

1 **Effect of high pressure processing before freezing and frozen storage on functional**
2 **properties of European hake (*Merluccius merluccius*)**

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16 **ABSTRACT**

17 High Pressure Processing (HPP) is a non-thermal technique of growing interest
18 for fish preservation. In this work, the effect on functional properties of European hake
19 (*Merluccius merluccius*) caused by HPP pre-treatment followed by freezing and frozen
20 storage at -10°C for 5 months were evaluated in an accelerated frozen storage study.
21 Following a central composite design, five pressure levels (150, 169.27, 300, 430.73, or
22 450 MPa) and frozen storage times (0, 0.32, 2.5, 4.68, or 5 months) were tested. Colour
23 parameters (L*, a* and b*) were measured in raw muscle. Expressible water and
24 mechanical texture parameters were evaluated on both raw and cooked muscles. Results
25 showed that a low-pressure level (150 MPa) allowed an adequate water holding capacity
26 for raw muscle up to 2.5 months of frozen storage time. Values of 150 or 169.27 MPa
27 did not cause significant changes on L* values of raw muscle when compared to non-
28 treated samples. Overall, HPP led to changes on texture parameters of fresh muscle
29 before and after cooking. However, 300 MPa and 5 months of frozen storage gave
30 adhesiveness for raw muscle like that of non-treated fresh muscle. Moreover, 150-300
31 MPa for 5 months of frozen storage allowed cohesiveness and chewiness values for
32 cooked muscle like those observed on cooked fresh hake showing that the HPP
33 improves the quality of frozen hake.

35 **Keywords:** *Merluccius merluccius*, small European hake (eliminar !?), high pressure,
36 freezing, frozen storage, color, functional properties, quality

37

38 **1. Introduction**

39 Recently, there is a growing interest for consumption of fresh or minimally
40 processed food. Fish and fish products are highly perishable foods. Their freshness is
41 lost rapidly during post-mortem autolytic degradation produced by microorganisms and
42 endogenous enzymes, causing an important loss of quality (Nielsen & Nielsen, 2006).
43 Furthermore, the degradation of proteins creates the favorable conditions for the growth
44 of microorganisms responsible for the formation of compounds which release
45 unpleasant odors (Chéret, Chapleau, Delbarre-Ladrat, Verrez-Bagnis, & Lamballerie,
46 2005). Seafood is more susceptible to post-mortem texture deterioration than meat from
47 land animals (Ashie & Simpson, 1996).

48 Freezing followed by frozen storage is one of the most used methods to preserve
49 the sensory and nutritional properties of fish (Erickson, 1997). During frozen storage,
50 the degree of fish deterioration is influenced by many factors including fish species,
51 storage temperature, time, and endogenous enzymatic activity (Teixeira et al., 2014). In
52 fatty fish species, the presence of highly unsaturated fatty acid and pro-oxidant
53 molecules can lead to important enzymatic and non-enzymatic rancidity (Ramalhosa et
54 al., 2012). Thermal processing can lead to inactivation of microbial growth and lipolytic
55 enzymes. However, it may damage vitamins, flavor compounds, and polyunsaturated
56 fatty acids (Yagiz et al., 2009).

57 High pressure processing (HPP) is a non-thermal technique of growing interest
58 for the processing and preservation of food. This processing system has advantages over
59 the traditional thermal processing because it retains better the organoleptic and
60 nutritional properties of food while inactivates microbial load leading to shelf-life
61 extension and safety enhancement (Álvarez-Virrueta et al., 2012; Cheftel & Culioli,
62 1997; Mújica-Paz, Valdez-Fragoso, Samson, Welti-Chanes, & Torres, 2011; Patterson,
63 Linton, & Doona, 2007; Ríos-Romero et al., 2012; Téllez-Luis, Ramírez, Pérez-Lamela,
64 Vázquez, & Simal-Gándara, 2001). Consumers usually associate the color of seafood
65 with freshness having better flavor and higher quality (Gormley, 1992). HPP has less
66 effect on the fish color than thermal processing (Hayashi, Kawamura, Nakasa, &
67 Okinaka, 1989; Hendrickx, Ludikhuyze, Van Den Broeck, & Weemaes, 1998).

68 HPP technology allows to inactivate a wide range of food enzymes reducing
69 their impact on food quality and nutritional value. HPP demonstrated its potential
70 application to control enzymes related to fresh seafood texture deterioration (Ashie &
71 Simpson, 1996). Oxidative endogenous enzymes can also be inactivated before further
72 storage and processing of fish products (Murchie et al., 2005). This is particularly
73 important for fatty fish species. Inhibition of endogenous enzymes was observed in
74 Atlantic mackerel (*Scomber scombrus*) and horse mackerel (*Trachurus trachurus*) when
75 HPP pre-treatment followed of freezing and frozen storage was applied (Fidalgo,
76 Saraiva, Aubourg, Vázquez, & Torres, 2014a, 2014b). An inhibition of lipid hydrolysis
77 was also observed in these fish species (Torres, Vázquez, Saraiva, Gallardo, &
78 Aubourg, 2013; Vázquez, Torres, Gallardo, Saraiva, & Aubourg, 2013) and recently
79 also in sardine (*Sardina pilchardus*) (Méndez et al., 2017).

80 HPP induces modifications on food functional properties. Therefore, it can be
81 used by the industry to create new product textures (Chapleau & De Lamballerie-Anton,
82 2003). Generally, foods are subjected to pressure levels in the range of 100 to 650 MPa.
83 Variables such as pressure level, pressure holding time, and pressurization rate, show an
84 influence on the functional and sensory properties and microbiological load of fish.
85 Furthermore, the effect of HPP on the fish quality varies with fish species.

86 HPP treatments also have negative effects on food quality. They affect
87 intermolecular bonds that may induce modifications in water, proteins, polysaccharides,
88 and lipids. The main effect is to produce changes in hydrophobic and electrostatic
89 interactions affecting secondary, tertiary, and quaternary structures in proteins (Chéret
90 et al., 2005).

