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Hematologically Important Mutations: The Autosomal Recessive Forms of Chronic Granulomatous Disease (Second Update)

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Keywords

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Mutations in the genes encoding four of the phagocyte NADPH oxidase components, p22-*phox*, p47-*phox*, p67-*phox* and 40-*phox*, cause the autosomal recessive forms of chronic granulomatous disease (CGD). These four forms of the disease collectively account for approximately one-third of all CGD cases. Many new mutations have been identified in these four genes since publication of the first updated version of the tables with these mutations (1). The remaining two-thirds of cases are caused by mutations in the X-linked gene for gp91-*phox*, *CYBB*; these mutations have been tabulated previously in this journal (2). The incidence of CGD as a whole is between 1 in 200,000 and 1 in 250,000 individuals.

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The protein p22-*phox* is one of two membrane-bound subunits of cytochrome *b*₅₅₈ (the other is gp91-*phox*), and mutations in the p22-*phox* gene (*CYBA*, located at 16q24, OMIM *608508) account for about 6% of CGD (Table 1). Also about 6% of CGD cases are caused by mutations in the gene for p67-*phox* (*NCF2*, 1q25, OMIM *608515), a cytosolic component of the superoxide-generating NADPH oxidase system (Table 2). The most common form of autosomal recessive CGD (about 20% of all cases) is caused by mutations in the gene for p47-*phox* (*NCF1*, 7q11.23, OMIM *608512), a second cytosolic component of the enzyme (Table 3). Only one patient has been described with mutations in *NCF4* (22q13.1, OMIM *601488), the gene encoding p40-*phox*, the third cytosolic NADPH oxidase component (Table 4). The type, position and number of the mutations in these four genes is depicted in figure 1. Tables 5-8 list apparently benign polymorphisms that have been identified in the *CYBA*, *NCF2*, *NCF1* and *NCF4* genes, respectively. It is important to realize that SNPs and other sequence variants available on the internet are not necessarily functionally neutral.

Unlike the other autosomal recessive and X-linked forms of the disease, in which there is a large heterogeneity among mutations, a single defect accounts for the vast majority of cases of p47-*phox*-deficiency. Of ~250 patients investigated worldwide at the DNA level, all but 53 patients in 42 families appear to be homozygous for a dinucleotide (GT) deletion (GT) at the start of exon 2 (3-19). Of the 42 families with exceptions, 20 had patients who were compound heterozygotes for the GT deletion and one additional mutation, and the others had patients with mutations other than GT on both alleles of *NCF1* (20 homozygous, 2 compound heterozygous). The GT-bearing allele of *NCF1* is therefore the most common CGD-causing allele in the population, carried by approximately 1 in 250 individuals. The reason for this predominance is that most normal individuals have two p47-*phox* pseudogenes, each of which co-localizes with the functional gene to 7q11.23 and carries GT. Recombination events between *NCF1* and these highly homologous pseudogenes lead to the incorporation of GT into *NCF1* (7, 20).

Additional information about the tabulated mutations and about CGD in general can be found in recent reviews (21-25) and in the cited literature. In the following tables we have used the standard notation for differentiating the various phenotypes of CGD (e.g., A22°, A22+, A67°, A67+, A67-, A47°, A40° and A40+). In this nomenclature the first letter refers to the mode of inheritance (autosomal recessive), the numeral indicates the *phox* component affected, and the superscript symbol indicates whether the protein is absent (°), diminished (-) or normal (+), based on immunoblot analysis. When this information is unavailable, that has been indicated as (?). The respective proteins can be non-functional, exert residual activity, or in case of (-) be fully functional. Online Mendelian Inheritance in Man (OMIM) numbers for A22, A67, A47 CGD are #233690, #233710, and #233700, respectively. Mutations added since the last updated versions of Tables 1-3 were published (1) are marked with an asterisk in the right hand column. The nucleotide numbering system we have used is based on the cDNA sequence and follows the convention that +1 is the A of the ATG initiator codon. This differs from the numbering of the GenBank sequences; for p22-*phox* (GenBank accession nos. M21186 and J03774) subtract 28 from the GenBank sequence number to make the initiator A +1; for p67-*phox* (accession no. M32011) subtract 67 from

the GenBank numbering; for p47-*phox* (GenBank accession nos. M25665 and M26193) subtract 12 from the GenBank numbering, and for p40-*phox* (accession no. NM_000631) subtract 184 from the GenBank numbering. The notation of the mutations and polymorphisms follows the recommendations of the Human Genome Variation Society (26) (see also www.hgvs.org/mutnomen). Where possible we have cross-referenced the mutations listed here with those in three CGD databases that list CGD patients by accession number. These databases contain additional biochemical, genetic and clinical information and are available at <http://www.uta.fi/imt/bioinfo/CYBAbase/> (or NCF1base/, or NCF2base/). In addition, information can also be found in the HGMD database at <http://www.hgdm.cf.ac.uk/ac.search.php>. The consequences of the mutations for protein composition have been checked with the Mutalyzer program (www.lovd.nl/mutalyzer) (27).

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References

1. Cross AR, Noack D, Rae J, Curnutte JT, Heyworth PG. Hematologically important mutations: The autosomal recessive forms of chronic granulomatous disease (first update). *Blood Cells Mol Dis*. 2000; 22:268–270.10.1006/bcmd.2000.0333 [PubMed: 9075578]
2. Heyworth PG, Curnutte JT, Rae J, Noack D, Roos D, van Koppen E, Cross AR. Hematologically important mutations: X-linked chronic granulomatous disease (second update). *Blood Cells Mol Dis*. 2001; 23:443–450.10.1006/bcmd.2000.0347 [PubMed: 9454688]
3. Casimir CM, Bu-Ghanim HN, Rodaway ARF, Bentley DL, Rowe P, Segal AW. Autosomal recessive chronic granulomatous disease caused by deletion at a dinucleotide repeat. *Proc Natl Acad Sci USA*. 1991; 88:2753–2757. [PubMed: 2011585]
4. Iwata M, Nunoi H, Yamazaki H, Nakano T, Niwa H, Tsuruta S, Ohga S, Ohmi S, Kanegasaki S, Matsuda I. Homologous dinucleotide (GT or TG) deletion in Japanese patients with chronic granulomatous disease with p47-*phox* deficiency. *Biochem Biophys Res Commun*. 1994; 199:1372–1377. [PubMed: 8147881]
5. Volpp BD, Lin Y. In vitro molecular reconstitution of the respiratory burst in B lymphoblasts from p47-*phox*-deficient chronic granulomatous disease. *J Clin Invest*. 1993; 91:201–207. [PubMed: 7678602]
6. Roos D, De Boer M, Kuribayashi F, Meischl C, Weening RS, Segal AW, Åhlin A, Nemet K, Hossle JP, Bernatowska-Matuszkiewicz E, Middleton-Price H. Mutations in the X-linked and autosomal recessive forms of chronic granulomatous disease. *Blood*. 1996; 87:1663–1681. [PubMed: 8634410]
7. Roesler J, Curnutte JT, Rae J, Barrett D, Patiño P, Chanock SJ, Görlach A. Recombination events between the p47-*phox* gene and its highly homologous pseudogenes are the main cause of autosomal recessive chronic granulomatous disease. *Blood*. 2000; 95:2150–2156. [PubMed: 10706888]
8. Noack D, Rae J, Cross AR, Ellis BA, Newburger PE, Curnutte JT, Heyworth PG. Autosomal recessive chronic granulomatous disease caused by defects in *NCF1*, the gene encoding the

phagocyte p47-*phox*: mutations not arising in the *NCF1* pseudogenes. *Blood*. 2001; 97:305–311. [PubMed: 11133775]

