

REVIEW

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Haploid genomes illustrate epigenetic constraints and gene dosage effects in mammals

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Abstract

Sequencing projects have revealed the information of many animal genomes and thereby enabled the exploration of genome evolution. Insights into how genomes have been repeatedly modified provide a basis for understanding evolutionary innovation and the ever increasing complexity of animal developmental programs. Animal genomes are diploid in most cases, suggesting that redundant information in two copies of the genome increases evolutionary fitness. Genomes are well adapted to a diploid state. Changes of ploidy can be accommodated early in development but they rarely permit successful development into adulthood. In mammals, epigenetic mechanisms including imprinting and X inactivation restrict haploid development. These restrictions are relaxed in an early phase of development suggesting that dosage regulation appears less critical. Here we review the recent literature on haploid genomes and dosage effects and try to embed recent findings in an evolutionary perspective.

Keywords: Haploid cells, Stem cells, Epigenetics, Development, Tumor, Evolution

Review

Haploid genomes in insects and mites

The information for the development of an organism is encoded in its genomic DNA sequence. In most animals each cell contains two copies of the genome making up a diploid chromosome set. Diploid genomes provide a buffer against deleterious effects of mutations and enable the maintenance of suboptimal alleles that could become advantageous if environmental conditions change. Furthermore, diploidy allows co-transmission of beneficial and suboptimal alleles from the same parent facilitating maintenance of a diverse genetic basis for selection to draw from. Advantages of diploidy might explain the rare observation of single copy genomes outside germ line development of animal species. In animal development, haploid genomes are largely limited to post-meiotic germ cells that show little proliferation and gene expression according to their specialized function in reproduction. Haploid genomes do occur in some social insects including ants, wasps and honeybees, where they determine male sex [1]. Haploidy presumably serves to purge deleterious mutations from the genome of males. Males are

largely dispensable compared with females as they are only during a brief reproductive period in these species. A small number of fit males can provide a copy of the genome that is largely free of deleterious mutations for the next generation. In addition, rare parthenogenetic haploid species have been described in mites and insects [2,3]. Parthenogenetic all-female species appear to be rare exceptions but they still illustrate that haploid genomes can support development of quite remarkably complex organisms. Notably, haploid cell lines have also been isolated from flies that do not normally show haploid development [4] indicating that the ability to accommodate a change of ploidy is widely maintained in insect species. The scaling of molecular networks and pathways relative to genome copy number is surprising given the complexity of interactions involved in the animal developmental programs. Balancing of genome copy number elevations could be related to evolutionary selection for robustness of regulatory networks, but this has not been investigated to date.

Limited haploid development in vertebrates

It is not hard to imagine that ploidy elevation can lead to problems for organismal development through different nuclear-cytoplasmic ratio or non-scaling gene dosage relations. However, evidence suggests that changes in genome copy number can be compatible with development in a

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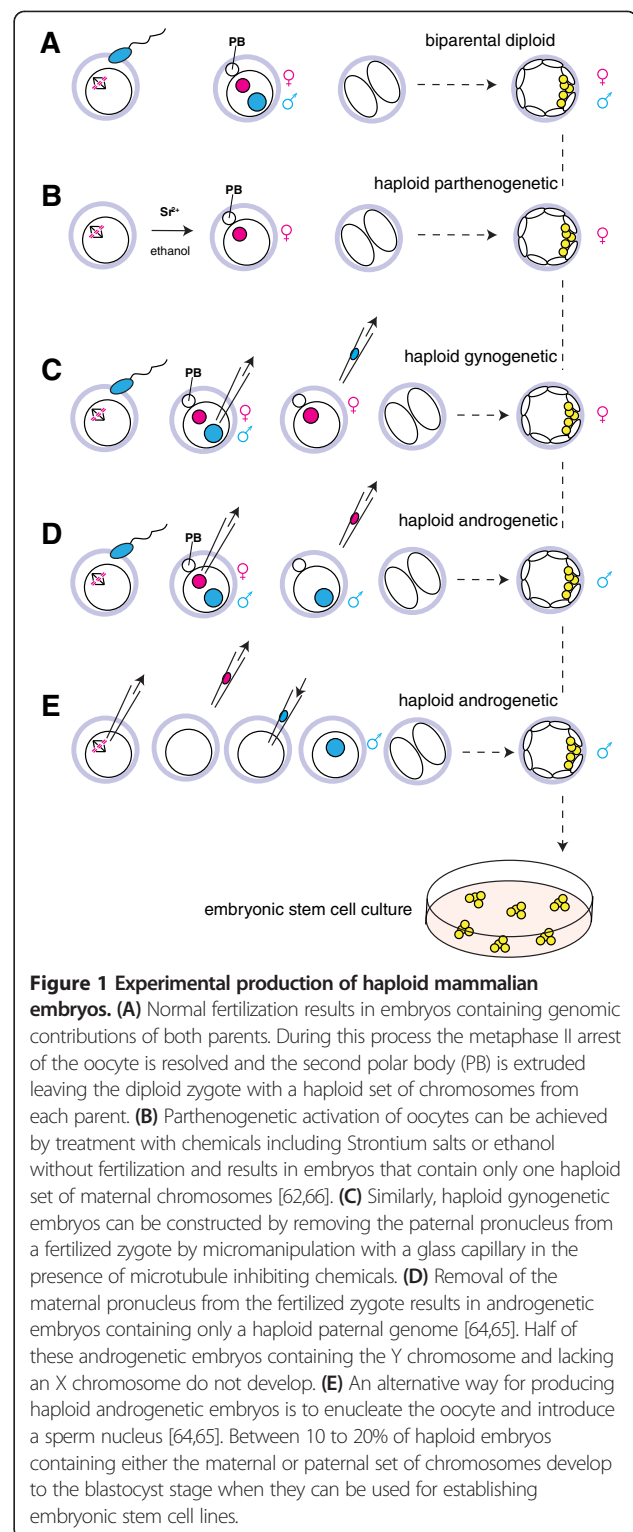
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range of organisms. Polyploid frogs and lizards can coexist with related diploid populations, and also interbreed in some cases [5-7]. A number of tetraploid amphibians and reptile species have been described [8]. Triploid vertebrates can arise through hybridization of diploid and tetraploid species or from nondisjunction of chromosomes in the egg after fertilization as is commercially applied in rainbow trout [9]. Even sexually reproductive triploid vertebrate species have been observed [7]. Notably, it has been possible to recreate ploidy elevation in the laboratory through fertilization of triploid eggs of parthenogenetic asexual vertebrates [6]. This suggests that little obstacles to ploidy elevation exist in vertebrates. Indeed there is evidence that two rounds of genome-wide duplications have occurred during vertebrate evolution [10,11], indicating that current vertebrate genomes are a relic from a polyploid stage [12]. Notably, haploid cell lines from frogs have been reported [13] showing that amphibians can accommodate both genome copy number elevation as well as reduction.

