

Homocysteine and vitamins in cardiovascular disease

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On the basis of recent retrospective and prospective studies, it is now widely accepted that increased total plasma homocysteine is a risk factor for cardiovascular disease. Impaired enzyme function as a result of genetic mutation or deficiency of the essential B vitamins folic acid, B₁₂, and B₆ can lead to hyperhomocysteinemia. Oxidized forms of homocysteine account for 98–99% of total plasma homocysteine. Although there is uncertainty as to whether increased homocysteine is causal or merely a proxy for cardiovascular disease, several lines of evidence suggest that it may play a role in atherothrombotic disease. Homocysteine appears to alter the anticoagulant properties of endothelial cells to a procoagulant phenotype. Mildly increased homocysteine causes dysfunction of the vascular endothelium. Folic acid effectively lowers homocysteine concentration in the plasma. Intervention studies are urgently needed to determine if lowering homocysteine is effective in decreasing the morbidity and mortality of cardiovascular disease.

Cardiovascular disease (CVD)¹ is the leading cause of mortality in the United States and in most Western countries. The emergence of new risk factors for heart disease and other major human illnesses are often greeted with skepticism in the scientific community. McCully and Wilson (1) proposed the “homocysteine theory of arteriosclerosis” in 1975 on the basis of pathological examinations of autopsy material from children with homocystinuria (2). However, only within the past 5 years has

homocysteine taken its place among other major risk factors such as cholesterol, smoking, and obesity. The fact that homocysteine is now widely accepted as a major independent risk factor for cardiovascular, cerebrovascular, and peripheral vascular disease is because, at least in part, of methodological advances in the determination of total plasma homocysteine that have occurred in the past decade and to a greater appreciation of the role of micronutrients in health and well-being.

Nearly 80 retrospective and prospective clinical studies have been published on homocysteine and vascular disease in the past 10 years. The vast majority of these studies support the observation that increased total plasma homocysteine is associated with an increased risk of CVD (3–10). However, whether homocysteine is causal in atherogenesis and thrombogenesis, or merely a proxy for the disease itself, remains to be determined. Secondary intervention trials that will effectively lower total plasma homocysteine in patients with CVD followed by the assessment of mortality and morbidity outcomes are only now beginning. It may be several years before the answer is known. In the meantime, it may be prudent to understand the relationship between homocysteine and folic acid, vitamin B₁₂, and vitamin B₆ because, in most individuals, optimal nutrition with respect to these vitamins is an effective means of maintaining lower concentrations of total plasma homocysteine.

The determinants of total plasma homocysteine are complex and involve demographic, genetic, and acquired factors. Acquired factors include both state-of-health and life-style considerations. Although the roles that homocysteine plays in atherogenesis, atherosclerosis, and thrombosis are unknown, several recent studies suggest that hyperhomocysteinemia can disrupt functions of the vascular endothelium that may lead to conversion of its usually anticoagulant surface to one that is procoagulant. Treatment of hyperhomocysteinemia and the design of clinical trials is considered with respect to the current policy to fortify the food supply with folic acid. Is homocysteine the “new cholesterol” of risk factors? Time will tell.

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¹ Nonstandard abbreviations: CVD, cardiovascular disease; CBS, cystathionine β -synthase; MTHFR, methylenetetrahydrofolate reductase; MS, methionine synthase; and Δ -MTHFR, thermolabile methylenetetrahydrofolate reductase.

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Total Plasma Homocysteine and Its Determination

In this review, it is important that the term "total plasma homocysteine" be understood in a biochemical sense, because this will have relevance in the discussions that follow on methodology and on pathophysiological mechanisms. Human plasma contains both reduced and oxidized species of homocysteine (Fig. 1). Chemical definitions were established by Butz and du Vigneaud over 65 years ago (11). The sulfhydryl or reduced form is called homocysteine and the disulfide or oxidized form is called homocystine. Disulfide forms also exist with cysteine and with proteins containing reactive cysteine residues (protein-bound homocysteine). The latter oxidized forms are referred to as mixed disulfides. The oxidized forms of homocysteine usually comprise 98–99% of total plasma homocysteine in human plasma, 80–90% of which is protein-bound (Fig. 1). There is no consensus as yet to designate multiple forms of homocysteine in plasma (12). Some investigators write "homocyst(e)ine and hyperhomocyst(e)inemia" to designate multiple plasma forms. This somewhat awkward and archaic usage should be abandoned. First, it is impossible to pronounce the parentheses in speaking, and we certainly don't use "cholesterol" when we refer to its multiple forms in plasma. No, we say and write "total cholesterol", or identify a specific chemical form by use of appropriate prefixes. Thus, "total homocysteine", "reduced homocysteine", "protein-bound homocysteine", "homocystine", and "homocysteine-cys-

teine mixed disulfide" are easily understood. Homocysteine, like cholesterol, has ever increasing generic meaning, and this should not be discouraged. Total homocysteine, therefore, is the sum total of all forms of homocysteine that exist in plasma or serum.

On the basis of work carried out by many investigators, the range of total homocysteine concentration in plasma from "healthy adults" is 5–15 $\mu\text{mol/L}$ (Table 1) (13). However, one must be cautious in the use of so-called "normal ranges" for homocysteine. We have shown that risk for coronary artery disease is represented by a continuum of total homocysteine concentration, with substantial risk occurring between 10 and 15 $\mu\text{mol/L}$ (14). Perhaps the upper limit of normal will be 10 $\mu\text{mol/L}$ or even lower, because it may be possible to achieve this "desirable level" by optimal nutrition with respect to folic acid, B₁₂, and B₆ (*vide infra*). Individuals with homocystinuria because of rare inborn errors of homocysteine metabolism have severe hyperhomocysteinemia, with total homocysteine concentrations approaching 500 $\mu\text{mol/L}$. It has been reported that up to 20% (~100 $\mu\text{mol/L}$) of the total homocysteine in these individuals is reduced homocysteine (15). Subjects with coronary artery, cerebrovascular, and peripheral vascular disease usually present with mild hyperhomocysteinemia (15–25 $\mu\text{mol/L}$) (3, 6, 8). However, if renal function is impaired, or if the subject has end-stage renal disease, total homocysteine can reach intermediate concentrations (25–50 $\mu\text{mol/L}$) (16, 17).

Methodologies to determine total plasma (or serum) homocysteine were first developed in the mid to late 1980s (18–23). Free homocysteine must first be generated in the plasma sample by chemical reduction of disulfide bonds. Commonly used reductants include 2-mercaptoethanol (19), dithiothreitol (24), sodium borohydride (25), *n*-tributylphosphine (21), and more recently, the water-soluble phosphine tris(2-carboxyethyl)phosphine (26). Homocysteine is then resolved from other low molecular weight thiols (cysteine, cysteinylglycine, and glutathione) by reversed-phase HPLC and determined directly by electrochromophore detection (20), or derivatized with a fluorochromophore, resolved by HPLC, and detected fluorometrically (27–29). Alternatively, after reductive generation, homocysteine can be derivatized for capillary gas chromatography and detected by mass spectrometry (19). Although there is relatively good agreement between different laboratories (13), standardized reference ranges do not exist, and there is no consensus on the use of a standardized calibrator. The differences seen between

Reduced:		
Homocysteine	$\begin{array}{c} \text{NH}_3^+ \\ \\ \text{OOCCHCH}_2\text{CH}_2\text{-SH} \end{array}$	1 %
Oxidized:		
Homocystine	$\begin{array}{c} \text{NH}_3^+ \\ \\ \text{OOCCHCH}_2\text{CH}_2\text{-S} \\ \\ \text{OOCCHCH}_2\text{CH}_2\text{-S} \\ \\ \text{NH}_3^+ \end{array}$	5-10 %
Mixed-disulfides:		
Protein-bound Homocysteine	$\begin{array}{c} \text{NH}_3^+ \\ \\ \text{OOCCHCHCH}_2\text{-S} \\ \quad \quad \quad \\ \text{Protein} \quad \quad \quad \text{S} \end{array}$	80-90 %
Cysteine-Homocysteine	$\begin{array}{c} \text{NH}_3^+ \\ \\ \text{OOCCHCH}_2\text{CH}_2\text{-S} \\ \\ \text{OOCCHCH}_2\text{-S} \\ \\ \text{NH}_3^+ \end{array}$	5-10 %

Fig. 1. Constituents of total plasma homocysteine and percentage of composition.

