

NANO EXPRESS

Open Access

Hybrid magnetite nanoparticles/*Rosmarinus officinalis* essential oil nanobiosystem with antibiofilm activity

Carmen Chifiriuc¹, Valentina Grumezescu², Alexandru Mihai Grumezescu^{2*}, Crina Saviuc¹, Veronica Lazăr¹ and Ecaterina Andronescu²

Abstract

Biofilms formed by fungal organisms are associated with drastically enhanced resistance against most antimicrobial agents, contributing to the persistence of the fungi despite antifungal therapy. The purpose of this study is to combine the unique properties of nanoparticles with the antimicrobial activity of the *Rosmarinus officinalis* essential oil in order to obtain a nanobiosystem that could be pelliculised on the surface of catheter pieces, in order to obtain an improved resistance to microbial colonization and biofilm development by *Candida albicans* and *C. tropicalis* clinical strains. The *R. officinalis* essential oils were extracted in a Neo-Clevenger type apparatus, and its chemical composition was settled by GC-MS analysis. Functionalized magnetite nanoparticles of up to 20 nm size had been synthesized by precipitation method adapted for microwave conditions, with oleic acid as surfactant. The catheter pieces were coated with suspended core/shell nanoparticles ($\text{Fe}_3\text{O}_4/\text{oleic acid}:\text{CHCl}_3$), by applying a magnetic field on nanofluid, while the CHCl_3 diluted essential oil was applied by adsorption in a secondary covering treatment. The fungal adherence ability was investigated in six multiwell plates, in which there have been placed catheters pieces with and without hybrid nanoparticles/essential oil nanobiosystem pellicle, by using culture-based methods and confocal laser scanning microscopy (CLSM). The *R. officinalis* essential oil coated nanoparticles strongly inhibited the adherence ability and biofilm development of the *C. albicans* and *C. tropicalis* tested strains to the catheter surface, as shown by viable cell counts and CLSM examination. Due to the important implications of *Candida* spp. in human pathogenesis, especially in prosthetic devices related infections and the emergence of antifungal tolerance/resistance, using the new core/shell/coated shell based on essential oil of *R. officinalis* to inhibit the fungal adherence could be of a great interest for the biomedical field, opening new directions for the design of film-coated surfaces with antibiofilm properties.

Background

The increasing occurrence of multidrug resistant, extensive drug resistant and pandrug resistant microbial strains has gradually rendered traditional antimicrobial treatment ineffective [1]. The prognosis is worsened by the formation of microbial biofilms on the biomaterials used in medicine due to their phenotypic resistance, even if the component cells tested in suspension (by the standard method) are susceptible to some antibiotics. Recent public announcements stated that 60% to 85% of

all microbial infections involve biofilms developed on natural, intact or damaged tissues (skin, mucosa, endothelial epithelia and teeth), or artificial devices (central venous catheters, peritoneal, urinary catheters, dental materials, cardiac valves, intrauterine contraceptive devices, contact lenses and other implants). The insertion of the prosthetic medical devices for different exploratory or therapeutical purposes, especially in severe pathological conditions, represents a risk factor for the occurrence of chronic infections in developed countries, being characterized by slow onset, middle intensity symptoms, chronic evolution, and resistance to antibiotic treatment [2]. In this context, finding and testing new preventive/therapeutic strategies for

* Correspondence: grumezescu@yahoo.com

²Faculty of Applied Chemistry and Materials Science, Politechnica University of Bucharest, Bucharest, Romania

