HUMAN ADENOVIRUS: VIRAL PATHOGEN WITH INCREASING IMPORTANCE

B. Ghebremedhin^{1,2,*}

¹ Faculty of Health, University of Witten/Herdecke, 58448 Witten, Germany

Received: December 12, 2013; Accepted: December 21, 2013

The aim of this review is to describe the biology of human adenovirus (HAdV), the clinical and epidemiological characteristics of adenoviral epidemic keratoconjunctivitis and to present a practical update on its diagnosis, treatment, and prophylaxis. There are two well-defined adenoviral keratoconjunctivitis clinical syndromes: epidemic keratoconjunctivitis (EKC) and pharyngoconjunctival fever (PCF), which are caused by different HAdV serotypes. The exact incidence of adenoviral conjunctivitis is still poorly known. However, cases are more frequent during warmer months. The virus is endemic in the general population, and frequently causes severe disease in immunocompromised patients, especially the pediatric patients. Contagion is possible through direct contact or fomites, and the virus is extremely resistant to different physical and chemical agents. The clinical signs or symptoms of conjunctival infection are similar to any other conjunctivitis, with a higher incidence of pseudomembranes. In the cornea, adenoviral infection may lead to keratitis nummularis. Diagnosis is mainly clinical, but its etiology can be confirmed using cell cultures, antigen detection, polymerase chain reaction or immunochromatography. Multiple treatments have been tried for this disease, but none of them seem to be completely effective. Prevention is the most reliable and recommended strategy to control this contagious infection.

Keywords: human adenovirus (HAdV), serogroups, epidemic keratoconjunctivitis (EKC), pharyngoconjunctival fever (PCF)

Introduction

Human adenoviruses (HAdVs) are classified in the genus Mastadenovirus, which contains seven known HAdV species HAdV-A to HAdV-G [1-3]. In 1953, adenoviruses were first isolated by Rowe and colleagues while studying the growth of polioviruses in adenoidal tissue [4]. During this study, a transmissible agent was identified which was capable of causing cytopathic changes in tissues in the absence of poliovirus. Traditionally, the HAdV species were classified by hemagglutination and serum neutralization reactions into different serotypes [1, 2]. Today, there have been reported 67 HAdV types. However, according to the genebank (http://hadvwg.gmu.edu/) [3] for human adenovirus genotype classification, HAdV type 68 - belonging to subgroup B – has been reported in 2012. The discovery and division of the HAdV types 52-68 are based on the genomic sequencing and bioinformatic analysis and different from the pre-existing 51 HAdV serotypes identified by traditional serological methods in composition and pathogenicity [1]. An additional serotype 52 constitutes a new HAdV species and has recently been reported after genomic sequencing and phylogenetic analysis of an isolate in the U.S. [5, 6]. New HAdV types have since been identified by several authors based on genomic data, including several emerging and recombinant viruses [7–14]. Most recently, a primate adenovirus from New World monkeys was detected which crossed the species barrier to infect humans [15, 16]. The majority of new HAdV types are homologous recombination within the same subgenus, and as a result, certain new serotypes acquire different pathogenicities. Recombination is a common evolutionary way for HAdV; however, the mechanism of recombination and the potential hazards to human beings remain unknown [1].

Adenoviruses can cause an array of clinical diseases, including conjunctivitis, gastroenteritis, hepatitis, myocarditis, and pneumonia (*Table 1*). Most of these occur in children younger than the age of 5 years and are generally self-limiting illnesses. They commonly present with mild gastrointestinal symptoms, respiratory symptoms,

² HELIOS Clinic Wuppertal, Institute of Medical Laboratory Diagnostics, 42283 Wuppertal, Germany

^{*}Corresponding author: Beniam Ghebremedhin, MD, MSc, University of Witten/Herdecke, Faculty of Health, Alfred-Herrhausen-Straße 50, 58448 Witten, HELIOS Clinic Wuppertal, Institute of Medical Laboratory Diagnostics, Heusnerstr. 40, 42283 Wuppertal, Germany; E-mail: bg.id@msn.com

Table 1. Adenovirus serotypes and associated clinical diseases [5–7, 17, 34–37]
--

HAvD subgroup	Serotype	Type of infection
A	12, 18, 31	gastrointestinal, respiratory, urinary
B, type 1	3, 7, 16, 21	keratoconjunctivitis, gastrointestinal, respiratory, urinary
B, type 2	11, 14, 34, 35	gastrointestinal, respiratory, urinary
C	1, 2, 5, 6	respiratory, gastrointestinal including hepatitis, urinary
D	8-10,13,15,17,19,20,22-30,32,33,36-39,42-49	keratoconjunctivitis, gastrointestinal
E	4	keratoconjunctivitis, respiratory
F	40, 41	gastrointestinal
G	52	gastrointestinal

or a combination of both. Globally, 5–7% of respiratory tract infections in pediatric patients are ascribed to HadV [17–19].