Haploid development in zebrafish can be experimentally induced by fertilization with inactivated sperm [14] or by fertilization of irradiated oocytes [15,16]. Haploid gynogenetic or androgenetic zebrafish embryos progress through embryonic development but do not reach the mature stage. This shows that in fish a haploid genome can direct embryonic growth and organogenesis but is incompatible with full adult development. Interestingly, haploid pluripotential embryonic cells from Medaka have been established [17]. These cells maintain an intact haploid karyotype in culture and can contribute to development through semicloning. Teleost fish have experienced a recent third genome duplication event and it is conceivable that haploid development could benefit from the approximation of an ancestral genome state before duplication [18,19]. These observations illustrate that developmental programs in fish, amphibian and reptile species can accommodate ploidy changes to variable degrees. It is conceivable that tolerance to ploidy changes is related to genome duplication events as a driver of evolutionary innovations in these branches [12]. Potentially more recent and complex developmental programs in higher vertebrates might introduce features that encounter greater problems with changes in ploidy.

Imprinting and X chromosome dosage restrict haploid development in mammals

In mammals, haploid development can be induced by activation of unfertilized oocytes to produce parthenogenetic haploid embryos [20-22] or by fertilization of enucleated oocytes to produce androgenetic haploid embryos [23] (Figure 1). Haploid mouse preimplantation embryos have also been obtained by mechanical bisection of zygotes [24] or by microsurgical removal of

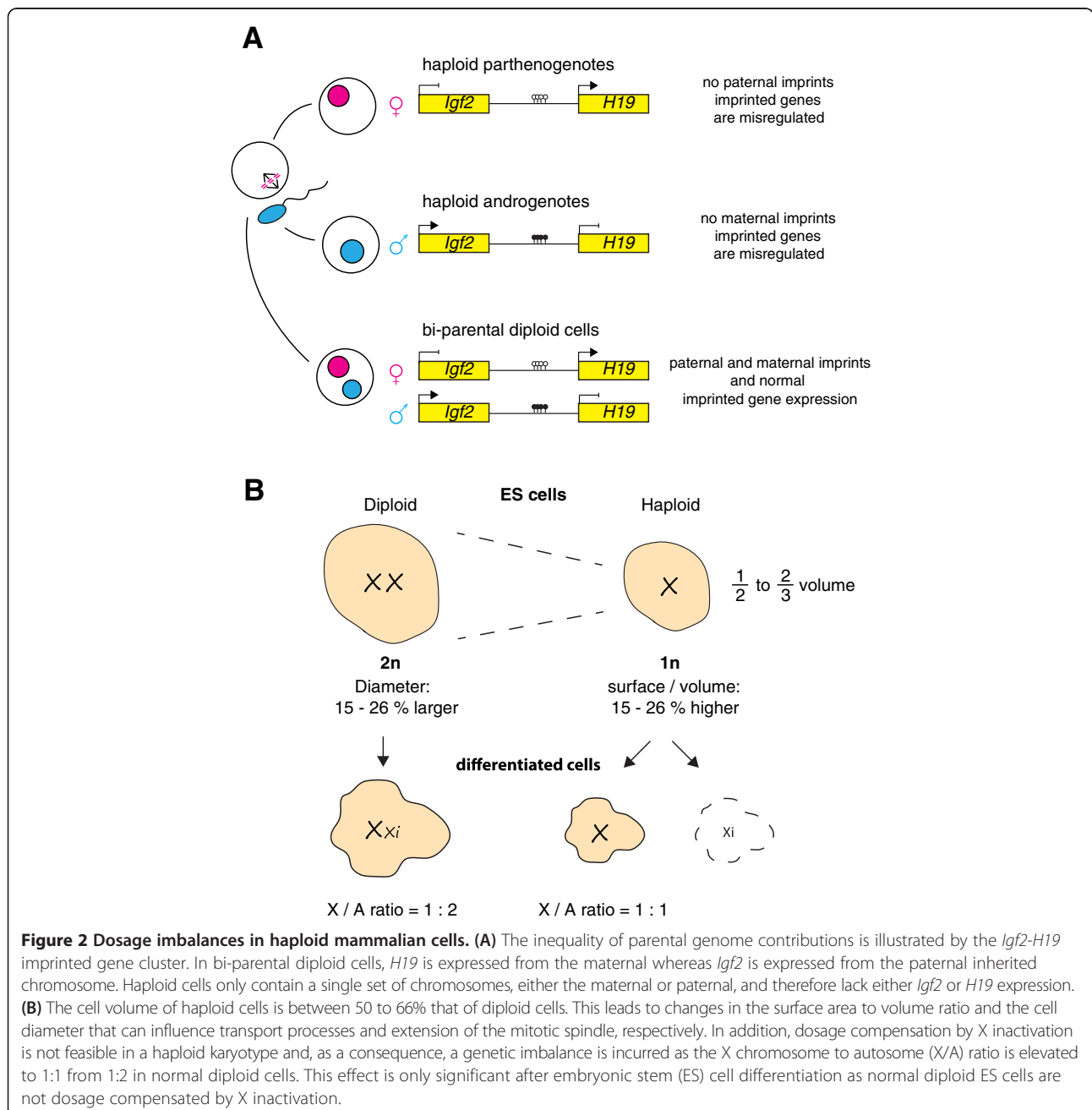


one pronucleus [25]. In mice, haploid cells have been observed until egg cylinder stage embryos [26], but haploid development beyond implantation is severely impaired. This is a direct consequence of the fact that the two parental contributions to the genome are not

equivalent in mammals (Figure 2A). Genomic imprinting restricts expression of certain genes to one parental allele [27-29]. As a consequence, both maternal and paternal chromosomes are required for successful development in mice [30-32]. Genomic imprinting can affect evolution though selective exposure of mutations in a functionally hemizygous state and has further been suggested to aid a greater variability in quantitative traits that could benefit species in changing environments [33]. Fixation of unbalanced parental contributions is hypothesized to be driven

by genetic effects including conflict over parental investment between both sexes [34,35].

Other examples for monoallelic expression in mammals include allelic exclusion of immunoglobulin loci [36], T-cell receptor genes and olfactory receptor genes. In addition, the majority of X-linked genes are expressed monoallelically. Compensation for X-linked gene dosage is required as a consequence of the mammalian XY sex chromosome system. In both males (XY) and females (XX), only a single X chromosome is transcriptionally



active [37]. This is achieved by transcriptional inactivation of one of the two X chromosomes in females through the process of X inactivation. The requirement of a single active X chromosome per diploid set of autosomes results in an X chromosome to autosome ratio of 1:2 that cannot be approximated within a haploid genome and causes immitigable dosage effects for haploid development in mammals (Figure 2B). Gene activity from the single X chromosome causes a two-fold relative increase in X-linked gene dosage. Alternatively, inactivation of the X chromosome leaves haploid cells nullisomic for X-linked genes, which is not compatible with survival [38]. Whereas early mouse embryos can tolerate a lack of dosage compensation, X inactivation becomes essential soon after implantation [39]. Genomic imprinting, monoallelic expression and X chromosome dosage impose genetic limits to haploid development in mammals.

Haploid phases in human tumors

It is a fact - despite rarely being consciously considered - that a diploid karyotype represents an exception rather than the rule in established cell cultures. Many permanent cell lines acquire aneuploidies in culture with gain and loss of chromosomes providing growth advantages possibly in combination with acquired mutations. Culture conditions might contribute significantly to the development of aneuploidies as growth requirements are less stringent than in development where growth depends on functioning tissues and organs. This is also true for mouse embryonic stem (ES) cells where aneuploidies accumulate with an increase in passage number [40]. Notably, aneuploidies are also observed in rare occasions of transmissible tumors in canines and Tasmanian devils suggesting that unusual and unexpected properties can result from karyotype changes [41,42]. Elevated levels of aneuploidy are also common in human tumors. These observations suggest that a diploid chromosome set is not essential for cell survival and deviations from a regular diploid genome might be advantageous in culture and tumors.

