM, Matsuoka Y, Chen H, Rowe T, et al. (2006) Lack of transmission of H5N1 avian-human reassortant influenza viruses in a ferret model. Proc Natl Acad Sci U S A 103: 12121–12126.
- Ungchusak K, Auewarakul P, Dowell SF, Kitphati R, Auwanit W, et al. (2005) Probable person-to-person transmission of avian influenza A (H5N1). N Engl J Med 352: 333–340.
- Oner AF, Bay A, Arslan S, Akdeniz H, Sahin HA, et al. (2006) Avian influenza A (H5N1) infection in eastern Turkey in 2006. N Engl J Med 355: 2179–2185.
- Kandun IN, Wibisono H, Sedyaningsih ER, Yusharmen, Hadisoedarsuno W, et al. (2006) Three Indonesian clusters of H5N1 virus infection in 2005. N Engl J Med 355: 2186–2194.
- Treanor JJ, Campbell JD, Zangwill KM, Rowe T, Wolff M (2006) Safety and immunogenicity of an inactivated subvirion influenza A (H5N1) vaccine. N Engl J Med 354: 1343–1351.
- Bresson JL, Perronne C, Launay O, Gerdil C, Saville M, et al. (2006) Safety and immunogenicity of an inactivated split-virion influenza A/Vietnam/ 1194/2004 (H5N1) vaccine: Phase I randomised trial. Lancet 367: 1657–1664.
- Treanor JJ, Wilkinson BE, Masseoud F, Hu-Primmer J, Battaglia R, et al. (2001) Safety and immunogenicity of a recombinant hemagglutinin vaccine for H5 influenza in humans. Vaccine 19: 1732–1737.
- Nicholson KG, Colegate AE, Podda A, Stephenson I, Wood J, et al. (2001) Safety and antigenicity of non-adjuvanted and MF59-adjuvanted influenza A/Duck/Singapore/97 (H5N3) vaccine: A randomised trial of two potential vaccines against H5N1 influenza. Lancet 357: 1937–1943.
- Stephenson I, Nicholson KG, Colegate A, Podda A, Wood J, et al. (2003) Boosting immunity to influenza H5N1 with MF59-adjuvanted H5N3 A/ Duck/Singapore/97 vaccine in a primed human population. Vaccine 21: 1687–1693.
- Stephenson I, Bugarini R, Nicholson KG, Podda A, Wood JM, et al. (2005) Cross-reactivity to highly pathogenic avian influenza H5N1 viruses after vaccination with nonadjuvanted and MF59-adjuvanted influenza A/Duck/ Singapore/97 (H5N3) Vaccine: A potential priming strategy. J Infect Dis 191: 1210–1215.
- 12. Wright PF, Dolin R, La Montagne JR (1976) From the National Institute of Allergy and Infectious Diseases of the National Institutes of Health, the Center for Disease Control, and the Bureau of Biologics of the Food and

Drug Administration. Summary of clinical trials of influenza vaccines–II. J Infect Dis 134: 633–638.

- Wright PF, Thompson J, Vaughn WK, Folland DS, Sell SH, et al. (1977) Trials of influenza A/New Jersey/76 virus vaccine in normal children: An overview of age-related antigenicity and reactogenicity. J Infect Dis 136 Suppl: S731–S741.
- Lin J, Zhang J, Dong X, Fang H, Chen J, et al. (2006) Safety and immunogenicity of an inactivated adjuvanted whole-virion influenza A (H5N1) vaccine: A phase I randomised controlled trial. Lancet 368: 991–997.
- Subbarao K, Chen H, Swayne D, Mingay L, Fodor E, et al. (2003) Evaluation of a genetically modified reassortant H5N1 influenza A virus vaccine candidate generated by plasmid-based reverse genetics. Virology 305: 192–200.
- Webby RJ, Perez DR, Coleman JS, Guan Y, Knight JH, et al. (2004) Responsiveness to a pandemic alert: Use of reverse genetics for rapid development of influenza vaccines. Lancet 363: 1099–1103.
- Lipatov AS, Webby RJ, Govorkova EA, Krauss S, Webster RG (2005) Efficacy of H5 influenza vaccines produced by reverse genetics in a lethal mouse model. J Infect Dis 191: 1216–1220.
- Nicolson C, Major D, Wood JM, Robertson JS (2005) Generation of influenza vaccine viruses on Vero cells by reverse genetics: An H5N1 candidate vaccine strain produced under a quality system. Vaccine 23: 2943–2952.
- Govorkova EA, Webby RJ, Humberd J, Seiler JP, Webster RG (2006) Immunization with reverse-genetics-produced H5N1 influenza vaccine protects ferrets against homologous and heterologous challenge. J Infect Dis 194: 159–167.