9. Vihinen, M.; Arredondo-Vega, FX.; Casanova, JL.; Etzioni, A.; Giliani, S.; Hammarström, L.; Hershfield, MS.; Heyworth, PG.; Hsu, AP.; Lähdesmäki, A.; Lappalainen, I.; Notarangelo, LD.; Puck, JM.; Reith, W.; Roos, D.; Schumacher, RF.; Schwarz, K.; Vezzoni, P.; Villa, A.; Väliäho, J.; Smith, CIE. Primary immunodeficiency mutation databases. In: Hall, JC.; Dunlap, JC.; Friedmann, T.; Giannelli, F., editors. *Advances in Genetics*. Vol. 43. 2000. in press
10. Jurkowska M, Kurenko-Deptuch M, Bal J, Roos D. The search for a genetic defect in Polish patients with chronic granulomatous disease. *Arch Immunol Ther Exp*. 2004; 52:441–446.
11. Prando-Andrade C, Agudelo-Florez P, Lopez JA, Aparecida de Souza Paiva M, Costa-Carvalho BT, Condino-Neto A. Autosomal chronic granulomatous disease: case report and mutation analysis of two Brazilian siblings. *J Pediatr (Rio J)*. 2004; 80:425–428. [PubMed: 15505740]
12. Agudelo-Flórez P, Cardoso Prando-Andrade C, López JA, Costa-Carvalho BT, Quezada A, Espinosa FJ, Aparacida de Souza Paiva M, Roxo P, Grumach A, Abe Jacob C, Salles Carneiro-Sampaio MM, Newburger PE, Condino-Neto A. Chronic granulomatous Disease in Latin American patients: clinical spectrum and molecular genetics. *Pediatr Blood Cancer*. 2006; 46:243–252. [PubMed: 16123991]
13. El Kares R, Barbouche MR, Elloumi-Zghal H, Bejaoui M, Chemli J, Mellouli F, Tebib N, Abdelmoula MS, Boukthir S, Fitouri Z, M'Rad S, Bouslama K, Touiri H, Abdelhak S, Dellagi MK. Genetic and mutational heterogeneity of autosomal recessive chronic granulomatous disease in Tunisia. *J Hum Genet*. 2006; 51:887–895. [PubMed: 16937026]
14. Roos D, de Boer M, Köker MY, Dekker J, Singh-Gupta V, Åhlin A, Palmblad J, Sanal Ö, Kurenko-Deptuch M, Jolles S, Wolach B. Chronic granulomatous disease caused by mutations other than the common GT deletion in *NCF1*, the gene encoding the p47^{phox} component of the phagocyte NADPH oxidase. *Hum Mutat*. 2006; 27:1218–1229. [PubMed: 16972229]
15. Köker MY, Sanal Ö, de Boer M, Teczan , Metin A, Ersoy F, Roos D. Mutations of chronic granulomatous disease in Turkish families. *Eur J Clin Invest*. 2007; 37:589–595. [PubMed: 17576211]
16. Wolach B, Gavrieli R, de Boer M, Gottesman G, Ben-Ari J, Rottem M, Schlesinger Y, Etzioni A, Roos D. Chronic granulomatous disease in Israel: functional and molecular studies of 38 patients. *Clin Immunol*. 2008; 129:103–114. [PubMed: 18708296]
17. Kannengiesser C, Gérard B, El Benna J, Henri D, Krovianski Y, Chollet-Martin S, Gougerot-Pocidallo MA, Elbim C, Grandchamp B. Molecular epidemiology of chronic granulomatous disease in a series of 80 kindreds: identification of 31 novel mutations. *Hum Mutat*. 2008; 29:E132–E149. [PubMed: 18546332]
18. Van de Vosse E, van Wengen A, van Geelen JA, de Boer M, Roos D, van Dissel JT. A novel mutation in *NCF1* in an adult CGD patient with a liver abscess as First presentation. *J Hum Genet*. 2009; 54:313–316. [PubMed: 19329991]
19. Bakri FG, Martel C, Khuri-Bulos N, Mahafza A, El-Khateeb MS, Al-Wahadneh AM, Hayajneh WA, Hamamy HA, Maquet E, Molin M, Stasia MJ. First report of clinical, functional, and molecular investigation of chronic granulomatous disease in nine Jordanian families. *J Clin Immunol*. 2009; 29:215–230. [PubMed: 18773283]
20. Görlach A, Lee PL, Roesler J, Hopkins PJ, Christensen B, Green ED, Chanock SJ, Curnutte JT. A p47-*phox* pseudogene carries the most common mutation causing p47-*phox*-deficient chronic granulomatous disease. *J Clin Invest*. 1997; 100:1907–1918. [PubMed: 9329953]
21. Roos, D.; Kuijpers, TW.; Curnutte, JT. Chronic granulomatous disease. In: Ochs, HD.; Smith, CIE.; Puck, JM., editors. *Primary immunodeficiency diseases, a molecular and genetic approach*. 2nd. Oxford University Press; New York: 2007. p. 525-549.
22. Stasia MJ, Li XJ. Genetics and immunopathology of chronic granulomatous disease. *Semin Immunopathol*. 2008; 30:209–235. [PubMed: 18509647]
23. Martire B, Rondelli R, Soresina A, Pignata C, Broccoletti T, Finocchi A, Rossi P, Gattorno M, Rabusin M, Azzari C, Dellepiane RM, Pietrogrande MC, Trizzino A, DiBartolomeo P, Martino S, Carpino L, Cossu F, Locatelli F, Maccario R, Pierani P, Putti MC, Stabile A, Notarangelo LD, Ugazio AG, Plebani A, De Mattia D. Clinical features, long-term follow-up and outcome of a

