

# High Prevalence of EMRSA-15 in Portuguese Public Buses: A Worrisome Finding

Roméo Rocha Simões<sup>1,2</sup>, Marta Aires-de-Sousa<sup>3</sup>, Teresa Conceição<sup>1</sup>, Filipa Antunes<sup>2</sup>, Paulo Martins da Costa<sup>2,4</sup>, Hermínia de Lencastre<sup>1,5\*</sup>

**1** Laboratory of Molecular Genetics, Instituto de Tecnologia Química e Biológica, Oeiras, Portugal, **2** Instituto de Ciências Biomédicas de Abel Salazar, Universidade do Porto, Oporto, Portugal, **3** Escola Superior de Saúde da Cruz Vermelha Portuguesa, Lisbon, Portugal, **4** CIIMAR - Centro Interdisciplinar de Investigação Marinha e Ambiental do Porto, Oporto, Portugal, **5** Laboratory of Microbiology, The Rockefeller University, New York, New York, United States of America

## Abstract

**Background:** The nosocomial prevalence of methicillin resistant *Staphylococcus aureus* (MRSA) in Portugal remains one of the highest in Europe and is currently around 50%. Transmission of *S. aureus*, including MRSA, occurs principally by direct human-to-human skin contact. However, *S. aureus* can survive for long periods on inanimate objects, which may represent an important reservoir for dissemination as well.

**Methodology/Principal Findings:** Between May 2009 and February 2010, handrails of 85 public urban buses circulating in Oporto, Portugal, were screened for the occurrence of MRSA. Twenty-two (26%) buses showed MRSA contamination. The molecular characterization of a total of 55 MRSA, by pulsed-field gel electrophoresis (PFGE), staphylococcal cassette chromosome (*SCC mec*) typing, *spa* typing, and multilocus sequence typing (MLST), clustered the isolates into three clonal types. However, the overwhelming majority ( $n = 50$ ; 91%) of the isolates belonged to a single clone (PFGE A, *spa* types t747, t032, t025 or t020, ST22, *SCC mec* type IVh) that exhibits the characteristics of the pandemic EMRSA-15, currently the major lineage circulating in Portuguese hospitals, namely in the Oporto region. Two additional clones were found but in much lower numbers: (i) PFGE B, ST5, *spa* type t002, *SCC mec* IVa ( $n = 3$ ), and (ii) PFGE C, *spa* type t008, ST8, *SCC mec* IVa ( $n = 2$ ). None of the 55 isolates was PVL positive.

**Conclusions/Significance:** Public buses in Oporto seem to be an important reservoir of MRSA of nosocomial origin, providing evidence that the major hospital-associated MRSA clone in Portugal is escaping from the primary ecological niche of hospitals to the community environment. Infection control measures are urgently warranted to limit the spread of EMRSA-15 to the general population and future studies are required to assess the eventual increase of MRSA in the Portuguese community, which so far remains low.

**Citation:** Simões RR, Aires-de-Sousa M, Conceição T, Antunes F, da Costa PM, et al. (2011) High Prevalence of EMRSA-15 in Portuguese Public Buses: A Worrisome Finding. PLoS ONE 6(3): e17630. doi:10.1371/journal.pone.0017630

**Editor:** Michael Otto, National Institutes of Health, United States of America

**Received:** January 6, 2011; **Accepted:** February 3, 2011; **Published:** March 2, 2011

**Copyright:** © 2011 Simões et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Funding:** This work was partially supported by Project TROCAR (TROCAR-HEALTH-F3-2008-223031) from the European Community to HL and by grant 095/BI-BI/2010 from the same Project to T.C. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. No additional external funding was received for this study.

