

This Health Hazard Evaluation (HHE) report and any recommendations made herein are for the specific facility evaluated and may not be universally applicable. Any recommendations made are not to be considered as final statements of NIOSH policy or of any agency or individual involved. Additional HHE reports are available at <http://www.cdc.gov/niosh/hhe/reports>

**HEALTH HAZARD EVALUATION
REPORT**

**HETA 92-139-XXXX
OHIO UNIVERSITY
JENNINGS HOME
ATHENS, OHIO**

PREFACE

The Hazard Evaluations and Technical Assistance Branch of NIOSH conducts field investigations of possible health hazards in the workplace. These investigations are conducted under the authority of Section 20(a)(6) of the Occupational Safety and Health Act of 1970, 29 U.S.C. 669(a)(6) which authorizes the Secretary of Health and Human Services, following a written request from any employer and authorized representative of employees, to determine whether any substance normally found in the place of employment has potentially toxic effects in such concentrations as used or found.

The Hazard Evaluations and Technical Assistance Branch also provides, upon request, medical, nursing, and industrial hygiene technical and consultative assistance (TA) to federal, state, and local agencies; labor; industry; and other groups or individuals to control occupational health hazards and to prevent related trauma and disease.

Mention of company names or products does not constitute endorsement by the National Institute for Occupational Safety and Health.

TABLE OF CONTENTS

I. SUMMARY	1
Keywords	2
II. INTRODUCTION	3
III. BACKGROUND	3
IV. MATERIALS AND METHODS	4
A. INDUSTRIAL HYGIENE EVALUATION	4
B. MEDICAL EVALUATION	7
V. EVALUATION CRITERIA	7
VI. RESULTS	11
A. ENVIRONMENTAL	11
B. MEDICAL	17
VII. CONCLUSIONS	17
VIII. REFERENCES	18
IX. AUTHORSHIP AND ACKNOWLEDGEMENTS	21
X. DISTRIBUTION AND AVAILABILITY OF REPORT	22

TABLE

Table 1. Environmental Sample Locations 4

LIST OF FIGURES

Figure 1. Basement Floor Plan with Sample Locations Indicated . . . 5

Figure 2. First Floor Plan with Sample Locations Indicated 6

Figure 3. Second Floor Plan with Sample Locations Indicated 6

Figure 4. Mean Carbon Dioxide Concentrations at Various Locations 12

Figure 5. Mean Temperature and Humidity Concentrations at Various Locations 13

Figure 6. ASHRAE Thermal Comfort Chart 14

Figure 7. Microorganism Concentrations at Selected Building Locations (resp
fraction is noted above bars) 15

**HETA 92-139-2274
NOVEMBER 1992
OHIO UNIVERSITY
JENNINGS HOME
ATHENS, OHIO**

**NIOSH INVESTIGATORS:
Kenneth F. Martinez,
M.S.E.E
Calvin K. Cook
Scott Deitchman, M.D.**

I. SUMMARY

On March 2 and 3, 1992, the National Institute for Occupational Safety and Health (NIOSH) conducted a health hazard evaluation at the Jennings Home, Ohio University, Athens, Ohio. This health hazard evaluation was conducted in response to a request by the Environmental Safety Coordinator, Department of Environmental Health and Safety, Ohio University, to investigate potential indoor environmental quality problems (specifically, possible microbiological contamination) suspected because of one employee's (index case) health complaints. The complaints specified in the request, included facial itching; eye and membrane irritation; nausea and diarrhea at the office.

The investigation included a physical inspection of the heating, ventilation, and air conditioning (HVAC) systems for the first and second floor; the collection and analysis of environmental air samples for analysis of cyclohexylamine and bioaerosols (specifically, saprophytic fungi, bacteria, and thermophilic actinomycetes); and measurement of carbon dioxide, temperature, and relative humidity (RH).

The physical inspection did not reveal any visible evidence that would indicate a significant or unusual microbial contamination source. Filters were free of accumulation, ventilation ducts and exterior insulation were in good shape, and heating/cooling coils were free of standing water and/or slime accumulation. Bioaerosol samples for fungi, bacteria, and thermophilic actinomycetes, collected using Andersen two-stage viable cascade impactors, did not support the conclusion of microbial sources of contamination. Bacterial concentrations ranged from 0 to 14 colony-forming units per cubic meter (CFU/m³) at both outdoor locations. Fungal concentrations ranged from 63 to 706 CFU/m³ in the conference room and complaint room, respectively. The samples collected in the conference and complaint room for thermophilic bacteria were 4 and 14 CFU/m³. Air samples had no detectable cyclohexylamine based on an analytical limit of detection of 0.01 mg/sampling volume (µg/liter for a 96 liter sampling volume). Mean carbon dioxide concentration was 525 parts per million (ppm) for the sample collected at the reception desk. Temperature measurements in the Jennings Home averaged 71°F; the RH measurements averaged 35%.

Questionnaires inquiring about the comfort and symptoms experienced in the building were distributed to all available employees in the Jennings Home. The questionnaires were returned to the NIOSH investigators. One individual, the index case, reported symptoms of runny nose and headaches. Specific quality complaints included lack of air circulation (stuffy feeling), noticeable and disturbing noises from the heat register. Evaluation of the medical records of the index case employee indicated that the bulk of these records were from pre-hospital and outpatient visits for reasons unrelated to working in this building.

This investigation did not find any current environmental exposures or conditions that would constitute a health hazard.

KEYWORDS: SIC 9441 (Administration of Social, Manpower, and Income Maintenance Programs), indoor environmental quality, bioaerosols, fungi, thermophilic actinomycetes, cyclohexylamine

II. INTRODUCTION

On March 2 and 3, 1992, the National Institute for Occupational Safety and Health (NIOSH) conducted a health hazard evaluation (HHE) at the Jennings Home at Ohio University, Athens, Ohio. This HHE was conducted in response to a request received on February 5, 1992, from the Environmental Safety Coordinator, Environmental Health and Safety, Ohio University, to investigate potential indoor environmental problems (specifically, possible microbiological contamination) suspected to be related to an employee's health complaints. This report presents the results of the environmental and medical assessment which includes a ventilation system inspection, air sampling for cyclohexylamine (CHA) and bioaerosols, and measurement of temperature, humidity, and carbon dioxide (CO₂).

