



Hepcidin and Anemia: A Tight Relationship

Alessia Pagani¹, Antonella Nai^{1,2}, Laura Silvestri^{1,2} and Clara Camaschella^{1*}

¹Division of Genetics and Cell Biology, San Raffaele Scientific Institute, Milan, Italy, ²Vita-Salute San Raffaele University, Milan, Italy

Hepcidin, the master regulator of systemic iron homeostasis, tightly influences erythrocyte production. High hepcidin levels block intestinal iron absorption and macrophage iron recycling, causing iron restricted erythropoiesis and anemia. Low hepcidin levels favor bone marrow iron supply for hemoglobin synthesis and red blood cells production. Expanded erythropoiesis, as after hemorrhage or erythropoietin treatment, blocks hepcidin through an acute reduction of transferrin saturation and the release of the erythroblast hormone and hepcidin inhibitor erythroferrone. Quantitatively reduced erythropoiesis, limiting iron consumption, increases transferrin saturation and stimulates hepcidin transcription. Deregulation of hepcidin synthesis is associated with anemia in three conditions: iron refractory iron deficiency anemia (IRIDA), the common anemia of acute and chronic inflammatory disorders, and the extremely rare hepcidin-producing adenomas that may develop in the liver of children with an inborn error of glucose metabolism. Inappropriately high levels of hepcidin cause iron-restricted or even iron-deficient erythropoiesis in all these conditions. Patients with IRIDA or anemia of inflammation do not respond to oral iron supplementation and show a delayed or partial response to intravenous iron. In hepcidin-producing adenomas, anemia is reverted by surgery. Other hepcidin-related anemias are the “iron loading anemias” characterized by ineffective erythropoiesis and hepcidin suppression. This group of anemias includes thalassemia syndromes, congenital dyserythropoietic anemias, congenital sideroblastic anemias, and some forms of hemolytic anemias as pyruvate kinase deficiency. The paradigm is non-transfusion-dependent thalassemia where the release of erythroferrone from the expanded pool of immature erythroid cells results in hepcidin suppression and secondary iron overload that in turn worsens ineffective erythropoiesis and anemia. In thalassemia murine models, approaches that induce iron restriction ameliorate both anemia and the iron phenotype. Manipulations of hepcidin might benefit all the above-described anemias. Compounds that antagonize hepcidin or its effect may be useful in inflammation and IRIDA, while hepcidin agonists may improve ineffective erythropoiesis. Correcting ineffective erythropoiesis in animal models ameliorates not only anemia but also iron homeostasis by reducing hepcidin inhibition. Some targeted approaches are now in clinical trials: hopefully they will result in novel treatments for a variety of anemias.

Keywords: anemia, iron, hepcidin, erythropoiesis, inflammation

OPEN ACCESS

Edited by:

Anna Bogdanova,
University of Zurich, Switzerland

Reviewed by:

Kostas Pantopoulos,
McGill University, Canada
Elizabeta Nemeth,
David Geffen School of Medicine at
UCLA, United States

*Correspondence:

Clara Camaschella
camaschella.clara@hsr.it

Specialty section:

This article was submitted to
Red Blood Cell Physiology,
a section of the journal
Frontiers in Physiology

Received: 31 August 2019

Accepted: 25 September 2019

Published: 09 October 2019

Citation:

Pagani A, Nai A, Silvestri L and
Camaschella C (2019) Hepcidin and
Anemia: A Tight Relationship.
Front. Physiol. 10:1294.
doi: 10.3389/fphys.2019.01294

INTRODUCTION

Anemia is one of the most common disorders worldwide and anemia due to iron deficiency is the prevalent form according to multiple analyses (review in Camaschella, 2019). This type of anemia results from the total body iron deficiency and the inability to supply the large amount of iron that the bone marrow consumes to produce an adequate number of red blood cells in order to maintain tissue oxygenation.

The iron availability is controlled by the liver peptide hormone hepcidin. The body iron increase causes the production of hepcidin, which is released in the circulation and acts on its receptor ferroportin, a transmembrane iron exporter protein highly expressed on enterocyte, macrophages, and hepatocytes. Hepcidin reduces the iron entry to plasma from absorptive duodenal cells and iron recycling macrophages by blocking iron export (Aschemeyer et al., 2018) and by degrading the iron exporter ferroportin (Nemeth et al., 2004). By regulating plasma iron and systemic iron homeostasis, the hepcidin/ferroportin axis strongly affects erythropoiesis, hence the possible development of anemia.

THE IRON-ERYTHROPOIESIS CONNECTION

The process of red blood cells production consumes approximately 80% of circulating iron for hemoglobin synthesis of maturing erythroblasts. Most iron (20–25 mg/daily) is recycled by macrophages, while a limited amount (1–2 mg daily) derives from intestinal absorption. The kidney hormone erythropoietin (EPO) controls the proliferation of erythroid progenitors, especially of CFU-e and at a lower degree of BFU-e, and the early phase of terminal erythropoiesis, while iron needs are increased in the late differentiation stages from proerythroblasts to reticulocyte, for the synthesis of heme incorporated into hemoglobin (Muckenthaler et al., 2017).