Table 1. Total plasma homocysteine: working ranges.

Normal range	5–15 $\mu\text{mol/L}$
Desirable (?)	<10 $\mu\text{mol/L}$
Hyperhomocysteinemia	
Mild	15–25 $\mu\text{mol/L}$
Intermediate	25–50 $\mu\text{mol/L}$
Severe	50–500 $\mu\text{mol/L}$

laboratories may be because of efficiency of disulfide bond reduction and use of different internal standards and calibrators. Immunoassays for total homocysteine were recently reported by Shipchandler and Moore (30) and by Frantzen et al. (31). Both of these assays use a mouse monoclonal antibody directed against S-adenosylhomocysteine, which is formed when homocysteine, again generated by reductive cleavage of plasma disulfides, is allowed to react with adenosine in the presence of S-adenosylhomocysteine hydrolase. Advances in the development of high-throughput assays for total homocysteine are likely to continue over the next several years.

Hyperhomocysteinemia and Links to CVD

Classical homocystinuria, a monogenic defect of homocysteine metabolism, was discovered in 1962 by Carson and Neill (32) and Gerritsen et al. (33). This rare disease, occurring in ~1 in every 100 000 to 200 000 births, is caused by deficiencies of cystathionine β -synthase (CBS), methylenetetrahydrofolate reductase (MTHFR), and methionine synthase (MS) and is inherited through an autosomal recessive mechanism. Individuals who are homozygous for CBS deficiency are unable to catabolize

homocysteine through the transsulfuration pathway (Fig. 2). Homozygous deficiency of either MTHFR or MS prevents conversion of homocysteine back to methionine through the methionine cycle (Fig. 2). The clinical hallmarks of homocystinuria include mental retardation, skeletal abnormalities, ectopic lenses, and premature atherosclerotic disease (34). These individuals excrete large amounts of homocysteine in the urine and have total homocysteine concentrations in the plasma from 50 to 500 $\mu\text{mol/L}$. The extremely rapid onset of arteriosclerosis and atherosclerosis seen in these subjects suggests that there may be a dose-time relationship between total plasma homocysteine and the onset and progression of CVD.

The association between mild hyperhomocysteinemia and CVD has been documented in numerous retrospective case-control studies. In 1976, before development of assays for total plasma homocysteine, Wilcken and Wilcken (35) reported higher concentrations of homocysteine-cysteine mixed disulfide in plasma from subjects with coronary artery disease 4 h after methionine loading compared with controls. Kang et al. (36) found that protein-bound homocysteine was higher in subjects with documented coronary artery disease. In an early study

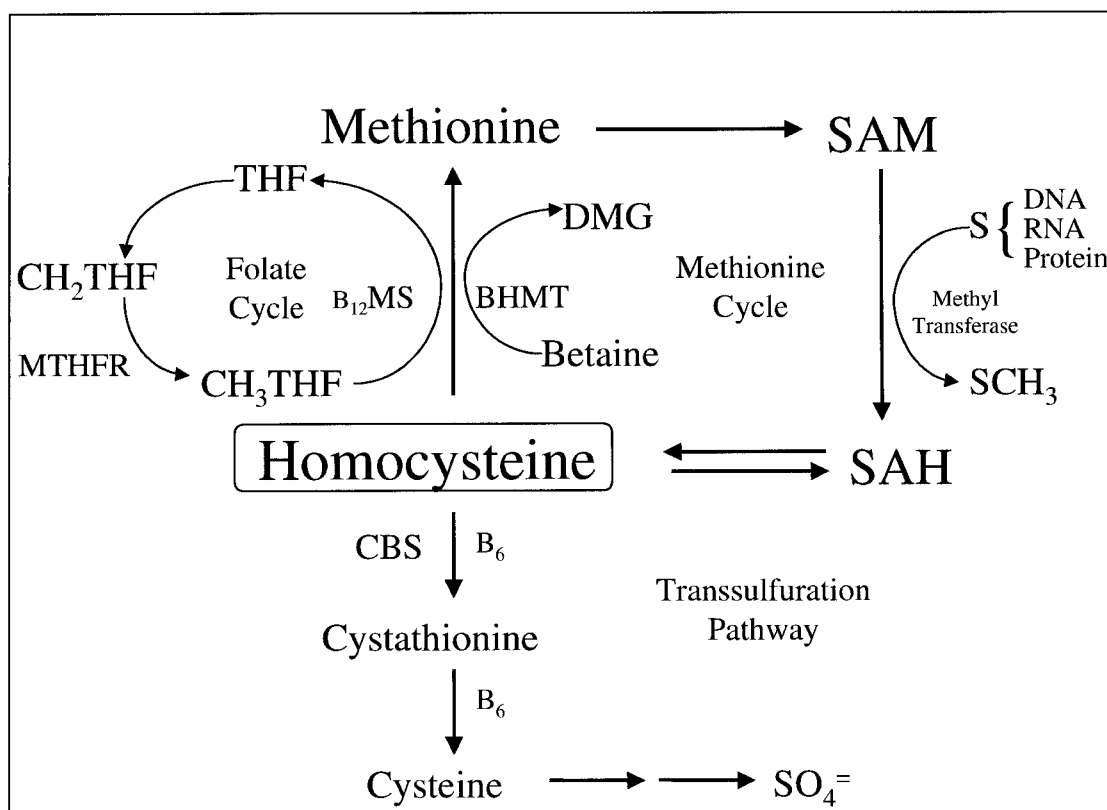


Fig. 2. Metabolism of homocysteine.

Remethylation: homocysteine is remethylated to methionine by B₁₂-dependent MS (B₁₂MS) in the presence of 5-methyltetrahydrofolate (CH₃THF). The latter is the product of 5,10-methylenetetrahydrofolate (CH₂THF) reduction by MTHFR. Homocysteine can also be remethylated by betaine:homocysteine methyltransferase (BHMT) in the liver and kidney. In the methionine cycle, dietary methionine is converted to S-adenosylmethionine (SAM), which serves as a methyl group donor substrate for methyltransferases. The other product of this reaction is S-adenosylhomocysteine (SAH), which is hydrolyzed by SAH hydrolase to homocysteine and adenosine. The methionine and folate cycle enzymes are widely distributed. Homocysteine also enters the catabolic transsulfuration pathway. The first enzyme in this pathway is B₆-dependent CBS. Cystathionine is converted to cysteine by B₆-dependent cystathionase. Cysteine is further catabolized to inorganic sulfate, which is excreted in the urine. The transsulfuration pathway has somewhat limited tissue distribution (liver, kidney, pancreas, and brain).

using basal total plasma homocysteine, Israelsson et al. (37) found an incidence of 24% of hyperhomocysteinemia in men who had suffered a myocardial infarction before the age of 55. Boushey et al. (38) did a metaanalysis on 27 studies (up to early 1995) relating homocysteine to CVD and 11 studies on folate as a determinant of plasma homocysteine. Increased total homocysteine was an independent graded risk factor for CVD, with odds ratios of 1.6 and 1.8 for a 5 $\mu\text{mol/L}$ increment in men and women, respectively. A strong inverse correlation was found between total homocysteine and serum folate. The authors speculated that increasing folate intake from foods, or folic acid by supplementation would substantially reduce annual mortality from coronary artery disease. More recently, Graham et al. (39) reported that increased total homocysteine was as strong as smoking and hyperlipidemia as an independent risk factor for CVD on the basis of a large multicenter European study involving 750 cases and 800 controls.

Results of prospective studies attempting to link hyperhomocysteinemia to CVD have been mixed. In the US Physicians' Health Study, Stampfer et al. (40) found higher total plasma homocysteine in cases compared with controls (11.1 ± 4.0 vs 10.5 ± 2.8 $\mu\text{mol/L}$, $P = 0.03$); however, the relative risk for myocardial infarction was 3.1 for men in the ≥ 95 th percentile, suggesting a threshold effect. Arnesen et al. (41), on the other hand, found that total plasma homocysteine was an independent risk factor for coronary artery disease, with no threshold concentration. Recently, total plasma homocysteine was found to be a strong predictor of mortality in Norwegian patients with coronary artery disease (42). However, prospective studies conducted by Alftan et al. (43) and by Evans et al. (44) were negative.

Secondary intervention studies are only just beginning in the United States and Europe. These studies, designed to lower total plasma homocysteine in patients with CVD by supplementation with combinations of folic acid, B_{12} , and B_6 , will assess long-term mortality and morbidity outcomes in treatment and placebo groups. It may be several years before the results of these clinical trials are known. A complicating factor for trials in this country is the addition of folic acid, a major nutritional determinant of total homocysteine, to the nation's food supply, as discussed below.