Some serotypes, such as HAdV type 14, have been associated with severe and potentially fatal outbreaks of pneumonia in residential facilities and military bases [20]. Of the seven HAdV subgroups (A to G), subgroup A has been associated with the gastrointestinal tract [21, 22], while subgroups B and C are more common to the respiratory tract [21], though conjunctivitis outbreaks have been associated with HAdV type 3 (subgroup B-1, *Table 1*). HAdV subgroup D overwhelmingly causes conjunctivitis [21, 23, 24], and subgroup E is associated with respiratory and ocular infections, but more commonly in conjunctivitis [21]. Finally, subgroups F and G are the causative agents of gastroenteritis [17, 21].

Adenovirus: biology, classification, and associated diseases

Adenoviruses are non-enveloped double-stranded DNA viruses that can infect a variety of human tissues. They range in size from 65 to 80 nm in diameter. The virion is composed of a protein capsid, made up of 252 capsomeres, and a nucleoprotein core that contains the DNA viral genome (26-46 kbp long, containing 23-46 protein-coding genes) and internal proteins. The capsid has an icosohedral shape, consisting of 240 hexon components and 12 pentons per virus particle [25, 26]. Each penton contains a base plate with fiber. The length of the fibers varies among the different serotypes [27]. DNA homology within the HAdV subgroups ranges from 48% to 99%. HAdV subgroup C serotypes revealed the highest DNA homology (up to 99%) as compared to other HAdV subgroups. However, the DNA homology between HAdV subgroups is less than 20% [17].

There is a varying degree of oncogenicity among the HAdV serotypes. For instance, HAdV subgroup A types, such as HAdV type 12, are highly oncogenic [28–30] with the ability to induce tumors in new born rodents within 4 months. On the other hand, HAdV subgroup B are weakly oncogenic, and finally HAdV subgroups C, E, and F are

not known to be oncogenic. However, it has been shown that the cells transformed by non-oncogenic HAdV can cause tumors in immunocompromised host, indicative of the host immune system rejecting those transformed by the non-oncogenic adenoviruses [31–33].

HAdV type 8 (HAdV-8) is a major causative agent of epidemic keratoconjunctivitis which is frequently associated with community, military, industrial, and nosocomial outbreaks. To date, HAdV-8A, B, E, and I have been found in Japan as variants of HAdV-8. In the recent time, two novel HAdV types causing nosocomial EKC were reported from Japan [38–40]. One of these has been misclassified as HAdV-8 in some cases because of its similarity to HAdV-8, according to neutralization test and phylogenetic analyses. However, the virus showed completely different restriction patterns from those of other published HAdV-8 genome types, revealing it is a novel serotype and is named as HAdV-54 [38].

Epidemic keratoconjunctivitis (EKC) is caused by a group of HAdVs of different serotypes that can also cause pharyngoconjunctival fever (PCF) and nonspecific follicular conjunctivitis. EKC is highly contagious, has a tendency to occur in epidemics, and has been reported worldwide such as epidemic outbreaks in hospitals, swimming pools, military bases, schools, and other community settings; in Asia, this disease is endemic [41–45]. In 1955, Jawetz et al. [25] were the first to attribute the etiology of EKC to the infection of ocular surface tissue caused by HAdV. EKC is one of the most common causes of acute conjunctivitis, with characteristic clinical features such as sudden onset of acute follicular conjunctivitis, with watery discharge, hyperemia, chemosis, and ipsilateral pre-auricular lymphadenopathy [46].

Epidemiology of epidemic keratoconjunctivitis (EKC)

EKC is one of the most frequent ocular diseases, exhibiting ubiquitous distribution. Due to its high frequency and that many of the cases do not obtain medical help, it is most difficult to acquire precise statistical data. The actual prevalence and incidence of EKC in countries, such as

28 B. Ghebremedhin

Germany, is well documented [41–44, 46–48]. EKC cases have to be reported to the medical authority in Germany. However, the prevalence and incidence of EKC at the international level are unknown. The infection is more common in adults, but all age groups can be affected with no specific gender affinity as has been reported by Adlhoch et al. [48]. Their analysis did not show a disproportionately affected gender or age group, but many infections were preceded by exposure to ophthalmological facilities, communal facilities, or public places. Seasonal patterns of infection have been demonstrated in the general pediatric population. This depends upon the HAdV serotypes and the population groups exposed. EKC is caused mainly by HAdV-3 (in HAdV-B), HAdV-4 (in HAdV-E), and HAdV-8, HAdV-19, and HAdV-37 (in HAdV-D). Among these, HAdV-8, HAdV-19, and HAdV-37 cause more severe conjunctivitis than the others [49]. HAdV-37 was first isolated from patients with keratoconjunctivitis in 1976 in The Netherlands and was reported by De Jong et al. as a newly identified serotype [27]. In northern Japan, HAdV-37 has been isolated every year since 1977 [50] and has caused large epidemics of conjunctivitis in the last two decades [49]. EKC outbreaks have been mainly associated to serotypes 8, 19, and 37, whereas PCF cases have been related more with serotypes 3, 7, and 11 [17, 36, 47–49, 51].