III. BACKGROUND

The administration offices of the College of Fine Arts, Ohio University, currently occupies the Jennings House. The building was originally constructed in 1900 and has a current usable capacity of approximately 2900 square feet. The occupancy of the building, however, increased and frequent traffic can be observed at peak times of the year (i.e., class registration). The heating, ventilating, and air-conditioning system is maintained by the University. Smoking is not permitted in the building spaces of the building.

According to the request, a single employee had "facial itching, eye and nose irritation, nausea and diarrhea when at work." Prior to the NIOSH investigation, the Environmental Safety Coordinator conducted an evaluation of the building for environmental and microbial contaminants. Environmental samples were collected for detection of carbon monoxide, CO₂, oxygen, hydrogen sulfide, and formaldehyde levels and for an evaluation of airborne microbial contamination (settling plates). Temperature and relative humidity (RH) were also measured. According to the Environmental Safety Coordinator, the environmental sample results were unremarkable with the exception of the settling plate samples. One of three settling plate samples was detected as having "numerous colonies," the other samples had "small numbers." It was noted at that point that NIOSH investigators were asked to further evaluate the building with an emphasis on microbial contamination.

IV. MATERIALS AND METHODS

A. INDUSTRIAL HYGIENE EVALUATION

Environmental samples were collected at selected locations throughout the building for CHA and bioaerosols (specifically, saprophytic fungi, bacteria, and thermophilic actinomycetes). In addition, direct measurement, at the selected locations, was conducted for CO₂, temperature, and humidity. Environmental sample locations are listed in Table 1 (refer to Figures 1 through 3 for building floor plan and sample placement). All samples were collected during the work shift of March 3, 1992.

To determine the concentrations of airborne microorganisms at selected locations in the building, the Andersen 2-stage viable cascade impactor was used at a flow rate of 28.3 liters per minute (lpm). The 50% effective cutoff diameter for the Andersen sampler is 8 μm - hence, larger non-respirable particles are collected on the top stage and smaller, respirable particles are collected on the bottom stage. Standard Plate Count and Malt Extract agars were used for the enumeration of bacteria (thermophilics included) and fungi, respectively. The sample plates for bacteria were incubated at 30°C. The sample plates for thermophilic bacteria were incubated at 55°C to promote the growth of thermotolerant bacteria (specifically, thermophilic actinomycetes - TA). The sample plates for fungi were incubated at 28°C. A sample time of 10 minutes was used at all sample locations. At each location, three samples were collected for bacteria and fungi. For thermophilic bacteria, single sample runs were made at two locations. Temperature and humidity were recorded for each sample run.

Table 1. Environmental Sample Locations

SAMPLE LOCATION	SAMPLE TYPE		
	Bioaerosols	CHA	Temperature, Humidity, and CO ₂
Next to HVAC unit (basement)		✓	✓
Reception desk (Room 109)			✓
Complaint office (Room 106)	✓	✓	✓
Kitchen (Room 103)			✓
Non-complaint office (Room 110A)		✓	
Non-complaint office (Room 203)		✓	✓
Conference room (Room 209)	✓	✓	✓
Outdoors	✓		✓
Complaint office (temporary space)			✓

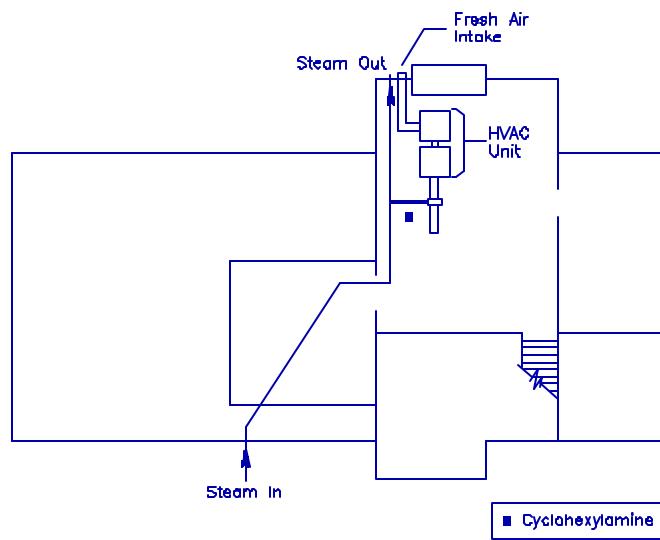


Figure 1. Basement Floor Plan with Sample Locations Indicated

Figure 2.
First Floor Plan
with Sample
Locations
Indicated

Humidity, and CO₂ were collected at each sample location for four rounds of beginning at 7:20 a.m., followed by subsequent sampling rounds at 8:56 a.m., and 2:25 a.m. Carbon dioxide was measured using a Gastech RI 411 dioxide monitor (Gastech, Inc., Newark, California) calibrated before the day's : collected using 800 parts per million (ppm) CO₂ in nitrogen (Alphagaz, Divisic Corporation, Cambridge, Maryland) as a calibrant. Temperature and RH were using a Vaisala HM 34 temperature and humidity meter (Vaisala Oy, Helsinki,

