

Histamine–Releasing Factor and Immunoglobulins in Asthma and Allergy

Toshiaki Kawakami,^{1,2*} Jun-ichi Kashiwakura,² Yuko Kawakami¹

¹ Division of Cell Biology, La Jolla Institute for Allergy and Immunology, La Jolla, California, USA ² Laboratory of Allergic Disease, RIKEN Center for Integrative Medical Sciences (IMS-RCAI), Yokohama, Kanagawa, Japan

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/3.0/) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Factors that can induce the release of histamine from basophils have been studied for more than 30 years. A protein termed histamine-releasing factor (HRF) was purified and molecularly cloned in 1995. HRF can stimulate histamine release and IL-4 and IL-13 production from IgE-sensitized basophils and mast cells. HRF-like activities were found in bodily fluids during the late phase of allergic reactions, implicating HRF in allergic diseases. However, definitive evidence for the role of HRF in allergic diseases has remained elusive. On the other hand, we found effects of monomeric IgE on the survival and activation of mast cells without the involvement of a specific antigen, as well as heterogeneity of IgEs in their ability to cause such effects. The latter property of IgE molecules seemed to be similar to the heterogeneity of IgEs in their ability to prime basophils in response to HRF. This similarity led to our recent finding that ~30% of IgE molecules can bind to HRF via their Fab interactions with two binding sites within the HRF molecule. The use of peptide inhibitors that block HRF-IgE interactions revealed an essential role of HRF to promote skin hypersensitivity and airway inflammation. This review summarizes this and more recent findings and provides a perspective on how they impact our understanding of allergy pathogenesis and potentially change the treatment of allergic diseases.

Key Words: Asthma; allergy; histamine-releasing factor; IgE; mast cell; basophil

INTRODUCTION

The mast cell has long been known to be a key effector cell type in IgE-mediated immediate hypersensitivity and allergic disorders.^{1,2} although recent studies have added their novel roles in host defense against bacteria and bee and snake venoms, as well as immunoregulatory roles.3 Mast cells are activated by cross-linking of IgE-bound high-affinity IgE receptors (FcERI) with multivalent antigen.⁴ Activated mast cells secrete preformed proinflammatory mediators, such as histamine, serotonin, nucleotides, proteases, and tumor necrosis factor (TNF)- α , and synthesize and secrete various cytokines, chemokines, and lipid mediators, such as leukotrienes and prostaglandins. The past two decades showed tremendous progress in our understanding of what happens inside the mast cells whose activation is triggered by IgE plus specific antigen.⁵⁻⁷ Thus, many scientists, particularly those interested in FcERI signal transduction research from outside the field, seem to be inclined to think that we have a very good understanding of how mast cells are activated through the allergen-IgE-FcERI axis.

In contrast to the long-held view of the requirement of multi-

valent antigen (or anti-IgE antibody) to cross-link IgE-bound FcERI, we and others have demonstrated that monomeric IgE binding to FcERI in the absence of specific antigen elicits numerous biological activities in mast cells: upregulated cell surface expression of FcERI,⁸⁻¹⁰ survival,^{11,12} increased histamine content,¹³ histamine release, leukotriene release, receptor internalization, DNA synthesis,¹⁴ increased responses to compound 48/80 and substance P,¹⁵ increased filamentous actin content,¹⁶ membrane ruffling,¹⁷ adhesion to fibronectin,¹⁸ and migration.¹⁹ Similar to cells stimulated with IgE plus antigen, FcERI cross-linking appears to be essential for mast cell activation induced by monomeric IgE.¹⁴

Correspondence to: Toshiaki Kawakami, MD, PhD, Division of Cell Biology, La Jolla Institute for Allergy and Immunology, 9420 Athena Circle, La Jolla, California 92037-1387, USA.

Tel: +1-858-752-6814; Fax: +1-858-752-6986; E-mail: toshi@liai.org Received: November 2, 2012; Revised: December 27, 2012 Accepted: January 15, 2013

• There are no financial or other issues that might lead to conflict of interest.

HETEROGENEITY OF IgE MOLECULES

Two papers published in 2001 showed the same effects of monomeric IgE, *i.e.*, survival promotion, as well as disparate results. Kalesnikoff et al.¹² showed that monomeric IgE could induce cytokine secretion and the corresponding signaling data, whereas Asai et al.¹¹ reported the absence of cytokine production. A follow-up study revealed that this difference stems from the difference in the IgE molecules used.¹⁴ Importantly, IgEs display a wide range of heterogeneity in their ability to induce the production and secretion of IL-6 and TNF- α and other activation events in mouse mast cells. At one end of the spectrum, highly cytokinergic (HC) IgEs can induce not only strong survival promotion but also all other activation events listed in the preceding paragraph, whereas, at the other end of the spectrum, poorly cytokinergic (PC) IgEs can induce only weak survival enhancement.¹⁴ More extensive receptor aggregation can be induced by HC IgEs than by PC IgEs. Polyclonal IgE from mice and humans suffering from atopic dermatitis (AD) can enhance survival and cytokine production in mast cell cultures, indicating that monomeric IgE effects are operative in polyclonal situations as well.²⁰

