## Review



## W in Melminth therapy or elimination: epidemiological, immunological, and clinical considerations

Linda J Wammes, Harriet Mpairwe, Alison M Elliott, Maria Yazdanbakhsh

#### Lancet Infect Dis 2014; 14: 1150-62

Published Online June 27, 2014 http://dx.doi.org/10.1016/ \$1473-3099(14)70771-6

Department of Parasitology, Leiden University Medical Center, Leiden, Netherlands (LI Wammes MD, Prof M Yazdanbakhsh PhD): MRC/Uganda Virus Research Institute, Uganda Research Unit on AIDS, Entebbe, Uganda (H Mpairwe PhD, Prof A M Elliott MD): and London School of Hygiene and Tropical Medicine, London, UK (Prof A M Elliott)

Correspondence to: Prof Maria Yazdanbakhsh. Department of Parasitology, Leiden University Medical Center, Leiden 2300 RC. Netherlands m.yazdanbakhsh@lumc.nl

Deworming is rightly advocated to prevent helminth-induced morbidity. Nevertheless, in affluent countries, the deliberate infection of patients with worms is being explored as a possible treatment for inflammatory diseases. Several clinical trials are currently registered, for example, to assess the safety or efficacy of Trichuris suis ova in allergies, inflammatory bowel diseases, multiple sclerosis, rheumatoid arthritis, psoriasis, and autism, and the Necator americanus larvae for allergic rhinitis, asthma, coeliac disease, and multiple sclerosis. Studies in animals provide strong evidence that helminths can not only downregulate parasite-specific immune responses, but also modulate autoimmune and allergic inflammatory responses and improve metabolic homoeostasis. This finding suggests that deworming could lead to the emergence of inflammatory and metabolic conditions in countries that are not prepared for these new epidemics. Further studies in endemic countries are needed to assess this risk and to enhance understanding of how helminths modulate inflammatory and metabolic pathways. Studies are similarly needed in non-endemic countries to move helminth-related interventions that show promise in animals, and in phase 1 and 2 studies in human beings, into the therapeutic development pipeline.

## Introduction

Parasitic worms have accompanied man throughout history.1 Infections with helminths such as roundworms, hookworms, whipworms, and schistosomes are often asymptomatic; only a few hosts carry high worm burdens and have overt clinical pathology.2 Moreover, mortality due to helminths is rare. These features suggest a long evolutionary coadaptation between these parasites and man. Key to this partnership is the immunological interaction between helminths and their mammalian hosts. Helminths polarise immune responses and modulate regulatory processes, which might account for their long-term survival within a host.3 In the course of the evolutionary relationship, helminths seem to have fundamentally affected the genetic composition of the host.<sup>4</sup> In heavily exposed human populations, helminths seem to have particularly promoted selection for genes that control the expression levels of cytokines. This adaptation might represent an effort to overcome the regulatory responses induced by helminths. In the absence of worms, such an adaptation could be detrimental and predispose individuals to immune-mediated diseases including allergies and autoimmunity.3

In the 20th century, great effort was put into the worldwide control of infectious diseases. However, the decrease in parasitic and other infectious diseases was associated with a substantial increase in prevalence of chronic inflammatory disorders such as asthma, autoimmune diseases (type 1 diabetes, multiple sclerosis), and inflammatory bowel disease.5 Although the prevalence of asthma and allergic disorders seems to have stabilised in developed countries, the prevalence has increased in developing countries.6 These epidemiological results accord with the hygiene hypothesis or derivatives, such as the so-called old friends hypothesis7 and the biodiversity hypothesis,8 which suggest that the removal of the regulatory effects of microorganisms and parasites-from populations genetically adapted to live with them-tends to lead to an imbalance in the immune system and an increase in immune-mediated diseases.9

Consequently, the question arises of whether helminths should be regarded as harmful pathogens or as beneficial commensals. In low-resource settings deworming is advocated to prevent worm-associated morbidity,10 whereas several research groups in high-income countries are investigating the therapeutic potential of worms and their secreted products in the treatment of inflammatory diseases. This paradox needs to be carefully considered because the practical implications are manifold.

In this Review we summarise present knowledge about immunological and metabolic changes associated with chronic helminth infections, the possible consequences of deworming with respect to inflammatory diseases, and the evidence as to whether a controlled use of worms to treat patients is beneficial, with a view to use helminthderived molecules as new therapeutics.

## Polarisation of immune responses: a doubleedged sword

The immune system is equipped with different cell types that recognise and eliminate pathogens. Innate lymphoid cells,11 dendritic cells, and T cells seem key to the control of different classes of incoming pathogens (figure 1). Type-1 immune responses protect against intracellular pathogens, type-2 responses combat helminths and ectoparasites, and type-17 cells seem to be important against extracellular bacteria and fungi.<sup>12</sup> These responses, spearheaded by T-helper cells, can inflict damage to tissues and organs if uncontrolled. Th1 and Th17 cells release pro-inflammatory cytokines that recruit and activate macrophages and neutrophils, which can attack pathogens. However, their inappropriate activation is associated with autoimmune and inflammatory diseases. Th2 cells trigger responses that disable, degrade, and dislodge parasites,3 but an overactivated Th2 immune response can lead to allergic disorders. Therefore, an important component of the immune system is the regulatory network, with regulatory T cells (Tregs), which are capable of controlling activated effector T cells through expression of inhibitory molecules, at the frontline.<sup>13</sup> Moreover, there is evidence for subtypes of antigenpresenting cells—including monocytes, macrophages, dendritic cells, and B cells—that contribute to suppression of the immune system, resolution of inflammation, and tissue repair, and act as rheostats for homoeostasis.<sup>14</sup>

## Helminth-induced immune regulatory network

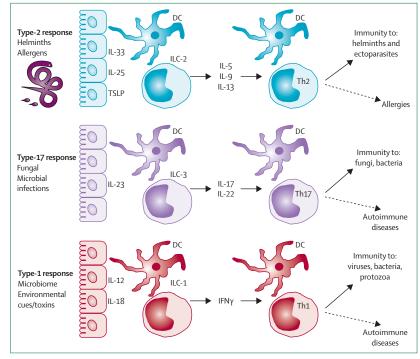
Studies in the 1970s established that, in individuals infected with helminths, the proliferative response of lymphocytes to parasite antigens was lower than in those exposed but not infected.<sup>15,16</sup> These investigations led to the concept that cellular immune hyporesponsiveness, induced by helminths to evade the host immune system,<sup>17</sup> is part of a sophisticated immune regulatory network that operates during helminth infections.18 Non-lymphocytic adherent cells can suppress antifilarial responses in patients with Brugia malayi microfilariae.19 These cells were probably representatives of regulatory antigen-presenting cells: alternatively activated macrophages, suppressory monocytes, or regulatory dendritic cells.<sup>20-22</sup> Subsequently, focus shifted to suppressory CD8+ T cells and then to regulatory CD4+ T cells.<sup>23</sup> Findings from animal models and several cross-sectional studies in human helminthiases have provided supportive evidence for the enhanced number and expanded functional capacity of Tregs during helminth infections.<sup>24,25</sup> Importantly, helminth-derived molecules have been identified that can drive Treg induction in mice.<sup>26,27</sup> Regulatory B cells, defined as B10 cells producing interleukin 10, were also identified during human helminth infections and are associated with the suppression of T-cell responses.28,29 Thus, helminths and their excretory-secretory molecules are endowed with the ability to act through a broad array of cellular mediators to temper host immune responses (figure 2).

On the basis of the ability of helminth infections to induce a regulatory network, helminths might protect against inflammatory diseases.<sup>33</sup> To investigate this hypothesis, human studies have so far included crosssectional association studies, anthelmintic trials, and trials of helminth therapy.

# Associations between helminth infections and inflammatory diseases

## Atopy and allergy-related diseases

Animal models show that helminths might protect against allergic airway inflammation.<sup>34</sup> However, results for the association between worms and allergy-related conditions from cross-sectional studies of people are inconclusive (reviewed by Leonardi-Bee and colleagues,<sup>35</sup> and Flohr and colleagues,<sup>36</sup> and updated in table 1, which does not contain literature previously reviewed). For atopy, assessed as a positive skin-prick test to a panel of allergens, an inverse association with worms has been noted, with occasional

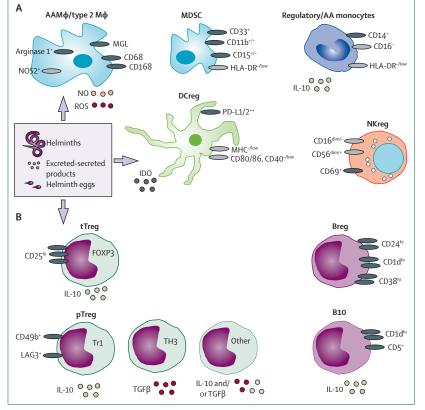


**Figure 1:** Polarisation of T-cell responses to incoming pathogens and environmental factors At mucosal surfaces, epithelial and immune cells detect changes or danger in the environment. Dependent on the nature of the insult, cytokines are produced, which can drive the expansion of group 1, 2, or 3 innate lymphoid cells (ILC-1, ILC-2, and ILC-3) that in turn are associated with the induction of T-helper cells (ILC-1 is associated with Th1, ILC-2 with Th2, and ILC-3 with Th17). The different T-helper cells (comparison of group 1, 2, or 3) the order of the insult invading microorganisms. However, when uncontrolled, similar T-cell responses can lead to pathological conditions (shown by broken arrows). DC=dendritic cell. IL=interlevkin. TSLP=thymic stromal lymphopoietin. IFN y=interferon gamma.

exceptions.<sup>36</sup> Results have been inconsistent for allergyrelated clinical syndromes, such as eczema, wheezing, and asthma. Factors that might affect this relationship are complex and include the timing, burden, or chronicity of helminth infections, helminth species, and host genetics.<sup>52</sup>

#### **Multiple sclerosis**

A 1966 case-control study highlighted the contribution of environmental factors in multiple sclerosis; the presence of piped water, a flush toilet, and sharing a room with one person or more were recorded as environmental factors associated with patients with multiple sclerosis compared with healthy controls.53 Moreover, in an ecological study, country prevalences of multiple sclerosis and Trichuris trichiura infections were almost mutually exclusive.54 Correale and Farez<sup>55</sup> compared helminth-infected and helminth-uninfected patients with multiple sclerosis and showed that new MRI lesions appeared less frequently in infected individuals during 5 years of follow-up. This difference seemed to be associated with an increased production of interleukin 10 and transforming growth factor- $\beta$  by peripheral blood mononuclear cells and the suppressive activity of Tregs.55 Furthermore, when a few patients infected with helminths were treated with anthelmintics because of their intestinal symptoms, the number of clinical relapses and MRI lesions increased, and



#### Figure 2: Helminth-induced regulatory cell network

> regulatory immune responses decreased in parallel.<sup>56</sup> The murine model for multiple sclerosis (experimental autoimmune encephalomyelitis) provided further evidence for a protective effect of helminths or their products<sup>37-60</sup> on the clinical course and inflammation in the CNS.

