

Homolog of protein kinase M ζ maintains context aversive memory and underlying long-term facilitation in terrestrial snail *Helix*

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It has been shown that a variety of long-term memories in different regions of the brain and in different species are quickly erased by local inhibition of protein kinase M ζ (PKM ζ), a persistently active protein kinase. Using antibodies to mammalian PKM ζ , we describe in the present study the localization of immunoreactive molecules in the nervous system of the terrestrial snail *Helix lucorum*. Presence of a homolog of PKM ζ was confirmed with transcriptomics. We have demonstrated in behavioral experiments that contextual fear memory disappeared under a blockade of PKM ζ with a selective peptide blocker of PKM ζ zeta inhibitory peptide (ZIP), but not with scrambled ZIP. If ZIP was combined with a “reminder” (20 min in noxious context), no impairment of the long-term contextual memory was observed. In electrophysiological experiments we investigated whether PKM ζ takes part in the maintenance of long-term facilitation (LTF) in the neural circuit mediating tentacle withdrawal. LTF of excitatory synaptic inputs to premotor interneurons was induced by high-frequency nerve stimulation combined with serotonin bath applications and lasted at least 4 h. We found that bath application of 2×10^{-6} M ZIP at the 90th min after the tetanization reduced the EPSP amplitude to the non-tetanized EPSP values. Applications of the scrambled ZIP peptide at a similar time and concentration didn’t affect the EPSP amplitudes. In order to test whether effects of ZIP are specific to the synapses, we performed experiments with LTF of somatic membrane responses to local glutamate applications. It was shown earlier that serotonin application in such an “artificial synapse” condition elicits LTF of responses to glutamate. It was found that ZIP had no effect on LTF in these conditions, which may be explained by the very low concentration of PKM ζ molecules in somata of these identified neurons, as evidenced by immunocytochemistry. Obtained results suggest that the *Helix* homolog of PKM ζ might be involved in post-induction maintenance of long-term changes in the nervous system of the terrestrial snail.

Keywords: invertebrates, reconsolidation, memory blockade, withdrawal behavior, memory maintenance, contextual fear conditioning

Introduction

It has been shown that a constitutively active fragment of the mammalian atypical protein kinase C ζ protein kinase M ζ (PKM ζ) plays a critical role in the persistence of long-term potentiation (LTP) in the mammalian hippocampus (Ling et al., 2002; Pastalkova et al., 2006), as well as in several forms of mammalian memory (Serrano et al., 2005; Shema et al., 2007; Gámiz and Gallo, 2011; recently reviewed in Glanzman, 2013). The ability of PKM ζ to maintain LTP and memory is due to its unique structure as an autonomously active protein kinase C isoform (Sacktor et al., 1993; Sacktor, 2011). PKM ζ protein persistently increases in the CA1 area of the hippocampus during LTP, and this increase correlates with the extent and duration of synaptic potentiation during LTP maintenance (Osten et al., 1996). The persistent activity of PKM ζ is both necessary and sufficient for maintaining synaptic LTP (Ling et al., 2002) and long-term memory storage (Pastalkova et al., 2006; Shema et al., 2011). Recently shown localization of PKM ζ within postsynaptic densities and spines (Hernández et al., 2013) supports the previously proposed (Sajikumar et al., 2005; Sacktor, 2011) “synaptic autotagging” model for explaining how this kinase stores and maintains the long-term memory.

Recent findings in invertebrate model organisms show an evolutionarily conserved role of atypical PKCs, homologous to mammalian PKM ζ , in memory acquisition and maintenance (Drier et al., 2002; Cai et al., 2011). An atypical PKC has been cloned from the nervous system of *Aplysia* (Bougie et al., 2009). This *Aplysia* PKC, PKC Apl III, can undergo proteolytic cleavage by calpain, thereby yielding a PKM fragment, PKM Apl III. It is also shown that 5-HT appears to activate PKM Apl III in motor neurons of *Aplysia* (Villareal et al., 2009). Recently it was demonstrated that long-term memory in *Aplysia* is maintained via a positive-feedback loop involving PKM Apl III-dependent protein phosphorylation (Cai et al., 2011).

In the present study, using as an animal model, a phylogenetically advanced pulmonate terrestrial snail *Helix*, which is capable of demonstrating associative forms of learning, and has well described neural circuits underlying its behavior (Balaban, 2002), we have investigated: (a) whether a homolog of PKM ζ exists in the neural transcriptome of this animal; (b) where this molecule is located in the nervous system; (c) whether the inhibitor of PKM ζ blocks the long-term (days) associative aversive memory in behavioral experiments; and (d) whether the inhibitor of PKM ζ blocks the long-term synaptic changes (hours) in identified premotor interneurons triggering the withdrawal responses.

Methods

Experiments were carried out in adult snails *Helix lucorum* L. (Crimea population) weighing 20–30 g. All animals were kept in terraria at temperature $22 \pm 2^\circ\text{C}$, in a 12:12 light/dark cycle. The snails were kept in an active state at least 2 weeks before the experiment in a wet environment and were fed regularly with cabbage. Two days before the training

session, the snails were deprived of food. Each snail was used in only one series of experiments. In total scores of 43, animals that survived the training and testing procedures and were in good health at least a week after the end of experiment were used for statistical evaluation in behavioral experiments. Isolated central nervous system (CNS) was used for electrophysiological experiments. Details of preparation and identification of neurons, are given elsewhere (Ierusalimsky et al., 1992; Balaban, 2002; Malyshev and Balaban, 2002). Injection of cold isotonic MgCl_2 was made before the CNS isolation to minimize pain. Experimental procedures were in compliance with the Guide for the Care and Use of Laboratory Animals published by the National Institutes of Health, and the protocol was approved by the Ethical Committee of the Institute of Higher Nervous Activity and Neurophysiology of Russian Academy of Sciences.