Symptomatology and clinical signs of epidemic keratoconjunctivitis (EKC)

EKC patients may complain about flu-like symptoms, including fever, malaise, respiratory symptoms, nausea, vomiting, diarrhoea, and myalgia. Recent history of ophthalmological examination or exposure within the family or at work has been documented in most cases. The incubation period is 2-14 days, and the person may remain infectious for 10–14 days after the onset of the symptoms. The ocular symptoms are mainly sudden onset of irritation, soreness, red eye, photophobia, foreign body sensation, and excessive tearing. More severe cases may include ocular and periorbital pain and decreased visual acuity. Symptoms tend to last for 7–21 days [17, 46, 49, 51, 52]. Ipsilateral pre-auricular lymphadenopathy is one of the classic findings of EKC. Decreased visual acuity is rarely present; it is usually present only if there is corneal involvement. Other clinical signs may include swelling and erythema of the lid, conjunctival hyperemia, chemosis, follicular reaction, mainly in the lower palpebral conjunctiva (the earliest and most common sign), papillary hypertrophy, or subconjunctival and petechial hemorrhage. In severe cases, membranous and pseudomembranous conjunctivitis can be seen in one [46, 52].

One of the distinguishing features of EKC is corneal involvement, which is usually mild and transient. Corneal involvement has been well documented 3–4 days after symptom onset as diffuse, fine epithelial keratitis that stains with fluorescein. This keratitis can persist for 2–3 weeks and in few cases corneal epithelial defect may occur. A week af-

ter the onset, focal epithelial keratitis develops with central ulceration and irregular borders with gray-white dots. These epithelial changes are related to active viral infection. These lesions persist for 1 to 2 weeks. About 2 weeks post-onset, subepithelial infiltrates can appear beneath the focal epithelial lesions, persisting for weeks to years. These resolve spontaneously, usually without scarring. In rare cases, disciform keratitis or anterior uveitis can occur. There is no change in corneal sensation [46, 52]. Bacterial superinfection is not frequent but could occur; mostly involved are gram-positive cocci such as *Streptococcus pyogenes*. Bacterial superinfection is particularly severe in pediatric patients and could lead to amblyopia [53].

Symptomatology and clinical signs of pharyngoconjunctival fever (PCF)

Pharyngoconjunctival fever (PCF) is a well-described syndrome attributed to HAdV subgroup B, particularly serotype 3 [53], which causes outbreaks, mainly among children. Outbreaks are frequent in schools, kindergartens, and summer camps. Acute onset of PCF comprises fever, pharyngitis, rhinitis, cervical adenopathies and bulbar and palpebral conjunctivitis with moderate follicular reaction which can last for 3–5 days. PCF associated ocular inflammation begins unilateral and generally becomes bilateral in the course of the disease. Bacterial superinfection and ocular complications are much less frequent than in EKC. The main sources of infection associated to PCF are contaminated waters of swimming pools and water reservoirs [46, 53, 54].

Immunology, transmission and spread of human adenovirus

Immune responses to adenovirus infection depend on various factors including the inoculation site, the HAdV serotype, and the antibody status of the host. HAdVs have several mechanisms for evading host immune responses. These include the inhibition of intrinsic cellular apoptosis in infected cells, inhibition of responses to interferon and tumor necrosis factor (TNF), and the prevention of MHC class I expression on cell surfaces [17–19, 55, 56].

Antibodies to adenovirus (mainly secretory IgA) are present in the upper respiratory tract within 3 days of infection. Approximately 7 days post-infection, antibodies can be detected in serum, and nasal secretions contain both secretory IgA and IgG [17, 18]. Antibodies are directed against the hexon (alpha component) of the viral capsid, which contains the antigenic component that is common to all mammalian adenoviruses [17, 19, 55, 56]. Typespecific neutralizing antibodies are produced in response to another hexon component as well as viral fibre antigens. These have been demonstrated 10 years after primary infection and might explain why re-infection with the same serotype is uncommon [17, 18].

There are three possible ways of HAdV interaction with the host cell:

- lytic infection (epithelial cells): the virus completes its multiplication cycle, producing cellular death and releasing new viruses, of which up to 5% are infectious;
- latent infection (lymphoid cells): only small amounts of virus are released, and the cellular death rate is offset by normal multiplication;
- oncogenic transformation: the HAdV DNA is included in the cellular genetic material and replicates inside it without producing new infectious virus.

Animal models of adenoviral pulmonary infection have shown histopathological changes that can be divided into two phases: the first with predominantly monocyte and macrophage infiltrates typical of non-specific, cytokine-mediated inflammation, whereas the second phase is characterized by predominantly T-lymphocyte infiltration [17, 57].

T-cell-mediated immunity seems to be of importance for recovery after acute HAdV infection, since immunocompromised individuals who lack effective cellular immunity are at a much higher risk of adenoviral infection. It has been suggested that both clinical and laboratory features of adenovirus infections are consistent with Th1 cell responses (production of TNF and interferon γ by T-cells) [17, 18]. Adenovirus-specific CD4+ lymphocytes can recognize conserved antigens across different adenovirus serotypes. Infection with one serotype can therefore produce T-cell-mediated immunity to infection with different serotypes. However, the neutralizing antibodies against HAdVs are serotype-specific [17-19]. The presence of adenovirus-specific CD4+ T-cells in most asymptomatic adults suggests that adenovirus-specific cellular responses are long lived [17, 58].