- large cohort of patients with chronic granulomatous disease: an Italian multicenter study. *Clin Immunol.* 2008; 126:155–164. [PubMed: 18037347]
24. Jones LB, McGrogan P, Flood TJ, Gennery AR, Morton L, Thrasher A, Goldblatt D, Parker L, Cant AJ. Chronic granulomatous disease in the United Kingdom and Ireland: a comprehensive national patient-based registry. *Clin Exp Immunol.* 2008; 152:211–218. [PubMed: 18410635]
 25. Van den Berg JM, van Koppen E, Åhlin A, Belohradsky BH, Bernatowska E, Corbeel L, Espanöl T, Fischer A, Kurenko-Deptuch M, Mouy R, Petropoulou T, Roesler J, Seger R, Stasia MJ, Valerius NH, Weening RS, Wolach B, Roos D, Kuijpers TW. Chronic granulomatous disease: the European experience. *PLoS ONE.* 2009; 4:e5234. [PubMed: 19381301]
 26. Den Dunnen JT, Antonarakis SE. Mutation nomenclature extensions and suggestions to describe complex mutations: a discussion. *Hum Mutat.* 2000; 15:7–12. [PubMed: 10612815]
 27. Wildeman M, van Ophuizen E, den Dunnen JT, Taschner PE. Improving sequence variant descriptions in mutation databases and literature using the Mutalyzer sequence variation nomenclature checker. *Hum Mutat.* 2008; 29:6–13. [PubMed: 18000842]
 28. Dinauer MC, Pierce EA, Bruns GAP, Curnutte JT, Orkin SH. Human neutrophil cytochrome *b* light chain (p22-*phox*). Gene structure, chromosomal location, and mutations in cytochrome-negative autosomal recessive chronic granulomatous disease. *J Clin Invest.* 1990; 86:1729–1737. [PubMed: 2243141]
 29. Yamada M, Ariga T, Kawamura N, Ohtsu M, Imajoh-Ohmi S, Ohshika E, Tatsuzawa O, Kobayashi K, Sakiyama Y. Genetic studies of three Japanese patients with p22-*phox*-deficient chronic granulomatous disease: detection of a possible common mutant *CYBA* allele in Japan and a genotype-phenotype correlation in these patients. *Br J Haematol.* 2000; 108:511–517. [PubMed: 10759707]
 30. Rae J, Noack D, Heyworth PG, Ellis BA, Curnutte JT, Cross AR. Molecular analysis of nine new families with chronic granulomatous disease caused by mutations in *CYBA*, the gene encoding p22-*phox*. *Blood.* 2000; 96:1106–1112. [PubMed: 10910929]
 31. Ishibashi F, Nunoi H, Endo F, Matsuda I, Kanegasaki S. Statistical and mutational analysis of chronic granulomatous disease in Japan with special reference to gp91-*phox* and p22-*phox* deficiency. *Hum Genet.* 2000; 106:473–481.10.1007/s004390000288 [PubMed: 10914676]
 32. De Boer M, Hartl D, Wintergerst U, Belohradsky BH, Roos D. A donor splice site mutation in intron 1 of *CYBA*, leading to chronic granulomatous disease. *Blood Cells Mol Dis.* 2005; 35:365–369. [PubMed: 16157492]
 33. Köker MY, van Leeuwen K, de Boer M, Çelmeli F, Metin A, Özgür TT, Sanal Ö, Roos D. Six different *CYBA* mutations including three novel mutations in ten families from Turkey, resulting in autosomal recessive chronic granulomatous disease. *Eur J Clin Invest.* 2009; 39:311–319. [PubMed: 19292887]
 34. Teimourian S, Zomorodian E, Badalzadeh M, Pouya A, Kannengiesser C, Mansouri D, Cheraghi T, Parvaneh N. Characterization of six novel mutations in *CYBA*: the gene causing autosomal recessive chronic granulomatous disease. *Br J Haematol.* 2008; 141:848–851. [PubMed: 18422995]
 35. Hossle JP, De Boer M, Seger RA, Roos D. Identification of allele-specific p22-*phox* mutations in a compound heterozygous patient with chronic granulomatous disease by mismatch PCR and restriction enzyme analysis. *Hum Genet.* 1994; 93:437–442. [PubMed: 8168815]
 36. Bagg A, Gonzales-Peralta R, Petrovic A, Sleasman JW. Novel *CYBA* gene mutation in a patient with chronic granulomatous disease associated with autoimmune hepatitis. *J Allergy Clin Immunol.* 2007; 119(Suppl. 1):S16.
 37. De Boer M, De Klein A, Hossle JP, Seger R, Corbeel L, Weening RS, Roos D. Cytochrome *b*₅₅₈-negative, autosomal recessive chronic granulomatous disease: two new mutations in the cytochrome *b*₅₅₈ light chain of the NADPH oxidase (p22-*phox*). *Am J Hum Genet.* 1992; 51:1127–1135. [PubMed: 1415254]
 38. Stasia MJ, Bordigoni P, Martel C, Morel F. A novel and unusual case of chronic granulomatous disease in a child with homozygous 36-bp deletion in the *CYBA* gene (A22°) leading to the activation of a cryptic splice site in intron 4. *Hum Genet.* 2002; 110:444–450. [PubMed: 12073015]

