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1	Hybridisation and hybrid speciation under global change
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54 Summary

55 An unintended consequence of global change is an increase in opportunities for hybridisation among previously isolated lineages. Here we illustrate how global change can facilitate the breakdown of 56 57 reproductive barriers and the formation of hybrids, drawing on the flora of the British Isles for 58 insight. Although global change may ameliorate some of the barriers preventing hybrid 59 establishment, for example by providing new ecological niches for hybrids, it will have limited effects 60 on environment-independent post-zygotic barriers. For example, genic incompatibilities and 61 differences in chromosome numbers and structure within hybrid genomes are unlikely to be 62 affected by global change. We thus speculate that global change will have a larger effect on eroding pre-zygotic barriers (eco-geographic isolation, phenology) than post-zygotic barriers, shifting the 63 64 relative importance of these two classes of reproductive barriers from what is usually seen in 65 naturally produced hybrids where pre-zygotic barriers are the largest contributors to reproductive 66 isolation. Although the long-term fate of neo-hybrids is still to be determined, the massive impact of 67 global change on the dynamics and distribution of biodiversity generates an unprecedented 68 opportunity to study large numbers of unpredicted, and often replicated, hybridisation 69 "experiments", allowing us to peer into the birth and death of evolutionary lineages. 70 71 Keywords: alien; allopolyploidy, genome duplication, global change, hybrid, invasive species, 72 reproductive isolation, speciation.

73

74 I. Introduction

75 'Global change', the term often used to describe the combination of planetary change and human 76 societal change, is having a profound effect on biodiversity across the globe. Climate change, 77 industrialization, environmental degradation, and global trade and travel have moved plants and 78 animals around the planet, breaking down previous geographic barriers to gene flow between 79 closely related species (Mooney & Cleland 2001). The loss of ecological and geographic barriers that 80 historically kept closely related species apart is creating unprecedented new opportunities for 81 hybridisation (Mable 2013; Chunco 2014, Brennan et al., 2015; Taylor et al., 2015), which could 82 potentially have a profound impact on biodiversity and ecosystems worldwide.

Hybridisation has been defined in many different ways, from crosses between genetically 83 84 distinct populations (Abbott et al., 2013), to crosses between genetically distinct taxa resulting in the 85 production of viable offspring (Mallet 2007), thus occurring both within and between species 86 (Rieseberg & Carney 1998). Here we focus on inter-specific hybridisation. Interspecific hybridisation 87 is a regular natural phenomenon and it is estimated that as many as 25% of plant species and 10% of 88 animal species hybridise naturally (Mallet 2007). Hybridisation has long attracted the interest of 89 evolutionary biologists, in part because it seems to undo the very process responsible for the 90 generation of species diversity (Dobzhansky 1937, Mayr 1942, Stebbins 1959, Grant 1971, Abbott et 91 al., 2013). However, the consequences of hybridisation between genetically distinct lineages can 92 result in a variety of outcomes that may influence diversity, including gene flow from one taxon to 93 another (introgression) (Rieseberg & Wendel, 1993), the displacement and/or extinction of one or 94 both parental taxa (Ellstrand & Elam, 1993), the fusion of previously divergent taxa (Grant & Grant, 95 2014), or the creation of new, stable hybrid taxa and, ultimately, speciation (Mallet 2007, Abbott et 96 al., 2013, Thomas 2015). Hybridisation and its consequences have been best studied in plants (Grant, 97 1971; Arnold, 1997; Rieseberg & Carney, 1998; Abbott et al., 2013), but the biological features and evolutionary mechanisms that contribute to the variation in the occurrence, persistence, and 98 99 evolution of hybrids are relatively little understood in both plants and animals. 100 Whether the incidence of hybridisation at a global scale is changing, and if so at what rate, is still

unclear, yet analyses of well-studied floras, such as the British Isles, indicate that a significant
fraction of hybrid taxa may involve introduced taxa (Stace *et al.*, 2015, Stace & Crawley 2015).
Among the flora of Britain and Ireland hybridisation involving introduced (non-native) taxa is well
documented (Stace *et al.*, 2015, Stace & Crawley 2015, Table 1), and a recent survey revealed that
33% of hybrid taxa (301/909) involve introduced (non-native) taxa (Preston & Pearman 2015).

106 Approximately half of these non-native hybrids have arisen spontaneously in the wild, while the

107 remainder were introduced as hybrids. Studying the origin and fate of these hybrids is timely and

important if we are to understand the consequences of ongoing changes in the distribution of globalbiodiversity.

110 Because hybrid formation does not equate to hybrid success, the long term consequences of 111 contemporary hybridisation under global change are hard to predict. Here we present an overview 112 of the consequences of this new era of increased hybridisation opportunities drawing on the flora of 113 Great Britain and Ireland because it is probably the best documented hybrid flora in the world. 114 Specifically, our review addresses the following questions: How does global change alter the 115 likelihood of hybrid formation? Does global change alter the relative importance of prezygotic and 116 postzygotic barriers in maintaining species apart? What is the incidence of human-made hybrids in current floras and is likely to increase or decrease under global change? What are the biological 117 118 characteristics that make some hybrids more likely to become established than others? Why are recently formed hybrids relatively common, but their allopolyploid derivatives rare? We suggest that 119 120 global change is clearly increasing opportunities for hybridisation, for example directly by moving 121 species around the world or indirectly by eroding phenological and ecological barriers. Human-made 122 hybrids, created for example for horticultural purposes, can also significantly contribute to current 123 floras as exemplified in the naturalised flora of Britain and Ireland. We speculate that this new 124 scenario is shifting the relative importance of prezygotic and postzygotic barriers from what is 125 generally seen in natural systems, and therefore the consequences of secondary contact in this 126 changing world may depend more heavily on postzygotic than on prezygotic barriers. The 127 persistence of newly formed hybrids, and their evolutionary fate, will hinge on mechanisms that stabilise hybrids including asexual reproduction, selection for increased fertility, polyploidy, and 128 129 ecological niche diversification, among others. In the next sections, we take a comprehensive 130 approach to understand the processes and mechanisms leading to: (1) hybrid formation, (2) the 131 establishment of hybrids, and (3) the ecological and evolutionary outcomes of hybridisation.

132

133 II. Hybrid formation

134 Understanding hybridisation and the potential for global change to alter its incidence requires 135 analysis of the conditions that allow hybrid formation and establishment. Speciation involves the 136 origin of barriers preventing gene flow between incipient species (Coyne & Orr 2004, Baack et al., 137 2015), but hybridisation bypasses these barriers. Isolating barriers are traditionally classified as those acting before (pre-zygotic) and after zygote formation (post-zygotic) (reviewed in Coyne & Orr 138 139 2004) (Fig. 1). In plants, pre-zygotic barriers include: (i) Pre-pollination barriers such as geographical 140 and ecological barriers (e.g. habitat preferences), flowering phenology and pollinator preference and 141 behaviour, which prevent or reduce the likelihood of interspecific mating; and (ii) post-pollination

barriers which act before zygote formation, such as unilateral interspecific incompatibility (Hiscock 142 et al., 1998), and gametophytic selection, in which conspecific pollen is favoured over heterospecific 143 144 pollen (Rieseberg & Willis 2007). The breakdown of geographic isolation is usually treated separately 145 from other isolating barriers (Coyne & Orr 2004) as it may reflect both biological differences in 146 habitat preference and non-biological and historical features causing species to occur in different 147 geographic locations (Dobzhansky 1937). However, we include geographic isolation in our discussion 148 because global change, including the movement of species around the globe, has had a direct impact 149 in increasing the opportunities for secondary contact between previously isolated taxa.

Although the potential for human-induced environmental change to influence the rate of hybridisation has long been recognised (e.g., Anderson 1948), the evidence to support this view is only just beginning to accumulate (Walther *et al.*, 2009, Garroway *et al.*, 2010, Hoffmann & Sgrò 2011, Campbell & Wendlandt 2013, Chunco 2014, Chown *et al.*, 2015). In this section, we discuss how hybrids may overcome pre-zygotic isolating barriers and geographic isolation, paying attention to those barriers that are likely to be affected by global change.

156 **1. Breakdown of geographic isolation**

171

157 The breakdown of geographic isolation when species increase their range, either as a consequence 158 of climate change (e.g. during periods of glaciation), or through anthropogenic dispersal, provides 159 historically isolated taxa with new opportunities for hybridisation. Human-mediated transport of 160 species, either accidental or deliberate, is the most dramatic of these and has been going on for 161 thousands of years, but has been accelerating rapidly in Europe and North America (Hulme et al., 162 2008). Increased international trade and travel in the current era of globalisation means that the 163 spread of non-native species is likely to increase (Hulme et al., 2008). The pathways of human-164 assisted dispersal are varied, and include accidental long-distance dispersal events that result in 165 translocation of terrestrial plant species across oceanic barriers, facilitated dispersal (e.g., along 166 roadsides and railroads), and intentional introductions (e.g., horticultural trade). Although long 167 distance dispersal events occur with low probability, theoretical analyses have shown that these events can accelerate migration rates in plants by an order of magnitude (Higgins & Richardson 168 169 1999), greatly increasing opportunities for secondary contact. 170 Long distance dispersal where propagules are transported as contaminants of commodities (e.g.,

instance, in the early 20th century, the River Tweed in the English-Scottish border hosted 384

grains, timber, and wool) and in ships' ballasts is well documented (Stace & Crawley 2015). For

173 introduced plant species growing along its banks (Myers & Bazely 2003). Many of these plants

- 174 originated in Europe, Asia, Australasia, and the Americas and were brought as wool contaminants,
- 175 which were then washed into local rivers during the cleaning process (Silvertown 2011). In addition,

176 anthropogenic activities can also facilitate range expansion by creating dispersal routes that can 177 then be exploited by natural means. Roads and railways provide corridors that can act as dispersal 178 routes allowing species to spread rapidly to new areas. The introduced hybrid Senecio squalidus (Oxford ragwort, Asteraceae), originally from Mount Etna, Sicily, escaped from cultivation in the 179 Oxford Botanic Garden in the late 18th century, but it was not until the development of railway lines 180 181 in the 1800's that it began spreading throughout the British Isles (Abbott et al., 2009). The spread of 182 S. squalidus in the British Isles has resulted in novel hybridisation events with native Senecio species. 183 Crosses between S. squalidus and S. vulgaris have given rise to three new fertile hybrid taxa via a 184 sterile triploid intermediate ('bridge') S. x baxteri: the tetraploid introgressant 'radiate goundsel', 185 Senecio vulgaris var. hibernicus, the allohexaploid S. cambrensis (Welsh groundsel), and the 186 tetraploid S. eboracensis (York radiate groundsel, Abbott & Lowe 2004). Additionally, hybridization 187 between S. squalidus and native S. viscosus has given rise to the sterile triploid S. subnebrodensis 188 (Lousley 1946).