Hepcidin regulation requires a crosstalk between liver endothelial sinusoidal cells (LSEC) that produce the bone morphogenetic proteins (BMPs) to activate the BMP-SMAD pathway and hepatocytes that produce and release hepcidin (Babitt et al., 2006; Rausa et al., 2015). BMP6 and BMP2 are the most important BMPs that upregulate hepcidin, while BMP6 expression is iron dependent (Andriopoulos et al., 2009; Meynard et al., 2009) BMP2 appears less iron-responsive (Canali et al., 2017; Koch et al., 2017).

Hepcidin levels are low in absolute iron deficiency and iron deficiency anemia. In these conditions, the iron stores are exhausted and the BMP-SMAD signaling is switched off at multiple levels. First, BMP6 expression is suppressed; next, the activity of *TMPRSS6*, a protease that cleaves the BMP co-receptor hemojuvelin (Silvestri et al., 2008), is strongly increased (Lakhal et al., 2011); and third, histone deacetylase3 (HDAC3) suppresses the hepcidin locus (Pasricha et al., 2017). In conditions of iron deficiency, the reduction of hepcidin production is an adaptation mechanism that facilitates dietary and pharmacological iron absorption (Camaschella and Pagani, 2018).

When anemia is severe, the coexisting hypoxia stimulates erythropoiesis through increased kidney synthesis and release of EPO. This leads to suppression of hepcidin transcription by erythroferrone (ERFE), an EPO target gene produced by erythroblasts (Kautz et al., 2014), by molecules (e.g., PDGF-BB) released by other tissues (Sonnweber et al., 2014), and likely by soluble components of transferrin receptors (TFR), sTFR1 (Beguin, 2003), and sTFR2 (Pagani et al., 2015). The final aim is to supply enough iron for the needs of an expanded erythropoiesis.

ANEMIAS WITH ABNORMAL HEPCIDIN LEVELS

Anemias may be classified on the basis of hepcidin levels as anemias with high and low hepcidin. It is intuitive that persistently high hepcidin levels, by blocking iron absorption, cause iron deficiency anemia because of decreased iron supply to erythropoiesis. Conversely, ineffective erythropoiesis characterizes the so-called *iron-loading anemias* that have low hepcidin levels and iron overload. These two groups of anemias are the outcome of opposite pathophysiology mechanisms (**Figure 1**). In the first group, anemia is due to the inhibitory effect exerted by hepcidin on iron absorption and recycling that leads to systemic iron deficiency; in the second group, anemia is due to hepcidin suppression by an expanded abnormal erythropoiesis (Camaschella and Nai, 2016).

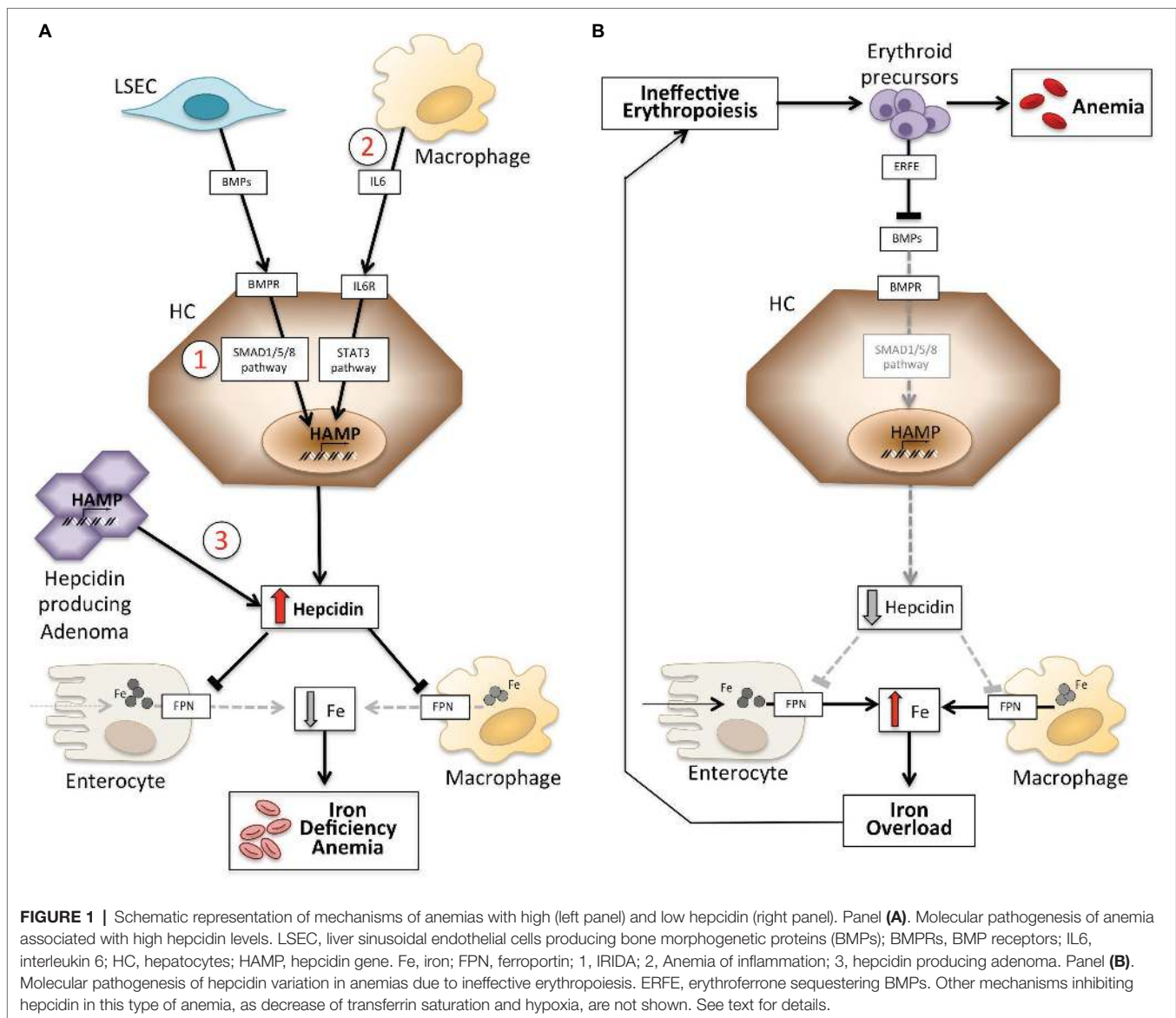
Anemia Associated With High Hepcidin Levels

