

Review Article

Human microbiome and its association with health and diseases[†]

Running title: Human microbiome and diseases

Asmaa Althani,^{1,2} Hany E. Marei,^{1,3*} Wedad S. Hamdi,¹ Gheyath K. Nasrallah,² Mohamed E. El Zowalaty,¹ Souhaila Al Khdor,⁴ Maha Al-Asmakh,² and Hassan Abdel-Aziz²

¹Biomedical Research Center, ²Department of Health Sciences, College of Arts and Science, Qatar University, Doha, 2713, Qatar, ³Department of Cytology and Histology, Faculty of Veterinary Medicine, Mansoura University, Mansoura 35116, Egypt, ⁴Infectious Diseases and Vaccine Development, Sidra Medical and Research Center, Qatar

*Corresponding author: Hany Marei, Biomedical Research Center, Qatar University, P.B. Box 2713, Qatar University, Doha, Qatar, Phone: (+ 974) 4403-6817, Email: hmady@qu.edu.qa

[†] This article has been accepted for publication and undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the Version of Record. Please cite this article as doi: [10.1002/jcp.25284]

Received 16 October 2015; Revised 16 November 2015; Accepted 9 December 2015
Journal of Cellular Physiology
This article is protected by copyright. All rights reserved
DOI 10.1002/jcp.25284

Abstract

Human microbiota are distinct communities of microorganisms that reside at different body niches. Exploration of the human microbiome has become a reality due to the availability of powerful metagenomics and metatranscriptomic analysis technologies. Recent advances in sequencing and bioinformatics over the past decade help provide a deep insight into the nature of the host-microbial interactions and identification of potential driver genes and pathways associated with human health, well-being, and predisposition to different diseases.

In the present review, we outline recent studies devoted to elucidate the possible link between the microbiota and various types of diseases. The present review also highlights the potential utilization of microbiota as a potential therapeutic option to treat a wide array of human diseases.

This article is protected by copyright. All rights reserved

Keywords Human microbiome; host-microbial interactions, dysbiosis, cancer, metabolism; obesity; neurodegenerative, gut microbiome; global health; prebiotics, probiotics, synbiotics; diseases, health, virome

Introduction

Humans are viewed as composites of human and microbial cells. Human microbiota are complex and dynamic microbial communities composed mainly of bacteria, but also includes protozoa, archaea, viruses, and fungi that reside in and on different body niches such as oral cavity, throat, esophagus, stomach, colon, urogenital tract, respiratory tract, and skin¹. The number of microbial cells inhabiting human body is estimated to exceed the *H.sapiens* cells by 10-fold and estimated at 350 trillion microbial cells². The colonic microbiota constitutes the most abundant microbial domain within the human body with the vast majority belonging to bacterial phyla; *Firmicutes* and *Bacteroidetes*³. The collective genomes of the complete human microbiota located at the different body sites are referred to as human microbiome⁴. Metagenome and metatranscriptome refers to the study of collective genes and RNA derived from a specific microbiome respectively. The term virome is used to describe the viral components (bacterial, archeal, eukaryotic virome, and virus-derived elements) while mycobiome refers to fungal organisms within human microbiota⁵⁻⁸. As shown in figure (1), the human microbiome is composed of complex communities of viral (virome), bacterial (microbiota) and fungal (mycobiota) and their associated genetic material. The interplay among human microbiome and host cells affects human health and contribute in the pathogenesis of various diseases.

Marked differences in the abundance and diversity of microbiota are observed in healthy human individuals along with the presence of a strong niche specialization both within and among individuals⁹. These differences may be explained due to a number of factors including differences in host genetics, feeding habits, life style, and early life microbial exposure¹⁰⁻¹². Moreover, changes in the composition of human microbiota have profound impacts on health and may predispose to different immunological and pathological conditions^{9,13,14}. These microbial

communities have a profound impact on human health and well-being, and each person's microbiome is thought to be unique. Differences in the microbiome composition can help explain why some people are more susceptible or resistant to certain diseases.

Exploration of the human microbiome has recently become possible due to the availability of powerful metagenomics and metatranscriptomic analysis protocols. Using these protocols, it become possible to have a better insight on the nature of the host-microbial interactions, and to identify potential driver genes, and pathways associated with human health and diseases.

This review highlights the recent advances in human microbiome and explores possible uses of different microbial genetic signatures in identifying possible disease risks. The exploration of novel therapeutic targets to improve human health, well-being, and to treat various diseases associated with microbiome dysbiosis are also mentioned.

Role of human microbiome in health

Accumulating evidence reveals that the gut microbiota plays a major role in promoting health, as a result of which it is often referred to as the 'forgotten organ'¹⁵. The relationship between the host and microbiota is symbiotic and mutualistic, each deriving benefits from the other. These two terms are similar but mutualism is defined as 'an interaction between species that is beneficial to both of them' and symbiosis as 'the living together of two organisms in close association'¹⁶. While the host provides the microbiota with a protected and nutrient-rich environment, the microbiota enhance, e.g., digestion, immunity and neuronal development.

Microbiota are key to maintaining homeostasis where it confers many benefits for the host such as pathogen displacement, development of the immune system, vitamin production and absorption of nutrients¹⁷. The influence of microbiota on health extends beyond the GI tract affecting almost every organ of the body^{18,19}. In the intestine, microbiota affect angiogenesis²⁰

and improve gut immunity and motility, as well as decreasing the permeability of the intestinal barrier. In distant organ such as the lungs, microbiota regulate immunological defense against viral infection²¹. Microbiota also influence behavior by reducing synaptic connectivity and elevating anxiety^{22,19} and perception of pain²³. In the liver, microbiota modulate hepatic metabolism in such a way as to decrease energy expenditure and promote adiposity²⁴. In addition, absence of gut microbiota leads to more bone mass in association with fewer osteoclasts surface area of bone²⁵. Recent studies also showed that microbiota are involved in the development of personalized medicine²⁶, in xenobiotic metabolism²⁷ and in regulating blood-tissue barriers²⁸⁻³⁰.

Human microbiome and diseases (Dysbiosis)

Alterations in the composition of microbiota can result from exposure to various environmental factors such as diet, xenobiotics, drugs, and pathogens as shown in figure (2), which eventually contribute to the pathogenesis of various metabolic, neurological, immunological, and cancer promoting diseases. The collective microbiota of the gut whose DNA contributes to the metagenome have links with inflammatory bowel disease (IBD), liver disorders, ankylosing spondylitis, neurodegenerative diseases, obesity and associated noncommunicable diseases (NCDs) including diabetes mellitus, hypertension, atherosclerosis, coronary heart disease, and neurodegenerative diseases beside other condition³¹⁻³⁵.