Aneuploidy in most tumors manifests itself in a shift of the modal average of chromosomes. Interestingly, hypodiploid, including rare near haploid tumor karyotypes, have been reported. Near haploid tumor cells have been observed in rare cases of leukemia [43-49], and have been less frequently reported in solid tumors [50-52]. Loss of chromosomes appears to be the primary event in near haploid acute lymphoid leukemia and correlates with poor prognosis [44,53]. Haploid karyotypes in tumors are not fully intact and often contain diploid genomic regions and chromosomal rearrangements [54]. This suggests a selective advantage of the haploid state, probably in the context of oncogenic mutations and rearrangements. A haploid phase where a single hit can

inactivate gene function could be explained by selection for loss of tumor suppressor genes during tumor development (Figure 3). However, it is unlikely that haploidy is a requirement for loss of tumor suppressor activity since this could also be achieved by selective loss of few chromosomes and maintenance of a largely diploid genome. Therefore, it cannot be ruled out that a haploid phase might contribute to tumor cell persistence in a different way, possibly involving gene dosage effects.

Establishment of haploid mammalian cell lines

Cells with near haploid and hypodiploid karyotypes have been adapted to growth in culture from a partially haploid chronic myeloid leukemia [43]. Apparently, these cultures were obtained at the blast phase after a long benign phase and repeated chemotherapeutic treatment suggesting significant selection of tumor cells before cultures were established. The haploid portion of the KBM7 cell line carries two copies of chromosomes 8 and 15 in addition to a BCR-ABL chromosomal translocation. Initially, the KBM7 cell line showed strong inclination to diploidization such that later passages had lost the haploid fraction of cells. However, a subclone (P1-55) from early passage KBM7 cells maintained a near haploid karyotype diploid only for chromosome 8 stably in culture

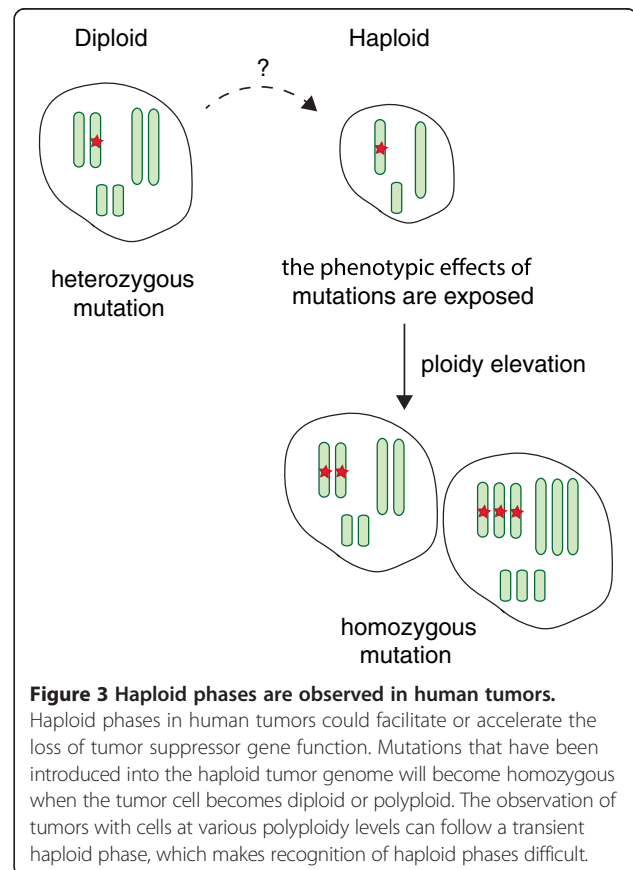


Figure 3 Haploid phases are observed in human tumors.

Haploid phases in human tumors could facilitate or accelerate the loss of tumor suppressor gene function. Mutations that have been introduced into the haploid tumor genome will become homozygous when the tumor cell becomes diploid or polyploid. The observation of tumors with cells at various polyploidy levels can follow a transient haploid phase, which makes recognition of haploid phases difficult.

[47]. Notably, the reduced rate of diploidization indicates a second and independent adaptation that has occurred after culture. Later work has attempted to change the cell type of the haploid cells for expansion of their use in genetic screening [55]. Introduction of viral vectors used for reprogramming of induced pluripotent stem cells resulted in an adherent cell line that had lost its hematopoietic character. Although pluripotency was not established, these HAP1 cells are of interest as they possess different growth properties including altered morphology and differential response to cell toxins [55]. This cell line also no longer contains a second copy of chromosome 8 suggesting a haploid karyotype, albeit with chromosomal translocations. These changes have also led to an increased rate of diploidization. These findings clearly illustrate that mammalian cells with a near haploid karyotype can proliferate and display distinct phenotypes in culture.

Pluripotent haploid cells from early mouse embryos

Following studies on haploid mammalian embryos, initial attempts to derive pluripotent ES cells from haploid mouse blastocysts resulted in the establishment of diploid cell lines [56]. This was surprising as both parthenogenetic and androgenetic diploid embryos can develop past the blastocyst state and survive beyond implantation [57,58]. Parthenogenetic embryos are lost around embryonic day 10 (E10) [58,59]. Similarly, embryos with impaired dosage compensation due to a mutation in the *Xist* gene develop beyond implantation [39,60]. These findings indicate that pre-implantation development is largely independent of dosage compensation and the presence of a bi-parental complement of imprints. However, pre-implantation development in parthenogenotes does not progress completely independent of X inactivation and delayed upregulation of *Xist* from one of the two maternal X chromosomes has been reported at the eight cell stage [61]. Recent improvements in ES cell culture techniques and innovation in flow cytometric cell sorting technology have finally facilitated the establishment of haploid parthenogenetic [62,63] and androgenetic [64,65] ES cell lines from mouse embryos (Figure 1). Haploid mouse ES cells proliferate in culture and maintain an intact haploid karyotype for more than 30 passages as evidenced by genomic analysis and developmental competence [66].