Direct measurements for temperature,

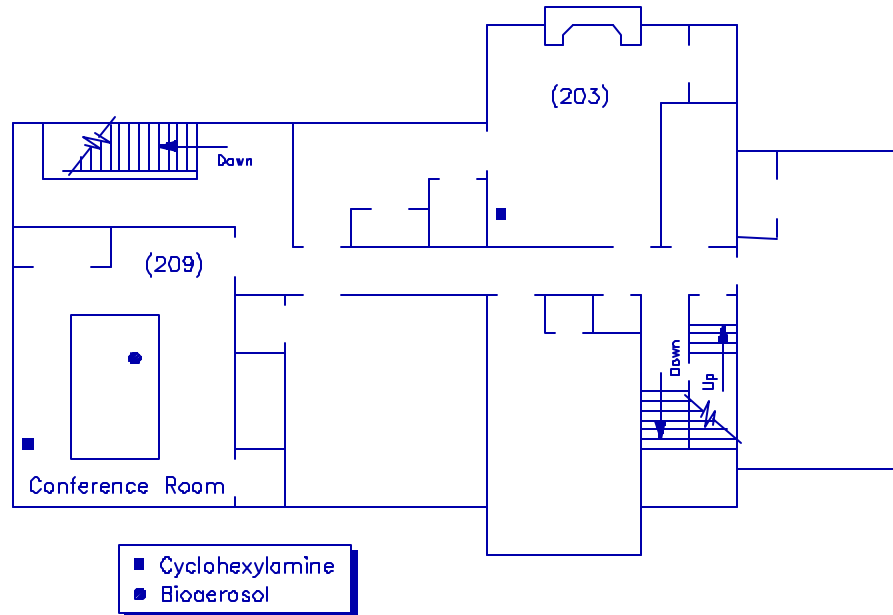


Figure 3. Second Floor Plan with Sample Locations Indicated

Steam is supplied to the building through a closed, common University piped steam system. CHA is added to the central boilers to retard corrosion system. Six full-shift environmental samples for CHA were collected on sorbent tubes with Gillian personal sampling pumps calibrated at a flow rate of 0.2 lpm prior to the field investigation. Samples for cyclohexane were collected to investigate the possibility of leaks in the steam heat exchanger. The sample tubes were analyzed using NIOSH Method 2010 with modifications. The silica gel tubes were desorbed (sonication) for 3 hours in 1.0 milliliter of 0.1M H₂SO₄, 10% methanol, and water. From the solution, 0.5 ml was removed and basified with 0.5 ml of 0.3M KOH. The samples were analyzed with a Hewlett-Packard Model 5730A gas chromatograph equipped with a nitrogen phosphorus detector; the column was a 30 meter (m) x 0.32 millimeter fused silica capillary coated internally with 0.25 micrometer (µm) of Stationary Phase.

B. MEDICAL EVALUATION

A medical evaluation was not initially involved in the investigation. However, questionnaires inquiring about symptoms experienced while in the building were disseminated to all available employees (six were returned to the NIOSH investigators) and a medical officer was assigned, after the site visit, to review the medical records of the employee mentioned in the request.

V. EVALUATION CRITERIA

NIOSH investigators have completed over 1100 investigations of the occurrence of indoor environmental problems in a wide variety of non-industrial settings. The majority of these investigations have been conducted since 1979.

The symptoms and health complaints reported to NIOSH by building occupants have been diverse and usually not suggestive of any particular medical diagnosis associated with a causative agent. A typical spectrum of symptoms has included headaches, unusual fatigue, varying degrees of itching or burning eyes, irritation of the skin, nasal congestion, dry or irritated throats and other respiratory irritations. Typically, the workplace environment has been implicated because workers report that their symptoms lessen or resolve when they leave the building. A number of published studies have reported high prevalences of symptoms among occupants of office buildings.²⁻⁶ Scientists investigating indoor environmental problems have concluded that there are multiple factors contributing to building-related occupant complaints. Among these factors are imprecisely defined characteristics of heating, ventilation, and air-conditioning (HVAC) systems, cumulative effects of exposure to low concentrations of multiple chemical pollutants, odors, elevated concentrations of particulate matter, microbiological contamination, and physical factors such as thermal comfort, lighting, and noise.⁹⁻¹⁴ Indoor environmental pollutants can originate from either outdoor sources or indoor sources.

There are also reports describing results which show that occupant perceptions of the indoor environment are more closely related than any measured indoor condition to the occurrence of symptoms.¹⁵⁻¹⁷ Some studies have shown relationships between psychological, social, and organizational factors in the workplace and the occurrence of symptoms and comfort complaints.¹⁷⁻²⁰

Less often, an illness may be found to be specifically related to something in the building environment. Some examples of potentially building-related illnesses include allergic rhinitis, allergic asthma, hypersensitivity pneumonitis, Legionnaires' disease, Pontiac fever, carbon monoxide poisoning, and reaction to boiler corrosion products. The first three conditions can be caused by various microorganisms or other materials. Legionnaires' disease and Pontiac fever are caused by *Legionella* species. Sources of carbon monoxide include vehicle exhaust and inadequately-ventilated kerosene heaters or other fuel-burning appliances. Exposure to boiler acid can occur if boiler steam is used for humidification or is released by accident.

Problems NIOSH investigators have found in the non-industrial indoor environment have included poor air quality due to ventilation system deficiencies, over concentrations of volatile organic chemicals from furnishings, machines, structural components, building and contents, tobacco smoke, microbiological contamination, air pollutants; comfort problems due to improper temperature and RH conditions, lighting, and unacceptable noise levels; adverse ergonomic conditions; and related psychosocial stressors. In most cases, however, these problems are directly linked to the reported health effects.

Standards specifically for the non-industrial indoor environment do not exist. The Occupational Safety and Health Administration (OSHA) and the American Conference of Governmental Industrial Hygienists (ACGIH) have published standards or recommended limits for occupational exposures.²¹⁻²³ With few exceptions, pollutant concentrations observed in non-industrial indoor environments fall well below these published occupational standards or recommended limits. The American Society of Heating, Refrigerating, and Air-Conditioning Engineers (ASHRAE) has published recommended building ventilation design and thermal comfort guidelines.^{24,25} The ACGIH has also developed a manual of guidelines for approaching investigations of building-related complaints that are caused by airborne living organisms or their effluents.²⁶

Measurement of indoor environmental contaminants has rarely proved to be useful in determining the cause of symptoms and complaints except where there are unusual sources, or a proven relationship between contaminants and specific building-related illnesses. The low-level concentrations of particles and volatile mixtures of organic materials usually found are difficult to interpret and usually impossible to causally link to observed and reported health symptoms. However, measuring ventilation and comfort indicators such as CO₂, temperature and humidity are proven useful in the early stages of an investigation in providing information about the proper functioning and control of HVAC systems.

NIOSH and the Environmental Protection Agency (EPA) jointly published a manual on building air quality, written to help prevent environmental problems in buildings and to solve problems when they occur.²⁷ This manual suggests that indoor environmental quality (IEQ) is a constantly changing interaction of a complex set of factors. The most important elements involved in the development of IEQ problems are: 1) the source of odors or contaminants; 2) a problem with the design or operation of the HVAC system; 3) a pathway between the contaminant source and the location of the complaint; 4) and the building occupants. A basic understanding of these factors is critical to preventing, investigating, and resolving IEQ problems.

The basis for measurements made during this evaluation are listed below.