### See Online for appendix Rheumatoid arthritis

Epidemiological studies in 1975 in South Africa showed that the prevalence of rheumatoid arthritis in urban areas was similar between black Africans and white populations, whereas in rural areas the prevalence was much lower for black Africans.<sup>61</sup> An association with parasitic infections was not proven, but several rodent studies have shown that helminth infections or helminth extracts can suppress or prevent arthritis.<sup>62</sup> A case-control study in India showed that no patient with rheumatoid arthritis harboured circulating microfilariae or filarial antigens, by contrast with 40% of healthy controls.<sup>63</sup>

#### Inflammatory bowel disease

The finding that the prevalence of inflammatory bowel disease was higher in northern than in southern parts of Europe and the USA led to formulation of the inflammatory bowel disease hygiene hypothesis.<sup>64</sup> The few studies of the association between helminths and inflammatory bowel disease have shown conflicting results.<sup>65-67</sup> However, these studies used self-reported helminth infection in early life or immunological markers of exposure to helminths, which are very uncertain measurements, to define helminth exposure. Studies in murine models have consistently shown that infection with worms protects against several forms of experimentally induced colitis.<sup>68</sup>

#### Diabetes

Type 1 diabetes is another autoimmune condition linked to the immune modulatory properties of helminths. In the non-obese diabetes mouse model, infection with several helminth species prevented type 1 diabetes.<sup>69</sup> Data from the Chennai Urban Rural Epidemiology Study (CURES) in India showed that antifilarial IgG4 antibody concentration, as proxy for current filarial infection, was greater in non-diabetics, which suggests that active filarial infection might protect against type 1 diabetes.<sup>70</sup>

Type 2 diabetes is regarded as an inflammatory disease,<sup>71</sup> but has a different pathogenesis and is affected by genetic, nutritional, and other lifestyle factors. In another CURES report, patients with type 2 diabetes had a lower prevalence of lymphatic filariasis than patients without diabetes.<sup>72</sup> Moreover, patients with type 2 diabetes and lymphatic filariasis had lower serum concentrations of pro-inflammatory cytokines than did patients without lymphatic filariasis. This finding suggests that filarial infections in type 2 diabetes can have regulatory functions, although the effect on disease severity was not investigated.

## Longitudinal studies of the consequences of deworming

## Atopy and allergy-related diseases

The effect of worms on allergic conditions has been studied in children from worm-endemic areas with different anthelmintic drugs for various periods of followup. Table 2 shows an overview of placebo-controlled randomised trials and the appendix provides additional details about study populations, interventions, and effect sizes. A cluster-randomised trial in 1632 children from Ecuador that assessed albendazole versus placebo once every 2 months for 1 year showed no effect on skin-prick test responses.75 A randomised trial in 1566 rural children from Vietnam of benzimidazoles versus placebo every 2 months for 12 months showed an increased risk of positive skin-prick test responses to allergens for those receiving benzimidazoles.76 A trial in Indonesia assessed albendazole treatment given every 3 months compared with placebo in 1364 children. A significant increase in

	Helminth species	Number of participants	Age (years)	Outcome	Effect size (CI)	Effect direction
Atopy						
Alcântara-Neves et al 2010, Brazil <sup>37</sup>	Trichuris trichiura; Ascaris lumbricoides; anti-ascaris IgE	283	1-4-2	aslgE	1·52 (0·66–3·50); 0·64 (1·30–1·39); 7·29 (3·90–13·64)	NS; NS; ↑
Calvert et al 2010, South Africa <sup>38</sup>	Ascaris spp	3322	8-12	Skin-prick test	0.63 (0.42-0.94)	↓
Choi et al 2011, Korea <sup>39</sup>	Clonorchis sinensis	1116	30-86	Skin-prick test	1.86 (1.20-2.87)	<b>↑</b>
Djuardi et al 2013, Indonesia40	Wuchereria bancrofti (mother in pregnancy)	126	4	Skin-prick test; aslgE	0-35 (0-07-1-70); 0-43 (0-16-1-15)	NS; NS
Hamid et al 2013, Indonesia41	Hookworm	315	5-15	Skin-prick test	0.46 (0.21–1.00)	↓
Mendonça et al 2012, Brazil <sup>42</sup>	Toxocara seropositivity	1148	4-11	Skin-prick test	0.74 (0.57–0.97)	↓
Moncayo et al 2012, Ecuador <sup>43</sup>	T trichiura; A lumbricoides; anti-ascaris IgE	149 wheezers, 227 controls*	7–19	Skin-prick test; Skin-prick test; aslgE	0·42 (0·17–0·99); 0·82 (0·41–1·64); 5·34 (2·49–11·45)	↓; NS; ↑
Rodrigues et al 2008, Brazil <sup>44</sup>	Ascaris (early childhood); trichuris (early childhood); any worm (early childhood); any worm† (late childhood)	1055	4.4-11.3	Skin-prick test	0·63 (0·44–0·90); 0·58 (0·37–0·92); 0·63 (0·45–0·87); 0·78 (0·57–1·06)	↓; ↓; ↓; NS
Supali et al 2010, Indonesia45	T trichiura; Brugia malayi	422; 574	2–90; 2–90	Skin-prick test	1.48 (0.79–2.76); 0.56 (0.35–0.88)	NS;↓
Wördemann et al 2008, Cuba <sup>46</sup>	A lumbricoides; hookworm; T trichiura; Enterobius vermicularis	1320	4-14	Skin-prick test	No statistically significant associations	NS
Asthma-related symptoms						
Alcântara-Neves et al 2010, Brazil <sup>37</sup>	T trichiura; A lumbricoides	682	1–4·2	Wheeze	2.60 (1.54–4.38); 1.09 (0.71–1.66)	↑; NS
Amberbir et al 2011, Ethiopia <sup>47</sup>	Any worm (hookworm, Ascaris spp, Trichuris spp)	876	3	Wheeze	0.74 (0.29–1.90)	NS
Bager et al 2012, Denmark⁴ <sup>8</sup>	History of treatment for <i>E vermicularis</i> with mebendazole	924749	0-14	Asthma	IRR 1·07 (1·00–1·13)	↑
Mendonça et al 2012, Brazil42	Toxocara seropositivity	1148	4-11	Wheeze and asthma	1.21 (0.92–1.60)	NS
Cobzaru et al 201249	Toxocara seropositivity	76 asthmatics, 88 controls*	5-16	Asthma	13.7 (6.31-29.86)	↑
Choi et al 2011, Korea <sup>39</sup>	C sinensis	1116	30-86	Wheeze; EIB	0·94 (0·54–1·62); 1·30 (0·87–1·92)	NS; NS
Kanobana et al 2013, Cuba⁵º	Toxocara antibodies; hookworm; A lumbricoides	958	5-14	Asthma	1·51 (1·01–2·26); 0·53 (0·32–0·87); 1·29 (0·73–2·28)	↑; ↓; NS
Moncayo et al 2012, Ecuador⁴³	T trichiura; A lumbricoides; anti-ascaris IgE	149 wheezers, 227 controls*	7–19	Wheeze	0·72 (0·44–1·18); 0·90 (0·55–1·48); 2·24 (1·33–3·78)	NS; NS; ↑
Walsh 2011, USA <sup>51</sup>	Toxocara antibodies	12174	17-65	FEV <sub>1</sub>	Adjusted mean difference; –73 mL (–128·1 to –17·9; ie, reduced FEV,, increased bronchoconstriction)	Ť
Wördemann et al 2008, Cuba <sup>46</sup>	A lumbricoides; hookworm; T trichiura; E vermicularis	1320	4-14	Skin-prick test	No statistically significant associations	NS
Eczema						
Amberbir et al 2011, Ethiopia <sup>47</sup>	Any worm (hookworm, Ascaris spp, Trichuris spp)	876	3	Eczema	0·39 (0·09–1·63)	NS
Wördemann et al 2008, Cuba46	A lumbricoides; hookworm; T trichiura; E vermicularis	1320	4–14	Atopic dermatitis	0·23 (0·08–0·68); no statistically significant associations <sup>‡</sup>	↓; NS; NS; NS
Rhinoconjunctivitis						
Amberbir et al 2011, Ethiopia <sup>47</sup>	Any worm (hookworm, Ascaris spp, Trichuris spp)	876	3	Rhinitis	0.49 (0.12–2.09)	NS
Wördemann et al 2008, Cuba46	A lumbricoides; hookworm; T trichiura; E vermicularis	1320	4–14	Rhinoconjunctivitis	No statistically significant associations‡	NS

If not indicated, effect size is odds ratio. aslgE=allergen-specific immunoglobulin E. EIB=exercise-induced bronchospasm. FEV<sub>i</sub>=forced expiratory volume in one s. IRR=incidence rate ratio. NS=not significant.\*Case-control study. †Ascaris spp (current infections) showed a significant inverse association with infection intensity (p=0-008) whereas the inverse trend for T *trichiura* was not significant (p=0-078). ‡Wördemann and colleagues, in their final model, also showed positive associations between rhinoconjunctivitis and history of hookworm (odds ratio 2-81 [95% CI 1-23-6-42]) or E *vermicularis* (odd ratio 1-34 [1-00-1-79]), and between atopic dermatitis and history of E *vermicularis* (1-86 [1-34-2-58]).