Intestinal Diseases

Due to the direct contact between the intestine and microbiota, it is predictable that alteration in the composition of microbiota could be involved in the pathogenesis of many intestinal disease such as Crohn's disease (CD) and ulcerative colitis (figure 3).

In CD, metagenomic analysis revealed a decrease in *Firmicutes*, in particular *F. prausnitzii*, and an increase in *Enterobacteriaceae*, especially the virulent invasive *E. coli*³⁶. Alteration of gut microbiota may affect mucosal health and immune system by acting on the epithelial barrier function, and regulation of the innate immune system³⁷. Reduction in the number of *F. prausnitzii* is associated with an increasing risk for the recurrence of ileal CD³⁸, another study confirmed an increase in the number of *F. prausnitzii* in pediatric CD³⁹. Ott and colleagues demonstrated that CD was associated with altered fungal profile with a marked increase in the diversity of fungal community. In pediatric inflammatory bowel disease (IBD), dominance of *Basidiomycota* species was recorded⁴⁰.

Viruses associated with gut bacteria may affect the pathogenesis of CD and disease-specific viromes had been related to CD and ulcerative colitis (UC)⁴¹. Previous studies demonstrated a significant increase of *Caudovirales* bacteriophages concomitant with a reduction in the relative abundance of bacterial species, indicating a possible involvement of virome in bacterial dysbiosis associated with CD and UC⁴².

Alterations in the homeostasis of gut microbiota may induce low-grade intestinal inflammation associated with irritable bowel syndrome (IBS)^{43,44}. IBS was associated with an increase in the numbers of *Ruminococcus*, *Clostridium*, and *Dorea*, with a marked reduction in *Bifidobacterium* and *Faecalibacterium* spp.⁴⁵. In comparison to normal population, an increase in the *Firmicutes* to *Bacteroidetes* ratio was evident in patients with IBS⁴⁶. Dysbiosis of the gut microbiota is thought to play a crucial role in the development of mucosal lesions⁴⁷ and intestinal inflammation is generally believed to be associated with a specific reduction in the *Bacteroidetes* and *Firmicutes* phyla specially reductions in the *Clostridium leptum* and *Clostridium coccoides*

groups⁴⁸. All of the aforementioned studies have certainly outlined a link between the gut microbiota and IBD.

Association between colorectal cancer (CRC) and the presence of specific causative organism has been suggested. For instance, the presence of high numbers of *Fusobacterium* in the gut microbiota has been linked to CRC^{49,50}. Interestingly, members of *Fusobacterium* has been associated with IBS⁵¹. The initiation of chronic inflammatory condition due to dysbiosis of gut microbiota lead to impairment of intestinal barrier, induction of inflammation through a host immune response, and in turn, increase in tumor growth⁵².

E. coli had been implicated in the initiation of CRC through polyketide synthase (pks) and mice mutants lacking the pks island had a decreased tumor growth and invasion compared to their wild-type pks+ counterparts⁴⁹. Although there were strong correlations between inflammation induced by the presence of specific types of microorganisms and CRC, it was clear that further investigations were required to further explore the role of bacterial induced inflammation in tumorigenesis and CRC.

Gastric diseases

Although gastric pH, peristalsis and mucus layer play an essential protective role in preventing bacterial colonization in the stomach, maintenance of gastric microbiota homeostasis is essential for the stomach health. Five major phyla have been detected in the stomach including *Bacteroidites*, *Actinobacteria*, *Fusobacteria* and *Proteobacteria*. Healthy human stomach is dominated by *Prevotella*, *Streptococcus*, *Veillonella*, *Rothia*, and *Haemophilus*; however, the composition of the gastric microbiota is dynamic and is affected by diet, drugs, and diseases⁵³.

Sequencing of the small subunit 16S rRNA revealed that *Helicobacter pylori* was the predominant phylotype in the stomach of chronic gastritis patients⁵⁴. The interaction between *H.*

pylori and other species within the gut microbiota might increase the risk of gastric cancer. Other bacterial genera such as *Streptococcus*, *Prevotella*, *Neisseria*, *Haemophilus* and *Porphyromonas* may also contribute to the development of gastritis. In this regard, the high number of *Streptococcus* genus was associated with antral gastritis⁵⁵.

Modifications of gastric microbiota have been associated with gastric cancer or precancerous conditions. Although gastric cancer is known to be a multifactorial disease, *H. pylori* infection was found to actively contribute to its progression, probably by induction of chronic atrophic gastritis leading to reduction of gastric acid secretion and initiation of inflammatory cytokines^{56,57}.

The exact role of microbiota in the origination of gastric cancer is not clear and is poorly understood. Using culture-based protocol, a comparatively large number of anaerobic bacteria such as *Clostridium* and *Bacteroides* was identified in patients with gastric cancer. Elevation of the pH in the gastric lumen due to reduction of acid-secreting cell number may influence colonization of microbiota within the gastric mucosa⁵⁸. Subsequent studies have cast doubts on the existence of significant differences in gastric microbiota between control and gastric cancer patients, and that the microbiota in gastric cancer patients was dominated by different species of the genera *Streptococcus*, *Lactobacillus*, *Veillonella*, and *Prevotella*⁵⁷.

Liver diseases

Disturbance to the gut microbiota as a result of extrinsic factors such as unbalanced diet and alcohol consumption had been reported to contribute to nonalcoholic fatty liver disease (NAFLD), steatohepatitis, alcoholic liver disease, and cirrhosis. Identifying specific microbial alterations associated with different liver diseases could improve our understanding of the role of

microbiota in the development of liver diseases, and hence could lead to the discovery of novel fecal biomarkers.

Nonalcoholic steatohepatitis (NASH) is characterized by the development of liver inflammation and fibrosis. Microbiota samples from NASH patients often yield a reduced number of *Ruminococcaceae* and a significantly higher percentage of *Clostridium coccooides*⁵⁹. The proportion of *Bacteroidaceae* was lower in samples from alcoholic patients than from nonalcoholic individuals⁶⁰. Aerobic and anaerobic bacterial cultures of jejunal aspirates from patients who chronically abuse alcohol were found to be associated with dysbiosis of jejunal microflora⁶¹.

At the preclinical level, ethanol intake in rats was associated with dysbiosis, overgrowth of bacteria along almost the entire gastrointestinal tract, and significant reductions in proportions of probiotic bacteria; *Lactobacillus*, *Pediococcus*, *Leuconostoc*, and *Lactococcus*⁶². The use of probiotics in patients of alcohol-induced liver injury lead to a marked improvement in liver functions⁶³. Modulation of the gut microbiome in response to alcohol intake might be supported by various factors including inhibition of intestinal motility, alterations in acid secretion, and modulation of the intestinal immune response^{64,65}.