The developmental stage from which mouse ES cells are derived appears to tolerate the loss of epigenetic regulation [67]. It has been reported that abrogation of DNA methylation [68], Polycomb complex function [69] and nuclear B type lamins [70] does not prevent proliferation and self-renewal of mouse ES cells. In contrast, respective mutations lead to defects in differentiated cells. ES cells are derived from cells of the inner cell

mass of the blastocyst that will develop into the epiblast. At these stages epigenetic patterns are reset and epigenetic regulation appears substantially different. For example, the cells of the early epiblast are not dosage compensated before X inactivation is initiated around the time of gastrulation in mice. The discovery of new culture conditions has facilitated the culture of ES cells in a naïve pluripotent ground state by inhibition of the mitogen activated protein (MAP) kinase and glycogen synthase kinase pathways [71]. These two inhibitor (2i) conditions are beneficial for obtaining ES cell lines with a high content of haploid cells [63]. Haploid ES cells have also been established or cultured in traditional serum containing media and Leukemia inhibitory factor (LIF), but with substantially reduced efficiency and increased rate of diploidization [62,66]. The question arises how 2i culture conditions contribute to the maintenance of a haploid karyotype. In serum-based culture conditions, ES cells are heterogeneous and at any given point in time only a fraction of cells express naïve pluripotency markers including *Nanog* and *Rex1*. In contrast, these markers are homogeneously expressed in all cells in 2i conditions [71-73]. Therefore, it is conceivable that, in the naïve ground state, selective pressure arising from gene dosage effects of a haploid genome are largely alleviated. Notably, culture in 2i medium also induces drastic changes in the epigenetic profiles of ES cells. It has been shown that Polycomb-associated histone H3 tri-methylation patterns are shifted in 2i conditions with reduced levels on promoters and greater enrichment over satellite repeats [74]. Furthermore, DNA methylation is substantially reduced in 2i medium compared to serum-based ES cell cultures [75]. This finding is consistent with low levels of DNA methylation in inner cell mass cells. Irrespective of reduced epigenetic modifications, genomic imprints are maintained in 2i medium [75]. Paternal imprints are further partially maintained in androgenetic haploid ES cells but are progressively lost with time in culture [64,65]. Loss of imprinting is not special to haploid ES cells but can also be associated with diploid ES cell cultures [76]. Haploid ES cells are competent to contribute to a wide range of tissues in chimeras [62-65]. However, contribution to development is only possible after diploidization *in vitro* or *in vivo*. Colonization of the female germ line and transmission of a transgene was observed for parthenogenetic haploid ES cells [66]. So far, the germ line competence of androgenetic haploid embryonic stem cells has been limited to early stages of primordial germ cells [64,65]. However, they have been used for semi-cloning and appear to contain functionally relevant paternal imprints. Live mice have been obtained from injection of haploid androgenetic ES cells into unfertilized oocytes indicating that they can substitute sperm cells [64,65]. Haploid ES cells

appear to have an intrinsic tendency for diploidization. The trigger for diploidization is not known but appears crucial for the establishment of stable differentiated cell types from haploid ES cells. Haploid ES cells proliferate with similar kinetics as diploid ES cells. Hence the need to replicate a genome that is half the normal size does not appear to increase proliferative potential. This observation is further in line with the observation that tetraploid ES cells show a similar rate of self-renewal as diploid ES cells [77]. This indicates that the replication of the genome is not the rate limiting step in cell division in ES cells. Alternatively, altered properties might compensate for differences in genome size. Interestingly, ploidy correlates with cell size. Haploid ES cells possess a volume that is approximately two-thirds of that of diploid cells (unpublished observation; Figure 2B). Therefore, a reduction in genome size leads to a concomitant reduction in the availability of resources due to a smaller cell volume.

Application of haploid cells in genetic screens

Diploid karyotypes of virtually all mammalian species have severely limited forward genetic approaches. The discovery of haploid mammalian cells has opened new possibilities for performing genetic screens in mammals (Figure 4). The use of haploid cells in screens was initially demonstrated using a near haploid KBM7-derived human tumor cell line [78]. For this a mutant library was generated by insertion of viral gene trap vectors in large pools of haploid cells. This library was subsequently exposed to various toxins and pathogens. Surviving cell clones were analyzed and mutations conferring resistance could be identified. Since then an impressive number of screens investigated host mechanisms utilized by pathogens [55,78-82]. In addition to understanding disease mechanisms, recent work has also provided insights into human cellular pathways [81]. Haploid ES cells could provide advantages through a largely intact genome that is free of tumor-specific mutations. This is especially important for dissection of developmental processes in forward genetic screens. The potential for using haploid ES cells in screens has been shown by proof-of-principle experiments identifying factors in the DNA mismatch repair pathway and mediating ricin toxicity [62,63]. Inherent tendency to diploidization is not an obstacle for screening as long as the mutations are inserted in a haploid state. Recently, haploid ES cells have also been used to investigate the resistance mechanism for the chemotherapeutic agent Olaparib [83]. It might be expected that future screens can utilize the pluripotent potential of haploid ES cells and the availability of reporter mouse lines for investigating molecular networks of gene regulation, cell signaling and development. This prospect suggests that haploid ES cells could become a tool for

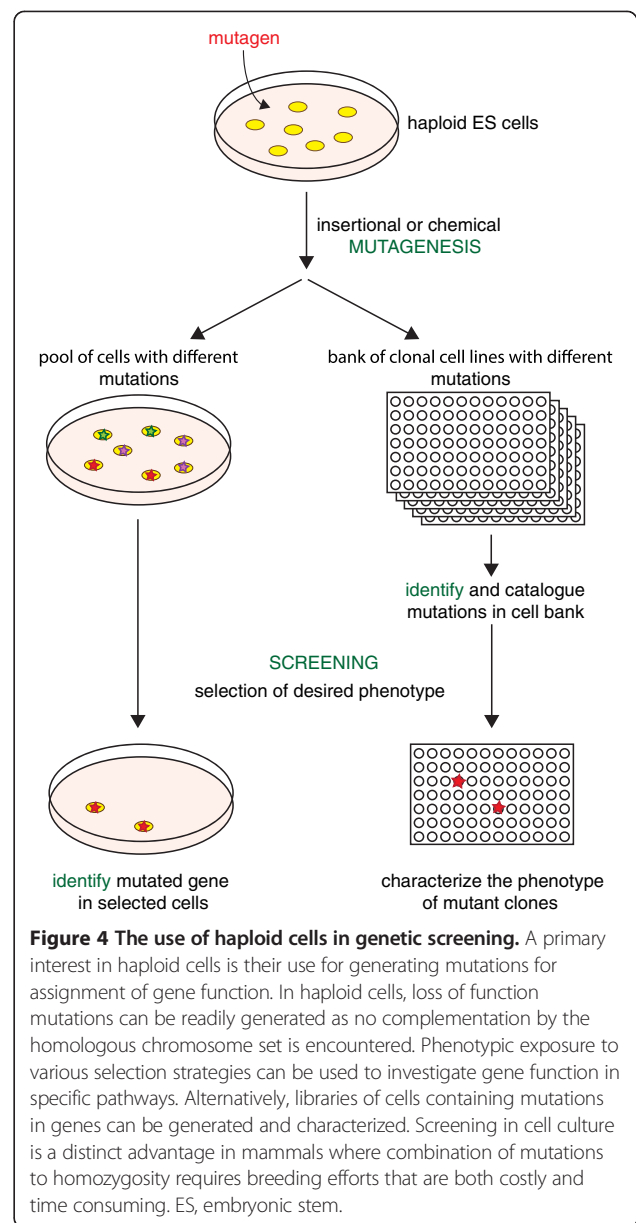


Figure 4 The use of haploid cells in genetic screening. A primary interest in haploid cells is their use for generating mutations for assignment of gene function. In haploid cells, loss of function mutations can be readily generated as no complementation by the homologous chromosome set is encountered. Phenotypic exposure to various selection strategies can be used to investigate gene function in specific pathways. Alternatively, libraries of cells containing mutations in genes can be generated and characterized. Screening in cell culture is a distinct advantage in mammals where combination of mutations to homozygosity requires breeding efforts that are both costly and time consuming. ES, embryonic stem.

performing developmental screens in culture similar to screens in haploid zebrafish [84].