This group includes two inherited rare disorders (iron refractory iron deficiency anemia and hepcidin-producing adenomas in an inborn error of glucose metabolism) and an acquired common condition: anemia of inflammation (**Table 1**).

Iron Refractory Iron Deficiency Anemia

Iron refractory iron deficiency anemia (IRIDA) is a rare recessive disorder characterized by hypochromic microcytic anemia, low transferrin saturation, and inappropriately normal/high hepcidin levels. It is caused by mutations of *TMPRSS6* (Finberg et al., 2008), a gene that encodes the type II serine protease, matriptase-2 (Du et al., 2008). Mutations of *TMPRSS6* are spread along the gene and may affect different domains especially the catalytic domain (De Falco et al., 2014). This transmembrane protease, highly expressed in the liver, inhibits hepcidin transcription by cleaving the cell surface BMP co-receptor hemojuvelin, thus attenuating the BMP signaling and hepcidin synthesis (Silvestri et al., 2008). *TMPRSS6* function is essential in iron deficiency to allow the compensatory mechanism of increased iron absorption.

IRIDA is present since birth and usually diagnosed in childhood. Compared with classic iron deficiency, iron parameters are atypical and raise the suspicion of the disease. The percent saturation of transferrin is strongly reduced (less than 10%) as in other forms of iron deficiency; however, at variance with iron deficiency, levels of serum ferritin are normal/increased (Camaschella, 2013; De Falco et al., 2013). This reflects an increased ferritin accumulation in macrophages, due to the high hepcidin levels that induce store iron sequestration.



None of the tests proposed for IRIDA diagnosis covers 100% of the cases. The genetic test identifies that *TMPRSS6* mutations, that in some cases (non-sense, frame-shift, and splicing mutations), are clearly causal. In other cases, as for previously unreported missense mutations, functional studies are needed to demonstrate causality (Silvestri et al., 2013). However, these tests are scarcely available. Serum hepcidin levels are usually increased/normal, independently of iron deficiency, and consistent with high/normal ferritin. It is important to exclude inflammation by concomitantly dosing C-reactive protein.

Some patients with a phenotype of refractory iron deficiency have been reported to have a single *TMPRSS6* mutated allele; here, the debate is whether they should be considered IRIDA or not. A spectrum of conditions can be envisaged ranging from classic severe IRIDA due to homozygous or compound heterozygous *TMPRSS6* mutations to increased susceptibility to iron deficiency conferred by single mutations/polymorphic changes.

One approach proposed to predict classic IRIDA is hepcidin normalization on other iron parameters, as ratios transferrin saturation (Tsat)/log hepcidin or Tsat/log Ferritin (Donker et al., 2016). According to other authors, most patients with a severe IRIDA phenotype have biallelic *TMPRSS6* mutations and, when unidentified, the second allele may be genetically occult (Heeney et al., 2018). In general terms, subjects with a single allele have a milder phenotype than those with two mutations and respond better to iron treatment (Donker et al., 2016). Interestingly, several *TMPRSS6* SNPs have been shown to provide susceptibility to iron deficiency in some populations (An et al., 2012) and in blood donors (Sorensen et al., 2019).

A digenic inheritance has been reported in a 5-year-old female originally found to have an atypical IRIDA genotype with one *TMPRSS6* (I212T) causal and one (R271Q) silent mutation (De Falco et al., 2014). She was later diagnosed *Fibrodysplasia ossificans progressiva* (FOP), a rare dominant

TABLE 1 | Anemias classified according to hepcidin levels.

