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Hepatoprotective and antioxidant effects of *Monochoria vaginalis* against acetaminophen-induced hepatotoxicity in rats

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SUMMARY

The present study was aimed to investigate the hepatoprotective and antioxidant activities of ethanol extract from Monochoria vaginalis (250 mg/kg and 500 mg/kg B/W) on acetaminophen (APAP) induced rat hepatic injury. Monochoria vaginalis is a traditional medicinal plant that is commonly used to treat and improve liver conditions in India and other Asian countries. The development of hepatotoxicity induced by APAP is promoted by oxidative stress. APAP treated group significantly (P < 0.01) elevated the serum enzymatic levels like glutamate oxaloacetate transaminase, glutamate pyruvate transaminase, alkaline phosphatase (SALP), total bilirubin and malondialdehyde (MDA), which were restored towards normalization significantly (P < 0.01) thanol extract of yonochoria vagin is (EEMV). In addition, the EEMV significantly (P < 0.01) elevated the decreased level of total protein and antioxidant enzymes such as superoxide dismutase, catalase, glutathione peroxidase, glutathione-s-transferase and reduced glutathione. Apart from these, histopathological changes also showed the protective nature of the EEMV against APAP induced hepatic damage in liver tissues. The activity of EEMV at 500 mg/kg B/W was comparable to the standard drug silymarin (25 mg/kg B/W). In conclusion, these data suggest that the EEMV possess hepatoprotective and antioxidant effects against APAP-induced hepatotoxicity and oxidative stress in rats.

Key words: Hepatoprotective; Monochoria vaginalis; Acetaminophen; Silymarin; Antioxidant

INTRODUCTION

Monochoria vaginalis commonly known as carpet weed belongs to family Pontederiaceae and distributed through out India, Juice of leaves used to treat cough and that of roots to address stomach and liver problems, asthma and tooth ache (Yoganarasimman *et al.*, 2000; Madhava chetty *et al.*, 2008). In siddha system root is used to treat cough, disease of pittam, venereal disease, thirst, fainting and fever (Yoganarasimman *et al.*, 2000). The whole part of plant is considered as a functional food and also been employed for the treatment of asthma and fever (Row *et al.*, 2003). Further, this plant is also resistant to several ALS inhibitors and in addition the mechanism of resistance was investigated with *in vitro* and *in*

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vivo ALS assay (Hwang *et al.*, 2001). *Monochoria* vaginalis was the first sulfonyl-urea-resistant weed species reported by Itoh *et al* method (1999). In addition, it has been reported that n-butanol fraction of *Monochoria* vaginalis exhibited the highest antioxidant activity (Zhou Yong-Jun *et al.*, 2007).

Acetaminophen [n-acetyl-para-aminophenol; paracetamol, acetaminophen (APAP)] is one of the most widely used over-the-counter drugs commonly used for the relief of fever and headaches due to its antipyretic and analgesic properties, and is a major ingredient in cold and flu remedies. Though APAP is generally considered safe for human use at recommended doses, potentially fatal liver damages occurred when an acute over-dose was used or even a normal dose was taken by rare individuals. Accordingly, APAP overdose is one the most common causes of drug poisoning world-wide. Excessive use of APAP can cause multiple organ damages, especially the liver and kidney (Yapar et al., 2007; Palani et al., 2009; Palani et al., 2010). The present study was therefore, aimed to evaluate the hepatoprotective and antioxidant effects of ethanol extract from Monochoria vaginalis (EEMV) activity against acetaminophen induced hepatotoxicity.

MATERIALS AND METHODS

Plant material

Aerial part of *Monochoria vaginalis* (pontederiaceae) was collected from Tirunelveli district, Tamil Nadu, India in the month of March. The plant material was taxonomically identified and authenticated by V.Chelladurai (Research Officer) Botany (C.C.R.A.S) Government of India. Voucher specimen (AECBT-01/2007-2008) has been retained in the Anna bioresearch foundation, Arunai engineering college, Tiruvannamalai, Tamilnadu, India.