Extensive research has been devoted to explain the phylogenetic analysis of gut microbiota associated with liver cirrhosis⁶⁶⁻⁷¹. Liver cirrhosis was associated with reduction in the number of beneficial bacteria with a concomitant increase of pathogenic microorganisms. Members of the *Prevotellaceae* family have significantly increased in patients with alcoholic cirrhosis compared with healthy individual⁶⁸. Factors contributing to modulation of intestinal microbiome include impaired motility of the small intestine, reduced bile flow, altered secretion of immunoglobulin A, and antimicrobial molecules⁷².

There has been a reciprocal interplay between gut microbiota and liver, where alcohol-induced liver diseases were reported to impair intestinal barrier by increasing systemic levels of IL1 β or tumor necrosis factor (TNF α), which disrupt tight junctions⁷³. Increasing intestinal leakage might facilitate the movement of microbial products from the lumen of the gut into other distal organs including the liver⁷³. In this respect, reduction of Gram-negative bacteria in the intestine due to the use of antibiotics has been associated with decrease in the levels of endotoxins and protection against liver disease after ethanol consumption⁷⁴.

The dysbiosis of gut microbiota lead to increase in the level of endotoxin and production of ammonia which has been implicated in the development of hepatic encephalopathy associated with liver cirrhosis⁷⁵. Interestingly, the number of bacterial species members of *Enterobacteriaceae* including *E. coli*, *Klebsiella*, *Proteus*, and *Enterobacter* surge in the microbiota of patients with cirrhosis⁶⁷.

Metabolic disorders

The gut microbiota plays important roles in modulating host metabolism, extraction of energy from ingested food, and synthesis of various metabolites and vitamins. Gut microbiota are also essential in the modulation of lipid absorption and deposition, polysaccharide content and the production of short-chain fatty acids which have a marked impact on food intake, inflammatory tone, or insulin signaling. Recent findings suggested that an altered gut microbial composition was associated with metabolic diseases including obesity, diabetes, or non-alcoholic fatty liver disease (figure 3)⁷⁶.

Changes to the gut microbiota play a critical role in the pathogenesis of obesity and diabetes. In humans it has been shown that gut microbiota composition differs between obese and lean subjects. Remarkably, inoculation of germ-free (GF) animals with gut microbiota derived from

obese controls significantly increase the deposition of fat, and was associated with increase in the insulin resistance⁷⁷. Leptin-deficient ob/ob obese mice displayed an alteration in the gut microbiota represented by a decrease in *Bacteroidetes* and a corresponding increase in *Firmicutes*⁷⁸.

Recent metagenomics studies showed the presence of reduced numbers of butyrate-producing *Clostridiales* and greater numbers of non-butyrate-producing ones in type 2 diabetes (T2DM), suggesting a protective role of butyrate-producing bacteria against T2DM^{79,80}. Furthermore, disruption of the gut barrier and microbiota-derived endotoxemia may contribute to the pathophysiology of T2DM and obesity. In this regard, modulation of the gut microbiota with antibiotics or prebiotics reduces the metabolic endotoxemia, decreases inflammatory markers, enhances gut permeability, and alleviates glucose intolerance. Moreover, the microbiome signature may act as an early diagnostic marker for T2DM, and may provide a novel therapeutic target against T2DM and obesity⁸¹⁻⁸³.

Lately, fecal microbiota transplantation (FMT) was reported to be highly successful therapeutic approach for the treatment of recurrent *Clostridium difficile* infection, a finding that might suggest a potential therapeutic protocol for metabolic syndrome⁸⁴. Inoculation of fecal microbiota via gastroduodenal tube from lean donors into obese subjects resulted in an increase in the abundance (2.5 fold increase) and diversity of gut microbiota, improvement of insulin sensitivity, increase in the proportion of butyrate producer *Roseburia intestinalis*, and decrease in the level of short chain fatty acids⁸⁵.

Dysbiotic gut microbiota has been found to play a role in obesity, other obesity related disorders such as type 2 diabetes (T2D), and metabolic syndrome⁸⁶⁻⁸⁸. The role of microbiota to weight gain and host metabolism is not completely understood. It was suggested that in obese individual, the

presence of specific microbial communities may increase the energy harvest and thus predispose to obesity^{86,88}. Other studies asserted that there can be several other mechanisms in which the microbial populations can influence weight gain and alteration of host metabolism⁸⁹.

Neurodegenerative diseases

Aging is associated with progressive changes in the gastrointestinal motility, defective gut-blood barrier, weakness in the immune function, and improper protein folding. Such age-related changes appear to have a great impact on the diversity of gut microbiota and may be linked to the age-relatedness of the neurodegenerations⁹⁰.

Amyloid protein has been linked to neurodegeneration particularly to Alzheimer's disease (AD) and the possibility of production of amyloid protein by human microbiota might raise great concerns about the possible role of microbiota in the induction of AD and other neurodegenerative diseases (NDD) including behavioral changes and autism⁹¹⁻⁹⁵. Nonfunctional amyloids is found in yeast and bacteria⁹³. There is paucity of information in literature on the potential role of amyloid protein in the pathogenesis of NDD. It was hypothesized that amyloids may induce or influence human neurodegeneration by three possible mechanisms including misfolding, neuroinflammation, and oxidative stress⁹⁶.

Antigen presenting cells such as epithelial microfold (M) cells and dendritic cells may uptake proteins produced by gut microbiota and transmit them into neurons in the myenteric plexus providing a means of communication between gut microbiota and CNS tissue⁹⁷. One misfolded molecule may elicit the misfolding of a different molecule to cause cross-seeding of A β -aggregation *in vitro*⁹⁸. Cross-seeding of neurodegenerative disorder proteins may be induced by environmental amyloids such as those produced by bacteria⁹⁹.

Bacterial amyloid has been demonstrated to activate a wide array of inflammatory molecules such as toll-like receptor-2 (TLR2), NFκB, TLR1, and CD14¹⁰⁰. Cerebral amyloid may mimic viral or bacterial infection resulting in glial cell activation through TLRs¹⁰¹. TLR2 activation lead to upregulation of Notch1 which play a crucial role in the development of AD¹⁰². It has been established that there is sterile inflammation in the brain in neurodegenerative disorders¹⁰³.

Peripheral inflammatory conditions may be involved in the induction of different form of neurodegeneration¹⁰⁴. It was reported that peripherally-induced inflammation induced damage of dopaminergic neurons as a response to the activation of complement pathway of microglial cells¹⁰⁵. CD14, which is involved in the activation of the TLR2/TLR1 complex lead to upregulation of NFκB expression and induction of oxidative toxicity that has been implicated in all neurodegenerative disorders¹⁰⁶. Bacterial amyloids are recognized through TLR2 mediated pathway leading to inflammation and oxidative toxicity that are the main induction factors to AD and PD¹⁰⁷.