189 In addition to facilitating long distance dispersal events, global change in the form of climate 190 change can increase previously existing areas of sympatry among species, or bring previously 191 isolated taxa together through shifts in their range (Hoffmann & Sgrò 2011, Brennan et al., 2015). 192 For example, changes in temperature or precipitation can result in increased range overlap, and the 193 creation or expansion of hybrid zones (Campbell & Wendlandt 2013, Taylor et al., 2015). Because hybrid zones often occur at the range limits of the parental species, changes to the geographic 194 195 boundaries where these species occur should have a particularly strong impact on the location and 196 extent of hybrid regions (Chunco 2014). There are a number of historical examples of this within the 197 British flora. For instance, the hybrid waterlily Nuphar x spenneriana (Nymphaeaceae) is postulated 198 to have arisen during the Late Glacial period ~10,000 years ago when the ranges of its parental 199 species N. pumila (tolerant of cold water) and N. lutea (preferring warmer water) overlapped 200 (Preston and Croft 1998). N x spenneriana then subsequently displaced N. lutea at its more northerly 201 locations due to its competitive advantage in tolerating cooler water. A similar story may account for 202 the current distribution of Circaea x intermedia (Onagraceae), a hybrid between C. alpina and C. 203 lutetiana which has all but displaced its cold-loving parent C. alpina in northern Britain since the ice 204 retreated (Marren 1999).

205 Climate change can also facilitate the spread of invasive populations which can then contribute 206 to hybridisation events either with native species, or with other invasive taxa. For example, warmer 207 climates may allow the migration of species from warmer regions into regions that were formally 208 too cold for their long-term survival as well as allowing introduced species from warmer regions to 209 overwinter more successfully and extend the growing season, thus facilitating the establishment and

- spread of non-native taxa (Walther *et al.,* 2009). The increases in populations of Orchis simia and O.
- 211 *purpurea* in Kent and their spread north into other counties of southern England provides a good
- example of the former scenario (Fay 2015). Despite the expected importance of climate change on
- increasing hybridisation rates (Chunco 2014, Brennan et al., 2015), most evidence in plants remains
- indirect (e.g., perceived changes on the extent of hybrid zones) or correlative (e.g., comparisons of
- 215 number of hybrids at different time points), and more detailed case studies are needed (e.g.,
- 216 Campbell & Wendlandt 2013).

217 **2. Erosion of ecological isolation barriers**

- 218 The production of hybrids may be prevented by pre-zygotic, ecological barriers that have evolved as 219 by-products of adaptation to their local environments (Coyne & Orr 2004, Baack et al., 2015) (Fig. 1). 220 For example, broadly sympatric species may still display ecological preferences for different habitats 221 (e.g., mesic vs. arid habitats and forested vs. open habitats), or be reproductively isolated by virtue 222 of flowering at different times. Habitat modification, including increased disturbance and 223 fragmentation, can erode ecological barriers and facilitate hybrid formation (Anderson 1948, 224 Stebbins 1950, Buggs 2007). Moreover, altered and disturbed habitats, e.g., arising as a consequence 225 of agricultural practices or urbanisation, can provide hybrids with new environments where they can 226 establish. Among the British flora Silene dioica x S. latifolia (= S. x hampeana, Caryophyllaceae) 227 (Marren 1999) provides a good example of how habitat modification may facilitate hybridisation, 228 while Senecio squalidus and S. cambrensis (Asteraceae) are examples of how hybrids can establish in 229 new environments (Abbott et al., 2009). Human-induced breakdown of ecological isolation has also 230 been implied in hybridisation between Centaurea erythraea and C. littorale (Gentianaceae). These 231 two species generally occur in different habitats, but human disturbance in sand dune systems is thought to have resulted in mixed populations (Ubsdell 1979), which can be found in coastal regions 232 233 in England and mainland Europe (Brys et al., 2014). Some of these populations harbour nearly sterile 234 F1 hybrids and backcrosses, which appear to differ in fertility (Ubsdell 1979). Nevertheless, 235 differences in floral morphology and mating system are efficient in reducing hybridisation, at least in 236 mixed populations in Belgium, where established hybrids occur at a frequency of approximately 1% 237 (Brys et al., 2014). Ultimately, the outcome of habitat alteration on hybridisation may be dependent on phylogenetic relationships between taxa. For instance, Brennan et al., (2015) suggest that habitat 238 239 alteration may be more important for hybridisation between younger taxa, as they are expected not 240 yet to have accumulated strong genetic barriers.
- Climate change affects the phenology of many species (Cleland *et al.*, 2007), for instance by
 causing plants to flower earlier. Changes in the timing of reproductive events, such as flowering, can
 directly alter the level of reproductive synchronisation between sympatric species, breaking down

temporal isolation barriers, and facilitating hybridisation. For temperature or precipitation changes
to increase hybridisation opportunities, it is necessary for one species to change its phenology while
the other remains stable or changes more slowly (Chunco 2014). Experimental data shows that this
situation may not be uncommon. For instance, artificial warming of experimental plots results in
more reproductive overlap among grassland species (Sherry *et al.,* 2007). The breakdown of
temporal isolation may be particularly important in sympatric species where phenological isolation is
an important reproductive barrier.

251 A further potential barrier to hybridisation in sympatric populations is isolation resulting from 252 pollinator preference (Ramsey et al., 2003). If global change affects the distribution of pollinators and/or their behaviour, we can predict that hybridisation could be facilitated by a breakdown of 253 254 pollinator isolation barriers (Campbell & Wendlandt 2013). Recent studies show that the distribution 255 of certain species of butterfly in the British Isles and Europe is changing and that their choices of 256 host (brood) plants are changing as a consequence (Bridle et al., 2013). If these changes mean that 257 pollinators visit a broader range of species, it is conceivable that this creates new opportunities for 258 hybridization. Paradoxically, changes in visitation preferences could hypothetically reduce 259 hybridisation, for instance if generalist pollinators change their preference to ignore rarer plant 260 species, thereby resulting in a reduction in heterospecific pollen transfer. In addition to changes in 261 visitation preferences, loss of pollinator diversity may also alter opportunities for hybridization. For 262 example, if pollinators that are specialised on individual plant species are lost, pollen transferred by 263 generalist pollinators may deposit a proportionally higher fraction of heterospecific pollen on stigmas. Clearly, the consequences of changes in pollinator distribution, diversity and abundance can 264 265 have complex repercussions on hybridization, and these may depend on the particular assemblage 266 of plant and pollinator species. Future studies, akin to that of Bridle et al., should seek to address 267 these questions.

268 **3. Artificial hybridisation**

269 Traditionally, artificial hybridisation is not considered in discussions of the ecological and 270 evolutionary significance of hybrids (Rieseberg 1995, Arnold 1997). However, hybrids can be 271 produced artificially and then become part of the natural environment. In the context of global 272 changes to species distribution, the importance of artificial hybrids may be substantial, at least at 273 ecological levels. For example, in the flora of the British Isles, approximately 17% (152/909) of 274 hybrids were introduced as hybrids (Preston & Pearman 2015). Moreover, some of these are 275 deliberate introductions of hybrids of agricultural or horticultural interest such as Mentha 276 (Lamiaceae), Mimulus (Phrymaceae), and Verbascum (Scrophulariaceae). Although most

277 horticultural varieties do not persist outside cultivation (Mack 2005), horticultural introductions do

contribute to the non-native component of many floras, including the British Isles (Clement & Foster1994, Stace 2010).

280 Artificial hybrids may have a disproportionate contribution to hybrid floras, as artificial 281 hybridisation can bypass other pre-zygotic barriers such as pollinator isolation, phenology, habitat 282 isolation and geographic isolation. Furthermore, artificial selection on horticultural hybrids may 283 coincidentally increase their probability of establishment and spread (Ellstrand & Schierenbeck 284 2000). For example, horticultural varieties are often selected on the basis of traits such as hardiness 285 and cold-tolerance (Milne & Abbott 2000), or ease of propagation and large plant size, which may 286 allow the establishment of cultivars beyond the ecological range of their wild parental species (Mack 287 2005). Indeed, it would be interesting to investigate whether artificial hybrids deliberately 288 introduced through the horticulture trade have a larger range than those that originated 289 spontaneously. Artificial crosses from both the horticultural trade and botanic gardens are a 290 relatively unappreciated source of plant hybridisation (Knobloch 1972, Ellstrand & Schierenbeck 291 2000), but the contribution of artificial hybrids to ecological and evolutionary phenomena is likely to 292 increase in an era of global change.

293 4. Weakening of gametic barriers

294 In some cases, pre-zygotic, post-pollination 'gametic' barriers must also be overcome for hybrid 295 formation. This type of post-pollination barrier can arise from gametophytic incompatibilities and 296 conspecific pollen precedence (Hiscock et al., 1998, Howard 1999, Husband et al., 2002). 297 Gametic/gametophytic barriers should be little affected by processes associated with global change, 298 unless these affect the relative receipt of heterospecific pollen (e.g., by increasing the number of 299 heterospecific matings). Because conspecific pollen precedence often depends on the relative 300 amounts of conspecific and heterospecific pollen received on stigmas (Howard 1999), an increase in 301 heterospecific pollen receipt may translate to higher rates of hybrid production. This could be the 302 case if an introduced species becomes invasive and comes to dominate a particular habitat where 303 related native species occur (Morales & Traveset 2009). Pollinators would then carry an increased 304 pollen load of the alien species with corresponding increased chance of cross-pollinating the native 305 species. Thus we predict that the main consequence of global change on pre-zygotic, post-306 pollination barriers will be through increasing the deposition of heterospecific pollen as non-native 307 species become more abundant, thereby weakening conspecific pollen precedence and facilitating 308 hybridization.

In summary, global change is clearly altering the opportunities for hybrid formation, and we
 would expect the incidence of hybridization to continue increasing. Increased opportunities for
 hybrid formation may result as a consequence of multiple and not mutually exclusive processes,

such as the global re-shuffling of species through human-mediated dispersal, shifting species 312 313 distributions and phenology as a consequence of climate change, the erosion of ecological barriers, 314 including the "hybridisation of the habitat", changes in pollinator preference and diversity, the 315 expansion of non-native species, and the artificial production and spread of hybrids. Although we 316 think that hybrid formation is likely to increase, an alternative view may be that we have reached 317 "peak" hybridisation, and the rate of hybridisation is slowing down. For example, it may be argued 318 that both species introductions and invasions reduce local biodiversity, and fewer species should 319 mean fewer hybridisation opportunities. Similarly, loss of habitat may reduce contact zones between 320 potentially hybridising taxa. However, we think that the effect of global change in promoting 321 hybridisation through the mechanisms reviewed in this section will by far exceed any hypothetical 322 reduction in hybridisation opportunities. As our records of local floras continues improving, 323 monitoring the temporal patterns of hybridization in floras around the world becomes feasible. Of 324 particular interest will be to monitor floras in developing countries where expansion of international 325 trade, and colossal changes in land use as economies grow, may fast-track the upward trend in the 326 formation of hybrids.