The difference between HC and PC IgEs appeared to be similar to the IgE+/IgE- dichotomy with regard to the priming ability of IgE in response to the relatively unknown cytokine-like protein histamine-releasing factor (HRF). HRF is an evolutionally conserved protein with a molecular mass of ~26 kDa.21 Prior to the purification and cloning of HRF in 1995, HRF-like activities had been reported from a variety of cellular sources including alveolar macrophages, platelets, vascular endothelial cells, B and T lymphocytes, mononuclear cell cultures, the U937 monocyte/macrophage-like cell line, and the RPMI 8866 B cell line.²² HRF-like activity was also found in nasal lavages, skin blister fluids, and bronchoalveolar lavage fluids during the late phase of allergic reactions.²³⁻²⁵ Without a signal sequence, the secretion of HRF is insensitive to brefeldin A or monensin, but it can be enhanced by TSAP6, a p53-inducible 5-6 transmembrane protein.²⁶ HRF can be found in exosomes, suggesting that it is secreted through a nonclassical exosome pathway.²⁶ Because human HRF can stimulate histamine release and cytokine (IL-4 and IL-13) production from IgE-sensitized (but not IgE-depleted) basophils and mast cells, 21,27,28 it is considered an IgE-dependent cytokine. MacDonald et al. revealed functional heterogeneity among human IgE molecules: IgE from HRF-responder (HRF-R) basophils derived from ~50% of atopic patients was termed IgE+, and IgE from nonresponders (HRF-NR) was termed IgE-.²⁹ However, heterogeneity in the carbohydrate portion of IgE molecules failed to distinguish between IgE+ and IgE-.³⁰ Interestingly, HRF responses in human basophils were shown to be negatively correlated with SHIP (SH2 domain-containing phosphatidylinositol 5'-phosphatase), but not Syk (spleen tyrosine kinase), levels,³¹ explaining some HRF-R subjects. Despite these efforts, the exact molecular basis of the IgE+/IgE- dichotomy remained an enigma for a number of years.

HRF BINDS A SUBSET OF IgE AND IgG MOLECULES

The most obvious interpretation of the two similar phenomena of IgE heterogeneity leads to the idea that some, but not all, IgE molecules interact with HRF. However, Wantke et al. thought that HRF does not bind to IgE, because they previously observed that HRF primed anti-IgE-antibody-induced histamine release from all basophils, irrespective of the type of IgE on the cell surface.³² So, they performed experiments using a rat basophilic leukemia cell line (RBL-SX38), which was engineered to express a functional human $Fc\epsilon RI$. They found that neither human nor mouse recombinant HRF caused histamine release in RBL-SX38 cells sensitized with IgE+. Additionally, priming the transfected RBL-SX38 cells or the parental cell line, RBL-2H3 cells, with human or mouse recombinant HRF did not increase anti-IgE-induced histamine release. Based on these results, the authors concluded that HRF does not bind to either IgE+ or IgE-.

Because we later (in 2003) found the heterogeneity of IgEs in their monomeric effects,¹⁴ we decided to pursue the most obvious possibility, namely IgE-HRF binding. However, being concerned this time that only a small proportion of IgEs might interact with HRF, we tested a relatively large panel of IgE mAbs. Simple ELISA experiments using GST-mHRF (glutathione S-transferase [GST] fused to mouse HRF) as a capturing agent, our experiments showed that six of 19 IgE mAbs tested bound GST-mHRF but not GST. Some of the HRF-reactive IgEs were also used to confirm HRF binding by two other methods. Similar proportions of IgG mAbs also bound GST-mHRF. Binding properties were not determined by antigen specificity or IgG isotype. Affinity measurement using the quartz crystal microbalance method³³ indicated that these interactions were of low affinity ($K_D = \sim 10^{-6}$ M).

DEVELOPMENT OF HRF INHIBITORS AND THEIR USE TO INHIBIT ALLERGIC INFLAMMATION

HRF-binding sites were mapped to Fab, not Fc, portions of IgE and IgG molecules. The HRF-binding sites seemed to be close to or to overlap at least partially with antigen-binding sites. On the other hand, IgE- (and IgG-) binding sites were mapped to the N-terminal 19 residues (N19) and the H3 (residues 107-135) portion of mHRF. Importantly, both GST-N19 and GST-H3 could inhibit HRF-IgE (or IgG) interactions in ELISA (Fig. 1). We also confirmed previous data showing that HRF can be present as a disulphide-bonded dimer,³⁴ suggesting that HRF should be able to induce cross-linking of HRF-reactive IgE-bound FcERI (Fig. 2), which was supported by mast cell activation induced by IgE plus HRF. This activation could be inhibited by GST-N19 or GST-H3. These results suggested that GST-N19 and GST-H3 may be useful reagents to probe the function of HRF in *in vivo* models