High-hepcidin anemias		
<i>Hereditary</i>	OMIM n.	Prevalence
Iron refractory iron deficiency anemia (IRIDA)	#206200	Rare
Hepcidin-producing adenomas*	#232200	Rare
<i>Acquired</i>		
Anemia of acute inflammation		Common**
Anemia of chronic inflammation (anemia of chronic disease)		Common
Low-hepcidin anemias		
<i>Hereditary – iron loading anemias</i>	OMIM n.	
β-thalassemia	#613985	Common***
Congenital dyserythropoietic anemia	#224100	Rare
Sideroblastic anemias	#300751	Rare
<i>Acquired</i>		
Low risk MDS with ringed sideroblasts		Rare

OMIM, online Mendelian Inheritance In Man; MDS, myelodysplastic syndromes. *Described in glycogen-storage-disease 1a.

**In hospitalized patients and in intensive care units.

***In people of Mediterranean or southern-east Asian origin.

disorder with ectopic bone formation in soft tissues due to mutated BMP type I receptor gene *ACVR1*, encoding ALK2 (Shore et al., 2006). The pathological allele *ALK2*^{R258S} is constitutively active since the mutation affects the glycine-serine-rich domain of the gene and renders the BMP/SMAD pathway overactive being unable to bind its specific inhibitor FKBP12 (Pagani et al., 2017).

This rare case is especially illustrative. First, since the ALK2 glycine-serine-rich domain interacts with FKBP12 and the mutation destabilizes the binding, it has revealed a previously unsuspected role for FKBP12 as a modulator of liver ALK2 and hepcidin (Colucci et al., 2017). Second, it has led to identify a link between activation of bone and liver BMP type I receptors. Third, the case strengthens the relevance of intact *TMPRSS6* in controlling the hepatic BMP/SMAD signaling, since no IRIDA was identified among other FOP patients with the same *ACVR1* mutation and presumably normal *TMPRSS6* (Pagani et al., 2017). Finally, this case is consistent with the concept that *TMPRSS6* haploinsufficiency cannot cause classic IRIDA.

The optimal treatment of IRIDA is undefined. Oral iron is ineffective, since it is not absorbed. The addition of vitamin C allows sporadic response. Intravenous iron induces a partial response usually at a slower rate in comparison with patients with acquired iron deficiency. EPO is ineffective in classic cases (De Falco et al., 2013; Heeney and Finberg, 2014).

Anemia of Hepcidin-Producing Adenomas

This is an extremely rare condition in adult patients affected by *glycogen storage disease 1a*, a recessive disorder due to deficiency of glucose-6 phosphatase, which catalyzes a reaction involved in both glycogenolysis and gluconeogenesis. A common dangerous disease symptom is hypoglycemia. The current treatment leads to prolonged survival of affected children up to adult age with the occurrence of several complications, such as anemia and liver adenomas. Anemia is microcytic and hypochromic, iron deficient, and refractory to oral iron treatment.

Anemia reverted after surgical adenoma resection. Adenoma tissue was found positive for hepcidin mRNA, while normal surrounding tissue showed hepcidin suppression, as expected because of the ectopic uncontrolled hepcidin production (Weinstein et al., 2002). The hematological features of patients resemble those of IRIDA as they share high hepcidin levels as a common mechanism of anemia.

Anemia of Inflammation

Anemia of inflammation (AI), previously known as anemia of chronic diseases, is a moderate normochromic-normocytic anemia that develops in conditions of systemic inflammation and immune activation. It occurs in several common disorders, including chronic infections, autoimmune diseases, advanced cancer, chronic kidney disease, congestive heart failure, chronic obstructive pulmonary disease, anemia of the elderly (at least partly), and graft versus host disease. AI is one of the most common anemias worldwide and the most frequent anemia in hospitalized patients. Acute inflammation contributes to the severity of anemia in intensive care units. Molecular mechanisms underlying AI are multiple and complex. Overproduction of cytokines such as IL1-β, TNF-α, and IL-6 by macrophages and INF-γ by lymphocytes blunts EPO production, impairs the erythropoiesis response, increases hepcidin levels, and may activate erythrophagocytosis, especially in the acute forms (Weiss and Goodnough, 2005; Ganz, 2019).

Hepcidin is activated by IL-6 through IL-6 receptor (IL-6R) and JAK2-STAT3 signaling. Full hepcidin activation requires an active BMP-SMAD pathway because inactivation of BMP signaling decreases hepcidin in animal models of inflammation (Theurl et al., 2011). The deregulation of systemic iron homeostasis causes macrophage iron sequestration and reduced absorption and recycling that leads to low saturation of transferrin and iron restriction of erythropoiesis and other tissues.

Traditional treatment of AI is based on reversibility/control of the underlying disease, whenever possible. If the disease is untreatable and anemia is mild, a careful evaluation of risks-benefits is needed to avoid side effects of any treatment. Pathophysiology-based treatments are limited to erythropoietin-like compounds and iron. The use of erythropoiesis stimulating agents (ESA) suppresses hepcidin by inducing erythropoiesis expansion. This approach is widely used in patients with chronic kidney disease, low-risk myelodysplastic syndromes, and cancer undergoing chemotherapy. However, a careful clinical control is necessary because high doses have cardiovascular side effects. The administration of intravenous iron may relieve iron restriction, caused by ESA-dependent expansion of erythropoiesis. Oral iron is usually ineffective since the high hepcidin levels counteract its intestinal absorption. Inhibitors of prolyl hydroxylase (hypoxia inducible factor, HIF stabilizers) are experimental in chronic kidney disease, to the aim of increasing endogenous EPO. Chronic treatment with red blood cells transfusions is not recommended because of transient effect and adverse reactions; it is limited to severe refractory anemia (Camaschella, 2019; Weiss et al., 2019).