Table 1: Completed studies that show associations between atopy or allergy-related diseases and helminths

cockroach reactivity (reactivity to skin-prick tests with cockroach allergens) was noted after 21 months but, overall, skin-prick test responses were not changed.<sup>79</sup> None of these three trials showed any effects of anthelmintic treatment on clinical allergy outcomes. Since clinical allergy is quite rare in these areas, the power of the studies might have been insufficient to detect significant effects. Moreover, differences in species of prevalent helminths, in coprevalence of additional immunomodulating infections,<sup>33,82,83</sup> in exposure to environmental pollutants, or in

	Study design and intervention	Outcomes		
Effects of anthelmintic treatment in healt	hy populations			
Van den Biggelaar et al 2004, Gabon <sup>73</sup>	A placebo-controlled trial of every 3 months praziquantel and mebendazole in school children (aged 5–14 years)	Increased rate of skin-prick test conversion in treated children and a greater decrease in total IgE		
Elliott et al 2005, Uganda <sup>74</sup>	A double-blind, placebo-controlled trial; single-dose albendazole in pregnancy, eczema events in the offspring	A weak effect on infantile eczema $\!\!\!\!^*$ but not statistically significant		
Cooper et al 2006, Ecuador (ISRCTN61195515) <sup>75</sup>	A non-blinded, cluster-randomised trial of albendazole twice every month for 1 year	No statistically significant effect on skin-prick test†, wheeze, rhinoconjunctivitis, eczema, VFD, or EIB		
Flohr et al 2010, Vietnam <sup>76</sup>	A double-blind, placebo-controlled trial among school children; single- dose mebendazole given once then albendazole given daily for 3 consecutive days once every 3 months	Increase in skin-prick test positivity; no statistically significant effect on changes in EIB†, flexural eczema, wheeze, or rhinitis		
Mpairwe et al 2011, Uganda (ISRCTN32849447, EMaBS) <sup>77</sup>	A double-blind, placebo-controlled trial; 2x2 factorial design; single dose albendazole and praziquantel in pregnancy, eczema events in the offspring	Increased rate of eczema among infants of mothers who received albendazole, and among infants of <i>Schistosoma mansoni</i> -infected mothers who received praziquantel		
Ndibazza et al 2012, Uganda (ISRCTN32849447, EmaBS) <sup>78</sup>	Double-blind, placebo-controlled trial of a single dose of albendazole given once every 3 months to pre-school children (age 15 months to 5 years)	No statistically significant effect on clinical eczema events‡		
Wiria et al 2013, Indonesia, (ISRCTN83830814, ImmunoSPIN) <sup>79</sup>	Household-based cluster-randomised, double-blind, placebo-controlled trial	Some evidence of an increase in skin-prick test response to cockroach at 21 months but no overall statistically significant effect on skin-prick test†, specific IgE, wheeze, or atopic dermatitis		
Effects of anthelminthics in asthmatics				
Lynch et al 1997, Venezuela <sup>∞</sup>	A non-blinded, controlled trial of albendazole once a month for 1 year among asthmatics	Statistically significant reduction in asthmatic crises, number of months on maintenance treatment, and salbutamol use in albendazole-treated group compared with previous year. No change in control group		
Almeida et al 2012, Brazil <sup>81</sup>	Double-blind, placebo-controlled trial, single treatment with albendazole and praziguantel among asthmatics	No statistically significant change in asthma severity scores, FEV,, or cytokin responses to house dust mites		

outcome. †Primary outcome. ‡Planned secondary outcome.

Table 2: Trials of anthelmintic treatment that examine the effects on atopy and allergy-related disease outcomes

duration and timing of treatment, could have had a major effect on trial outcomes.

Early-life exposures affect the development of physiological and immunological processes,<sup>84</sup> and therefore the timing of an intervention might be an important element to determine trial outcomes. In support of this notion, a study in Cuba showed that infants growing up in the economic crisis in the 1990s had a reduced risk of developing asthma and rhinoconjunctivitis later in life.85 Moreover, a trial in 2507 pregnant women in Uganda that compared single doses of albendazole and praziquantel versus matching controls (in a 2×2 factorial design) showed that albendazole during pregnancy was associated with an increased risk of eczema in infancy77 and in the first 5 years of life.78 Praziquantel during pregnancy was also associated with an increased risk of eczema in infancy, but only in children whose mothers were infected with Schistosoma mansoni.77 However, albendazole treatment given every 3 months to children was not associated with an increased risk of eczema in early childhood.78 These results suggest that in-utero events might be more important in priming or programming the child's immune system than events in early childhood. Surprisingly, the Ugandan study showed no beneficial effects of anthelmintic treatment during pregnancy on the immune responses to early childhood vaccines, and on none of the anticipated benefits for birthweight,

resistance to infections, or improved child develoment  $^{86,87}$  that would have compensated for the noted adverse effect on eczema.  $^{77,78}$ 

The time needed before an intervention has an effect could be important. A study in Ecuador compared skin-prick test reactivity and allergy-related symptoms in children (aged 6-16 years) from communities that had received 15-17 years of periodic ivermectin treatment with those from adjacent untreated communities.88 The prevalence of skin reactivity to allergens in children from long-term treated communities was double that of children from untreated communities. Children from the treated communities also had a higher prevalence of recent eczema symptoms.<sup>88</sup> Although the study was limited by the fact that the communities were not randomised, the results suggest that long-term interventions against helminths might be needed not only to change responsiveness to allergens or allergyrelated clinical outcomes but also to affect other inflammatory disorders.

The studies described so far investigated cohorts representing the general population. Another approach has been to examine effects of anthelmintic treatment in people who already have an allergy-related disease. In a small trial of individuals aged 5–50 years with a history of asthma in the last 12 months from a schistosomiasis-endemic area in Brazil,<sup>81</sup> study participants received a single dose of albendazole and praziquantel, or placebo.

Investigators recorded no differences in asthma severity between the treatment groups during the 3 month followup period. All participants were then treated with both drugs and worsening of asthma symptoms was recorded at 15 months, but there was no comparison group for this part of the study.<sup>81</sup> Larger trials investigating the effect of the treatment of worms in people with established allergyrelated diseases are warranted.

## Effect on other inflammatory diseases

The effect of deworming on other chronic inflammatory diseases has not been extensively studied, partly because (and inherent to the hypothesis) the prevalence of helminth infections and these conditions show little overlap. Bager and colleagues<sup>48</sup> assessed the effect of anthelmintic treatment retrospectively on chronic inflammatory diseases, in a population cohort in Denmark.48 In this study 14% of more than 900000 children were prescribed mebendazole, for probable Enterobius vermicularis infection (pinworm), a disease that is still endemic in the USA and Europe.<sup>89</sup> Incidence for asthma, type 1 diabetes, juvenile arthritis, and inflammatory bowel disease was not significantly higher in treated children.48 However, the authors suggest that mebendazole was usually prescribed on the basis of symptoms rather than parasitological diagnosis of pinworm infection. Moreover, enterobiasis in these children might not have been sufficiently chronic to induce immune regulation, and the treatment prescribed might have abrogated the possible benefits.

### Immunological consequences

Several studies have assessed immune responses after anthelmintic treatment. Within clinical trials, Th1 and Th2 cytokine production to helminth antigens was enhanced after albendazole treatment in children (average age of 9 years) in Ecuador,<sup>90</sup> and after praziquantel treatment of pregnant women in Uganda.<sup>91</sup> However, the effect of deworming on regulatory responses was not consistent. A decrease in interleukin 10 might be expected after clearance of immunoregulatory helminth infections, but production of S mansoni-specific interleukin 10 was higher in women treated with praziquantel than in those given placebo in the Ugandan study<sup>91</sup> (in line with earlier studies, which were not placebo-controlled<sup>92,93</sup>). By contrast, other studies have shown a decrease in interleukin-10 concentrations after anthelmintic treatment,76,90 which paralleled an increase in allergic reactivity measured by a skin-prick test.76 More advanced statistical methods, such as latent class analysis, might help to understand how complex immune response patterns are associated with disease outcomes; defined immune phenotypes can be incorporated into regression models with other important factors included in these associations, such as environmental conditions.<sup>94</sup> A complicating factor in the study of immune responses after anthelmintic treatment is that dead worms, or products released by dying worms, can stimulate immune reactivity too; thus treatment might boost responses by the release of parasite antigens into the host circulation and removal of parasite-induced immunosuppression.

Helminth treatment might result in an increased T-cell response to some non-helminth antigens,<sup>95,96</sup> but very few studies have investigated allergen-specific cellular immune responses after anthelmintic treatment. The trial in Ecuador recorded no differences in cytokine production in response to cockroach and housemite (*Dermatophagoides pteronyssinus*) antigens after repeated treatment with albendazole.<sup>90</sup> Additional trials are therefore needed to explore this issue.

## Considerations for anthelmintic treatment

In conclusion, although anthelmintic trials are the design of choice to establish cause and effect, or to minimise confounding and the problem of reverse causation, they are based on several assumptions whose validity is unknown. First, the studies assume that the effect of worms is immediately removed after treatment and that development of allergy symptoms follows soon thereafter; however, the protective effects of worms might persist long after anthelmintic treatment. Second, any recorded effect might be due to the anthelmintic drug itself, or from the broader range of effects of the drug, and not because of the elimination of worms. Albendazole binds to tubulin and thereby interferes with the formation of microtubules in the cytoskeleton.<sup>97</sup> As a result, albendazole can affect protozoa,<sup>98,99</sup> fungi,<sup>100</sup> and mammalian cells.<sup>101</sup> In the trial in Uganda, maternal treatment with albendazole was associated with an increased incidence of infantile eczema, even in the children of mothers with no evidence of helminth infection." Therefore, results from anthelmintic trials should be interpreted with caution and it might be helpful to examine effects of a variety of anthelmintic drugs in additional trials.