Animals

Studies were carried out using Wistar albino male rats (150 – 200 g), obtained from Indian Veterinary Preventive medicine (IVPM), Ranipet, Tamilnadu, India. The animals were grouped and housed in polyacrylic cages ($38 \times 23 \times 10$ cm) with not more than six animals per cage and maintained under standard laboratory conditions (temperature $25 \pm 2^{\circ}$ C) with dark and light cycle (12/12 h). The animals were fed with standard pellet diet supplied by poultry research station, Nandhanam, India and fresh water *ad libitum*. All the animals were acclimatized to laboratory condition for a week before commencement of experiment. All procedures described were reviewed and approved by the University animal's ethical committee.

Drugs and chemicals

Silymarin was purchased from Micro labs, Tamilnadu. and the rest of the chemicals utilized were of analytical grade and were obtained from Ranbaxy research laboratory, Hyderabad, India.

Extraction

The aerial part of *Monochoria vaginalis* was dried under shade and then powdered with a mechanical grinder to obtain a coarse powder. Equal quantity of powder was passed through 40 mesh sieve and extracted with ethanol (90% v/v) in soxhlet apparatus at 60°C (Chattopadhyay, 2003). The solvent was completely removed by rotary vacuum evaporator. The extract was freeze dried and stored in a vacuum desiccator's.

Experimental treatments

Animals were divided into five groups of six animals each. Group I treated with vehicle (distilled water) was kept as normal. Group II treated with acetaminophen (APAP) of 750 mg/kg body weight for 7 days orally and was kept as toxin control. Group III and IV were treated with EEMV at two different doses of 250 and 500 mg/ kg body wt plus APAP for 7 days. Group V were fed with standard drug silymarin 25 mg/kg plus APAP daily for seven days. The extract/silymarin was administered by oral gavages 1 h before APAP administration (Deepak et al., 2007).

Preparation of serum from blood

After 24 h, animals were sacrificed by chloroform anesthesia. Blood was collected by heart puncture. The blood samples of each animal were taken and allowed to clot for 45 min at room temperature. Serum was separated by centrifugation at $600 \times g$ for 15 min and analyzed for various biochemical parameters including serum glutamate oxaloacetate transaminases (SGOT), serum glutamate pyruvate transaminases (SGPT), (Reitman and Frankel, 1957) alkaline phosphatase (ALP), (King *et al.*, 1934) and bilirubin (Malloy *et al.*, 1937).

Preparation of liver homogenate

Hepatic tissues were homogenized in KCl [10 mM] phosphate buffer (1.15%) with ethylenediamine tetra acetic acid (EDTA; pH 7.4) and centrifuged at $12,000 \times g$ for 60 min. The supernatant was used for assay of the marker enzymes [glutathione peroxidase, glutathione-s-transferase, superoxide dismutase and Catalase (CAT)], reduced glutathione, malondialdehyde (MDA) content and protein estimation.

Biochemical estimation of markers of oxidative stress

MDA content was Measured according to the earlier

method reported (Zhang, 1992). superoxide dismutase (SOD) activity was determined according to previous report (Rai *et al.*, 2006). Catalase (CAT) activity was determined from the rate of decomposition of H_2O_2 by the reported method (Bergmeyer *et al.*, 1974). Glutathione peroxidase (GPX) activity was determined by measuring the decrease in glutathione (GSH) content after incubating the sample in the presence of H_2O_2 and NaN₃ (Hafemann *et al.*, 1974). Reduced glutathione was measured according to the method of Ellman (1959). Protein content in the tissue was determined by earlier method reported (Lowry *et al.*, 1951), using bovine serum albumin (BSA) as the standard.

Histopathological study

On completion of closing regimen animals were sacrificed and the liver dissected out. Paraffin sections were prepared for histological examination and following standard procedure (Galighor and Kozloff, 1976). Hematoxylin-eosin stained sections were observed.

Statistical analysis

The obtained results were analyzed for statistical significance using one way ANOVA followed by Dunnet test statistical software for comparison with control group and acetaminophen treated

Table 1. Effect of ethanol extract of Monochoria vaginalis and silymarin on serum enzymes (SGPT, SGOT and
ALP), total bilirubin and total protein on acetaminophen induced hepatotoxicity in rats. Values are mean ± S.D. (n
= 6). ** $P < 0.01$, * $P < 0.05$, respectively.