Full list of author information is available at the end of the article

prosthetic devices associated infections have become a top priority at the international level. Two main strategies have been employed for the prevention of catheter associated infections: a) development of biomaterials with antiadhesive properties using physico-chemical methods [3-5] and b) incorporation or coating biomaterials which significantly reduce the microbial adherence, e.g., *in vitro* urinary catheters impregnated with silver ions. However, recent research is raising the risk of selecting resistance by coating catheters with antimicrobial substances [6]. Nanotechnology is expected to open some new ways to fight and prevent diseases using atomic scale tailoring of materials [7]. There are a lot of reports on the antimicrobial and antibiofilm properties of different types of nanoparticles, especially heavy metal containing ones (silver, copper, gold and ZnO) ([8-10], as well as the core/shell nanosystems (e.g., CoFe_2O_4 /oleic acid, Fe_3O_4 /oleic acid and Fe_3O_4 /PEG600) [11,12]. Although the potential of natural compounds of vegetal origin to be used as therapeutical remedies is known since longtime, their use is still empirical, but the problem of microbial resistance as well as the negative impact of the chemical substances discharged in the external environment on the ecological balance has reinforced the studies concerning the characterization of the chemical structures of vegetal products and the active doses, aiming to understand their specific mechanisms of action. The efficiency of essential oils obtained from different plant species and their synergic effects as alternative strategies for the treatment of severe infections caused by highly resistant bacteria was already proved [13,14]. The purpose of this study is to combine the unique properties of nanoparticles with the antimicrobial activity of the *Rosmarinus officinalis* essential oil in order to obtain a nanobiosystem that could be pelliculised on the surface of catheter pieces, in order to obtain an improved resistance to microbial colonization and biofilm development.

Methods

Extraction and analysis of *R. officinalis* essential oils

The essential oil microwave assisted extraction was performed in a Neo-Clevenger type apparatus, and its chemical composition was settled by GC-MS analysis. Gas chromatographic analysis was performed using an Agilent 6890 Series GC System gas chromatograph (Agilent Technologies Inc., Santa Clara, CA, USA) fitted with a splitless injector for a low background with an injector liner split/splitless under a column head pressure of 12.5 psi and H_2 as carrier gas at a flow rate of 1.2 ml/min. Oven temperature was programmed from 50 to 300°C at 5°C/min. Injector and detector temperatures were 250°C. A capillary column DB5-MS fused-silica J&W Scientific Inc. (Krackeler Scientific, Inc., Albany,

NY) (30 m × 0.25 mm i.d.; 0.25 μm film) was used. Detection was carried out with a 5973 mass-selective single quadrupole detector (Agilent technologies). Operation control and the data process were carried out by Agilent Technologies ChemStation software. The mass spectrometer was calibrated before use with perfluorotributylamine as a calibration standard.