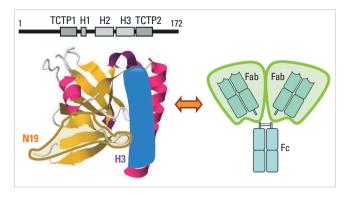


Fig. 1. Interaction sites between HRF and IgG. Ig-binding sites were mapped to the N-terminal 19 residues (N19) and the H3 (residues 107-135) portion of mHRF, while HRF binds to the Fab portion of Igs. The top left and bottom left panels show the domain structure and the 3-D structure of human HRF (Protein Data Bank, 1YZ1), respectively. The N19 and H3 portions are highlighted. The TCTP1 portion is omitted because it does not form electron-dense structures. The right panel depicts a cartoon of an IgG molecule.

of allergic diseases. However, they are required to not affect HRF's intracellular functions (see Box) because those functions are essential for fundamental cellular properties such as proliferation and survival. Fortunately, neither GST-N19 nor GST-H3 affected the growth or survival of various cultured cells; this was shown to be due to their failure to enter the cell.

Using these HRF inhibitors, we could show that passive cutaneous anaphylaxis (PCA) induced by intradermal antigen challenge in mice primed with HRF-reactive IgE can be substantially ameliorated by GST-N19 or GST-H3. Combined pretreatment with GST-N19 and GST-H3 largely inhibited PCA reactions. We also demonstrated that pretreatment with GST-N19 before allergen (i.e., ovalbumin [OVA]) challenge strongly inhibited OVAinduced mast cell-dependent airway inflammation, as revealed by reduced eosinophilia in the airway, reduced immune cell infiltration in the lungs, reduced IL-5 and IL-13 in the lungs, and reduced airway hyperresponsiveness. Synthetic N19 peptide also showed similar efficacy to inhibit airway inflammation. However, GST-N19 was ineffective in inhibiting T cell-dependent (but mast cell-independent) airway inflammation induced by OVA challenge in OVA+alum-sensitized mice. A partially IgE-dependent model induced by intranasal administration of Aspergillus fumigatus extract was partially inhibited by GST-N19. Therefore, these results suggest that mast cells are target cells for HRF to promote airway and skin inflammation induced by IgE.

MECHANISM OF HRF-MEDIATED INFLAMMATION

The efficacy of HRF inhibitors in the suppression of PCA induced by HRF-reactive IgE (anti-TNP IgE mAb C38-2) and antigen (TNP-BSA) can be understood as follows: IgE-antigen interactions may be inhibited by HRF inhibitors because the interaction of HRF-reactive IgE with HRF was at least partially in-

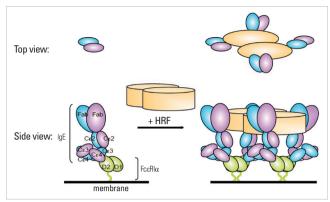


Fig. 2. Working model of HRF-mediated FccRI crosslinking. IgE binds FccRI α chain via the interaction between Cc3 and D2 domains. HRF can exist as a dimer, and one HRF molecule can bind to two molecules of IgE via interactions with the N19 and H3 regions of HRF. The top view (Top) of IgE at the level of Fab and the side view (Bottom) of IgE and IgE-bound FccRI α chain are shown on the left. After binding the HRF dimer, four FccRI α chain-nucleated complexes will be formed (Right). The cytoplasmic portion of FccRI α as well as β and γ chains of FccRI are omitted for clarity.

hibited by monovalent TNP hapten. However, it is much more difficult to understand how HRF affects airway inflammation because airway inflammation involves a complex interplay of various types of cells and many soluble and insoluble factors. Therefore, we pursued the development of a simpler protocol to study HRF-mediated inflammation. Intranasal administration of recombinant mHRF in naïve mice has been shown to induce weak lung inflammation. Similar to transgenic mice overexpressing HRF in a Clara cell-specific manner,³⁵ this lung inflammation consists mainly of macrophages. Using various mutant mice, we could show that this inflammation required B cells (source of immunoglobulins [Igs]) and mast cells (target cells), as well as FcRy. FcRy is shared by multiple Fc receptors including FcERI, FcyRI, FcyRIIIA, and FcyRIV.^{36,37} Among the Igs and Fc receptors, IgE and FcERI were the predominant contributors to the HRF effects, as HRF-induced lung inflammation was abrogated in naïve FccRI--- mice. These results were consistent with the idea that IgE (and IgG) was the long-sought receptor for HRF. Global gene expression analysis was instrumental in further strengthening this idea. The expression of 196 genes was up- or down-regulated by over three fold by HRF in the lungs of naïve WT mice. Upregulated genes encode Th1-, Th2-, and Th17-associated cytokines and various chemokines, potentially accounting for the recruitment of monocytes/macrophages, neutrophils, eosinophils, and other immune cells. In contrast, only a small fraction of these genes (39 of 196 genes) fluctuated by more than three fold in *Fc* $\epsilon RIa^{-/-}$ mice; furthermore, fewer (11 of 196 genes) genes were changed in $FcR\gamma^{-/-}$ mice. These results suggest that HRF executes its action largely, if not exclusively, by engaging IgE- and IgG-bound Fc receptors and promotes airway inflammation. Collectively, these findings solved three major HRF-related problems. First, the HRF receptor was identified. Second, the key role of HRF in allergic disease models was demonstrated. Third, the IgE+/IgE- dichotomy is now understood as the dependency of IgEs on their reactivity with HRF. What about other HRF functions? By definition, the functions of HRF should be limited to those of the extracellularly secreted protein, but not those of the intracellular protein TCTP (see Box).