**Competing Interests:** The authors have declared that no competing interests exist.

\* E-mail: lencash@mail.rockefeller.edu

## Introduction

Methicillin-resistant *Staphylococcus aureus* (MRSA) is one of the most important hospital-associated (HA-MRSA) pathogens, responsible for increased patient morbidity and mortality, length of hospitalization and higher healthcare costs. More recently, MRSA has emerged worldwide as a community-associated (CA-MRSA) pathogen, generating an additional public health concern. Although CA-MRSA are genetically different from nosocomial MRSA [1], the distinction between the two groups is blurring, since nowadays, CA-MRSA show multidrug resistance and are endemic in many hospitals [2,3,4,5].

The nosocomial prevalence of MRSA in Europe varies considerably. The current prevalence of HA-MRSA in Portugal (49.1%) is among the highest in the continent (EARSS Annual report 2009 [http://www.ecdc.europa.eu/en/publications/Publications/1011\\_SUR\\_annual\\_EARS\\_Net\\_2009.pdf](http://www.ecdc.europa.eu/en/publications/Publications/1011_SUR_annual_EARS_Net_2009.pdf)). Considering this prob-

lematic situation in Portuguese hospitals, different investigations have been performed to evaluate the extension of CA-MRSA in the country. A first study in the late 1990s and another one a decade later, including isolates from nasal swabs of young healthy individuals and nasopharyngeal swabs of children attending day care centers, respectively, reported a prevalence of MRSA lower than 1% in the Portuguese healthy community [6,7]. More recently, a study involving children attending the pediatric emergency department of a hospital due to skin and soft tissue infections, identified for the first time in Portugal a single CA-MRSA isolate ST80-IV producing the Pantone Valentine leukocidin [8]. Therefore, although MRSA is a major problem in Portuguese hospitals, the prevalence of CA-MRSA, among children and youth, seems to remain low in the country.

Transmission of *S. aureus*, including MRSA, occurs principally by direct human-to-human skin contact. However, *S. aureus* can survive for long periods on inanimate objects, which may

constitute an important reservoir for dissemination as well [9]. Therefore hand-touch surfaces in public transport vehicles, such as handrails, may represent a potential reservoir. Although methicillin susceptible *S. aureus* and methicillin resistant coagulase negative isolates have been found in public buses in London and Belgrade, respectively, MRSA was not detected in any of the studies [10,11]. Nevertheless, ambulances [12], patient homes [13], public areas of hospitals [14], and environmental surfaces at emergency medical responders facilities [15] were found to represent possible reservoirs of MRSA.

The aim of the present study was to explore the extension of public buses as a reservoir of MRSA in Portugal, which shows the second highest prevalence of nosocomial MRSA in Europe. The molecular characterization of the isolates and comparison with the MRSA clones spread among Portuguese hospitals provided insights into the origin of the isolates.

## Materials and Methods

### Screened vehicles

Between May 2009 and February 2010, handrails of 85 public urban buses circulating in Oporto, Portugal, were screened for the occurrence of MRSA. The participating buses were assigned to 12 different lines/routes (Table 1). On average, the vehicles have 35 seating places and 90 standing ones. Superficial cleaning of the buses occurs every day but disinfection happens every three months only.

### Sampling and bacterial isolates

Samples were collected, soon after the vehicles had ended transportation and before any putative cleaning, using sterile gauzes moistened in Brain Heart Infusion broth - BHI (Oxoid, United Kingdom, Basingstoke) supplemented with 0.1% Tween 80 (Merck, Germany, Darmstadt). In each bus, a single gauze was used to sample a large surface of different handrails. Samples were kept for a maximum of two hours at 4°C in BHI broth until

processing in the laboratory, where each sample was incubated at 37°C for approximately two hours. Subsequently, aliquots of 60 and 500 µ were inoculated onto three to five BBL™ CHROMagar™ Staph aureus plates (BD, NJ USA, Franklin Lakes), dried at 44°C for 10 minutes, and supplemented with 2 µg/ml of oxacillin. After 24 to 48 h of incubation at 37°C, all colonies exhibiting typical *S. aureus* morphology (mauve-colored colonies) were selected for antimicrobial susceptibility testing and storage. The plates with 500 µl of inoculum were incubated in an inverted position and partially open for the first 30 minutes. If any doubt subsisted concerning the species, isolates were tested for coagulase with a rapid latex agglutination test (BioMérieux, France, Marcy l'Etoile).