A. CARBON DIOXIDE

Carbon dioxide (CO₂) is a normal constituent of exhaled breath and, if measured, may be useful as a screening technique to evaluate whether adequate levels of fresh air are being introduced into an occupied space. The ANSI/ASHRAE Standard 62-1989, Ventilation for Acceptable Indoor Air Quality, recommends outdoor air supply rates of 20 cubic feet per minute per person (cfm/person) for office spaces and conference rooms, 15 cfm/person for reception areas, and 10 cfm/person for smoking lounges, and provides estimated maximum occupancy figures for each area.²⁴

Indoor CO₂ concentrations are normally higher than the generally constant ambient CO₂ concentration (range 300-350 ppm). When indoor CO₂ concentrations exceed 1000 ppm in areas where the only known source is human breath, inadequate ventilation is suspected. Elevated CO₂ concentrations indicate that other indoor contaminants may also be increased.

B. TEMPERATURE AND RELATIVE HUMIDITY

The perception of comfort is related to one's metabolic heat production, the transfer of heat to the environment, physiological adjustments, and body temperatures. Heat transfer from the body to the environment is influenced by factors such as temperature, humidity, air movement, personal activities, and clothing. ANSI/ASHRAE Standard 55-1981 specifies conditions in which more than 90% of the occupants would be expected to find the environment thermally comfortable.²⁵

C. CYCLOHEXYLAMINE

Cyclohexylamine is a colorless to slightly yellow liquid with a strong, fishy odor. It has been used in the production of rubber-processing chemicals, insecticides, plasticizers, and dry-cleaning soaps; additionally, it has been used as a corrosion inhibitor in boiler feed water. Cyclohexylamine is a severe irritant of the eyes and has a moderate sensitizing potential. The ACGIH Threshold Limit Value (TLV), the NIOSH Recommended Exposure Limit (REL), and the OSHA

Recommended Exposure Limit (PEL) are 10 ppm ($\approx 40 \text{ mg/m}^3$) over an work shift.^{21, 23, 27}

VI. RESULTS

A. ENVIRONMENTAL

The HVAC system consisted of two separate units; the first floor was serviced by a unit in the basement and the second floor was serviced by a unit in the first floor. These units are similar in design to home HVAC systems with a variable volume heating component; heat is provided by coils connected to the University steam supply. The HVAC units are designed to provide a constant volume of air during the thermostatically controlled "on" cycle. The original design of the HVAC system specified 100% air recirculation in the interior spaces. However, in response to health complaints, the basement HVAC unit was ducted to the outside to provide fresh air (percentage unknown) to the offices (including the complaint room) serviced by this unit. Each HVAC unit was designed with coarse filters at the upstream end of the fan.

Smoke tubes were used to document the airflow patterns through-out the building. General air movement in the offices resulted in airflow into the offices from the hallway. This was expected due to the placement of the return air duct at the end of the hallway. Stagnant pockets of air were observed through-out the building. The HVAC unit was cycled "off."

Physical inspections were conducted on both HVAC units. The inspections did not reveal any visible evidence that would indicate a microbial contamination. Specifically, the filters appeared free of debris accumulation; the ventilation ducts and exterior insulation were in good shape; and the heating coils, and the areas beneath, were absent of standing water and/or "slime" accumulation. In addition, environmental samples for bioaerosols were collected at the request of the Environmental Safety Manager.

The results of the CO₂ monitoring performed on March 3, 1992, are graphically presented in Figure 4. The mean CO₂ concentrations ranged from 375 ppm for the outside sample to 525 ppm for the sample collected at the reception desk. All of the mean CO₂ concentrations were below the ASHRAE recommended limit of 1000 ppm.²⁴ However, the occupancy rate in the complaint room was less than the limiting criteria of 7 occupants per 1000 square feet in ASHRAE 62-1989. Observation of the individual data points (Appendix B) did not indicate a large variation in the concentrations over the course of the survey. It should be noted that the complaint room was vacant at the time of the survey; therefore, the CO₂ value may not be representative. However, it is doubtful that the occupancy of the office by a single individual would greatly influence the concentration from those observed during the survey. This is validated by the small CO₂ variations among the other offices sampled.

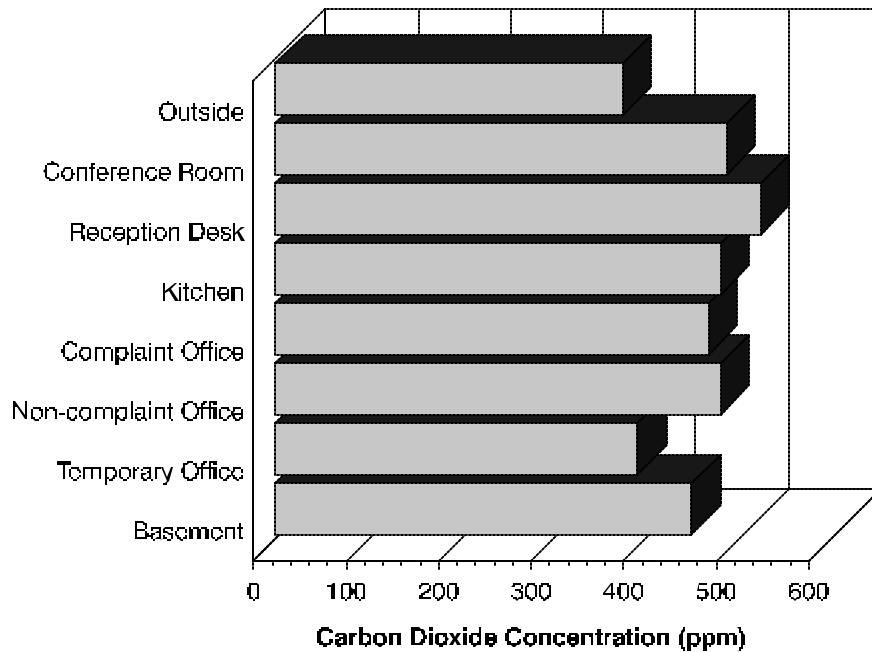


Figure 4. Mean Carbon Dioxide Concentrations at Various Locations

The concentration of CO₂ in offices designed for single occupancy can be significantly elevated by intermittent occupant load--commonly observed in university environments due to student traffic. For example, the mean CO₂ concentration in the Reception Desk area was higher than all other sample locations (refer to Figure 4). In addition, the variance was greater than all other locations (the standard deviation was 46 ppm compared to less than 46 ppm for all other sample locations) suggesting significant fluctuation in the CO₂ concentrations over the work day. The increased concentrations and variance can be mainly attributed to the increased student traffic observed (at the time of the survey) in the reception area and its proximity to other interior building locations. Increasing the fresh air supply in areas expected to receive greater student traffic can help to reduce concentration variation and ensure that levels remain below recommended limits.