CAN WE EXPALIN ALL KNOWN PROPERTIES OF HRF?

Much of the 'HRF enigma' has been solved. However, HRF is also known as a B cell growth factor.³⁸ HRF can stimulate IL-8 secretion from GM–CSF-primed eosinophils.³⁹ HRF can inhibit PMA+A23187-stimulated cytokine gene expression in T cells.⁴⁰ More recently, HRF has been shown to stimulate bronchial epithelial cells to produce IL-8 and GM-CSF.⁴¹ It is tempting to speculate that these HRF functions could occur through HRF's interactions with B cell antigen receptor, FccRI- or FccRII (CD23)bound IgE, or FcγR -bound IgG. Examination of these possibilities is highly warranted.

HRF AS A SUPERANTIGEN OR AUTOANTIGEN

The broad reactivity of HRF with the Fab portion of Igs (~30% of Igs) and its ability to activate mast cells indicate that HRF can be considered a 'mast cell superantigen'. In addition to typical superantigens (such as staphylococcal enterotoxin B), which can bind to MHC II and a substantial subset of T cell receptors, there are B cell superantigens (such as Staphylococcal protein A, Peptostreptococcus magnus protein L, and HIV gp120), which target large clonal sets of B cells. B cell superantigens interact with the conserved antigen receptor variable (V)-region sites, which are different from antigen-binding sites.⁴² Some B cell superantigens can also activate mast cells and/or basophils by interacting with FccRI-bound IgE.43-45 However, unlike the B cell superantigens, the HRF-binding sites at least partially overlap with antigen-binding sites. Our sequence analysis of a limited number of IgE and IgG molecules indicated that HRF-reactive Igs use specific V gene families of κ (kappa) light chain.⁴⁶ The exact binding sites must be analyzed through structural studies.

More recently, we found that IgEs' binding of autoantigens correlates with differences in HC versus PC properties.⁴⁷ Most mouse monoclonal HC IgEs exhibited polyreactivity to doublestranded (ds) DNA, single-stranded (ss) DNA, β -galactosidase, thyroglobulin, and/or HRF. In contrast, mouse PC IgEs failed to react with these antigens. A human monoclonal HC IgE also displayed polyreactivity to HRF, dsDNA, and ssDNA. Interestingly, sera from AD patients showed increased reactivity to ssD-NA and β -galactosidase and increased levels of HRF. Some AD patients, but not healthy individuals, had substantial serum levels of HRF-reactive IgE. Sera from AD patients with high titers of DNA-reactive IgE could induce several fold more IL-8 secretion in human mast cells than could sera from healthy individuals. These results show that most HC, but not PC, IgEs exhibit polyreactivity to autoantigens, supporting the autoimmune mechanism in the pathogenesis of AD. It has been thought that autoimmunity underlies allergic diseases.⁴⁸ Consistent with this notion, the prevalence of allergies and autoimmune diseases has increased in parallel.^{49,50} Recent studies support the concept that IgE autoreactivity may play a pathogenic role in severe and chronic forms of atopy.48,51 Acute allergic reactions following exposure to exogenous allergens can be understood as immediate-type inflammation triggered by degranulation of mast cells via allergen-IgE-FcERI interactions, whereas chronic allergic inflammation with Th1 characteristics can occur and persist in the absence of exogenous allergens. IgE-mediated presentation of autoallergens may activate autoreactive Th1 cells to release proinflammatory cytokines.^{52,53} Several environmental allergens share striking structural and immunologic similarities with human proteins.⁵⁴⁻⁵⁸ A large number (\geq 140 proteins) of IgE-reactive autoantigens were found in AD patients, and some of them were shown to have the abili-ty to activate basophils.⁵⁹ Therefore, the autoimmune mechanism could be an important part of the disease pathogenesis in a subset of AD patients.

FUTURE PERSPECTIVE

Our studies imply that elevated levels of HRF and/or HRF-reactive Igs activate mast cells and basophils and amplify inflam-

Box. Intracellular functions of HRF (TCTP) versus extracellular functions of HRF

Despite the name "translationally controlled tumor protein", TCTP expression is subject to both transcriptional and translational control. TCTP within a cell plays a critical role in the fundamental processes of cell-cycle progression, proliferation, survival, and malignant transformation. TCTP is a Ca²⁺-binding protein.⁶⁰⁻⁶³ It is involved in the elongation step of protein synthesis by interacting with both eEF1A (a small GTPase) and eEF1Bβ (a guanine nucleotide exchange factor).⁶⁴⁻⁶⁶ *Drosophila* TCTP acts as the guanine nucleotide-exchange factor for Rheb (Ras homologue enriched in brain), a Ras superfamily GTPase that regulates the TSC1-mTOR pathway.⁶⁷ TCTP interacts with Mcl-1^{68,69} and Bcl-xL,⁷⁰ anti-apoptotic members of the Bcl-2 family. Recent studies identified mutual antagonism between p53 tumor suppressor and TCTP, with TCTP promoting p53 degradation via MDM2-mediated ubiquitination of p53 and with p53 directly repressing TCTP transcription.^{71,72} Another conserved property of TCTP is its interaction with microtubules and mitochondria.⁷³ Readers interested in this topic are referred to Bommer.⁷⁴