Synthesis and characterization of hybrid nanomaterial core/shell/coated shell

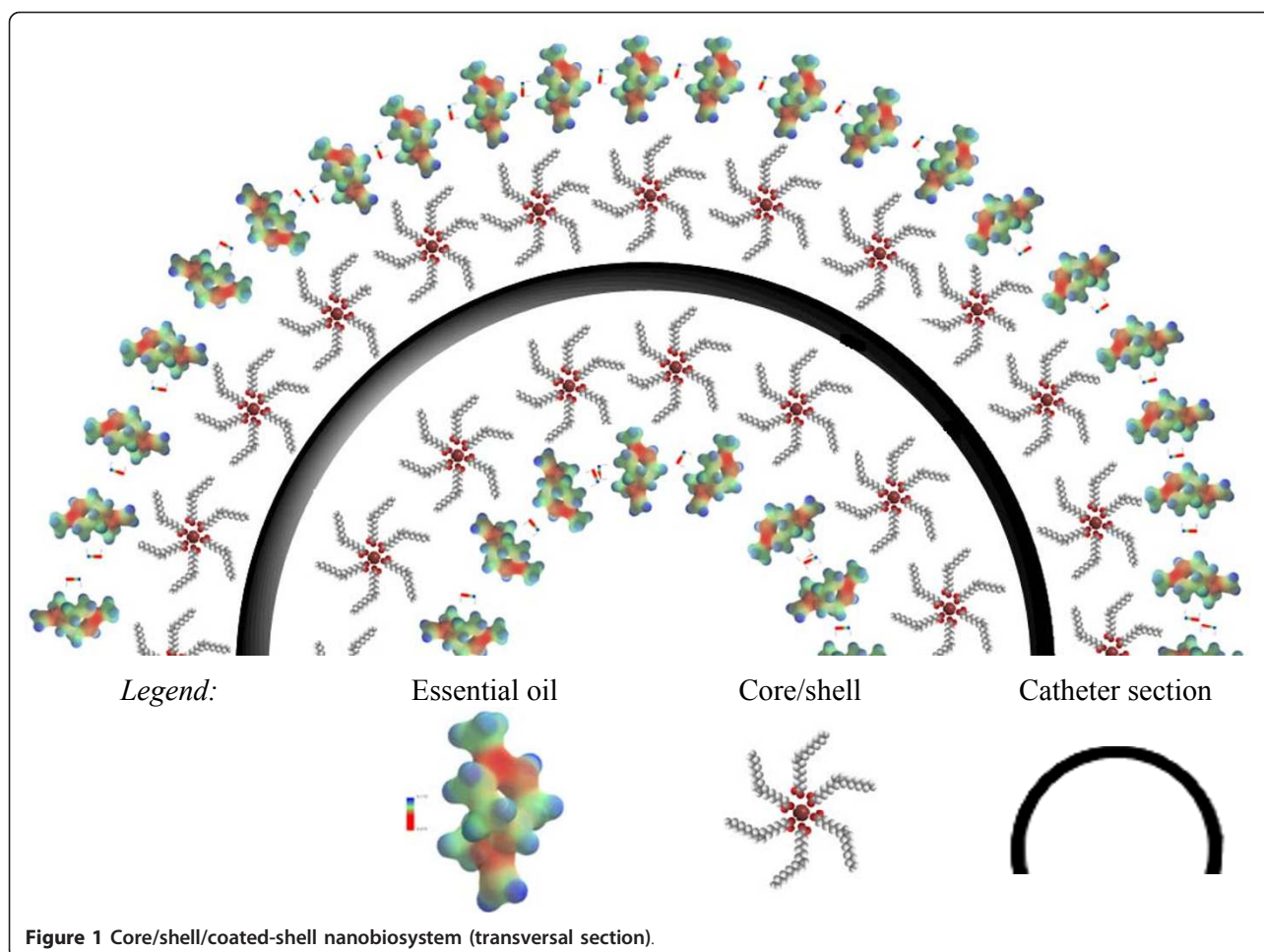
Functionalized magnetite nanoparticles had been synthesized by precipitation method adapted for microwave conditions, with oleic acid as surfactant. High resolution transmission electron microscopy was used as a primary characterization method for Fe_3O_4 /oleic acid- core/shell nanostructure with dimensions not exceeding 20 nm, according to our previous studies [15,16]. The nanobiosystem was obtained by coating the prosthetic device with suspended core/shell nanoparticles (Fe_3O_4 /oleic acid: CHCl_3 0.33% (w/v)). The layer of core/shell nanostructure on prosthetic device was achieved by submerging the catheter pieces in 5 ml of the nanofluid represented by Fe_3O_4 /oleic acid: CHCl_3 0,33% (w/v). The nanoparticles were aligned in a magnetic field of 100 Kgf applied for 1 s, and thereafter, the catheter pieces have been extemporaneously dried at room temperature. The rapid drying was facilitated by the convenient volatility of chloroform. In order to achieve core/shell/coated-shell type samples, the extra-shell (CHCl_3 diluted essential oil, 160 μL/mL) was applied by adsorption in a secondary covering treatment, performed by submerging again for 1 s the catheter samples previously coated with Fe_3O_4 /oleic acid in 5 ml of CHCl_3 diluted essential oil, followed by extemporaneous drying at room temperature (Figure 1). The coated prosthetic devices were then sterilized by ultraviolet irradiation for 15 min.

Fungal model

The artificial monospecific biofilms were developed using two strains of *C. tropicalis* and *C. albicans* recently isolated from clinical specimens and identified by using Vitek II (bioMérieux, Inc., Durham, NC automatic system and previously tested for their susceptibility to currently used antifungals (voriconazole, itraconazole, caspofungin, amphotericin B, fluconazole and flucytosin) and to some essential oils [13,14,17].

Microbial adherence to the inert and modified prosthetic devices

The microbial adherence ability was investigated in six multiwell plates, in which there have been placed catheters pieces of 1 cm with and without nanobiosystem. Plastic wells were filled with liquid medium, inoculated



with 300 μ L 0.5 McFarland microbial suspension and incubated for 72 h at 30°C. After 24 h, the culture medium was removed, the catheters were washed three times in phosphate buffered saline (PBS) in order to remove the non-adherent strains, and fresh Glucose broth was added. Also viable cell counts (VCCs) have been achieved for both working variants (coated and uncoated catheter pieces) at each 24 h in order to assess the biofilm forming ability of the two strains. The adhered cells have been removed from the catheter sections by vortexing and brief sonication and serial dilutions ranging from 10^{-1} to 10^{-6} of the obtained inocula have been spotted on Sabouraud agar, incubated for 24 h at 30°C and assessed for VCCs [18].