Determinants of Hyperhomocysteinemia

The determinants of total plasma homocysteine are complex and involve demographic, genetic, and acquired factors (Table 2). It is likely that gene-nutrient interactions will be important determinants in subjects who carry one or more mutations in genes that regulate homocysteine metabolism. Thus, genetic background, nutrition, state-of-health, life-style, gender, and age influence the homeostasis of homocysteine.

Table 2. Determinants of hyperhomocysteinemia.

Demographic
Age
Sex
Ethnic origin
Genetic
MTHFR
MS
CBS
Acquired
B-vitamin deficiency (folate, B_{12} , and B_6)
State of health
Impaired renal function
End-stage renal disease
Heart and other organ transplants
Hypothyroidism
Lifestyle
Smoking
Alcohol (excessive)
Lack of exercise
Coffee (excessive)

DEMOGRAPHIC FACTORS

Total plasma homocysteine appears to increase throughout life. In Norwegian children, ages 8–12 years, total homocysteine concentrations were 5–6 $\mu\text{mol/L}$, with no reported differences for gender (45). Nygård et al. (46) found that in Norwegian men and women 40–42 years old, geometric mean values for total homocysteine were 10.8 and 9.1 $\mu\text{mol/L}$, respectively. In Norwegians 65–67 years old, the values increased to 12.3 and 11.0 $\mu\text{mol/L}$. In a predominantly female group of French centenarians, mean total plasma homocysteine was 26.7 ± 9.6 $\mu\text{mol/L}$ (47). It is now clear that men have higher total plasma homocysteine concentrations than women of the same age (46); however, the reasons for this difference and when the divergence begins (presumably after puberty) are unknown. South African black children (7–15 years old) had significantly higher homocysteine concentrations than South African white children (5.8 ± 1.8 vs 5.1 ± 0.8 $\mu\text{mol/L}$, respectively); however, Ubbink et al. (48) found that black men had significantly lower concentrations than white men (9.7 ± 3.4 vs 12.0 ± 6.7 $\mu\text{mol/L}$, respectively) and that young black men were able to metabolize homocysteine more efficiently than young white men, based on methionine-loading studies (49). Comparable studies have not been reported in this country.

GENETIC FACTORS

Studies in the early 1990s established that genetic factors were important contributors to the hyperhomocysteinemia seen in patients with coronary artery disease (50–53). Recent evidence suggests that genetic factors, in combination with acquired or environmental factors, lead to increased total plasma homocysteine (46, 54).

MTHFR converts 5,10-methylenetetrahydrofolate to

5-methyltetrahydrofolate in the presence of NADPH (Fig. 2). 5-Methyltetrahydrofolate in turn serves as a substrate for B₁₂-dependent methionine synthase. MTHFR is a flavin-dependent enzyme consisting of two identical 77-kDa subunits with N-terminal catalytic domains and C-terminal regulatory domains (55). Approximately 50 cases of severe MTHFR deficiency have been reported (56), with neurological or vascular complications appearing in the first or second decade of life. The gene for the enzyme has been cloned (57), and 14 mutations causing severe deficiency have been described (58, 59). A thermolabile variant of the enzyme, designated Δ -MTHFR in this review, was first reported by Kang et al. (60). Subsequent studies by the same group found a high incidence of Δ -MTHFR in patients with coronary artery disease (61, 62). In a more recent study by Engbersen et al. (63), 28% of hyperhomocysteinemic patients with premature vascular disease had the thermolabile enzyme. A C677T gene mutation, which produces an alanine-to-valine substitution, is the cause of Δ -MTHFR. The allele frequency of this mutation is widespread: 38% in the French Canadian population (64) and 25–39% in other populations (65–72), but only 10% in African Americans (73). Many reports support the hypothesis that individuals who are homozygous for Δ -MTHFR may be at greater risk for CVD (65–68, 71), but others do not (69, 72), irrespective of folate status (70, 74). Although Ma et al. (75) found no increased risk for myocardial infarction in men because of Δ -MTHFR, they did find higher total homocysteine concentrations in homozygous men with low folate status. A similar gene-nutrient interaction was found by Jacques et al. (76) who studied 365 individuals from the NHLBI Family Heart Study. In patients with end-stage renal disease, Bostom et al. (77) found 33% higher total homocysteine in pooled homozygote and heterozygote patients who had plasma folate values below the median when compared with patients with wild-type MTHFR genotype.

Vitamin B₁₂-dependent MS is found in most cells and tissues (78). Inborn errors that affect B₁₂ absorption, systemic transport, intracellular transport, and conversion to the coenzyme form (methyl-B₁₂) will produce loss of MS activity and hyperhomocysteinemia (56, 79). Mutations in the structural gene for MS itself are only now being described (80–82). Human MS cDNA contains an open reading frame of 3798 nucleotides encoding a polypeptide of 1265 amino acids (predicted molecular mass, 140 kDa). On the basis of complementation studies using patient fibroblast cell lines, two types of MS-associated genetic diseases, *cblE* and *cblG*, have been described (83). The *cblE* class may involve a reducing component of the MS system (84), whereas *cblG* is thought to be caused by defective MS apoenzyme (85, 86). Several MS mutations have now been identified in *cblG* patients (81, 87); however, the prevalence of these mutations in the general population and their contribution to hyperhomo-

cysteinemia in heterozygous individuals remain to be determined.

Entry of homocysteine into the transsulfuration pathway is catalyzed by B₆-dependent CBS, which has limited tissue distribution (78). Several hundred cases of CBS deficiency, the most common form of homocystinuria, have been described (34). The human cDNA for CBS, cloned by Kraus et al. (88) in 1993, contains 2554 nucleotides encoding a subunit of 551 amino acids with a predicted molecular mass of 63 kDa. When bacterial (89, 90) and yeast (91, 92) expression systems have been used, close to 40 mutations have been identified in the CBS gene. Obligate heterozygotes for CBS deficiency might be at greater risk for CVD because of possible increases in basal and postmethionine load total homocysteine (35, 51, 93). However, the reliability of assessing heterozygosity based on CBS enzyme activity measurements and postmethionine-loading studies has been questioned (94). A number of genotyping studies have been done; however, they have failed to identify CBS polymorphisms that correlate with CVD (68, 95, 96). Clearly, additional studies are needed to assess the role of CBS polymorphism in atherosclerotic disease.

ACQUIRED AND LIFE-STYLE FACTORS

The source of homocysteine in the diet is L-methionine. Fruits and vegetables generally contain 0.9–1.2 g of methionine per 100-g serving. Peaches and grapes (3.6 g/100 g) are exceptions. Nuts and cereal grains contain 1.4–1.8 g/100 g, except Brazil nuts, which contain 5.8 g/100 g. Sources of animal protein have a higher methionine content: meat and fish, 2.7 g/100 g, eggs, 3.2 g/100 g, and cow's milk, 2.9 g/100 g. By comparison, human milk has only 1.4 g/100 g.

Dietary insufficiency or malabsorption of folate, vitamin B₁₂, or vitamin B₆ will lead to hyperhomocysteinemia and an increased risk of CVD (97–103). Even in well-nourished generally healthy populations, serum folate and B₁₂ inversely correlate with total homocysteine (29, 104). The B vitamins drive homocysteine metabolism, with 5-methyltetrahydrofolate serving as substrate for B₁₂-dependent MS in the remethylation of homocysteine back to methionine and vitamin B₆ (as pyridoxal-5-phosphate) as a cofactor for CBS in the transsulfuration pathway (Fig. 2). The large Hordaland study in Norway found that increased total plasma homocysteine was associated with smoking, high blood pressure, increased cholesterol, and sedentary life-style (46). Alcoholics have higher total homocysteine concentrations, perhaps because of malnourishment and malabsorption (105, 106). Heavy coffee consumption is associated with higher homocysteine concentrations as well (54).

Certain disease states produce higher total homocysteine concentrations. Patients with end-stage renal disease have intermediate hyperhomocysteinemia and an increased risk for vascular disease (16, 17). Heart transplant recipients have mild to intermediate hyperhomocysteine-

mia (107,108), which may in part be related to renal insufficiency (109). Hypothyroidism produces increased total plasma homocysteine (110), but treatment with L-thyroxine will normalize homocysteine concentrations (111).