91 The pressure levels and holding times needed to reduce the microbial load
92 denature myofibrillar proteins and produce changes in visual appearance, protein
93 functionality, and mechanical properties of fish muscle (Uresti, Velazquez, Vázquez,
94 Ramírez, & Torres, 2005). A change in the myofibrillar protein profiles was observed in
95 sea bass fillets. Proteins with molecular weight below 30 kDa increased and those with
96 lower isoelectric point values decreased. This could explain the whitish color and the
97 decrease of water holding capacity of fish muscle observed (Teixeira et al., 2014). In
98 chilled coho salmon (*Oncorhynchus kisutch*), an increase of hydrostatic pressure caused
99 a lipid oxidation development augmentation (Aubourg, Tabilo-Munizaga, Reyes,
100 Rodríguez, & Pérez-Won, 2010) and a marked protein damage in the sarcoplasmic
101 fraction (Ortea, Rodríguez, Tabilo-Munizaga, Pérez-Won, & Aubourg, 2010). Therefore

102 a carefully selection of the conditions used in HHP pre-treatments is necessary to
103 minimize undesirable changes in the fish muscle quality.

104 The effect of HPP treatments followed of freezing and frozen storage on the
105 functional and sensory properties of fish species such as Atlantic mackerel (*Scomber*
106 *scombrus*) (Aubourg, Torres, Saraiva, Guerra-Rodríguez, & Vázquez, 2013) and horse
107 mackerel (*Trachurus trachurus*) (Torres, Saraiva, Guerra-Rodríguez, Aubourg, &
108 Vazquez, 2014) were studied. Results of these studied showed a species-dependent
109 effect. This can be due to differences in fat composition.

110 The European hake (*Merluccius merluccius*) is a lean fish, widely consumed in
111 many countries because of its high nutritional value and recognised health benefits.
112 There is not studies about the effects of HPP pre-treatments of freezing and frozen
113 storage on this fish species. Therefore, the aim of this work was to study the changes on
114 color and functional properties of frozen small European hake (*Merluccius merluccius*)
115 caused by HPP pre-treatments before freezing and frozen storage for up to 5 months.
116 The effect of HPP on functional properties was evaluated in raw and cooked fish.

117 Quizás haya que reducir algo la mención de nuersto paper previos, y centrarnos en los
118 más directamente relacionados, y mencionados en Res y Disc.

119 Y creo que faltaría, en cambio, más énfasis en la problemática propia de merluza. Ahí se
120 puede tomar algo del paper enviado; de la Introducción: el gran problema de la
121 congelación de merluza sería la formación de FA y DMA y su repercussion en la
122 textura. Copia, pega y cambia algo lo que veas de esa Introducción

123

124 **2. Materials and methods**

125 *2.1. Raw fish, processing, storage and sampling*

126 European hake (*Merluccius merluccius*) was obtained at the Vigo harbour
127 (Pontevedra, más conocido es Galicia, no ?, northwest Spain), being caught close to the
128 Galician coast and immediately transported to the “Plataforma Tecnológica
129 Multidisciplinar Alta Pressão” (University of Aveiro, Portugal) in a refrigerated truck
130 for HPP treatment within 6 h after catch. Samples were packed in polyethylene bags and
131 vacuum sealed at 400 mbar. The length and weight of the specimens ranged 27.5-29.5
132 cm and 180-205 g, respectively.

133 HPP treatments were performed in a 55-L high pressure unit (WAVE
134 6000/55HT; NC Hyperbaric, Burgos, Spain). The pressure levels studied were 150,
135 169.27, 300, 430.73 and 450 MPa. In all cases, the holding time of the pressure was 2

136 min. Water applied as the pressurising medium at 3 MPa s^{-1} yielded 50, 56.42, 100,
137 143.58 and 150 s as the come-up times for the 150, 169.27, 300, 430.73 and 450 MPa
138 treatments, respectively. While decompression time was less than 3 s. Inlet water was
139 adjusted to holding temperature conditions during HPP treatment at room temperature
140 (20 °C). After HPP processing, individual small hakes were kept frozen at -20 °C for 48
141 h before storage at -10 °C and sampling after 0, 0.32, 2.5, 4.68 and 5 months of frozen
142 storage. The relatively high temperature (-10°C), as compared to commercial practices
143 (-18°C), was chosen as an accelerated storage test to determine in less time the effect of
144 the HPP pre-treatment.

145 For analysis, fish samples were thawed at 4 °C for 24 h, eviscerated, bones
146 removed manually and then filleted. Frozen samples with no HPP treatment (frozen
147 controls) were subjected to the same freezing and frozen storage conditions. Fresh fish
148 with no HPP treatment (fresh controls) was also analyzed. The analytical procedures
149 described below were carried out on the muscle, raw or cooked. Cooked fish was
150 prepared in an oven at 200 °C for 10 min reaching at least 68°C at the centre point.

151

152 *2.2. Expressible water content and colour*

153 The expressible water content was determined for raw and cooked samples
154 following the procedure described by Uresti, Lopez-Arias, Ramirez, & Vazquez (2003).
155 Four analyses were realized for each treatment and replicate. Colour was only
156 determined in raw samples following the procedure described by the same authors. A
157 colorimeter ColorStriker meter (Mathai, Hannover, Germany) was used. Values of L*,
158 a*, and b* were calculated based on illuminant C and the 2° standard observer. Nine
159 measures were realized for each treatment and replicate.

160

161 *2.3. Texture profile analysis (TPA)*

162 Texture profile for each treatment was studied in raw and cooked fish samples.
163 Measures were realized at room temperature. A TA-XTplus texturometer (Stable Micro
164 System, Viena Court, UK) equipped with a 50-mm diameter cylindrical aluminium
165 probe (P/50) was used. Samples were cut into small cubes (2 x 2 x 1.5 cm) which were
166 compressed to 75% of the original height at a 60 mm/min compression speed. Five
167 textural parameters (hardness, adhesiveness, springiness, cohesiveness and chewiness)
168 were determined and seven samples were analysed for each treatment and replicate

169 (Cortez-Vega, Fonseca, Feisther, Silva, & Prentice, 2013; Palmeira, Mársico, Monteiro,
170 Lemos, & Conte Junior, 2016; Silva, Lourenço, & Pena, 2017).