Direct examination of biofilm architecture by CLSM

In order to evaluate the biofilm formation on coated and uncoated catheters, a confocal laser scanning microscopy (CLSM) was used. After 48 and 72 h of incubation, the samples were removed from the plastic wells, washed three times with PBS, fixed with cold methanol, and dried before microscopic examination. Samples

were visualized in reflection and transmission mode by using a Leica microscope (TCS-SP CSLM model), equipped with PL FLUOTAR (40X NA0.7, electronic zoom 1), and an He-Ne laser tuned on 633 nm wavelength. A lateral resolution of about 600 nm was achieved [19]. The Leica software was used for examining the surface topography.

Results and discussions

The increasing resistance of *C. albicans* towards the existent antifungal compounds and the reduced number of available drugs led to the search of new therapeutic alternatives among plants and their essential oils, empirically used due to their antifungal proprieties. Plant oils traditionally used for domestic and therapeutic purposes are increasingly claimed to have broad spectrum antimicrobial properties. Some essential oils have been suggested to have potent antimicrobial activity, including antihelminthic, skin infections and insect bites, chicken pox, colds, flu and measles sinus congestion, asthma, bronchitis, pneumonia, tuberculosis, and cholera properties, due to their phenolic, alcoholic, and

terpenoid constituents. Agarwal et al. (2008) investigated the inhibitory effect of 30 plant oils against one biofilm forming *C. albicans* strain isolated from a clinical sample, out of which eucalyptus, peppermint, ginger grass, and clove oils proved to be fungistatic, fungicidal, and antibiofilm agents [20].

Previous studies indicated the antimicrobial activity of essential-oil rich fractions of *R. officinalis* against gram-positive (*Staphylococcus aureus* and *Bacillus subtilis*), gram-negative bacterial strains (*Escherichia coli* and *Pseudomonas aeruginosa*) but also against yeasts (*C. albicans*) and molds (*Aspergillus niger*), the main components found in the oil (80%) being represented by alpha-Pinene, 1,8-cineole, camphor, verbenone, and borneol [21].

The identified phytocomponents from *R. officinalis* essential oil analyzed in our study are listed in Table 1. The principal components percentage was 40.596% Eucalyptol, 11.389% Camphor, 10.19% Caryophyllene, and alpha-Pinene 18.42% from the total area.

The microbial biofilm is defined as a sessile microbial community composed of cells irreversibly attached to a substratum and between them, embedded in a matrix of extracellular polymeric substances produced by themselves and which presents a modified phenotype with respect to their rate of growth as well as gene transcription [22]. The biofilm cells are resistant to all kinds of antimicrobial substances: antibiotics, antiseptics, disinfectants; this type of resistance, consecutive to biofilm formation is phenotypical, behavioral, and more recently, called tolerance [23]. Tolerance is defined as the ability to survive from bactericidal factors without necessarily expressing a genetic resistance mechanism. The microbial species of clinical interest, often involved