327III. Hybrid establishment: Overcoming intrinsic and extrinsic post-

328 zygotic barriers

329 After hybrids are formed, their short-term fate is partly determined by their ability to overcome both 330 intrinsic (e.g., environment-independent low viability and sterility), and extrinsic (e.g., ecological 331 selection) post-zygotic isolation barriers (Coyne & Orr 2004) (Fig. 1). In natural settings, pre-zygotic 332 barriers may contribute more to total reproductive isolation than post-zygotic barriers (Lowry et al., 333 2008, Baack et al., 2015). However, given the potential for global change to by-pass pre-zygotic 334 barriers and geographic isolation, as argued in the previous section, we speculate that post-zygotic 335 isolation may be the most important hurdle to overcome in the early stages of neo-hybridisation. 336 Intrinsic post-zygotic isolation barriers have been relatively well characterised at the genetic level, in 337 comparison to pre-zygotic barriers (Lowry et al., 2008, Widmer et al., 2009, Rieseberg & Blackman 338 2010). As in animal systems, post-zygotic isolation barriers are expected to increase as a function of 339 genetic divergence (Coyne & Orr 2004), although evidence of this from plant systems is limited 340 (Moyle et al., 2004, Scopece et al., 2007).

5. Escaping hybrid inviability and sterility

Hybrids are not uniformly unfit (Arnold & Hodges 1995, Rieseberg & Carney 1998), and within a
single hybrid taxon (or sometimes a single hybrid cross) it is possible to find individuals with lower,
similar or higher fitness relative to their parents (Taylor *et al.*, 2009). Nevertheless, many hybrids

have to overcome severe intrinsic post-zygotic barriers before they can become established (Lowry *et al.*, 2008).

347 Post-zygotic barriers include intrinsic hybrid inviability and sterility (Fig. 1), and may be 348 caused by a variety of mechanisms (Rieseberg & Blackman 2010, Baack et al., 2015), including 349 differences in chromosome structure and ploidy level (Stebbins 1971, Rieseberg 2001, Levin 2002), 350 nuclear-nuclear and nuclear-cytoplasmic genic incompatibilities (Lowry et al., 2008), and parental 351 genomic conflict during seed development (Köhler et al., 2010) (Fig. 1). Genic incompatibilities 352 associated with reduced hybrid fertility and viability have been extensively studied at the genetic 353 level in many plant hybrids (Lowry et al., 2008). The most widely accepted model for the 354 accumulation of such hybrid incompatibilities is the Dobzhansky-Muller (DM) model, in which 355 allopatric populations fix different (and mutually incompatible) alleles at one or more loci (Rieseberg & Willis 2007, Baack et al., 2015). An example of a post-zygotic hybridisation barrier that arises 356 357 through genic incompatibilities is hybrid necrosis, which resembles plant responses to stress 358 including pathogens (Bomblies & Weigel 2007). Hybrid necrosis can be debilitating or lethal, and 359 generally acts in early (F_1 and sometimes F_2) hybrid generations (Bomblies & Weigel 2007). Other 360 examples of genic-based, intrinsic post-zygotic barriers in inter-specific crosses include hybrid 361 inviability in Iris (Iridaceae) (Martin et al., 2008), and hybrid sterility in Mimulus (Phrymaceae) 362 (Fishman & Willis 2001) and Solanum (Solanaceae) (Moyle & Graham 2005). Postzygotic barriers can 363 thus affect hybrids at different stages (e.g., immediately after zygote formation or in later hybrid generations) and with different consequences (e.g., killing or debilitating hybrids, or rendering them 364 365 partially or completely sterile).

366 Among the strongest postzygotic barriers are those due to differences in the structure and 367 number of chromosomes of hybridising species (Levin 2002, Husband 2004, Karlsdóttir et al., 2008, 368 Chapman & Abbott 2010). Hybrid viability can be severely affected by differences in the number of 369 chromosomes between hybridising species, especially in interploidy crosses (Stebbins 1958). 370 Hybridisation between a diploid and a tetraploid is often associated with endosperm failure, a 371 phenomenon known as triploid block (Bretagnolle & Thompson 1995, Köhler et al., 2010). However, 372 triploid block may represent a porous hybridisation barrier (Scott *et al.*, 2013). For example, hybrid 373 inviability associated with triploid block is often asymmetric, meaning that the viability of the hybrid 374 depends on whether a given taxon is the maternal or paternal parent (Ramsey & Schemske 1998). In 375 the British Isles a good example of this asymmetry is Nasturtium x sterile (2n = 48 (45-58),376 Brassicaceae), where attempts to produce this well-established hybrid through artificial crosses are 377 successful only if *N. microphyllum* (2n =64) is used as the maternal parent and *N. officinale* (2n = 32)

as the paternal (Howard & Manton 1946, Stace *et al.*, 2015). The British flora has many examples of

379 young, established hybrids produced from parents of different ploidy, including a number of species 380 of recent origin, in which at least one of the parents is a non-native taxon such as Anchusa 381 ochroleuca x A. officinalis (Boraginaceae), Brassica napus x B. rapa (Brassicaceae), Fallopia 382 baldschuanica x F. japonica (Polygonaceae), Gaultheria mucronata x G. shallon (Ericaceae), Mimulus 383 guttatus x M. luteus (Phrymaceae), Rorippa austriaca x R. sylvestris (Brassicaceae), Rumex crispus x 384 R. frutescens/R. obovatus, Rumex cristatus x R. palustris (Polygonaceae), Senecio squalidus x S. 385 vulgaris (Asteraceae), Spartina alterniflora x S. maritima (Poaceae), and a number of Verbascum 386 (Scrophulariaceae) hybrids (Stace et al., 2015). These taxa provide excellent opportunities to further 387 investigate how inter-ploidy hybrids overcome viability barriers such as triploid block, and to what 388 extent natural hybrid populations have an asymmetric origin.

389 Once a viable hybrid has been produced, a subsequent major challenge is to overcome 390 partial or complete sexual sterility. Interploidy hybrids are often sterile (or have strongly reduced 391 fertility), as a consequence of problems in chromosome pairing during meiosis, leading to 392 unbalanced aneuploid gametes, which are often non-functional (Ramsey & Schemske 1998, Comai 393 2005)(Fig. 2). This pairing problem is expected to be most conspicuous in triploids and hybrids with 394 an odd-number set of chromosomes (Griffiths et al., 2000, Comai 2005), but may also arise in other 395 hybrids in which meiotic pairing results in univalents or odd-numbered multivalents (De Storme & 396 Mason 2014). However, occasionally, interploidy hybrids, such as triploids, produce viable gametes 397 (Ramsey & Schemske 1998, Husband 2004, De Storme & Mason 2014) (Fig. 2). Viable gametes could 398 be produced through multiple routes (Fig. 2), including the production of gametes with the somatic 399 number of chromosomes (unreduced gametes; Köhler et al., 2010, De Storme & Mason 2014, Mason 400 & Pires 2015). Although unreduced gametes are produced at a low rate in non-hybrids (0.0056), 401 their rate of production in hybrids is 50 times higher (0.275) (Ramsey & Schemske 1998). This 402 difference in the rate of production of unreduced gametes is consistent with the observation that 403 polyploids are more common in interspecific hybrid crosses than in crosses within species (Ramsey & 404 Ramsey 2014). Incidentally, the production of unreduced gametes may also help hybrids to bypass 405 the triploid block. For example, mating between the unreduced gametes of a diploid and a tetraploid 406 would produce a hexaploid zygote, without the need to go through a triploid stage. Interestingly, it 407 has been suggested that environmental stress, such as extremes of temperature, may increase the 408 rate at which unreduced, and potentially viable, gametes are formed (De Storme & Mason 2014). 409 Together, this leads to the tantalising idea that global change could facilitate hybridization between 410 diploids and tetraploids—with or without triploids stages—that in the absence of extreme weather 411 events would have remained reproductively isolated.

412 Even in species with the same number of chromosomes, hybrid fertility can be negatively affected by structural differences between parental species including chromosomal re-arrangements 413 414 (e.g., fusions, fissions, deletions, insertions and inversions) (Rieseberg 2001), and differences in the size of homologous chromosomes (Levin 2002). Reduced fertility among species with structural 415 416 chromosomal differences is often caused by pairing irregularities during meiosis, which result in 417 unbalanced, and unviable, gametes (Rieseberg 2001, Levin 2002). The strength of this type of 418 chromosomal barrier thus depends on the level of differentiation between the hybridising genomes, 419 and the extent to which these differences cause pairing irregularities (Levin 2002). Bivalent pairing 420 during meiosis may partly reflect structural similarity of the hybridising genomes, although recent 421 work in both auto- and allopolyploid systems suggests that it may also be under the control of a few 422 genes (Hollister 2015). Chromosomal re-arrangements have been shown to contribute to hybrid sterility in many plant groups (Rieseberg & Carney 1998), and they may be particularly important in 423 424 mediating secondary contact between previously allopatric species (Rieseberg 2001). However, 425 artificial selection on fertility of initially near-sterile hybrids has shown that increased chromosome 426 pairing and more stable meiosis can evolve very rapidly and drastically improve hybrid fertility 427 (Grant 1966). Indeed, some hybrid derivatives between species with divergent chromosome 428 structure have overcome initial reductions in fertility, and formed evolutionarily stable lineages 429 (Rieseberg et al., 1996).

430 6. Persistence of viable but sexually sterile hybrids

Hybrids may prevail even when the initial F1 is highly sterile, if they are able to propagate by other
means (clonally, apomictically, Fig. 3; Table 1) or by evolving higher sexual fertility (Grant 1966). In
addition, polyploid derivatives produced via unreduced gametes (Fig. 2) or somatic mutation can
have increased fertility (Rieseberg 2001, Stathos & Fishman 2014); the evolutionary fate of these
neo-polyploids is discussed in the *Allopolyploid speciation* section.