Anemias Associated With Low Hepcidin Levels

Ineffective erythropoiesis and low or inappropriately normal hepcidin levels, with consequent iron overload, are typical features of the “iron-loading anemias.” The prototype is β -thalassemia, a genetic recessive disease due to β -globin gene mutations that cause anemia and excess α -globin chain production. The latter precipitates as hemichromes in the bone marrow, damaging maturing erythroid precursors and leading to ineffective erythropoiesis. This occurs in non-transfusion-dependent thalassemia or thalassemia intermedia, whose erythropoiesis is characterized by the prevalence of immature cells that release erythroferrone to inhibit liver hepcidin expression. Hepcidin levels are usually greater in transfusion-dependent thalassemia, where endogenous ineffective erythropoiesis is at least partially suppressed by transfusions (Camaschella and Nai, 2016).

Hepcidin suppression is mediated by the increased cytokine erythroferrone (ERFE), a member of the TNF- α family encoded by *ERFE* gene, synthesized by erythroblasts upon EPO stimulation (Kautz et al., 2014). ERFE is released into the circulation and sequesters BMPs, especially BMP6 (Arezes et al., 2018), attenuating the hepcidin signaling in response to iron. In addition, an epigenetic suppression occurs at the hepcidin locus by histone deacetylase HDAC3 (Pasricha et al., 2017). When anemia causes hypoxia, other mediators such as PDGF-BB (Sonnweber et al., 2014), which is released by different cell types, suppress hepcidin.

Hepcidin levels are decreased by a special mechanism in low-risk myelodysplasia with ringed sideroblasts, a clonal disorder due to mutations of the spliceosome gene *SF3B1*. Iron accumulates in mitochondria, leading to ineffective erythropoiesis and systemic iron overload. An abnormally spliced, elongated ERFE protein is more powerful than wild type ERFE in suppressing hepcidin (Bondu et al., 2019) and causing transfusion-independent iron loading.

TARGETED THERAPIES FOR HEPCIDIN-RELATED ANEMIAS

The identification of molecular mechanisms responsible of the previously discussed anemias has stimulated research in developing targeted therapies to replace current symptomatic treatment (Sebastiani et al., 2016; Crielaard et al., 2017). Approaches differ according to the type of anemia and the aim of decreasing or increasing hepcidin levels or their effects (Table 2).

Experimental Therapies to Decrease Hepcidin Levels/Increase Ferroportin Function

Except for hepcidin producing tumors, which have to be surgically removed, compounds that antagonize hepcidin or its effects may be useful in all anemias characterized by high hepcidin levels. Their main application would be in chronic inflammatory diseases in order to reverse hypoferrremia and anemia. Several experimental therapies aimed at manipulating the hepcidin

TABLE 2 | Experimental therapies targeting the hepcidin-ferroportin axis.

	Mechanism	Compounds
Compounds that decrease hepcidin or increase ferroportin function		
Class I	Reduction of the signaling pathway stimulating hepcidin	Anti IL6-R, anti IL-6 Anti-BMP6 MoAb* BMPR inhibitors Anti-HJV MoAb Non anticoagulant heparins
Class II	Hepcidin binders	Anti-HAMP MoAb Oligonucleotides aptamers
Class III	Interfering with hepcidin-FPN interaction	Anti-FPN MoAb, GDP
Compounds that increase hepcidin or decrease ferroportin function		
Class I	Hepcidin mimics	Hepcidin analogues* Minihepcidin
Class II	Activating hepcidin Blocking the hepcidin inhibitor Blocking the hepcidin receptor	BMPs (preclinical studies) Anti-TMPRSS6 (siRNA, ASO*) FPN Inhibitors*
Class III	Others	Human transferrin infusions Protoporphyrin IX (inhibition of HO) Bone marrow TFR2 inactivation

*BMPR, BMP receptor; HAMP, hepcidin gene; HJV, hemojuvelin; MoAb, monoclonal antibody; FPN, ferroportin; siRNA, small interfering RNA; ASO, antisense oligonucleotides; GDP, guanosine 5' diphosphate; HO, heme oxygenase; TFR2, transferrin receptor 2. Compounds indicated by * are in clinical trials.*

pathway and its function have been investigated in preclinical studies. Hepcidin antagonists are inhibitors of hepcidin synthesis/regulators (Ganz, 2019), hepcidin binders that block its function, and compounds that interfere with hepcidin-ferroportin interaction (Table 2). Some compounds are in clinical trials especially in chronic kidney disease (Sheetz et al., 2019). In IRIDA, manipulation of the hepcidin pathway has been proposed in preclinical studies with the use of anti-HJV MoAb (Kovac et al., 2016).