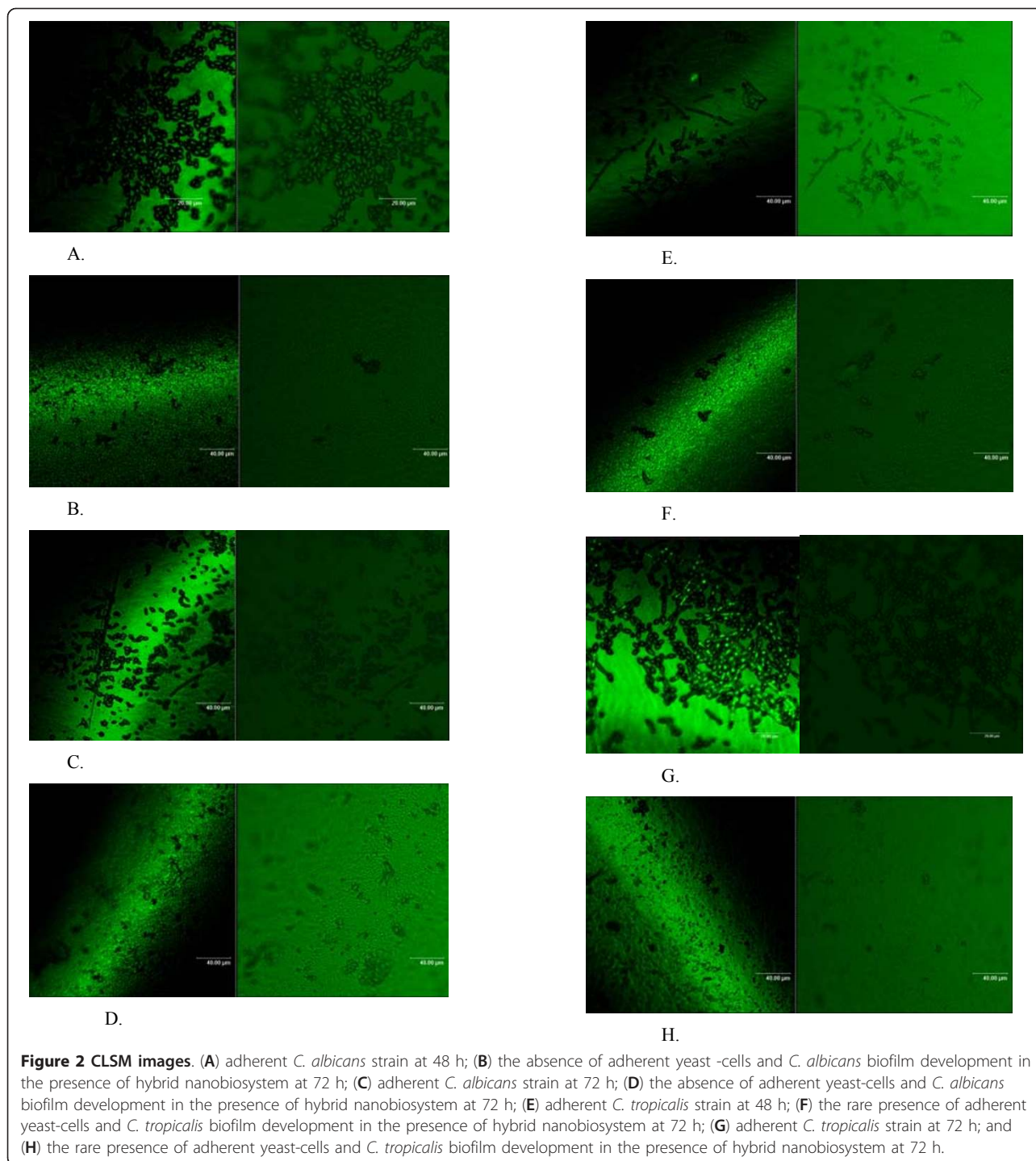
in biofilm associated diseases, are belonging to a very large spectrum, from the gram positive (*Staphylococcus epidermidis* and *Staphylococcus aureus*) to the gram negative pathogens (*Pseudomonas aeruginosa*, and *Escherichia coli*) and to different members of the *Candida* genus. Biofilms formed by fungal organisms are associated with drastically enhanced resistance against most antimicrobial agents, contributing to the persistence of the fungi despite antifungal therapy [20,24].

Although *C. albicans* is the predominant etiologic agent of candidiasis, other *Candida* species that tend to be less susceptible to the commonly used antifungal drugs such as *C. krusei*, *C. glabrata*, *C. lusitaniae*, and the newest *Candida* species, *C. dubliniensis*, have emerged as substantial opportunistic pathogens [25]. The outlined work investigates the potential of *R. officinalis* essential oil stabilized in nanolayer and its effect on biofilm formation of *C. albicans* and *C. tropicalis* strains, previously isolated from clinical samples. Confocal laser scanning microscopy (CLSM) was used for the analysis of fungal biofilms developed on the uncoated catheters and on the catheters pelliculised with the plant oils included in the nanosystem. Visualization of control biofilms revealed the ability of the tested strains to form biofilms on the glass coverlips. The visualization of fungal biofilms developed at 48 h revealed the presence of yeast-phase cells attached to the inert substratum and the microcolony yeast basal layer formation, as well as the expansion of the biofilm architecture by the occurrence of hyphal and pseudohyphal forms, that diminished at 72 h (Figure 2). Visualization of the biofilm structure in the presence of the tested hybrid nanosystem revealed an important reduction in adhering cells and biofilm development as compared to the untreated biofilm, the adhered yeast-cells being practically absent at 48 and 72 h for both tested strains (Figure 2). This suggests that despite the relative minimal diffusion due to the inclusion of the essential oil in the nanoparticles layer, the vegetal compounds may be exerting a metabolic interference in biofilm.