39. Porter CD, Parkar MH, Kinnon C. Identification of a donor splice site mutation leading to loss of p22-phox exon 5 in autosomal chronic granulomatous disease. *Hum Mutat.* 1996; 7:374. [PubMed: 8723692]
40. Dinauer MC, Pierce EA, Erickson RW, Muhlebach TJ, Messner H, Orkin SH, Seger RA, Curnutte JT. Point mutation in the cytoplasmic domain of the neutrophil p22-*phox* cytochrome *b* subunit is associated with a nonfunctional NADPH oxidase and chronic granulomatous disease. *Proc Natl Acad Sci USA.* 1991; 88:11231–11235. [PubMed: 1763037]
41. Patiño PJ, Rae J, Noack D, Erickson RW, Ding J, Garcia de Olarte D, Curnutte JT. Molecular characterization of autosomal recessive chronic granulomatous disease caused by a defect of the NADPH oxidase component p67-*phox*. *Blood.* 1999; 94:2505–2514. [PubMed: 10498624]
42. Noack D, Rae J, Cross AR, Munoz J, Salmen S, Mendoza JA, Rossi N, Curnutte JT, Heyworth PG. Autosomal recessive chronic granulomatous disease caused by novel mutations in *NCF-2*, the gene encoding the p67-*phox* component of phagocyte oxidase. *Hum Genet.* 1999; 105:460–467.10.1007/s004399900152 [PubMed: 10598813]
43. Yu G, Hong DK, Dionis KY, Rae J, Heyworth PG, Curnutte JT, Lewis DB. The continuing diagnostic challenge of autosomal recessive chronic granulomatous disease. *Clin Immunol.* 2008; 128:117–126. [PubMed: 18625437]
44. Åhlin A, De Boer M, Roos D, Leusen J, Smith CIE, Sundin U, Rabbani H, Palmblad J, Elinder G. Prevalence, genetics and clinical presentation of chronic granulomatous disease in Sweden. *Acta Paediatr.* 1995; 84:1386–1394. [PubMed: 8645957]
45. Leusen JHW, De Klein A, Hilarius PM, Åhlin A, Palmblad J, Smith CIE, Diekmann D, Hall A, Verhoeven AJ, Roos D. Disturbed interaction of p21-*rac* with mutated p67-*phox* causes chronic granulomatous disease. *J Exp Med.* 1996; 184:1243–1249. [PubMed: 8879195]
46. Köker MY, Sanal Ö, van Leeuwen K, de Boer M, Metin A, Patiro lu T, Özgür TT, Tezcan I, Roos D. Four different *NCF2* mutations in six families from Turkey and an overview of *NCF2* gene mutations. *Eur J Clin Invest.* 2009; 39:942–951. [PubMed: 19624736]
47. De Boer M, Hilarius-Stokman PM, Hossle JP, Verhoeven AJ, Graf N, Kenney RT, Seger R, Roos D. Autosomal recessive chronic granulomatous disease with absence of the 67-kD cytosolic NADPH oxidase component: identification of mutation and detection of carriers. *Blood.* 1994; 83:531–536. [PubMed: 8286749]
48. Tanugi-Cholley LC, Issartel JP, Lunardi J, Freycon F, Morel F, Vignais PV. A mutation located at the 5' splice junction sequence of intron 3 in the p67-*phox* gene causes the lack of p67-*phox* mRNA in a patient with chronic granulomatous disease. *Blood.* 1995; 85:242–249. [PubMed: 7803798]
49. Khan HA, Good RA, Tangsinmankong N, Rae J, Noack D, Heyworth P, Day NK, Bahna S. P67-phox deficient chronic granulomatous disease due to heterozygous defects in exons 4 and 12 of the *NCF2* gene. *J Allergy Clin Immunol.* 2002; 109(Suppl. 1):S278.
50. Gentsch M, Kaczmarczyk A, van Leeuwen K, de Boer M, Kaus-Drobek M, Dagher MC, Kaiser P, Arkwright P, Gahr M, Rösen-Wolff A, Bochtler M, Secord E, Saifi M, Maddalena A, Dbaibo G, Bustamante J, Casanova JL, Roos D, Roesler J. Alu-repeat-induced deletions within the *NCF2* gene cause p67-phox-deficient chronic granulomatous disease (CGD). *Hum Mutat.* 2009 Dec 1. Epub ahead of print.
51. Nunoi H, Iwata M, Tatsuzawa S, Onoe Y, Shimizu S, Kanegasaki S, Matsuda I. AG dinucleotide insertion in a patient with chronic granulomatous disease lacking cytosolic 67-kD protein. *Blood.* 1995; 86:329–333. [PubMed: 7795241]
52. Bonizzato A, Russo MP, Donini M, Dusi S. Identification of a double mutation (D160V-K161E) in the p67-*phox* gene of a chronic granulomatous disease patient. *Biochem Biophys Res Commun.* 1997; 231:861–863. [PubMed: 9070911]
53. Al-Muhsen S, Al-Hemidan A, Al-Sheri A, Al-Harbi A, Al-Ghoniaim A, Al-Saud B, Al-Mousa H, Al-Dhekri H, Arnaout R, Al-Mohsen I, Alsmadi O. Ocular manifestations in chronic granulomatous disease in Saudi Arabia. *J Am Ass Pediatr Ophtalmol Strabismus.* 2009; 13:396–399.
54. Borgato L, Bonizzato A, Lunardi C, Dusi S, Andrioli G, Scarperi A, Corrocher R. A 1.1-kb duplication in the p67-*phox* gene causes chronic granulomatous disease. *Hum Genet.* 2001; 108:504–510. [PubMed: 11499676]

55. Aoshima M, Nunoi H, Shimazu M, Shimizu S, Tatsuzawa O, Kenney RT, Kanegasaki S. Two-exon skipping due to a point mutation in p67-*phox*-deficient chronic granulomatous disease. *Blood*. 1996; 88:1841–1845. [PubMed: 8781442]
56. De Boer M, Singh V, Dekker J, Di Rocco M, Goldblatt D, Roos D. Prenatal diagnosis in two families with autosomal, p47^{phox}-deficient chronic granulomatous disease due to a novel point mutation in *NCF1*. *Prenatal Diagn*. 2002; 22:235–240.
57. Jakobsen MA, Pedersen SS, Barington T. Detection of non- GT *NCF1* mutations in chronic granulomatous disease. *Genet Test Mol Biomarkers*. 2009; 13:505–510. [PubMed: 19663600]
58. Kabuki T, Kawai T, Kin Y, Joh K, Ohashi H, Kosho T, Yachie A, Kanegane H, Miyawaki T, Ohishi T. A case of Williams syndrome with p47-*phox*-deficient chronic granulomatous disease. *Nihon Rinsho Meneki Gakkai Kaishi*. 2003; 26:299–303. [PubMed: 14635404]
59. Matute JD, Arias AA, Wright NAM, Wrobel I, Waterhouse CCM, Li XJ, Marchal CC, Stull ND, Lewis DB, Steele M, Kellner JD, Yu W, Meroueh SO, Nauseef WM, Dinuer MC. A new genetic subgroup of chronic granulomatous disease with autosomal recessive mutations in p40^{phox} and selective defects in neutrophil NADPH oxidase activity. *Blood*. 2009
60. Bedard K, Attar H, Bonnefont J, Jaquet V, Borel C, Plastre O, Stasia MJ, Antonarakis SE, Krause KH. Three common polymorphisms in the *CYBA* gene form a haplotype associated with decreased ROS formation. *Hum Mutat*. 2009; 30:1123–1133. [PubMed: 19388116]
61. Olsson LM, Lindqvist AK, Källberg H, Padyukov L, Burkhardt H, Alfredsson L, Klareskog L, Holmdahl R. A case-control study of rheumatoid arthritis identifies an associated single nucleotide polymorphism in the *NCF4* gene, supporting a role for the NADPH-oxidase complex in autoimmunity. *Arthritis Res Ther*. 2007; 9:R98. [PubMed: 17897462]