Conclusions

The ability to derive haploid ES cells might be facilitated by a distinct developmental state. Epigenetic mechanisms are largely dispensable in preimplantation mouse embryos. This likely reflects a period of resetting the genome to attain pluripotency. In addition, cell size in preimplantation embryos changes in a remarkable fashion through successive cleavage divisions of the oocyte during which overall embryo growth is negligible. This leads to progressively smaller cell sizes. It is conceivable that regulatory networks have been adapted to cope with changing

cell sizes and, thus, are robust against dosage effects. Indeed, ES cells appear to tolerate considerable changes in gene expression profiles. Gene expression profiles in Polycomb-deficient ES cells are substantially changed but do not abrogate self-renewal [69]. Notably, a recent comparison of serum and 2i culture has also identified surprisingly large differences in gene expression [74]. These observations suggest that regulatory networks in ES cells are robust to disturbances in gene expression patterns. This robustness could contribute to the scaling of pathways with different levels of ploidy.

The extent to which differentiated cell types can be maintained with a haploid karyotype remains unknown. Induction of haploid ES cells to differentiation conditions inexorably leads to rapid diploidization. An indication that haploid karyotypes are compatible at least with early developmental cell fates comes from reports showing that haploid epiblast stem cells and primitive endoderm-like cells have been established from haploid ES cells in culture [64,66]. These reports are consistent with the observation that haploid cells can contribute to E6.5 post implantation embryos before diploidization [63-65] and have been observed in egg cylinder stage embryos [26].

Development of haploid embryos is affected by requirements for imprinted gene expression and dosage compensation. Haploid ES cells can contribute to the development of chimeric embryos after diploidization but are unable to support ES cell derived mice in a tetraploid complementation assay [66]. Imprinting defects are illustrated by the inability of diploid parthenogenotes to progress through development beyond E10 [59]. Interestingly, it has been possible to generate bimaternal embryos that can develop normally from fully grown oocytes and non-growing oocytes that contain double deletions in the H19 differentially methylated region and the Dlk1-Dio3 intergenic germ line-derived imprinting control region [85]. It is interesting to consider if similar manipulations could improve the stability and differentiation potential of parthenogenetic haploid cells. The imprints that inhibit androgenote growth are yet to be determined.

The dosage compensation problem is more difficult to resolve as a half dose of X chromosome linked genes would be required in the case of a single set of autosomes (Figure 2A). The relative expression balance for X-linked and autosomal genes is assumed to be maintained in evolution through upregulation of the active X chromosome relative to autosomes following Y chromosome erosion and a switch to a single active X chromosome [86]. The mechanism of X upregulation is presently not well understood. Recent results suggest that the Males absent on the first (MOF) histone acetyltransferase contributes to the upregulation of a subset of X-linked genes [87]. Interference with the mechanism of X upregulation could potentially be considered for reducing the X-linked

gene dosage in haploid cells. Not all X-linked genes appear to be upregulated and subject to dosage compensation [88]. Expression reduction, possibly by RNAi-mediated strategies, could therefore also be considered. Genes whose products contribute to multi-subunit complexes appear most critical, as loss of stoichiometry can topple the balance of fine-tuned regulatory networks and protein-complex formation [89-91]. Restoration of X dosage, and hence stoichiometry, could be an effective means for enhancing haploid cell stability and developmental performance.

The observation of haploid phases in human tumors suggests that certain oncogenic signals can stabilize a haploid karyotype. Notably, overexpression of X linked genes has been implicated as a driver of tumorigenesis [92,93]. Future work will be needed to establish a connection between oncogenic transformation and changes in ploidy. This could yield important insights into dosage sensitive pathways in mammals and also be relevant for understanding certain human tumors. Dosage balance is less critical in differentiated cells and aneuploidies are tolerated in tumors and cell cultures to some extent. Dosage regulation could be critical in a developmental window but be less stringent in preimplantation development and at the end of the developmental program. An interesting question is if haploid cells can be generated directly from somatic diploid cells. Loss of chromosomes has been experimentally induced by interfering with centromere function [94]. Loss of chromosomes often appears to lead to aneuploidies that are not compatible with cell survival and proliferation. It appears that, in contrast to tumor cells, relative gene dosage imbalances are more detrimental to survival of untransformed cells than haploidy. This suggests that reduction of a diploid to a near haploid karyotype in a single instance or rapid succession of manipulations would be required. It is hard to imagine how this could be achieved with current technology. Induction of meiosis could in principle be considered as an alternative strategy. However, meiosis is an elaborate process that requires pairing of homologous chromosomes which in animals has not been observed outside the germ line. Recent advances in culture systems suggest that the generation of germ cells might become feasible. Protocols for deriving oocytes [95,96] and sperm [97] from ES cells have been reported. These methods could be useful for establishing haploid cells from ES cells or germ line precursor cells. Lastly, the still elusive mechanism that cancer cells use to reduce the genome by half might be applied for experimental induction of haploidy in cell cultures. Undoubtedly, future research will contribute to methods for establishing haploid cells and rebalancing gene dosage that could finally lead to an increased developmental potential.

Independently, haploid ES cells might provide a tool for studying allelic differences in genomic imprinting. The ability to establish haploid androgenotes and parthenogenotes will allow the maintenance of the two parental genome contributions in separate cell cultures and facilitate the functional investigation of parental marks. Although the haploid cell state is, with the exception of gametes, either artificial or associated with malignancies, it holds the promise of teaching us about genomic balance and dosage effects. Haploid embryonic cells will have important implications for understanding gene regulatory networks and genome evolution and will provide a powerful genetic screening platform.

Abbreviations

2i: Two inhibitor; E10: Embryonic day 10; ES: Embryonic stem.

Competing interests

The authors declare that they have no conflict of interest in writing this review, but wish to make it known that a patent application covering haploid embryonic stem cells has been filed (patent publication number WO2012117254 A1).

Authors' contributions

ML has researched the topic and written the text of the review. AW has written the text and advised on the content and structure of the review. Both authors read and approved the final manuscript.