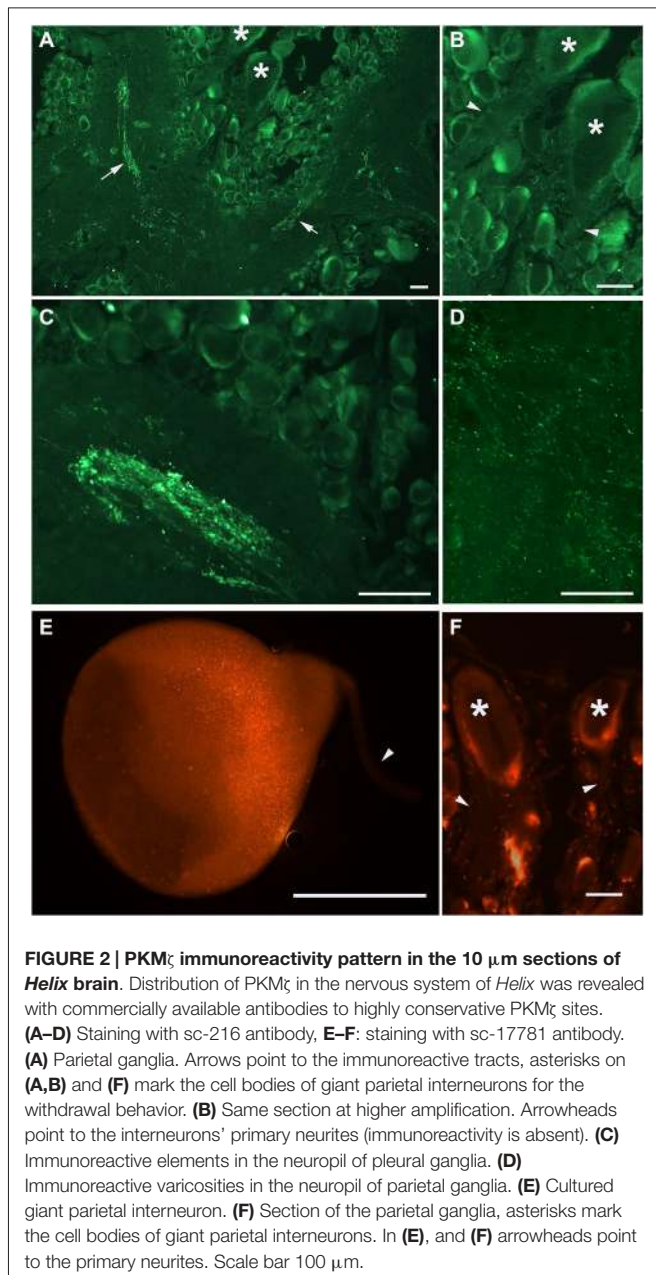
Behavioral Experiments

Apparatus and Analysis of Behavior

In the experimental set-up (Context 1), the snail was tethered by its shell in a manner allowing it to crawl on a ball that rotated freely in a water solution containing 0.01% NaCl (**Figure 3A**, right inset). The ball was covered with aluminium foil to complete an electrical circuit between the animal's foot and a carbon electrode placed in the water. Electric shock was delivered using a 1–4 mA, 1 s current through a macroelectrode applied manually to the dorsal surface of the snail's foot (**Figure 3A**). Punctate mechanical stimuli were applied with calibrated von Frey hairs, permitting delivery of pressures ranging from 6 (estimated as weak) to 68 gr/mm^2 (estimated as noxious). After several pilot series, the behavioral response, the intensity (25 gr/mm^2), and the location of tactile stimulation were chosen. Ommatophores (posterior tentacles) withdrawal in response to chosen intensity of tactile stimulation of the rostral part of skin 4–5 mm behind the posterior tentacles was at the level of 10–30% of maximal in normal animals. In pilot experiments, it was shown that responses to such test stimulation were sensitized after noxious stimuli, and this part of the foot skin was chosen as the standard place for tactile stimulation. An investigator, blind to the experimental histories of the animals, applied the tactile stimuli to the snail's skin and video recorded the tentacle withdrawal. To quantify and average the results, we analyzed off-line the distance between tip and base of the tentacle and scored the withdrawal amplitude as a percentage of initial length of the tentacle in each trial.

Learning and Reminder Protocol

Before training, each snail was exposed for 30 min daily for 2 days to the experimental set-up. Then the first test session (T) was performed for all groups (first day, **Figure 3A**). Blind testing was performed for each snail in two alternating contexts (Context 1 was a ball floating in water, while the Context 2 was a flat glass similar to glass of terrarium where the snails were kept between the experimental sessions, see inset on **Figure 3A**). After obtaining the pre-training scores, all groups of snails received five electrical shocks per day with 20–30 min



Next day after the second test session (T1), one group of snails (G2) was reminded of training by placing the snails for 20 min (Reminder) in the same Context 1 where they were previously shocked (on the ball, **Figure 3A**). Twenty minutes before the reminder, the snails were injected either with ZIP or scrambled ZIP (scrZIP, 0.4 mg in 0.2 ml of saline plus 0.5 ml of saline to equalize volume per snail weighing 20–30 g). On the second day after a session of drug injections or “reminding”, the third test session (T2) was performed for all parallel groups in two different contexts (detailed protocol in Balaban et al., 2011, 2014).

Drugs and Injections

ZIP (TOCRIS) and scrambled ZIP were dissolved in sterile Ringer saline [in mM: 100 NaCl, 4 KCl, 7 CaCl₂, 5 MgCl₂, and

10 Tris-HCl buffer (pH 7.8)]. Estimated final concentrations in the hemolymph of free behaving animals of ZIP and scrZIP were 2×10^{-6} M. Selected concentrations were effective in our electrophysiological experiments in snails without obvious toxic effects. For calculating final concentrations in the nervous system, each gram of the snail body weight was scored as 1 ml.

Drugs for *in vivo* injections were prepared in deionized water as a stock solution at a concentration 28.6-fold greater than required. Because the snails used in these experiments were comparable in weight ($20\text{g} \pm 2$), 0.7 ml of the drug solutions were injected into the hemocoel, thereby achieving a required concentration in the animals' body ($0.7\text{ ml} \times 28.6 = 20\text{ ml}$).

Intracoelemic injections were performed with a fine needle via an insensitive part of the foot skin normally hidden under the shell. During injections, the snails stopped locomotion and lowered the ommatophores, which was most likely due to the experimenter raising the shell. However, the snails never showed a generalized withdrawal into the shell.

Electrophysiological Experiments

Intracellular recordings from isolated brain ganglia were made using standard electrophysiological techniques. Identified withdrawal premotor interneurons of the parietal ganglia (Pa3 and Pa2; Balaban, 2002) were penetrated with sharp glass microelectrodes filled with 2 M potassium acetate (tip resistance, 15–20 M Ω). The cutaneous and intestinal nerves were stimulated via plastic suction electrodes with 3 ms stimuli. Intensity of stimuli was adjusted in each experiment to elicit complex EPSPs of 5–8 mV amplitude. Intracellular signals were recorded with preamplifiers (Axoclamp 2B, Axon Instruments, CA, USA), digitized, and stored on a computer (Digidata 1400A A/D converter and Axoscope 10.0 software, both from Axon Instruments, CA, USA).

In the first series of experiments (homosynaptic facilitation) we used test stimulation of the cutaneous nerve, and potentiation was elicited by tetanization of the same cutaneous nerve associated with bath serotonin application. Experimental protocol started with five test stimuli with a 10 min interstimulus interval, followed by tetanization (three high-frequency 10 Hz trains of stimuli; duration of each train 1 min; with 5 min intervals between trains; test stimulus amplitude was increased 10 fold), followed by posttetanic testing with single stimuli of normal amplitude and a 10 min interstimulus interval for several hours. Serotonin was applied to the experimental bath just before each tetanization train to a final bath concentration of 10^{-5} M with washout in 4 min after each tetanization train. Ninety minutes after the last tetanization train, the saline flow (0.1 ml/min) of the perfusion system was switched off, and ZIP or scrZIP stock solution was applied to a final bath concentration of 2×10^{-6} M.

In the second series of experiments (heterosynaptic facilitation) we also used test stimulation of the second cutaneous nerve but potentiation was elicited by tetanization of the intestinal nerve associated with serotonin application to the experimental bath. Otherwise, experimental protocol was

identical to the described protocol above for homosynaptic facilitation.