Mechanisms of Blood Vessel Injury

It is usually assumed that reduced homocysteine is the atherogenic form of homocysteine in circulation. But recall that it contributes only 1% of the basal total plasma homocysteine in normocysteinemic and mildly hyperhomocysteinemic individuals (Fig. 1). However, in addition to basal concentrations, we should also consider the transient hyperhomocysteinemia that occurs after eating. Within 2 h after methionine loading, reduced homocysteine reaches a maximum and then declines as oxidized forms (homocystine and mixed disulfides) become increased and reach peak values in 6 h (112). The transient hyperhomocysteinemia seen after methionine loading also occurs after meals, and its magnitude is proportional to protein consumption (113). Abnormal methionine loading in which peak total plasma homocysteine concentrations exceed an overall reference range have been seen in up to 40% of subjects, predominantly female (37, 39, 114). Silberberg et al. (115), however, recommend that age- and gender-specific reference ranges be used to prevent overdiagnosis.

It can be hypothesized that reduced homocysteine directly alters vascular cell function. Because reduced homocysteine undergoes oxidation in vivo, one could argue that the homocysteine oxidation products such as hydrogen peroxide, superoxide anion radical, and other reactive oxygen species are the injurious agents. Thus, a second hypothesis is that homocysteine is acting indirectly through its oxidation and formation of reactive oxygen species.

Many of the in vitro studies using cultured endothelial and smooth muscle cells from human and animal vessels during the past 10 years used exceedingly high concentrations of reduced homocysteine (5–10 mmol/L), far exceeding physiological and pathophysiological concentrations of reduced homocysteine (and for that matter total plasma homocysteine). Another problem is that many of these studies failed to demonstrate specificity for homocysteine and in many cases found a “general thiol effect”. Keep in mind that total plasma cysteine is 20- to 30-fold higher than total plasma homocysteine and that the concentration of reduced cysteine is ~70-fold higher than reduced homocysteine (5.0 ± 3.6 vs 0.07 ± 0.02 $\mu\text{mol/L}$) (112). There is little or no evidence in the literature that cysteine is atherogenic. Nevertheless, it is widely believed that homocysteine can alter the surface properties of endothelial cells by changing their phenotype from anticoagulant to procoagulant (116).

Wang et al. (117) recently reported that 10–50 $\mu\text{mol/L}$ D,L-homocysteine (but not L-cysteine or L-cystine) in the presence of 50 $\mu\text{mol/L}$ adenosine inhibited the growth of

vascular endothelial cells by a mechanism involving decreased carboxymethylation of p21^{ras}. This is one of the few reports in which physiologically relevant concentrations of homocysteine have been used and in which thiol specificity has been demonstrated. Upchurch and co-workers reported that homocysteine modulated the expression of glutathione peroxidase (118) and nitric oxide synthase (119) in bovine aortic endothelial cells; however, relatively high concentrations of homocysteine were used. Tsai and co-workers (120,121) reported that homocysteine was mitogenic to smooth muscle cells by a mechanism involving synergistic induction of cyclin A mRNA expression with serum. However, other thiols, such as cysteine and glutathione, could replace homocysteine, suggesting a general thiol effect on the stimulation of smooth muscle cell proliferation. Matrix proteins accumulate in atherosclerotic plaques, and recently Majors et al. (122) reported that relatively low concentrations of homocysteine (50–300 $\mu\text{mol/L}$) stimulate collagen production in cultured rabbit aortic smooth muscle cells.

There have been some interesting in vivo studies in both humans and animal models that support the hypothesis of impaired endothelial cell function in the presence of hyperhomocysteinemia. Van den Berg et al. (123) assessed endothelial dysfunction in young patients with peripheral arterial occlusive disease and mild hyperhomocysteinemia by measuring plasma von Willebrand factor, thrombomodulin, and tissue plasminogen activator. Plasma concentrations of the first two were above reference values at baseline but decreased after treatment of the hyperhomocysteinemia with pyridoxine and folic acid. Tawakol et al. (124) observed impaired endothelial-dependent vasodilation in subjects with mild hyperhomocysteinemia. Linear regression analysis revealed that total plasma homocysteine was the only meaningful predictor of flow-mediated vasodilation. No attempt to correct impaired vasodilation with cofactor therapy was reported. Similar results were obtained by Woo et al. (125) in subjects with intermediate hyperhomocysteinemia.

Using an animal model, Lentz et al. (126) recently tested the hypothesis that diet-induced mild hyperhomocysteinemia would lead to vascular dysfunction. In cynomolgus monkeys on a diet with high methionine and low folate for 4 weeks, total plasma homocysteine was 10.6 ± 2.6 $\mu\text{mol/L}$. Monkeys on a regular diet had total plasma homocysteine concentrations of 4.0 ± 0.2 $\mu\text{mol/L}$. Impaired endothelial-mediated vasodilation was observed in the hyperhomocysteinemic animals. However, in atherosclerotic animals maintained on an atherosclerotic diet for 17 months, normalization of their moderate hyperhomocysteinemia by administration of folic acid, B₁₂, and B₆ was insufficient to restore normal vascular function (127).

Work in our laboratory has focused on the metabolism of homocysteine in vascular cells and tissues. Using direct enzyme assays and Western and Northern blotting, we found that cultured human aortic endothelial cells do not express an active form of CBS (128) and thus apparently

cannot initiate homocysteine catabolism through the transsulfuration pathway (Fig. 2). Their inability to express a major pathway for homocysteine could render them highly susceptible to mild increases of total homocysteine seen in the hyperhomocysteinemia of CVD. Our laboratory has also investigated the role of homocysteine in possible early events in atherogenesis. We found that extremely low concentrations of reduced homocysteine (10–50 $\mu\text{mol/L}$) up-regulated the expression of the chemokine monocyte chemoattractant protein 1 in cultured human aortic endothelial cells and that the up-regulation was specific for homocysteine (129). At maximal dose-response for homocysteine (50 $\mu\text{mol/L}$), cysteine, cystine, methionine, and homocystine had no effect.

Treatment of Hyperhomocysteinemia

The severe hyperhomocysteinemia seen in subjects with classical homocystinuria often responds to treatment. Approximately 50% of homocystinurics with CBS deficiency respond to pyridoxine (vitamin B₆) (34). In patients with congenital deficiency of transcobalamin, the serum protein that delivers B₁₂ to cells throughout the body, weekly injections of vitamin B₁₂ will normalize their hyperhomocysteinemia and allow them to lead relatively normal lives. Deficiencies of MTHFR and MS are usually more refractory to treatment (56, 130). Nevertheless, many of these homozygous monogenic diseases respond quite well to treatment. Can the mild hyperhomocysteinemia associated with vascular disease be treated? The answer is emphatically yes.

The treatment of mild hyperhomocysteinemia is based on the hypothesis that the nutritional status of the micro-nutrients that drive homocysteine metabolism, folate, B₁₂, and B₆, is suboptimal and that by dietary supplementation with folic acid, cyanocobalamin (vitamin B₁₂), and pyridoxine hydrochloride (vitamin B₆), either singly or in combination, one can optimize homocysteine metabolism and lower not only basal total plasma homocysteine, but abnormal methionine-loading hyperhomocysteinemia as well. This hypothesis has been confirmed by many investigators. In 1988, Brattström et al. (131) showed that 5 mg of folic acid per day for 14 days was very effective in lowering total homocysteine in healthy subjects, whereas 40 mg of pyridoxine HCl or 1 mg of cyanocobalamin had little or no effect when overall effects of the three treatment groups were compared. Brattström et al. (132) later reported that administration of 10 mg of folic acid plus 240 mg of pyridoxine HCl per day for 4 weeks reduced mean total plasma homocysteine by 53% and abnormal methionine loading by 39% in 20 patients with premature cerebral and peripheral occlusive disease. The same group showed that patients who had suffered a myocardial infarction were able to lower their total homocysteine concentrations with 2.5 mg of folic acid daily for 6 weeks (133).