Experimental Therapies to Increase Hepcidin Levels/Decrease Ferroportin Function

Increasing hepcidin levels may not only reduce iron overload but also partially control ineffective erythropoiesis in *iron loading anemias*. β -thalassemia is the most studied among these conditions (Casu et al., 2018; Gupta et al., 2018). Proposed drugs are hepcidin analogs (some in clinical trials), hepcidin modulators, especially TMPRSS6 inhibitors, or compounds that interfere with hepcidin-ferroportin interaction decreasing iron export (Table 2).

While compounds that increase hepcidin reduce ineffective erythropoiesis due to the vicious cycle between ineffective erythropoiesis and iron loading (Camaschella and Nai, 2016), drugs that favor erythroid precursor maturation, as the activin receptor IIB ligand trap, luspatercept, not only improve anemia but also ameliorate iron homeostasis by reducing hepcidin inhibition (Piga et al., 2019).

Some targeted approaches now in clinical trials will hopefully result in novel treatments for a variety of anemias.

CONCLUSION

The spectacular advances in understanding the regulation of iron metabolism and hepcidin allowed a better understanding of erythropoiesis control, since together with erythropoietin iron is a fundamental factor for erythroid cells maturation. Conditions that lead to anemia can be associated with high and low hepcidin levels. In both instances, contrasting hepcidin deregulation may ameliorate/correct anemia in preclinical models, offering new tools that are already or will be soon clinically explored for the treatment of specific anemias.