## Helminth therapy in human beings

A more direct approach, which avoids the pitfalls described, is to study the effects of helminths with live infective stages, or to mimic the effects through helminth-derived molecules. Helminth therapy began in the 1990s with use of *Trichuris suis* ova and later *Necator americanus* larvae.

### **Trichuris infections**

*T* suis is a pig whipworm that colonises the human gut for a short period. *T* suis ova have been particularly studied as a therapy in inflammatory bowel disease.<sup>68</sup> After two open-label trials assessing the safety of *T* suis infection in patients with inflammatory bowel disease showed promising results (about 70% remission in Crohn's disease<sup>102,103</sup>), Summers and colleagues<sup>104</sup> set out to study the effect of *T* suis ova in a first placebocontrolled, double-blind, randomised trial including 54 patients with ulcerative colitis.<sup>104</sup> The ulcerative colitis

disease activity index in the T suis ova group improved significantly compared with the placebo group; however, the number of remissions was not significantly different. Another group characterised the local immune responses surrounding trichuris worms, by studying a patient who self-medicated with T trichiura, the human whipworm.<sup>105</sup> In this patient, during colitis, the T cells producing only interleukin 17 were abundant, whereas after trichuris infection more multifunctional T cells induced, producing cytokines were including interleukin 22.105 In a murine asthma model, treatment with interleukin 22 improved airway constriction and limited airway inflammation.106 In addition to the induction of regulatory cells, trichuris worms seem to modify the cytokine signature of local inflammatory cells. Interleukin 22, together with type-2 cytokines, might contribute to tissue repair and restore gut homoeostasis.107 However, an accumulation of interleukin 17 and interleukin 22 coexpressing cells was associated with colorectal cancer, a result that warrants careful consideration of this molecule.108 In October, 2013, the outcome of a trial undertaken in 250 patients with inflammatory bowel disease showed no strong beneficial effect of T suis ova.<sup>109</sup> The full details of the results are yet to be published.

A safety trial of *T* suis ova in patients with multiple sclerosis was a starting point for a planned phase 2 trial. The trial followed up five patients with relapsing–remitting multiple sclerosis after inoculation with *T* suis ova.<sup>110</sup> Although most patients had mild gastrointestinal symptoms, the number of new lesions shown by MRI was lower during *T* suis ova treatment than before treatment, or after treatment was discontinued. This result was not accompanied by a change in circulating Tregs or alternatively activated monocytes, which suggests that such cells might be recruited in affected tissues and absent in peripheral blood.

A randomised controlled trial of *T* suis ova in 100 patients with allergic rhinitis showed that the therapy induced gastrointestinal symptoms and *T* suis-specific antibody responses without any effect on rhinitis symptom scores, medication use, or skin-prick test reactivity.<sup>111</sup> However, this trial has been criticised because the time between infection with *T* suis ova and the start of the hay fever season might have been too short for sufficient regulatory responses to develop.<sup>112</sup>

As of June, 2014, 18 clinical trials are registered to assess the safety or efficacy of *T suis* ova in allergies, inflammatory bowel diseases, multiple sclerosis, rheumatoid arthritis, psoriasis, and autism (table 3).

	Sponsor	Phase	Status	Condition		
Interventions involving Trichuris suis ova						
EUCTR2007-006099-12-DK	Statens Serum Institut, Denmark	2	Completed	Allergic rhinitis		
NCT01070498	Beth Israel, Boston, USA	1	Completed	Food allergy		
NCT01279577/EUCTR2006-000720-13-DE	Dr Falk Pharma, Frankfurt, Germany	2	Ongoing	Crohn's disease		
ACTRN12608000241336	Asphelia Pharaceuticals, San Diego, USA	1	Not yet recruiting	Crohn's disease		
NCT01434693	Coronado Biosciences, USA	1	Completed	Crohn's disease		
NCT01576471	Coronado Biosciences, USA	2	Ongoing	Crohn's disease		
NCT01433471	New York University, New York, USA	2	Recruiting	Ulcerative colitis		
NCT01953354	NIAID, Bethesda, Maryland, USA	2	Recruiting	Ulcerative colitis		
NCT01413243/EUCTR2009-015319-41-DE	Charité, Berlin, Germany	2	Recruiting	Multiple sclerosis		
NCT00645749	University of Wisconsin, Madison, USA	2	Ongoing	Multiple sclerosis		
NCT01006941	Rigshospitalet, Copenhagen, Denmark	2	Completed	Multiple sclerosis		
EUCTR2011-006344-71-DE	Immanuel Hospital, Berlin, Germany	2	Unknown	Rheumatoid arthritis		
NCT01836939	Mount Sinai School of Medicine, New York, USA	2	Ongoing	Psoriasis		
NCT01948271	Tufts Medical Center, Boston, USA	1	Recruiting	Psoriasis		
NCT02011269	Coronado Biosciences, USA	2	Not yet recruiting	Psoriasis		
NCT01040221	Montefiore, New York, USA	1	Recruiting	Autism		
NCT01734941	Hadassah Medical Organization, Jerusalem, Israel	2	Recruiting	Autism		
NCT02140112	Coronado Biosciences, USA	2	Recruiting	Autism		
Hookworm larvae intervention						
NCT00232518	University of Nottingham, UK	2	Completed	Allergic rhinoconjunctivitis		
NCT00469989	University of Nottingham, UK	2	Completed	Asthma		
NCT00671138	Princess Alexandra Hospital, Brisbane, Australia	2	Unknown	Coeliac disease		
NCT01661933	Princess Alexandra Hospital, Brisbane, Australia	1&2	Recruiting	Coeliac disease		
NCT01470521/EUCTR2008-005008-24-GB	University of Nottingham, UK	2	Recruiting	Multiple sclerosis		
NIAID=National Institute of Allergy and Infectious Diseases. 						

#### Necator americanus larvae

Live Namericanus larvae (human hookworm) can establish long-term infections in man (with longevity of several years). The safety of N americanus larvae was assessed in a dose-ranging study in the UK. Inoculation with 50 larvae or more resulted in substantial gastrointestinal symptoms.113,114 However, inoculation with ten larvae was sufficient to achieve an infection intensity equivalent to 50 eggs per gram of faeces in healthy volunteers and induced a modest immunological response, as measured by eosinophil counts, IgE concentrations, and hookwormspecific IgG concentrations.114

Further safety studies showed that in patients with allergic rhinitis, the lung passage of hookworm larvae did not cause deterioration in airway reactivity<sup>115</sup> and that hookworm-induced type-2 responses did not potentiate an allergen-specific IgE response.<sup>116</sup> The randomised controlled trial of N americanus infection in patients with asthma, with ten larvae, did not show any beneficial effects against asthma symptoms.117 A randomised controlled trial of N americanus infection in patients with coeliac disease showed no effect on the clinical response to wheat challenge.118 However, these trials were undertaken with a maximum of only 16 volunteers in each group. The immunological responses in peripheral blood and the mucosal tissue were investigated after hookworm therapy. In participants infected with hookworm, unstimulated mucosal cells produced less interferon-y and interleukin 17 than in uninfected participants, whereas cells stimulated with the wheat protein gliadin showed a greater production of Th2 cytokine.<sup>119</sup> When hookworm-specific responses were assessed, peripheral blood mononuclear cells and mucosal cells produced higher concentrations of Th2 cytokines in hookworm-infected participants.<sup>120</sup> In parallel, a strong downregulation of interleukin 23, thought to originate from an innate cell source, was reported that could account for the suppression of Th17 responses noted in the earlier study.<sup>119</sup> These data show that helminth therapy might be able to change local and systemic immune responses. However, many more and larger studies are needed, and to be completed in a standardised manner, to firmly establish the extent of immune modulation that is achievable and its potential to change clinical outcomes. Five clinical trials are registered to use N americanus larvae for allergic rhinitis, asthma, coeliac disease, and multiple sclerosis (table 3).

#### Challenges in helminth immunotherapy

Helminthic therapy has some drawbacks. *N americanus*, and possibly T suis, could have pathogenic effects in people, particularly at high doses. Patients undergoing helminth infection should be monitored closely for infection intensity and for possible extraintestinal manifestations of the infection.<sup>121</sup> The long-term results of helminth immunotherapy have not yet been assessed; 24 weeks has been the longest follow-up time for clinical, parasitological, and immunological outcomes. The question remains whether these parameters would change after a longer period of time. The advantage of the introduction of hookworm infections is that only occasional inoculations would be needed, whereas T suis ova would need to be given every 2-3 weeks because it is not a natural human infection. However, this implies that hookworm infections are less controllable because they lead to chronic infestations. The timing of infection is another issue since evidence suggests that the protective immune modulatory effects might be most effectively established in early life.78,84 Moreover, the full development of immunomodulatory effects might take years. In this respect, introduction of parasites as a preventive measure in early life would be the most effective way to control inflammatory diseases.

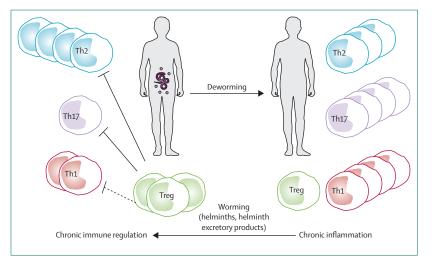
Although immune regulatory responses are desired to counteract inflammatory disorders, they could be detrimental for other immune-associated conditions; defence against incoming pathogens might be impaired, and anti-tumour immune responses could be compromised. Efforts by the scientific community are being made to inhibit Tregs in cancer by immunotherapy.122 Moreover, immunosuppressive agents, such as glucocorticoids, and the increasingly prescribed tumour necrosis factor (TNF) inhibitors for inflammatory bowel disease and rheumatoid arthritis, are associated with an increased risk of (myco)bacterial and some viral infections, such as herpes zoster.<sup>123,124</sup> However, helminthinduced immunoregulation might be more selective than present forms of immunotherapy. For example, evidence shows that B cells can escape Treg control when toll-like receptors (TLR) 4 and TLR 9 are triggered, which happens during viral and bacterial infections.125 Further studies of the effect of helminth co-infection on susceptibility to other infections and to cancer, which can be undertaken in endemic settings, will be helpful, alongside helminth therapy trials in non-endemic settings, to assess these aspects of the safety of helminth therapy.