171

172 2.4. Statistical analysis

173 The experimental design was statistically analyzed using the Design Expert®
174 7.1.1 software (Stat-Ease, Inc., Minneapolis, MN). The set of experiments followed a
175 central composite design (CCD). A CCD has three groups of design points: (a) two-
176 level factorial or fractional factorial design points; (b) axial points (sometimes called
177 "star" points); (c) center points. This design has 5 levels of the independent variables
178 with desirable statistical properties. The following second order polynomial model was
179 used as a first approach to analyse the experimental data:

$$180 \quad y^i = b_0^i + b_1^i x_1 + b_2^i x_2 + b_3^i x_1 x_2 + b_4^i x_1^2 + b_5^i x_2^2$$

181 where x_i ($i = 1-2$) are the code variables for pressure level and storage time; y^i ($i = 1-$
182 15) are the dependent variables (raw expressible water, cooked expressible water, L^* ,
183 a^* , b^* , raw hardness, adhesiveness, springiness, cohesiveness and chewiness and
184 cooked hardness, adhesiveness, springiness, cohesiveness and chewiness); and, b_0^i ,
185 $b_0^i \dots b_5^i$ are regression coefficients estimated from the experimental data by multiple
186 linear regression. Model terms were selected or rejected based on p-values at 95%
187 confidence level determined by analysis of variance (ANOVA). Partial models of the
188 quadratic model were also obtained and analysed by ANOVA.

189

190 3. Results and discussion

191

192 3.1. Expressible water

193 This parameter is related to the fish flesh water holding capacity and affects to
194 the product juiciness perceived by consumers. Therefore, fish processing should have a
195 minimum effect on this parameter to retain an acceptable product sensory quality.

196 The values of expressible water of raw and cooked samples without HPP
197 (control) for the frozen storage time studied are shown in Table 1. The average values
198 for fresh fish were $33.92 \pm 2.24\%$ (w/w) before cooked and $36.66 \pm 1.60\%$ (w/w) for
199 cooked samples. The frozen storage of fresh fish led to changes in the expressible water
200 values of both raw and cooked muscle. After frozen storage for 5 months at -10°C ,
201 expressible water increased up to 42.72% (w/w) before cooking and 40.70 (w/w) after
202 cooking.

203 The expressible water values of raw muscle subjected at HPP pre-treatments are
204 shown in Table 2. They were higher (34.32-46.88% w/w) than those observed in fresh
205 muscle without HPP pre-treatment. In order to assess the relative influence of pressure
206 level and frozen time on the value of expressible water, a multifactor ANOVA was
207 carried out. A significant model ($p < 0.0001$) was obtained to predict the effect of
208 pressure level and frozen storage time on expressible water. Experiment 1 was excluded
209 because it was detected as an outlier point. ANOVA showed that expressible water was
210 mainly affected by the quadratic pressure level term (F-value=13.52) and the pressure
211 level-frozen storage time interaction term (F=7.79) followed of linear pressure level and
212 linear frozen storage time terms. Quadratic frozen storage exerted less influence. The
213 determination coefficient (r^2) of the model was 0.9289. The prediction of the model
214 obtained for the effect of the pressure level and frozen storage time on expressible water
215 is shown in Figure 1a.

216 The employment of HPP pre-treatment followed of freezing and frozen storage
217 can lead to a significant increase of expressible water if high pressure levels and frozen
218 storage times are selected. However, fish subjected to a hydrostatic pressure of 150 MPa
219 followed of 2.5 months of frozen storage yielded an expressible water value of 37.86 %
220 (w/w). The use of this pressure and frozen storage time produced a water holding
221 capacity enough for a desirable juiciness.

222 At 150 MPa, studies on other fish species showed comparable results.
223 Expressible water values lower than 40% (w/w) was obtained for Atlantic mackerel
224 (*Scomber scombrus*) (Aubourg et al., 2013). In sea bass fillets, the water holding
225 capacity of the samples treated at 100 MPa were not different from non-treated samples.
226 With high pressure levels (250 and 400 MPa) and holding times (5-30 min), water
227 holding capacity decreased significantly (Teixeira et al., 2014). An expressible water
228 value of 38.7% was considered optimal for low-salt restructured fish products from
229 Atlantic mackerel (Martelo-Vidal, Mesas, & Vazquez, 2012).

230 HPP pre-treatments gave expressible water values for cooked muscle between
231 30.10 and 36.32% (w/w) (Table 2). The effect of pressure level and frozen storage time
232 on expressible water of the cooked fish was evaluated by multifactor ANOVA.
233 However, the model obtained was not significant (F-value = 3.00).

234

235 3.2. *Flesh colour*

236 The mean colour parameters for fresh muscle were 59.58 for L*, 0.449 for a*
237 and 0.542 for b*. The effect of frozen storage time of raw fish on colour parameters was
238 evaluated. The results obtained are showed in Table 1. The L* values did not
239 significantly change after frozen storage. However, important changes in the values of
240 a* and b* were observed. After fish frozen storage, the values of a* were lower and the
241 values of b* higher than those observed on fresh muscle. For instance, after 5 months of
242 frozen storage, the values of a* and b* were -1.091 and 2.931, respectively.

243 The values for L*, a* and b* obtained for the HPP pre-treatments and frozen
244 storage times studied are showed in Table 2. The values of L* ranged between 65.44
245 and 79.06. The effect of HPP pre-treatment and frozen storage time on L* value of fish
246 was evaluated by ANOVA. A significant linear model was obtained (F-value 9.36). The
247 value of r² was acceptable (0.7006). The effect of pressure level on L* value of the raw
248 muscle was significant (F-value=18.67). However, the effect of frozen storage time was
249 negligible. The prediction of the model for the effect of the pressure level and frozen
250 storage time on L* value is shown in Figure 1b. The L value significantly increased
251 with pressure level, reaching a value of 79.06 at 450 MPa. Low pressure levels (150 and
252 169.27 MPa) did not cause differences when compared to non-treated samples.

253 An increase on the L* value with pressure was also observed on the muscle of
254 Atlantic mackerel (*Scomber scombrus*) (Aubourg et al., 2013), sea bass (*D. labrax*)
255 (Tironi, Lebail, & De Lamballerie, 2007) and horse mackerel (*Trachurus trachurus*)
256 (Torres et al., 2014). This HPP effect could be interesting if whiteness is a desirable
257 parameter by consumers. The effects of HPP pre-treatment and frozen storage time on
258 a* and b* parameters were also evaluated by multifactor ANOVA. Significant models
259 were not obtained.