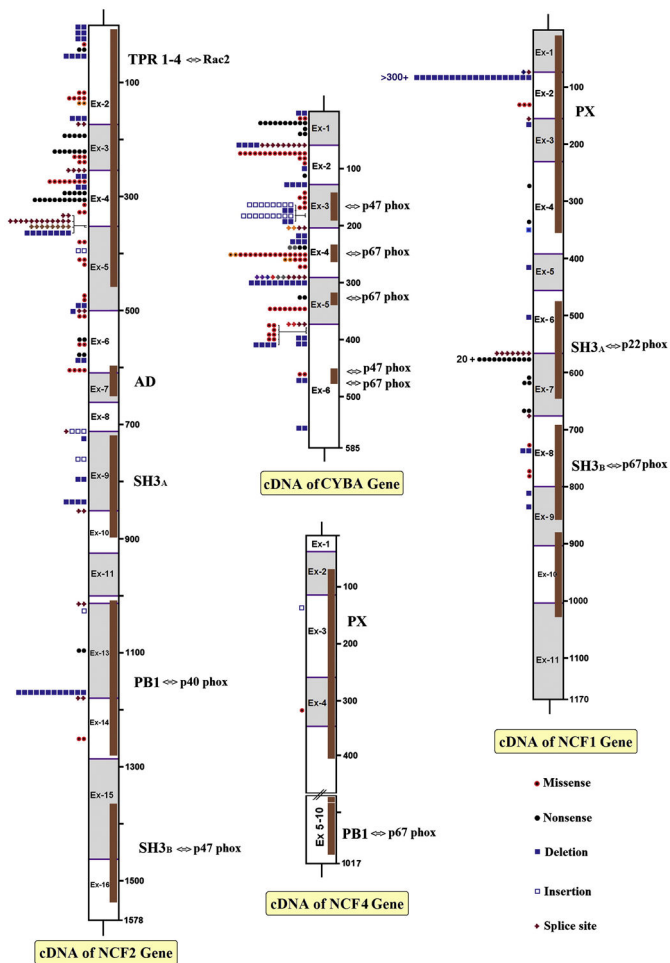


Figure 1. Schematic overview of mutations in *NCF2*, *CYBA*, *NCF4* and *NCF1*
 For each cDNA, the exon positions and the corresponding protein domains have been depicted. For some of the protein domains, their interaction with other proteins has been indicated. The PX domains interact with phosphatidylinositol-phosphates. The type of mutations (explained in the right hand corner), their position and number of mutated alleles are indicated. Splice site mutations are given at the exon borders.

Table 1

Mutations in the p22-phox gene CYBA.

Nucleotide change	Mutation	Amino acid or mRNA change	CGD type	Reference	Families (alleles) ^d	Accession number(s) ^g
g. del>10 kb	Deletion	NA ^b	A22°	[28]	1(2)	
c.2T>A	Missense	p.Met1Lys	A22°	unpubl. ^d	1(2) ^c	*
c.7C>T	Nonsense	p.Gln3X	A22°	[29] unpubl.	5(10)	H0023 H0026
c.26G>A	Nonsense	p.Trp9X	A22°	[30]	1(1)	H0017
c.27G>A	Nonsense	p.Trp9X	A22°	[31]	1(2)	H0027
c.58+4_-+7delAGTG ^e	Splice site	ins. 79 bp in intron 1 p.Ile20SerfsX77	A22°	[31,32] unpubl.	5(9)	H0028 H0039
g.exon2_3del	Deletion	p.Ile20ArgfsX6	A22°	[30]	1(2)	
g.exon2_5del	Deletion	p.Ile20SerfsX68	A22°	unpubl.	1(2) ^c	*
c.70G>A	Missense	p.Gly24Arg	A22°	[16,29-31,33] unpubl.	9(14)	H0021 H0024 H0025 H0028 H0030 H0034
c.71G>A	Missense	p.Gly24Glu	A22°	[16]	1(2)	*
c.74G>T	Missense	p.Gly25Val	A22°	[30]	1(1)	H0017
c.77delT	Deletion	p.Ile26ThrfsX48	A22°	unpubl.	1(1)	*
c.107G>A	Nonsense	p.Trp36X	A22°	[30]	1(1)	H0021
g.exon3_5del	Deletion	p.Ile43MetfsX68	A22°	[34]	1(2)	*
g.exon3_6del	Deletion	NA	A22°	[33]	1(2)	*
c.152T>A	Missense	p.Leu51Gln	A22°	unpubl.	1(1)	*
c.155T>C	Missense	p.Leu52Pro	A22°	[30]	1(2)	H0014
c.158A>T	Missense	p.Glu53Val	A22°	[35]	1(1)	H0005
c.164C>G	Missense	p.Pro55Arg	A22°	[16]	1(2)	*
c.166dupC	Insertion	p.Arg56ProfsX157	A22°	[30,33] unpubl.	5(8)	H0019 H0020
c.171delG	Deletion	p.Lys58ArgfsX16	A22°	unpubl.	1(2)	*

Nucleotide change	Mutation	Amino acid or mRNA change	CGD type	Reference	Families (alleles) ^d	Accession number(s) ^e
c.171dupG	Insertion	p.Lys58GlufsX155	A22°	[16,35] unpubl.	5(9)	H0005 H0032 H0036 H0037
c.173delA #	Deletion	p.Lys58ArgfsX16	A22°	[34]	1(2)	H0041 *
c.203+1G>T ^e	Splice site	del. exon 3 p.Ile43_Trp68delinsMet	A22°	[30]	1(2)	H0016
c.204-2A>G ^e	Splice site	del. exon 4? p.Gly69_Leu96del?	A22°	[36]	1(2)	*
c.223delG #	Deletion	p.Ala75ProfsX3	A22°	[34]	1(2)	H0042 *
c.246delC	Deletion	p.Phe83LeufsX108	A22°	[28,30]	2(3)	H0001 H0015
c.261C>G	Nonsense	p.Tyr87X	A22°	unpubl.	1(2)	*
c.261C>A	Nonsense	p.Tyr87X	A22°	unpubl.	1(2)	*
c.268C>T	Missense	p.Arg90Trp	A22°	[30] unpubl.	8(14)	H0018 H0019 H0020
c.268C>G	Missense	p.Arg90Gly	A22°	unpubl.	1(2)	*
c.269G>A	Missense	p.Arg90Gln	A22°	[28,37]	2(3)	H0001 H0006 H0007 H0008
c.269G>C	Missense	p.Arg90Pro	A22°	unpubl.	1(2)	*
c.281A>G	Missense	p.His94Arg	A22°	[37]	1(2)	H0012
c.287+1G>A ^e	Splice site	del. exon 4 p.Gly69_Leu96del	A22°	[37] unpubl.	2(4)	H0009 H0035
c.287+1G>T ^e	Splice site	del. exon 4 p.Gly69_Leu96del	A22°	unpubl.	1(2)	*
c.287+2T>C ^e	Splice site	del. exon 4? p.Gly69_Leu96del?	A22°	unpubl.	1(1) ^f	*
c.288-6_296del16 ^e	Splice site	del. exon 5 p.Leu97ArgfsX68	A22°	unpubl.	1(1)	*
c.288-15_308del36 ^e	Splice site	intron4ins179/exon5del21	A22°	[38]	1(2)	H0038
c.288G>T	Splice site	del. exon 5 p.Leu97ArgfsX68	A22°	unpubl.	1(2)	*
c.295_301delGTGCCCG	Deletion	p.Val99ProfsX90	A22°	[13,17,19] unpubl.	5(10) ^c	H0040 *
c.339C>A	Nonsense	p.Cys113X	A22°	[33]	1(2)	*