#### Helminth-derived molecules

Because helminth infections have clinical and pathological results, focus is shifting towards helminth-derived molecules to substitute treatment using whole parasites.<sup>126</sup> Several helminthic products with immune-modulating properties have been defined.<sup>127</sup> Although studies in animal models have shown promise, no helminth-derived molecule has been given to human beings. The filarialderived glycoprotein ES-62 is the best characterised candidate molecule for therapeutic trials. This phosphorylcholine-coupled glycoprotein (first described in 1989128) has beneficial effects in a mouse model of arthritis, 129 and reduces the production of pro-inflammatory cytokines in synovial cells from patients with rheumatoid arthritis.<sup>129</sup> Additionally, filarial-derived glycoprotein ES-62 inhibits mast cell histamine release, which shows that it might protect against allergic diseases.130

Heligmosomoides polygyrus excreted-secreted products suppress murine allergic airway inflammation.<sup>131</sup> AvCystatin, a molecule secreted by *Acanthocheilonema viteae*, inhibits the development of allergic airway inflammation and acute colitis in mice.<sup>132</sup> In-vitro Th2 responses of peripheral blood mononuclear cells in patients allergic to grass pollen are substantially reduced by the addition of AvCystatin to cultures.<sup>133</sup> Furthermore, although less well characterised, soluble products from *S mansoni*, *T suis*, and *Trichinella spiralis* can suppress clinical signs of murine experimental autoimmune encephalomyelitis by modulation of dendritic cells.<sup>134</sup>

Extracts from S mansoni adult worms and excretedsecreted products of the canine hookworm Ankylostoma caninum have shown beneficial effects in murine models of colitis.135 Although treatment with S mansoni extracts did not improve the clinical score of colitis, it diminished local inflammation and myeloid cell infiltration in colon tissue. In parallel, lower Th1 and Th17 responses and enhanced expressions of interleukin 10 and transforming growth factor-β in T cells were recorded in local tissues.135 These results show that changed immune responses do not always lead to clinical improvements, and a longer follow-up might be needed for a clinical improvement to be detectable. Finally, lacto-N-fucopentaose III (LNFPIII; a LewisX-containing glycan that is found in S mansoni eggs) suppresses experimental autoimmune encephalomyelitis by enhancement of interleukin 10 and Th2 cytokines,<sup>136</sup> and improves psoriasis by reducing interferon-γ production in the skin.137 Taken together, these encouraging results from animal models warrant further studies and possible clinical trials, to assess their beneficial effects.



#### Figure 3: Immunological effects of deworming and worming

Helminth-infected individuals (left panel) express enhanced Th2 responses but these are kept under control by the increased number or functional capacity of Tregs. After deworming (right panel), removal of immune suppression could lead to overt inflammation, which is characteristic of several inflammatory diseases; Th1, Th2, or Th17 responses are more active as Tregs decrease in number or function. Treatment with experimental helminth infection or with helminth-derived immunomodulatory molecules could restore the immune regulation noted during natural chronic helminth infections. Th=T-helper cells. Tregs=T-regulatory cells.

## An emerging frontier: immunometabolism and helminth infections

Immunometabolism is an emerging specialty, which investigates the interaction between nutrients, metabolism, and the immune system.138 Macrophages might have a central role in the crosstalk between the immune system and organs controlling whole-body energy metabolism. Classically activated macrophages in adipose tissue can produce pro-inflammatory cytokines, such as TNF, which interfere with the insulin-signalling pathway and lead to the development of insulin resistance, whereas alternatively activated macrophages improve insulin sensitivity.<sup>139</sup> Interleukin 4, a key Th2 cytokine implicated in helminth immunity, plays an important part in the development and maintenance of alternatively activated macrophages in adipose tissue140 and in the control of peripheral insulin sensitivity.141 Helminth infections might therefore have beneficial effects on metabolic disorders. First, worms, as multicellular complex organisms, use host nutrients for their survival; and second, they are the strongest natural stimuli for type-2 immune responses, which was confirmed by the identification of several helminth-derived molecules that can skew immune responses towards Th2 cells.142,143 A study in mice by Wu and colleagues<sup>140</sup> showed that eosinophils were a key source of alternatively activated macrophage-inducing interleukin 4 in adipose tissue, and that helminthinduced eosinophilia resulted in alternatively activated macrophage induction and a sustained improvement in glucose tolerance.<sup>140</sup> Further, LNFPIII, an immunomodulatory glycan<sup>144</sup> present in human milk<sup>145</sup> and on S mansoni eggs, improves whole-body glucose tolerance in high fat diet-induced obese mice through the restoration of insulin sensitivity in white adipose tissue, partly through increased interleukin-10 production by macrophages and dendritic cells.<sup>146</sup> These studies emphasise the possible metabolic advantages of harbouring helminths that, if supported in people, would open new possibilities for helminths and helminthderived molecules as therapeutics to control a group of diseases that are major causes of morbidity worldwide.

### Search strategy and selection criteria

We searched PubMed for articles in English published from Jan 1, 1950, to Feb 20, 2014, with the search terms: "helminth", "worm", "immunology", "allergy", "asthma", "atopy", "inflammatory diseases", "multiple sclerosis", "inflammatory bowel diseases", "rheumatoid arthritis", "diabetes", "randomised clinical trial", and "helminthic therapy". Animal studies were excluded, except for those that were in-vivo studies of helminth infections or helminthderived molecules and inflammatory diseases. We included all helminth therapy trials but excluded deworming trials that were not placebo-controlled.

### **Conclusions and future perspectives**

In summary, a paradox exists between efforts to deworm populations with helminth-associated morbidities, and initiatives to test helminthic therapy on patients with hyperinflammatory diseases (figure 3). Large-scale deworming activities are mainly implemented in the tropics and subtropics, whereas helminthic therapy trials in patients with hyperinflammatory diseases are done in affluent countries.

Murine models support the hypothesis that helminths or their products could be beneficial for inflammatory conditions. Human studies in poor-resource settings have been less consistent, which could be accounted for by the presence of other modulating infections and because the clinical effects of deworming might take time to establish. Further deworming trials should take these issues into account and plan for longer follow-up periods. Mass deworming programmes are advocated,147 which creates opportunities to investigate prospectively whether deworming leads to an increased prevalence of allergic and other inflammatory diseases. Well designed trials, in the context of large-scale deworming programmes, would allow the (as-yet uncertain) benefits of this intervention in human populations to be assessed,148,149 and weighed against potential adverse effects on inflammatory and metabolic disease risks. Human trials of the therapeutic use of helminths in resource-rich settings might elucidate the role of helminths in human physiology, metabolism, and immunology. Although some positive results have so far been reported in inflammatory bowel disease and multiple sclerosis, not much benefit has been seen in the treatment of asthma and allergies. Further trials need to be less modest in the number of patients included, the duration of helminth infection, and, if safety data allow, the dose of infection used, while accounting for the possibility of accumulating low-dose infections.

#### Contributors

LJW did the literature searches, and drafted and finalised the manuscript and the figures; HM assisted with the literature search and drafting of the manuscript and tables; and AME and MY developed the concept, inputted into the literature search, and reviewed the content.

#### **Declaration of interests**

AME reports grants from Wellcome Trust and from the European Union. LJW, HM, and MY declare no competing interests.

#### Acknowledgments

We thank Bruno Guigas for advice on topics surrounding immune metabolism and Katja Polman for reading the manuscript critically.

#### References

- Hoeppli R. The knowledge of parasites and parasitic infections from ancient times to the 17th century. *Exp Parasitol* 1956; 5: 398–419.
- 2 Bundy DAP, Medley GF. Immuno-epidemiology of human geohelminthiasis: ecological and immunological determinants of worm burden. *Parasitology* 1992; **104** (suppl): S105–19.
- 3 Allen JE, Maizels RM. Diversity and dialogue in immunity to helminths. *Nat Rev Immunol* 2011; **11**: 375–88.
- 4 Fumagalli M, Pozzoli U, Cagliani R, et al. Parasites represent a major selective force for interleukin genes and shape the genetic predisposition to autoimmune conditions. J Exp Med 2009; 206: 1395–408.