TCTP and HRF are the same protein. The extracellular functions of the protein are the focus of this review. As discussed in the text, they include the activation of mast cells and basophils whose FccRI molecules are occupied with HRF-reactive IgE.

mation.^{46,47} Although it is unknown whether HRF can activate mast cells and basophils under homeostatic conditions (or physiological levels and the repertoire of IgE and IgG), HRF-mediated mast cell activation should contribute to the amplification of allergic reactions when IgE, IgG, and HRF levels are high enough at the allergic inflammatory site. It is noteworthy that there is a level of HRF in normal serum that does not induce noticeable inflammation. Thus, there should be a mechanism(s) by which normal levels of HRF do not harm the animal. As a relatively large fraction (~30%) of Igs can bind HRF, changes in the antigen repertoire of Igs may cause a change in the levels of HRF-reactive Igs. Thus, it will be interesting to investigate the potential involvement of HRF in other pathological conditions such as autoimmune or infectious diseases. Overall, recent studies encourage us to further understand the mechanism by which HRF plays a proinflammatory role in allergy and other diseases. As human IgE also can interact with human HRF, attempts to translate the most recent observations to human pathology will be a major focus in the future, raising hope for improving the arsenal with which we combat allergic diseases.

REFERENCES

- 1. Galli SJ. Mast cells and basophils. Curr Opin Hematol 2000;7:32-9.
- Galli SJ, Maurer M, Lantz CS. Mast cells as sentinels of innate immunity. Curr Opin Immunol 1999;11:53-9.
- Galli SJ, Grimbaldeston M, Tsai M. Immunomodulatory mast cells: negative, as well as positive, regulators of immunity. Nat Rev Immunol 2008;8:478-86.
- 4. Metzger H. The receptor with high affinity for IgE. Immunol Rev 1992;125:37-48.
- 5. Kalesnikoff J, Galli SJ. New developments in mast cell biology. Nat Immunol 2008;9:1215-23.
- 6. Gilfillan AM, Rivera J. The tyrosine kinase network regulating mast cell activation. Immunol Rev 2009;228:149-69.
- Rivera J, Fierro NA, Olivera A, Suzuki R. New insights on mast cell activation via the high affinity receptor for IgE. Adv Immunol 2008; 98:85-120.
- 8. Furuichi K, Rivera J, Isersky C. The receptor for immunoglobulin E on rat basophilic leukemia cells: effect of ligand binding on receptor expression. Proc Natl Acad Sci U S A 1985;82:1522-5.
- 9. Hsu C, MacGlashan D Jr. IgE antibody up-regulates high affinity IgE binding on murine bone marrow-derived mast cells. Immunol Lett 1996;52:129-34.
- Yamaguchi M, Lantz CS, Oettgen HC, Katona IM, Fleming T, Miyajima I, Kinet JP, Galli SJ. IgE enhances mouse mast cell Fc(epsilon) RI expression in vitro and in vivo: evidence for a novel amplification mechanism in IgE-dependent reactions. J Exp Med 1997;185:663-72.
- 11. Asai K, Kitaura J, Kawakami Y, Yamagata N, Tsai M, Carbone DP, Liu FT, Galli SJ, Kawakami T. Regulation of mast cell survival by IgE. Immunity 2001;14:791-800.
- 12. Kalesnikoff J, Huber M, Lam V, Damen JE, Zhang J, Siraganian RP, Krystal G. Monomeric IgE stimulates signaling pathways in mast cells that lead to cytokine production and cell survival. Immunity 2001;14:801-11.
- 13. Tanaka S, Takasu Y, Mikura S, Satoh N, Ichikawa A. Antigen-inde-

pendent induction of histamine synthesis by immunoglobulin E in mouse bone marrow-derived mast cells. J Exp Med 2002;196:229-35.