Folic acid is clearly the most effective form of supplementation to reduce total homocysteine concentrations in

subjects with mild hyperhomocysteinemia. But is it safe for all individuals and are lower doses effective as well? Before 1986, the recommended daily allowance for folic acid was 400 μg per day. The recommended daily allowance was then changed to 180 μg per day for women and 200 μg per day for men. Women who have folate deficiency at the time of conception are at much greater risk for giving birth to children with neural tube defects. For this reason, the Food and Drug Administration in this country has approved the addition of folic acid to cereals and flour products (134). Individuals will now be consuming on average an additional 100 μg of folic acid a day. Will this have an impact on total plasma homocysteine? Recent studies from John Scott's laboratory in Dublin suggest that it will. Ward et al. (135) showed that even 100 μg of folic acid daily for 6 weeks can lower total homocysteine concentrations in healthy individuals, but that 200 μg per day was near optimal. These doses are clearly safe for most individuals. However, the elderly with undiagnosed B₁₂ deficiency could experience adverse effects by folic acid supplementation. Stabler et al. (136) have recently reviewed the problem that could affect millions of individuals in this country. To protect against unrecognized B₁₂ deficiency, cyanocobalamin could be combined with folic acid supplementation. Approximately 1–2% of an oral dose of cyanocobalamin is absorbed by passive diffusion, even in subjects with pernicious anemia. Bostom et al. (137) found that patients with renal failure require much more aggressive B-vitamin therapy to achieve total plasma homocysteine lowering.

Studies on classical homocystinuria provide strong evidence that homocysteine is causal; however, the role that homocysteine plays in the usually mild hyperhomocysteinemia associated with coronary artery disease, cerebrovascular disease, and peripheral vascular disease is still unclear. Clinical trials are urgently needed to determine if lowering total plasma homocysteine will reduce the mortality and morbidity of CVD.

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References

1. McCully KS, Wilson RB. Homocysteine theory of arteriosclerosis. *Atherosclerosis* 1975;22:215–27.
2. McCully KS. Vascular pathology of homocysteinemia: implications for the pathogenesis of arteriosclerosis. *Am J Pathol* 1969;56:111–28.
3. Ueland PM, Refsum H, Brattström L. Plasma homocysteine and cardiovascular disease. In: Francis RB, ed. *Atherosclerotic cardiovascular disease, hemostasis, and endothelial function*. New York: Marcel Dekker, Inc., 1992:183–236.

4. Kang S-S, Wong PWK, Malinow MR. Hyperhomocyst(e)inemia as a risk factor for occlusive vascular disease. *Annu Rev Nutr* 1992;12:279–98.
5. Masser PA, Taylor LM Jr, Porter JM. Importance of elevated plasma homocysteine levels as a risk factor for atherosclerosis. *Ann Thorac Surg* 1995;58:1240–6.
6. Green R, Jacobsen DW. Clinical implications of hyperhomocysteinemia. In: Bailey LB, ed. *Folate in health and disease*. New York: Marcel Dekker, Inc., 1995:75–122.
7. Verhoef P, Stampfer MJ. Prospective studies of homocysteine and cardiovascular disease. *Nutr Rev* 1995;53:283–8.
8. Mayer EL, Jacobsen DW, Robinson K. Homocysteine and coronary atherosclerosis. *J Am Coll Cardiol* 1996;27:517–27.
9. Guba SC, Fink LM, Fonseca V. Hyperhomocysteinemia—an emerging and important risk factor for thromboembolic and cardiovascular disease. *Am J Clin Pathol* 1996;106:709–22.
10. Miner SES, Evrosvki J, Cole DEC. Clinical chemistry and molecular biology of homocysteine metabolism: an update. *Clin Biochem* 1997;30:189–201.
11. Butz LW, du Vigneaud V. The formation of a homologue of cystine by the decomposition of methionine with sulfuric acid. *J Biol Chem* 1932;99:135–42.
12. Mudd SH, Levy HL. Plasma homocyst(e)ine or homocysteine? [Letter]. *N Engl J Med* 1995;333:325.
13. Ueland PM, Refsum H, Stabler SP, Malinow MR, Andersson A, Allen RH. Total homocysteine in plasma or serum: methods and clinical applications. *Clin Chem* 1993;39:1764–79.
14. Robinson K, Mayer EL, Miller DP, Green R, van Lente F, Gupta A, et al. Hyperhomocysteinemia and low pyridoxal phosphate. Common and independent reversible risk factors for coronary artery disease. *Circulation* 1995;92:2825–30.
15. Mansoor MA, Ueland PM, Aarsland A, Svoldal AM. Redox status and protein binding of plasma homocysteine and other aminothiols in patients with homocystinuria. *Metabolism* 1993;42:1481–5.
16. Dennis VW, Robinson K. Homocysteinemia and vascular disease in end-stage renal disease. *Kidney Int* 1996;50(Suppl 57):S11–7.
17. Bostom AG, Lathrop L. Hyperhomocysteinemia in end-stage renal disease: prevalence, etiology, and potential relationship to arteriosclerotic outcomes. *Kidney Int* 1997;52:10–20.
18. Refsum H, Helland S, Ueland PM. Radioenzymic determination of homocysteine in plasma and urine. *Clin Chem* 1985;31:624–8.
19. Stabler SP, Marcell PD, Podell ER, Allen RH. Quantitation of total homocysteine, total cysteine, and methionine in normal serum and urine using capillary gas chromatography-mass spectrometry. *Anal Biochem* 1987;162:185–96.
20. Malinow MR, Kang S-S, Taylor LM, Wong PWK, Coull B, Inahara T, et al. Prevalence of hyperhomocyst(e)inemia in patients with peripheral arterial occlusive disease. *Circulation* 1989;79:1180–8.
21. Araki A, Sako Y. Determination of free and total homocysteine in human plasma by high-performance liquid chromatography with fluorescence detection. *J Chromatogr* 1987;422:43–52.
22. Jacobsen DW, Gatautis VJ, Green R. Determination of plasma homocysteine by high-performance liquid chromatography with fluorescence detection. *Anal Biochem* 1989;178:208–14.
23. Refsum H, Ueland PM, Svoldal AM. Fully automated fluorescence assay for determining total homocysteine in plasma. *Clin Chem* 1989;35:1921–7.
24. Andersson A, Isaksson A, Brattström L, Hultberg B. Homocysteine and other thiols determined in plasma by HPLC and thiol-specific postcolumn derivatization. *Clin Chem* 1993;39:1590–7.
25. Smolin LA, Sneider JA. Measurement of total plasma cysteamine using high-performance liquid chromatography with electrochemical detection. *Anal Biochem* 1988;168:374–9.
26. Gilfix BM, Blank DW, Rosenblatt DS. Novel reductant for determination of total plasma homocysteine. *Clin Chem* 1997;43:687–8.
27. Vester B, Rasmussen K. High performance liquid chromatography method for rapid and accurate determination of homocysteine in plasma and serum. *Eur J Clin Chem Clin Biochem* 1991;29:549–54.
28. Fiskerstrand T, Refsum H, Kvalheim G, Ueland PM. Homocysteine and other thiols in plasma and urine: automated determination and sample stability. *Clin Chem* 1993;39:263–71.
29. Jacobsen DW, Gatautis VJ, Green R, Robinson K, Savon SR, Secic M, et al. Rapid HPLC determination of total homocysteine and other thiols in serum and plasma: sex differences and correlation with cobalamin and folate levels in normal subjects. *Clin Chem* 1994;40:873–81.
30. Shipchandler MT, Moore EG. Rapid, fully automated measurement of plasma homocyst(e)ine with the Abbott IMx analyzer. *Clin Chem* 1995;41:991–4.
31. Frantzen F, Faaren AL, Alfheim I, Nordhei AK. Enzyme conversion immunoassay for determining total homocysteine in plasma or serum. *Clin Chem* 1998;44:311–6.
32. Carson NAJ, Neill DW. Metabolic abnormalities detected in a survey of mentally backward individuals in Northern Ireland. *Arch Dis Child* 1962;37:505–13.
33. Gerritsen T, Vaughn JG, Waisman HA. The identification of homocystine in the urine. *Biochem Biophys Res Commun* 1962;9:493–6.
34. Mudd SH, Levy HL, Skovby F. Disorders of transsulfuration. In: Scriver CR, Beaudet AL, Sly WS, Valle D, eds. *The metabolic and molecular basis of inherited disease*, 7th ed. New York: McGraw-Hill, 1995:1279–327.
35. Wilcken DEL, Wilcken B. The pathogenesis of coronary artery disease: a possible role for methionine metabolism. *J Clin Invest* 1976;57:1079–82.
36. Kang S-S, Wong PWK, Cook HY, Norusis M, Messer JV. Protein-bound homocysteine: a possible risk factor for coronary artery disease. *J Clin Invest* 1986;77:1482–6.
37. Israelsson B, Brattström L, Hultberg BL. Homocysteine and myocardial infarction. *Atherosclerosis* 1988;71:227–33.
38. Boushey CJ, Beresford SAA, Omenn GS, Motulsky AG. A quantitative assessment of plasma homocysteine as a risk factor for vascular disease—probable benefits of increasing folic acid intakes. *JAMA* 1995;274:1049–57.
39. Graham IM, Daly LE, Refsum HM, Robinson K, Brattström L, Ueland PM, et al. Plasma homocysteine as a risk factor for vascular disease—the European concerted action project. *JAMA* 1997;277:1775–81.
40. Stampfer MJ, Malinow MR, Willet WC, Newcomer LM, Upson B, Ullmann D, et al. A prospective study of plasma homocyst(e)ine and risk of myocardial infarction in US physicians. *JAMA* 1992;268:877–81.
41. Arnesen E, Refsum H, Bonna KH, Ueland PM, Forde OH, Nordrehaug JE. Serum total homocysteine and coronary heart disease. *Int J Epidemiol* 1995;24:704–9.
42. Nygård O, Nordrehaug JE, Refsum H, Ueland PM, Farstad M, Vollset SE. Plasma homocysteine levels and mortality in patients with coronary artery disease. *N Engl J Med* 1997;337:230–6.
43. Alfthan G, Pekkanen J, Jauhiainen M, Pitkaniemi J, Karvonen M, Tuomilehto J, et al. Relation of serum homocysteine and lipoprotein(a) concentrations to atherosclerotic disease in a prospective Finnish population based study. *Atherosclerosis* 1994;106:9–19.
44. Evans RW, Shaten BJ, Hempel JD, Cutler JA, Kuller LH. Homo-