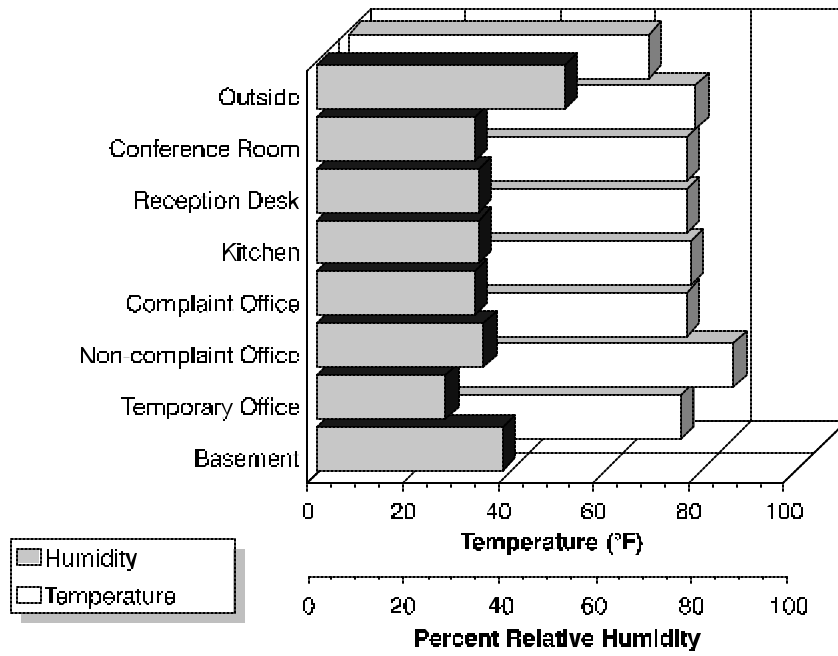


Figure 5. Mean Temperature and Humidity Concentrations at Various Locations

The results of monitoring for temperature and humidity are graphically in Figure 5. The mean temperature measurements ranged from 63°F at a sampling location outside of the building to 81°F in the temporary office (located in a separate building). The mean temperatures in the Jennings were fairly stable from office to office over the work day (overall mean of 73°F with a standard deviation of 1°F). The mean humidity measurements ranged from 10% at the sampling location outside of the building to 27% in the temporary office. Similar to the temperature measurements, the RHs in the Jennings Home were fairly stable from office to office over the work day (overall mean of 35% with a standard deviation of 2%). The indoor temperatures and RHs, with the exception of the temporary office, are within the limits recommended by ASHRAE thermal comfort chart (Figure 6). This chart specifies the acceptable (and will feel thermally comfortable) ranges of operative temperature and humidity for persons clothed in typical summer and winter clothing, performing mainly sedentary activity.²⁴ The temperature in the temporary office was beyond the comfort range established for the winter months.

The air samples for cyclohexylamine were all non-detectable based on an analytical limit of detection of 0.01 mg/sample, which equates to a minimum detectable concentration of 0.1 µg/liter, assuming a sampling volume of 96 liters.

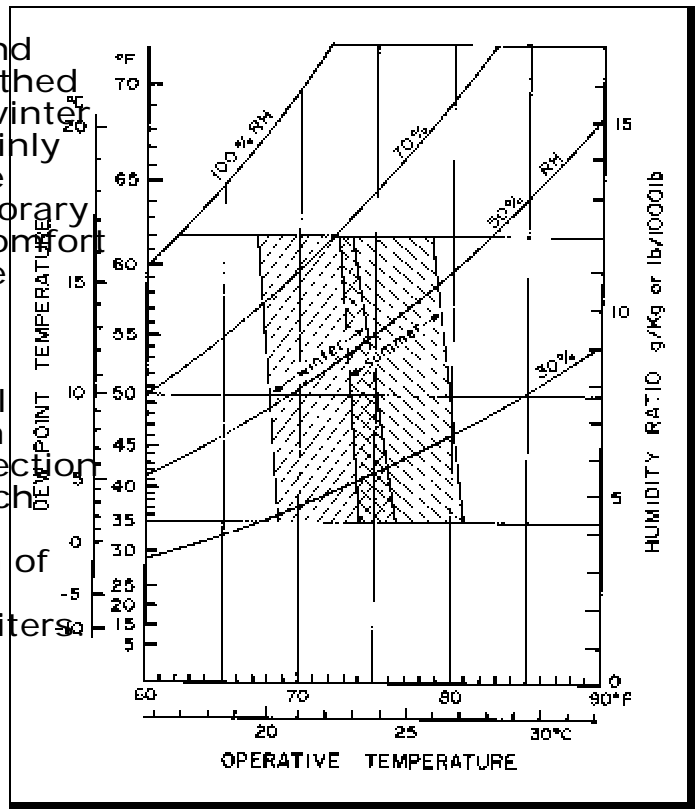


Figure 6. ASHRAE Thermal Comfort Chart

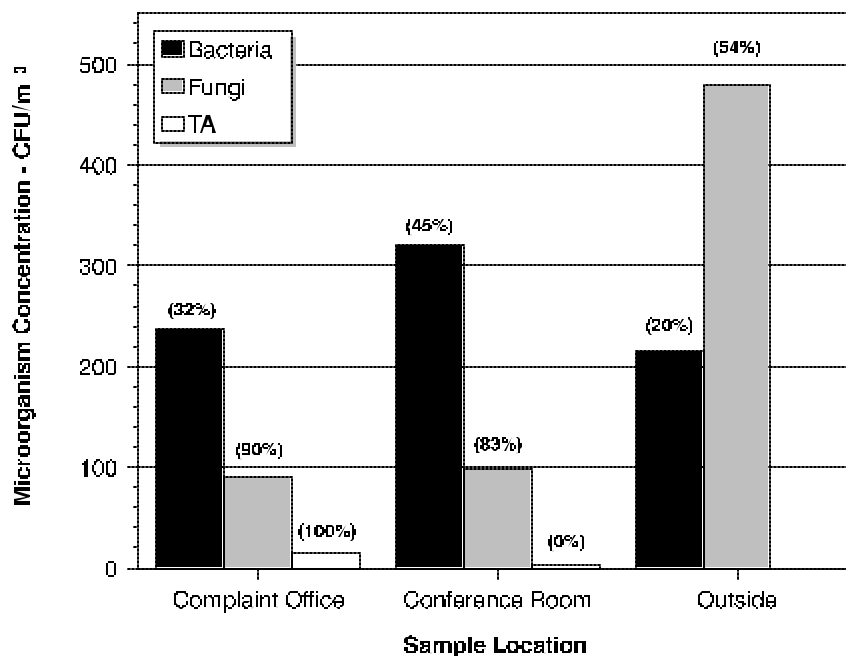


Figure 7. Microorganism Concentrations at Selected Building Locations (respirable fraction is noted above bars)