Diagnostics of epidemic keratoconjunctivitis (EKC)

The diagnosis of EKC is usually made on the basis of anamnesis and clinical findings. However, HAdV detection is traditionally performed by cell culture or antigen detection methods. Most HAdVs grow readily in cell culture and can be detected in various specimen types including nasopharyngeal, throat and conjunctival swabs, urine, cerebrospinal fluid (CSF), and stool. However, some types can take up to 4 weeks to isolate, and diarrheagenic types 40 and 41 of the subgroup F show difficulty to grow in routine cell culture. Direct antigen detection from clinical specimens may be used for acute infections, but provides markedly low sensitivity compared to culture. Less commonly, serology is used to detect a rise in adenovirus-specific antibodies following infection [2, 46, 59–61]. Due to the high mortality rates from invasive disease, individual adapted screening of urine stool and blood for adenovirus

in high-risk patients using PCR technology might present a sensible diagnostic approach to detect acute infection much earlier [2, 46, 61, 62].

Differential diagnosis should include the following pathological processes:

- allergic conjunctivitis: symptoms are generally similar in the early stages, but allergic conjunctivitis is more frequently bilateral and symmetrical; itching is the most characteristic symptoms of allergic conjunctivitis, whereas the foreign body feeling would indicate adenoviral origin,
- herpetic conjunctivitis: typically unilateral symptoms and more frequently associated with pain; could include bacterial superinfection with greater frequency than adenoviral conjunctivitis; herpetic conjunctivitis has a self-limited course of 8–9 days,
- Chlamydia inclusion conjunctivitis: exhibits larger follicle sizes, mainly in the inferior sac fundus; patients may refer *Chlamydia* genital-urinary infection history.

Differential diagnostic considerations

Besides EKC and PCF the following diagnoses should be considered in the differential diagnosis [17–19, 46] of:

- 1. acute follicular conjunctivitis:
 - acute trachoma
 - acute inclusion conjunctivitis
 - primary herpes simplex conjunctivitis
 - acute hemorrhagic conjunctivitis
 - infectious mononucleosis
 - neonatal inclusion conjunctivitis, etc.
- 2. subepithelial corneal opacities:
 - herpes simplex infection
 - herpes zoster infection
 - infectious mononucleosis
 - Epstein-Barr virus infection, etc.

Therapy of epidemic keratoconjunctivitis (EKC)

EKC is a self-limited disease that exhibits complete resolution within 3 weeks in most cases. To date, there is no causative therapy or specific drug against HAdV-associated diseases. EKC treatment is focused on managing patient symptoms and avoiding the appearance of complications while the patient immune system resolves the infection. However, the following treatments are available: some molecular compounds and cyctokines which are active against HAdV replication without adverse effects exhibited benefits: zalcitabine, sanilbudine, cidofovir, interferon beta, antiosteopontine peptide and N-chlorotaurine, although there is lack of randomized clinical trials of EKC [63–70].

30 B. Ghebremedhin

Animal studies have demonstrated that the use of topical corticosteroids could increase the replication rate of HAdV in the conjunctiva and prolong the duration of infection. However, the use of topical corticosteroids should be restricted to complicated cases with pseudomembranes or subepithelial infiltrates, in cases where visual acuity could be impaired significantly [63–67, 71, 72].

Randomized clinical trial in humans demonstrated that the use of topical cyclosporine A in combination with cidofovir did not significantly diminish the incidence of subepithelial infiltrates in comparison with cidofovir on its own. Topical cyclosporine A should only be considered in patients with subepithelial infiltrates that produce significant visual acuity loss and do not respond to topical corticoid therapy [73, 74]. One of the ophthalmic broad spectrum antiseptics is povidone-iodine. However, a clinical trial carried out in children did not find differences in the rate of healing between those treated with 1.25% povidone iodine or with antibiotic [75]. The use of topical antibiotics to prevent bacterial superinfection should be reserved for high-risk pediatric patients [73, 75].

Prevention and infection control/prophylaxis

As there is no efficient antiviral treatment against HAdV, prophylaxis is essential to control the infections caused by this pathogen. Washing of hands and disinfection of instruments do not appear to be sufficient to control the propagation of EKC outbreaks [44, 46, 76, 77]. A prospective study demonstrated that the application of a hospital infection control protocol against adenovirus can diminish the incidence of epidemic outbreaks and isolated adenoviral conjunctivitis cases through a four-year period [46, 78]. Recently, Dart et al. [79] demonstrated again that the application of an EKC identification and control protocol diminishes the incidence of hospital contagion.

Current and future utilities: therapeutic use of adenoviruses

Vector-based utility of adenovirus has been demonstrated for gene therapy and vaccine applications although some limitations due to adenovirus have to be overcome: modification of the virus capsid, of the ligands, administration routes, and the use of genome modified adenovirus or development of chimeric or alternative serotype adenovirus [80]. Oncolytic adenoviruses have been generated as vectors and present a new modality to treat cancer. In 2005, the first oncolytic adenovirus was approved for the treatment of head-and-neck cancer by the Chinese authority. The vectors demonstrated modest anti-tumor effect when applied as a single agent; their efficacy improved when they were combined with another modality, including vector design, delivery techniques, and supplemental therapy [81, 82].