Conclusion

Recent advances in metagenomics and metatranscriptomics tools coupled with the availability of rapid and cost-effective sequencing platforms have revolutionized the field of microbiome research. However, it remains imperative to completely understand the strengths and limitations of current genetic methods used to study the human gut microbiome. Advances in bioinformatics and high-throughput sequencing techniques facilitate the identification of the abundance and diversity of human microbiota in the different body niches, and help to decipher the possible link between microbiome and different disease conditions. Such goals are important prerequisites to identify novel diagnostic and therapeutic targets that will help to alleviate and cure different microbiome-associated diseases.

Deciphering the possible inter-individual variations in the microbial composition within different body regions and identification of the potential role of these human microbiota in the induction of different disease conditions is expected to hasten the application of microbiota-driven personalized medicine. In addition, it is possible to identify novel antibiotics against the emerging antibiotic resistant microbiota that may be present in different human body niches. Altogether, this comprehensive picture on human microbiome will help understand the current therapeutics available for modulating the gut microbiome composition for the prevention and treatment of various NCDs and determine whether the dysbiosis and reduced microbial diversity seen in many NCDs is causal or a consequence of those diseases.

Current status of microbiome research discloses the need to map the different types and complexity of microbiota among different human populations globally and to demonstrate the possible reciprocal interplay between some developmental factors, different environmental factors such as food, lifestyle, exposure to different environmental hazards and microbiota.

Research has shown the possible role of microbiota in induction of different disease conditions and in exploring possible novel therapeutic and preventative strategies to improve and reverse the microbiome-associated diseases as well as promote global human health.

Acknowledgments:

Authors would like to thank (Qatar University, and BRC, etc) for supporting this work.

Authors Contributions: AAT and HM conceived the study. HM drafted the manuscript. MEZ and MA designed the schematic diagrams. MEZ, SAK and MA contributed in parts in writing the manuscript. MEZ, SAK, NGK, HAZ and MA have thoroughly and critically revised the manuscript. All authors have read and revised the manuscript before submission.

Conflict of Interests: None to disclose.

References:

- 1 Cénit, M., Matzaraki, V., Tigchelaar, E. & Zhernakova, A. Rapidly expanding knowledge on the role of the gut microbiome in health and disease. *Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease***1842**, 1981-1992 (2014).
- 2 Ley, R. E., Peterson, D. A. & Gordon, J. I. Ecological and evolutionary forces shaping microbial diversity in the human intestine. *Cell***124**, 837-848 (2006).
- 3 Belenguer, A. *et al.* Rates of production and utilization of lactate by microbial communities from the human colon. *FEMS microbiology ecology***77**, 107-119 (2011).
- 4 Cho, I. & Blaser, M. J. The human microbiome: at the interface of health and disease. *Nature Reviews Genetics***13**, 260-270 (2012).
- 5 Qin, J. *et al.* A human gut microbial gene catalogue established by metagenomic sequencing. *nature***464**, 59-65 (2010).
- 6 Virgin, H. W. The virome in mammalian physiology and disease. *Cell***157**, 142-150 (2014).
- 7 Park, H. K. *et al.* Characterization of the fungal microbiota (mycobiome) in healthy and dandruff-afflicted human scalps. *PLoS One***7**, e32847 (2012).
- 8 Paulino, L. C., Tseng, C.-H., Strober, B. E. & Blaser, M. J. Molecular analysis of fungal microbiota in samples from healthy human skin and psoriatic lesions. *Journal of clinical microbiology***44**, 2933-2941 (2006).
- 9 Eckburg, P. B. *et al.* Diversity of the human intestinal microbial flora. *science***308**, 1635-1638 (2005).
- 10 Johnson, C. L. & Versalovic, J. The human microbiome and its potential importance to pediatrics. *Pediatrics***129**, 950-960 (2012).
- 11 Laparra, J. M. & Sanz, Y. Interactions of gut microbiota with functional food components and nutraceuticals. *Pharmacological Research***61**, 219-225 (2010).
- 12 Lozupone, C. A., Stombaugh, J. I., Gordon, J. I., Jansson, J. K. & Knight, R. Diversity, stability and resilience of the human gut microbiota. *Nature***489**, 220-230 (2012).
- 13 Blaser, M. J. Harnessing the power of the human microbiome. *Proceedings of the National Academy of Sciences***107**, 6125-6126 (2010).
- 14 Peterson, J. *et al.* The NIH human microbiome project. *Genome research***19**, 2317-2323 (2009).
- 15 O'Hara, A. M. & Shanahan, F. The gut flora as a forgotten organ. *EMBO reports***7**, 688-693, doi:10.1038/sj.embor.7400731 (2006).
- 16 Ghosh, A. R. Appraisal of microbial evolution to commensalism and pathogenicity in humans. *Clinical medicine insights. Gastroenterology***6**, 1-12, doi:10.4137/CGast.S11858 (2013).
- 17 Moens, E. & Veldhoen, M. Epithelial barrier biology: good fences make good neighbours. *Immunology***135**, 1-8, doi:10.1111/j.1365-2567.2011.03506.x (2012).
- 18 Korecka, A. & Arulampalam, V. The gut microbiome: scourge, sentinel or spectator? *Journal of oral microbiology***4**, doi:10.3402/jom.v4i0.9367 (2012).
- 19 Al-Asmakh, M., Anuar, F., Zadjali, F., Rafter, J. & Pettersson, S. Gut microbial communities modulating brain development and function. *Gut microbes***3**, 366-373, doi:10.4161/gmic.21287 (2012).
- 20 Stappenbeck, T. S., Hooper, L. V. & Gordon, J. I. Developmental regulation of intestinal angiogenesis by indigenous microbes via Paneth cells. *Proceedings of the National Academy of Sciences of the United States of America***99**, 15451-15455, doi:10.1073/pnas.202604299 (2002).
- 21 Ichinohe, T. *et al.* Microbiota regulates immune defense against respiratory tract influenza A virus infection. *Proceedings of the National Academy of Sciences of the United States of America***108**, 5354-5359, doi:10.1073/pnas.1019378108 (2011).