The antibiofilm effect of the tested nanobiosystem was also proved by the significant reduction of viable cell number in the presence of the protective pellicle of the hybrid nanobiosystem. In the case of *C. albicans* biofilms, although a slight reduction was observed in the early phase of biofilm development (at 24 h), a clear inhibitory effect was observed in the case of 48- and 72-h biofilms, the VCCs drastically dropping with approximately 85% to 98%, as compared to the uncoated controls (Figure 3). Comparatively, in case of *C. tropicalis* biofilm, the most intensive decrease of viable cells adhered to the catheter sections was observed at 24 h (Figure 3). These results could be probably related to the earlier production of

Table 1 Chemical composition of *R.officinalis* L. essential oils

Peak	Compound	RT	Relative content (%)
1	Tricyclene	4.557	0.527
2	Alpha-pinene	4.96	18.42
3	Camphene	5.274	3.532
4	Beta-pinene	6.08	8.162
5	Alpha-phellandrene	6.852	0.164
6	Eucalyptol	7.793	40.596
7	Alpha-terpinene	8.509	0.49
8	Camphor	11.073	11.389
9	Borneol	11.621	0.286
10	Alpha-terpineol	12.394	0.327
11	Bornyl acetate	15.136	2.602
12	Copaene	17.487	0.201
13	Caryophyllene	18.685	10.19
14	Caryophyllene oxide	22.615	0.873



pseudohyphae and germ tubes by *C. tropicalis* (48 h) as compared with *C. albicans* (72 h). These structures could represent additional contact and adherence points to inert or cellular substrata and also sources of genetic variability and adaptation to different limitative conditions, being responsible, for example, for the high tolerance to antifungal substances [26].

Conclusions

The *R. officinalis* essential oil coated nanoparticles strongly inhibited the adherence ability and biofilm development on catheter surface of the *C. albicans* and *C. tropicalis* tested strains, as shown by VCCs and CLSM examination. Due to the important implications of *Candida* spp. in human pathogenesis, especially in

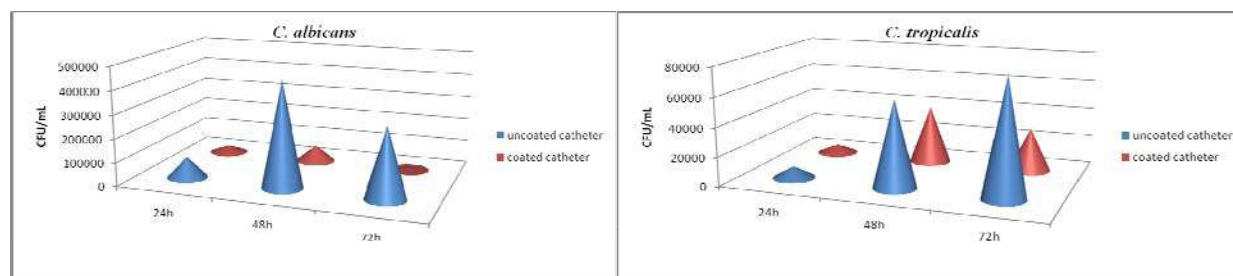


Figure 3 The viable cell counts of fungal cells. The viable cell counts of fungal cells adhered and embedded in biofilms formed on the catheter surface (control versus coated catheters).

prosthetic devices related infections and antifungal tolerance/resistance, using the new core/shell/coated shell based on essential oil of *R. officinalis* to inhibit fungal adherence to prosthetic device could be of a great interest for the biomedical field. These materials-based approaches to control of fungal adherence could provide both (i) new tools to study mechanisms of fungal virulence and biofilm formation, and (ii) approaches to the design of film-coated surfaces or to treat the surfaces of solid and fiber-based materials that prevent or disrupt the formation of fungal biofilms.