Nucleotide change	Mutation	Amino acid or mRNA change	CGD type	Reference	Families (alleles) ^d	Accession number(s) ^g
c.354C>A	Missense	p.Ser118Arg	A22°	[28,30] unpubl.	4(8)	H0003 H0010 H0022
c.369+1G>C ^e	Splice site	del. exon 5 p.Leu97ArgfsX68	A22°	[6,39]	1(2)	H0002
c.369+1G>A ^e	Splice site	del exon 5 p.Leu97ArgfsX68	A22°	[34]	1(2)	H0045 H0046
c.370G>T	Missense	p.Ala124Ser	A22°	[17]	1(2)	*
c.371C>T	Missense	p.Ala124Val	A22°	[31]	1(1)	H0030
c.373G>A	Missense	p.Ala125Thr	A22°	[34]	1(2)	H0047
c.385G>A	Missense	p.Glu129Lys	A22?	unpubl.	1(2)	*
c.385_388delGAGC	Deletion	p.Glu129SerfsX61	A22°	[34]	2(4)	H0043 H0044
c.399delC	Deletion	p.Ile134SerfsX57	A22?	unpubl.	1(2) ^c	*
c.408delC	Deletion	p.Lys137SerfsX54 [#]	A22°	[33]	1(2)	*
c.467C>A	Missense	p.Pro156Gln	A22+	[40]	1(2)	H0011
c.472_484del	Deletion	p.Pro160AlafsX27	A22?	[1]	1(2) ^c	H0031
c.571_604del	Deletion	p.Thr191ProfSX	A22°	[31]	1(2)	
Mutations in <i>CYBA</i>						
			Total number of different alleles		Total number of alleles	
Deletions			16 alleles	(29.1%)	42 alleles	(24.3%)
Nonsense mutations			7 alleles	(12.7%)	20 alleles	(11.6%)
Splice site mutations			11 alleles	(20.0%)	29 alleles	(16.8%)
Missense mutations			19 alleles	(34.6%)	65 alleles	(37.5%)
Insertions			2 alleles	(3.6%)	17 alleles	(9.8%)
Total 55 different allelic mutations					Total 87 families with 173 identified alleles in the 96 patients	

^aNumber of different families with patients with this mutation (number of alleles carrying this mutation).

^bNot applicable.

^cOne patient presumed homozygous for this mutation.

^dUnpublished data from the authors' laboratories.

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^e Position of introns in *CYBA*: intron 1 c.58_59; intron 2 c.128_129; intron 3 c.203_204; intron 4 c.287_288; intron 5 c.369_370.

^f Patient is heterozygous for this mutation and for an unidentified mutation in the other allele.

^g Accession number in database at <http://www.uta.fi/mt/bioinfo/CYBAbase/>.

* New mutations since ref. [1].

Corrected after consultation of the authors.

Table 2

Mutations in the p67-*phox* gene *NCF-2*.

Nucleotide change	Mutation	Amino acid or mRNA change	CGD type	Families (alleles)	Reference	Accession number <i>h</i>
c.-547-?_174+?del <i>g</i>	Deletion	p..Met1_Lys58del	A67°	1(2)	Unpubl.	*
c.-274-?_174 + ?del~400 <i>g</i>	Deletion	p..Met1_Lys58del	A67°	1(2)	Unpubl.	*
c.-274-?_174 + ?del~400 <i>g</i>	Deletion	p..Met1_Lys58del	A67°	1(2)	Unpubl.	*
c.1A>G	Missense	p..Met1Val	A67°	1(1)	Unpubl.	*
c.29G>A	Nonsense	p..Trp10X	A67°	1(2)	Unpubl.	*
c.55_63delAAAGGAC <i>a</i>	Deletion	p..Lys19_Asp21del	A67°	4(4)	[41-43] unpubl.	M0004 M0015
c.125A>G	Missense	p..Asn42Ser	A67°	1(2)	Unpubl.	*
c.130G>C	Missense	p..Gly44Arg	A67?	2(4) <i>b</i>	[1,43]	
c.130G>T	Missense	p..Gly44Cys	A67°	1(2)	Unpubl.	*
c.172_174delAAG	Deletion	p..Lys58del	A67+	2(3) <i>c</i>	[44,45] unpubl.	M0009
c.175-1G>A <i>g</i>	Splice site	del, exon 3? p..Ala59IlefsX2?	A67°	1(2)	Unpubl.	*
c.196C>T	Nonsense	p..Arg66X	A67°	3(5)	[42]	
c.229C>T	Nonsense	p..Arg77X	A67°	4(7)	[46], unpubl.	*
c.230G>A	Missense	p..Arg77Gln	A67°	3(3)	[42], unpubl.	M0016
c.233G>A	Missense	p..Gly78Glu	A67°	1(2)	[47]	M0002
c.257+2T>C <i>g</i>	Splice site	del, exon3 p..Ala59IlefsX2	A67°	2(4)	[13,48]	M0010 M0020
c.258-?_366 + ?del~1100 <i>g</i>	Deletion	p..Tyr87CysfsX22	A67?	1(2)	Unpubl.	*
c.279C>G <i>d</i>	Missense	p..Asp93Glu	A67°	4(8)	[46], unpubl.	*
c.287_289delAAG	Deletion	p..Glu96del	A67-	1(2)	Unpubl.	*
c.298C>T	Nonsense	p..Gln100X	A67°	5(5)	[42], unpubl.	M0016
c.304C>T	Nonsense	p..Arg102X	A67°	6(11) <i>b</i>	[16,41,46,48] unpubl.	M0001
c.305G>C	Missense	p..Arg102Pro	A67+?	1(1)	Unpubl.	*
c.323A>T	Missense	p..Asp108Val	A67-?	1(2)	[43]	*
c.364_366+2delGAGGT <i>g</i>	Splice site	del, exon 4? p..Tyr87CysfsX2?	A67°	1(2)	[17]	*