REFERENCES

- An, P., Wu, Q., Wang, H., Guan, Y., Mu, M., Liao, Y., et al. (2012). TMPRSS6, but not TF, TFR2 or BMP2 variants are associated with increased risk of iron-deficiency anemia. *Hum. Mol. Genet.* 21, 2124–2131. doi: 10.1093/hmg/dds028
- Andriopoulos, B. Jr., Corradini, E., Xia, Y., Faasse, S. A., Chen, S., Grgurevic, L., et al. (2009). BMP6 is a key endogenous regulator of hepcidin expression and iron metabolism. *Nat. Genet.* 41, 482–487. doi: 10.1038/ng.335
- Arezes, J., Foy, N., McHugh, K., Sawant, A., Quinkert, D., Terraube, V., et al. (2018). Erythroferone inhibits the induction of hepcidin by BMP6. *Blood* 132, 1473–1477. doi: 10.1182/blood-2018-06-857995
- Aschemeyer, S., Qiao, B., Stefanova, D., Valore, E. V., Sek, A. C., Ruwe, T. A., et al. (2018). Structure-function analysis of ferroportin defines the binding site and an alternative mechanism of action of hepcidin. *Blood* 131, 899–910. doi: 10.1182/blood-2017-05-786590
- Babitt, J. L., Huang, F. W., Wrighting, D. M., Xia, Y., Sidis, Y., Samad, T. A., et al. (2006). Bone morphogenetic protein signaling by hemojuvelin regulates hepcidin expression. *Nat. Genet.* 38, 531–539. doi: 10.1038/ng1777
- Beguín, Y. (2003). Soluble transferrin receptor for the evaluation of erythropoiesis and iron status. *Clin. Chim. Acta* 329, 9–22. doi: 10.1016/S0009-8981(03)00005-6
- Bondu, S., Alary, A. S., Lefevre, C., Houy, A., Jung, G., Lefebvre, T., et al. (2019). A variant erythroferone disrupts iron homeostasis in SF3B1-mutated myelodysplastic syndrome. *Sci. Transl. Med.* 11:pii:eaav5467. doi: 10.1126/scitranslmed.aav5467
- Camaschella, C. (2013). How I manage patients with atypical microcytic anaemia. *Br. J. Haematol.* 160, 12–24. doi: 10.1111/bjh.12081
- Camaschella, C. (2019). Iron deficiency. *Blood* 133, 30–39. doi: 10.1182/blood-2018-05-815944
- Camaschella, C., and Nai, A. (2016). Ineffective erythropoiesis and regulation of iron status in iron loading anaemias. *Br. J. Haematol.* 172, 512–523. doi: 10.1111/bjh.13820
- Camaschella, C., and Pagani, A. (2018). Advances in understanding iron metabolism and its crosstalk with erythropoiesis. *Br. J. Haematol.* 182, 481–494. doi: 10.1111/bjh.15403
- Canali, S., Wang, C. Y., Zumbrennen-Bullough, K. B., Bayer, A., and Babitt, J. L. (2017). Bone morphogenetic protein 2 controls iron homeostasis in mice independent of Bmp6. *Am. J. Hematol.* 92, 1204–1213. doi: 10.1002/ajh.24888
- Casu, C., Nemeth, E., and Rivella, S. (2018). Hepcidin agonists as therapeutic tools. *Blood* 131, 1790–1794. doi: 10.1182/blood-2017-11-737411
- Colucci, S., Pagani, A., Pettinato, M., Artuso, I., Nai, A., Camaschella, C., et al. (2017). The immunophilin FKBP12 inhibits hepcidin expression by binding the BMP type I receptor ALK2 in hepatocytes. *Blood* 130, 2111–2120. doi: 10.1182/blood-2017-04-780692
- Crielaard, B. J., Lammers, T., and Rivella, S. (2017). Targeting iron metabolism in drug discovery and delivery. *Nat. Rev. Drug Discov.* 16, 400–423. doi: 10.1038/nrd.2016.248
- De Falco, L., Sanchez, M., Silvestri, L., Kannengiesser, C., Muckenthaler, M. U., Iolascon, A., et al. (2013). Iron refractory iron deficiency anemia. *Haematologica* 98, 845–853. doi: 10.3324/haematol.2012.075515
- De Falco, L., Silvestri, L., Kannengiesser, C., Moran, E., Oudin, C., Rausa, M., et al. (2014). Functional and clinical impact of novel TMPRSS6 variants in iron-refractory iron-deficiency anemia patients and genotype-phenotype studies. *Hum. Mutat.* 35, 1321–1329. doi: 10.1002/humu.22632
- Donker, A. E., Schaap, C. C., Novotny, V. M., Smeets, R., Peters, T. M., van den Heuvel, B. L., et al. (2016). Iron refractory iron deficiency anemia: a heterogeneous disease that is not always iron refractory. *Am. J. Hematol.* 91, E482–E490. doi: 10.1002/ajh.24561
- Du, X., She, E., Gelbart, T., Truksa, J., Lee, P., Xia, Y., et al. (2008). The serine protease TMPRSS6 is required to sense iron deficiency. *Science* 320, 1088–1092. doi: 10.1126/science.1157121
- Finberg, K. E., Heeney, M. M., Campagna, D. R., Aydinok, Y., Pearson, H. A., Hartman, K. R., et al. (2008). Mutations in TMPRSS6 cause iron-refractory iron deficiency anemia (IRIDA). *Nat. Genet.* 40, 569–571. doi: 10.1038/ng.130
- Ganz, T. (2019). Anemia of inflammation. *N. Engl. J. Med.* 381, 1148–1157. doi: 10.1056/NEJMra1804281
- Gupta, R., Musallam, K. M., Taher, A. T., and Rivella, S. (2018). Ineffective erythropoiesis: anemia and iron overload. *Hematol. Oncol. Clin. North Am.* 32, 213–221. doi: 10.1016/j.hoc.2017.11.009
- Heeney, M. M., and Finberg, K. E. (2014). Iron-refractory iron deficiency anemia (IRIDA). *Hematol. Oncol. Clin. North Am.* 28, 637–652. doi: 10.1016/j.hoc.2014.04.009
- Heeney, M. M., Guo, D., De Falco, L., Campagna, D. R., Olbina, G., Kao, P. P., et al. (2018). Normalizing hepcidin predicts TMPRSS6 mutation status in patients with chronic iron deficiency. *Blood* 132, 448–452. doi: 10.1182/blood-2017-03-773028
- Kautz, L., Jung, G., Valore, E. V., Rivella, S., Nemeth, E., and Ganz, T. (2014). Identification of erythroferone as an erythroid regulator of iron metabolism. *Nat. Genet.* 46, 678–684. doi: 10.1038/ng.2996
- Koch, P. S., Olsavszky, V., Ulbrich, F., Sticht, C., Demory, A., Leibing, T., et al. (2017). Angiocrine Bmp2 signaling in murine liver controls normal iron homeostasis. *Blood* 129, 415–419. doi: 10.1182/blood-2016-07-729822
- Kovac, S., Boser, P., Cui, Y., Ferring-Appel, D., Casarubea, D., Huang, L., et al. (2016). Anti-hemojuvelin antibody corrects anemia caused by inappropriately high hepcidin levels. *Haematologica* 101, e173–e176. doi: 10.3324/haematol.2015.140772
- Lakhal, S., Schodde, J., Townsend, A. R., Pugh, C. W., Ratcliffe, P. J., and Mole, D. R. (2011). Regulation of type II transmembrane serine proteinase TMPRSS6 by hypoxia-inducible factors: new link between hypoxia signaling and iron homeostasis. *J. Biol. Chem.* 286, 4090–4097. doi: 10.1074/jbc.M110.173096
- Meynard, D., Kautz, L., Darnaud, V., Canonne-Hergaux, F., Coppin, H., and Roth, M. P. (2009). Lack of the bone morphogenetic protein BMP6 induces massive iron overload. *Nat. Genet.* 41, 478–481. doi: 10.1038/ng.320
- Muckenthaler, M. U., Rivella, S., Hentze, M. W., and Galy, B. (2017). A red carpet for iron metabolism. *Cell* 168, 344–361. doi: 10.1016/j.cell.2016.12.034
- Nemeth, E., Tuttle, M. S., Powelson, J., Vaughn, M. B., Donovan, A., Ward, D. M., et al. (2004). Hepcidin regulates cellular iron efflux by binding to ferroportin and inducing its internalization. *Science* 306, 2090–2093. doi: 10.1126/science.1104742
- Pagani, A., Colucci, S., Boccardi, R., Bertamino, M., Dufour, C., Ravazzolo, R., et al. (2017). A new form of IRIDA due to combined heterozygous mutations of TMPRSS6 and ACVR1A encoding the BMP receptor ALK2. *Blood* 129, 3392–3395. doi: 10.1182/blood-2017-03-773481

AUTHOR CONTRIBUTIONS

AP drafted the paper. CC developed the final version. AN and LS contributed to writing and to critical review the manuscript. All the authors approved the final version.