- 14. Kitaura J, Song J, Tsai M, Asai K, Maeda-Yamamoto M, Mocsai A, Kawakami Y, Liu FT, Lowell CA, Barisas BG, Galli SJ, Kawakami T. Evidence that IgE molecules mediate a spectrum of effects on mast cell survival and activation via aggregation of the FcepsilonRI. Proc Natl Acad Sci U S A 2003;100:12911-6.
- 15. Yamada N, Matsushima H, Tagaya Y, Shimada S, Katz SI. Generation of a large number of connective tissue type mast cells by culture of murine fetal skin cells. J Invest Dermatol 2003;121:1425-32.
- Oka T, Hori M, Tanaka A, Matsuda H, Karaki H, Ozaki H. IgE aloneinduced actin assembly modifies calcium signaling and degranulation in RBL-2H3 mast cells. Am J Physiol Cell Physiol 2004;286: C256-63.
- 17. Pandey V, Mihara S, Fensome-Green A, Bolsover S, Cockcroft S. Monomeric IgE stimulates NFAT translocation into the nucleus, a rise in cytosol Ca2+, degranulation, and membrane ruffling in the cultured rat basophilic leukemia-2H3 mast cell line. J Immunol 2004;172:4048-58.
- Lam V, Kalesnikoff J, Lee CW, Hernandez-Hansen V, Wilson BS, Oliver JM, Krystal G. IgE alone stimulates mast cell adhesion to fibronectin via pathways similar to those used by IgE + antigen but distinct from those used by Steel factor. Blood 2003;102:1405-13.
- Kitaura J, Eto K, Kinoshita T, Kawakami Y, Leitges M, Lowell CA, Kawakami T. Regulation of highly cytokinergic IgE-induced mast cell adhesion by Src, Syk, Tec, and protein kinase C family kinases. J Immunol 2005;174:4495-504.
- 20. Kashiwakura J, Kawakami Y, Yuki K, Zajonc DM, Hasegawa S, Tomimori Y, Caplan B, Saito H, Furue M, Oettgen HC, Okayama Y, Kawakami T. Polyclonal IgE induces mast cell survival and cytokine production. Allergol Int 2009;58:411-9.
- 21. MacDonald SM, Rafnar T, Langdon J, Lichtenstein LM. Molecular identification of an IgE-dependent histamine-releasing factor. Science 1995;269:688-90.
- 22. MacDonald SM, Lichtenstein LM. Histamine-releasing factors and heterogeneity of IgE. Springer Semin Immunopathol 1990;12:415-28.
- 23. Warner JA, Pienkowski MM, Plaut M, Norman PS, Lichtenstein LM. Identification of histamine releasing factor(s) in the late phase of cutaneous IgE-mediated reactions. J Immunol 1986;136:2583-7.
- 24. MacDonald SM, Lichtenstein LM, Proud D, Plaut M, Naclerio RM, MacGlashan DW, Kagey-Sobotka A. Studies of IgE-dependent histamine releasing factors: heterogeneity of IgE. J Immunol 1987;139: 506-12.
- 25. MacDonald SM. Histamine releasing factors and IgE heterogeneity. In: Middleton E, Reed CE, Ellis EF, Adkinson NF, Yunginger JW, Busse WW, editors. Allergy: principles and practice. 4th ed. St. Louis, MO: Mosby-Year Book Inc.; 1993.
- 26. Amzallag N, Passer BJ, Allanic D, Segura E, Théry C, Goud B, Amson R, Telerman A. TSAP6 facilitates the secretion of translationally controlled tumor protein/histamine-releasing factor via a nonclassical pathway. J Biol Chem 2004;279:46104-12.
- 27. Schroeder JT, Lichtenstein LM, MacDonald SM. An immunoglobulin E-dependent recombinant histamine-releasing factor induces interleukin-4 secretion from human basophils. J Exp Med 1996;183: 1265-70.
- 28. Schroeder JT, Lichtenstein LM, MacDonald SM. Recombinant histamine-releasing factor enhances IgE-dependent IL-4 and IL-13 secretion by human basophils. J Immunol 1997;159:447-52.