Acknowledgements

This paper is supported by the Sectoral Operational Programme Human Resources Development, financed from the European Social Fund, and by the Romanian Government under the contract number POSDRU/86/1.2/S/58146 (MASTERMAT) and by the Romanian CNCSIS research project, Human Resources Project no. 135/2010, contract no. 76/2010 and ideas contract no. 154/05.10.2011. The authors thank to Prof. G. Stanciu, Head of Center for Microscopy-Microanalysis and Information Processing and PhD Student Radu Hristu for their assistance in obtaining the CLSM images.

Author details

¹Faculty of Biology, University of Bucharest, Bucharest, Romania ²Faculty of Applied Chemistry and Materials Science, Politehnica University of Bucharest, Bucharest, Romania

Authors' contributions

CC conceived the study, provided the fungal strains, and drafted the manuscript together with AG. AG and VG performed the synthesis and characterization of hybrid nanomaterial core/shell/coated shell. CS obtained the essential oil and performed the biological analyses, EA and VL participated in the design of the study and coordination. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Received: 10 January 2012 Accepted: 10 April 2012

Published: 10 April 2012

References

1. Falagas ME, Karageorgopoulos DE: Pandrug resistance (PDR), extensive drug resistance (XDR), and multidrug resistance (MDR) among gram-negative bacilli: need for international harmonization in terminology. *Clin Infect Dis* 2008, **46**:1121-1122.
2. Donlan RM, Costerton JW: Biofilms: Survival mechanisms of clinically relevant microorganisms. *Clinical Microbiol Rev* 2002, **15**:167-193.
3. Stripple H, Westman R, Holm D: Development and environmental improvements of plastics for hydrophilic catheters in medical care: an environmental evaluation. *J Clean Prod* 2008, **16**:1764-1776.
4. Witjes JA, et al: Multicenter, double-blind, randomized, parallel group study comparing polyvinyl chloride and polyvinyl chloride-free catheter materials. *J Urol* 2009, **182**:2794-2798.
5. Perni S, Pratten J, Wilson M, Piccirillo C, Parkin IP: Prokopovich: antimicrobial properties of light-activated polyurethane containing indocyanine green. *P J Biomater Appl* 2011, **25**:387-400.
6. Timsit J, Dubois Y, Minet C, Bonadona A, Lugosi M, Ara-Somohano C, Hamidfar R, Schwebel R: New materials and devices for preventing biofilm associated infections. *Annals of Intensive Care* 2011, **1**:34.
7. Singh M, Singh S, Prasad S, Gambhir IS: Nanotechnology in medicine and antibacterial effect of silver nanoparticles. *Digest J Nanomaterials and Biostructures* 2008, **3**:115-122.
8. Jun Sung Kim, Eunye Kuk, Kyeong Nam Yu, Jong-Ho Kim, Sung Jin Park, Hu Jang Lee, So Hyun Kim, Young Kyung Park, Yong Ho Park, Cheol-Yong Hwang, Yong-Kwon Kim, Yoon-Sik Lee, Dae Hong Jeong, Myung-Haing Cho: Antimicrobial effects of silver nanoparticles. *Nanomedicine: Nanotechnology, Biology, and Medicine* 2007, **3**:95-101.
9. Rai A, Prabhune A, Perry CC: Antibiotic mediated synthesis of gold nanoparticles with potent antimicrobial activity and their application in antimicrobial coatings. *J Mater Chem* 2010, **20**:6789-6799.
10. Gajjar P, Pettee B, Britt DW, Huang W, Johnson WP, Anderson AJ: Antimicrobial activities of commercial nanoparticles against an environmental soil microbe, *Pseudomonas putida* KT2440. *J Biol Eng* 2009, **3**:9.
11. Chifiriuc MC, Lazar V, Bleotu C, Calugarescu I, Grumezescu AM, Mihaiescu DE, Mogoşanu DE, Buteica AS, Buteica E: Bacterial adherence to the cellular respectively inert substrate in the presence of magnetic CoFe₂O₄ and Fe₃O₄/oleic acid - core/shell nanoparticle. *Digest J Nanomaterials and Biostructures* 2011, **6**:37-42.
12. Saviuc C, Grumezescu AM, Holban A, Chifiriuc C, Mihaiescu D, Lazar V: Hybrid nanostructured material for biomedical applications. *Biointerface Res Appl Chem* 2011, **1**:64-71.
13. Saviuc C, Grumezescu AM, Oprea E, Radulescu V, Dascalu L, Chifiriuc MC, Bucur M, Banu O, Lazar V: Antifungal activity of some vegetal extracts on *Candida* biofilms developed on inert substratum. *Biointerface Res Appl Chem* 2011, **1**:15-23.
14. Saviuc C, Grumezescu AM, Holban A, Bleotu C, Chifiriuc C, Balaure P, Lazar V: Phenotypical studies of raw and nanosystem embedded *Eugenia carryophyllata* buds essential oil antibacterial activity on *Pseudomonas aeruginosa* and *Staphylococcus aureus* strains. *Biointerface Res Appl Chem* 2011, **1**:111-118.
15. Buteică AS, Mihaiescu DE, Grumezescu AM, Vasile BS, Popescu A, Mihaiescu OM, Cristescu R: The anti-bacterial activity of magnetic nanofluid: Fe₃O₄/oleic acid/cephalosporins core/shell/adsorption shell proved on *S. aureus* and *E. coli* and possible applications as drug delivery systems. *Digest J Nanomaterials and Biostructures* 2010, **5**:927.
16. Mihaiescu DE, Grumezescu AM, Mogoşanu DE, Traistaru V, Balaure PC, Buteica A: Hybrid organic/inorganic nanomaterial for controlled cephalosporins release. *Biointerface Res Appl Chem* 2011, **1**:41-47.