Nucleotide change	Mutation	Amino acid or mRNA change	CGD type	Families (alleles)	Reference	Accession number <i>h</i>
c.366+1G>A <i>g</i>	Splice site	del. exon 3_4 p.Ala59_Glu122del	A67°	8(12)	[17,41–43] unpubl.	M0011 M0018
c.366+1G>C <i>g</i> , <i>g</i>	Splice site	del. exon 4? p.Tyr87CysfsX22?	A67°	4(8)	Unpubl.	*
c.366+240L_502–527 del1380 <i>g</i>	Deletion	del. exon 5 p.Val123_Trp167del	A67°	4(8)	[50]	*
c.383C>T	Missense	p.Ala128Val	A67°	1(2)	[42]	M0013
c.398_399dupAG	Insertion	p.Lys134ArgfsX12	A67°	1(2)	[51]	M0006
c.409T>A	Missense	p.Trp137Arg	A67–	1(2)	Unpubl.	*
c.419C>G	Missense	p.Ala140Asp	A67°	1(1)	Unpubl.	*
c.[479A>T;481A>G]	Dbl. missense	p.AspLys160_161 ValGlu	A67°	1(1) <i>f</i>	[52]	M0012
c.482delA	Deletion	p.Lys161ArgfsX16	A67?	1(2) <i>b</i>	Unpubl.	*
c.488_501delTTGGAGTGTCTCTGG <i>g</i>	Deletion/splice site	del. exon 5 p.Val123_Trp167del	A67°	1(1)	Unpubl.	*
c.502–1G>T <i>g</i>	Splice site	del. exon 6? p.Lys168_Thr203del?	A67°	1(2)	Unpubl.	*
c.505C>G	Missense	p.Gln169Glu	A67?	1(2)	Unpubl.	*
c.550C>T	Nonsense	p.Arg184X	A67?	1(2)	Unpubl.	*
c.551G>C	Missense	p.Arg184Pro	A67°	1(2)	Unpubl.	*
c.576C>T	Nonsense	p.Gln192X	A67?	1(2) <i>b</i>	[53]	*
c.586_588delAAAG	Deletion	p.Lys196del	A67+	1(2)	Unpubl.	*
c.605C>T	Missense	p.Ala202Val	A67–	2(4)	[46] unpubl.	*
c.714–?_924+?dup~1100 <i>g</i>	Insertion	p.Glu309GlyfsX15	A67°	2(3) <i>f</i>	[54] unpubl.	*
c.714–1G>T <i>g</i>	Splice site	del. exon 9? p.Ala239ArgfsX59?	A67°	1(1)	Unpubl.	*
c.728delA	Deletion	p.Glu243GlyfsX28	A67°	1(1)	[41]	*
c.767_768dupAA	Insertion	p.Glu257LysfsX15	A67°	1(2)	Unpubl.	*
c.799_800delGT	Deletion	p.Val267LeufsX8	A67°	1(2)	[17]	*
c.835_836delAC	Deletion	p.Thr279GlyfsX16	A67°	2(4) <i>b</i>	[42] unpubl.	M0017
c.855+1G>A <i>g</i>	Splice site	del. exon 8_9 p.Ala224_Gln285del	A67°	1(2)	[55]	M0007
c.1026G>A <i>g</i>	Splice site	del. exon 12 p.Gln335SerfsX38	A67°	2(2)	[49]	*

Nucleotide change	Mutation	Amino acid or mRNA change	CGD type	Families (alleles)	Reference	Accession number ^h
c.1034dupA	Insertion	p.Leu346AlafsX36	A67 [?]	1(1)	Unpubl.	*
c.1099C>T	Nonsense	p.Gln367X	A67 [?]	1(2) ^b	Unpubl.	*
c.1171_1175delAAGCT	Deletion	p.Lys391GlufsX9	A67 [°]	6(12)	[16,19,41]	M0005
c.1179-2A>T ^g	Splice site	del. exon 14? p.Ser393ArgfsX54?	A67 [°]	1(2)	Unpubl.	*
c.1256A>T	Missense	p.Asn419Ile	A67 [°]	1(2)	[13]	M0019
Mutations in <i>NCF2</i>						
Number of different alleles			Total number of alleles			
Deletions	14 alleles		(25.9%)	48 alleles		(28.1%)
Nonsense mutations	8 alleles		(14.8%)	36 alleles		(21.0%)
Splice site mutations	11 alleles		(20.4%)	38 alleles		(22.2%)
Missense mutations	17 alleles		(31.5%)	41 alleles		(24.0%)
Insertions	4 alleles		(7.4%)	8 alleles		(4.7%)
Total 54 different allelic mutations			Total 83 families with 171 identified alleles in the 95 patients			

^a Always in combination with c.1183C>T (polymorphism, rs13306575) on the same allele.

^b One patient presumed homozygous for this mutation.

^c One patient is heterozygous for this mutation and for an undefined deletion of 11-13 kb in the other allele [44,45].

^d Always in combination with c.366+1G>C on the same allele.

^e Always in combination with c.279C>G on the same allele.

^f One patient is heterozygous for this mutation and for an unidentified mutation in the other allele [54].

^g Positions of introns in *NCF2*: intron 1 c.-510_-274 in 5' UTR; intron 2 c.174_175; intron 3 c.257_258; intron 4 c.366_367; intron 5 c.501_502; intron 6 c.609_610; intron 7 c.669_670; intron 8 c.713_714; intron 9 c.855_856; intron 10 c.924_925; intron 11 c.1000_1001; intron 12 c.1026_1027; intron 13 c.1178_1179; intron 14 c.1290_1291; intron 15 c.1468_1469.

^h Accession number in database at <http://www.uta.fi/mt/bioinfo/NCF2base/>.

Table 3

Mutations in the p47-*phox* gene *NCF1*.

Nucleotide change	Mutation	Amino acid or mRNA change	CGD type	Families (alleles)	Reference	Accession number(s) ^a
c.72+1G>A ^e	Splice site	del. exon 1? p.Met1_Tyr24del?	A47°	1(1)	[8]	N0031
c.72+3G>T ^e	Splice site	del. exon 1? p.Met1_Tyr24del?	A47°	1(1)	[8]	N0032
c.75_76delGT	Deletion	p.Tyr26HisfsX26	A47°	>300 homozygous, 20 heterozygous ^b	[3–19] unpubl.	N0004–7,9–23, 27–29, 31–35, 40–63, 69,70
c.125G>A	Missense	p.Arg42Gln	A47°	3(3)	[8] unpubl.	N0034N00035
c.153+1G>A ^e	Splice site	c.153+1_+73ins ^c p.Lys52MetX24	A47°	1(1)	[14]	N0068
c.154–283_451+821 del2858 ^e	Deletion	del. exon 3_5 p.Lys52ThrfsX82	A47°	1(1)	[14]	N0061
c.271C>T	Nonsense	p.Gln91X	A47°	1(1)	[14]	N0027
c.333T>A	Nonsense	p.Cys111X	A47°	1(1)	[9,14]	N0028 N0056 N0057 N0058
c.353_354delCC insAA	Deletion/insertion	p.Phe118X	A47°	1(1)	[10,14]	N0059
c.417_451+650 del685 ^e	Deletion	del. exon 5 p.Thr133HisfsX66	A47°	1(1)	unpubl.	*
c.502delG	Deletion	p.Glu168ArgfsX19	A47°	1(1)	[5]	N0002
c.574G>A ^e	Splice site	del. exon 6+7 ^d p.Asp151_Ala227del	A47°	4(7) unpubl.	[8, 14]	N0036 N0064 N0065 N0066 N0067
c.579G>A	Nonsense	p.Trp193X	A47°	17(31)	[14,16,56] unpubl.	N0024 N0025 N0026 N0037 N0038 N0039 N0060 N0068
c.604C>T	Nonsense	p.Arg202X	A47°	1(1)	[17]	*
c.612G>A	Nonsense	p.Trp204X	A47 ^f	1(2) ^f	[57]	*
c.678T>G	Nonsense	p.Tyr226X	A47°	1(2)	[17]	*