### Antimicrobial susceptibility testing

Antimicrobial susceptibility testing was performed by the disk diffusion method, following the Clinical and Laboratory Standards Institute guidelines [16] for a panel of 21 antimicrobial agents (Oxoid, United Kingdom, Basingstoke): oxacillin, ampicillin, cefoxitin, amoxicillin-clavulanic acid, ciprofloxacin, erythromycin, azithromycin, imipenem, kanamycin, tobramycin, gentamicin, quinupristin-dalfopristin, tetracycline, trimethoprim-sulfamethoxazole, rifampin, chloramphenicol, nitrofurantoin, clindamycin, linezolid, teicoplanin, and vancomycin. Methicillin resistance was confirmed on all isolates by PCR amplification of the *mecA* gene, as previously described [17].

### Pulsed-field gel electrophoresis

Pulsed-field gel electrophoresis (PFGE) was performed as previously described [18] on all MRSA isolates. The resulting *SmaI* restriction patterns were analysed by both visual inspection and computer analysis with Bionumerics version 6.1 software (Applied Maths, Sint-Martens-Latem, Belgium). Dendrogram was generated using an optimization of 0.5% and tolerance of 1.25% [19]. Similarity coefficients of 80% and 95% were used to define PFGE types and subtypes, respectively [19].

**Table 1.** Buses lines data and PFGE types found in the different vehicles contaminated with MRSA.

Bus line	Hospitals <sup>(b)</sup>	No. of buses contaminated with MRSA	PFGE types found in each contaminated bus (date of screening) <sup>(c)</sup>			Total number of MRSA isolates
1	1	3	A1 (16/06/2009)	A1 (16/06/2009)	A3 (27/10/2009)	8
2	1	1	A2 (19/01/2010)			1
3;7 <sup>(a)</sup>	2	1	A1 (23/09/2009)			1
4	0	1	A1 (11/11/2009)			3
5;9 <sup>(a)</sup>	1	1	A1 (24/11/2009)			3
6	1	1	A1 (11/02/2010)			3
7	1	2	A1 (16/06/2009)	A1, A6, C (11/11/2009)		8
8	2	3	A2 (11/11/2009)	A2 (11/11/2009)	A1, A4 (11/02/2010)	8
9	1	1	A2 (05/01/2010)			2
10	1	2	C (14/10/2009)	A1 (05/01/2010)		3
11	2	3	A1 (24/11/2009)	A5, B (05/01/2010)	A1 (11/02/2010)	10
12	2	1	A3 (05/01/2010)			3
12;7 <sup>(a)</sup>	3	2	A1 (16/06/2009)			2
12		22				55

<sup>(a)</sup>Two bus lines were assigned to the same vehicle during the screening day.

<sup>(b)</sup>Number of hospitals on each bus route.

<sup>(c)</sup>Each column corresponds to different buses assigned to the same bus line. Date of screening: day/month/year.

doi:10.1371/journal.pone.0017630.t001

**spa typing, MLST and SCCmec typing**

spa typing was performed on at least one representative isolate of each PFGE subtype, as previously described [20], and spa types were assigned through the Ridom web server (http://spaserver.ridom.de). Multilocus sequence typing (MLST) was carried out as described previously [21], on a representative isolate of each PFGE type. Allelic profiles and sequence types (ST) were assigned using the MLST database (http://www.mlst.net).

Staphylococcal cassette chromosome (SCC) mec was typed by the multiplex PCR strategy described by Milheiro et al. [22] in each representative isolate of a different PFGE subtype-spa type association. Additionally, all SCCmec type IV isolates were subtyped as previously described [23].

**PVL detection**

Panton Valentine leukocidin (PVL) detection was performed on all MRSA isolates according to a previous published protocol [24].