In the third series (artificial synapse) we used two intracellular electrodes, one for recording and another for stimulation. Before recording, somata of the same premotor parietal giant interneurons were synaptically isolated by cutting the ganglia neuropile at a distance of 400 micrometers from the soma where the single giant neurite of these cells starts branching (our unpublished observations). All synaptic inputs of parietal withdrawal interneurons are contained in the neuropil of parietal ganglia (Balaban, 2002), and were cut out in this type of preparation, therefore diminishing possible polysynaptic effects of transmitter applications. Input resistance of the cell was monitored using fixed amplitude hyperpolarization current impulses of 2 s duration. In this series glutamate (Glu) was dissolved to a concentration of 2×10^{-3} M in Ringer solution with 0.15% of vital dye Fast Green and sucked in the glass pipette with ~ 1 μ m tip. Aliquots were stored in a freezer and were warmed at room temperature for at least 30 min before use. Applications of Glu to the neural membrane were made using picoinjector (PV830 PicoPump, WPI) and pressure 70–80 psi, adjusting minimal pulse duration in order to get stable responses in a neuron with 3–10 mV amplitude. Preparations in this series of experiments were perfused (2 ml/min, bath volume 1.5 ml) using a 1 mm suction tube located close to the recorded cell and pipette with Glu, thus washing out the Glu in less than 0.5 s after the application. This timing was estimated visually with a Fast Green applied together with Glu. After the first 15 applications of Glu with 1/min intervals, serotonin was added three times in final bath concentration 10^{-5} M for 10 min, then 10 min washout. During all this time the Glu was applied repeatedly. ZIP was added 90 min after the end of the last serotonin application. After ZIP application, the perfusion system was switched to the closed cycle.

Transcriptome Analysis

Transcriptome of *Helix lucorum taurica* L. was prepared from eight nervous systems processed separately on Illumina HiSeq 2000. We extracted RNA from each sample, using an RNAaqueous™ micro kit (Ambion, Austin, TX, USA). The quality and quantity of RNA were assessed using an Agilent 2100 Bioanalyzer (Agilent Technologies, CA, USA). RNA samples were subsequently used in cDNA library construction and Illumina sequencing. DNA was processed as described in the TruSeq DNA Sample Preparation Guide (Illumina). The library was quantified using fluorimetry with Qubit (Invitrogen) and real-time PCR and diluted up to final concentration of 8 pM. The diluted library was clustered on a paired-end flowcell (TruSeq PE Cluster Kit v3) using a cBot instrument and sequenced using a HiSeq2000 sequencer with the TruSeq SBS Kit v3-HS with a read length of 101 bp from each end. Paired-end libraries with 250-bp inserts were constructed with read length 100 + 100 bp and ~ 200 mln reads in total. The transcriptome assembly was constructed *de novo* using Trinity software (Grabherr et al., 2011). Multiple sequence alignment (MSA) was constructed by DIALIGN-PFAM online tool with default parameters (Al Ait et al., 2013).

Immunohistochemistry

Two commercially available antibodies to PKM ζ were selected: sc-216, and sc-11781 (Santa Cruz Biotechnology). Both were produced to the highly conservative (C-terminus) part of PKM ζ (see **Figure 1**). The brains of eight adult snails were subjected to the immunochemical procedure, as well as the individually identified four withdrawal interneurons mechanically isolated with the primary neurite. The brains were fixed in 4% paraformaldehyde in 0.1 PBS. The duration of fixation was 2 h at 4°C and it was followed by washes in PBS. The brains were mounted in Paraplast and sectioned at a thickness of 10 μ m. Prior to incubation with the primary antibody solution, sections were washed in the blocking solution for 2 h. The blocking solution contained 0.5% Triton X-100, 0.01% sodium azide, 5% normal goat serum (Sigma) and 1% BSA (Sigma) in PBS. The staining procedure was: primary antibody for 24–48 h; wash for 2 h; secondary antibody for 24 h; wash for 2 h. The secondary antibody was either Alexa-488 goat anti-rabbit (sc-216) or Alexa-546 goat anti-mouse (sc-17781) conjugated antibody. Finally, the sections were embedded in Aqua Poly/Mount (Polyscience). Cultured neurons were processed as a whole-mount. The preparations were examined with an AxioPlan microscope (Zeiss, Germany) supplied with the program KS-300 for analysis of visual images. To test the specificity of immunostaining, the primary antiserum was omitted. No staining resulted in this series.

Statistical Evaluation of Data

Blind testing was performed at different time intervals as shown in inset of **Figure 3**. Comparison between groups was made only for parallel groups of animals in one experimental series. We used nonparametric Mann-Whitney rank sum test to compare performance of two groups of snails, and Wilcoxon signed-rank test was used for comparison of performance of the same group. On all figures significant differences in performance are indicated.

Results

Helix Sequence Homology to the *Aplysia* and Mammalian PKM ζ

Transcriptome of *Helix lucorum taurica* L. was prepared from eight nervous systems processed separately on Illumina HiSeq 2000. Paired-end libraries with 250-bp inserts were constructed with read length 100 + 100 bp and ~ 200 mln reads in total. The transcriptome assembly was constructed *de novo* using Trinity software (Grabherr et al., 2011). The search resulted in finding many described genes of mollusks, and among them a homolog of the *Aplysia* atypical PKC with high identity in amino acids sequence (**Figure 1**). *Helix* atypical PKC sequence showed high homology with pulmonate *Lymnaea* (94% aa identity) and opisthobranch *Aplysia* (91% aa identity) molluscan sequences.

Localization of PKM ζ in the Nervous System

Our next step was aimed to analyze the distribution of PKM ζ in the nervous system of *Helix*. We selected two commercially

available antibodies (sc-216, and sc-11781) both produced to the highly conservative (C-terminus) part of PKM ζ (see **Figure 1**), and immunohistochemically analyzed distribution of the PKM ζ -reactive sites. Mainly, the results were similar with both antibodies used. It appeared that a small amount of immunoreactive fluorescence can be seen in cytoplasm of most neurons including the parietal giant interneurons used in electrophysiological experiments (stars on **Figures 2A,B,F**). No fluorescence was observed in large neurites of these cells (arrowheads on **Figures 2B,F**), while in the neuropil regions glomeruli-like areas with intensive specific fluorescent small blobs and neurite-like conglomerates were observed (**Figures 2C,D**). Obtained results suggest that the PKM ζ -immunoreactive molecules are present in neurons of *H. lucorum*, both in cell bodies and in places where the majority of synapses are supposed to be present. In cultured giant parietal neurons, no immunoreactivity was observed in the primary neurites (arrowhead on **Figure 2E**), while the immunoreactive product in the cell body was detected in small granules (**Figure 2E**).

Behavioral Experiments

In this series of experiments, we have investigated involvement of PKM ζ in maintenance of contextual memory in terrestrial snail *Helix*. Three groups of snails were randomly tested in two different contexts (ball and glass, see inset on **Figure 2A**) before the training session (T, see protocol in **Figure 3A**). Percentage of maximal withdrawal to tactile stimulation was scored. Then the snails were trained (shocked) for 10 days to remember the context in which they were shocked (Context 1—on the ball) and tested with 1 day rest interval for aversive context memory (T1 on **Figure 3A**). After that, group 1 was injected with ZIP without a reminder, group 2 was injected with ZIP 30 min before the reminder, and was reminded (20 min in noxious context, no shocks during reminder) of the context in which they were shocked. Group 3 was injected with scrZIP without the reminder. All groups were tested 24 h later for maintenance of context memory (T2, **Figure 3A**).