- 22 Diaz Heijtz, R. *et al.* Normal gut microbiota modulates brain development and behavior. *Proceedings of the National Academy of Sciences of the United States of America***108**, 3047-3052, doi:10.1073/pnas.1010529108 (2011).
- 23 Amaral, F. A. *et al.* Commensal microbiota is fundamental for the development of inflammatory pain. *Proceedings of the National Academy of Sciences of the United States of America***105**, 2193-2197, doi:10.1073/pnas.0711891105 (2008).
- 24 Nicholson, J. K. *et al.* Host-gut microbiota metabolic interactions. *Science***336**, 1262-1267, doi:10.1126/science.1223813 (2012).
- 25 Sjogren, K. *et al.* The gut microbiota regulates bone mass in mice. *Journal of bone and mineral research : the official journal of the American Society for Bone and Mineral Research***27**, 1357-1367, doi:10.1002/jbmr.1588 (2012).
- 26 Wilson, I. D. Drugs, bugs, and personalized medicine: pharmacometabonomics enters the ring. *Proceedings of the National Academy of Sciences of the United States of America***106**, 14187-14188, doi:10.1073/pnas.0907721106 (2009).
- 27 Bjorkholm, B. *et al.* Intestinal microbiota regulate xenobiotic metabolism in the liver. *PloS one***4**, e6958, doi:10.1371/journal.pone.0006958 (2009).
- 28 Al-Asmakh, M. *et al.* The gut microbiota and developmental programming of the testis in mice. *PloS one***9**, e103809, doi:10.1371/journal.pone.0103809 (2014).
- 29 Braniste, V. *et al.* The gut microbiota influences blood-brain barrier permeability in mice. *Science translational medicine***6**, 263ra158, doi:10.1126/scitranslmed.3009759 (2014).
- 30 Al-Asmakh, M. & Hedin, L. Microbiota and the control of blood-tissue barriers. *Tissue barriers***3**, e1039691, doi:10.1080/21688370.2015.1039691 (2015).
- 31 Pflughoeft, K. J. & Versalovic, J. Human microbiome in health and disease. *Annual Review of Pathology: Mechanisms of Disease***7**, 99-122 (2012).
- 32 Preidis, G. A. & Versalovic, J. Targeting the human microbiome with antibiotics, probiotics, and prebiotics: gastroenterology enters the metagenomics era. *Gastroenterology***136**, 2015-2031 (2009).
- 33 Sonnenburg, J. L. & Fischbach, M. A. Community health care: therapeutic opportunities in the human microbiome. *Science translational medicine***3**, 78ps12-78ps12 (2011).
- 34 Al-Asmakh, M., Anuar, F., Zadjali, F., Rafter, J. & Pettersson, S. Gut microbial communities modulating brain development and function. *Gut microbes***3**, 366-373 (2012).
- 35 Lau, E., Carvalho, D. & Freitas, P. Gut Microbiota: Association with NAFLD and Metabolic Disturbances. *BioMed Research International* (2015).
- 36 Baumgart, M. *et al.* Culture independent analysis of ileal mucosa reveals a selective increase in invasive *Escherichia coli* of novel phylogeny relative to depletion of Clostridiales in Crohn's disease involving the ileum. *The ISME journal***1**, 403-418 (2007).
- 37 Pickard, J. M. *et al.* Rapid fucosylation of intestinal epithelium sustains host-commensal symbiosis in sickness. *Nature* (2014).
- 38 Sokol, H. *et al.* *Faecalibacterium prausnitzii* is an anti-inflammatory commensal bacterium identified by gut microbiota analysis of Crohn disease patients. *Proceedings of the National Academy of Sciences***105**, 16731-16736 (2008).
- 39 Hansen, R. *et al.* Microbiota of de-novo pediatric IBD: increased *Faecalibacterium prausnitzii* and reduced bacterial diversity in Crohn's but not in ulcerative colitis. *The American journal of gastroenterology***107**, 1913-1922 (2012).
- 40 Ott, S. J. *et al.* Fungi and inflammatory bowel diseases: alterations of composition and diversity. *Scandinavian journal of gastroenterology***43**, 831-841 (2008).
- 41 Mukhopadhyay, I. *et al.* The fungal microbiota of de-novo paediatric inflammatory bowel disease. *Microbes and Infection***20**, 1e7 (2014).

- 42 Norman, J. M. *et al.* Disease-specific alterations in the enteric virome in inflammatory bowel
disease. *Cell***160**, 447-460 (2015).
- 43 Brint, E. K., MacSharry, J., Fanning, A., Shanahan, F. & Quigley, E. M. Differential expression of
toll-like receptors in patients with irritable bowel syndrome. *The American journal of
gastroenterology***106**, 329-336 (2011).
- 44 Ponnusamy, K., Choi, J. N., Kim, J., Lee, S.-Y. & Lee, C. H. Microbial community and metabolomic
comparison of irritable bowel syndrome faeces. *Journal of medical microbiology***60**, 817-827
(2011).
- 45 Rajilić–Stojanović, M. *et al.* Global and deep molecular analysis of microbiota signatures in fecal
samples from patients with irritable bowel syndrome. *Gastroenterology***141**, 1792-1801 (2011).
- 46 Jeffery, I. B. *et al.* An irritable bowel syndrome subtype defined by species-specific alterations in
faecal microbiota. *Gut***61**, 997-1006 (2012).
- 47 Manichanh, C., Borruel, N., Casellas, F. & Guarner, F. The gut microbiota in IBD. *Nature Reviews
Gastroenterology and Hepatology***9**, 599-608 (2012).
- 48 Sokol, H. *et al.* Specificities of the fecal microbiota in inflammatory bowel disease. *Inflammatory
bowel diseases***12**, 106-111 (2006).
- 49 Arthur, J. C. *et al.* Intestinal inflammation targets cancer-inducing activity of the microbiota.
*science***338**, 120-123 (2012).
- 50 McCoy, A. N. *et al.* Fusobacterium is associated with colorectal adenomas. *PLoS one***8**, e53653
(2013).
- 51 Guinane, C. M. *et al.* Microbial composition of human appendices from patients following
appendectomy. *MBio***4**, e00366-00312 (2013).
- 52 Grivennikov, S. I. *et al.* Adenoma-linked barrier defects and microbial products drive IL-23/IL-17-
mediated tumour growth. *Nature***491**, 254-258 (2012).
- 53 Li, X.-X. *et al.* Bacterial microbiota profiling in gastritis without Helicobacter pylori infection or
non-steroidal anti-inflammatory drug use. *PLoS One***4**, e7985 (2009).
- 54 De Vries, A. C. & Kuipers, E. J. Helicobacter pylori infection and nonmalignant diseases.
*Helicobacter***15**, 29-33 (2010).
- 55 Nardone, G. & Compare, D. The human gastric microbiota: Is it time to rethink the pathogenesis
of stomach diseases? *United European Gastroenterology Journal*, 2050640614566846 (2015).
- 56 de Vries, A. C., Kuipers, E. & Rauws, E. Helicobacter pylori Eradication and Gastric Cancer: When
Is the Horse Out of the Barn? *The American journal of gastroenterology***104**, 1342-1345
(2009).
- 57 Dicksved, J. *et al.* Molecular characterization of the stomach microbiota in patients with gastric
cancer and in controls. *Journal of medical microbiology***58**, 509-516 (2009).
- 58 Guerre, J., Vedel, G., Gaudric, M., Paul, G. & Cornuau, J. [Bacterial flora in gastric juice taken at
endoscopy in 93 normal subjects]. *Pathologie-biologie***34**, 57-60 (1986).
- 59 Mouzaki, M. *et al.* Intestinal microbiota in patients with nonalcoholic fatty liver disease.
*Hepatology***58**, 120-127 (2013).
- 60 Mutlu, E. A. *et al.* Colonic microbiome is altered in alcoholism. *American Journal of Physiology-
Gastrointestinal and Liver Physiology***302**, G966-G978 (2012).
- 61 Bode, J., Bode, C., Heidelbach, R., Dürr, H. & Martini, G. Jejunal microflora in patients with
chronic alcohol abuse. *Hepato-gastroenterology***31**, 30-34 (1984).
- 62 Yan, A. W. *et al.* Enteric dysbiosis associated with a mouse model of alcoholic liver disease.
*Hepatology***53**, 96-105 (2011).
- 63 Kirpich, I. A. *et al.* Probiotics restore bowel flora and improve liver enzymes in human alcohol-
induced liver injury: a pilot study. *Alcohol***42**, 675-682 (2008).