- 5 Bach JF. The effect of infections on susceptibility to autoimmune and allergic diseases. N Engl J Med 2002; 347: 911–20.
- 6 Pearce N, Aït-Khaled N, Beasley R, et al, and the ISAAC Phase Three Study Group. Worldwide trends in the prevalence of asthma symptoms: phase III of the international study of asthma and allergies in childhood (ISAAC). *Thorax* 2007; **62**: 758–66.
- 7 Rook GA. 99th Dahlem conference on infection, inflammation and chronic inflammatory disorders: darwinian medicine and the 'hygiene' or 'old friends' hypothesis. *Clin Exp Immunol* 2010; 160: 70–79.
- 8 Hanski I, von Hertzen L, Fyhrquist N, et al. Environmental biodiversity, human microbiota, and allergy are interrelated. *Proc Natl Acad Sci USA* 2012; 109: 8334–39.
- 9 Strachan DP. Hay fever, hygiene, and household size. BMJ 1989; 299: 1259–60.
- 10 WHO. Preventive chemotherapy in human helminthiasis: coordinated use of drugs in control interventions: a manual for health professionals and programme managers. WHO: Geneva, 2006. http://whqlibdoc.who.int/publications/2006/9241547103\_eng. pdf (accessed Feb 19, 2014).
- 11 Spits H, Artis D, Colonna M, et al. Innate lymphoid cells—a proposal for uniform nomenclature. *Nat Rev Immunol* 2013; 13: 145–49.
- 12 Zhu J, Paul WE. CD4 T cells: fates, functions, and faults. Blood 2008; 112: 1557–69.
- 13 Belkaid Y. Regulatory T cells and infection: a dangerous necessity. Nat Rev Immunol 2007; 7: 875–88.
- 14 Germain RN. Maintaining system homeostasis: the third law of Newtonian immunology. Nat Immunol 2012; 13: 902–06.
- 15 Ottesen EA, Hiatt RA, Cheever AW, Sotomayor ZR, Neva FA. The acquisition and loss of antigen-specific cellular immune responsiveness in acute and chronic schistosomiasis in man. *Clin Exp Immunol* 1978; 33: 37–47.
- 16 Ottesen EA, Weller PF, Heck L. Specific cellular immune unresponsiveness in human filariasis. *Immunology* 1977; 33: 413–21.
- 17 Maizels RM, Bundy DAP, Selkirk ME, Smith DF, Anderson RM. Immunological modulation and evasion by helminth parasites in human populations. *Nature* 1993; 365: 797–805.
- 18 Maizels RM, Yazdanbakhsh M. Immune regulation by helminth parasites: cellular and molecular mechanisms. *Nat Rev Immunol* 2003; 3: 733–44.
- 19 Piessens WF, Ratiwayanto S, Tuti S, et al. Antigen-specific suppressor cells and suppressor factors in human filariasis with Brugia malayi. N Engl J Med 1980; 302: 833–37.
- 20 Babu S, Kumaraswami V, Nutman TB. Alternatively activated and immunoregulatory monocytes in human filarial infections. *J Infect Dis* 2009; **199:** 1827–37.
- 21 Li Z, Liu G, Chen Y, Liu Y, Liu B, Su Z. The phenotype and function of naturally existing regulatory dendritic cells in nematode-infected mice. *Int J Parasitol* 2011; 41: 1129–37.
- 22 Van Dyken SJ, Locksley RM. Interleukin-4- and interleukin-13mediated alternatively activated macrophages: roles in homeostasis and disease. Annu Rev Immunol 2013; 31: 317–43.
- 23 MacDonald TT. Suppressor T cells, rebranded as regulatory T cells, emerge from the wilderness bearing surface markers. *Gut* 2002; 51: 311–12.
- 24 Maizels RM, Smith KA. Regulatory T cells in infection. *Adv Immunol* 2011; **112**: 73–136.
- 25 Metenou S, Nutman TB. Regulatory T cell subsets in filarial infection and their function. *Front Immunol* 2013; 4: 305.
- 26 Grainger JR, Smith KA, Hewitson JP, et al. Helminth secretions induce de novo T cell Foxp3 expression and regulatory function through the TGF-β pathway. J Exp Med 2010; 207: 2331–41.
- 27 Zaccone P, Burton OT, Gibbs SE, et al. The S mansoni glycoprotein w-1 induces Foxp3 expression in NOD mouse CD4<sup>+</sup> T cells. Eur [ Immunol 2011; 41: 2709–18.
- 28 Correale J, Farez M, Razzitte G. Helminth infections associated with multiple sclerosis induce regulatory B cells. *Ann Neurol* 2008; 64: 187–99.
- 29 Van der Vlugt LE, Labuda LA, Ozir-Fazalalikhan A, et al. Schistosomes induce regulatory features in human and mouse CD1d(hi) B cells: inhibition of allergic inflammation by IL-10 and regulatory T cells. *PLoS One* 2012; 7: e30883.

- 30 Gabrilovich DI, Nagaraj S. Myeloid-derived suppressor cells as regulators of the immune system. Nat Rev Immunol 2009; 9: 162–74.
- 31 Jewett A, Man YG, Tseng HC. Dual functions of natural killer cells in selection and differentiation of stem cells; role in regulation of inflammation and regeneration of tissues. J Cancer 2013; 4: 12–24.
- 32 Gagliani N, Magnani CF, Huber S, et al. Coexpression of CD49b and LAG-3 identifies human and mouse T regulatory type 1 cells. *Nat Med* 2013; 19: 739–46.
- 33 Yazdanbakhsh M, Kremsner PG, Van Ree R. Allergy, parasites, and the hygiene hypothesis. *Science* 2002; **296**: 490–94.
- 34 Smits HH, Everts B, Hartgers FC, Yazdanbakhsh M. Chronic helminth infections protect against allergic diseases by active regulatory processes. *Curr Allergy Asthma Rep* 2010; 10: 3–12.
- 35 Leonardi-Bee J, Pritchard D, Britton J. Asthma and current intestinal parasite infection: systematic review and meta-analysis. *Am J Respir Crit Care Med* 2006; **174**: 514–23.
- 36 Flohr C, Quinnell RJ, Britton J. Do helminth parasites protect against atopy and allergic disease? *Clin Exp Allergy* 2009; **39**: 20–32.
- 37 Alcântara-Neves NM, Badaró SJ, dos Santos MC, Pontes-de-Carvalho L, Barreto ML. The presence of serum anti-Ascaris lumbricoides IgE antibodies and of Trichuris trichiura infection are risk factors for wheezing and/or atopy in preschoolaged Brazilian children. Respir Res 2010; 11: 114.
- 38 Calvert J, Burney P. Ascaris, atopy, and exercise-induced bronchoconstriction in rural and urban South African children. J Allergy Clin Immunol 2010; 125: 100–05.
- 39 Choi MH, Chang YS, Lim MK, et al. *Clonorchis sinensis* infection is positively associated with atopy in endemic area. *Clin Exp Allergy* 2011; 41: 697–705.
- 40 Djuardi Y, Supali T, Wibowo H, et al. The development of TH2 responses from infancy to 4 years of age and atopic sensitization in areas endemic for helminth infections. *Allergy Asthma Clin Immunol* 2013; **9**: 13.
- 41 Hamid F, Wiria AE, Wammes LJ, et al. Risk factors associated with the development of atopic sensitization in Indonesia. *PLoS One* 2013; 8: e67064.
- 42 Mendonça LR, Veiga RV, Dattoli VC, et al. Toxocara seropositivity, atopy and wheezing in children living in poor neighbourhoods in urban Latin American. *PLoS Negl Trop Dis* 2012; **6**: e1886.
- 43 Moncayo AL, Vaca M, Oviedo G, et al. Effects of geohelminth infection and age on the associations between allergen-specific IgE, skin test reactivity and wheeze: a case-control study. *Clin Exp Allergy* 2013; 43: 60–72.
- 44 Rodrigues LC, Newcombe PJ, Cunha SS, et al, and the Social Change, Asthma and Allergy in Latin America. Early infection with *Trichuris trichiura* and allergen skin test reactivity in later childhood. *Clin Exp Allergy* 2008; 38: 1769–77.
- 45 Supali T, Djuardi Y, Wibowo H, Van Ree R, Yazdanbakhsh M, Sartono E. Relationship between different species of helminths and atopy: a study in a population living in helminth-endemic area in Sulawesi, Indonesia. Int Arch Allergy Immunol 2010; 153: 388–94.
- 46 Wördemann M, Diaz RJ, Heredia LM, et al. Association of atopy, asthma, allergic rhinoconjunctivitis, atopic dermatitis and intestinal helminth infections in Cuban children. *Trop Med Int Health* 2008; 13: 180–86.
- 47 Amberbir A, Medhin G, Erku W, et al. Effects of *Helicobacter pylori*, geohelminth infection and selected commensal bacteria on the risk of allergic disease and sensitization in 3-year-old Ethiopian children. *Clin Exp Allergy* 2011; **41**: 1422–30.
- 48 Bager P, Vinkel Hansen A, Wohlfahrt J, Melbye M. Helminth infection does not reduce risk for chronic inflammatory disease in a population-based cohort study. *Gastroenterology* 2012; 142: 55–62.
- 49 Cobzaru RG, Rîpă C, Leon MM, Luca MC, Ivan A, Luca M. Correlation between asthma and *Toxocara canis* infection. *Rev Med Chir Soc Med Nat Iasi* 2012; 116: 727–30.
- 50 Kanobana K, Vereecken K, Junco Diaz R, et al. Toxocara seropositivity, atopy and asthma: a study in Cuban schoolchildren. *Trop Med Int Health* 2013; 18: 403–06.
- 51 Walsh MG. Toxocara infection and diminished lung function in a nationally representative sample from the United States population. *Int J Parasitol* 2011; 41: 243–47.
- 52 Cooper PJ. Interactions between helminth parasites and allergy. Curr Opin Allergy Clin Immunol 2009; 9: 29–37.