- cyst(e)ine, risk of cardiovascular disease in the Multiple Risk Factor Intervention Trial. *Arterioscler Thromb Vasc Biol* 1997;17:1947–53.
45. Tonstad S, Refsum H, Sivertsen M, Christophersen B, Ose L, Ueland PM. Relation of total homocysteine and lipid levels in children to premature cardiovascular death in male relatives. *Pediatr Res* 1996;40:47–52.
 46. Nygård O, Vollset SE, Refsum H, Stensvold I, Tverdal A, Nordrehaug JE, et al. Total plasma homocysteine and cardiovascular risk profile—the Hordaland homocysteine study. *JAMA* 1995;274:1526–33.
 47. Faure-Delanef L, Quere I, Chasse JF, Guerassimenko O, Lesaulnier M, Bellet H, et al. Methylene tetrahydrofolate reductase thermolabile variant and human longevity. *Am J Hum Genet* 1997;60:999–1001.
 48. Ubbink JB, Delport R, Vermaak WJH. Plasma homocysteine concentrations in a population with a low coronary heart disease prevalence. *J Nutr* 1996;126(Suppl):1254S–7S.
 49. Ubbink JB, Vermaak WJH, Delport R, van der Merwe A, Becker PJ, Potgieter H. Effective homocysteine metabolism may protect South African blacks against coronary heart disease. *Am J Clin Nutr* 1995;62:802–8.
 50. Williams RR, Malinow MR, Hunt SC, Upson B, Wu LL, Hopkins PN, et al. Hyperhomocyst(e)inemia in Utah siblings with early coronary artery disease. *Coron Artery Dis* 1990;1:681–5.
 51. Clarke R, Daly L, Robinson K, Naughten E, Cahalane S, Fowler B, et al. Hyperhomocysteinemia: an independent risk factor for vascular disease. *N Engl J Med* 1991;324:1149–55.
 52. Genest JJ, McNamara JR, Upson B, Salem DN, Ordovas JM, Schaefer EJ, et al. Prevalence of familial hyperhomocyst(e)inemia in men with premature coronary artery disease. *Arterioscler Thromb* 1991;11:1129–36.
 53. Wu LL, Wu J, Hunt SC, James BC, Vincent GM, Williams RR, et al. Plasma homocyst(e)ine as a risk factor for early familial coronary artery disease. *Clin Chem* 1994;40:552–61.
 54. Nygård O, Refsum H, Ueland PM, Stensvold I, Nordrehaug JE, Kvåle G, et al. Coffee consumption and plasma total homocysteine: the Hordaland homocysteine study. *Am J Clin Nutr* 1997;65:136–43.
 55. Matthews RG, Vanoni MA, Hainfeld JF, Wall J. Methylene tetrahydrofolate reductase. Evidence for spatially distinct subunit domains obtained by scanning transmission electron microscopy and limited proteolysis. *J Biol Chem* 1984;259:11647–50.
 56. Rosenblatt DS. Inherited disorders of folate transport and metabolism. In: Scriver CR, Beaudet AL, Sly WS, Valle D, eds. *The metabolic and molecular basis of inherited disease*, 7th ed. New York: McGraw-Hill, 1995:3111–49.
 57. Goyette P, Sumner JS, Milos R, Duncan AMV, Rosenblatt DS, Matthews RG, et al. Human methylene tetrahydrofolate reductase: isolation of cDNA, mapping and mutation identification. *Nat Genet* 1994;7:195–200.
 58. Goyette P, Frosst P, Rosenblatt DS, Rozen R. Seven novel mutations in the methylene tetrahydrofolate reductase gene and genotype/phenotype correlations in severe methylene tetrahydrofolate reductase deficiency. *Am J Hum Genet* 1995;56:1052–9.
 59. Goyette P, Christensen B, Rosenblatt DS, Rozen R. Severe and mild mutations in cis for the methylene tetrahydrofolate reductase (MTHFR) gene, and description of five novel mutations in MTHFR. *Am J Hum Genet* 1996;59:1268–75.
 60. Kang S-S, Zhou J, Wong PWK, Kowalisyn J, Strokosch G. Intermediate homocysteinemia: a thermolabile variant of methylene tetrahydrofolate reductase. *Am J Hum Genet* 1988;43:414–21.
 61. Kang S-S, Wong PWK, Zhou J, Sora J, Lessick M, Ruggie N, et al. Thermolabile methylene tetrahydrofolate reductase in patients with coronary artery disease. *Metabolism* 1988;37:611–3.
 62. Kang S-S, Wong PWK, Susmano A, Sora J, Norusis M, Ruggie N. Thermolabile methyl tetrahydrofolate reductase: an inherited risk factor for coronary artery disease. *Am J Hum Genet* 1991;48:536–45.
 63. Engbersen AMT, Franken DG, Boers GHJ, Stevens EMB, Trijbels FJM, Blom HJ. Thermolabile 5,10-methylene tetrahydrofolate reductase as a cause of mild hyperhomocysteinemia. *Am J Hum Genet* 1995;56:142–50.
 64. Frosst P, Blom HJ, Milos R, Goyette P, Sheppard CA, Matthews RG, et al. A candidate genetic risk factor for vascular disease: a common mutation in methylene tetrahydrofolate reductase. *Nat Genet* 1995;10:111–3.
 65. Izumi M, Iwai N, Ohmichi N, Nakamura Y, Shimoike H, Kinoshita M. Molecular variant of 5,10-methylene tetrahydrofolate reductase risk factor of ischemic heart disease in the Japanese population. *Atherosclerosis* 1996;121:293–4.
 66. De Franchis R, Mancini FP, D'Angelo A, Sebastio G, Fermo I, De Stefano V, et al. Elevated total plasma homocysteine and 677C→T mutation of the 5,10-methylene tetrahydrofolate reductase gene in thrombotic vascular disease. *Am J Hum Genet* 1996;59:262–4.
 67. Harmon DL, Woodside JV, Yarnell JWG, McMaster D, Young IS, McCrum EE, et al. The common 'thermolabile' variant of methylene tetrahydrofolate reductase is a major determinant of mild hyperhomocysteinemia. *Q J Med* 1996;89:571–7.
 68. Kluijtmans LAJ, Van den Heuvel LPWJ, Boers GHJ, Frosst P, Stevens EMB, Van Oost BA, et al. Molecular genetic analysis in mild hyperhomocysteinemia: a common mutation in the methylene tetrahydrofolate reductase gene is a genetic risk factor for cardiovascular disease. *Am J Hum Genet* 1996;58:35–41.
 69. Wilcken DEL, Wang XL, Sim AS, McCredie RM. Distribution in healthy and coronary populations of the methylene tetrahydrofolate reductase (MTHFR) C₆₇₇T mutation. *Arterioscler Thromb Vasc Biol* 1996;16:878–82.
 70. Schmitz C, Lindpaintner K, Verhoef P, Gaziano JM, Buring J. Genetic polymorphism of methylene tetrahydrofolate reductase and myocardial infarction—a case-control study. *Circulation* 1996;94:1812–4.
 71. Gallagher PM, Meleady R, Shields DC, Tan KS, McMaster D, Rozen R, et al. Homocysteine and risk of premature coronary heart disease—evidence for a common gene mutation. *Circulation* 1996;94:2154–8.
 72. Adams M, Smith PD, Martin D, Thompson JR, Samani NJ. Genetic analysis of thermolabile methylene tetrahydrofolate reductase as a risk factor for myocardial infarction. *Q J Med* 1996;89:437–44.
 73. McAndrew PE, Brandt JT, Pearl DK, Prior TW. The incidence of the gene for thermolabile methylene tetrahydrofolate reductase in African Americans. *Thromb Res* 1996;83:195–8.
 74. Van Bockxmeer FM, Mamotte CDS, Vasikaran SD, Taylor RR. Methylene tetrahydrofolate reductase gene and coronary artery disease. *Circulation* 1997;95:21–3.
 75. Ma J, Stampfer MJ, Hennekens CH, Frosst P, Selhub J, Horsford J, et al. Methylene tetrahydrofolate reductase polymorphism, plasma folate, homocysteine, and risk of myocardial infarction in US physicians. *Circulation* 1996;94:2410–6.
 76. Jacques PF, Bostom AG, Williams RR, Ellison RC, Eckfeldt JH, Rosenberg IH, et al. Relation between folate status, a common mutation in methylene tetrahydrofolate reductase, and plasma homocysteine concentrations. *Circulation* 1996;93:7–9.
 77. Bostom AG, Shemin D, Lapane KL, Nadeau MR, Sutherland P, Chan J, et al. Folate status is the major determinant of fasting