- 64 Bode, C. & Bode, J. C. Alcohol's role in gastrointestinal tract disorders. *Alcohol health and research world***21**, 76-83 (1997).
- 65 Wegener, M. *et al.* Gastrointestinal transit of solid-liquid meal in chronic alcoholics. *Digestive diseases and sciences***36**, 917-923 (1991).
- 66 Bajaj, J. S. *et al.* Colonic mucosal microbiome differs from stool microbiome in cirrhosis and hepatic encephalopathy and is linked to cognition and inflammation. *American Journal of Physiology-Gastrointestinal and Liver Physiology***303**, G675-G685 (2012).
- 67 Bajaj, J. S. *et al.* Linkage of gut microbiome with cognition in hepatic encephalopathy. *American Journal of Physiology-Gastrointestinal and Liver Physiology***302**, G168-G175 (2012).
- 68 Chen, Y. *et al.* Characterization of fecal microbial communities in patients with liver cirrhosis. *Hepatology***54**, 562-572 (2011).
- 69 Lu, H. *et al.* Intestinal microbiota was assessed in cirrhotic patients with hepatitis B virus infection. *Microbial ecology***61**, 693-703 (2011).
- 70 Wu, Z.-W. *et al.* Assessment of the fecal lactobacilli population in patients with hepatitis B virus-related decompensated cirrhosis and hepatitis B cirrhosis treated with liver transplant. *Microbial ecology***63**, 929-937 (2012).
- 71 Xu, M. *et al.* Changes of fecal Bifidobacterium species in adult patients with hepatitis B virus-induced chronic liver disease. *Microbial ecology***63**, 304-313 (2012).
- 72 Teltschik, Z. *et al.* Intestinal bacterial translocation in rats with cirrhosis is related to compromised Paneth cell antimicrobial host defense. *Hepatology***55**, 1154-1163 (2012).
- 73 Turner, J. R. Intestinal mucosal barrier function in health and disease. *Nature Reviews Immunology***9**, 799-809 (2009).
- 74 Adachi, Y., Moore, L. E., Bradford, B. U., Gao, W. & Thurman, R. G. Antibiotics prevent liver injury in rats following long-term exposure to ethanol. *Gastroenterology***108**, 218-224 (1995).
- 75 Tranah, T. H., Vijay, G. K. M., Ryan, J. M. & Shawcross, D. L. Systemic inflammation and ammonia in hepatic encephalopathy. *Metabolic brain disease***28**, 1-5 (2013).
- 76 Hur, K. Y. & Lee, M.-S. Gut Microbiota and Metabolic Disorders. *Diabetes & Metabolism Journal***39**, 198-203 (2015).
- 77 Turnbaugh, P. J., Bäckhed, F., Fulton, L. & Gordon, J. I. Diet-induced obesity is linked to marked but reversible alterations in the mouse distal gut microbiome. *Cell host & microbe***3**, 213-223 (2008).
- 78 Ley, R. E. *et al.* Obesity alters gut microbial ecology. *Proceedings of the National Academy of Sciences of the United States of America***102**, 11070-11075 (2005).
- 79 Karlsson, F. H. *et al.* Gut metagenome in European women with normal, impaired and diabetic glucose control. *Nature***498**, 99-103 (2013).
- 80 Qin, J. *et al.* A metagenome-wide association study of gut microbiota in type 2 diabetes. *Nature***490**, 55-60 (2012).
- 81 De Kort, S., Keszthelyi, D. & Masclee, A. Leaky gut and diabetes mellitus: what is the link? *Obesity Reviews***12**, 449-458 (2011).
- 82 Everard, A. *et al.* Responses of gut microbiota and glucose and lipid metabolism to prebiotics in genetic obese and diet-induced leptin-resistant mice. *diabetes***60**, 2775-2786 (2011).
- 83 Everard, A. *et al.* Microbiome of prebiotic-treated mice reveals novel targets involved in host response during obesity. *The ISME journal* (2014).
- 84 van Nood, E. *et al.* Duodenal infusion of donor feces for recurrent *Clostridium difficile*. *New England Journal of Medicine***368**, 407-415 (2013).
- 85 Dixon, M. F., Genta, R. M., Yardley, J. H. & Correa, P. Classification and grading of gastritis: the updated Sydney system. *The American journal of surgical pathology***20**, 1161-1181 (1996).