260

261 3.3. Textural profile analysis of raw samples

262 Table 3 shows the effect of frozen storage time up to 5 months on the texture
263 profile of raw muscle without HPP pre-treatment. Hardness of fresh muscle was 5152 g-
264 force and significantly increased when it was frozen stored. After 4.98 and 5 months of
265 frozen storage, hardness increased to 9314 g-force and 7764 g-force, respectively.

266 Texture parameters of raw muscle subjected at different HPP pre-treatments are
267 shown in Table 4. Hardness ranged from 7,246 to 14,763 (en general habría que poner
268 la coma a los miles, no ?)g-force. The effect of HPP pre-treatment and frozen storage
269 time on raw fish hardness were evaluated by multifactor ANOVA. A significant model

270 was obtained. Hardness was highly affected by linear (F-value = 93.09) and quadratic
271 pressure level (F-value = 20.35) terms. Terms corresponding to linear and quadratic
272 frozen storage times and pressure level-frozen storage time interaction, exerted a minor
273 effect on the hardness of muscle. Prediction of the model ($r^2 = 0.9603$) is shown in
274 Figure 2a. High pressure levels caused a strong increase on hardness. The hardness of
275 raw samples (5152 g-force) increased to 14623 g-force at 450 MPa.

276 An increase in hardness with the pressure level was also observed in other
277 species like Atlantic mackerel (*Scomber scombrus*). The effect of frozen storage time
278 subsequent to HPP treatment was negligible in that case (Aubourg et al., 2013). In cod
279 (*Gadus morhua*), hardness increased with the pressure level while only minor changes
280 were observed during frozen storage (Matser, Stegeman, Kals, & Bartels, 2000).
281 Hardness of horse mackerel (*Trachurus trachurus*) was highly affected by pressure
282 level and frozen storage time (Torres et al., 2014).

283 The adhesiveness of fresh muscle was -175.7 g·s and decreased with the frozen
284 storage time, reaching a value of -56.9 g·s after 5 months of frozen storage at -10°C.
285 When HPP pre-treatments were applied, the adhesiveness values ranged from -50.3
286 to -392.2 g·s. The multifactor ANOVA analysis of the effect of HPP pre-treatment and
287 frozen storage time on adhesiveness of raw muscle resulted in a significant model.
288 Adhesiveness was mainly affected by the linear (F-value = 8.22) and quadratic pressure
289 levels (F-value=6.72) and quadratic frozen storage time (F-value =7.15) terms. A minor
290 influence was observed for the linear frozen storage time and pressure level-frozen
291 storage time interaction terms. The prediction of the model ($r^2 = 0.8501$) obtained is
292 shown in Figure 2b. The model obtained can be used to select a similar adhesiveness to
293 that of fresh fish.

294 Comparing with other species, the adhesiveness of horse mackerel (*Trachurus*
295 *trachurus*) was highly affected by the linear and quadratic pressure level term.
296 However, the effect of frozen storage time on adhesiveness was negligible (Torres et al.,
297 2014). HPP pre-treatments caused a significant negative effect on the adhesiveness of
298 Atlantic mackerel (*Scomber scombrus*) when high pressure levels and long storage
299 times were used (Aubourg et al., 2013).

300 Springiness of fresh muscle (0.232) increased with the frozen storage time in
301 non-treated samples, reaching a value of 0.377 after 5 months. Springiness values for
302 HPP pre-treated raw muscle ranged from 0.294 to 0.494. Similar ranges were found for
303 HPP pre-treated and frozen Atlantic mackerel (*Scomber scombrus*) (0.189-0.346)

304 (Aubourg et al., 2013), HPP pre-treated and frozen horse mackerel (*Trachurus*
305 *trachurus*) (0.224-0.492) (Torres et al., 2014) and restructured fish products (0.20-0.60)
306 from gilthead sea bream (*Sparus aurata*) (Andrés-Bello, García-Segovia, Ramírez, &
307 Martínez-Monzó, 2011).

308 For springiness, the ANOVA led to an F-value of 6.67 excluding experiment 10
309 as outlier, which implied that the linear model was significant. The value of r^2 was
310 0.6558. The evaluation of the F-values showed that springiness was affected by pressure
311 level (F-value = 5.62) and frozen storage time (F-value = 5.07; p-value = 0.0590).
312 Prediction of the model obtained for the effect of the pressure level and frozen storage
313 on springiness is shown in Figure 2c. The two variables affected **positively the**
314 springiness values. Pressure levels between 150 and 300 MPa gave springiness values in
315 the range 0.301-0.388 like the range observed in frozen muscle controls (0.326-0.377).
316 Contrary, previous studies on horse mackerel (*Trachurus trachurus*) showed that
317 springiness was not affected by the variation of the pressure treatment and frozen
318 storage time (Torres et al., 2014). This fact can show a species-dependent effect.

319 Cohesiveness of fresh muscle (0.247) increased with the frozen storage time at -
320 10°C, reaching a value of 0.414 for 5 months. HPP pre-treatment followed of freezing
321 and frozen storage gave cohesiveness values ranging between 0.294 and 0.476. A
322 similar range of cohesiveness was observed for other fish products such as restructured
323 fish products from gilthead sea bream (*Sparus aurata*) where values between 0.30 and
324 0.40 were obtained (Andrés-Bello et al., 2011).

325 The prediction linear model was significant ($r^2=0.8581$). Cohesiveness was
326 affected by both frozen storage time (F-value = 30.01) and pressure level (F-value =
327 20.49). Prediction of the model is showed in Figure 3a. HPP pre-treatments caused an
328 important increase on cohesiveness when a high pressure and long storage time were
329 selected. At 430.73 MPa and 4.68 months of frozen storage, the cohesiveness value was
330 0.476. This effect was also observed in another fish species. HPP pre-treatments caused
331 a significant increase on cohesiveness of Atlantic mackerel (*Scomber scombrus*) when
332 high pressure and long storage time were selected (Aubourg et al., 2013).

333 Chewiness of fresh muscle (320 g) increased to 1213 g for 5 months of storage
334 at -10°C. HPP pre-treated samples showed a wide range of chewiness values (713-3583
335 g). The multifactor ANOVA analysis led to a significant linear model (F-value=10.00).
336 Chewiness was more affected by the pressure level (F-value = 17.14) than by the frozen
337 storage time. The value of r^2 for the model was 0.7144. Prediction of the model

338 obtained for the effect of the pressure level and frozen storage on chewiness is shown in
339 Figure 3b. The changes on chewiness with the time of frozen storage were more
340 pronounced when high pressures are selected. For example, at 430.73 MPa, 0.38 and
341 4.68 months of frozen storage gave chewiness values of 2255 and 3583, respectively.