- 53 Leibowitz U, Antonovsky A, Medalie JM, Smith HA, Halpern L, Alter M. Epidemiological study of multiple sclerosis in Israel. II. Multiple sclerosis and level of sanitation. *J Neurol Neurosurg Psychiatry* 1966; 29: 60–68.
- 54 Fleming JO, Cook TD. Multiple sclerosis and the hygiene hypothesis. *Neurology* 2006; 67: 2085–86.
- 55 Correale J, Farez M. Association between parasite infection and immune responses in multiple sclerosis. Ann Neurol 2007; 61: 97–108.
- 56 Correale J, Farez MF. The impact of parasite infections on the course of multiple sclerosis. J Neuroimmunol 2011; 233: 6–11.
- 57 La Flamme AC, Ruddenklau K, Bäckström BT. Schistosomiasis decreases central nervous system inflammation and alters the progression of experimental autoimmune encephalomyelitis. *Infect Immun* 2003; 71: 4996–5004.
- 58 Sewell D, Qing Z, Reinke E, et al. Immunomodulation of experimental autoimmune encephalomyelitis by helminth ova immunization. *Int Immunol* 2003; 15: 59–69.
- 59 Walsh KP, Brady MT, Finlay CM, Boon L, Mills KH. Infection with a helminth parasite attenuates autoimmunity through TGF-betamediated suppression of Th17 and Th1 responses. *J Immunol* 2009; 183: 1577–86.
- 60 Donskow-Łysoniewska K, Krawczak K, Doligalska M. Heligmosomoides polygyrus: EAE remission is correlated with different systemic cytokine profiles provoked by L4 and adult nematodes. Exp Parasitol 2012; 132: 243–48.
- Solomon L, Robin G, Valkenburg HA. Rheumatoid arthritis in an urban South African Negro population. Ann Rheum Dis 1975; 34: 128–35.
- 62 Matisz CE, McDougall JJ, Sharkey KA, McKay DM. Helminth parasites and the modulation of joint inflammation. J Parasitol Res 2011; 2011: 942616.
- 63 Panda AK, Ravindran B, Das BK. Rheumatoid arthritis patients are free of filarial infection in an area where filariasis is endemic: comment on the article by Pineda et al. *Arthritis Rheum* 2013; 65: 1402–03.
- 64 Elliott DE, Urban JF Jr, Argo CK, Weinstock JV. Does the failure to acquire helminthic parasites predispose to Crohn's disease? FASEB J 2000; 14: 1848–55.
- 55 Kabeerdoss J, Pugazhendhi S, Subramanian V, Binder HJ, Ramakrishna BS. Exposure to hookworms in patients with Crohn's disease: a case-control study. *Aliment Pharmacol Ther* 2011; 34: 923–30.
- 66 Castiglione F, Diaferia M, Morace F, et al. Risk factors for inflammatory bowel diseases according to the "hygiene hypothesis": a case-control, multi-centre, prospective study in Southern Italy. *J Crohn's Colitis* 2012; 6: 324–29.
- 67 Chu KM, Watermeyer G, Shelly L, et al. Childhood helminth exposure is protective against inflammatory bowel disease: a case control study in South Africa. *Inflamm Bowel Dis* 2013; 19: 614–20.
- 68 Weinstock JV, Elliott DE. Translatability of helminth therapy in inflammatory bowel diseases. Int J Parasitol 2013; 43: 245–51.
- 69 Zaccone P, Cooke A. Helminth mediated modulation of type 1 diabetes (T1D). Int J Parasitol 2013; 43: 311–18.
- 70 Aravindhan V, Mohan V, Surendar J, et al. Decreased prevalence of lymphatic filariasis among subjects with type-1 diabetes. *Am J Trop Med Hyg* 2010; 83: 1336–39.
- 71 Donath MY, Shoelson SE. Type 2 diabetes as an inflammatory disease. *Nat Rev Immunol* 2011; **11**: 98–107.
- 72 Aravindhan V, Mohan V, Surendar J, et al. Decreased prevalence of lymphatic filariasis among diabetic subjects associated with a diminished pro-inflammatory cytokine response (CURES 83). *PLoS Negl Trop Dis* 2010; 4: e707.
- 73 Van den Biggelaar AH, Rodrigues LC, Van Ree R, et al. Long-term treatment of intestinal helminths increases mite skin-test reactivity in Gabonese schoolchildren. *J Infect Dis* 2004; 189: 892–900.
- 74 Elliott AM, Mpairwe H, Quigley MA, et al. Helminth infection during pregnancy and development of infantile eczema. JAMA 2005; 294: 2032–34.
- 75 Cooper PJ, Chico ME, Vaca MG, et al. Effect of albendazole treatments on the prevalence of atopy in children living in communities endemic for geohelminth parasites: a clusterrandomised trial. *Lancet* 2006; 367: 1598–603.

- 76 Flohr C, Tuyen LN, Quinnell RJ, et al. Reduced helminth burden increases allergen skin sensitization but not clinical allergy: a randomized, double-blind, placebo-controlled trial in Vietnam. *Clin Exp Allergy* 2010; **40**: 131–42.
- 77 Mpairwe H, Webb EL, Muhangi L, et al. Anthelminthic treatment during pregnancy is associated with increased risk of infantile eczema: randomised-controlled trial results. *Pediatr Allergy Immunol* 2011; 22: 305–12.
- 78 Ndibazza J, Mpairwe H, Webb EL, et al. Impact of anthelminthic treatment in pregnancy and childhood on immunisations, infections and eczema in childhood: a randomised controlled trial. *PLoS One* 2012; 7: e50325.
- 79 Wiria AE, Hamid F, Wammes LJ, et al. The effect of three-monthly albendazole treatment on malarial parasitemia and allergy: a household-based cluster-randomized, double-blind, placebo-controlled trial. *PLoS One* 2013; 8: e57899.
- 80 Lynch NR, Palenque M, Hagel I, DiPrisco MC. Clinical improvement of asthma after anthelminthic treatment in a tropical situation. *Am J Respir Crit Care Med* 1997; 156: 50–54.
- 81 Almeida MC, Lima GS, Cardoso LS, et al. The effect of antihelminthic treatment on subjects with asthma from an endemic area of schistosomiasis: a randomized, double-blinded, and placebocontrolled trial. J Parasitol Res 2012; 2012: 296856.
- 82 Matricardi PM, Rosmini F, Riondino S, et al. Exposure to foodborne and orofecal microbes versus airborne viruses in relation to atopy and allergic asthma: epidemiological study. *BMJ* 2000; **320**: 412–17.
- 83 Pelosi U, Porcedda G, Tiddia F, et al. The inverse association of salmonellosis in infancy with allergic rhinoconjunctivitis and asthma at school-age: a longitudinal study. *Allergy* 2005; 60: 626–30.
- 84 von Mutius E, Vercelli D. Farm living: effects on childhood asthma and allergy. Nat Rev Immunol 2010; 10: 861–68.
- 85 Van der Werff SD, Polman K, Ponce MC, et al. Childhood atopic diseases and early life circumstances: an ecological study in Cuba. *PLoS One* 2012; 7: e39892.
- 86 Ndibazza J, Muhangi L, Akishule D, et al. Effects of deworming during pregnancy on maternal and perinatal outcomes in Entebbe, Uganda: a randomized controlled trial. *Clin Infect Dis* 2010; 50: 531–40.
- 87 Webb EL, Mawa PA, Ndibazza J, et al. Effect of single-dose anthelminitic treatment during pregnancy on an infant's response to immunisation and on susceptibility to infectious diseases in infancy: a randomised, double-blind, placebo-controlled trial. *Lancet* 2011; 377: 52–62.
- 88 Endara P, Vaca M, Chico ME, et al. Long-term periodic anthelmintic treatments are associated with increased allergen skin reactivity. *Clin Exp Allergy* 2010; 40: 1669–77.
- 89 Burkhart CN, Burkhart CG. Assessment of frequency, transmission, and genitourinary complications of enterobiasis (pinworms). Int | Dermatol 2005; 44: 837–40.
- 90 Cooper PJ, Moncayo AL, Guadalupe I, et al. Repeated treatments with albendazole enhance Th2 responses to Ascaris lumbricoides, but not to aeroallergens, in children from rural communities in the Tropics. J Infect Dis 2008; 198: 1237–42.
- 91 Tweyongyere R, Mawa PA, Ngom-Wegi S, et al. Effect of praziquantel treatment during pregnancy on cytokine responses to schistosome antigens: results of a randomized, placebo-controlled trial. *J Infect Dis* 2008; **198**: 1870–79.
- 92 Van den Biggelaar AH, Borrmann S, Kremsner P, Yazdanbakhsh M. Immune responses induced by repeated treatment do not result in protective immunity to *Schistosoma haematobium*: interleukin (IL)-5 and IL-10 responses. *J Infect Dis* 2002; **186**: 1474–82.
- 93 Wright VJ, Ame SM, Haji HS, et al. Early exposure of infants to GI nematodes induces Th2 dominant immune responses which are unaffected by periodic anthelminthic treatment. *PLoS Negl Trop Dis* 2009; 3: e433.
- 94 Figueiredo CA, Amorim LD, Alcantara-Neves NM, et al. Environmental conditions, immunologic phenotypes, atopy, and asthma: new evidence of how the hygiene hypothesis operates in Latin America. J Allergy Clin Immunol 2013; 131: 1064-68.
- 95 Cooper PJ, Chico M, Sandoval C, et al. Human infection with Ascaris lumbricoides is associated with suppression of the interleukin-2 response to recombinant cholera toxin B subunit following vaccination with the live oral cholera vaccine CVD 103-HgR. Infect Immun 2001; 69: 1574–80.