- 86 Ley, R. E. Obesity and the human microbiome. *Current opinion in gastroenterology***26**, 5-11
(2010).
- 87 Tilg, H. & Kaser, A. Gut microbiome, obesity, and metabolic dysfunction. *The Journal of clinical
investigation***121**, 2126 (2011).
- 88 Turnbaugh, P. J. *et al.* A core gut microbiome in obese and lean twins. *nature***457**, 480-484
(2009).
- 89 Clarke, S. F. *et al.* The gut microbiota and its relationship to diet and obesity: new insights. *Gut
microbes***3**, 186-202 (2012).
- 90 Claesson, M. J. *et al.* Gut microbiota composition correlates with diet and health in the elderly.
*Nature***488**, 178-184 (2012).
- 91 Hufnagel, D. A., Tükel, Ç. & Chapman, M. R. Disease to dirt: the biology of microbial amyloids.
*Plos Pathog***9**, e1003740 (2013).
- 92 Oli, M. W. *et al.* Functional amyloid formation by *Streptococcus mutans*. *Microbiology***158**, 2903-
2916 (2012).
- 93 Schwartz, K. & Boles, B. R. Microbial amyloids—functions and interactions within the host.
*Current opinion in microbiology***16**, 93-99 (2013).
- 94 Hsiao, E. Y. *et al.* Microbiota modulate behavioral and physiological abnormalities associated
with neurodevelopmental disorders. *Cell***155**, 1451-1463 (2013).
- 95 Kang, D.-W. *et al.* Reduced incidence of *Prevotella* and other fermenters in intestinal microflora
of autistic children. *PLoS One***8**, e68322 (2013).
- 96 Bokranz, W., Wang, X., Tschäpe, H. & Römling, U. Expression of cellulose and curli fimbriae by
Escherichia coli isolated from the gastrointestinal tract. *Journal of Medical Microbiology***54**,
1171-1182 (2005).
- 97 Mabbott, N. A., Donaldson, D. S., Ohno, H., Williams, I. R. & Mahajan, A. Microfold (M) cells:
important immunosurveillance posts in the intestinal epithelium. *Mucosal immunology***6**, 666-
677 (2013).
- 98 Morales, R. *et al.* Molecular cross talk between misfolded proteins in animal models of
Alzheimer's and prion diseases. *The Journal of Neuroscience***30**, 4528-4535 (2010).
- 99 Friedland, R. P. Mechanisms of Molecular Mimicry Involving the Microbiota in
Neurodegeneration. *Journal of Alzheimer's disease: JAD***45**, 349-362 (2015).
- 100 Tükel, Ç. *et al.* Toll-like receptors 1 and 2 cooperatively mediate immune responses to curli, a
common amyloid from enterobacterial biofilms. *Cellular microbiology***12**, 1495-1505 (2010).
- 101 Trudler, D., Farfara, D. & Frenkel, D. Toll-like receptors expression and signaling in glia cells in
neuro-amyloidogenic diseases: towards future therapeutic application. *Mediators of
inflammation***2010**: 497987 (2010).
- 102 Palaga, T. *et al.* Notch signaling is activated by TLR stimulation and regulates macrophage
functions. *European journal of immunology***38**, 174-183 (2008).
- 103 Akiyama, H. *et al.* Inflammation and Alzheimer's disease. *Neurobiology of aging***21**, 383-421
(2000).
- 104 Raby, A.-C. *et al.* Soluble TLR2 reduces inflammation without compromising bacterial clearance
by disrupting TLR2 triggering. *The Journal of Immunology***183**, 506-517 (2009).
- 105 Bodea, L.-G. *et al.* Neurodegeneration by Activation of the Microglial Complement–Phagosome
Pathway. *The Journal of Neuroscience***34**, 8546-8556 (2014).
- 106 Yan SD, Y. S., Chen X, Fu J, Chen M, Kuppusamy P, Smith MA, Perry G, Godman GC, Nawroth P,
et al. Non-enzymatically glycosylated tau in Alzheimer's disease induces neuronal oxidant stress
resulting in cytokine gene expression and release of amyloid b-peptide. *Nature medicine***1**, 693-
699 (1995).

- 107 Holmes, C. Review: systemic inflammation and Alzheimer's disease. *Neuropathology and applied neurobiology***39**, 51-68 (2013).