**Results and Discussion**

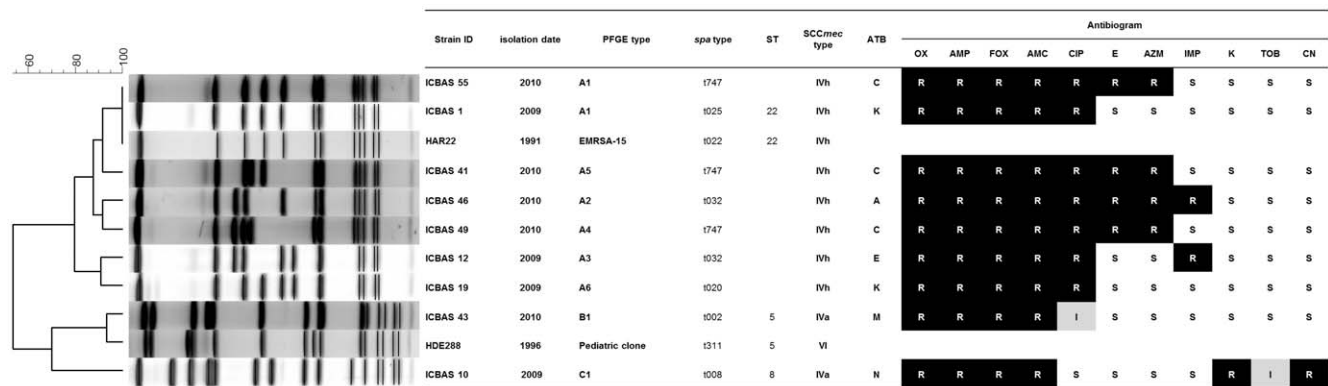
A total of 55 MRSA was recovered from 22 (26%) out of 85 buses. To our knowledge, this is the first study reporting MRSA in public urban buses. The molecular characterization of the 55 MRSA strains clustered the isolates into three clonal types (Figure 1). However, the overwhelming majority (n = 50; 91%) belonged to a single clone (PFGE A, spa types t747, t032, t025 or t020, ST22, SCCmec type IVh), which corresponds to the internationally disseminated EMRSA-15. In a study performed in 2006 in 11 Portuguese hospitals scattered all over the country, EMRSA-15 accounted for 54% of the total isolates [25]. The clone was detected in all hospitals and was the major lineage in seven of them, namely in the region of Oporto; EMRSA-15 represented 50% and 85% of the isolates in the two hospitals studied in Oporto and 82% in a hospital in Braga [25], a city located 50 km apart from Oporto. Moreover, a study performed in another hospital in Oporto showed that EMRSA-15 represented 79% of the isolates collected between 2003 and 2005 [26]. Recently, EMRSA-15 was also found as the prevalent clonal type (87%) in the Portuguese Azores islands [27]. The fact that 91% of the isolates recovered in the present study correspond to the major MRSA clonal type currently spread in all Portuguese hospitals [25,27] is noteworthy and evidences the dissemination of a

nosocomial MRSA clone from the hospital to the community environment. This situation may be explained, in part, by the fact that all but one bus lines included in the study pass near at least one hospital (Table 1), which in a country with a prevalence of nosocomial MRSA of 49.1% constitutes a plausible MRSA source. In addition, in Portugal, discharged patients are not decolonized for MRSA and screenings of health care workers colonization are not a routine practice. Hence, the contamination of the buses handrails most probably originated from the hands of health care workers, discharged patients, as well as outpatients and/or hospital visitors departing the hospitals.