Summary

EKC is an ocular surface infection produced by diverse HAdV serotypes, a DNA virus without envelope highly resistant to physical and chemical agents that is contaged through direct contact or fomites. The most frequent clinical syndromes are EKC and PCF. EKC gives rise to severe ocular surface inflammation, which can be complicated with the formation of pseudomembranes or subepithelial infiltrates caused by cellular immune reaction against virus antigen. The diagnosis is mainly clinical although the etiology can be confirmed by different diagnostic approaches, such as cell culture, antigen detection, and PCR. There is no efficient antiviral drug against HAdV. Therefore, symptomatic treatment is recommended with conservative measures and topical nonsteroid anti-inflammatory drugs. If complications arise, the use of topical corticoid therapy could be indicated. Prevention is crucial to control the propagation of this adenoviral infection.

Conflict of interest

No conflict of interest has been declared by the author.

References

- 1. Huang GH, Xu WB: Recent advance in new types of human adenovirus. Bing Du Xue Bao 29, 342–348 (2013)
- 2. Buckwalter SP, Teo R, Espy MJ, Sloan LM, Smith TF, Pritt BS: Real-time qualitative PCR for 57 human adenovirus types from multiple specimen sources. J Clin Microbiol 50, 766–771 (2012)
- Human Adenovirus Working Group. Human Adenovirus Genotype Classification. (http://hadvwg.gmu.edu/) (accessed Dec. 7th, 2013)
- Rowe WP, Huebner RJ, Gilmore LK, Parrot RH, Ward TG: Isolation of a cytopathogenic agent from human adenoids undergoing spontaneous degeneration in tissue culture. Proc Soc Exp Biol Med 84, 570–573 (1953)
- Jones MS 2nd, Harrach B, Ganac RD, Gozum MM, Dela Cruz WP, Riedel B, Pan C, Delwart EL, Schnurr DP: New adenovirus species found in a patient presenting with gastroenteritis. J. Virol 81, 5978–5984 (2007)
- Madisch I, Hofmayer S, Moritz C, Grintzalis A, Hainmueller J, Pring-Akerblom P, Heim A: Phylogenetic analysis and structural predictions of human adenovirus penton proteins as a basis for tissue-specific adenovirus vector design. J Virol 81, 8270–8281 (2007)
- Aoki K, Ishiko H, Konno T, Shimada Y, Hayashi A, Kaneko H, Ohguchi T, Tagawa Y, Ohno S, Yamazaki S: Epidemic keratoconjunctivitis due to the novel hexon-chimeric-intermediate 22,37/H8 human adenovirus. J Clin Microbiol 46, 3259–3269 (2008)
- 8. Kaneko H, Aoki K, Ishida S, Ohno S, Kitaichi N, et al.: Recombinant analysis of intermediate human adenovirus type 53 in Japan by complete genome sequence. J. Gen Virol 92, 1251–1259 (2011)
- Kaneko H, Aoki K, Ohno S, Ishiko H, Fujimoto T, Kikuchi M, et al.: Complete genome analysis of a novel intertypic