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Groups and treatment	Dose	SGOT	SGPT	SALP	Total Bilirubin	Total Protein
Groups and treatment	(mg/kg)	(IU/L)	(IU/L)	(IU/L)	(mg/dl)	(mg/dl)
Group I- Normal (NaCl 0.9% w/v)	5 ml/kg	45.20 ± 1.47	48.16 ± 2.73	14.00 ± 0.45	1.64 ± 0.05	7.5 ± 0.18
Group II Acetami- nophen Control	750 mg/kg	$110 \pm 4.28^{**}$	97.83 ± 2.75**	28.68 ± 1.23**	$6.42 \pm 0.22^{**}$	$5.01 \pm 0.19^{**}$
Group III- EEMV + Acetaminophen	250 mg/kg + 750 mg/kg	98.63 ± 2.00*	88.26 ± 2.38*	$24.64 \pm 0.55^*$	$5.55 \pm 0.07^{**}$	$5.18 \pm 0.15^{*}$
Group IV- EEMV + Acetaminophen	500 mg/kg + 750 mg/kg	79.65 ± 1.66**	71.48 ± 2.47**	16.36 ± 0.29**	3.41 ± 0.31**	$5.43 \pm 0.09^{**}$
Group V- Silymarin + acetaminophen	25 mg/kg + 750 mg/kg	60.26 ± 2.59**	55.17 ± 1.10**	16.09 ± 0.43**	$1.90 \pm 0.05^{**}$	6.51 ± 0.13**

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group. P < 0.05 was considered as significant.

RESULTS

The effect of EEMV on serum marker enzymes is presented in Table 1, Figs. 1, 2 and 4. The serum levels of GOT, GPT, ALP and total bilirubin were markedly significantly (P < 0.01) elevated and that of protein levels were significantly (P < 0.01) decreased in acetaminophen treated animals,

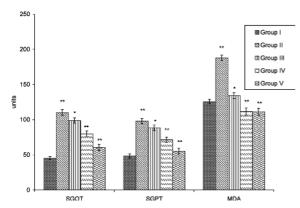


Fig. 1. Effect of ethanolic extract of *Monochoria vaginalis* and silymarin (standard drug, (25 mg/kg)) on serum levels of SGOT (IU/l), SGPT (IU/l) and MDA (nM/mg of protein) [Lipid peroxidation (LPO)] level of hepatic tissue during acetaminophen treated hepatotoxicity and oxidative stress in rats. Values are mean \pm S.D. (n = 6). ** P < 0.01, * P < 0.05, respectively.

indicating liver damage. Administration of ethanol extract of *Monochoria vaginalis* at the doses of 250 and 500 mg/kg remarkably significantly (P < 0.05; P < 0.01) prevented hepatotoxicity induced by acetaminophen.

Acetaminophen treatment caused a significant (P < 0.01) decrease in the level of SOD, catalase,

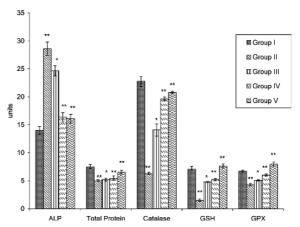


Fig. 2. Effect of ethanolic extract of *Monochoria vaginalis* and silymarin (standard drug, (25 mg/kg)) on serum levels of alkaline phosphatase (ALP) (IU/L) & total protein and hepatic levels of CAT (U/mg protein), GSH (U/mg protein) and GPX (mg of glutathione utilized/min/mg protein) during acetaminophen treated hepatotoxicity and oxidative stress in rats. Values are mean ± S.D. (n = 6). ** P < 0.01, * P < 0.05, respectively.

Table 2. Effect of ethanol extract of *Monochoria vaginalis* and silymarin on antioxidants levels (SOD, CAT, MDA, GSH, GPX & GST) in acetaminophen induced heapatotoxicity in rats. Values are mean \pm S.D. (n = 6). ** P < 0.01, * P < 0.05, respectively.