Two additional clones were found in the present study but slightly represented, in agreement with the current situation in Portuguese hospitals: clone B - PFGE B, ST5, spa type t002, SCCmec IVa (n = 3; 6%) and clone C - PFGE C, spa type t008, ST8, SCCmec IVa (n = 2; 4%). Clone B is very similar to the Pediatric MRSA clone (Figure 1), which was described for the first time in a pediatric hospital in Lisbon [28]. Although the pediatric clone was a major clone in 1992–1997 in that particular hospital, it was detected in a single isolate only in the national study performed in 2006 [25]. Clonal type ST8-IV has been previously described mainly associated with CA-MRSA strains and in many cases coupled with PVL genes. In our study none of the 55 isolates was PVL positive.

PFGE subtype A1, the predominant (58%) subtype of PFGE type A, was found in all but one (n = 11) bus lines and during the whole study period (Table 1). The fact that the buses are only superficially cleaned every day, disinfected only every three months and that in a given day one to three different lines could be assigned to a same vehicle, may explain the fact that clonal type A1 is widely disseminated in this public transport network. Interestingly, in two cases, different PFGE types/subtypes were recovered in a same bus; PFGE types A1, A6 and C in one line 7 bus and PFGE types A5 and B in one line 11 bus (Table 1), evidencing the coexistence of different clonal types in a single vehicle.

Two previous studies have tried to demonstrate that public transports are a potential *S. aureus* reservoir, but both have failed to identify MRSA [10,11]. The high salt concentrations in the selective media used in one of the studies were suggested to have inhibited the growth of MRSA [29]. In the present work, plating onto a chromogenic agar with a pre-enrichment step using a



**Figure 1. Characterization of representative MRSA isolates and comparison with MRSA pandemic clones.** From left to right: (i) dendrogram, showing the estimated relationships of PFGE types based on Bionumerics analysis, including representatives of two international pandemic MRSA clones (EMRSA-15 and Pediatric clone); (ii) list of isolates; (iii) isolation date; (iv) PFGE type; (v) spa type; (vi) MLST sequence type (ST); (vii) SCCmec type; (viii) antibiotype (ATB); and (ix) antibiogram. R - resistance, I - intermediate susceptibility, S - susceptibility; Antibiotic abbreviations: OX - oxacillin, AMP - ampicillin, FOX - ceftiofexim, AMC - amoxicillin-clavulanic acid, CIP - ciprofloxacin, E - erythromycin, AZM - azithromycin, IMP - imipenem, K - kanamycin, TOB - tobramycin, CN - gentamicin. doi:10.1371/journal.pone.0017630.g001

nonselective broth (BHI), might have contributed to the recovery of a significant number of MRSA isolates. Eventually, the screening could have been improved if using a semi-selective enrichment broth as described recently by Bocher and collaborators [30]. The second study published attributed the absence of MRSA in public buses to the low prevalence of MRSA carriage in the local healthy population [10]. However, in the present study public buses were found to constitute an important MRSA reservoir in Portugal despite the low MRSA colonization or infection in the community [6,7]. Nevertheless, since the screenings for CA-MRSA were performed in the Southern region of Portugal (Lisbon and Montemor-o-Novo) future studies are required to assess the prevalence of EMRSA-15 among the general population in the region of Oporto where public buses were recently found to be highly contaminated with this MRSA lineage.

The smaller number of buses screened in the two previous studies, i.e. two buses in the London study [10] and 55 in the Belgrade work [11], may also be an explanation for the failure to identify MRSA. Additionally, a higher frequency of disinfection could also have contributed to the absence of MRSA. However, and although we do not possess information concerning the Belgrade buses, the London study reported that the surfaces sampled were infrequently cleaned sites [10]. EMRSA-15, the predominant clone highly spread in Oporto buses, harbors SCCmec type IV, one of the smaller cassettes, and consequently shows faster growth rates and resistance to a limited number of antimicrobial agents (Figure 1). Therefore, although it is a hospital-associated clone, EMRSA-15 shows similar traits to CA-MRSA that may favor its survival and persistence in the community. Although ST22-IV isolates had been previously found in the community in different animal species, particularly in dogs and cats, but also in turtles, bats and pet birds [31], and found to be able to circulate in human Australian remote communities [32], the first documented isolation of EMRSA-15 among healthy individuals with no HA-MRSA risk factors was only recently published from Ireland [33]. Consequently, the high prevalence of MRSA in Portuguese public buses might represent a first step for the massive spread of EMRSA-15 into the general population.