- total plasma homocysteine levels in maintenance dialysis patients. *Atherosclerosis* 1996;123:193–202.
78. Finkelstein JD. Methionine metabolism in mammals. *J Nutr Biochem* 1990;1:228–37.
 79. Fenton WA, Rosenberg LE. Inherited disorders of cobalamin transport and metabolism. In: Scriver CR, Beaudet AL, Sly WS, Valle D, eds. *The metabolic and molecular basis of inherited disease*, 7th ed. New York: McGraw-Hill, 1995:3129–49.
 80. Li YN, Gulati S, Baker PJ, Brody LC, Banerjee R, Kruger WD. Cloning, mapping, and RNA analysis of the human methionine synthase gene. *Hum Mol Genet* 1996;5:1851–8.
 81. Leclerc D, Campeau E, Goyette P, Adjalla CE, Christensen B, Ross M, et al. Human methionine synthase: cDNA cloning and identification of mutations in patients of the *cbIG* complementation group of folate/cobalamin disorders. *Hum Mol Genet* 1996;5:1867–74.
 82. Chen LH, Liu ML, Hwang HY, Chen LS, Korenberg J, Shane B. Human methionine synthase—cDNA cloning, gene localization and expression. *J Biol Chem* 1997;272:3628–34.
 83. Watkins D, Rosenblatt DS. Genetic heterogeneity among patients with methylcobalamin deficiency. Definition of two complementation groups, *cbIE* and *cbIG*. *J Clin Invest* 1988;81:1690–4.
 84. Rosenblatt DS, Cooper BA, Pottier A, Lue-Shing H, Matiaszuk N, Grauer K. Altered vitamin B12 metabolism in fibroblasts from a patient with megaloblastic anemia and homocystinuria due to a new defect in methionine biosynthesis. *J Clin Invest* 1984;74:2149–56.
 85. Watkins D, Rosenblatt DS. Functional methionine synthase deficiency (*cbIE* and *cbIG*): clinical and biochemical heterogeneity. *Am J Med Genet* 1989;34:427–34.
 86. Sillaots SL, Hall CA, Hurlteloup V, Rosenblatt DS. Heterogeneity in *cbIG*: differential retention of cobalamin on methionine synthase. *Biochem Med Metab Biol* 1992;47:242–9.
 87. Gulati S, Baker P, Li YN, Fowler B, Kruger WD, Brody L, et al. Defects in human methionine synthase in *cbIG* patients. *Hum Mol Genet* 1996;5:1859–65.
 88. Kraus JP, Le K, Swaroop M, Ohura T, Tahara T, Rosenberg LE, et al. Human cystathionine β -synthase cDNA: sequence, alternative splicing and expression in cultured cells. *Hum Mol Genet* 1993;2:1633–8.
 89. Kozich V, Kraus JP. Screening for mutations by expressing patient cDNA segments in *E. coli*: homocystinuria due to cystathionine β -synthase deficiency. *Hum Mutat* 1992;1:113–23.
 90. De Franchis R, Kozich V, McInnes RR, Kraus JP. Identical genotypes in siblings with different homocystinuric phenotypes: identification of three mutations in cystathionine β -synthase using an improved bacterial expression system. *Hum Mol Genet* 1994;3:1103–8.
 91. Kruger WD, Cox DR. A yeast system for expression of human cystathionine β -synthase: structural and functional conservation of the human and yeast genes. *Proc Natl Acad Sci U S A* 1994;91:6614–8.
 92. Kruger WD, Cox DR. A yeast assay for functional detection of mutations in the human cystathionine β -synthase gene. *Hum Mol Genet* 1995;4:1155–61.
 93. Boers GHJ, Smals AGH, Trijbels FJM, Fowler B, Bakkeren JAJM, Schoonderwaldt HC, et al. Heterozygosity for homocystinuria in premature peripheral and cerebral occlusive arterial disease. *N Engl J Med* 1985;313:709–15.
 94. McGill JJ, Mettler G, Rosenblatt DS, Scriver CR. Detection of heterozygotes for recessive alleles. Homocyst(e)inemia: paradigm of pitfalls in phenotypes. *Am J Med Genet* 1990;36:45–52.
 95. Kozich V, Kraus E, De Franchis R, Fowler B, Boers GHJ, Graham I, et al. Hyperhomocysteinemia in premature arterial disease: examination of cystathionine β -synthase alleles at the molecular level. *Hum Mol Genet* 1995;4:623–9.
 96. Tsai MY, Bignell M, Schwichtenberg K, Hanson NQ. High prevalence of a mutation in the cystathionine β -synthase gene. *Am J Hum Genet* 1996;59:1262–7.
 97. Pancharuniti N, Lewis CA, Sauberlich HE, Perkins LL, Go PCP, Alvarez JO, et al. Plasma homocyst(e)ine, folate, and vitamin B₁₂ concentrations and risk for early onset coronary artery disease. *Am J Clin Nutr* 1994;59:940–8.
 98. Hopkins PN, Wu LL, Wu J, Hunt SC, James BC, Vincent GM, et al. Higher plasma homocyst(e)ine and increased susceptibility to adverse effects of low folate in early familial coronary artery disease. *Arterioscler Thromb Vasc Biol* 1995;15:1314–20.
 99. Morrison HI, Schaubel D, Desmeules M, Wigle DT. Serum folate and risk of fatal coronary heart disease. *JAMA* 1996;275:1893–6.
 100. Chasan-Taber L, Selhub J, Rosenberg IH, Malinow MR, Terry P, Tishler PV, et al. A prospective study of folate and vitamin B₆ and risk of myocardial infarction in US physicians. *J Am Coll Nutr* 1996;15:136–43.
 101. Verhoef P, Kok FJ, Kruyssen DACM, Schouten EG, Witteman JCM, Grobbee DE, et al. Plasma total homocysteine, B vitamins, and risk of coronary atherosclerosis. *Arterioscler Thromb Vasc Biol* 1997;17:989–95.
 102. Verhoef P, Stampfer MJ, Buring JE, Gaziano JM, Allen RH, Stabler SP, et al. Homocysteine metabolism and risk of myocardial infarction: relation with vitamins B₆, B₁₂, and folate. *Am J Epidemiol* 1996;143:845–59.
 103. Rimm EB, Willet WC, Hu FB, Sampson L, Colditz GA, Manson JE, et al. Folate and vitamin B₆ from diet and supplements in relation to risk of coronary heart disease among women. *JAMA* 1998;279:359–64.
 104. Ubbink JB, Vermaak WJH, van der Merwe A, Becker PJ. Vitamin B-12, vitamin B-6, folate nutritional status in men with hyperhomocysteinemia. *Am J Clin Nutr* 1993;57:47–53.
 105. Hultberg B, Berglund M, Andersson A, Frank A. Elevated plasma homocysteine in alcoholics. *Alcohol Clin Exp Res* 1993;17:687–9.
 106. Cravo ML, Gloria LM, Selhub J, Nadeau MR, Camilo ME, Resende MP, et al. Hyperhomocysteinemia in chronic alcoholism: correlation with folate, vitamin B-12, and vitamin B-6 status. *Am J Clin Nutr* 1996;63:220–4.
 107. Berger PB, Jones JD, Olson LJ, Edwards BS, Frantz RP, Rodeheffer RJ, et al. Increase in total plasma homocysteine concentration after cardiac transplantation. *Mayo Clin Proc* 1995;70:125–31.
 108. Gupta A, Moustapha A, Jacobsen DW, Goormastic M, Tuzcu EM, Hobbs R, et al. Hyperhomocysteinemia and vascular complications in heart transplant recipients: relationships with folate and vitamin B₆. *Transplantation* 1998;65:544–50.
 109. Ambrosi P, Barlatier A, Habib G, Garcon D, Kreitman B, Roland PH, et al. Hyperhomocysteinemia in heart transplant recipients. *Eur Heart J* 1994;15:1191–5.
 110. Nedrebo B, Ericsson U-B, Ueland PM, Refsum H, Lien EA. Plasma levels of the atherogenic amino acid homocysteine in hyper- and hypothyroid patients [Abstract]. *Ir J Med Sci* 1995;164 (Suppl): 13.
 111. Hussein WI, Jacobsen DW, Green R, Faiman C. Hyperhomocysteinemia and hypothyroidism: normalization with L-thyroxine replacement [Abstract]. *Proceedings, Am Assoc Clin Endocrinol, Orlando, 1998.*
 112. Mansoor MA, Svardal AM, Schneede J, Ueland PM. Dynamic relation between reduced, oxidized, and protein-bound homocysteine and other thiol components in plasma during methionine loading in healthy men. *Clin Chem* 1992;38:1316–21.