- 29. MacDonald SM. Human recombinant histamine-releasing factor. Int Arch Allergy Immunol 1997;113:187-9.
- 30. Kleine-Tebbe J, Kagey-Sobotka A, MacGlashan DW Jr, Lichtenstein LM, MacDonald SM. Lectins do not distinguish between heterogenous IgE molecules as defined by differential activity of an IgE-dependent histamine releasing factor. J Allergy Clin Immunol 1996; 98:181-8.
- 31. Vonakis BM, Gibbons S Jr, Sora R, Langdon JM, MacDonald SM. Src homology 2 domain-containing inositol 5' phosphatase is negatively associated with histamine release to human recombinant histamine-releasing factor in human basophils. J Allergy Clin Immunol 2001;108:822-31.
- 32. Wantke F, MacGlashan DW, Langdon JM, MacDonald SM. The human recombinant histamine releasing factor: functional evidence that it does not bind to the IgE molecule. J Allergy Clin Immunol 1999;103:642-8.
- Ozeki T, Verma V, Uppalapati M, Suzuki Y, Nakamura M, Catchmark JM, Hancock WO. Surface-bound casein modulates the adsorption and activity of kinesin on SiO2 surfaces. Biophys J 2009;96:3305-18.
- 34. Kim M, Min HJ, Won HY, Park H, Lee JC, Park HW, Chung J, Hwang ES, Lee K. Dimerization of translationally controlled tumor protein is essential for its cytokine-like activity. PLoS One 2009;4:e6464.
- 35. Yeh YC, Xie L, Langdon JM, Myers AC, Oh SY, Zhu Z, Macdonald SM. The effects of overexpression of histamine releasing factor (HRF) in a transgenic mouse model. PLoS One 2010;5:e11077.
- Ravetch JV, Bolland S. IgG Fc receptors. Annu Rev Immunol 2001; 19:275-90.
- Nimmerjahn F, Bruhns P, Horiuchi K, Ravetch JV. FcgammaRIV: a novel FcR with distinct IgG subclass specificity. Immunity 2005;23: 41-51.
- Kang HS, Lee MJ, Song H, Han SH, Kim YM, Im JY, Choi I. Molecular identification of IgE-dependent histamine-releasing factor as a B cell growth factor. J Immunol 2001;166:6545-54.
- Bheekha-Escura R, MacGlashan DW, Langdon JM, MacDonald SM. Human recombinant histamine-releasing factor activates human eosinophils and the eosinophilic cell line, AML14-3D10. Blood 2000;96:2191-8.
- 40. Vonakis BM, Sora R, Langdon JM, Casolaro V, MacDonald SM. Inhibition of cytokine gene transcription by the human recombinant histamine-releasing factor in human T lymphocytes. J Immunol 2003;171:3742-50.
- 41. Yoneda K, Rokutan K, Nakamura Y, Yanagawa H, Kondo-Teshima S, Sone S. Stimulation of human bronchial epithelial cells by IgE-dependent histamine-releasing factor. Am J Physiol Lung Cell Mol Physiol 2004;286:L174-81.
- 42. Silverman GJ, Goodyear CS. Confounding B-cell defences: lessons from a staphylococcal superantigen. Nat Rev Immunol 2006;6:465-75.
- Patella V, Casolaro V, Björck L, Marone G. Protein L. A bacterial Igbinding protein that activates human basophils and mast cells. J Immunol 1990;145:3054-61.
- 44. Patella V, Florio G, Petraroli A, Marone G. HIV-1 gp120 induces IL-4 and IL-13 release from human Fc epsilon RI+ cells through interaction with the VH3 region of IgE. J Immunol 2000;164:589-95.
- 45. Patella V, Giuliano A, Florio G, Bouvet JP, Marone G. Endogenous superallergen protein Fv interacts with the VH3 region of IgE to induce cytokine secretion from human basophils. Int Arch Allergy Immunol 1999;118:197-9.
- 46. Kashiwakura JC, Ando T, Matsumoto K, Kimura M, Kitaura J, Ma-

tho MH, Zajonc DM, Ozeki T, Ra C, MacDonald SM, Siraganian RP, Broide DH, Kawakami Y, Kawakami T. Histamine-releasing factor has a proinflammatory role in mouse models of asthma and allergy. J Clin Invest 2012;122:218-28.

- 47. Kashiwakura J, Okayama Y, Furue M, Kabashima K, Shimada S, Ra C, Siraganian RP, Kawakami Y, Kawakami T. Most highly cytokinergic IgEs have polyreactivity to autoantigens. Allergy Asthma Immunol Res 2012;4:332-40.
- 48. Valenta R, Mittermann I, Werfel T, Garn H, Renz H. Linking allergy to autoimmune disease. Trends Immunol 2009;30:109-16.
- 49. Black P. Why is the prevalence of allergy and autoimmunity increasing? Trends Immunol 2001;22:354-5.
- 50. Rabin RL, Levinson AI. The nexus between atopic disease and autoimmunity: a review of the epidemiological and mechanistic literature. Clin Exp Immunol 2008;153:19-30.
- Valenta R, Seiberler S, Natter S, Mahler V, Mossabeb R, Ring J, Stingl G. Autoallergy: a pathogenetic factor in atopic dermatitis? J Allergy Clin Immunol 2000;105:432-7.
- 52. Aichberger KJ, Mittermann I, Reininger R, Seiberler S, Swoboda I, Spitzauer S, Kopp T, Stingl G, Sperr WR, Valent P, Repa A, Bohle B, Kraft D, Valenta R. Hom s 4, an IgE-reactive autoantigen belonging to a new subfamily of calcium-binding proteins, can induce Th cell type 1-mediated autoreactivity. J Immunol 2005;175:1286-94.
- 53. Mittermann I, Reininger R, Zimmermann M, Gangl K, Reisinger J, Aichberger KJ, Greisenegger EK, Niederberger V, Seipelt J, Bohle B, Kopp T, Akdis CA, Spitzauer S, Valent P, Valenta R. The IgE-reactive autoantigen Hom s 2 induces damage of respiratory epithelial cells and keratinocytes via induction of IFN-gamma. J Invest Dermatol 2008;128:1451-9.
- Crameri R, Faith A, Hemmann S, Jaussi R, Ismail C, Menz G, Blaser K. Humoral and cell-mediated autoimmunity in allergy to Aspergillus fumigatus. J Exp Med 1996;184:265-70.
- 55. Mayer C, Appenzeller U, Seelbach H, Achatz G, Oberkofler H, Breitenbach M, Blaser K, Crameri R. Humoral and cell-mediated autoimmune reactions to human acidic ribosomal P2 protein in individuals sensitized to Aspergillus fumigatus P2 protein. J Exp Med 1999;189:1507-12.
- 56. Schmid-Grendelmeier P, Flückiger S, Disch R, Trautmann A, Wüthrich B, Blaser K, Scheynius A, Crameri R. IgE-mediated and T cellmediated autoimmunity against manganese superoxide dismutase in atopic dermatitis. J Allergy Clin Immunol 2005;115:1068-75.
- 57. Spitzauer S, Schweiger C, Sperr WR, Pandjaitan B, Valent P, Muhl S, Ebner C, Scheiner O, Kraft D, Rumpold H, Valenta R. Molecular characterization of dog albumin as a cross-reactive allergen. J Allergy Clin Immunol 1994;93:614-27.
- 58. Valenta R, Duchêne M, Pettenburger K, Sillaber C, Valent P, Bettelheim P, Breitenbach M, Rumpold H, Kraft D, Scheiner O. Identification of profilin as a novel pollen allergen; IgE autoreactivity in sensitized individuals. Science 1991;253:557-60.
- Zeller S, Rhyner C, Meyer N, Schmid-Grendelmeier P, Akdis CA, Crameri R. Exploring the repertoire of IgE-binding self-antigens associated with atopic eczema. J Allergy Clin Immunol 2009;124: 278-85, 285.e1-7.
- 60. Feng Y, Liu D, Yao H, Wang J. Solution structure and mapping of a very weak calcium-binding site of human translationally controlled tumor protein by NMR. Arch Biochem Biophys 2007;467:48-57.
- 61. Graidist P, Yazawa M, Tonganunt M, Nakatomi A, Lin CC, Chang JY, Phongdara A, Fujise K. Fortilin binds Ca2+ and blocks Ca2+-dependent apoptosis in vivo. Biochem J 2007;408:181-91.