Allied to the fact that antimicrobial agents may still be bought over the counter in Portugal, the country shows one of the highest

rates of outpatient antibiotic sales among the European Union [34], which may constitute sufficient selective pressure to maintain this MRSA clone in the community environment. In 2007, the annual outpatient antibiotics consumption in Portugal was estimated as 21.86 daily doses per 1000 people per day and the most used antibiotics were penicillins (52%), followed by macrolides (18%), quinolones (13%) and cephalosporins (10%) [35]. These correlated to the antimicrobial agents that showed decreased susceptibility among the isolates contaminating the buses in Oporto (Figure 1).

In addition, fat of hand sweat, warm temperatures, and appropriate humidity conditions observed in Portuguese buses circulating in the city of Oporto may also have played a role in the survival of *S. aureus* on the handrails of the vehicles. Tolba et al. demonstrated that *S. aureus* can easily be recovered from metallic surfaces and can survive for long periods in adequate conditions [36]. However, since the buses in Oporto circulate several times a day near at least one hospital, the MRSA isolates are probably being constantly (several times a day) inoculated on the handrails, which makes it difficult to distinguish between long time survival and recent inoculation.

In summary, public buses in Oporto seem to be an important reservoir of MRSA of nosocomial origin, providing evidence that the major hospital-associated MRSA clone in Portugal is escaping from the primary ecological niche of hospitals to the community environment. Consequently, infection control measures are urgently warranted to limit the spread of EMRSA-15 to the general population and future studies are required to assess the eventual increase of MRSA in the Portuguese community, which so far remains low.

## Acknowledgments

We are grateful to the public transports network for allowing the study and for the given assistance during the screening.

## Author Contributions

Conceived and designed the experiments: MAS PMC HL. Performed the experiments: RRS TC FA. Analyzed the data: TC RRS MAS PMC HL. Wrote the paper: MAS RRS HL.