A graphical summary of the bioaerosol air sampling results (bacteria, fungi, and TA) is presented in Figure 7. Microorganism concentrations are indicated by the height of the bars and the respirable fraction is shown above the bars in parentheses. The samples in the complaint office had a mean bacteria content of 237 CFU/m³ and a standard deviation of 122 CFU/m³; this is statistically similar (α level of 0.05) to the samples collected outdoors (mean of 216 CFU/m³ and a standard deviation of 192 CFU/m³). The percentage of respirable particles (defined by the sampling method as the 50% effective cut-off diameter at 8 μ m) were low for all sample locations. The respirable fraction in the complaint office (mean of 32% with a standard deviation of 12%) and the non-complaint office (mean of 45% with a standard deviation of 25%) were slightly greater than that encountered in the outdoor samples (20% with a standard deviation of 1%). Observation of the taxa ranking (see the Appendix) indicates dissimilar species encountered indoors versus outdoors. The bacterial component of the outdoor samples were composed of various *Bacillus* species; common gram-positive soil flora. However, indoors the bacteria were predominated by non-pathogenic, gram-positive species *Micrococcus*, *Corynebacterium*, and *Staphylococcus*. These bacterial species are commonly associated with desquamated human skin and are a direct indication of occupant activity.²⁸ These bacterial species identified in the complaint and non-complaint offices have not been documented in the outdoor samples.

as causative agents in hypersensitivity pneumonitis or other related illnesses.

The complaint office had a mean fungal content of 90 CFU/m³ and a standard deviation of 11 CFU/m³, and the non-complaint office had a mean fungal content of 99 CFU/m³ and a standard deviation of 52 CFU/m³; these were statistically below (α level at 0.05) the concentrations encountered outdoors (mean 286 CFU/m³ and a standard deviation of 286 CFU/m³). In addition, the results for the complaint office were statistically similar (α level at 0.05) to those of the non-complaint office. The percentage of respirable particles were high for the non-complaint office (mean of 90% with a standard deviation of 7%) and the complaint office (mean of 83% with a standard deviation of 11%) and greater than that encountered in the outdoor sample (mean of 50% with a standard deviation of 6%). Observation of the taxa ranking does not indicate **amplification** of fungal species that have typically been associated with health effects (*Aspergillus*, *Penicillium*, *Sporobolomyces*, *Alternaria*, etc.).²⁶ In order to illicit immunologic responses in susceptible individuals, a microorganism must be present in the environment (reservoir), capable of propagating at concentrations necessary to induce responses (amplification), and available as an aerosol to the susceptible individual (dissemination).¹²

Cladosporium was the most predominant species in the outdoor samples and this is consistent with "normal" outdoor taxa ranking.²⁸ *Alternaria* was present in the top plate (non-respirable fraction) of some of the outdoor samples and may be a result of *Alternaria's* larger spores in comparison to those of *Cladosporium*.²⁸ *Penicillium*, *Ulocladium*, and yeasts were also identified in small concentrations in the outdoor air. Indoors, the air was predominated by *Cladosporium* and *Penicillium*; lower concentrations of other common saprophytic molds were also identified. *Penicillium* was the most numerous species on the bottom stage (respirable fraction) of most indoor samples; however, the concentrations, when compared to those encountered outdoors, were very similar supporting the conclusion of no amplification. *Gliocladium* was identified as the predominant species in most of the samples in the complaint office which would indicate a possible indoor reservoir. However, *Gliocladium* has not been associated with cases of hypersensitivity pneumonitis and the concentrations encountered were low.

Results for thermophilic actinomycetes (TA) were only available as single samples in the complaint office (14 CFU/m³) and non-complaint office (14 CFU/m³). Although these numbers are not typical of levels which have been implicated in cases of hypersensitivity pneumonitis, their existence as an indicator of a problem cannot be ruled out. The current knowledge concerning the typical concentrations of TA's in the ambient environment is extremely limited due to the inability to detect TA concentrations at "elevated levels" during the survey period. This may be the result of an unfavorable operating state in the building ventilation system (TA's prefer warm temperatures and a humid environment) specific to the survey period.

B. *MEDICAL*

Medical records of the employee mentioned in the request were supplied to the NIOSH investigator and reviewed by a NIOSH medical officer. The bulk of the records are from previous hospital and outpatient visits for reasons unrelated to working at this building. None of the medical tests described in these records suggests an environmentally-induced disease. However, there are few any specific tests for such diseases. Carboxyhemoglobin was measured on a single test and was less than 1%, which suggests there was no exposure to carbon monoxide at the time of that test.

The six questionnaires did not reveal work-related health complaints from any of the individuals with one exception. One person reported symptoms of recurrent headaches. Specified air quality complaints included lack of air circulation (stagnant feeling), noticeable odors, and disturbing noises from the heat register.

VII. CONCLUSIONS

The request was prompted by concerns of a single worker regarding the environmental quality and health symptoms including tearing eyes; itching of the skin of the throat, neck, and cheeks; mucous drainage; occasional chest pain; nausea; and diarrhea. The symptoms began in mid-January of 1991, with occurrences entering the building and subsidence out of the building. There are reports of interior water damage and pesticide use (over the past 4 years) and smoking is not permitted in the building. A portable humidification unit was in operation for 4 years in the reception area but its use has since been discontinued as it was removed. Visual inspection of the unit indicated inadequate maintenance. Evaluation of the medical records of the affected employee and environmental air sampling for CO₂, temperature, humidity, bioaerosols, and cyclohexylamine support the conclusion of an environmental causative agent.