436 When sexual reproduction is severely impaired, asexual reproduction can allow individuals 437 to persist and spread (Grant 1971). In the British Isles, perennial hybrids that can propagate through 438 clonal reproduction are more widespread than those that cannot (Preston & Pearman 2015; Table 439 2), showing that the capacity for vegetative reproduction is correlated with spatial coverage. 440 Furthermore, analysis of introduced taxa in the recently published alien flora of the British Isles 441 (Stace & Crawley 2015) indicates that approximately 20% of introduced hybrid taxa reproduce 442 exclusively via clonal propagation, compared to only 8% of non-hybrid introduced taxa (Table 2). 443 Examples of sterile hybrid lineages that persist through vegetative propagation include Circaea x 444 intermedia (Onagraceae), Drosera x obovata (Droseraceae) and Stachys x ambigua (Lamiaceae)

445 (Stace *et al.*, 2015). Some of these sterile lineages, for instance the triploid hybrid *Spartina x*

446 townsendii have not spread widely (Strong & Ayres 2013). In contrast other clonal taxa such as the sterile triploid hybrid Mimulus x robertsii have spread more extensively (Preston et al., 2002, Vallejo-447 448 Marín & Lye 2013; Box 2). The ability to disperse over larger geographic areas through vegetative 449 propagation alone, may depend on the characteristics of the habitat. For instance, hybrids growing 450 along dynamic habitats (e.g., rivers and streams) may disperse clonal propagules more extensively 451 than those where dispersal relies on clonal growth on solid ground. For hybrids exploiting dynamic riparian habitats such as those occupied by introduced Mimulus populations in the British Isles, 452 453 global change may facilitate their spread. For example, if extreme weather events result in more 454 intense or frequent flooding, changes in water flow regimes could help spread clonal propagules 455 further. Thus clonal propagation provides a (temporary) escape route to sexual sterility in hybrids, 456 and extreme weather events brought by global change may amplify its effects on the spatial spread 457 of some hybrids.

458 Another way in which asexuality can bypass sexual failure is through the evolution of 459 agamospermy, i.e., the production of seeds without sex (Fig. 3). Agamospermy (often referred to as 460 apomixis, Whitton et al., 2008) is frequently associated with polyploidy and to a lesser extent 461 hybridisation (Grant 1971, Briggs & Walters 1997, Otto & Whitton 2000). Seeds produced through 462 apomixis are genetically identical to the parental plant, and because there is no sexual reproduction 463 they are reproductively isolated from progenitor taxa, so act as 'good' biological species. For this 464 reason, collections of similar apomictic lineages are sometimes called microspecies, agamospecies, 465 agamocomplexes or syngameons (Briggs & Walters 1997). The flora of the British Isles includes numerous examples of cryptohybrids (ancient hybrids stabilised by apomixis) including syngameons 466 467 in the genera Euphrasia (Orobanchaceae), Sorbus (Rosaceae; see Box 1), Rubus (Rosaceae), 468 Hieracium (Asteraceae), Taraxacum (Asteraceae) and Rosa (Rosaceae), all of which are relatively 469 understudied. However, apomixis does not necessarily result in the complete loss of sex. Many 470 facultative apomictic species, such as Sorbus (Box 1), display sexual 'leakiness', and produce viable 471 pollen and ovules that can contribute to subsequent hybridisation events (Ludwig et al., 2013). A 472 further example of variation in reproductive system in apomicts is Hypericum perforatum 473 (Hypericaceae). This species is a facultative agamospermous tetraploid (2n = 32), where sexual 474 reproduction results in diploid offspring, and agamospermy results in tetraploid and hexaploid plants 475 (Barcaccia et al., 2006). H. perforatum hybridises with other taxa, including diploid and tetraploid H. 476 maculatum (2n = 16, 32) with which it produces 3x, 4x, and 5x hybrids, which show variable levels of 477 pollen and seed fertility. Yet, it is not known whether these hybrids reproduce sexually, 478 apomictically, or using a combination of both. An unusual form of reproduction via seeds that has 479 evolved in hybrids with odd-numbered chromosomes, occurs in species such as Rosa canina (2n = 5x

480 = 35). The R. canina complex contains a large number of species in which a permanent state of odd-481 ploidy is maintained by fusion of gametes with complementary genomes (Grant 1971). Permanent 482 chromosomal heterozygosity results from fertilisation and syngamy between pollen with 7 483 chromosomes and ovules with 28 chromosomes (Grant 1971). Determining the incidence and 484 genetic consequences of occasional bouts of sexuality and recombination in hybrids with facultative 485 apomixis and other unusual forms of reproduction is of key importance to understand their fate, as 486 lack of sex can have both potential benefits and costs for the maintenance of hybrids at ecological 487 and evolutionary timescales.

488 One of the potential benefits that arises from bypassing regular sexual reproduction is that the genetic constitution of the hybrids can be preserved ('fixed heterozygosity'), since gene 489 490 combinations are not broken down by meiosis and recombination. Viable F1 hybrids have the 491 capacity to show increased vigour (heterosis) relative to their parents (Barton 2001), and this hybrid 492 vigour can be maintained in the absence of sex and recombination. Therefore, clonality and apomixis 493 may be important not only for the persistence of hybrids, but also as a mechanism to maintain 494 hybrid vigour. In the context of global change, the advantages of asexual reproduction, including 495 fixed heterozygosity, may be balanced by a limited ability to deal with rapidly changing 496 environments due to low genotypic diversity. Yet, in recently formed asexual hybrid populations, 497 genotypic diversity may still occur as remnants of the initial hybridisation event (e.g., in hybrids 498 formed from diverse parental stock) or as a consequence of multiple origins of the same hybrid 499 (Mimulus x robertsii; Vallejo-Marin and Lye 2013), which may provide enough raw material for short-500 term evolutionary change through genotypic selection. Moreover, many highly asexual populations 501 can preserve significant levels of genetic and genotypic diversity (Vallejo-Marín et al., 2010), as even 502 rare bouts of sexual reproduction can significantly increase genetic variation (Bengtsson 2003). 503 Populations of asexual hybrids may also be able to deal with rapid environmental change through 504 phenotypic plasticity (Nicotra et al., 2010). Additionally, epigenetic variation can also contribute to 505 rapid adaptation to environmental challenges, as suggested by the ecologically differentiated 506 populations of the invasive species Fallopia japonica which display abundant epigenetic diversity, 507 despite genetic uniformity (Kilvitis et al., 2014). The variety of mechanisms available to asexual 508 hybrid populations to adapt to different environments may make hybrids more resilient to the rapid 509 modifications brought by global change.

510 **7. Ecological inviability**

Hybrids may suffer from low fitness when they are unable to find a suitable ecological niche, even if
they are not affected by intrinsic developmental problems (Coyne & Orr 2004). For instance, hybrids
which have intermediate phenotypes might be selected against in parental habitats, resulting in

ecological hybrid inviability (Baack et al., 2015). A potential example of such an extrinsic barrier to 514 515 hybridisation in the British Isles flora is the hybrid between Geum urbanum and G. Rivale, (G. x 516 intermedium (Rosaceae). Although the parents of G. x intermedium have broadly overlapping 517 distributions they occupy slightly different habitats: G. urbanum occurs in well-drained soils, in areas 518 partially shaded or sometimes in open disturbed habitats, while G. rivale is usually found in wetter 519 soils and more open habitats (Stace et al., 2015). Experimental crosses have shown that F1 hybrids 520 are viable and fertile, and that seeds produced in zones of contact contain a variety of hybrid 521 genotypes (Ruhsam et al., 2013). Yet, among adult plants only F1s and backcrosses to G. rivale are 522 observed (Ruhsam et al., 2011). Using an experimental garden, Ruhsam et al. (2013) showed that 523 hybrids have no intrinsic low fitness, raising the possibility that ecological selection may be filtering 524 out certain hybrid classes (Ruhsam et al., 2013). The agent of selection is unknown, but it is possible 525 that the intermediate morphology of hybrids makes them poorly suited for establishing in parental 526 environments, and explain why some advanced generation hybrid are rare despite obvious intrinsic 527 post-zygotic barriers. The role that ecological barriers play in preventing the establishment of 528 recently formed hybrids remains a relatively unexplored.

529 IV. Outcomes of hybridisation

530 The long term outcomes of hybridisation can vary from genetic homogenisation of the parental taxa 531 (genetic swamping; Rieseberg & Ellstrand 1993, Wolf et al., 2001), transfer of neutral and adaptive 532 genetic variation across species (introgression and genetic rescue; Rieseberg & Wendel 1993, 533 Ellstrand & Schierenbeck 2000, Baskett & Gomulkiewicz 2011, Gomulkiewicz & Shaw 2013, Hamilton 534 & Miller 2016), and the evolutionary persistence of hybrids as autonomous entities (with or without 535 speciation; Abbott et al., 2013). The persistence of hybrids over evolutionary time can be achieved 536 through four different, non-mutually exclusive pathways: (1) the stabilisation of hybrid zones (Barton 537 & Hewitt 1989, Baack & Rieseberg 2007), (2) spatial displacement of parental taxa, (3) the expansion 538 of hybrids into new ecological niches (hybrids by virtue of transgressive segregation may be pre-539 adapted to more extreme habitats, Rieseberg 1991, Rieseberg et al., 1999), and (4) hybrid speciation 540 (Rieseberg & Willis 2007, Abbott et al., 2013) (Fig. 3).

Global change through its effects on habitat modification, change in climate, and alterations
in ecological interactions, has the potential to alter the outcomes of hybridisation (Campbell &
Wendlandt 2013, Chunco 2014). Probably the most obvious example of such an effect is the
facilitation of hybrid persistence through "hybridisation of the habitat" in which parents occupy
different ecological niches and hybrids a third, intermediate, niche (Anderson 1948, Arnold *et al.*,
2012). Under the bounded hybrid superiority model, in which hybrids enjoy a higher fitness in
intermediate habitats but suffer from lower fitness in parental ones (Moore 1977, Barton & Hewitt

548 1985), the creation and distribution of these intermediate habitats should influence not only hybrid formation, but also their persistence. Despite the intuitive appeal of the effects of global change on 549 550 plant hybridisation outcomes, there are few good examples from the British Isles flora. Nevertheless, 551 one tantalizing case for the potential of hybridisation to result in adaptive introgression comes from 552 hybridisation between non-native species of Rhododendron (Ericaceae). Rhododendron ponticum is 553 widespread in the British Isles and displays the signature of hybridisation with other species, 554 including the more cold-tolerant R. catawbiense (Milne & Abbott 2000). Introgression from R. 555 catawbiense to R. ponticum is more prevalent in colder regions in eastern Scotland than elsewhere 556 in Britain. Given this, Milne and Abbott (2000) raised the possibility that gene flow may confer 557 increased cold-tolerance. However, the hypothesis that hybridisation results in increased cold 558 tolerance in R. ponticum, and that selection (either natural or artificial) is responsible for the 559 geographic distribution of introgression patterns remains to be tested. The success of hybrids under 560 the novel ecological conditions brought by global change will depend on both the availability and type of new habitats and ecological niches, and on whether hybrids are better equipped to deal with 561 these new environments, either via the ability to thrive in "intermediate" habitats or through the 562 563 expression of new adaptive phenotypes, e.g., due to transgressive segregation in hybrids.