## References

- Naimi TS, LeDell KH, Como-Sabetti K, Borchardt SM, Boxrud DJ, et al. (2003) Comparison of community- and health care-associated methicillin-resistant *Staphylococcus aureus* infection. *Jama* 290: 2976–2984.
- Udo EE, Sarkhoo E (2010) The dissemination of ST80-SCCmec-IV community-associated methicillin resistant *Staphylococcus aureus* clone in Kuwait hospitals. *Ann Clin Microbiol Antimicrob* 9: 31.
- Gonzalez BE, Rueda AM, Shelburne SA, 3rd, Musher DM, Hamill RJ, et al. (2006) Community-associated strains of methicillin-resistant *Staphylococcus aureus* as the cause of healthcare-associated infection. *Infect Control Hosp Epidemiol* 27: 1051–1056.
- Moran GJ, Krishnadasan A, Gorwitz RJ, Fosheim GE, McDougal LK, et al. (2006) Methicillin-resistant *S. aureus* infections among patients in the emergency department. *N Engl J Med* 355: 666–674.
- McAdams RM, Ellis MW, Trevino S, Rajnik M (2008) Spread of methicillin-resistant *Staphylococcus aureus* USA300 in a neonatal intensive care unit. *Pediatr Int* 50: 810–815.
- Sá-Leão R, Sanches IS, Couto I, Alves CR, de Lencastre H (2001) Low prevalence of methicillin-resistant strains among *Staphylococcus aureus* colonizing young and healthy members of the community in Portugal. *Microb Drug Resist* 7: 237–245.
- Tavares DA, Sá-Leão R, Miragaia M, de Lencastre H (2010) Large screening of CA-MRSA among *Staphylococcus aureus* colonizing healthy young children living in two areas (urban and rural) of Portugal. *BMC Infect Dis* 10: 110.
- Conceição T, Aires-de-Sousa M, Pona N, Brito MJ, Barradas C, et al. (2010) High prevalence of ST121 in community-associated methicillin-susceptible *Staphylococcus aureus* lineages responsible for skin and soft tissue infections in Portuguese children. *Eur J Clin Microbiol Infect Dis* 30: 293–297.
- Neely AN, Maley MP (2000) Survival of enterococci and staphylococci on hospital fabrics and plastic. *J Clin Microbiol* 38: 724–726.
- Otter JA, French GL (2009) Bacterial contamination on touch surfaces in the public transport system and in public areas of a hospital in London. *Lett Appl Microbiol* 49: 803–805.
- Stepanovic S, Cirkovic I, Djukic S, Vukovic D, Svabic-Vlahovic M (2008) Public transport as a reservoir of methicillin-resistant staphylococci. *Lett Appl Microbiol* 47: 339–341.
- Roline CE, Crumpecker C, Dunn TM (2007) Can methicillin-resistant *Staphylococcus aureus* be found in an ambulance fleet? *Prehosp Emerg Care* 11: 241–244.
- Allen KD, Anson JJ, Parsons LA, Frost NG (1997) Staff carriage of methicillin-resistant *Staphylococcus aureus* (EMRSA 15) and the home environment: a case report. *J Hosp Infect* 35: 307–311.
- Manning N, Wilson AP, Ridgway GL (2004) Isolation of MRSA from communal areas in a teaching hospital. *J Hosp Infect* 56: 250–251.
- Sexton JD, Reynolds KA (2010) Exposure of emergency medical responders to methicillin-resistant *Staphylococcus aureus*. *Am J Infect Control* 38: 368–373.
- CLSI (2009) Performance standards for antimicrobial disk susceptibility tests; approved standard, 10th edn. WaynePA: CLSI publication M02-A10. Clinical and Laboratory Standards Institute.
- Murakami K, Minamide W, Wada K, Nakamura E, Teraoka H, et al. (1991) Identification of methicillin-resistant strains of staphylococci by polymerase chain reaction. *J Clin Microbiol* 29: 2240–2244.
- Chung M, de Lencastre H, Matthews P, Tomasz A, Adamsson I, et al. (2000) Molecular typing of methicillin-resistant *Staphylococcus aureus* by pulsed-field gel electrophoresis: comparison of results obtained in a multilaboratory effort using identical protocols and MRSA strains. *Microb Drug Resist* 6: 189–198.