- recombinant human adenovirus causing epidemic keratoconjunctivitis in Japan. J Clin Microbiol 49, 484–490 (2011)
- Kaneko H, Suzutani T, Aoki K, Kitaichi N, Ishida S, et al.: Epidemiological and virological features of epidemic keratoconjunctivitis due to new human adenovirus type 54 in Japan. Br J Ophthalmol 95, 32–36 (2011)
- Robinson CM, Singh G, Henquell C, Walsh MP, Peigue-Lafeuille H, Seto D, et al.: Computational analysis and identification of an emergent human adenovirus pathogen implicated in a respiratory fatality. Virology 409, 141–147 (2011)
- Seto D, Chodosh J, Brister JR, Jones MS: Using the whole genome sequence to characterize and name human adenoviruses. J. Virol. 85, 5701–5702 (2011)
- 13. Walsh MP, Walsh MP, Chintakuntlawar A, Robinson CM, Madisch I, Harrach B, et al.: Evidence of molecular evolution driven by recombination events influencing tropism in a novel human adenovirus that causes epidemic keratoconjunctivitis. PLoS One 4, e5635 (2009)
- Walsh MP, Seto J, Jones MS, Chodosh J, Xu W, Seto D: Computational analysis identifies human adenovirus type 55 as a re-emergent acute respiratory disease pathogen. J Clin Microbiol 48, 991–993 (2010)
- 15. Kohl C, Vidovszky MZ, Mühldorfer K, Dabrowski PW, Radonić A, Nitsche A, et al.: Genome analysis of bat adenovirus 2: indications of interspecies transmission. J Virol 86, 1888–1892 (2012)
- Wevers D, Metzger S, Babweteera F, Bieberbach M, et al.: Novel adenoviruses in wild primates: a high level of genetic diversity and evidence of zoonotic transmissions. J Virol 85, 10774–10784 (2011)
- 17. Walls T, Shankar AG, Shingadia D: Adenovirus: an increasingly important pathogen in paediatric bone marrow transplant patients. Lancet Infect Dis 3, 79–86 (2003)
- 18. Singh-Naz N, Rodriguez W: Adenoviral infections in children. Adv Pediatr Infect Dis 11, 365–388 (1996)
- Mahr J, Gooding L: Immune evasion by adenoviruses. Immunol Rev 168, 121–130 (1999)
- Lewis PF, Schmidt MA, Lu X, Erdman DD, Campbell M, et al.: A community-based outbreak of severe respiratory illness caused by human adenovirus serotype 14. J Infect Dis 199, 1427–1434 (2009)
- 21. Chang SY, Lee CN, Lin PH, et al.: A community derived outbreak of adenovirus type 3 in children in Taiwan between 2004 and 2005. J Med Virol 80, 102–112 (2008)
- 22. Harrach B, Benkö M, Both G, Brown M, Davis A, Echavarría M, Hess M, Jones M, Kajon A, Lahmkuhl H, Mautner V, Mittal S, Wadell G: Family Adenoviridae. In: King A, Adams M, Carstens E, Lefkowitz E (2011): Virus taxonomy: classification and nomenclature of viruses. Ninth report of the International Committee on Taxonomy of Viruses. Elsevier, San Diego, CA, pp. 95–111
- Maranhao AG, Soares CC, Albuquerque MC, Santos N: Molecular epidemiology of adenovirus conjunctivitis in Rio de Janeiro, Brazil, between 2004 and 2007. Rev Inst Med Trop Sao Paulo 51, 227–229 (2009)
- Sambursky RP, Fram N, Cohen EJ: The prevalence of adenoviral conjunctivitis at the Wills Eye Hospital Emergency Room. Optometry 78, 236–239 (2007)
- 25. Jawetz E, Kimura S, Nicholas AN, Thygeson P, Hanna L: New type of APC virus from epidemic keratoconjunctivitis. Science 122, 1190–1191 (1955)

- 26. Swenson PD, Wadell G, Allard A, Hierholzer JC (2003): Adenoviruses. In: Manual of Clinical Microbiology, 8 ed, vol 2. ASM Press, Washington, DC
- 27. De Jong JC, Wermenbol AG, Verweij-Uijterwaal MW, Slaterus KW, Wertheim-Van Dillen P, Van Doornum GJ, et al.: Adenoviruses from human immunodeficiency virus-infected individuals, including two strains that represent new candidate serotypes Ad50 and Ad51 of species B1 and D, respectively. J Clin Microbiol 37, 3940–3945 (1999)
- 28. Trentin JJ, Yabe Y, Taylor G: The quest for human cancer viruses. Science 137, 835–841 (1962)
- Yabe Y, Samper L, Bryan E, Taylor G, Trentin JJ: Oncogenic effect of human adenovirus type 12 in mice. Science 143, 46–47 (1964)
- 30. Yabe Y, Trentin JJ, Taylor G: Cancer induction in hamsters by human type 12 adenovirus. Effect of age and of virus dose. Proc Soc Exp Biol Med 111, 343–344 (1962)
- 31. Gallimore PH: Tumour production in immunosuppressed rats with cells transformed in vitro by adenovirus type 2. J Gen Virol 16, 99–102 (1972)
- 32. Gallimore PH, McDougall JK, Chen LB: In vitro traits of adenovirus-transformed cell lines and their relevance to tumorigenicity in nude mice. Cell 10, 669–678 (1977)
- 33. Van der Eb AJ, Mulder C, Graham FL, Houweling A: Transformation with specific fragments of adenovirus DNAs. I. Isolation of specific fragments with transforming activity of adenovirus 2 and 5 DNA. Gene 2, 115–132 (1977)
- 34. Feigin R, Cherry J (1998): Textbook of paediatric infectious diseases. 4th ed: Philadelphia: WB Saunders Company
- 35. Hierholzer J: Adenovirus in the immunocompromised host. Clin Microbiol Rev 5, 262–274 (1992)
- 36. Erdman D, Xu W, Gerber S, et al.: Molecular epidemiology of adenovirus type 7 in the United States, 1966–2000. Emerg Infect Dis 8, 269–277 (2002)
- 37. Meyer-Rüsenberg B, Loderstädt U, Richard G, Kaulfers PM, Gesser C: Epidemic Keratoconjunctivitis the current situation and recommendations for prevention and treatment. Dtsch Ärztebl Int 108, 475–480 (2011)
- 38. Ishiko H, Shimada Y, Konno T, Hayashi A, Ohguchi T, Tagawa Y, Aoki K, Ohno S, Yamazaki S: Novel human adenovirus causing nosocomial epidemic keratoconjunctivitis. J Clin Microbiol 46, 2002–2008 (2008)
- 39. Kaneko H, Iida T, Ishiko H, Ohguchi T, Ariga T, Tagawa Y, Aoki K, Ohno S, Suzutani T: Analysis of the complete genome sequence of epidemic keratoconjunctivitis-related human adenovirus type 8, 19, 37 and a novel serotype. J Gen Virol 90, 1471–1476 (2009)
- Aoki K, Ishiko H, Konno T, Shimada Y, Hayashi A, Kaneko H, Ohguchi T, Tagawa Y, Ohno S, Yamazaki S: Epidemic keratoconjunctivitis due to the novel hexon-chimeric-intermediate 22,37/H8 human adenovirus. J Clin Microbiol 46, 3259–3269 (2008)
- 41. Martone WJ, Hierholzer JC, Keenlyside RA, Fraser DW, D'Angelo LJ, Winkler KG: An outbreak of adenovirus type 3 disease at a private recreation center swimming pool. Am J Epidemiol 111, 229–327 (1980)
- D'Angelo LJ, Hierholzer JC, Keenlyside RA, Anderson LJ, Martone WJ: Pharyngoconjunctival fever caused by adenovirus type 4: Report of a swimming pool-related outbreak with recovery of virus from pool water. J Infect Dis 140, 42–47 (1979)