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References

1. Normark BB: The evolution of alternative genetic systems in insects. *Annu Rev Entomol* 2003, **48**:397–423.
2. Weeks AR, Marec F, Breeuwer JA: A mite species that consists entirely of haploid females. *Science* 2001, **292**:2479–2482.
3. Beukeboom LW, Kamping A, Louter M, Pijnacker LP, Katju V, Ferree PM, Werren JH: Haploid females in the parasitic wasp *Nasonia vitripennis*. *Science* 2007, **315**:206.
4. Debec A: Evolution of karyotype in haploid cell lines of *Drosophila melanogaster*. *Exp Cell Res* 1984, **151**:236–246.
5. Christiansen DG, Jakob C, Arioli M, Roethlisberger S, Reyer HU: Coexistence of diploid and triploid hybrid water frogs: population differences persist in the apparent absence of differential survival. *BMC Ecol* 2010, **10**:14.
6. Lutes AA, Baumann DP, Neaves WB, Baumann P: Laboratory synthesis of an independently reproducing vertebrate species. *Proc Natl Acad Sci U S A* 2011, **108**:9910–9915.
7. Stock M, Lamatsch DK, Steinlein C, Epplen JT, Grosse WR, Hock R, Klapperstuck T, Lampert KP, Scheer U, Schmid M, Scharl M: A bisexually reproducing all-triploid vertebrate. *Nat Genet* 2002, **30**:325–328.
8. Comai L: The advantages and disadvantages of being polyploid. *Nat Rev Genet* 2005, **6**:836–846.
9. Thorgaard GH, Gall GA: Adult triploids in a rainbow trout family. *Genetics* 1979, **93**:961–973.
10. Ohno S: Gene duplication and the uniqueness of vertebrate genomes circa 1970–1999. *Semin Cell Dev Biol* 1999, **10**:517–522.
11. Nakatani Y, Takeda H, Kohara Y, Morishita S: Reconstruction of the vertebrate ancestral genome reveals dynamic genome reorganization in early vertebrates. *Genome Res* 2007, **17**:1254–1265.
12. Freeling M, Thomas BC: Gene-balanced duplications, like tetraploidy, provide predictable drive to increase morphological complexity. *Genome Res* 2006, **16**:805–814.
13. Freed JJ, Mezger-Freed L: Stable haploid cultured cell lines from frog embryos. *Proc Natl Acad Sci U S A* 1970, **65**:337–344.
14. Postlethwait JH, Johnson SL, Midson CN, Talbot WS, Gates M, Ballinger EW, Africa D, Andrews R, Carl T, Eisen JS, et al: A genetic linkage map for the zebrafish. *Science* 1994, **264**:699–703.
15. Brandhorst BP, Corley-Smith GE: Production of haploid and diploid androgenetic zebrafish. *Methods Mol Biol* 2004, **254**:255–270.
16. Corley-Smith GE, Lim CJ, Brandhorst BP: Production of androgenetic zebrafish (*Danio rerio*). *Genetics* 1996, **142**:1265–1276.
17. Yi M, Hong N, Hong Y: Generation of medaka fish haploid embryonic stem cells. *Science* 2009, **326**:430–433.
18. Jaillon O, Aury JM, Brunet F, Petit JL, Stange-Thomann N, Mauceli E, Bouneau L, Fischer C, Ozouf-Costaz C, Bernot A, Nicaud S, Jaffe D, Fisher S, Lutfalla G, Dossat C, Segurens B, Dasilva C, Salanoubat M, Levy M, Boudet N, Castellano S, Anthouard V, Jubin C, Castelli V, Katinka M, Vacherie B, Biemont C, Skalli Z, Cattolico L, Poulain J, et al: Genome duplication in the teleost fish *Tetraodon nigroviridis* reveals the early vertebrate proto-karyotype. *Nature* 2004, **431**:946–957.
19. Meyer A, Van de Peer Y: From 2R to 3R: evidence for a fish-specific genome duplication (FSGD). *Bioessays* 2005, **27**:937–945.
20. Graham CF: Parthenogenetic mouse blastocysts. *Nature* 1970, **226**:165–167.
21. Tarkowski AK, Witkowska A, Nowicka J: Experimental parthenogenesis in the mouse. *Nature* 1970, **226**:162–165.
22. Braden AW, Austin CR: Reactions of unfertilized mouse eggs to some experimental stimuli. *Exp Cell Res* 1954, **7**:277–280.
23. Kono T, Sotomaru Y, Sato Y, Nakahara T: Development of androgenetic mouse embryos produced by in vitro fertilization of enucleated oocytes. *Mol Reprod Dev* 1993, **34**:43–46.
24. Tarkowski AK, Rossant J: Haploid mouse blastocysts developed from bisected zygotes. *Nature* 1976, **259**:663–665.
25. Modlinski JA: Haploid mouse embryos obtained by microsurgical removal of one pronucleus. *J Embryol Exp Morphol* 1975, **33**:897–905.
26. Kaufman MH: Chromosome analysis of early postimplantation presumptive haploid parthenogenetic mouse embryos. *J Embryol Exp Morphol* 1978, **45**:85–91.
27. DeChiara TM, Robertson EJ, Efstratiadis A: Parental imprinting of the mouse insulin-like growth factor II gene. *Cell* 1991, **64**:849–859.
28. Bartolomei MS, Zemel S, Tilghman SM: Parental imprinting of the mouse H19 gene. *Nature* 1991, **351**:153–155.
29. Barlow DP, Stoger R, Herrmann BG, Saito K, Schweifer N: The mouse insulin-like growth factor type-2 receptor is imprinted and closely linked to the *Tme* locus. *Nature* 1991, **349**:84–87.
30. Solter D: Differential imprinting and expression of maternal and paternal genomes. *Annu Rev Genet* 1988, **22**:127–146.
31. McGrath J, Solter D: Completion of mouse embryogenesis requires both the maternal and paternal genomes. *Cell* 1984, **37**:179–183.
32. Cattanaach BM, Kirk M: Differential activity of maternally and paternally derived chromosome regions in mice. *Nature* 1985, **315**:496–498.
33. Beaudet AL, Jiang YH: A rheostat model for a rapid and reversible form of imprinting-dependent evolution. *Am J Hum Genet* 2002, **70**:1389–1397.
34. Moore T, Haig D: Genomic imprinting in mammalian development: a parental tug-of-war. *Trends Genet* 1991, **7**:45–49.
35. Hurst LD, McVean GT: Growth effects of uniparental disomies and the conflict theory of genomic imprinting. *Trends Genet* 1997, **13**:436–443.
36. Chess A: Mechanisms and consequences of widespread random monoallelic expression. *Nat Rev Genet* 2012, **13**:421–428.
37. Schulz EG, Heard E: Role and control of X chromosome dosage in mammalian development. *Curr Opin Genet Dev* 2013, **23**:109–115.
38. Wutz A, Rasmussen TP, Jaenisch R: Chromosomal silencing and localization are mediated by different domains of *Xist* RNA. *Nat Genet* 2002, **30**:167–174.
39. Marahrens Y, Panning B, Dausman J, Strauss W, Jaenisch R: *Xist*-deficient mice are defective in dosage compensation but not spermatogenesis. *Genes Dev* 1997, **11**:156–166.