342

343 **3.4. Textural profile analysis of cooked samples**

344 Textural parameters for cooked muscle samples without HPP pre-treatment are
345 shown in Table 3. When the samples of fresh controls were cooked, hardness (8060 g-
346 force), springiness (0.42), cohesiveness (0.45), and chewiness (1541) were higher than
347 those of raw samples, mainly chewiness. However, adhesiveness was lower after
348 cooking (-93 g·s). This behaviour was also observed for any time of frozen storage
349 considered.

350 All textural parameters for cooked muscle were affected by the frozen storage
351 time. However, the effect on springiness and cohesiveness was very small. After
352 freezing and frozen storage, hardness (11661-19601 g-force), springiness (0.42-0.593),
353 cohesiveness (0.45-0.539), and chewiness (3181-5440 g-force) increased and
354 adhesiveness (-14.4-(-32.1)) decreased.

355 The results on the cooked texture profile of HPP are shown in Table 5. Using
356 multifactor ANOVA, no significant models were obtained for hardness, adhesiveness
357 and springiness. The values range for these parameters were: hardness: 8669-18405 g-
358 force, adhesiveness: -29.9-(-259.6), and springiness: 0.336-0.591.

359 Significant models for cohesiveness (F-value=10.75; $r^2=0.9307$) and chewiness
360 (F-value=7.70; $r^2=0.9059$) were obtained. Cohesiveness (0.428-0.548) was mainly
361 affected by the pressure level-frozen storage time interaction (F-value=16) and
362 quadratic pressure level terms (F=12.19). The linear and quadratic frozen storage time
363 terms exercised a minor influence and the effect of linear pressure level term was
364 negligible. The prediction of model for the effect of pressure level and frozen storage
365 time on cohesiveness is shown in Figure 4a. The model obtained can be used to select a
366 value of springiness like that of cooked fresh muscle without HPP pre-treatment.
367 Therefore, a pressure of 300 MPa and a time of frozen storage of 5 months gave
368 cohesiveness like that of cooked fresh muscle (not HPP treated).

369 Chewiness was mainly affected by quadratic pressure level (F=15.76) term
370 followed of linear (F-value=8.01) and quadratic (F-value=7.68) frozen storage time
371 terms. The linear pressure level and pressure level-frozen storage time interaction terms

372 exercised less influence on chewiness. The prediction of model for the effect of pressure
373 level and frozen storage time on chewiness of cooked muscle is shown in Figure 4b.
374 The model obtained can be used to select a value of chewiness like that observed in
375 cooked fresh muscle (non-treated). A pressure of 300 MPa followed by freezing and
376 frozen storage during 5 months yielded a chewiness value like that of cooked fresh
377 muscle without HPP pre-treatment.

378

379 **4. Conclusions**

380 Frozen storage of European hake (*Merluccius merluccius*) at -10°C led to
381 important changes on colour parameters of raw muscle and texture profile of both raw
382 and cooked muscles of European hake. However, lightness of raw fish was not affected.
383 The frozen storage for 5 months of European hake caused important changes on the
384 water holding capacity of both raw and cooked muscles. A pretreatment at 300 MPa
385 before frozen storage at -10°C for 5 months gave cohesiveness like that of cooked fresh
386 muscle showing that the HPP improves the quality of frozen hake.

387

388 **ACKNOWLEDGMENTS**

389 The work was supported by the Consejo Superior de Investigaciones Científicas
390 (CSIC) (Spain) through the Research Project 2017-70E032, by ...

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528 **Table 1**

529 Effect of frozen storage time at -10°C on expressible water and colour of European hake
 530 (*Merluccius merluccius*) without HPP pre-treatment.

Frozen time (months)	Expressible water % (w/w)		L*	a*	b*
	raw	cooked	raw	raw	raw
0	33.92	36.66	59.58	0.449	0.542
0.32	37.14	33.53	61.73	-0.063	4.567
2.5	35.00	34.26	63.08	-0.876	3.170
4.68	38.63	34.47	60.16	-0.370	3.136
5	42.72	40.70	63.52	-1.091	2.931

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549 **Table 2**

550 Effect of high pressure processing (HPP) as a pre-treatment and frozen storage time on
 551 expressible water and colour of European hake (*Merluccius merluccius*). Experimental
 552 treatment use codes P and F for Pressure (MPa) and frozen storage time (months),
 553 respectively.

Treatment	Expressible	Expressible	L*	a*	b*
	water % (w/w)	water % (w/w)	raw	raw	raw
1 (P150F2.5)	37.86	32.25	66.15	-1.821	4.106
2 (P169.27F0.32)	34.32	33.87	66.14	-0.659	0.003
3 (P169.27F4.68)	44.46	34.01	65.44	-1.261	2.963
4 (P300F0)	44.14	36.32	72.49	-1.994	0.412
5 (P300F2.5)	43.51	33.31	77.61	-2.258	-0.039
6 (P300F2.5)	41.38	30.84	68.78	-0.676	3.018
7 (P300F2.5)	44.46	30.82	72.29	-1.199	4.051
8 (P300F5)	46.88	34.74	73.21	-1.440	4.540
9 (P430.73F0.32)	37.66	30.10	75.07	-2.833	0.716
10 (P430.73F4.68)	43.20	33.15	73.31	-2.097	2.969
11 (P450F2.5)	40.33	30.24	79.06	-1.971	4.023

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555 **En lugar de F, parece más intuitivo emplear T, de time)**

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562 **Table 3**

563 Effect of frozen storage time on the raw and cooked texture profile of European hake

564 (*Merluccius merluccius*) without HPP pre-treatment.