Prior to the training session, the behavioral responses in two contexts did not differ significantly in all groups (panel “before learning” in **Figure 3B**). On the second day after a 10-day session of electric shocks in Context 1, the context conditioning was observed as a highly significant difference of behavioral response amplitudes in two contexts in all groups (panel “after learning”, **Figure 3B**, $p < 0.001$ for all groups, Wilcoxon Signed Rank Test, $z = 3.6$ for G1, $z = 3.9$ for G2, $z = 3.5$ for G3). On the day following testing of context memory, a session of “reminding” (no shocks, just 20 min in Context 1) and drug injections with or without reminding was performed. Next day testing of long-term context memory demonstrated that ZIP injection without a reminder completely abolished the memory in G1, while ZIP + reminder and scrZIP without reminder had no significant effect on context memory (panel “next day after injections”, **Figure 3B**, $p < 0.001$ for G2 and G3, Wilcoxon Signed Rank Test, $z = 3.7$ for G2, $z = 3.4$ for G3). The results evidence that blockade of PKM ζ results in a loss of consolidated

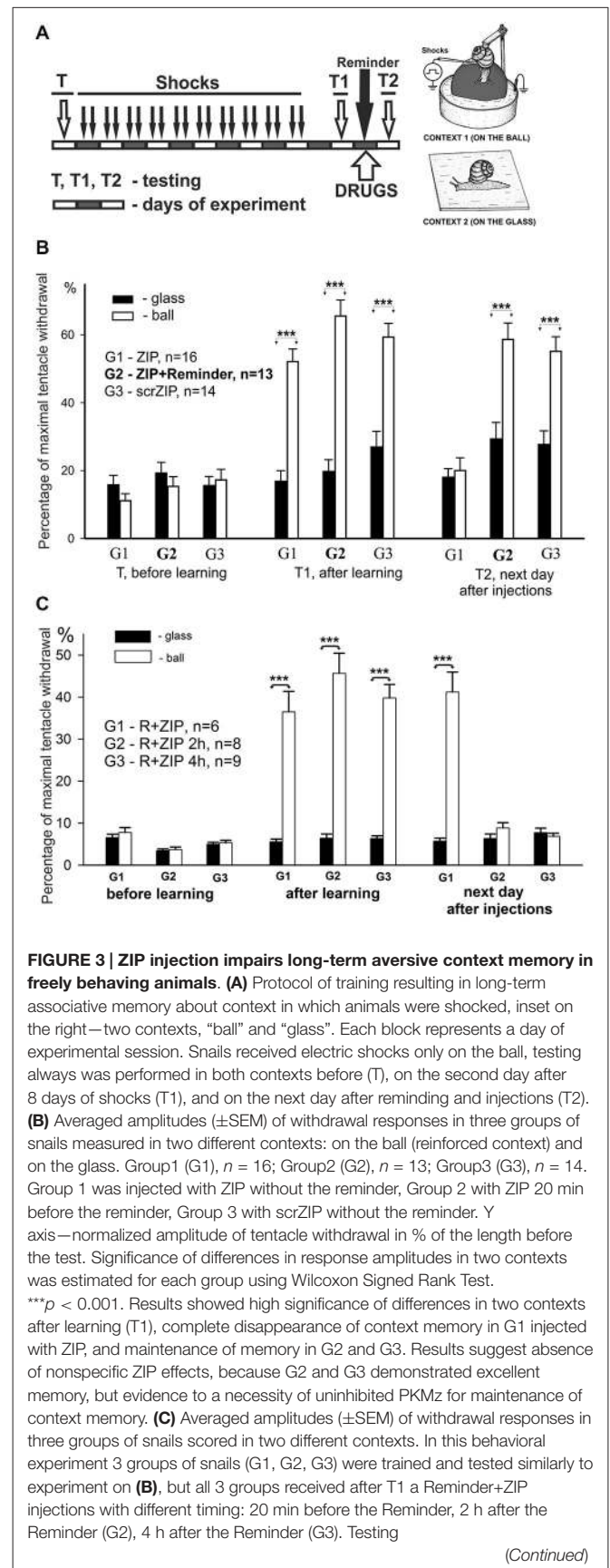


FIGURE 3 | Continued

on the next day after Reminder+injections showed excellent maintenance of memory in G1 (similar to obtained in G2 in experiment shown on “B”), and disappearance of significant context memory in G2 injected with ZIP 2 h after the Reminder, and complete disappearance of memory in G3 injected with ZIP 4 h after the Reminder. Results suggest that ZIP can be effective in conditions of reconsolidation when a new memories/PKM ζ molecules are supposed to be formed if the timing of ZIP effect is compatible with timing of new molecules of PKM ζ (2–4 h after reconsolidation procedure).

memory, while the same blockade in conditions of reminding and further reconsolidation of memory (which is assumed to take place in several hours) results in maintaining or, more probably, renewal of memory.

We decided to test whether ZIP would be effective at the time after reminding when memory starts to re-consolidate. With the aim to describe the time window of reconsolidation in our model, we repeated the experiments in another groups of snails in conditions that completely repeat conditions of G2 (**Figure 3B**), but injected ZIP not only 20 min before the reminder, but also at 2 h after the reminder, and at 4 h after the reminder. We have found that injection of ZIP just before the reminding did not influence the snail withdrawal as in the previous series of experiments. At the same time, injection of PKM ζ blocker 2 and 4 h after the reminding completely blocked the contextual memory (**Figure 3C,G2,G3**). Therefore, these results demonstrate with great significance that ZIP injection starts to be effective 2 h and more after the reminder, which completely corresponds to the assumption that long-term memory reconsolidation starts about 2 h after the reminder, and that the molecular substrate for ZIP is formed about this time.

Electrophysiological Experiments on Isolated CNS

Long-term facilitation (LTF) of excitatory synaptic inputs from sensory neurons to giant premotor interneurons triggering withdrawal in *Helix* is supposed to be a basis of aversion learning and memory in terrestrial snails (Balaban, 2002). We investigated whether the PKM ζ takes part in maintenance of LTF in the neural circuit of tentacle and body withdrawal. Normally, the LTF of excitatory synaptic inputs to identified premotor interneurons induced by high-frequency nerve stimulation combined with serotonin bath applications or extracellular stimulation of serotonergic cells lasts at least 4 h (Balaban et al., 2004).