The building HVAC unit is designed to provide tempered air during the "occupied" periods as monitored by interior thermostats. This intermittent load may not be capable of providing adequate fresh air (per ASHRAE guidelines) during peak occupancy (during class registration). Although CO₂ measurements at the time of the investigation did not indicate deficiencies in the ventilation system, during seasonal periods when the ventilation system minimally operates and occupancy increases, additional monitoring for CO₂ should be conducted. If monitored concentrations surpass the ASHRAE criterion of 1000 ppm, efforts should be made to increase the fresh air supply during times of increased occupancy.

VIII. REFERENCES

1. Kreiss KK, Hodgson MJ [1984]. Building associated epidemics. In: W Dudley CS, Copenhaver ED, eds. Indoor air quality. Boca Raton, FL: pp 87-108.
2. Gammage RR, Kaye SV, eds. [1985]. Indoor air and human health: Part of the Seventh Life Sciences Symposium. Chelsea, MI: Lewis Publishers
3. Woods JE, Drewry GM, Morey PR [1987]. Office worker perceptions of quality effects on discomfort and performance. In: Seifert B, Esdorn H et al., eds. Indoor air '87, Proceedings of the 4th International Conference on Air Quality and Climate. Berlin Institute for Water, Soil and Air Hygiene
4. Skov P, Valbjorn O [1987]. Danish indoor climate study group. The "sick building syndrome" in the office environment: The Danish town hall study. *Environ Int* 13:399-349.
5. Burge S, Hedge A, Wilson S, Bass JH, Robertson A [1987]. Sick building syndrome: a study of 4373 office workers. *Ann Occup Hyg*. 31:493-502.
6. Kreiss K [1989]. The epidemiology of building-related complaints and symptoms. *Occupational Medicine: State of the Art Reviews*. 4(4):575-592.
7. Norbäck D, Michel I, Widstrom J [1990]. Indoor air quality and person related to the sick building syndrome. *Scan J Work Environ Health*. 16:1-6.
8. Morey PR, Shattuck DE [1989]. Role of ventilation in the causation of sick building syndrome associated illnesses. *Occupational Medicine: State of the Art Reviews* 642.
9. Mendell MJ, Smith AH [1990]. Consistent pattern of elevated symptoms in poorly conditioned office buildings: A reanalysis of epidemiologic studies. *American Journal of Public Health*. 80(10):1193-1199.
10. Molhave L, Bachn B, Pedersen OF [1986]. Human reactions to low concentrations of volatile organic compounds. *Environ Int*. 12:167-176.
11. Fanger PO [1989]. The new comfort equation for indoor air quality. *Indoor Air* 31(10):33-38.
12. Burge HA [1989]. Indoor air and infectious disease. *Occupational Medicine: State of the Art Reviews*. 4(4):713-722.
13. Robertson AS, McInnes M, Glass D, Dalton G, Burge PS [1989]. Building sickness, are symptoms related to the office lighting? *Ann Occup Hyg*. 59.
14. Levin H [1989]. Building materials and indoor air quality. *Occupational Medicine: State of the Art Reviews*. 4(4):667-694.

15. Wallace LA, Nelson CJ, Dunteman G [1991]. Workplace characteristics with health and comfort concerns in three office buildings in Washington, D.C. Geshwiler M, Montgomery L, and Moran M, eds. Healthy buildings. Proceedings of the ASHRAE/ICBRSD conference IAQ'91. Atlanta, GA. The American Society of Heating, Refrigerating, and Air-Conditioning Engineers, Inc.
16. Haghghat F, Donnini G, D'Addario R [1992]. Relationship between objective and subjective discomfort as perceived and as measured objectively. *Indoor Environ.*
17. NIOSH [1991]. Hazard evaluation and technical assistance report: Library of Congress Madison Building, Washington, D.C. Cincinnati, OH: U.S. Department of Health and Human Services, Public Health Service, Centers for Disease Control and Prevention, National Institute for Occupational Safety and Health, NIOSH Report No. 92-100. 364-2104 - Vol. III.
18. Skov P, Valbjørn O, Pedersen BV [1989]. Influence of personal characteristics, environmental factors, and psychosocial factors on the sick building syndrome. *Work Environ Health.* 15:286-295.
19. Boxer PA [1990]. Indoor air quality: A psychosocial perspective. *J Occup Environ Hygiene.* 32(5):425-428.
20. Baker DB [1989]. Social and organizational factors in office building-associated illness. *Occupational Medicine: State of the Art Reviews.* 4(4):607-616.
21. CDC [1992]. NIOSH recommendations for occupational safety and health. Compendium of policy documents and statements. Cincinnati, OH: U.S. Department of Health and Human Services, Public Health Service, Centers for Disease Control, National Institute for Occupational Safety and Health. (NIOSH) Publication No. 92-100.
22. Code of Federal Regulations [1989]. OSHA Table Z-1-A. 29 CFR 1910.1000. Washington, DC: U.S. Government Printing Office, Federal Register.
23. ACGIH [1991]. 1991-1992 Threshold limit values for chemical substances and physical agents and biological exposure indices. Cincinnati, OH: American Conference of Governmental Industrial Hygienists.
24. ASHRAE [1990]. Ventilation for acceptable indoor air quality. Atlanta, GA: American Society of Heating, Refrigerating, and Air-conditioning Engineers. ANSI/ASHRAE Standard 62-1989.
25. ASHRAE [1981]. Thermal environmental conditions for human occupancy. Atlanta, GA: American Society for Heating, Refrigerating, and Air-conditioning Engineers. ANSI/ASHRAE Standard 55-1981.

26. ACGIH [1989]. Guidelines for the assessment of bioaerosols in the indoor environment. Cincinnati, OH: American Conference of Governmental Industrial Hygienists.
27. Hathaway GJ, Proctor NH, Hughes JP, Fischman ML [1991]. Proctor & Fischman's Chemical Hazards of the Workplace. Third Edition. New York, NY: Van Nostrand Reinhold.
28. Miller RD [1992]. Microbiological laboratory report dated April 4, 1992 regarding analysis of airborne bacteria, fungi, and thermophilic actinobacteria samples collected at Jennings House, Ohio University. Louisville, KY: University of Louisville, School of Medicine, Department of Microbiology and Immunology.
29. NIOSH [1984]. NIOSH manual of analytical methods. Vol. 3. Cincinnati, OH: National Institute for Occupational Safety and Health, DHHS (NIOSH) Publication No. 84-100.