- 96 Elias D, Wolday D, Akuffo H, Petros B, Bronner U, Britton S. Effect of deworming on human T cell responses to mycobacterial antigens in helminth-exposed individuals before and after bacille Calmette-Guérin (BCG) vaccination. *Clin Exp Immunol* 2001; 123: 219–25.
- 97 Lacey E. Mode of action of benzimidazoles. *Parasitol Today* 1990; 6: 112–15.
- 98 Skinner-Adams TS, Davis TM, Manning LS, Johnston WA. The efficacy of benzimidazole drugs against *Plasmodium falciparum* in vitro. *Trans R Soc Trop Med Hγg* 1997; 91: 580–84.
- 99 MacDonald LM, Armson A, Thompson AR, Reynoldson JA. Characterisation of benzimidazole binding with recombinant tubulin from Giardia duodenalis, Encephalitozoon intestinalis, and Cryptosporidium parvum. Mol Biochem Parasitol 2004; 138: 89–96.
- 100 Cruz MC, Edlind T. beta-Tubulin genes and the basis for benzimidazole sensitivity of the opportunistic fungus *Cryptococcus neoformans. Microbiology* 1997; 143: 2003–08.
- 101 Mizuno K, Toyoda Y, Fukami T, Nakajima M, Yokoi T. Stimulation of pro-inflammatory responses by mebendazole in human monocytic THP-1 cells through an ERK signaling pathway. *Arch Toxicol* 2011; 85: 199–207.
- 102 Summers RW, Elliott DE, Qadir K, Urban JF Jr, Thompson R, Weinstock JV. *Trichuris suis* seems to be safe and possibly effective in the treatment of inflammatory bowel disease. *Am J Gastroenterol* 2003; 98: 2034–41.
- 103 Summers RW, Elliott DE, Urban JF Jr, Thompson R, Weinstock JV. Trichuris suis therapy in Crohn's disease. Gut 2005; 54: 87–90.
- 104 Summers RW, Elliott DE, Urban JF Jr, Thompson RA, Weinstock JV. *Trichuris suis* therapy for active ulcerative colitis: a randomized controlled trial. *Gastroenterology* 2005; 128: 825–32.
- 105 Broadhurst MJ, Leung JM, Kashyap V, et al. IL-22+ CD4+ T cells are associated with therapeutic *Trichuris trichiura* infection in an ulcerative colitis patient. *Sci Transl Med* 2010; 2: 60ra88.
- 106 Taube C, Tertilt C, Gyülveszi G, et al. IL-22 is produced by innate lymphoid cells and limits inflammation in allergic airway disease. *PLoS One* 2011; 6: e21799.
- 107 Allen JE, Wynn TA. Evolution of Th2 immunity: a rapid repair response to tissue destructive pathogens. *PLoS Pathog* 2011; 7: e1002003.
- 108 Kirchberger S, Royston DJ, Boulard O, et al. Innate lymphoid cells sustain colon cancer through production of interleukin-22 in a mouse model. J Exp Med 2013; 210: 917–31.
- 109 Lu L, Nanus M, Forman S. Coronado biosciences announces topline results from its TRUST-I phase 2 clinical trial of TSO for the treatment of Crohn's Disease. Oct 14, 2013. http://globenewswire. com/news-release/2013/10/14/580190/10052399/en/Coronado-Biosciences-Announces-Top-Line-Results-From-Its-TRUST-I-Phase-2-Clinical-Trial-of-TSO-for-the-Treatment-of-Crohn-s-Disease.html (accessed March 1, 2014).
- 110 Fleming JO, Isaak A, Lee JE, et al. Probiotic helminth administration in relapsing-remitting multiple sclerosis: a phase 1 study. *Mult Scler* 2011; 17: 743–54.
- 111 Bager P, Arnved J, Ronborg S, et al. *Trichuris suis* ova therapy for allergic rhinitis: a randomized, double-blind, placebo-controlled clinical trial. J Allergy Clin Immunol 2010; 125: 123–30.
- 112 Summers RW, Elliott DE, Weinstock JV. Trichuris suis might be effective in treating allergic rhinitis. J Allergy Clin Immunol 2010; 125: 766–67.
- 113 Croese J, O'neil J, Masson J, et al. A proof of concept study establishing *Necator americanus* in Crohn's patients and reservoir donors. *Gut* 2006; 55: 136–37.
- 114 Mortimer K, Brown A, Feary J, et al. Dose-ranging study for trials of therapeutic infection with *Necator americanus* in humans. *Am J Trop Med Hyg* 2006; **75**: 914–20.
- 115 Feary J, Venn A, Brown A, et al. Safety of hookworm infection in individuals with measurable airway responsiveness: a randomized placebo-controlled feasibility study. *Clin Exp Allergy* 2009; **39**: 1060–68.
- 116 Blount D, Hooi D, Feary J, et al. Immunologic profiles of persons recruited for a randomized, placebo-controlled clinical trial of hookworm infection. Am J Trop Med Hyg 2009; 81: 911–16.
- 117 Feary JR, Venn AJ, Mortimer K, et al. Experimental hookworm infection: a randomized placebo-controlled trial in asthma. *Clin Exp Allergy* 2010; 40: 299–306.

- 118 Daveson AJ, Jones DM, Gaze S, et al. Effect of hookworm infection on wheat challenge in celiac disease—a randomised double-blinded placebo controlled trial. *PLoS One* 2011; 6: e17366.
- 119 McSorley HJ, Gaze S, Daveson J, et al. Suppression of inflammatory immune responses in celiac disease by experimental hookworm infection. *PLoS One* 2011; 6: e24092.
- 120 Gaze S, McSorley HJ, Daveson J, et al. Characterising the mucosal and systemic immune responses to experimental human hookworm infection. *PLoS Pathog* 2012; 8: e1002520.
- 121 Van Kruiningen HJ, West AB. Potential danger in the medical use of *Trichuris suis* for the treatment of inflammatory bowel disease. *Inflamm Bowel Dis* 2005; **11**: 515.
- 122 Zou W. Regulatory T cells, tumour immunity and immunotherapy. Nat Rev Immunol 2006; 6: 295–307.
- 123 Ford AC, Peyrin-Biroulet L. Opportunistic infections with antitumor necrosis factor-α therapy in inflammatory bowel disease: meta-analysis of randomized controlled trials. Am J Gastroenterol 2013; 108: 1268–76.
- 124 Gardam MA, Keystone EC, Menzies R, et al. Anti-tumour necrosis factor agents and tuberculosis risk: mechanisms of action and clinical management. *Lancet Infect Dis* 2003; **3**: 148–55.
- 125 Adjobimey T, Satoguina J, Oldenburg J, Hoerauf A, Layland LE. Co-activation through TLR4 and TLR9 but not TLR2 skews Tregmediated modulation of Igs and induces IL-17 secretion in Treg: B cell co-cultures. *Innate Immun* 2014; **20**: 12–23.
- 126 Harnett W, Harnett MM. Helminth-derived immunomodulators: can understanding the worm produce the pill? *Nat Rev Immunol* 2010; **10**: 278–84.
- 127 Zaccone P, Cooke A. Vaccine against autoimmune disease: can helminths or their products provide a therapy? *Curr Opin Immunol* 2013; 25: 418–23.
- 128 Harnett W, Worms MJ, Kapil A, Grainger M, Parkhouse RM. Origin, kinetics of circulation and fate in vivo of the major excretory-secretory product of Acanthocheilonema viteae. Parasitology 1989; 99: 229–39.
- 129 McInnes IB, Leung BP, Harnett M, Gracie JA, Liew FY, Harnett W. A novel therapeutic approach targeting articular inflammation using the filarial nematode-derived phosphorylcholine-containing glycoprotein ES-62. J Immunol 2003; 171: 2127–33.
- 130 Melendez AJ, Harnett MM, Pushparaj PN, et al. Inhibition of Fc epsilon RI-mediated mast cell responses by ES-62, a product of parasitic filarial nematodes. *Nat Med* 2007; 13: 1375–81.
- 131 McSorley HJ, O'Gorman MT, Blair N, Sutherland TE, Filbey KJ, Maizels RM. Suppression of type 2 immunity and allergic airway inflammation by secreted products of the helminth *Heligmosomoides polygyrus. Eur J Immunol* 2012; 42: 2667–82.
- 132 Schnoeller C, Rausch S, Pillai S, et al. A helminth immunomodulator reduces allergic and inflammatory responses by induction of IL-10-producing macrophages. *J Immunol* 2008; 180: 4265–72.
- 133 Daniłowicz-Luebert E, Steinfelder S, Kühl AA, et al. A nematode immunomodulator suppresses grass pollen-specific allergic responses by controlling excessive Th2 inflammation. *Int J Parasitol* 2013; 43: 201–10.

- 134 Kuijk LM, Klaver EJ, Kooij G, et al. Soluble helminth products suppress clinical signs in murine experimental autoimmune encephalomyelitis and differentially modulate human dendritic cell activation. *Mol Immunol* 2012; 51: 210–18.
- 135 Ruyssers NE, De Winter BY, De Man JG, et al. Therapeutic potential of helminth soluble proteins in TNBS-induced colitis in mice. *Inflamm Bowel Dis* 2009; 15: 491–500.
- 136 Zhu B, Trikudanathan S, Zozulya AL, et al. Immune modulation by Lacto-N-fucopentaose III in experimental autoimmune encephalomyelitis. *Clin Immunol* 2012; 142: 351–61.
- 137 Atochina O, Harn D. Prevention of psoriasis-like lesions development in fsn/fsn mice by helminth glycans. *Exp Dermatol* 2006; 15: 461–68.
- 138 Pearce EL, Pearce EJ. Metabolic pathways in immune cell activation and quiescence. *Immunity* 2013; **38**: 633–43.
- 139 Olefsky JM, Glass CK. Macrophages, inflammation, and insulin resistance. Annu Rev Physiol 2010; 72: 219–46.
- 140 Wu D, Molofsky AB, Liang HE, et al. Eosinophils sustain adipose alternatively activated macrophages associated with glucose homeostasis. *Science* 2011; 332: 243–47.
- 141 Ricardo-Gonzalez RR, Red Eagle A, Odegaard JI, et al. IL-4/STAT6 immune axis regulates peripheral nutrient metabolism and insulin sensitivity. Proc Natl Acad Sci USA 2010; 107: 22617–22.
- 142 Whelan M, Harnett MM, Houston KM, Patel V, Harnett W, Rigley KP. A filarial nematode-secreted product signals dendritic cells to acquire a phenotype that drives development of Th2 cells. *J Immunol* 2000; **164:** 6453–60.
- 143 Everts B, Perona-Wright G, Smits HH, et al. Omega-1, a glycoprotein secreted by *Schistosoma mansoni* eggs, drives Th2 responses. J Exp Med 2009; 206: 1673–80.
- 144 Atochina O, Da'dara AA, Walker M, Harn DA. The immunomodulatory glycan LNFPIII initiates alternative activation of murine macrophages in vivo. *Immunology* 2008; 125: 111–21.
- 145 Stahl B, Thurl S, Zeng J, et al. Oligosaccharides from human milk as revealed by matrix-assisted laser desorption/ionization mass spectrometry. Anal Biochem 1994; 223: 218–26.
- 146 Bhargava P, Li C, Stanya KJ, et al. Immunomodulatory glycan LNPPIII alleviates hepatosteatosis and insulin resistance through direct and indirect control of metabolic pathways. *Nat Med* 2012; 18: 1665–72.
- 147 Utzinger J. A research and development agenda for the control and elimination of human helminthiases. *PLoS Negl Trop Dis* 2012; 6: e1646.
- 148 Taylor-Robinson DC, Jones AP, Garner P. Deworming drugs for treating soil-transmitted intestinal worms in children: effects on growth and school performance. *Cochrane Database Syst Rev* 2007; 4: CD000371.
- (49) Awasthi S, Peto R, Read S, Richards SM, Pande V, Bundy D, and the DEVTA (Deworming and Enhanced Vitamin A) team. Population deworming every 6 months with albendazole in 1 million pre-school children in north India: DEVTA, a cluster-randomised trial. *Lancet* 2013; 381: 1478–86.