Parameters	Group I- Nor- mal (NaCl 0.9% w/v)	Group II- Ace- taminophen Control(750 mg/kg)	taminophen	Group IV- EEMV + Ace- taminophen (500 mg/kg + 750 mg/kg)	Group V- Sily- marin + Ace- taminophen (25 mg/kg + 750 mg/kg)
SOD(units of activity/mg protein)	0.99 ± 0.04	$0.65 \pm 0.06^{**}$	$0.74 \pm 0.01^*$	$0.84 \pm 0.06^{**}$	$1.03 \pm 0.02^{**}$
CAT (micromoles of H ₂ O ₂	22.83 ± 4.66	$6.28 \pm 0.46^{**}$	$14.08\pm0.47^{*}$	$19.60 \pm 1.02^{**}$	$20.80 \pm 1.02^{**}$
decomposed/mg protein/min)					
LP (nanomoles of MDA formed/mg protein/h)	125.5 ± 9.41	187.83 ± 4.95**	134.12 ± 3.54*	111.60 ± 6.55**	111.17 ± 8.40**
GSH (μg/mg protein)	7.07 ± 1.18	$1.52 \pm 0.27^{**}$	$4.78 \pm 0.87^{*}$	5.19 ± 1.77**	$7.64 \pm 1.49^{**}$
GPX (micrograms of glutathione utilized/min/mg protein)	6.72 ± 0.81	4.32 ± 0.27**	$5.06 \pm 1.10^{*}$	$6.02 \pm 0.48^{**}$	$7.98 \pm 0.04^{**}$
GST(Units/mg protein)	0.39 ± 0.08	$0.17 \pm 0.08^{**}$	$0.29\pm0.05^{*}$	$0.33 \pm 0.07^{**}$	$0.38 \pm 0.02^{**}$

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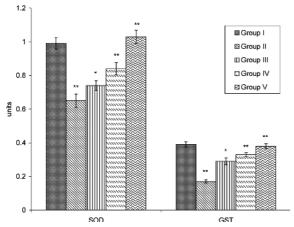


Fig. 3. Effect of ethanolic extract of *Monochoria vaginalis* and silymarin (standard drug, (25 mg/kg) on hepatic levels of SOD (units of activity/mg protein) & GST (Units/mg protein) during acetaminophen treated hepatotoxicity and oxidative stress in rats. Values are mean \pm S.D. (n = 6). ** P < 0.01, * P < 0.05, respectively.

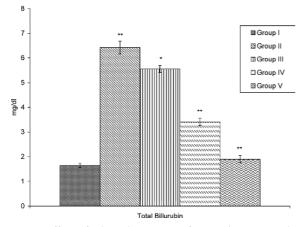


Fig. 4. Effect of ethanolic extract of *Monochoria vaginalis* and silymarin (standard drug, (25 mg/kg)) on serum levels of total bilurubin (mg/dl) during acetaminophen treated hepatotoxicity and oxidative stress in rats. Values are mean \pm S.D. (n = 6). ** P < 0.01, * P < 0.05, respectively.

GPX, GSH and glutathione-s-transferase (GST) in liver tissue when compared with control group. The treatment of EEMV at the doses of 250 and 500 mg/kg resulted in a significant (P < 0.05; P <0.01) increase of SOD, catalase, GPX, GSH and GST when compared to Group II (Table 2; Figs. 2 and 3). The standard drug, silymarin treated animals also showed a significant (P < 0.01) increase in antioxidant enzymes levels compared to Group II. Analysis of MDA levels by thiobarbituric acid reaction showed a significant (P < 0.01) increase in the acetaminophen treated rats. Treatment with EEMV (250 mg/kg and 500 mg/kg) significantly (P < 0.01; P < 0.01) prevented the increase in MDA level which was brought to near normal (Fig. 1).

Normal photomicrograph of liver tissue of control rat showing normal hepatic cells with

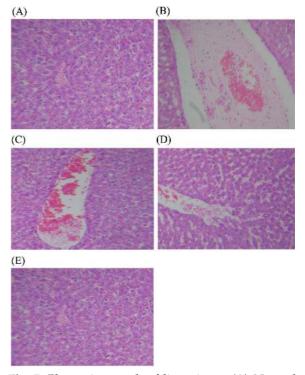


Fig. 5. Photomicrograph of liver tissue. (A) Normal photomicrograph of liver tissue of control rat showing normal hepatic cells with central vein and sinusoidal dilation (H & E, 100×). (B) The acetaminophen treated animals showed severe centrilobular necrosis and fatty infiltration. (C) Histology of liver from rat which received Monochoria vaginalis ethanolic extract at 250 mg/kg. (Group III) showing mild degenerative changes and absence of centrilobular necrosis (H & E, 100×). (D) Histology of liver from rat which received Monochoria vaginalis ethanolic extract at 500 mg/kg (Group IV) showing normal hepatocytes with mild inflammation (H & E, 100×). E) Liver section of Rat treated with silymarin at 25 mg/kg showed less vacule formation reduced sinusoidal dilation, less disarrangements and degeneration of hepatocytes (H & E, 100×).

central vein and sinusoidal dilation (Fig. 5A). The acetaminophen treated animals showed severe centrilobular necrosis and fatty infiltration (Fig. 5B). Histology of liver from rat which received *Monochoria vaginalis* ethanolic extract at 250 mg/kg (Group III) showing mild degenerative changes and absence of centrilobular necrosis (Fig. 5C).