IX. AUTHORSHIP AND ACKNOWLEDGEMENTS

Report Prepared By:	Kenneth F. Martinez, M.S.E.E. Industrial Hygiene Engineer Industrial Hygiene Section
	Calvin K. Cook Industrial Hygienist Industrial Hygiene Section
	Scott Deitchman, M.D. Occupational Physician Medical Section
Analytical Support:	Richard D. Miller, Ph.D. University Microbiological Associates, Inc. Louisville, Kentucky
	Datachem Laboratories Cincinnati, Ohio

Report Formatted By: Donna M. Humphries
Office Automation Assistant

Originating Office: Hazard Evaluation and Technical
Assistance Branch
Division of Surveillance, Hazard
Evaluations, and Field Studies

X. DISTRIBUTION AND AVAILABILITY OF REPORT

Copies of this report may be freely reproduced and are not copyrighted. A limited number of copies of this report will be available for a period of 90 days from the date of this report from the NIOSH Publications Office, 4676 Columbia Parkway, Cincinnati, Ohio 45226. To expedite your request, include a self-addressed mailing label along with your request. After this time, copies may be purchased from the National Technical Information Service (NTIS), 5285 Port Royal Road, Springfield, Virginia 22161. Information regarding the NTIS stock number may be obtained from the NIOSH Publications Office at the Cincinnati address.

Copies of this report have been sent to:

1. Department of Environmental Health & Safety, Ohio University
2. OSHA Region V

For the purpose of informing affected employees, copies of this report shall be posted by the employer in a prominent place accessible to the employees for a period of 30 calendar days.

APPENDIX

Individual Results for Temperature, Humidity, and CO₂

LOCATION	TIME	TEMP (°F)	% HUMIDITY	CO ₂ (ppm)
Basement	07:34 am	68	39	475
Basement	09:09 am	68	39	400
Basement	11:14 am	71	40	425
Basement	02:32 pm	71	39	500
Non-complaint Office	07:29 am	71	31	450
Non-complaint Office	09:04 am	70	34	500
Non-complaint Office	11:09 am	70	36	450
Non-complaint Office	02:35 pm	73	37	525
Complaint Office	07:22 am	69	35	475
Complaint Office	08:58 am	72	32	475
Complaint Office	11:05 am	72	32	450
Complaint Office	02:27 pm	74	34	475
Temporary Office	09:35 am	83	24	400
Temporary Office	11:23 am	80	28	425
Temporary Office	02:43 pm	80	30	350
Kitchen	07:26 am	71	32	525
Kitchen	09:02 am	70	33	450
Kitchen	11:07 am	70	35	450
Kitchen	02:30 pm	73	35	500
Outside	07:18 am	47	70	425
Outside	09:10 am	53	62	350
Outside	11:23 am	71	40	375
Outside	02:38 pm	81	35	350
Reception Desk	07:20 am	65	38	500
Reception Desk	08:56 am	72	32	500
Reception Desk	11:02 am	73	33	625
Reception Desk	02:25 pm	75	34	475
Conference Room	07:30 am	72	31	450
Conference Room	09:06 am	71	33	525
Conference Room	11:10 am	73	33	500
Conference Room	02:36 pm	76	33	475

Individual Results for Bioaerosols

SAMPLE NUMBER	SAMPLE TYPE	SAMPL E LOCATI ON	TIME OF DAY	COMBI NED CONC (CFU/m ³)	PERCENT RESPIRA BLE	GENUS AND/OR SPECIES RANKING	
						TOP	BOTTOM
OU-1B	B	A	08:18	360	28%	Corynbacterium > Micrococcus + Yeasts	Bacillus sp. > Micrococcus sp. + Yeasts
OU-2B	B	A	08:35	152	23%	Staphylococcus sp. + Yeasts	Staphylococcus sp. + Yeasts
OU-3B	B	A	08:50	198	46%	Micrococcus luteus + Yeasts	Micrococcus luteus + Yeasts
OU-5B	B	B	09:26	427	71%	Micrococcus luteus	M. luteus > Staph. hominis
OU-6B	B	B	09:44	173	23%	Cornybacterium sp. + Yeasts	Cornybacterium sp. + Yeasts
OU-7B	B	B	10:01	361	40%	Micrococcus luteus	Micrococcus luteus
OU-9B	B	C	10:44	438	19%	Bacillus sp.	Bacillus sp.
OU-10	B	C	11:00	109	19%	Bacillus sp.	Bacillus sp.
OU-11	B	C	11:25	102	21%	Bacillus insolitus > Bacillus sp.	Bacillus insolitus > Bacillus sp.
OU-1F	F	A	08:18	85	84%	Cladisporium = Penicillium + Yeasts	Penicillium > Aspergillus > Cladisporium
OU-5F	F	A	08:35	85	84%	Cladisporium = Alternaria = Yeasts	Cladisporium > Penicillium > Aspergillus
OU-7F	F	A	08:50	85	95%	Gliocladium	Penicillium > Cladisporium > Aspergillum
OU-4F	F	A	09:08	106	96%	Gliocladium	Gliocladium > Penicillium > Cladisporium
OU-6F	F	B	09:26	176	84%	Yeasts > Alternaria	Penicillium > Cladisporium > Aspergillus >
OU-8F	F	B	09:44	78	91%	Cladisporium = Yeasts	Penicillium > Cladisporium > Aspergillus
OU-9F	F	B	10:01	63	67%	Cladiporium = Yeast = Penicillium = Epicoccum	Penicillium > Cladisporium
OU-10	F	B	10:20	78	91%	Cladisporium = Penicillium	Penicillium > Cladisporium
OU-11	F	C	10:44	706	50%	Alternaria > Cladisporium >> Penicillium	Cladiporium >> Alternaria > Penicillium
OU-12	F	C	11:00	346	56%	Cladiporium > Penicillium > Yeasts > Alternaria >	Cladiporium >> Alternaria >> Penicillium
OU-13	F	C	11:25	141	62%	Cladiporium >> Penicillium > Ulocladium	Cladiporium >> Penicillium
OU-14	F	C	11:38	728	48%	Alternaria > Cladisporium	Cladisporium
OU-4B	T B	A	09:08 am	14	100%		Streptomyces sp.
OU-8B	T B	B	10:20 am	4	0%	Streptomyces sp.	

Sample Type:
 B = Bacteria
 F = Fungi
 T B = Thermophilic Bacteria

Sample Location:
 A = Complaint Office
 B = Conference Room
 C = Outside