32 B. Ghebremedhin

43. Guyer B, O'Day DM, Hierholzer JC, Schaffner W: Epidemic keratoconjunctivitis: a community outbreak of mixed adenovirus type 8 and type 19 infection. J Infect Dis 132, 142–150 (1975)

- Keenlyside RA, Hierholzer JC, D'Angelo LJ: Keratoconjunctivitis associated with adenovirus type 37: an extended outbreak in an ophthalmologist office. J Infect Dis 147, 191–198 (1983)
- 45. Hillenkamp J, Reinhard T, Ross RS, et al.: Topical treatment of acute adenoviral keratoconjunctivitis with 0.2% cidofovir and 1% cyclosporine: a controlled clinical pilot study. Arch Ophthalmol 119, 1487–1491 (2001)
- González-López JJ, Morcillo-Laiz R, Muñoz-Negrete FJ: Adenoviral keratoconjunctivitis: an update. Arch Soc Esp Oftalmol 88, 108–115 (2013)
- Schrauder A, Altmann D, Laude G, Claus H, Wegner K, Köhler R, Habicht-Thomas H, Krause G: Epidemic conjunctivitis in Germany, 2004. Euro Surveill 11, 185–187 (2006)
- 48. Adlhoch C, Schöneberg I, Fell G, Brandau D, Benzler J: Increasing case numbers of adenovirus conjunctivitis in Germany, 2010. Euro Surveill. 15(45). pii: 19707 (2010)
- 49. Ariga T, Shimada Y, Shiratori K, Ohgami K, et al.: Five new genome types of adenovirus type 37 caused epidemic keratoconjunctivitis in Sapporo, Japan, for more than 10 years. J Clin Microbiol 43, 726–732 (2005)
- Aoki K, Kanazono N, Ishi K, Kato K, Ohtsuka H: Clinicoepidemiological study of keratoconjunctivitis due to adenovirus type 37 (Ad 37) in Sapporo, Japan. Nihon Ganka Gakkai Zasshi 89, 294–298 (1985)
- Yamadera S, Yamashita K, Akatsuka M, Kato N, Tokunaga M, Sakae I: Adenovirus type 7 outbreaks in Japan in 1998. Jpn J Infect Dis 53, 22–23 (2000)
- Kimura R, Migita H, Kadonosono K, Uchio E: Is it possible to detect the presence of adenovirus in conjunctiva before the onset of conjunctivitis? Acta Ophthalmol 87, 44–47 (2009)
- Kuo SC, Shen SC, Chang SW, Huang SC, Hsiao CH: Corneal superinfection in acute viral conjunctivitis in young children. J Pediatr Ophthalmol Strabismus 45, 374–376 (2008)
- Aronson B, Aronson S, Sobel G, Walker D: Pharyngoconjunctival fever; report of an epidemic outbreak. AMA J Dis Child 92, 596–612 (1956)
- 55. Wold W, Hermiston T, Tollefson A: Adenoviral proteins that subvert host defences. Trends Microbiol 2, 437–443 (1994)
- 56. Gooding LR, Wold WS: Molecular mechanisms by which adenoviruses counteract antiviral immune mechanisms. Crit Rev Immunol 10, 53–71 (1990)
- Prince GA, Porter DD, Jenson AB, Horswood RL, Chanock RM, Ginsberg HS: Pathogenesis of adenovirus type 5 pneumonia in cotton rats (Sigmodon hispidus). J Virol 67, 101–111 (1993)
- 58. Flomenberg P, Piaskowski V, Truitt R, Casper J: Characterisation of human proliferative T cell responses to adenovirus. J Infect Dis 171, 1090–1096 (1995)
- Sambursky R, Tauber S, Schirra F, Kozich K, Davidson R, Cohen EJ: The RPS adeno detector for diagnosing adenoviral conjunctivitis. Ophthalmology 113, 1758–1764 (2006)
- 60. Hiroi S, Izumi M, Takahashi K, Morikawa S, Kase T: Isolation and characterization of a novel recombinant human