- 62. Kim M, Jung Y, Lee K, Kim C. Identification of the calcium binding sites in translationally controlled tumor protein. Arch Pharm Res 2000;23:633-6.
- 63. Sanchez JC, Schaller D, Ravier F, Golaz O, Jaccoud S, Belet M, Wilkins MR, James R, Deshusses J, Hochstrasser D. Translationally controlled tumor protein: a protein identified in several nontumoral cells including erythrocytes. Electrophoresis 1997;18:150-5.
- 64. Cans C, Passer BJ, Shalak V, Nancy-Portebois V, Crible V, Amzallag N, Allanic D, Tufino R, Argentini M, Moras D, Fiucci G, Goud B, Mirande M, Amson R, Telerman A. Translationally controlled tumor protein acts as a guanine nucleotide dissociation inhibitor on the translation elongation factor eEF1A. Proc Natl Acad Sci U S A 2003; 100:13892-7.
- 65. Fleischer TC, Weaver CM, McAfee KJ, Jennings JL, Link AJ. Systematic identification and functional screens of uncharacterized proteins associated with eukaryotic ribosomal complexes. Genes Dev 2006;20:1294-307.
- 66. Langdon JM, Vonakis BM, MacDonald SM. Identification of the interaction between the human recombinant histamine releasing factor/translationally controlled tumor protein and elongation factor-1 delta (also known as eElongation factor-1B beta). Biochim Biophys Acta 2004;1688:232-6.
- 67. Hsu YC, Chern JJ, Cai Y, Liu M, Choi KW. Drosophila TCTP is essential for growth and proliferation through regulation of dRheb GT-Pase. Nature 2007;445:785-8.
- 68. Zhang D, Li F, Weidner D, Mnjoyan ZH, Fujise K. Physical and

functional interaction between myeloid cell leukemia 1 protein (MCL1) and Fortilin. The potential role of MCL1 as a fortilin chaperone. J Biol Chem 2002;277:37430-8.

- 69. Liu H, Peng HW, Cheng YS, Yuan HS, Yang-Yen HF. Stabilization and enhancement of the antiapoptotic activity of mcl-1 by TCTP. Mol Cell Biol 2005;25:3117-26.
- 70. Yang Y, Yang F, Xiong Z, Yan Y, Wang X, Nishino M, Mirkovic D, Nguyen J, Wang H, Yang XF. An N-terminal region of translationally controlled tumor protein is required for its antiapoptotic activity. Oncogene 2005;24:4778-88.
- Rho SB, Lee JH, Park MS, Byun HJ, Kang S, Seo SS, Kim JY, Park SY. Anti-apoptotic protein TCTP controls the stability of the tumor suppressor p53. FEBS Lett 2011;585:29-35.
- 72. Amson R, Pece S, Lespagnol A, Vyas R, Mazzarol G, Tosoni D, Colaluca I, Viale G, Rodrigues-Ferreira S, Wynendaele J, Chaloin O, Hoebeke J, Marine JC, Di Fiore PP, Telerman A. Reciprocal repression between P53 and TCTP. Nat Med 2012;18:91-9.
- 73. Rinnerthaler M, Jarolim S, Heeren G, Palle E, Perju S, Klinger H, Bogengruber E, Madeo F, Braun RJ, Breitenbach-Koller L, Breitenbach M, Laun P. MMI1 (YKL056c, TMA19), the yeast orthologue of the translationally controlled tumor protein (TCTP) has apoptotic functions and interacts with both microtubules and mitochondria. Biochim Biophys Acta 2006;1757:631-8.
- 74. Bommer UA. Cellular function and regulation of the translationally controlled tumour protein TCTP. Open Allergy J 2012;5:19-32.