Frozen time (months)	Hardness (g)	Adhesiveness (g·s)	Springiness	Cohesiveness	Chewiness (g)
	Raw	Raw	Raw	Raw	Raw
0	5152	-175,7	0,232	0,247	320
0.32	6511	-126,1	0,266	0,255	489
2.5	7106	-97,2	0,326	0,343	783
4.68	9314	-97,6	0,358	0,384	1260
5	7764	-56,9	0,377	0,414	1213
	Cooked	Cooked	Cooked	Cooked	Cooked
0	8060	-93,0	0,42	0,45	1541
0.32	19601	-32,1	0,593	0,464	5440
2.5	15875	-25,9	0,524	0,486	4045
4.68	15096	-14,4	0,577	0,537	4678
5	11661	-20,3	0,487	0,539	3181

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567 **Corregir puntos y comas en anglosajón**

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582 **Table 4**

583 Effect on the raw muscle texture of high pressure processing as a pre-treatment for
 584 frozen European hake (*Merluccius merluccius*). Experimental treatment use codes P and
 585 F for Pressure (MPa) and frozen storage time (months), respectively.

Experiments	Hardness (g)	Adhesiveness (g·s)	Springiness	Cohesiveness	Chewiness (g)
1 (P150F2.5)	9669	-152,0	0,359	0,294	1005
2 (P169.27F0.32)	7246	-120,1	0,301	0,328	713
3 (P169.27F4.68)	8000	-50,3	0,346	0,358	991
4 (P300F0)	8195	-244,8	0,294	0,301	773
5 (P300F2.5)	10642	-392,2	0,388	0,368	1581
6 (P300F2.5)	9357	-388,7	0,333	0,320	1036
7 (P300F2.5)	9448	-237,7	0,346	0,346	1119
8 (P300F5)	9241	-188,0	0,356	0,406	1333
9 (P430.73F0.32)	13270	-349,7	0,489	0,347	2255
10 (P430.73F4.68)	14763	-129,0	0,494	0,476	3583
11 (P450F2.5)	14623	-287,9	0,401	0,364	2187

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588 **Corregir puntos y comas en anglosajón**

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605 **Table 5**

606 Effect on the cooked muscle texture of high pressure processing as a pre-treatment for
607 frozen European hake (*Merluccius merluccius*). Experimental treatment use codes P and
608 F for Pressure (MPa) and frozen storage time (months), respectively.

Experiments	Hardness (g)	Adhesiveness (g·s)	Springiness	Cohesiveness	Chewiness (g)
1 (P150F2.5)	17904	-29,9	0,54	0,504	4995
2 (P169.27F0.32)	15435	-51,8	0,54	0,496	4227
3 (P169.27F4.68)	12182	-65,7	0,517	0,548	3555
4 (P300F0)	12805	-259,6	0,428	0,428	2393
5 (P300F2.5)	12890	-183,0	0,468	0,471	2851
6 (P300F2.5)	15649	-156,6	0,467	0,465	3435
7 (P300F2.5)	14672	-120,7	0,517	0,468	3622
8 (P300F5)	8669	-237,3	0,336	0,45	1497
9 (P430.73F0.32)	18405	-132,2	0,591	0,436	4959
10 (P430.73F4.68)	13950	-197,8	0,546	0,537	4295
11 (P450F2.5)	15149	-215,8	0,539	0,483	4122

609 **Corregir puntos y comas en anglosajón**

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612 FIGURE LEGENDS

613

614 **Fig 1.** Model prediction for the effect of pressure level (MPa) and frozen storage time
615 (months) on a) expressible water of raw muscle and b) lightness parameter (L) of
616 European hake (*Merluccius merluccius*).

617

618 **Fig. 2.** Model prediction for the effect of pressure level (MPa) and frozen storage time
619 (month) on a) hardness, b) adhesiveness, c) springiness of raw muscle of European hake
620 (*Merluccius merluccius*).

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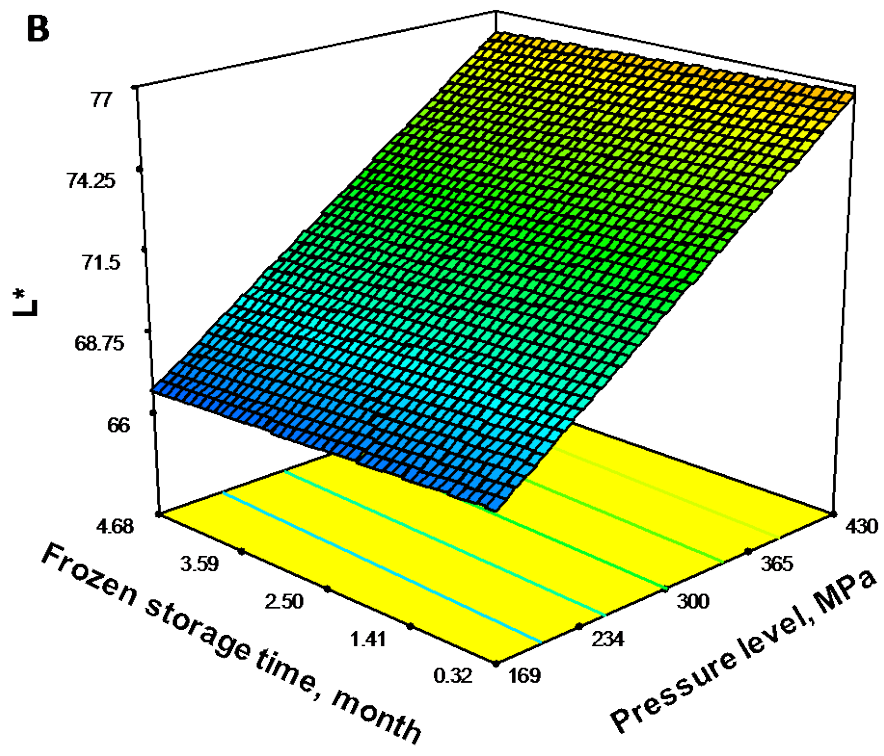
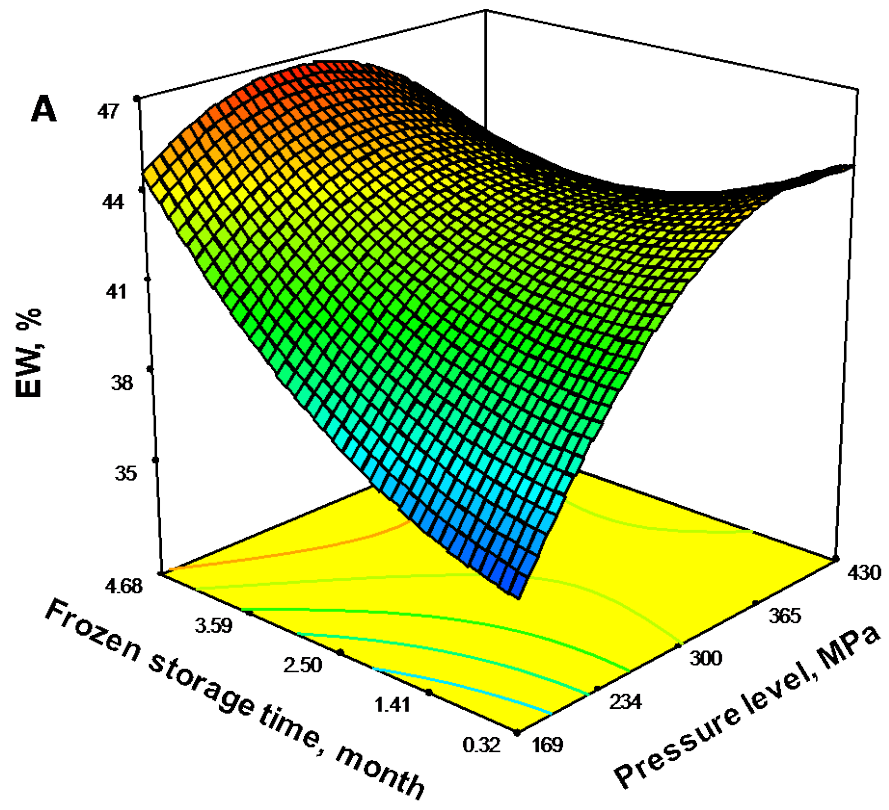
622 **Fig. 3.** Model prediction for the effect of pressure level (MPa) and frozen storage time
623 (month) on a) cohesiveness and b) chewiness of raw muscle of European hake
624 (*Merluccius merluccius*).