In present experiments, we compared effects of ZIP or scrZIP on dynamics of LTF of complex excitatory synaptic inputs to premotor withdrawal interneurons induced by high-frequency tetanization (homosynaptic—of the tested nerve, or heterosynaptic—of another nerve) combined with several serotonin bath applications (five applications for heterosynaptically induced LTF and three applications for homosynaptically induced facilitation, see Section Methods). The protocol described above (**Figure 4A**) typically induced a strong and long-lasting increase in complex EPSP amplitude (examples on **Figure 4B**, averaged data on **Figure 4C**, open

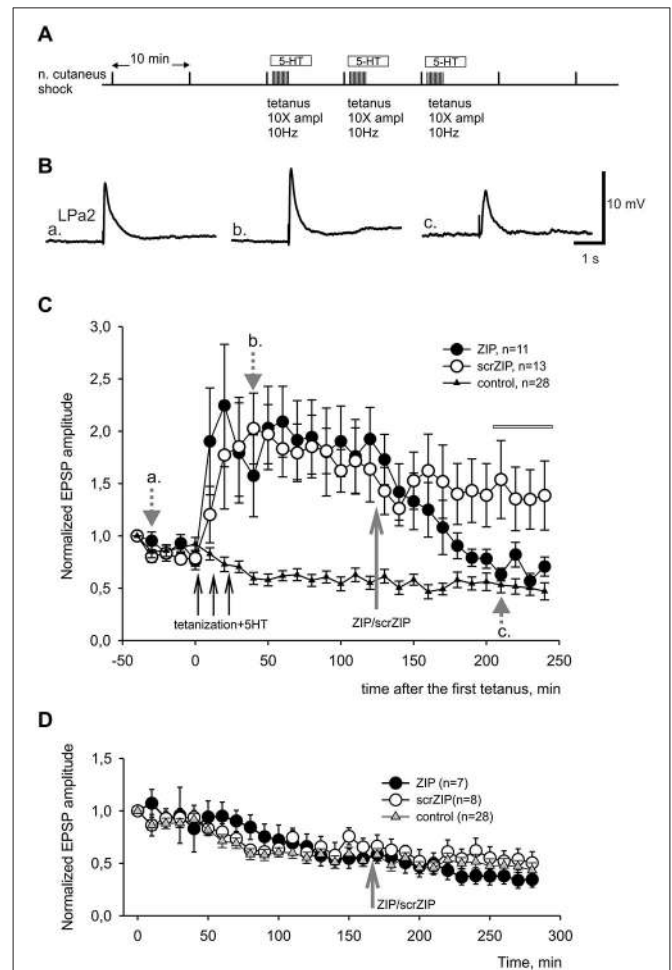
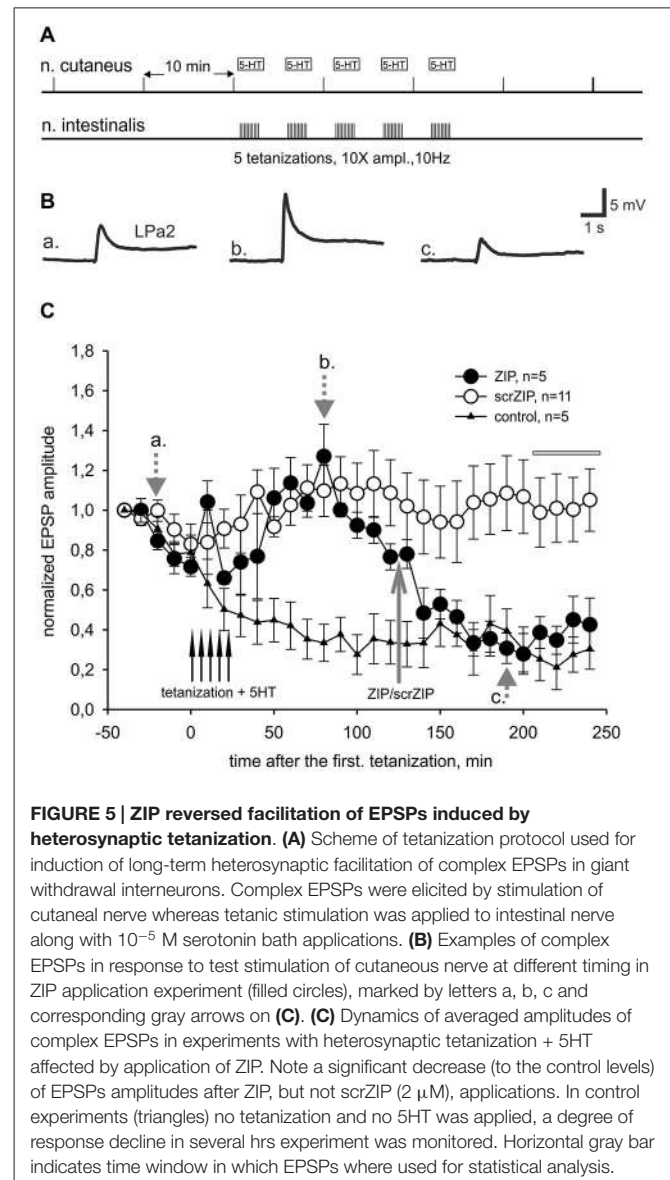


FIGURE 4 | ZIP reversed facilitation of EPSPs induced by tetanization of homosynaptic input. (A) Protocol of experiment, tetanization was applied to the same nerve with 10fold increased amplitude. Tetanic stimulation trains were applied along with 10^{-5} M serotonin bath applications. **(B)** Examples of complex EPSPs in response to stimulation of cutaneous nerve at different timing in experiment, marked by letters a, b, and corresponding gray arrows on **(C)**. **(C)** Averaged results of changes in amplitudes of complex EPSPs after tetanization and after application of ZIP or scrZIP applied at final concentration $2 \mu\text{M}$ 90 min after the tetanization. In control experiments (triangles) no tetanization and no 5HT was applied, a degree of response decline in several hours experiment was monitored. Averaged data are given as mean \pm SEM. **(D)** Averaged results of changes in amplitudes of complex EPSPs after application of ZIP or scrZIP without tetanization and 5HT. Note a significant (to the control levels) decrease of EPSPs amplitudes after ZIP, but not scrZIP, applications. Horizontal gray bar indicates time window in which EPSPs were used for statistical analysis.

and closed circles). Repeated nerve stimulation with single stimuli with 10 min intervals without tetanization and without serotonin application (control experiments) produced some habituation of responses which is typical for this preparation (**Figure 4C**, triangles). We have found that application of ZIP in final bath concentration of $2 \mu\text{M}$ at 90th min after the end of tetanization led to a significant decrease of complex EPSPs amplitudes, almost returning them to the level of nontetanized control 80 min after application (**Figure 4C**, filled circles).

At the same time, application of scrZIP did not influence the LTF (**Figure 4C**, open circles). Average EPSP amplitude, measured in the interval 210–240 min after the first tetanization in experiments with scrZIP application was $140.6 \pm 15.8\%$ of baseline before the tetanization, whereas at the same time period, under ZIP application the averaged EPSP amplitude significantly decreased to $68.1 \pm 4.5\%$, ($p < 0.001$, Mann-Whitney rank sum test). However, it was still significantly ($p < 0.01$) greater than averaged amplitude of responses in control (non-tetanized) experiments ($46.5 \pm 3.2\%$). In order to check if ZIP/scrZIP can affect the amplitude of non-tetanized EPSPs we performed a separate series of experiments where ZIP/scrZIP were applied at the same time point as in experiments with LTF but tetanization along with 5-HT treatment were omitted. No significant changes in average EPSP amplitude were found after both ZIP and scrZIP applications in these experiments (**Figure 4D**). In some experiments, the withdrawal interneurons were impaled with two electrodes and cell input resistance was monitored by applying negative current steps through one electrode while measuring membrane responses with the other one. No significant changes in the cell input resistance were detected after ZIP or scrZIP applications (data not shown).