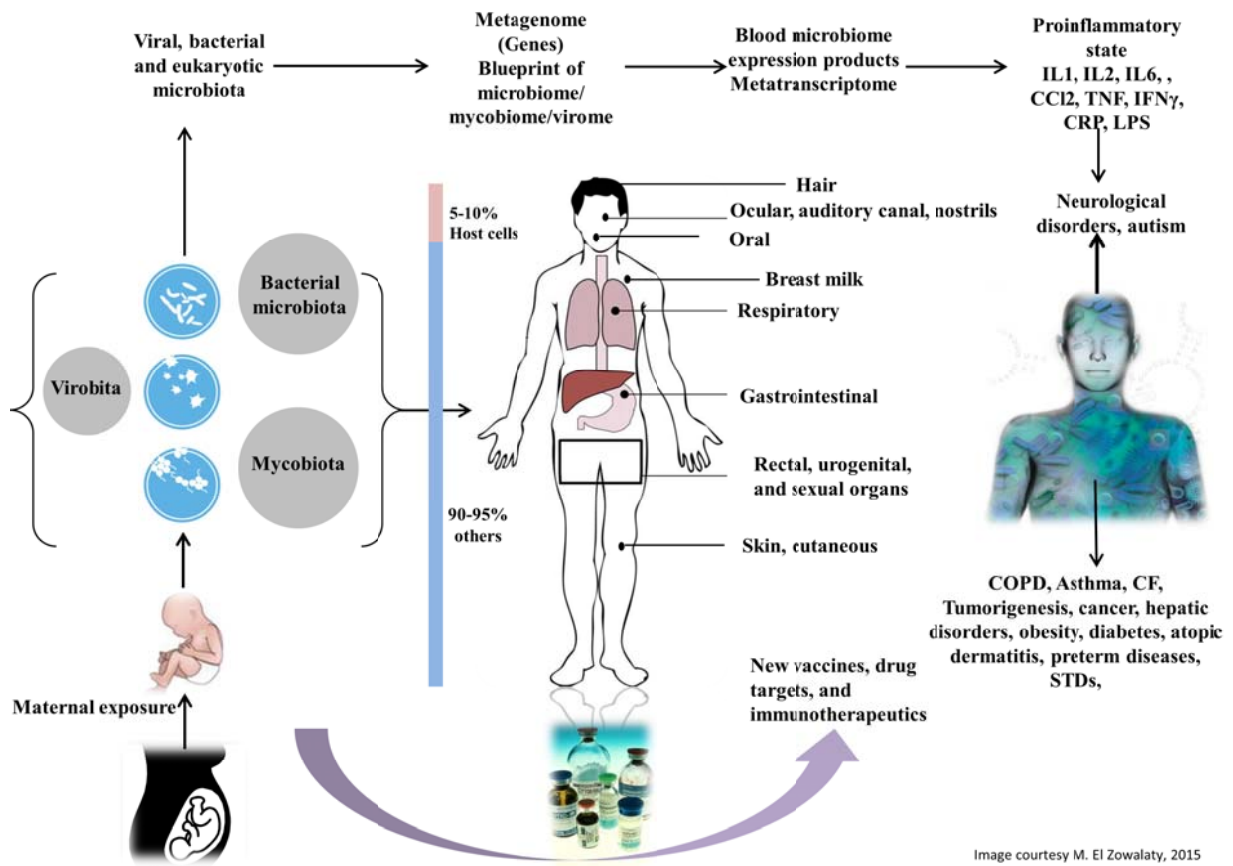


Image courtesy M. El Zowalaty, 2015

Figure 1

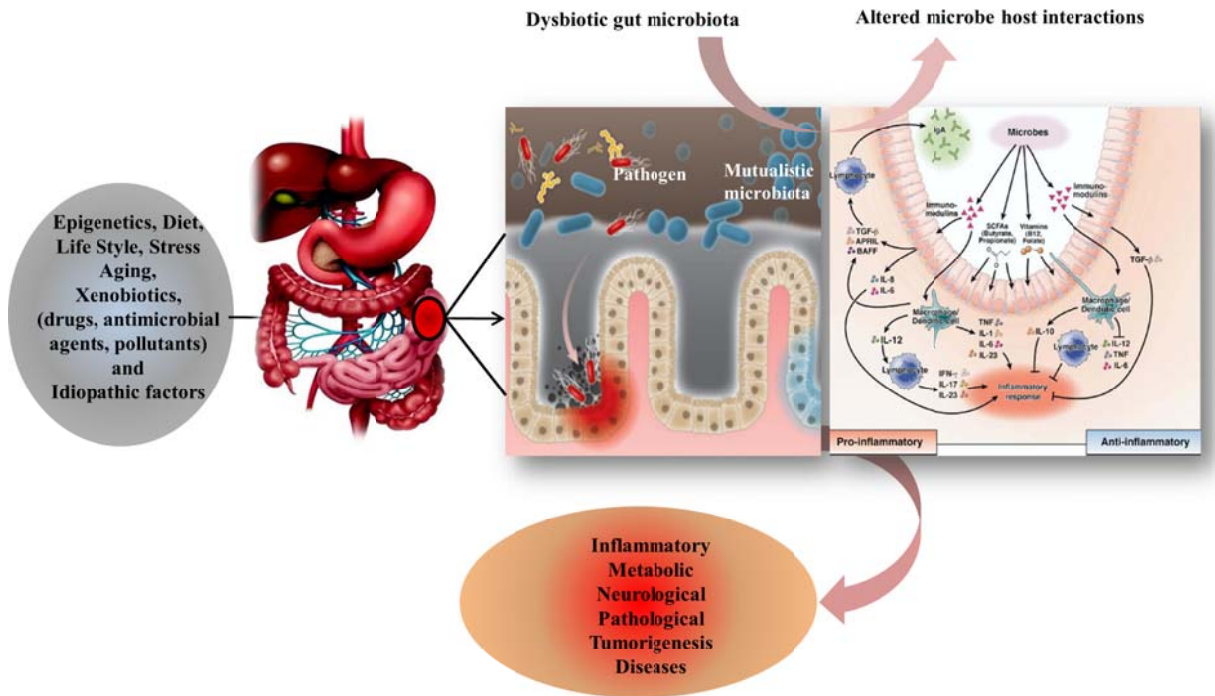


Image courtesy M. El Zowalaty, 2015

Figure 2

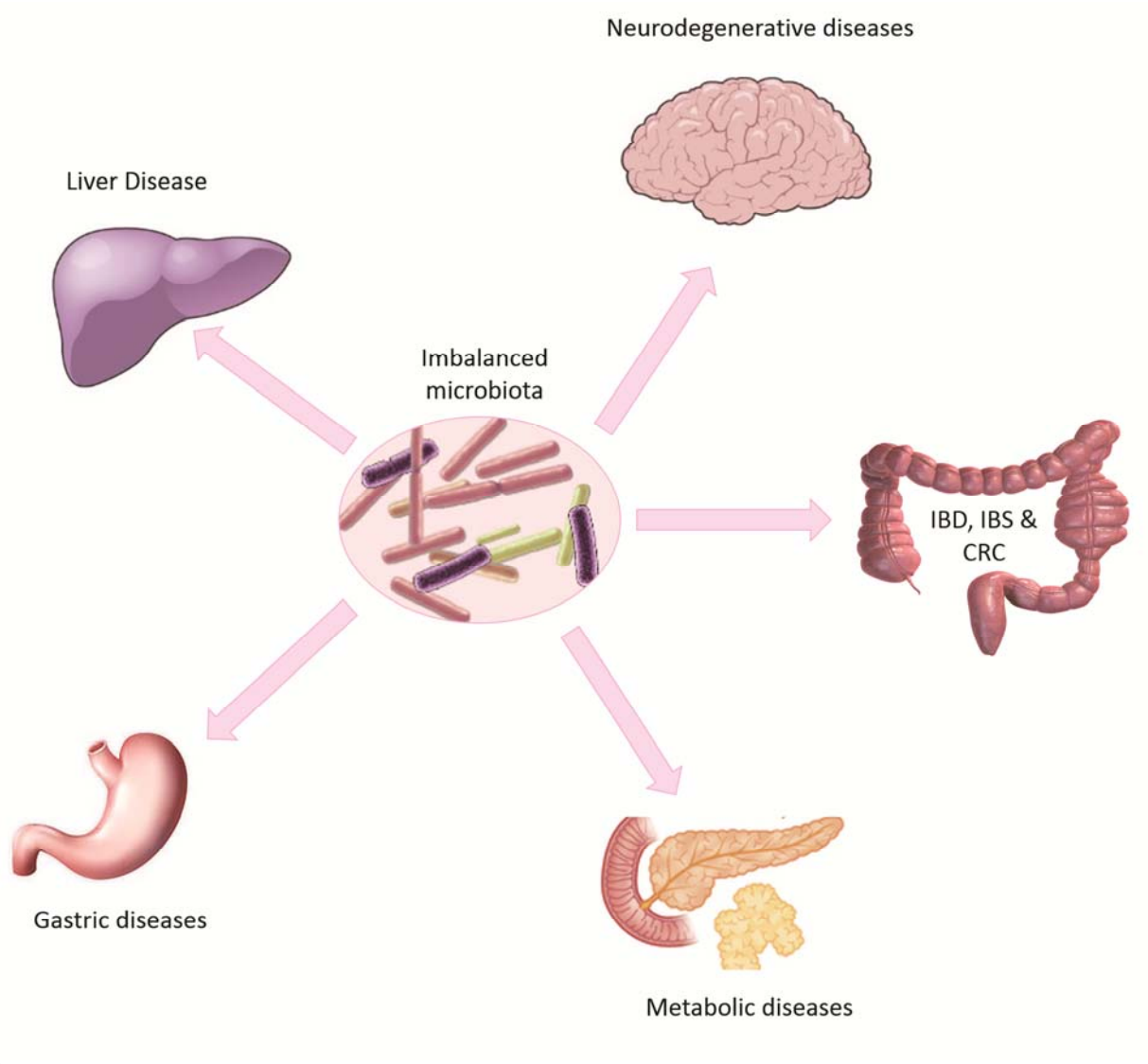


Figure 3: A diagram showing the microbiota and associated diseases has been added