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626 **Fig. 4.** Model prediction for the effect of pressure level (MPa) and frozen storage time
627 (month) on a) cohesiveness and b) chewiness of cooked muscle of European hake
628 (*Merluccius merluccius*)

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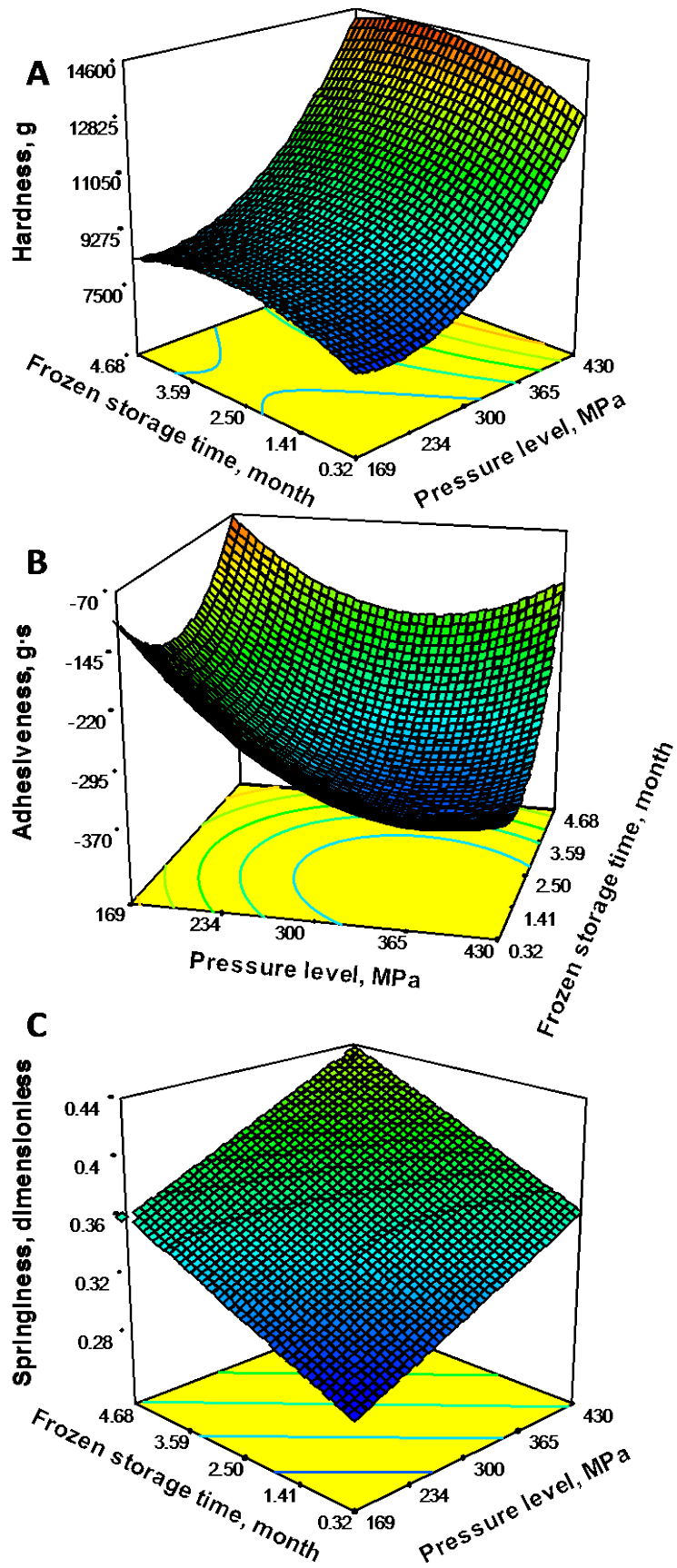