In our behavioral experiments, the snails were tested by applying the tactile test stimuli to the skin of the head of the animal, which corresponded to cutaneous nerve stimulation in the first series of electrophysiological experiments. However, electrical shocks used in our learning paradigm as negative reinforcement were applied to the rear part of the snail body. Although current was apparently flowing through the whole body of the animal, maximum strength of stimulation definitely was not in the area of the tactile stimulation testing. To reproduce our behavioral situation on the level of isolated CNS more adequately, we performed a series of electrophysiological experiments in which the LTF of EPSPs in parietal giant interneurons, induced by cutaneous nerve stimulation, was elicited by tetanization of the intestinal nerve which innervates caudal part of the snail's body (heterosynaptic plasticity). In pilot experiments we found that three tetanizations of intestinal nerve combined with 5-HT applications were insufficient to induce stable LTF in Pa2/Pa3 neurons, while five tetanizations induced facilitation lasting for several hours (**Figures 5A–C**). Similar to results obtained in experiments with homosynaptic facilitation, we have found that application of ZIP 90 min after the last tetanization of intestinal nerve reduced average amplitude of complex EPSPs induced by cutaneous nerve stimulation to the level of non-tetanized controls in the time window 210–240 min after the first tetanization ($39.4 \pm 4.3\%$ and $26.3 \pm 4.7\%$ to initial amplitude, accordingly). Under the scrZIP application, average amplitude of complex EPSPs elicited by test stimuli of cutaneous nerve 210 min after tetanizations was $101.4 \pm 8.0\%$ and was significantly larger than the amplitude of responses under ZIP application in the same time period ($p < 0.01$, **Figure 5C**). Therefore, the results obtained suggest that PKM ζ may participate in maintaining heterosynaptic LTF in terrestrial snail nervous system.



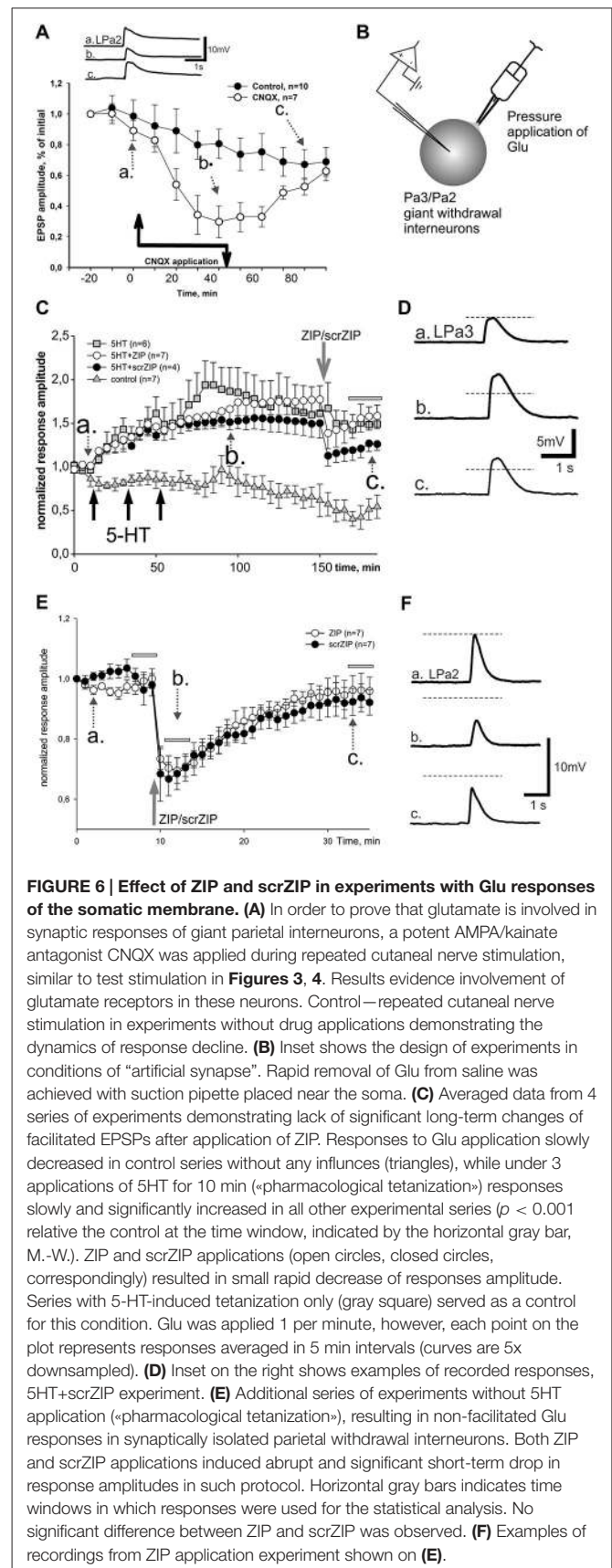
“Artificial Synapse” Experiments

As we showed earlier, primary mechanosensory neurons, presynaptic to giant withdrawal interneurons, send their processes to the second cutaneous nerve (Malyshev and Balaban, 2002) and use glutamate as a neurotransmitter (Bravarenko et al., 2003). Therefore it is very likely that complex EPSPs, induced by stimulation of cutaneous nerve in our current experiments, are mediated by glutamate, at least partially. To test this suggestion we performed series of experiments with pharmacological blockade of glutamatergic synaptic transmission. We have found that application of competitive AMPA/kainate receptor antagonist CNQX in a final bath concentration 5×10^{-6} M greatly reduced the amplitude of complex EPSPs induced by second cutaneous nerve stimulation (**Figure 6A**). Washout of CNQX showed slow restoration of responses to control levels. Obtained data (**Figure 6A**) demonstrate significant involvement of Glu in synaptic responses elicited by stimulation of second

cutaneous nerve. Similar results were obtained for the complex EPSPs induced by intestinal nerve stimulation (data not shown).

In order to test the involvement of postsynaptic mechanisms in the maintenance of LTF independently of the presynaptic ones, we designed experiments in which the transmitter was directly applied to somatic membrane of “synaptically isolated” (see Section Methods) giant withdrawal interneurons Pa2 or Pa3 (see inset on **Figure 6B**). Earlier it was shown in *Helix* that applications of serotonin in such an “artificial synapse” preparation lead to LTF of Glu-induced responses (Balaban et al., 2004). Averaged results demonstrate that in control experiments Glu applications at a 1/min rate elicit slow habituation of responses (**Figure 6C**, triangles; examples shown in **Figure 6D**). Three applications of serotonin in final bath concentration 10^{-5} M for 10 min, with a following 10 min washout led to a highly significant increase of Glu-PSPs (**Figure 6C**, squares). Adding ZIP (in final bath concentration $2 \mu\text{M}$) at the 90th min after the end of the last serotonin application resulted in an abrupt and transient decrease of the Glu responses amplitudes followed by a slow increase (**Figure 6C**, open circles). Finally, average amplitudes of responses, facilitated by serotonin application in the time window 170–185 min from the start of experiment were $147.8 \pm 9.3\%$ of baseline and did not differ significantly from the facilitated response amplitudes in the same time window under ZIP application ($155.2 \pm 7.3\%$). Surprisingly, we have found that application of scrambled ZIP in the experiments with serotonin-induced facilitation of Glu responses also produced a similar short-term drop in the response amplitudes (**Figure 6C**, filled circles).