564 8. Hybrid speciation

565 Although hybridisation may result in the reversal of speciation ("speciation undone", Grant & Grant 566 2014, Taylor et al., 2006), in some cases new species can be formed following hybridisation events 567 (Mallet 2007, Abbott et al., 2013). Hybrid speciation can occur with or without whole genome duplication (allopolyploid or homoploid speciation, respectively) (Soltis & Soltis, 2009). The use of 568 569 genetic and genomic tools has helped to dramatically increase our understanding of hybrid 570 speciation at both homoploid (Rieseberg 1991, 1997, 2006, Gross et al., 2007) and allopolyploid 571 levels (Hegarty & Hiscock, 2008, Soltis et al., 2014a). Below we present a brief overview of these two 572 modes of hybrid speciation, focusing on those speciation events that have been facilitated by the 573 breakdown of reproductive barriers due to global change, particularly the introduction of non-native 574 species into the British Isles flora.

575 (a) Allopolyploid hybrid speciation

The origin of a new species through hybridisation and polyploidisation can occur rapidly, as taxa with
different chromosome numbers are usually characterised by post-zygotic reproductive barriers
(Stebbins 1971). In principle, a hybrid can give rise to a new species in a single generation via
polyploidisation (Ramsey & Ramsey 2014) leading to a "cataclysmic origin of species" (Dobzhansky
1937, p. 192). Such abrupt speciation is well documented for new British allopolyploid species in the

genera Senecio, Spartina, and Mimulus, all of which have evolved in the last 200 years (Ainouche et
al., 2004, Hegarty et al., 2012, Vallejo-Marín 2012), and all of these cases involve hybrids that owe
their origin to global change.

584 The genus Senecio (ragworts and groundsels) provides some of the best examples of recent 585 hybrid speciation and 'evolution in action' among the British Flora. Within the last 100 years three 586 new polyploid taxa have arisen in the UK as a consequence of hybridization between native tetraploid 587 Senecio vulgaris (common groundsel) and the introduced invasive diploid species S. squalidus (Abbott 588 & Lowe 2004, Hegarty et al., 2012), providing one of the first examples of hybridization induced by 589 global change. Hybridisation between these two species gave rise to three allopolyploid taxa -590 allohexaploid S. cambrensis (Welsh groundsel) tetraploid S. eboracensis (York radiate groundsel) and 591 tetraploid S. vulgaris var. hibernicus (radiate groundsel, a stabilized introgressant form of S. vulgaris). 592 The latter of these hybrid taxa occurs sporadically throughout the UK, but S. eboracensis, first 593 discovered in a York car park, is now probably extinct (Lowe & Abbott 2000, 2003, Abbott et al., 2009). 594 Senecio cambrensis, which was discovered in North Wales in 1948 (Rosser 1955) is locally common in 595 North Wales (Hegarty et al., 2012) and most likely arose following a genome duplication event in the 596 sterile triploid hybrid S. x baxteri, as hexaploid plants with similar morphology to the wild form of S. 597 cambrensis can be produced by treating synthetic triploid S. vulgaris x S. squalidus hybrids with 598 colchicine (Weir & Ingram 1980, Hegarty et al., 2005). In 1982 S. cambrensis was found growing in 599 Edinburgh (Abbott et al., 1983) and subsequent molecular marker analyses revealed that this 600 represented an independent second origin in Scotland (probably during the 1970s) rather than 601 dispersal from Wales (Ashton & Abbott 1992, Harris & Ingram 1992). Unfortunately, the Edinburgh 602 lineage may now be extinct as the species has not been recorded in Edinburgh since 1993 (Abbott & 603 Forbes 2002). Independent origins of recently formed allopolyploids are common as has been shown 604 for the allotetraploid Tragopogon of the USA (Soltis et al., 2004) - themselves another example of 605 hybridization facilitated by global change.

606 Another recently discovered example of rapid allopolyploid speciation in the British Isles is 607 Mimulus peregrinus, which has evolved at least twice, independently in southern Scotland and the 608 Orkney Isles (Vallejo-Marín et al., 2015; Box 2). As in the case of Senecio, M. peregrinus owes its 609 origin to global change bringing together previously isolated species. This species is derived from a 610 sexually-sterile triploid hybrid (M. x robertsii), which is the product of hybridisation between two 611 non-native species: diploid *M. guttatus* (native to North America) and tetraploid *M. luteus* (native to 612 South America). The triploid hybrid persists in the wild and can form large, naturalised populations 613 where it reproduces clonally (Vallejo-Marín & Lye 2013). Unlike the triploid, the allohexaploid 614 derivatives are pollen and seed fertile (Vallejo-Marín 2012). The mechanism for the formation of the allohexaploid from triploid hybrids is unknown, but one likely route is through mating between
unreduced gametes (Husband 2004, Mason & Pires 2015). *Mimulus peregrinus* is an example of how
a sterile hybrid taxon has escaped the sterility barrier through genome duplication to form a fertile
species.

619 Given the young age (<200 years) of allopolyploids such as Senecio cambrensis (Hegarty et 620 al., 2012), Spartina anglica (Ainouche et al., 2004), and M. peregrinus (Vallejo-Marín 2012), their 621 long term persistence is still uncertain. Allopolyploid species often go through severe bottlenecks 622 during their formation (Soltis et al., 2014a), as potentially a single allopolyploid hermaphroditic 623 individual can give rise to a new allopolyploid taxon. This initially low population size may impose 624 severe ecological barriers, such as costs due to matings between individuals with different ploidies 625 (minority cytotype exclusion, Levin 1975; Figure 3), as well as simple stochastic extinction. Yet allopolyploids tend to be formed recurrently, and most allopolyploid species have multiple origins 626 627 (Soltis et al., 2014a). To the extent that these independently originated allopolyploids are inter-628 fertile (Modliszewski & Willis 2012), interpopulation crosses could, in principle, increase genetic and 629 phenotypic variation in nascent lineages, as appears to have been the case in S. cambrensis (Abbott 630 et al., 2007). Indeed, inter-population crosses between accessions from the two known localities for 631 M. peregrinus produce viable and fertile offspring (Vallejo-Marín et al., unpublished), although 632 whether this results in increased phenotypic diversity remains to be established. Determining the 633 degree of phenotypic and ecological diversity of recently allopolyploids seems fundamental to 634 predict their long term fate, particularly as global change continues to alter environments and 635 presents novel challenges to their long term survival.

636 It has long been recognised that hybridisation and polyploidy have the potential to generate 637 ecological novelty (Levin 1983, Otto 2007, Soltis et al., 2014a). The development of genomic tools for 638 recently formed allopolyploids opens the opportunity to take these early hypotheses and investigate 639 the underlying genetic changes associated with ecological innovation. Unlike the case of homoploid 640 hybrids, which have been subject of intense scrutiny at the interphase between ecology and 641 genomics (Yakimowski & Rieseberg 2014), genomic approaches have been rarely used to study the 642 ecology of natural populations of allopolyploids (Ramsey & Ramsey 2014, Soltis et al., 2014b). A rare 643 example of such a study from the British flora is a recent study of the allopolyploid species complex 644 Dactylorhiza majalis, D. traunsteinerii, and D. ebudensis (Orchidaceae) by Paun et al., (2011). 645 Analysis of genome-wide markers (cDNA-AFLPs) showed that these allopolyploids have higher 646 variation in gene expression than their diploid relatives. Paun et al., (2011) suggest that variable 647 gene expression, and potentially having more plastic phenotypes, have facilitated the colonisation of 648 different ecological niches by these three allopolyploid orchids. A similar situation may be

responsible for the success of *S. cambrensis* which also shows altered patterns of gene expression relative to its parents (Hegarty *et al.*, 2006) and also changed patterns of DNA methylation (Hegarty *et al.*, 2011) suggesting the possibility of an epigenetic element to the generation of phenotypic novelty in allopolyploids. Studying the genomic basis and evolution of ecological novelty in polyploids is particularly timely, as we have increasing access to genomic tools for non-model organisms, and because understanding ecological adaptation becomes urgent as populations face the challenge to adapt or perish in the new environments brought by global change.

656 Beyond the earliest stages of allopolyploid speciation, the British Isles flora contains several 657 examples of older allopolyploid taxa, including Mentha spp., Rubus, Euphrasia, Rorippa, and 658 Nasturtium to name just a few (Stace et al., 2015). A particularly intriguing case is the rare Scottish 659 primrose, Primula scotica (2n = 6x = 54, Section Aleuretia), endemic to the north of Scotland and 660 Orkney Isles. Early genetic work showed clear evidence that P. scotica was formed through an 661 allopolyploidisation event (Glover & Abbott 1995). Furthermore, P. scotica is thought to have given rise to another allo-octopolyploid species, *P. scandinavica* (2n = 8x = 72), through a hybridisation 662 event with P. farinosa (Guggisberg et al., 2009), and it may even be involved in the origin of the 14x-663 664 polyploid *P. stricta* (Guggisberg *et al.*, 2006). Currently, the distribution of *P. scotica*'s putative 665 parents, P. farinosa and P. halleri (Guggisberg et al., 2009), and its potential derivatives P. 666 scandinavica and P. stricta, does not overlap with P. scotica, suggesting that they may occupy 667 different ecological niches, a hypothesis that could be tested using ecological niche modelling (e.g., 668 McIntyre 2012). Recent work on polyploid taxa in *Primula* Sect. *Aleuretia* has shown that polyploids in this section tend to have a smaller distribution and narrower niche breadths than diploids 669 670 (Theodoridis et al., 2013), which could indicate that genome duplication is associated with habitat specialisation. *Primula scotica* and other taxa in Section Aleuretia are wonderful examples of how 671 672 allopolyploid taxa can not only persist over evolutionary time, but also continue to speciate and 673 diversify. The development of genomic tools in Primula, including a draft reference genome for P. 674 veris (Nowak et al., 2015), has the potential to elevate this group as a system for the study of 675 evolutionarily established allopolyploids. Notably, the timing of the formation of some of these 676 Primula allopolyploids seems to coincide with large-scale environmental changes brought by 677 historical climate change (e.g., end of glacial maxima), which spurs further speculation on the 678 incidence and importance of allopolyploid speciation in times of environmental upheaval.