FUNDING

This paper was supported in part by an EHA Advanced Research Grant in 2018 to AP. Cariplo Foundation Young Investigator Grant n° 2017-0916 to AN.

- Pagani, A., Vieillevoye, M., Nai, A., Rausa, M., Ladli, M., Lacombe, C., et al. (2015). Regulation of cell surface transferrin receptor-2 by iron-dependent cleavage and release of a soluble form. *Haematologica* 100, 458–465. doi: 10.3324/haematol.2014.118521
- Pasricha, S. R., Lim, P. J., Duarte, T. L., Casu, C., Oosterhuis, D., Mleczo-Sanecka, K., et al. (2017). Hepcidin is regulated by promoter-associated histone acetylation and HDAC3. *Nat. Commun.* 8:403. doi: 10.1038/s41467-017-00500-z
- Piga, A., Perrotta, S., Gamberini, M. R., Voskaridou, E., Melpignano, A., Filosa, A., et al. (2019). Luspatercept improves hemoglobin levels and blood transfusion requirements in a study of patients with beta-thalassemia. *Blood* 133, 1279–1289. doi: 10.1182/blood-2018-10-879247
- Rausa, M., Pagani, A., Nai, A., Campanella, A., Gilberti, M. E., Apostoli, P., et al. (2015). Bmp6 expression in murine liver non parenchymal cells: a mechanism to control their high iron exporter activity and protect hepatocytes from iron overload? *PLoS One* 10:e0122696. doi: 10.1371/journal.pone.0122696
- Sebastiani, G., Wilkinson, N., and Pantopoulos, K. (2016). Pharmacological targeting of the hepcidin/ferroportin axis. *Front. Pharmacol.* 7:160. doi: 10.3389/fphar.2016.00160
- Sheetz, M., Barrington, P., Callies, S., Berg, P. H., McColm, J., Marbury, T., et al. (2019). Targeting the hepcidin-ferroportin pathway in anaemia of chronic kidney disease. *Br. J. Clin. Pharmacol.* 85, 935–948. doi: 10.1111/bcp.13877
- Shore, E. M., Xu, M., Feldman, G. J., Fenstermacher, D. A., Cho, T. J., Choi, I. H., et al. (2006). A recurrent mutation in the BMP type I receptor ACVR1 causes inherited and sporadic fibrodysplasia ossificans progressiva. *Nat. Genet.* 38, 525–527. doi: 10.1038/ng1783
- Silvestri, L., Pagani, A., Nai, A., De Domenico, I., Kaplan, J., and Camaschella, C. (2008). The serine protease matriptase-2 (TMPRSS6) inhibits hepcidin activation by cleaving membrane hemojuvelin. *Cell Metab.* 8, 502–511. doi: 10.1016/j.cmet.2008.09.012
- Silvestri, L., Rausa, M., Pagani, A., Nai, A., and Camaschella, C. (2013). How to assess causality of TMPRSS6 mutations? *Hum. Mutat.* 34, 1043–1045. doi: 10.1002/humu.22321
- Sonnweber, T., Nachbaur, D., Schroll, A., Nairz, M., Seifert, M., Demetz, E., et al. (2014). Hypoxia induced downregulation of hepcidin is mediated by platelet derived growth factor BB. *Gut* 63, 1951–1959. doi: 10.1136/gutjnl-2013-305317
- Sorensen, E., Rigas, A. S., Didriksen, M., Burgdorf, K. S., Thorner, L. W., Pedersen, O. B., et al. (2019). Genetic factors influencing hemoglobin levels in 15,567 blood donors: results from the Danish blood donor study. *Transfusion* 59, 226–231. doi: 10.1111/trf.15075
- Theurl, I., Schroll, A., Sonnweber, T., Nairz, M., Theurl, M., Willenbacher, W., et al. (2011). Pharmacologic inhibition of hepcidin expression reverses anemia of chronic inflammation in rats. *Blood* 118, 4977–4984. doi: 10.1182/blood-2011-03-345066
- Weinstein, D. A., Roy, C. N., Fleming, M. D., Loda, M. F., Wolfsdorf, J. I., and Andrews, N. C. (2002). Inappropriate expression of hepcidin is associated with iron refractory anemia: implications for the anemia of chronic disease. *Blood* 100, 3776–3781. doi: 10.1182/blood-2002-04-1260
- Weiss, G., Ganz, T., and Goodnough, L. T. (2019). Anemia of inflammation. *Blood* 133, 40–50. doi: 10.1182/blood-2018-06-856500
- Weiss, G., and Goodnough, L. T. (2005). Anemia of chronic disease. *N. Engl. J. Med.* 352, 1011–1023. doi: 10.1056/NEJMra041809

Conflict of Interest: CC is a consultant of Vifor Pharma, Celgene, and Novartis.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Pagani, Nai, Silvestri and Camaschella. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.