- adenovirus species D. J Med Microbiol 61, 1097-1102 (2012)
- Adhikary AK, Ushijima H, Fujimoto T: Human adenovirus type 8 genome typing. J Med Microbiol 61, 1491–1503 (2012)
- 62. Heim A, Ebnet C, Harste G, Pring-Akerblom P: Rapid and quantitative detection of human adenovirus DNA by real-time PCR. J Med Virol 70, 228–239 (2003)
- 63. Romanowski EG, Roba LA, Wiley L, Araullo-Cruz T, Gordon YJ: The effects of corticosteroids of adenoviral replication. Arch Ophthalmol 114, 581–585 (1996)
- Romanowski EG, Yates KA, Gordon YJ: Short-term treatment with a potent topical corticosteroid of an acute ocular adenoviral infection in the New Zealand white rabbit. Cornea 20, 657–660 (2001)
- 65. Romanowski EG, Yates KA, Gordon YJ: Antiviral prophylaxis with twice daily topical cidofovir protects against challenge in the adenovirus type 5/New Zealand rabbit ocular model. Antiviral Res 52, 275–280 (2001)
- Romanowski EG, Yates KA, Gordon YJ: Topical corticosteroids of limited potency promote adenovirus replication in the Ad5/NZW rabbit ocular model. Cornea 21, 289–291 (2002)
- 67. Romanowski EG, Yates KA, Gordon YJ: The in vitro and in vivo evaluation of ddC as a topical antiviral for ocular adenovirus infections. Invest Ophthalmol Vis Sci 50, 5295–5299 (2009)
- 68. Hillenkamp J, Reinhard T, Ross RS, Bohringer D, Cartsburg O, Roggendorf M, et al.: The effects of cidofovir 1% with and without cyclosporin a 1% as a topical treatment of acute adenoviral keratoconjunctivitis: a controlled clinical pilot study. Ophthalmology 109, 845–850 (2002)
- 69. Teuchner B, Nagl M, Schidlbauer A, Ishiko H, Dragosits E, Ulmer H, et al.: Tolerability and efficacy of N-chlorotaurine in epidemic keratoconjunctivitis a double-blind, randomized, phase-2 clinical trial. J Ocul Pharmacol Ther 21, 157–165 (2005)
- 70. Sundmacher R, Wigand R, Cantell K: The value of exogenous interferon in adenovirus keratoconjunctivitis. Graefes Arch Clin Exp Ophthalmol 218, 139–140 (1982)
- 71. Maĭchuk IuF, Iani EV: Locferon: new eyedrops for treatment eye adenoviral diseases. Vestn Oftalmol 115, 32–33 (1999)
- 72. Yamazaki ES, Ferraz CA, Hazarbassanov RM, Allemann N, Campos M: Phototherapeutic keratectomy for the treatment of corneal opacities after epidemic keratoconjunctivitis. Am J Ophthalmol 151, 35–43.e1 (2011)
- Pelletier JS, Stewart K, Trattler W, Ritterband DC, Braverman S, Samson CM, et al.: A combination povidone-iodine 0.4%/dexamethasone 0.1% ophthalmic suspension in the treatment of adenoviral conjunctivitis. Adv Ther 26, 776–783 (2009)
- 74. Levinger E, Slomovic A, Sansanayudh W, Bahar I, Slomovic AR: Topical treatment with 1% cyclosporine for subepithelial infiltrates secondary to adenoviral keratoconjunctivitis. Cornea 29, 638–640 (2010)
- Isenberg SJ, Apt L, Valenton M, Del Signore M, Cubillan L, Labrador MA, et al.: A controlled trial of povidone-io-dine to treat infectious conjunctivitis in children. Am J Ophthalmol 134, 681–688 (2002)
- 76. Ford E, Nelson KE, Warren D: Epidemiology of epidemic keratoconjunctivitis. Epidemiol Rev 9, 244–261 (1987)

- 77. Nauheim RC, Romanowski EG, Araullo-Cruz T, Kowalski RP, Turgeon PW, Stopak SS, et al.: Prolonged recovery of desiccated adenovirus 19 from various surfaces. Ophthalmology 97, 1450–1453 (1990)
- 78. Gottsch JD: Surveillance and control of epidemic keratoconjunctivitis. Trans Am Ophthalmol Soc 94, 539–587 (1996)
- 79. Dart JKG, El-Amir AN, Maddison T, Desai P, Verma S, Hughes A, et al.: Identification and control of nosocomial adenovirus keratoconjunctivitis in an ophthalmic department. Br J Ophthalmol 93, 18–20 (2009)
- 80. Seregin SS, Amalfitano A: Improving adenovirus based gene transfer: strategies to accomplish immune evasion. Viruses 2, 2013–2036 (2010)
- 81. Ghosh SS, Gopinath P, Ramesh A: Adenoviral vectors: a promising tool for gene therapy. Appl Biochem Biotechnol 133, 9–29 (2006)
- 82. Toth K, Wold WSM: Increasing the efficacy of oncolytic adenovirus vectors. Viruses 2, 1844–1866 (2010)