Recently formed hybrids, including those potentially associated with global change, are
much more common than the new allopolyploid species they have generated (Stace *et al.*, 2015,
Thomas 2015), posing the question, why? Polyploids are thought to be most commonly formed by
fusion of unreduced gametes (Levin 2013), and the rate of unreduced gamete formation in hybrids

can be extremely high in some plant groups (> 25%, Ramsey & Schemske 1998), indicating ample 683 684 opportunities for their formation. In the case of sterile hybrids, one key hurdle to overcome in order 685 for an allopolyploidisation event to result in a new taxon is the restoration of sexual fertility. In some 686 cases, genome duplication can restore fertility when sterility arises from structural or numeric 687 differences in the parental chromosome sets (Stebbins 1958, Coyne & Orr 2004). However, if sterility 688 is rooted in genic incompatibilities between the parents, genome duplication alone will not be 689 sufficient for fertility restoration. Thus one would expect that those hybrids that have produced 690 allopolyploid species should tend to show few or no genic incompatibilities. This hypothesis can be 691 experimentally tested by inducing polyploidy in sterile hybrids (Hegarty et al., 2013) and assessing 692 their level of fertility (Stathos & Fishman 2014). If synthetic polyploids are as sterile as their parents, 693 then genic incompatibilities can be inferred (Coyne & Orr 2004). If polyploidy restores fertility, the 694 absence of allopolyploids in nature could be explained by ecological or perhaps stochastic causes.

695 Ultimately, determining why allopolyploids are rarer than their hybrid ancestors remains an 696 open question. Their absence is puzzling given the high rate of unreduced gamete production in 697 hybrids (Ramsey and Schemske 1998). Beyond the barrier of overcoming sexual sterility in 698 allopolyploids, other extrinsic post-zygotic mechanisms such as minority cytotype disadvantage 699 (Levin 1975), and ecological competition with their hybrid ancestors, may pose significant barriers to 700 the establishment of recently formed allopolyploids. It is also possible that our records of neo-701 allopolyploids will continue to accumulate as more recently formed hybrids come of age (Thomas 702 2015). Recently formed allopolyploids may be difficult to spot in botanical surveys, particularly in the 703 initial stages when they are quite rare. Large-scale searches for cryptic allopolyploids among recently 704 formed hybrids, using direct or indirect measurements of genome size such as flow cytometry (e.g., 705 Castro et al., 2012), and pollen and stomata size, may provide an effective way to establish if recent 706 allopolyploids are indeed as rare as they appear to be.

707

(b) Homoploid hybrid speciation

708 In contrast to allopolyploid speciation, homoploid hybrid speciation appears less common, although 709 the number of confirmed cases of homoploid hybrid speciation is increasing (Gross & Rieseberg 710 2005, Abbott et al., 2013, Schumer et al., 2014). This apparent rarity of homoploid speciation may 711 therefore simply reflect the difficulty in detecting it (Abbott et al., 2013). Homoploid hybrid 712 speciation requires the evolution of a hybrid that is reproductively isolated from its parents in the 713 absence of differences in ploidy level (Gross & Rieseberg 2005, Yakimowski & Rieseberg 2014). 714 Reproductive isolation between parental and hybrid taxa could thus be based on differences in 715 chromosome structure, ecological divergences, spatial isolation, or a combination of the above 716 (Rieseberg 1997). In the British Isles flora, the only confirmed case of recent homoploid hybrid

22

speciation is *Senecio squalidus* (Oxford ragwort) although the place of origin of the hybrid plants was
Sicily (reviewed in Abbott *et al.*, 2013). *S. squalidus* is a recent homoploid hybrid species, which
evolved in the UK following its introduction from a *S. aethnensis* x *S. chrysanthemifolius* hybrid zone
on Mt Etna ~300 years ago.

721 Senecio aethnensis is endemic to of high altitudes of Mt Etna, whereas S. chrysanthemifolius, a 722 native of Sicily, is more widespread at lower altitudes. At mid altitudes on the volcano the 723 distribution of the two species frequently overlaps leading to the formation of stable hybrid zones 724 around the circumference of the volcano (Brennan et al., 2009). Material from this hybrid zone was 725 introduced to the Oxford Botanic Garden in the early 1700s from where plants subsequently 726 escaped and colonized the masonry of college walls and roadsides. During the industrial revolution 727 the clinker beds of the expanding railway network provided an ideal habitat (akin to the volcanic 728 slopes of Mt Etna) for S. squalidus to thrive and spread. During the next 300 years S. squalidus 729 diverged phenotypically and to a lesser extent genetically in allopatric isolation from its parental 730 species, such that it now meets the criteria for a new homoploid species (James & Abbott 2005, 731 Abbott et al., 2013). S. squalidus and its parental species are divergent in their morphology, 732 flowering phenology, and in ecologically important traits even though they are genetically very 733 similar and completely interfertile (Chapman et al., 2013, Osborne et al., 2013). Nevertheless, the 734 ~300 years of allopatric isolation have allowed S. squalidus to adapt to the cooler climate and non-735 volcanic soils of the UK, resulting in the ecological and phenotypic divergence seen today. 736 Other potential candidates for homoploid hybrids in the British flora are found in the complex 737 assemblage of interspecific hybrids in annual, hemiparasitic Euphrasia. Species of Euphrasia in the 738 British Isles include both diploid and tetraploid taxa, which are strongly reproductively isolated 739 across ploidy levels (Yeo 1968). However, hybrids within ploidy levels are often fertile (Stace et al.,

2015), and Yeo (1968) suggests that some of these hybrids may have produced new hybrid taxa. The
high fertility of within-ploidy hybrids means that for homoploid speciation to occur, reproductive

isolation between parental and hybrid taxa will depend on ecological divergence and spatial isolation

743 (Yeo 1968, Rieseberg 1997). Future studies of *Euphrasia* will determine whether the hypothesis of

homoploid hybrid speciation holds for this group.

Taken at face value, homoploid hybrid speciation seems to be rarer than polyploid hybrid
speciation (Abbott *et al.*, 2013). The apparent higher facility of allopolyploids to form and establish
may be linked, in part, to the reproductive isolation barriers introduced by differences in ploidy level
between derivative and parental taxa. But allopolyploid taxa also have the added feature of
genome-wide redundancy conferred by polyploidisation (Soltis *et al.*, 2014a). It is tempting to
speculate that genomic redundancy is causally linked to evolutionary success in allopolyploids, for

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- 751 example by allowing duplicated subgenomes to differentiate and specialise in different functions
- (subfunctionalisation and neofunctionalisation; Ohno 1970; Soltis et al., 2014b). This higher
- evolutionary potential of allopolyploids may not be immediately realised upon genome duplication
- (or seen in synthetic allopolyploids; Hegarty *et al.*, 2006), but instead may accumulate as lineages
- diverge. The higher incidence of allopolyploids *vs.* homoploid hybrid species may thus reflect an
- evolutionary advantage over the longer term in taxa with highly redundant and malleable genomes.
- 757 Whether recently formed allopolyploids can keep up with rapid global change remains to be seen.

758 V. Outlook

759 Hybridisation and hybrid speciation permeates the evolutionary history of plants, and is certainly not 760 a novel phenomenon. However, the accelerating transformation of the world's biota brought about 761 by global change make the study of hybridisation both current and urgent if we are to understand 762 the consequences of eroding reproductive barriers between evolutionarily distinct lineages. For 763 instance, hybridisation can threaten the conservation of local biodiversity by altering the genetic 764 integrity of native species through introgression, and genetically swamp rare ones (Vilà et al., 2000, 765 Wolf et al., 2001, Brennan et al., 2015, Chown et al., 2015, Taylor et al., 2015). Changes to ecological 766 communities due to hybridisation involving non-native species can also change ecological 767 communities, and hybrids can compete for pollination services and fruit dispersers (Vilà et al., 2000). 768 Moreover, hybrids themselves can become significant invasive pests (Ellstrand & Schierenbeck 2000) 769 with negative effects for local economies and biodiversity. On the other hand, the consequences of 770 hybridisation and its effects on biodiversity need not necessarily always be negative since 771 hybridisation can spur accelerated rates of speciation and thus increase biodiversity (Thomas 2015). 772 More studies are needed to gauge whether the balance of neo-hybridisation is positive or negative. 773 Although the ecological and evolutionary consequences of recently formed hybrids are hard to 774 predict, many hybrids are probably innocuous to local ecosystems as we know them. Regardless of 775 their perceived costs and benefits, recently formed hybrids provide us with the unique opportunity 776 to study, in real time, the breakdown of reproductive barriers and the processes allowing newly 777 formed hybrids to become established, and in some cases speciate (Mallet 2007, Abbott et al., 778 2013). The dynamic and rapidly changing nature of our planet's biota has created a global 779 experiment on hybridisation. We now have the timely opportunity to use this unplanned experiment 780 to study the basic biological phenomena responsible for the birth and death of species. Capitalizing 781 on this opportunity needs a multidisciplinary approach, combining classic taxonomic studies and 782 catalogues of hybrid floras (Stace et al., 2015) with ecological experiments, phylogenetic analysis,

783 quantitative genetics, and bioinformatics to exploit the new genomic resources available for non-

784 model organisms.

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791 **References**

792 Abbott R, Albach D, Ansell S, Arntzen JW, Baird SJE, Bierne N, Boughman JW, Brelsford A, Buerkle

- 793 CA, Buggs R, Butlin RK, Dieckmann U, Eroukhmanoff F, Grill A, Cahan SH, Hermansen JS,
- 794 Hewitt G, Hudson AG, Jiggins C, Jones J, Keller B, Marczewski T, Mallet J, Martínez-Rodríguez
- 795 P, Moest M, Mullen S, Nichols R, Nolte AW, Parisod C, Pfennig K, Rice AM, Ritchie MG, Seifert
- 796 B, Smadja CM, Stelkens R, Szymura JM, Vainola R, Wolf JBW, and Zinner D. 2013.

797 Hybridization and speciation. J. Evol. Biol. 26: 229-246. 10.1111/j.1420-9101.2012.02599.x.

- 798 Abbott RJ, Brennan AC, James JK, Forbes DG, Hegarty MJ, and Hiscock SJ. 2009. Recent hybrid
- origin and invasion of the British Isles by a self-incompatible species, Oxford ragwort (*Senecio squalidus* L., Asteraceae). *Biol. Invasions* 11: 1145-1158. 10.1007/s10530-008-9382-3.

Abbott RJ, and Forbes DG. 2002. Extinction of the Edinburgh lineage of the allopolyploid neospecies,
 Senecio cambrensis Rosser (Asteraceae). Heredity 88: 267-269.