Nucleotide change	Mutation	Amino acid or mRNA change	CGD type	Families (alleles)	Reference	Accession number(s) ^a
c.682+1G>C ^e	Splice site	del. exon 7 p.Trp193_Gly228del	A47°	1(1)	[14]	N0062 N0063
c.730G>A	Missense	p.Glu244Lys	A47°	1(1)	unpubl.	*
c.734_748del115	Deletion	p.Val245_Glu249del	A47°	1(2)	unpubl.	*
c.784G>A	Missense	p.Gly262Ser	A47°	1(1)	[8]	N0033
c.789G>C	Missense	p.Trp263Cys	A47°	1(1)	unpubl.	*
c.811delG	Deletion	p.Val271SerfsX105	A47°	1(1)	[8]	
c.838delC	Deletion	p.Leu280CysfsX96	A47°	1(1)	[18]	*
Mutations in <i>NCF1</i>						
Number of different alleles			Total number of alleles			
Deletions	7 alleles		(30.4%)	7 alleles		(11.1%)
Nonsense mutations	6 alleles		(26.1%)	38 alleles		(60.3%)
Splice site mutations	5 alleles		(21.7%)	11 alleles		(17.5%)
Missense mutations	4 alleles		(17.4%)	6 alleles		(9.5%)
Deletion/insertions	1 allele		(4.4%)	1 allele		(1.6%)
Total 23 different allelic mutations (including delta-GT)			Total 42 families with 6 (other than delta-GT) in 3 identified alleles the 53 patients			

^a Accession number in database at <http://www.uta.fi/mt/bioinfo/NCF1base/>.

^b One patient is a compound heterozygote for this mutation and for an undefined chromosomal microdeletion on the other allele [58].

^c Activation of cryptic donor splice site in intron 2, leading to incorporation of 73 nucleotides from the 5' side of intron 2 into mRNA, including the mutated G>A at position +1 of intron 2. At the protein level, this mutation predicts incorporation of 24 aberrant amino acids after His51, followed by a stop codon at position 76 [14].

^d In addition, these patients show evidence of mRNA for p47phox from which the last 22 bp at the 3' region of exon 6 has been skipped (r.552_574del), as well as mRNA in which intron 6 has been included and in which the mutated exon 6 is expressed (r.[intron6+L_exon6-1ins; 574 g>q]) [14].

^e Positions of introns in *NCF1*: intron 1 c.72_73; intron 2 c.153_154; intron 3 c.229_230; intron 4 c.395_396; intron 5 c.451_452; intron 6 c.574_575; intron 7 c.682_683; intron 8 c.801_802; intron 9 c.905_906; intron 10 c.1051_1052.

^f Patient presumed to be homozygous for the mutation.

Table 4

Mutations in the p40-*phox* gene *NCF4*.

Nucleotide change	Mutation	Amino acid or mRNA change	CGD type	Families (alleles)	Reference
c.143_152dup10	Insertion	p.Lys52ArgfsX79	A40 ^o	1 (1)	[59] *
c.314C>A	Missense	p.Arg105Gln	A40 ⁺	1 (1)	[59] *

Table 5Polymorphisms in the p22-*phox* gene *CYBA*.

Polymorphic nucleotide	Amino acid change	Reference
c.59-37A/G	NA	[30]
c.36A/G	p.Glu12Glu	[60]
c.179A/C	p.Lys60Thr	[30]
c.214C/T	p.His72Tyr	[28,60]
c.288-138ins50	NA	[13]
c.381T/C	p.Arg127Arg	[60]
c.403G/A	p.Glu135Lys	[30]
c.480G/A	p.Pro160Pro	[30,37,60]
c.512A/G	p.Glu171Gly	[60]
c.521C/T	p.Ala174Val	[28,30,31,60]
c.579G/T	p.Glu193Asp	[60]
c.612A/G (+24 of 3' UT region)	NA	[37,60]

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Table 6Polymorphisms in the p67-*phox* gene *NCF2*.

Polymorphic nucleotide	Amino acid change	Reference
c.-185G/A	NA	[41,42]
c.-181G/A	NA	[41,42]
c.-24C/T	NA	[41,42]
c.235A/G	p.Met79Val	[41]
c.542A/G	p.Lys181Arg	[41,42,51]
c.606G/A	p.Ala202Ala	[42]
c.895C/T	p.Leu299Leu	[41,47,51]
c.925-21G/A	NA	[41]
c.983G/A	p.Arg328Lys	[41,47,51]
c.1105G/A	p.Gly369Arg	[42]
c.1167C/A	p.His389Gln	[41,42]
c.1183C/T	p.Arg395Trp	[41-43]

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Table 7Polymorphisms in the p47-*phox* gene *NCF1*.

Polymorphic nucleotide ^a	Amino acid change	Reference
c.66G/C	p.Glu22His	Unpubl.
c.73G/A	p.Val25Met	Unpubl.
c.345C/T	p.Leu115Leu	[8]
c.468C/T	p.Ile156Ile	Unpubl.
c.558A/G	p.Val186Val	Unpubl.
c.621G/A	p.Ala206Ala	Unpubl.
c.825C/T	p.Phe275Phe	Unpubl.
c.849A/G	p.Ser283Ser	Unpubl.
c.936C/T	p.His312His	Unpubl.

^aIdentification of polymorphic sites in *NCF1* is complicated by the p47-*phox* pseudogenes, which contain several differences from the functional gene; the referenced polymorphism was identified after amplification of *NCF1* with primers that do not bind to the pseudogenes [8]. More synonymous polymorphisms can be expected to be introduced into *NCF1* by recombination with the pseudogenes [20].

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Table 8Polymorphisms in the p40-*phox* gene *NCF4*.

Polymorphic nucleotide ^a	Amino acid change	Reference
c.32+1258G/T	N.A.	[61]
c.33-1101T/C	N.A.	[61]
c.33-728T/C	N.A.	[61]
c.118-360G/A	N.A.	[61]
c.342+202G/C	N.A.	[61]
c.342+342G/T	N.A.	[61]
c.342+1326G/A	N.A.	[61]
c.343-1378A/G	N.A.	[61]
c.343-339A/G	N.A.	[61]
c.528+16G/A	N.A.	[61]
c.627+711G/A	N.A.	[61]
c.627+1040G/T	N.A.	[61]
c.628-1193G/A	N.A.	[61]
c.758+57A/T	N.A.	[61]

^aPositions of introns in *NCF4*: intron 1 c.32_33; intron 2 c.117_118; intron 3 c.271_272; intron 4 c.342_343; intron 5 c.470_471; intron 6 c.528_529; intron 7 c.627_628; intron 8 c.758_759; intron 9 c.824_825.

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