17. Saviuc C, Grumezescu AM, Chifiriuc MC, Bleotu C, Stanciu G, Hristu R, Mihaiescu D, Lazăr V: *In vitro* methods for the study of microbial biofilms. *Biointerface Res Appl Chem* 2011, **1**:31-40.
18. Grumezescu AM, Saviuc C, Chifiriuc MC, Hristu R, Mihaiescu DE, Balaure P, Stanciu G, Lazar V: **Inhibitory Activity of Fe₃O₄/Oleic Acid/Usnic Acid-Core/Shell/Extra-Shell Nanofluid on *S. aureus* Biofilm Development**, *Transactions on NanoBioScience*, DOI:10.1109/TNB.2011.2178263.
19. Banu O, Bleotu C, Chifiriuc MC, Savu B, Stanciu G, Antal C, Alexandrescu M, Lazăr V: **Virulence factors of *Staphylococcus aureus* and *Pseudomonas aeruginosa* strains involved in the etiology of cardiovascular infections**. *Biointerface Res Appl Chem* 2011, **1**:72.
20. Agarwal V, Lal P, Pruthi V: **Prevention of *Candida albicans* biofilm by plant oils**. *Mycopathologia* 2008, **165**:13-19.
21. Santoyo S, Caverio S, Jaime L, Ibañez E, Señoráns FJ, Reglero G: **Chemical composition and antimicrobial activity of *Rosmarinus officinalis* L. essential oil obtained via supercritical fluid extraction**. *J Food Prot* 2005, **684**:790-795.
22. Costerton JW: **Biofilms: the bacterial way to persist**, Abstracts book of The International symposium and the 43 rd ESCMID post-graduate course- Bacterial Adaptation Mechanisms: Biofilms, Hypermutability and antibiotic Resistance, Palma de Mallorca, Spain. 2007, **9**:12.
23. Lazar V: **Quorum sensing in biofilms-how to destroy the bacterial citadels or their cohesion/power?** *Anaerobe* 2011, **17**:280-285.
24. Lazar V, Chifiriuc C: **Medical significance and new therapeutical strategies for biofilm associated infections**. *Rom Arch of Microb & Immunol* 2010, **69**:125-138.
25. Jabra-Rizk MA, Falkler WA, Meiller TF: **Fungal biofilms and drug resistance**. *Emerg Infect Dis* 2004, **10**:1.
26. Lazăr V: **Microbial Adherence**. Bucharest: Romanian Academy Press; 2003, ISBN: 973-27-0992-8.

doi:10.1186/1556-276X-7-209

Cite this article as: Chifiriuc et al.: Hybrid magnetite nanoparticles/*Rosmarinus officinalis* essential oil nanobiosystem with antibiofilm activity. *Nanoscale Research Letters* 2012 **7**:209.

Submit your manuscript to a SpringerOpen[®] journal and benefit from:

- Convenient online submission
- Rigorous peer review
- Immediate publication on acceptance
- Open access: articles freely available online
- High visibility within the field
- Retaining the copyright to your article

Submit your next manuscript at ► springeropen.com