We have performed additional series of experiments with non-facilitated Glu responses recorded in somata of parietal interneurons and found that both ZIP and scrZIP induced abrupt and significant (but short-term) drops in response amplitudes from 98.8 ± 1.6 – 70.2 ± 2.1 for ZIP and from 98.4 ± 2.2 – 68.6 ± 2.9 for scrambled ZIP (**Figures 6E,F**). In 20 min after ZIP or scrZIP applications, amplitudes of the responses were restored to $96.1 \pm 2.7\%$ and $92.8 \pm 2.5\%$, accordingly. As it was described in the Methods section, experiments with Glu-induced responses were carried out in “synaptically isolated” preparations, consisting of upper segments of parietal ganglia containing bodies of parietal withdrawal interneurons. It is possible that the transient inhibiting effect of ZIP/scrZIP application was an artifact of this type of preparation. In order to clarify this situation we carried out a special series of experiments on isolated CNS in the configuration which was used in the main series of our experiments with homo- and heterosynaptic facilitation but added repeated measurements of input resistance of the recorded cell and Glu application on the soma (see inset on **Figure 7**). We applied test pulses to the second cutaneous nerve eliciting complex EPSPs in withdrawal interneurons every 5 min. Between regular nerve stimulations we applied pulses of Glu to the soma of the same neuron every minute. We have found that applications of ZIP or scrambled ZIP in concentrations of $2 \mu\text{M}$ produced a transient drop in Glu-responses in parietal interneurons but no changes in the amplitude of complex EPSPs elicited in the same cell by the cutaneous nerve stimulation or any changes



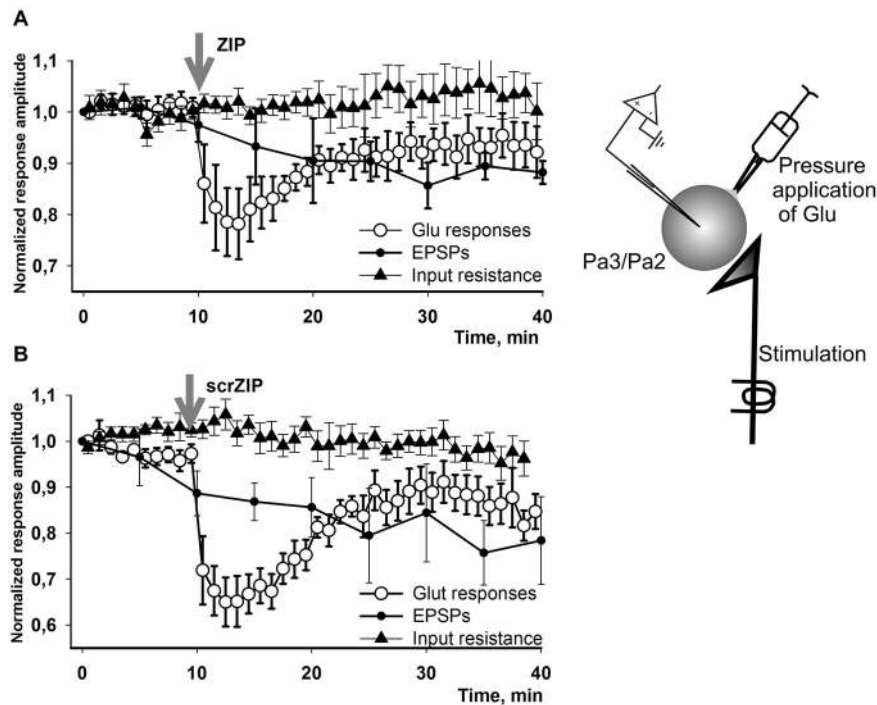


FIGURE 7 | Effect of ZIP and scrZIP in experiments with Glu responses of the somatic membrane in conditions of preserved synaptic inputs.

In a special series of experiments on isolated CNS in configuration which was used in the main series of our experiments with homo- and heterosynaptic facilitation (Figures 3, 4) we applied test pulses to the second cutaneous nerve eliciting EPSPs in parietal interneurons every 5 min. Between regular nerve stimulations we applied pulses of Glu to the soma of the same neuron every minute. Before each test stimulus the input resistance

of the recorded cell was measured. (A,B)—applications of ZIP or scrambled ZIP in concentrations 2 μ M produced a transient drop in Glu-responses in withdrawal interneurons that do not affect the EPSPs elicited by the cutaneous nerve stimulation in the same cell. No significant changes were observed in the input resistance throughout the experiment. EPSPs showed some habituation which is typical for this preparation. Note that the amplitudes of Glu responses in 20 min were restored almost to the level existing before ZIP/scrZIP application.

in input resistance (Figures 7A,B). Therefore, it appears that both peptides (ZIP and scrambled ZIP) have some transient non-specific effects on the responses induced by the application of glutamate on the somatic membrane of parietal withdrawal interneurons. Extrasynaptic glutamate receptors are presumably involved in these transient changes.

Discussion

Activity of PKM ζ Homolog Maintains Associative Long-term Memory in *Helix*

In our experiments it was shown that a homolog of PKM ζ exists in the neural transcriptome, and that the molecules immunoreactive to the conservative sites of PKM ζ are present in the neurons and neuropile of *Helix lucorum*. In behavioral experiments it was shown that associative contextual fear memory disappeared under blockade of PKM ζ by a selective inhibitor ZIP, but not under scrambled ZIP. If ZIP application was combined with a “reminder” (20 min in “noxious” context, no shocks), no impairment of the long-term context memory was observed. Testing of ZIP effectiveness at the time after reminding when memory starts to re-consolidate demonstrate that ZIP injection starts to be effective 2 h and more after the

reminding, which completely corresponds to the assumption that long-term memory reconsolidation starts about 2 h after the reminder and it is the time when the molecular substrate for ZIP is formed (Debiec et al., 2002; Duvarci et al., 2005). Obtained results suggest that blockade of PKM ζ impairs existing fear memory in terrestrial snails, while the “reminder” addresses this memory and starts the process of its reconsolidation even under transient (estimated time of ZIP presence in active state in the hemeolymph is not more than 4 h) blockade of PKM ζ synthesis. It was recently shown in *Helix* that 4 h after the reminder under protein synthesis blockade, the memory still exists, but is completely absent in the same animals 24 h later (Balaban et al., 2014). A recent study in marine mollusk *Aplysia* can help in explaining such a profile of protein-dependency of memory maintained by atypical PKM (Bougie et al., 2012). To activate cleavage of PKM Apl III isoform, activity-induced protein synthesis of calpain is required at the early phase of memory formation.