- Horimoto T, Takada A, Fujii K, Goto H, Hatta M, et al. (2006) The development and characterization of H5 influenza virus vaccines derived from a 2003 human isolate. Vaccine 24: 3669–3676.
- Lu X, Tumpey TM, Morken T, Zaki SR, Cox NJ, et al. (1999) A mouse model for the evaluation of pathogenesis and immunity to influenza A (H5N1) viruses isolated from humans. J Virol 73: 5903–5911.
- Takada A, Kuboki N, Okazaki K, Ninomiya A, Tanaka H, et al. (1999) Avirulent avian influenza virus as a vaccine strain against a potential human pandemic. J Virol 73: 8303–8307.
- Lipatov AS, Hoffmann F, Salomon R, Yen HL, Webster RG (2006) Crossprotectiveness and immunogenicity of influenza A/Duck/Singapore/3/ 97(H5) vaccines against infection with A/Vietnam/1203/04(H5N1) virus in ferrets. J Infect Dis 194: 1040–1043.
- Li S, Liu C, Klimov A, Subbarao K, Perdue ML, et al. (1999) Recombinant influenza A virus vaccines for the pathogenic human A/Hong Kong/97 (H5N1) viruses. J Infect Dis 179: 1132–1138.
- Lu X, Edwards LE, Desheva JA, Nguyen DC, Rekstin A, et al. (2006) Crossprotective immunity in mice induced by live-attenuated or inactivated vaccines against highly pathogenic influenza A (H5N1) viruses. Vaccine 24: 6588–6593.
- Desheva JA, Lu XH, Rekstin AR, Rudenko LG, Swayne DE, et al. (2006) Characterization of an influenza A H5N2 reassortant as a candidate for live-attenuated and inactivated vaccines against highly pathogenic H5N1 viruses with pandemic potential. Vaccine 24: 6859–6866.
- Suguitan AL Jr, McAuliffe J, Mills KL, Jin H, Duke G, et al. (2006) Live, attenuated influenza A H5N1 candidate vaccines provide broad crossprotection in mice and ferrets. PLoS Med 3: e360. doi:10.1371/journal. pmed.0030360
- Rimmelzwaan GF, Claas EC, van Amerongen G, de Jong JC, Osterhaus AD (1999) ISCOM vaccine induced protection against a lethal challenge with a human H5N1 influenza virus. Vaccine 17: 1355–1358.
- Gao W, Soloff AC, Lu X, Montecalvo A, Nguyen DC, et al. (2006) Protection of mice and poultry from lethal H5N1 avian influenza virus through adenovirus-based immunization. J Virol 80: 1959–1964.
- Hoelscher MA, Garg S, Bangari DS, Belser JA, Lu X, et al. (2006) Development of adenoviral-vector-based pandemic influenza vaccine against antigenically distinct human H5N1 strains in mice. Lancet 367: 475–481.
- 31. Qiao CL, Yu KZ, Jiang YP, Jia YQ, Tian GB, et al. (2003) Protection of chickens against highly lethal H5N1 and H7N1 avian influenza viruses with a recombinant fowlpox virus co-expressing H5 haemagglutinin and N1 neuraminidase genes. Avian Pathol 32: 25–32.
- Karaca K, Swayne DE, Grosenbaugh D, Bublot M, Robles A, et al. (2005) Immunogenicity of fowlpox virus expressing the avian influenza virus H5 gene (TROVAC AIV-H5) in cats. Clin Diagn Lab Immunol 12: 1340–1342.
- 33. Veits J, Wiesner D, Fuchs W, Hoffmann B, Granzow H, et al. (2006) Newcastle disease virus expressing H5 hemagglutinin gene protects chickens against Newcastle disease and avian influenza. Proc Natl Acad Sci U S A 103: 8197–8202.
- 34. Ge J, Deng G, Wen Z, Tian G, Wang Y, et al. (2007) Newcastle disease virusbased live attenuated vaccine completely protects chickens and mice from lethal challenge of homologous and heterologous H5N1 avian influenza viruses. J Virol 81: 150–158.
- Kodihalli S, Goto H, Kobasa DL, Krauss S, Kawaoka Y, et al. (1999) DNA vaccine encoding hemagglutinin provides protective immunity against H5N1 influenza virus infection in mice. J Virol 73: 2094–2098.
- Epstein SL, Tumpey TM, Misplon JA, Lo CY, Cooper LA, et al. (2002) DNA vaccine expressing conserved influenza virus proteins protective against H5N1 challenge infection in mice. Emerg Infect Dis 8: 796–801.