40. Longo L, Bygrave A, Grosveld FG, Pandolfi PP: **The chromosome make-up of mouse embryonic stem cells is predictive of somatic and germ cell chimaerism.** *Transgenic Res* 1997, **6**:321–328.
41. Pearce AM, Swift K: **Allograft theory: transmission of devil facial-tumour disease.** *Nature* 2006, **439**:549.
42. Weber WT, Nowell PC, Hare WC: **Chromosome studies of a transplanted and a primary canine venereal sarcoma.** *J Natl Cancer Inst* 1965, **35**:537–547.
43. Andersson BS, Beran M, Pathak S, Goodacre A, Barlogie B, McCredie KB: **Ph-positive chronic myeloid leukemia with near-haploid conversion in vivo and establishment of a continuously growing cell line with similar cytogenetic pattern.** *Cancer Genet Cytogenet* 1987, **24**:335–343.
44. Brodeur GM, Williams DL, Look AT, Bowman WP, Kalwinsky DK: **Near-haploid acute lymphoblastic leukemia: a unique subgroup with a poor prognosis?** *Blood* 1981, **58**:14–19.
45. Gancberg D, Dargent JL, Verhest A, Kentos A, Feremans W, Lohrisch C, Kornreich A: **Near haploid blast phase in a chronic myeloid leukemia detected by fluorescence in situ hybridization using a BCR-ABL probe.** *Cancer Genet Cytogenet* 2001, **128**:172–174.
46. Gibbons B, MacCallum P, Watts E, Rohatiner AZ, Webb D, Katz FE, Secker-Walker LM, Temperley IJ, Harrison CJ, Campbell RH, et al: **Near haploid acute lymphoblastic leukemia: seven new cases and a review of the literature.** *Leukemia* 1991, **5**:738–743.
47. Kotecki M, Reddy PS, Cochran BH: **Isolation and characterization of a near-haploid human cell line.** *Exp Cell Res* 1999, **252**:273–280.
48. Stark B, Jeison M, Gobuzov R, Krug H, Glaser-Gabay L, Luria D, El-Hasid R, Harush MB, Avrahami G, Fisher S, Stein J, Zaizov R, Yaniv I: **Near haploid childhood acute lymphoblastic leukemia masked by hyperdiploid line: detection by fluorescence in situ hybridization.** *Cancer Genet Cytogenet* 2001, **128**:108–113.
49. Sunil SK, Prakash PN, Hariharan S, Vinod G, Preethi RT, Geetha N, Ankathil R: **Adult acute lymphoblastic leukemia with near haploidy, hyperdiploidy and Ph positive lines: a rare entity with poor prognosis.** *Leuk Lymphoma* 2006, **47**:561–563.
50. Asperg F, Mertens F, Bauer HC, Lindholm J, Mitelman F, Mandahl N: **Near-haploidy in two malignant fibrous histiocytomas.** *Cancer Genet Cytogenet* 1995, **79**:119–122.
51. Orndal C, Mandahl N, Carlen B, Willen H, Wennerberg J, Heim S, Mitelman F: **Near-haploid clones in a malignant fibrous histiocytoma.** *Cancer Genet Cytogenet* 1992, **60**:147–151.
52. Sukov WR, Ketterling RP, Wei S, Monaghan K, Blunden P, Mazzara P, Raghavan R, Oliviera AM, Wiktor AE, Keeney GL, Van Dyke DL: **Nearly identical near-haploid karyotype in a peritoneal mesothelioma and a retroperitoneal malignant peripheral nerve sheath tumor.** *Cancer Genet Cytogenet* 2010, **202**:123–128.
53. Safavi S, Forestier E, Golovleva I, Barbany G, Nord KH, Moorman AV, Harrison CJ, Johansson B, Paulsson K: **Loss of chromosomes is the primary event in near-haploid and low-hypodiploid acute lymphoblastic leukemia.** *Leukemia* 2013, **27**:248–250.
54. Corver WE, Ruano D, Weijers K, den Hartog WC, van Nieuwenhuizen MP, de Miranda N, van Eijk R, Middeldorp A, Jordanova ES, Oosting J, Kapiteijn E, Hovens G, Smit J, van Wezel T, Morreau H: **Genome haploidisation with chromosome 7 retention in oncocytic follicular thyroid carcinoma.** *PLoS One* 2012, **7**:e38287.
55. Carette JE, Raaben M, Wong AC, Herbert AS, Obernosterer G, Mulherkar N, Kuehne AI, Kranzusch PJ, Griffin AM, Ruthel G, Dal Cin P, Dye JM, Whelan SP, Chandran K, Brummelkamp TR: **Ebola virus entry requires the cholesterol transporter Niemann-Pick C1.** *Nature* 2011, **477**:340–343.
56. Kaufman MH, Robertson EJ, Handside AH, Evans MJ: **Establishment of pluripotential cell lines from haploid mouse embryos.** *J Embryol Exp Morphol* 1983, **73**:249–261.
57. Kaufman MH, Barton SC, Surani MA: **Normal postimplantation development of mouse parthenogenetic embryos to the forelimb bud stage.** *Nature* 1977, **265**:53–55.
58. Obata Y, Ono Y, Akuzawa H, Kwon OY, Yoshizawa M, Kono T: **Post-implantation development of mouse androgenetic embryos produced by in-vitro fertilization of enucleated oocytes.** *Hum Reprod* 2000, **15**:874–880.
59. Kono T, Obata Y, Wu Q, Niwa K, Ono Y, Yamamoto Y, Park ES, Seo JS, Ogawa H: **Birth of parthenogenetic mice that can develop to adulthood.** *Nature* 2004, **428**:860–864.
60. Csankovszki G, Panning B, Bates B, Pehrson JR, Jaenisch R: **Conditional deletion of Xist disrupts histone macroH2A localization but not maintenance of X inactivation.** *Nat Genet* 1999, **22**:323–324.
61. Nesterova TB, Barton SC, Surani MA, Brockdorff N: **Loss of Xist imprinting in diploid parthenogenetic preimplantation embryos.** *Dev Biol* 2001, **235**:343–350.
62. Elling U, Taubenschmid J, Wirnsberger G, O'Malley R, Demers SP, Vanhaelen Q, Shukalyuk AI, Schmauss G, Schramek D, Schuetzen F, von Melchner H, Ecker JR, Stanford WL, Zuber J, Stark A, Penninger JM: **Forward and reverse genetics through derivation of haploid mouse embryonic stem cells.** *Cell Stem Cell* 2011, **9**:563–574.
63. Leeb M, Wutz A: **Derivation of haploid embryonic stem cells from mouse embryos.** *Nature* 2011, **479**:131–134.
64. Li W, Shuai L, Wan H, Dong M, Wang M, Sang L, Feng C, Luo GZ, Li T, Li X, Wang L, Zheng QY, Sheng C, Wu HJ, Liu Z, Liu L, Wang L, Wang XJ, Zhao XY, Zhou Q: **Androgenetic haploid embryonic stem cells produce live transgenic mice.** *Nature* 2012, **490**:407–411.
65. Yang H, Shi L, Wang BA, Liang D, Zhong C, Liu W, Nie Y, Liu J, Zhao J, Gao X, Li D, Xu GL, Li J: **Generation of genetically modified mice by oocyte injection of androgenetic haploid embryonic stem cells.** *Cell* 2012, **149**:605–617.
66. Leeb M, Walker R, Mansfield B, Nichols J, Smith A, Wutz A: **Germline potential of parthenogenetic haploid mouse embryonic stem cells.** *Development* 2012, **139**:3301–3305.
67. Leeb M, Wutz A: **Establishment of epigenetic patterns in development.** *Chromosoma* 2012, **121**:251–262.
68. Tsumura A, Hayakawa T, Kumaki Y, Takebayashi S, Sakaue M, Matsuoka C, Shimotohno K, Ishikawa F, Li E, Ueda HR, Nakayama J, Okano M: **Maintenance of self-renewal ability of mouse embryonic stem cells in the absence of DNA methyltransferases Dnmt1, Dnmt3a and Dnmt3b.** *Genes Cells* 2006, **11**:805–814.
69. Leeb M, Pasini D, Novatchkova M, Jaritz M, Helin K, Wutz A: **Polycomb complexes act redundantly to repress genomic repeats and genes.** *Genes Dev* 2010, **24**:265–276.
70. Kim Y, Sharov AA, McDole K, Cheng M, Hao H, Fan CM, Gaiano N, Ko MS, Zheng Y: **Mouse B-type lamins are required for proper organogenesis but not by embryonic stem cells.** *Science* 2011, **334**:1706–1710.
71. Ying QL, Wray J, Nichols J, Battle-Morera L, Doble B, Woodgett J, Cohen P, Smith A: **The ground state of embryonic stem cell self-renewal.** *Nature* 2008, **453**:519–523.
72. Chambers I, Silva J, Colby D, Nichols J, Nijmeijer B, Robertson M, Vrana J, Jones K, Grotewold L, Smith A: **Nanog safeguards pluripotency and mediates germline development.** *Nature* 2007, **450**:1230–1234.
73. Wray J, Kalkan T, Gomez-Lopez S, Eckardt D, Cook A, Kemler R, Smith A: **Inhibition of glycogen synthase kinase-3 alleviates Tcf3 repression of the pluripotency network and increases embryonic stem cell resistance to differentiation.** *Nat Cell Biol* 2011, **13**:838–845.
74. Marks H, Kalkan T, Menafrá R, Denisov S, Jones K, Hofemeister H, Nichols J, Kranz A, Stewart AF, Smith A, Stunnenberg HG: **The transcriptional and epigenomic foundations of ground state pluripotency.** *Cell* 2012, **149**:590–604.
75. Leitch HG, McEwen KR, Turp A, Encheva V, Carroll T, Grabole N, Mansfield W, Nashun B, Knezovich JG, Smith A, Surani MA, Hajkova P: **Naive pluripotency is associated with global DNA hypomethylation.** *Nat Struct Mol Biol* 2013, **20**:311–316.
76. Schumacher A, Doerfler W: **Influence of in vitro manipulation on the stability of methylation patterns in the Snurf/Snrpn-imprinting region in mouse embryonic stem cells.** *Nucleic Acids Res* 2004, **32**:1566–1576.
77. Fujikawa-Yamamoto K, Miyagoshi M, Yamagishi H: **Establishment of a tetraploid cell line from mouse H-1 (ES) cells highly polyploidized with demecolcine.** *Cell Prolif* 2007, **40**:327–337.
78. Carette JE, Guimaraes CP, Varadarajan M, Park AS, Wuethrich I, Godarova A, Kotecki M, Cochran BH, Spooner E, Ploegh HL, Brummelkamp TR: **Haploid genetic screens in human cells identify host factors used by pathogens.** *Science* 2009, **326**:1231–1235.
79. Carette JE, Guimaraes CP, Wuethrich I, Blomen VA, Varadarajan M, Sun C, Bell G, Yuan B, Muellner MK, Nijman SM, Ploegh HL, Brummelkamp TR: **Global gene disruption in human cells to assign genes to phenotypes by deep sequencing.** *Nat Biotechnol* 2011, **29**:542–546.