Activity of PKM ζ Homolog Maintains Long-term Facilitation in *Helix*

In electrophysiological experiments we tried to prove that PKM ζ takes part in maintenance of LTF in the neural circuit

underlying tentacle withdrawal. LTF of excitatory synaptic inputs to giant parietal premotor interneurons was induced by high-frequency stimulation combined with serotonin bath applications and lasted at least 4 h. We found that bath application of 2×10^{-6} M ZIP at 90th min after the tetanization reduced the EPSP amplitude to the non-tetanized EPSP values. Application of scrambled ZIP peptide at similar time and concentrations didn't affect the EPSP amplitudes. Our results largely correspond to published in *Aplysia* (Cai et al., 2011) effects of ZIP on LTF of synaptic connections between co-cultured sensory and motor neurons as well as to data obtained in rodents where ZIP disrupted persistence of LTP in hippocampus (Ling et al., 2002; Pastalkova et al., 2006). Therefore, obtained here results support the idea of atypical PKC involvement in post-induction maintenance of long-term plasticity in *Helix*.

PKM ζ Homolog is not Involved in LTF of “Artificial Synapse” in *Helix*

In order to test whether effects of ZIP can be reproduced in a model situation, we performed experiments with LTF of somatic membrane responses to local glutamate applications. It was shown earlier that serotonin applications in such “artificial synapse” conditions elicit LTF of responses (Balaban et al., 2004). In the present study it was shown that ZIP had no effect on LTF in these conditions, which may be explained by the very low concentration of PKM ζ molecules in somata of these identified neurons, as was evidenced by immunocytochemistry. Transient (minutes) decrease of the response to glutamate under ZIP and scrZIP is described for the first time and it may be of interest to investigate the mechanism of this phenomenon at the channel level.

The drugs were introduced via intrahemocoel injections and had access to the entire CNS. Therefore, although facilitation of synaptic inputs to the withdrawal interneurons in the parietal ganglia has been demonstrated to mediate long-term sensitization (Balaban, 2002), and although we have shown here that inhibition of PKM ζ disrupts maintenance of both long-term contextual aversive memory *in vivo* and LTF of the synaptic inputs to the parietal withdrawal interneurons *in vitro*, we cannot rule out the possibility that our behavioral results were attributable, at least in part, to actions of ZIP on central sites other than the synapses between sensory neurons and interneurons. The present results add to the accumulating evidence that PKM ζ plays crucial roles in the persistence of long-term memory and long-term synaptic plasticity in both vertebrates and invertebrates (Drier et al., 2002; Ling et al., 2002; Serrano et al., 2005; Pastalkova et al., 2006; Shema et al., 2007).

Possible Mechanism of PKM ζ -Mediated Maintenance of Synaptic Facilitation in *Helix*

How does PKM ζ maintain synaptic facilitation which we believe underlie associated long-term memory in *Helix*? According to T. Sacktor's model, the increase in synaptic strength in mammalian models of synaptic plasticity is based on PKM ζ -mediated increase in trafficking of AMPA receptors into the

postsynaptic membrane (Sacktor, 2012). As it was shown in *Aplysia*, mostly by data from D. Glanzman's laboratory, up-regulation of trafficking of AMPA receptors plays crucial role in the maintenance of both synaptic plasticity and behavioral learning in this mollusk (Chitwood et al., 2001; Li et al., 2005; Glanzman, 2010). Induction of associative learning as well as induction of synaptic facilitation in our experiments, similar to the situation in *Aplysia*, relies heavily on serotonin modulation of glutamatergic synapses. By application of selective blockers it was demonstrated that synaptic inputs to withdrawal interneurons, which are subject to change during induction of avoidance learning in terrestrial snails (Balaban, 2002), are mediated by glutamate (Bravarenko et al., 2003). Therefore, it is very likely that synaptic facilitation in our experiments is also mediated by constant increase of insertion of AMPA-like receptors into the postsynaptic membrane. We can further hypothesize that serotonin, which was used for induction of long-term synaptic facilitation in *Helix*, induced calpain-mediated cleavage of *Helix* ortholog of PKC ζ to form PKM ζ which underlies increased trafficking of AMPA receptors. Such a mechanism was recently reported for *Aplysia* (Bougie et al., 2009, 2012) and we believe it can also play a role in the mechanisms of synaptic plasticity in *Helix*. It should be noted here that *Aplysia* PKM ζ (and presumably in *Helix* as well) is not formed by alternative translation from mRNA of an atypical isoform of PKC as it happens in mammals (Hernandez et al., 2003), but rather, it is produced by calpain-dependent cleavage of PKC ζ ortholog (Bougie et al., 2009).

Specificity of ZIP and the Role of PKM ζ in Learning and Memory

Nowadays, much evidence suggests an involvement of PKM ζ in different types of synaptic plasticity and learning and memory in rodents, insects and mollusks (for review see, for example, Sacktor, 2012). Recently, in the cockroach it was shown that systemic injections of either chelerythrine or ZIP erase long-term olfactory memories, but have no effect on memory acquisition during conditioning (Deng et al., 2015). On the other hand, several other groups reported that activity of PKM ζ might not be necessary for synaptic plasticity. Thus, it was demonstrated that mice with a knockout for the PKM ζ gene exhibit normal hippocampal synaptic plasticity, learning and memory (Lee et al., 2013; Volk et al., 2013), suggesting existence of other targets for ZIP, presumably the isoforms of PKM ζ . It was also reported that scrambled ZIP with a changed amino-acid sequence can also block PKM ζ , although not as effectively as ZIP itself (Lee et al., 2013). In our experiments, scrZIP was not able to reverse potentiated EPSPs to the base level, and was also unable to erase memory in the learned snails. However, we observed similar transient reduction of glutamate responses, evoked by application of glutamate to the soma of neurons, both by ZIP and scrZIP, which suggests existence of some similar properties in these two peptides. Nevertheless, despite disagreement on the role of PKM ζ itself in processes of learning in memory, nobody argues that the short peptide ZIP can erase stored memories in a variety of experimental animals—from mollusks

to mammals. This fact, without any doubts, requires future investigations.

PKM ζ Expression in CNS

Using the sc-216 antibody, PKM ζ was detected previously in membranes, cytoplasm and in the nucleus of cultured non-nervous PC12 cells (Crisanti et al., 2005). In embryonic hippocampal neurons, both sc-216 and sc-17781 antibodies detected PKM ζ both in cell bodies and at non-axon-forming neurites (Parker et al., 2013). This pattern of staining coincides with our data. In adult rat brain, it was shown that PKM ζ is widely expressed in the forebrain including hippocampus and neocortex (Hernández et al., 2013). It was detected in the cell somata and dendrites, including postsynaptic densities. We suggest that these results match to the granules we have seen in cultured neurons of the snail. Thus, cell soma staining (granules) and absence of staining in axons seems to be a common case within different brains. Therefore, our data show similar patterns of PKM ζ immunoreactivity distribution in the

neurons of rodents and mollusk which suggests a common role for PKM ζ in neural functions in these two classes of animals.

Conclusion

We have demonstrated that inhibiting an isoform of PKM ζ in terrestrial snail eliminates the long-term memory, as well as the specific form of long-term synaptic plasticity that underlies the learning in the well-studied invertebrate *Helix*.

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