- 803 Abbott RJ, Ireland HE, and Rogers HJ. 2007. Population decline despite high genetic diversity in the
- 804 new allopolyploid species *Senecio cambrensis* (Asteraceae). *Molecular Ecology*, **16**: 1023-1033.
- 805 **Abbott RJ, and Lowe AJ. 2004.** Origins, establishment and evolution of new polyploid species:
- Senecio cambrensis and S. eboracensis in the British Isles. Biological Journal of the Linnean
 Society 82: 467-474.
- Abbott RJ, Noltie HJ, and Ingram R. 1983. The origin and distribution of *Senecio cambrensis* Rosser
 in Edinburgh. *Transactions of the Botanical Society of* Edinburgh 44: 103-106.
- 810 Ainouche M, Baumel A, Salmon A, and Yannic G. 2004. Hybridization, polyploidy and speciation in
- 811 *Spartina* (Poaceae). *New Phytol.* **161**: 165-172. 10.1046/j.1469-8137.2003.00926.x.
- 812 Anderson E. 1948. Hybridization of the habitat. *Evolution*: 1-9.
- 813 Arnold ML. 1997. Natural Hybridization and Evolution. Oxford University Press.

- 814 Arnold ML, Ballerini ES, and Brothers AN. 2012. Hybrid fitness, adaptation and evolutionary
- 815 diversification: lessons learned from Louisiana Irises. *Heredity* **108**: 159-166.

816 10.1038/hdy.2011.65.

- Arnold ML, and Hodges SA. 1995. Are natural hybrids fit or unfit relative to their parents? *Trends in Ecology & Evolution* 10: 67-71. 10.1016/S0169-5347(00)88979-X.
- 819 Ashton PA, and Abbott RJ. 1992. Multiple origins and genetic diversity in the newly arisen
- allopolyploid species, *Senecio cambrensis* Rosser (Compositae). *Heredity* **68**: 25-32.
- 821 Baack E, Melo MC, Rieseberg LH, and Ortiz-Barrientos D. 2015. The origins of reproductive isolation
- 822 in plants. *New Phytologist.* **207**: 968-984.
- Baack EJ, and Rieseberg LH. 2007. A genomic view of introgression and hybrid speciation. *Current* Opinion in Genetics & Development 17: 513-518.
- Bakker FT, Lei D, Yu J, Mohammadin S, Wei Z, Kerke S, Gravendeel B, Nieuwenhuis M, Staats M,
- 826 and Alquezar-Planas DE. 2016. Herbarium genomics: plastome sequence assembly from a
- range of herbarium specimens using an Iterative Organelle Genome Assembly pipeline.
- Biological Journal of the Linnaean Society **117:** 33-43. doi: 10.1111/bij.12642
- 829 Barcaccia G, Arzenton F, Sharbel TF, Varotto S, Parrini P, and Lucchin M. 2006. Genetic diversity
- and reproductive biology in ecotypes of the facultative apomict *Hypericum perforatum* L.

831 *Heredity*, **96**: 322-334.

- 832 Barton NH. 2001. The role of hybridization in evolution. *Molecular Ecology* 10: 551-568.
- Barton NH, and Hewitt G. 1985. Analysis of hybrid zones. *Annual Review of Ecology and Systematics*:
 113-148.
- Barton NH, and Hewitt GM. 1989. Adaptation, speciation and hybrid zones. *Nature* 341: 497-503.
 doi: 10.1038/341497a0
- Baskett ML, and Gomulkiewicz R. 2011. Introgressive hybridization as a mechanism for species
 rescue. *Theoretical Ecology* 4: 223-239.
- Bengtsson BO. 2003. Genetic variation in organisms with sexual and asexual reproduction. *Journal of Evolutionary Biology* 16: 189-199.
- Bomblies K, and Weigel D. 2007. Hybrid necrosis: autoimmunity as a potential gene-flow barrier in
 plant species. *Nature Reviews Genetics* 8: 382-393.
- 843 Bradshaw HD, Wilbert SM, Otto KG, and Schemske DW. 1995. Genetic-mapping of floral traits
- associated with reproductive isolation in monkeyflowers (*Mimulus*). *Nature* **376**: 762-765.
- 845 Brandvain Y, Kenney AM, Flagel L, Coop G, and Sweigart AL. 2014. Speciation and introgression
- between *Mimulus nasutus* and *Mimulus guttatus*. *PLoS Genetics*. 10(6): e1004410. doi:
- 847 10.1371/journal.pgen.1004410

- 848 Brennan AC, Bridle JR, Wang A, Hiscock SJ, and Abbott RJ. 2009. Adaptation and selection in the
- 849 *Senecio* (Asteraceae) hybrid zone on Mount Etna, Sicily. *New Phytologist* **183**: 702-717. doi:
- 850 10.1111/j.1469-8137.2009.02944.x

851 Brennan AC, Woodward G, Seehausen O, Muñoz-Fuentes V, Moritz C, Guelmami A, Abbott RJ, and

- 852 Edelaar P. 2015. Hybridization due to changing species distributions: adding problems or
- solutions to conservation of biodiversity during global change? *Evolutionary Ecology Research*16: 475-491.
- 855 Bretagnolle FA, and Thompson J. 1995. Gametes with the somatic chromosome number:
- 856 mechanisms of their formation and role in the evolution of autopolyploid plants. *New*
- 857 *Phytologist.* **129**:1-22. doi: 10.1111/j.1469-8137.1995.tb03005.x
- 858 Bridle JR, Buckley J, Bodsworth EJ, and Thomas CD. 2013. Evolution on the move: specialization on
- 859 widespread resources associated with rapid range expansion in response to climate change.
- 860 *Proceedings of the Royal Society. B: Biological Sciences.* **281**: 20131800. doi:
- 861 10.1098/rspb.2013.1800.
- Briggs D, and Walters SM. 1997. Plant Variation and Evolution. Cambridge, U.K.: Cambridge
 University Press.
- 864 Brys R, Broeck AV, Mergeay J, and Jacquemyn H. 2014. The contribution of mating system variation
- to reproductive isolation in two closely related *Centaurium* species (Gentianaceae) with a
 generalized flower morphology. *Evolution* 68: 1281-1293.
- 867 Buggs R. 2007. Empirical study of hybrid zone movement. *Heredity* 99: 301-312.
- Campbell DR, and Wendlandt C. 2013. Altered precipitation affects plant hybrids differently than
 their parental species. *American Journal of Botany* 100: 1322-1331. doi: 10.3732/ajb.1200473
- 870 **Castro M, Castro S, and Loureiro J. 2012.** Genome size variation and incidence of polyploidy in
- 871 Scrophulariaceae *sensu lato* from the Iberian Peninsula. *AoB plants*. **2012**: pls037. doi:
- 872 10.1093/aobpla/pls037
- Chapman MA, and Abbott RJ. 2010. Introgression of fitness genes across a ploidy barrier. *New Phytologist.* 186: 63-71. doi: 10.1111/j.1469-8137.2009.03091.x
- Chapman MA, Hiscock SJ, and Filatov DA. 2013. Genomic divergence during speciation driven by
 adaptation to altitude. *Molecular Biology and Evolution* 30: 2553-2567. doi:
- 877 10.1093/molbev/mst168
- 878 Chown SL, Hodgins KA, Griffin PC, Oakeshott JG, Byrne M, and Hoffmann AA. 2015. Biological
- 879 invasions, climate change and genomics. *Evolutionary Applications* **8**: 23-46.
- 880 10.1111/eva.12234.

881 Chunco AJ. 2014. Hybridization in a warmer world. *Ecology and Evolution* 4: 2019-2031. doi: 882 10.1002/ece3.1052 883 Cleland EE, Chuine I, Menzel A, Mooney HA, and Schwartz MD. 2007. Shifting plant phenology in 884 response to global change. Trends in Ecology and Evolution 22: 357-365. 885 Clement EJ, and Foster M. 1994. Alien Plants of the British Isles. London: Botanical Society of the 886 British Isles. 887 **Comai L. 2005.** The advantages and disadvantages of being polyploid. *Nature Reviews Genetics* 6: 888 836-846. 889 Coyne JA, and Orr HA. 2004. Speciation. Sunderland, Massachusetts, U.S.A.: Sinauer Associates. 890 De Storme N, Mason A. 2014. Plant speciation through chromosome instability and ploidy change: 891 Cellular mechanisms, molecular factors and evolutionary relevance. Current Plant Biology 1: 10-33. 892 893 Dobzhansky TG. 1937. Genetics and the Origin of Species. New York: Columbia University Press. 894 Dodsworth S. 2015. Genome skimming for next-generation biodiversity analysis. Trends in Plant 895 Sciences 20: 525-527. Ellstrand NC, and Schierenbeck KA. 2000. Hybridization as a stimulus for the evolution of 896 897 invasiveness in plants? Proceedings of the National Academy of Sciences U. S. A. 97: 7043-7050. 898 Ellstrand NC, and Elam DR. 1993. Population genetic consequences of small population size: 899 implications for plant conservation. Annual Review of Ecology and Systematics 24: 217-242. 900 Fay MF 2015. British and Irish orchids in a changing world. Curtis's Botanical Magazine 32: 3-23. 901 Fishman L, and Willis JH. 2001. Evidence for Dobzhansky-Muller incompatibilities contributing to the 902 sterility of hybrids between Mimulus guttatus and M. nasutus. Evolution 55: 1932-1942. 903 Garroway CJ, Bowman J, Cascaden TJ, Holloway GL, Mahan CG, Malcolm JR, Steele MA, Turner G, 904 and Wilson PJ. 2010. Climate change induced hybridization in flying squirrels. Global Change 905 *Biology* **16**: 113-121. 906 Glover BJ, and Abbott RJ. 1995. Low genetic diversity in the Scottish endemic Primula scotica Hook. 907 New Phytologist **129**: 147-153. 908 Gomulkiewicz R, and Shaw RG. 2013. Evolutionary rescue beyond the models. Philosophical 909 Transactions of the Royal Society of London. B: Biological Sciences. 368: 20120093. doi: 910 10.1098/rstb.2012.0093. 911 Grant PR, and Grant BR. 2014. Evolutionary biology: Speciation undone. Nature 507: 178-179. 912 Grant V. 1966. Selection for vigor and fertility in the progeny of a highly sterile species hybrid in 913 Gilia. Genetics 53: 757-776. 914 Grant V. 1971. Plant Speciation. New York: Columbia University Press.