Histology of liver from rat which received Monochoria vaginalis ethanolic extract at 500 mg/kg (Group IV) showing normal hepatocytes with mild inflammation (Fig. 5D). Liver section of rat treated with silymarin at 25 mg/kg showed less vacule formation reduced sinusoidal dilation, less disarrangements and degeneration of hepatocytes (Fig. 5E). All these results indicate a hepatoprotective potential by the EEMV.

DISCUSSION

Acetaminophen (*N*-acetyl-*p*-aminophenol, Paracetamol), a widely used analgesic and antipyretic drug is known to cause hepatotoxicity in experimental animals and humans at high doses (Prescott *et al.*, 1971; Mitchell, 1988; Kuma and Rex, 1991; Eriksson *et al.*, 1992; Thompsen *et al.*, 1995). The laboratory features of hepatotoxicity induced by APAP resemble other kinds of acute inflammatory liver disease with prominent increase of GOT, GPT, and ALP levels (Davidson and Eastham, 1966).

In the present study, the serum level of hepatic enzymes GOT, GPT, ALP and total bilirubin levels were increased and reflected the hepatocellular damage in the APAP-induced hepatotoxicity animal model. This is indicative of cellular leakage and loss of functional integrity of cell membrane in liver (Drotman *et al.*, 1978). However the total protein level was decreased. There was a significant (P < 0.01) restoration of these enzyme levels on administration of the EEMV in a dose dependent manner and also by silymarin at a dose of 25 mg/kg. The reversal of increased serum enzymes in acetaminophen induced liver damage by the EEMV may be due to the prevention of the leakage of intracellular enzymes by its membrane stabilizing activity. This is in agreement with the commonly accepted view that serum levels of transaminases return to normal with the healing of hepatic parenchyma and the regeneration of hepatocytes (Thabrew and Joice, 1987; Maiti *et al.*, 2005).

Effective control of ALP, bilirubin and total protein levels points towards an early improvement in the secretary mechanism of the hepatic cells, as well as repair of hepatic tissue damage caused by APAP. This indicates the anti-lipid per oxidation and/or adaptive nature of the systems as brought about by EEMV against the damaging effects of free radical produced by APAP.

Previous studies have demonstrated that oxidative stress is a major mechanism in the development of APAP-induced hepatotoxicity (Lin et al., 1998; Ahmed and Khater, 2001; Shanmugasundaram and Venkataraman, 2006). In the present study, the data suggested that high dosage of APAP in the liver could lead to decreased levels of antioxidant enzymes (SOD, CAT, GPx) and present a significant level of hepatotoxicity in the course of the treatment. However, the EEMV could raise the levels of SOD, CAT, and GPX against the APAP-induced oxidative stress mediated by ROS and RNS. Both reductions of GST & GSH activity APAP-treated rats as observed in this study indicate the damage to the hepatic cells. Administration of EEMV extract promoted the reactivation of hepatic glutathione reductase enzyme in APAPtreated rats. The restoration of GSH level to such APAP treated rats due to the protective effect of after the administration of EEMV. Furthermore, the level of MDA was increased in the group receiving APAP administration, but treatment with the EEMV reduced the amount of MDA. This result indicated that decreasing the formation of lipid peroxidation is also one of the events in preventing the oxidative toxicity by APAP.

In conclusion, ethanol extract of Monochoria

vaginalis significantly protects against liver injuries as well as oxidative stress, resulting in improved serum biochemical parameters such as SGOT, SGPT and alkaline phosphatase (SALP). The reduced levels of parameters of SOD, CAT, GSH, GPX, and GST in acetaminophen-treated rats were significantly increased by treatment with EEMV.

These findings suggest the potential use of the EEMV as a novel therapeutically useful hepatoprotective agent. Further studies to characterize the active principles and to elucidate the mechanism are in progress.

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