19. Faria NA, Carriço JA, Oliveira DC, Ramirez M, de Lencastre H (2008) Analysis of typing methods for epidemiological surveillance of both methicillin-resistant and methicillin-susceptible *Staphylococcus aureus* strains. *J Clin Microbiol* 46: 136–144.
20. Harmsen D, Claus H, Witte W, Rothganger J, Turnwald D, et al. (2003) Typing of methicillin-resistant *Staphylococcus aureus* in a university hospital setting by using novel software for *spa* repeat determination and database management. *J Clin Microbiol* 41: 5442–5448.
21. Aires-de-Sousa M, Parente CE, Vieira-da-Motta O, Bonna IC, Silva DA, et al. (2007) Characterization of *Staphylococcus aureus* isolates from buffalo, bovine, ovine, and caprine milk samples collected in Rio de Janeiro State, Brazil. *Appl Environ Microbiol* 73: 3845–3849.
22. Milheiro C, Oliveira DC, de Lencastre H (2007) Update to the multiplex PCR strategy for assignment of *mec* element types in *Staphylococcus aureus*. *Antimicrob Agents Chemother* 51: 3374–3377.
23. Milheiro C, Oliveira DC, de Lencastre H (2007) Multiplex PCR strategy for subtyping the staphylococcal cassette chromosome *mec* type IV in methicillin-resistant *Staphylococcus aureus*: ‘SCC*mec* IV multiplex’. *J Antimicrob Chemother* 60: 42–48.
24. Lina G, Piemont Y, Godail-Gamot F, Bes M, Peter MO, et al. (1999) Involvement of Panton-Valentine leukocidin-producing *Staphylococcus aureus* in primary skin infections and pneumonia. *Clin Infect Dis* 29: 1128–1132.
25. Aires-de-Sousa M, Correia B, de Lencastre H (2008) Changing patterns in frequency of recovery of five methicillin-resistant *Staphylococcus aureus* clones in Portuguese hospitals: surveillance over a 16-year period. *J Clin Microbiol* 46: 2912–2917.
26. Amorim ML, Faria NA, Oliveira DC, Vasconcelos C, Cabeda JC, et al. (2007) Changes in the clonal nature and antibiotic resistance profiles of methicillin-resistant *Staphylococcus aureus* isolates associated with spread of the EMRSA-15 clone in a tertiary care Portuguese hospital. *J Clin Microbiol* 45: 2881–2888.
27. Conceição T, Tavares A, Miragaia M, Hyde K, Aires-de-Sousa M, et al. (2010) Prevalence and clonality of methicillin-resistant *Staphylococcus aureus* (MRSA) in the Atlantic Azores islands: predominance of SCC*mec* types IV, V and VI. *Eur J Clin Microbiol Infect Dis* 29: 543–550.
28. Sá-Leão R, Santos Sanches I, Dias D, Peres I, Barros RM, et al. (1999) Detection of an archaic clone of *Staphylococcus aureus* with low-level resistance to methicillin in a pediatric hospital in Portugal and in international samples: relics of a formerly widely disseminated strain? *J Clin Microbiol* 37: 1913–1920.
29. Kassem II (2009) Concerning public transport as a reservoir of methicillin-resistant staphylococci. *Lett Appl Microbiol* 48: 268.
30. Bocher S, Middendorf B, Westh H, Mellmann A, Becker K, et al. (2010) Semi-selective broth improves screening for methicillin-resistant *Staphylococcus aureus*. *J Antimicrob Chemother* 65: 717–720.
31. Cuny C, Friedrich A, Kozytka S, Layer F, Nubel U, et al. (2010) Emergence of methicillin-resistant *Staphylococcus aureus* (MRSA) in different animal species. *Int J Med Microbiol* 300: 109–117.
32. O’Brien FG, Lim TT, Chong FN, Coombs GW, Enright MC, et al. (2004) Diversity among community isolates of methicillin-resistant *Staphylococcus aureus* in Australia. *J Clin Microbiol* 42: 3185–3190.
33. Mollaghan AM, Lucey B, Coffey A, Cotter L (2010) Emergence of MRSA clone ST22 in healthy young adults in the community in the absence of risk factors. *Epidemiol Infect* 138: 673–676.
34. Cars O, Molstad S, Melander A (2001) Variation in antibiotic use in the European Union. *Lancet* 357: 1851–1853.
35. Ramalinho I, Cabrita J, Ribeirinho M, Vieira I Evolução do consumo de antibióticos em Portugal Continental (2000 – 2007). Lisbon: [http://www.infarmed.pt/portal/page/portal/INFARMED/MONITORIZACAO\\_DO\\_MERCADO/OBSERVATORIO/ESTUDOS\\_REALIZADOS\\_PROTOCOLOS/Evolu%E7%E3o\\_Consumo\\_Ab\\_Portugal.pdf](http://www.infarmed.pt/portal/page/portal/INFARMED/MONITORIZACAO_DO_MERCADO/OBSERVATORIO/ESTUDOS_REALIZADOS_PROTOCOLOS/Evolu%E7%E3o_Consumo_Ab_Portugal.pdf).
36. Tolba O, Loughrey A, Goldsmith CE, Millar BC, Rooney PJ, et al. (2007) Survival of epidemic strains of nosocomial- and community-acquired methicillin-resistant *Staphylococcus aureus* on coins. *Am J Infect Control* 35: 342–346.