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633 Figure 1

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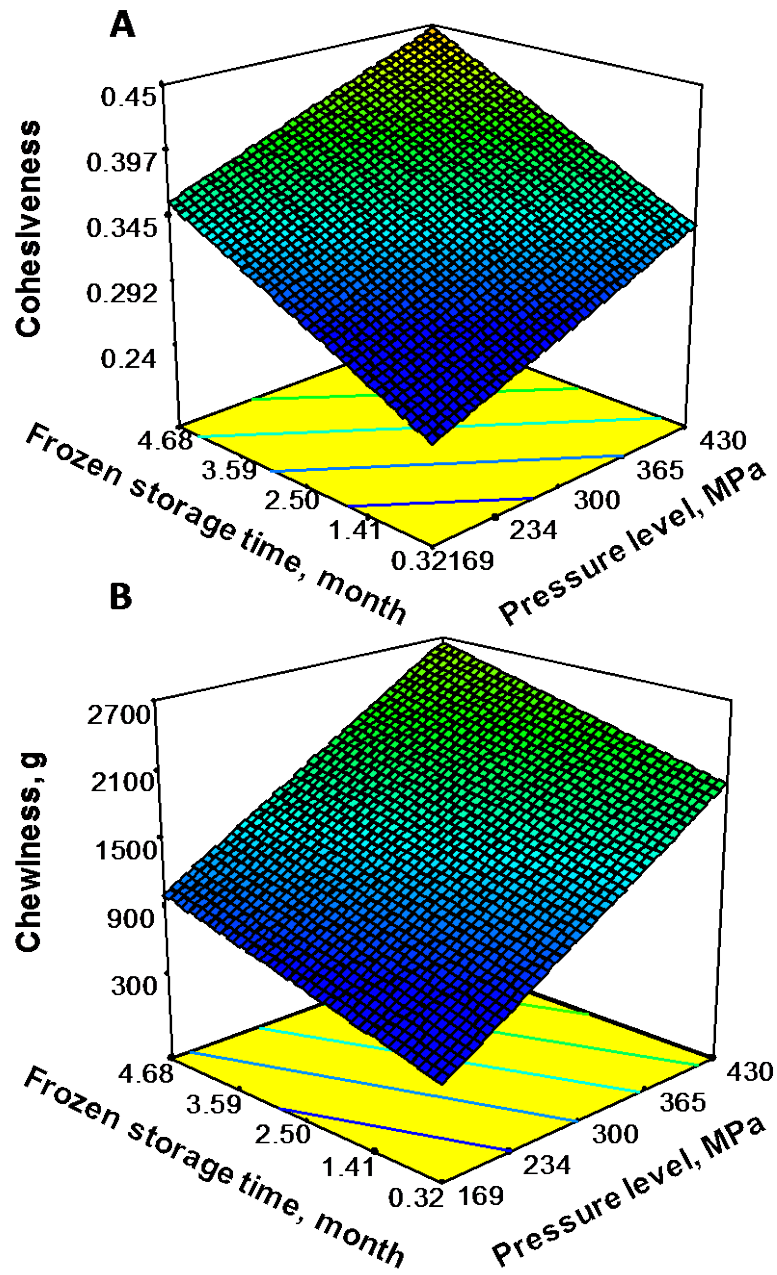


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636 Figure 2

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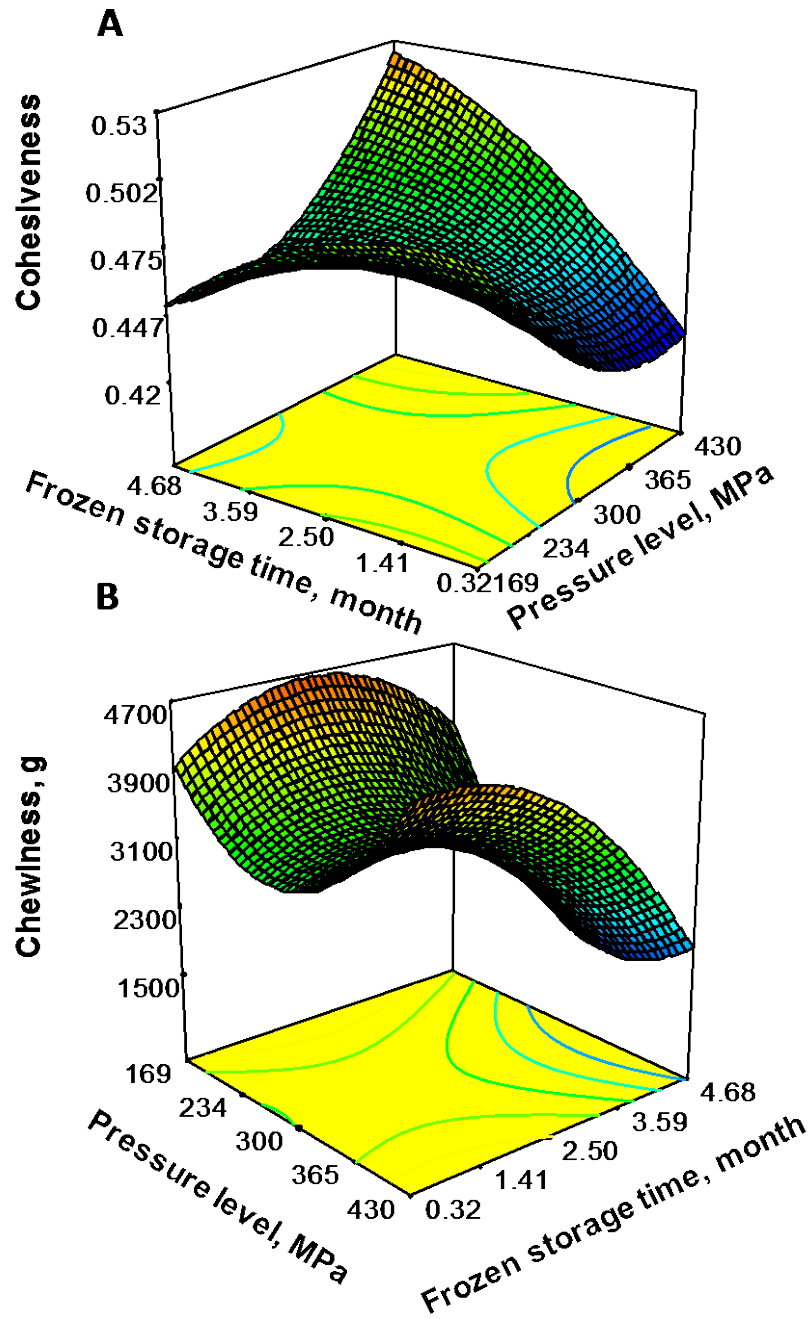


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641 Figure 3

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646 Figure 4

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648 **Creo que se pone months, en lugar de month. Eliminar "dimensionless" en Fig 2**