- 113.** Guttormsen AB, Schneede J, Fiskerstrand T, Ueland PM, Refsum HM. Plasma concentrations of homocysteine and other amino-thiol compounds are related to food intake in healthy human subjects. *J Nutr* 1994;124:1934–41.
- 114.** Bostom AG, Jacques PF, Nadeau MR, Williams RR, Ellison RC, Selhub J. Post-methionine load hyperhomocysteinemia in persons with normal fasting total plasma homocysteine: initial results from the NHLBI Family Heart Study. *Atherosclerosis* 1995;116:147–51.
- 115.** Silberberg J, Crooks R, Fryer J, Wlodarczyk J, Nair B, Guo XW, et al. Gender differences and other determinants of the rise in plasma homocysteine after L-methionine loading. *Atherosclerosis* 1997;133:105–10.
- 116.** D'Angelo A, Selhub J. Homocysteine and thrombotic disease. *Blood* 1997;90:1–11.
- 117.** Wang H, Yoshizumi M, Lai K, Tsai J-C, Perrella MA, Haber E, et al. Inhibition of growth and p21^{ras} methylation in vascular endothelial cells by homocysteine but not cysteine. *J Biol Chem* 1997;272:25380–5.
- 118.** Upchurch GR Jr, Welch GN, Fabian AJ, Freedman JE, Johnson JL, Keaney JF Jr, et al. Homocyst(e)ine decreases bioavailable nitric oxide by a mechanism involving glutathione peroxidase. *J Biol Chem* 1997;272:17012–7.
- 119.** Upchurch GR Jr, Welch GN, Fabian AJ, Pigazzi A, Keaney JF Jr, Loscalzo J. Stimulation of endothelial nitric oxide production by homocyst(e)ine. *Atherosclerosis* 1997;132:177–85.
- 120.** Tsai J-C, Perrella MA, Yoshizumi M, Hsieh C-M, Haber E, Schlegel R, et al. Promotion of vascular smooth muscle growth by homocysteine: a link to atherosclerosis. *Proc Natl Acad Sci U S A* 1994;91:6369–73.
- 121.** Tsai J-C, Wang H, Perrella MA, Yoshizumi M, Sibinga NES, Tan LC, et al. Induction of cyclin A gene expression by homocysteine in vascular smooth muscle cells. *J Clin Invest* 1996;97:146–53.
- 122.** Majors A, Ehrhart LA, Pezacka EH. Homocysteine as a risk factor for vascular disease—enhanced collagen production and accumulation by smooth muscle cells. *Arterioscler Thromb Vasc Biol* 1997;17:2074–81.
- 123.** Van den Berg M, Boers GHJ, Franken DG, Blom HJ, Van Kamp GJ, Jakobs C, et al. Hyperhomocysteinemia and endothelial dysfunction in young patients with peripheral arterial occlusive disease. *Eur J Clin Invest* 1995;25:176–81.
- 124.** Tawakol A, Omland T, Gerhard M, Wu JT, Creager MA. Hyperhomocyst(e)inemia is associated with impaired endothelium-dependent vasodilation in humans. *Circulation* 1997;95:1119–21.
- 125.** Woo KS, Chook P, Lolin YI, Cheung BN, Chan RN, Sun YY, et al. Hyperhomocyst(e)inemia is a risk factor for arterial endothelial dysfunction in humans. *Circulation* 1997;96:2542–4.
- 126.** Lentz SR, Sobey CG, Piegors DJ, Bhopatkar MY, Faraci FM, Malinow MR, et al. Vascular dysfunction in monkeys with diet-induced hyperhomocyst(e)inemia. *J Clin Invest* 1996;98:24–9.
- 127.** Lentz SR, Malinow MR, Piegors DJ, Bhopatkar-Teredesai M, Faraci FM, Heistad DD. Consequences of hyperhomocyst(e)inemia on vascular function in atherosclerotic monkeys. *Arterioscler Thromb Vasc Biol* 1997;17:2930–4.
- 128.** Jacobsen DW, Savon SR, Stewart RW, Robinson K, Green R, Kottke-Marchant K, et al. Limited capacity for homocysteine catabolism in vascular cells and tissues: a pathophysiologic mechanism for arterial damage in hyperhomocysteinemia? [Abstract]. *Circulation* 1995;91:29.
- 129.** Poddar R, Sivasubramanian N, Robinson K, Jacobsen DW. Homocysteine modulates the expression of a specific cytokine (monocyte chemoattractant protein 1) in human aortic endothelial cells [Abstract]. *Circulation* 1997;96:1286.
- 130.** Cooper BA, Rosenblatt DS. Inherited defects of vitamin B₁₂ metabolism. *Annu Rev Nutr* 1987;7:291–320.
- 131.** Brattström L, Israelsson B, Jeppsson J-O, Hultberg BL. Folic acid—an innocuous means to reduce plasma homocysteine. *Scand J Clin Lab Invest* 1988;48:215–21.
- 132.** Brattström L, Israelsson B, Norrving B, Bergqvist D, Thorne J, Hultberg B, et al. Impaired homocysteine metabolism in early-onset cerebral and peripheral occlusive disease—effects of pyridoxine and folic acid. *Atherosclerosis* 1990;81:51–60.
- 133.** Landgren F, Israelsson B, Lindgren A, Hultberg B, Andersson A, Brattström L. Plasma homocysteine in acute myocardial infarction: homocysteine-lowering effect of folic acid. *J Intern Med* 1995;237:381–8.
- 134.** Food and Drug Administration. Food standards: amendment of standards of identity for enriched grain products to require addition of folic acid. Final rule. *Fed Regist* 1996;61:8781–807.
- 135.** Ward M, McNulty H, McPartlin J, Strain JJ, Weir DG, Scott JM. Plasma homocysteine, a risk factor for cardiovascular disease, is lowered by physiological doses of folic acid. *Q J Med* 1997;90:519–24.
- 136.** Stabler SP, Lindenbaum J, Allen RH. Vitamin B-12 deficiency in the elderly: current dilemmas. *Am J Clin Nutr* 1997;66:741–9.
- 137.** Bostom AG, Shemin D, Lapane KL, Hume AL, Yoburn D, Nadeau MR, et al. High dose B-vitamin treatment of hyperhomocysteinemia in dialysis patients. *Kidney Int* 1996;49:147–52.