80. Rosmarin DM, Carette JE, Olive AJ, Starnbach MN, Brummelkamp TR, Ploegh HL: **Attachment of Chlamydia trachomatis L2 to host cells requires sulfation.** *Proc Natl Acad Sci U S A* 2012, **109**:10059–10064.
81. Jae LT, Raaben M, Riemersma M, van Beusekom E, Blomen VA, Velds A, Kerkhoven RM, Carette JE, Topaloglu H, Meinecke P, Wessels MW, Lefeber DJ, Whelan SP, van Bokhoven H, Brummelkamp TR: **Deciphering the glycosylome of dystroglycanopathies using haploid screens for lassa virus entry.** *Science* 2013, **340**:479–483.
82. Reiling JH, Clish CB, Carette JE, Varadarajan M, Brummelkamp TR, Sabatini DM: **A haploid genetic screen identifies the major facilitator domain containing 2A (MFSD2A) transporter as a key mediator in the response to tunicamycin.** *Proc Natl Acad Sci U S A* 2011, **108**:11756–11765.
83. Pettitt SJ, Rehman FL, Bajrami I, Brough R, Wallberg F, Kozarewa I, Fenwick K, Assiotis I, Chen L, Campbell J, Lord CJ, Ashworth A: **A Genetic Screen Using the PiggyBac Transposon in Haploid Cells Identifies Parp1 as a Mediator of Olaparib Toxicity.** *PLoS One* 2013, **8**:e61520.
84. Wiellette E, Grinblat Y, Austen M, Hirsinger E, Amsterdam A, Walker C, Westerfield M, Sive H: **Combined haploid and insertional mutation screen in the zebrafish.** *Genesis* 2004, **40**:231–240.
85. Kawahara M, Wu Q, Takahashi N, Morita S, Yamada K, Ito M, Ferguson-Smith AC, Kono T: **High-frequency generation of viable mice from engineered bi-maternal embryos.** *Nat Biotechnol* 2007, **25**:1045–1050.
86. Nguyen DK, Distèche CM: **Dosage compensation of the active X chromosome in mammals.** *Nat Genet* 2006, **38**:47–53.
87. Deng X, Berletch JB, Ma W, Nguyen DK, Hiatt JB, Noble WS, Shendure J, Distèche CM: **Mammalian X upregulation is associated with enhanced transcription initiation, RNA half-life, and MOF-mediated H4K16 acetylation.** *Dev Cell* 2013, **25**:55–68.
88. Lin F, Xing K, Zhang J, He X: **Expression reduction in mammalian X chromosome evolution refutes Ohno's hypothesis of dosage compensation.** *Proc Natl Acad Sci U S A* 2012, **109**:11752–11757.
89. Birchler JA, Veitia RA: **Gene balance hypothesis: connecting issues of dosage sensitivity across biological disciplines.** *Proc Natl Acad Sci U S A* 2012, **109**:14746–14753.
90. Levasseur A, Pontarotti P: **The role of duplications in the evolution of genomes highlights the need for evolutionary-based approaches in comparative genomics.** *Biol Direct* 2011, **6**:11.
91. Pessia E, Makino T, Bailly-Bechet M, McLysaght A, Marais GA: **Mammalian X chromosome inactivation evolved as a dosage-compensation mechanism for dosage-sensitive genes on the X chromosome.** *Proc Natl Acad Sci U S A* 2012, **109**:5346–5351.
92. Ganesan S, Richardson AL, Wang ZC, Iglehart JD, Miron A, Feunteun J, Silver D, Livingston DM: **Abnormalities of the inactive X chromosome are a common feature of BRCA1 mutant and sporadic basal-like breast cancer.** *Cold Spring Harb Symp Quant Biol* 2005, **70**:93–97.
93. Yildirim E, Kirby JE, Brown DE, Mercier FE, Sadreyev RI, Scadden DT, Lee JT: **Xist RNA is a potent suppressor of hematologic cancer in mice.** *Cell* 2013, **152**:727–742.
94. Ravi M, Chan SW: **Haploid plants produced by centromere-mediated genome elimination.** *Nature* 2010, **464**:615–618.
95. Hubner K, Fuhrmann G, Christenson LK, Kehler J, Reinbold R, De La Fuente R, Wood J, Strauss JF 3rd, Boiani M, Scholer HR: **Derivation of oocytes from mouse embryonic stem cells.** *Science* 2003, **300**:1251–1256.
96. Hayashi K, Ogushi S, Kurimoto K, Shimamoto S, Ohta H, Saitou M: **Offspring from oocytes derived from in vitro primordial germ cell-like cells in mice.** *Science* 2012, **338**:971–975.
97. Hayashi K, Ohta H, Kurimoto K, Aramaki S, Saitou M: **Reconstitution of the mouse germ cell specification pathway in culture by pluripotent stem cells.** *Cell* 2011, **146**:519–532.

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