- 915 Griffiths AJ, Miller JH, Lewontin RC, and Suzuki DT. 2000. An Introduction to Genetic Analysis. New
 916 York: WH Freeman.
- 917 Gross BL, and Rieseberg LH. 2005. The ecological genetics of homoploid hybrid speciation. *Journal of* 918 *Heredity.* 96: 241-252. 10.1093/jhered/esi026
- 919 Gross BL, Turner KG, and Rieseberg LH. 2007. Selective sweeps in the homoploid hybrid species
 920 *Helianthus deserticola*: evolution in concert across populations and across origins. *Molecular* 921 *Ecology* 16: 5246-5258. 10.1111/j.1365-294X.2007.03574.x
- Guggisberg A, Mansion G, Kelso S, and Conti E. 2006. Evolution of biogeographic patterns, ploidy
 levels, and breeding systems in a diploid–polyploid species complex of Primula. *New Phytologist*
- **171**: 617-632.
- Guggisberg A, Mansion G, and Conti E. 2009. Disentangling reticulate evolution in an arctic–alpine
 polyploid complex. *Systematic Biology*. 58: 55-73. doi: 10.1093/sysbio/syp010
- Hamilton JA, and Miller JM. 2016. Adaptive introgression: A resource for management and genetic
 conservation in a changing climate. *Conservation Biology* 30: 33–41. doi: 10.1111/cobi.12574
- Harris SA, and Ingram R. 1992. Molecular systematics of the genus *Senecio* L. 1. Hybridization in a
 British polyploid complex. *Heredity* 69: 1-10.
- 931 Hegarty MJ, Batstone TOM, Barker GL, Edwards KJ, Abbott RJ, and Hiscock SJ. 2011. Nonadditive
- 932 changes to cytosine methylation as a consequence of hybridization and genome duplication in
 933 Senecio (Asteraceae). Molecular Ecology, 20: 105-113.
- Hegarty MJ, Jones JM, Wilson ID, Barker GL, Coghill JA, Sanchez-Baracaldo P, Buggs RJA, Liu G,
- 935 **Abbott RJ, Edwards KJ, Hiscock SJ. 2005.** Development of anonymous cDNA microarrays to
- study changes to the *Senecio* floral transcriptome during hybrid speciation. *Molecular Ecology* **14**: 2493-2510.
- 938 Hegarty MJ, Barker GL, Wilson ID, Abbott RJ, Edwards KJ, Hiscock SJ. 2006 Transcriptome shock
- 939 after interspecific hybridization in *Senecio* is ameliorated by genome duplication. *Current Biology* 16:940 1652-1659.
- Hegarty MJ, Hiscock SJ. 2008. Genomic clues to the evolutionary success of polyploid plants. *Current*Biology 18: R435-R444.
- 943 Hegarty MJ, Abbott RJ, and Hiscock SJ. 2012. Allopolyploid speciation in action: The origins and
- 944 evolution of *Senecio cambrensis*. Pages 245-270 In: Soltis PS, and Soltis DE, eds. *Polyploidy and*
- 945 *genome evolution.* Berlin: Springer-Verlag. 245-270.
- 946 Hegarty M, Coate J, Sherman-Broyles S, Abbott R, Hiscock S, and Doyle J. 2013. Lessons from
- 947 natural and artificial polyploids in higher plants. Cytogenetics and Genome Research. 140: 204-
- 948 225. doi: 10.1159/000353361

- Higgins SI, and Richardson DM. 1999. Predicting plant migration rates in a changing world: the role
 of long-distance dispersal. *American Naturalist* 153: 464-475.
- 951 Hiscock SJ, Doughty J, and Dickinson HG. 1998. Unilateral incompatibility and the S (self-
- 952 incompatibility) locus. In: Owens SJ and Rudall PJ. Reproductive Biology in Systematics,
- 953 *Conservation and Economic Botany.* Richmond, Surrey, UK: Royal Botanic Gardens, Kew. 31-46.
- Hoffmann AA, and Sgrò CM. 2011. Climate change and evolutionary adaptation. *Nature* 470: 479485.
- Hollister JD. 2015. Polyploidy: adaptation to the genomic environment. *New Phytologist* 205: 10341039. doi: 10.1111/nph.12939
- Howard DJ. 1999. Conspecific sperm and pollen precedence and speciation. *Annual Review of Ecology and Systematics* 30: 109-132.
- Howard H, and Manton I. 1946. Autopolyploid and allopolyploid watercress with the description of
 a new species. Annals of Botany 10: 1-13.
- Hulme PE, Bacher S, Kenis M, Klotz S, Kühn I, Minchin D, Nentwig W, Olenin S, Panov V, and Pergl
 J. 2008. Grasping at the routes of biological invasions: a framework for integrating pathways
- 964 into policy. *Journal of Applied Ecology* **45**: 403-414.
- Husband BC. 2004. The role of triploid hybrids in the evolutionary dynamics of mixed-ploidy
 populations. *Biological Journal of the Linnean Society* 82: 537-546.
- Husband BC, Schemske DW, Burton TL, and Goodwillie C. 2002. Pollen competition as a unilateral
 reproductive barrier between sympatric *Chamerion angustifolium*. *Proceedings of the Royal* Society of London Series B: Biological Sciences 269: 2565-2571.
- 970 James JK, and Abbott RJ. 2005. Recent, allopatric, homoploid hybrid speciation: The origin of
- 971 *Senecio squalidus* (Asteraceae) in the British Isles from a hybrid zone on Mount Etna, Sicily.
- 972 *Evolution* **59**: 2533-2547.
- 973 Karlsdóttir L, Hallsdóttir M, Thórsson AT, and Anamthawat-Jónsson K. 2008. Characteristics of
 974 pollen from natural triploid *Betula* hybrids. *Grana* 47: 52-59.

975 Kilvitis HJ, Alvarez M, Foust CM, Schrey AW, Robertson M., and Richards CL, 2014. Ecological

- 976 epigenetics. *In*: Landy CR, and Aubin-Hort N (eds), *Ecological Genomics: Ecology and the*
- 977 *Evolution of Genes and Genomes.* Springer Netherlands. pp. 191-210.
- 978 Knobloch IW. 1972. Intergeneric hybridization in flowering plants. *Taxon*: 97-103.
- 879 Köhler C, Scheid OM, and Erilova A. 2010. The impact of the triploid block on the origin and
- 980 evolution of polyploid plants. *Trends in Genetics* **26**: 142-148. 10.1016/j.tig.2009.12.006.
- 981 Levin DA. 1975. Minority cytotype exclusion in local plant populations. *Taxon*: 35-43.

982 Levin DA. 1983. Polyploidy and novelty in flowering plants. American Naturalist: 1-25.

983 http://www.jstor.org/stable/2461003

- Levin DA. 2002. *The Role of Chromosomal Change in Plant Evolution*. Oxford, UK: Oxford University
 Press.
- Levin DA. 2013. The timetable for allopolyploidy in flowering plants. *Annals of Botany*, 112: 12011208.
- Lousley J. 1946. A new hybrid Senecio from the London area. *Report of the Botanical Society and Exchange Club of the British Isles* 12: 869-874.
- Lowe AJ, and Abbott RJ. 2000. Routes of origin of two recently evolved hybrid taxa: Senecio vulgaris
 var. hibernicus and York radiate groundsel (Asteraceae). American Journal of Botany 87: 1159 1167.
- Lowe AJ, and Abbott RJ. 2003. A new British species, *Senecio eboracensis* (Asteraceae), another
 hybrid derivative of *S. vulgaris* L. and *S. squalidus* L. *Watsonia* 24: 375-388.
- Lowry DB, Modliszewski JL, Wright KM, Wu CA, and Willis JH. 2008. The strength and genetic basis
 of reproductive isolating barriers in flowering plants. *Philosophical Transactions of the Royal Society B-Biological Sciences* 363: 3009-3021.
- 998 Ludwig S, Robertson A, Rich TCG, Djordjevic M, Cerovic R, Houston L, Harris SA, Hiscock SJ (2013)
- 999 Breeding systems, hybridization and continuing evolution in Avon Gorge Sorbus. Annals of
 1000 Botany 111: 563-575.
- Mable BK. 2013. Polyploids and hybrids in changing environments: winners or losers in the struggle
 for adaptation? *Heredity* 110: 95-96. doi: 10.1038/hdy.2012.105
- Mack RN. 2005. Predicting the identity of plant invaders: future contributions from horticulture.
 HortScience 40: 1168-1174.
- 1005 Mallet J. 2007. Hybrid speciation. *Nature* 446: 279-283. doi: 10.1038/nature05706.
- 1006 Marren P. 1999. Britain's Rare Flowers. London: A & C Black Publishers Ltd.
- Martin NH, Sapir Y, and Arnold ML. 2008. The genetic architecture of reproductive isolation in
 Louisiana irises: Pollination syndromes and pollinator preferences. *Evolution* 62: 740-752.
- Mason AS, and Pires JC. 2015. Unreduced gametes: meiotic mishap or evolutionary mechanism?
 Trends in Genetics 31: 5-10. doi: 10.1016/j.tig.2014.09.011
- 1011 Mayr E. 1942. Systematics and the Origin of Species, from the Viewpoint of a Zoologist. Cambridge,
- 1012 Massachusetts: Harvard University Press.
- 1013 McIntyre PJ. 2012. Polyploidy associated with altered and broader ecological niches in the *Claytonia*

1014 *perfoliata* (Portulacaceae) species complex. *American Journal of Botany* **99**: 655-662. doi:

1015 10.3732/ajb.1100466.

- 1016 Milne RI, and Abbott RJ. 2000. Origin and evolution of invasive naturalized material of
- 1017 *Rhododendron ponticum* L. in the British Isles. *Molecular Ecology* **9**: 541-556. doi:
- 1018 10.1046/j.1365-294x.2000.00906.x.
- Modliszewski JL, and Willis JH. 2012. Allotetraploid *Mimulus sookensis* are highly interfertile despite
 independent origins. *Molecular Ecology* 21: 5280-5298. doi: 10.1111/j.1365-294X.2012.05706.x.
- Mooney H, and Cleland E. 2001. The evolutionary impact of invasive species. *Proceedings of the National Academy of Sciences, U. S. A.* 98: 5446-5451. 10.1073/pnas.091093398.
- Moore WS. 1977. An evaluation of narrow hybrid zones in vertebrates. *Quarterly Review of Biology* 52: 263-277.
- Morales CL, and Traveset A. 2009. A meta-analysis of impacts of alien vs. native plants on pollinator
 visitation and reproductive success of co-flowering native plants. *Ecology Letters* 12: 716-728.
- 1027 Moyle LC, and Graham EB. 2005. Genetics of hybrid incompatibility between Lycopersicon
- 1028 esculentum and L. hirsutum. Genetics **169**: 355-373. doi: 10.1534/genetics.104.029546
- Moyle LC, Olson MS, and Tiffin P. 2004. Patterns of reproductive isolation in three angiosperm
 genera. *Evolution* 58: 1195-1208.
- Myers JH, and Bazely D. 2003. Ecology and Control of Introduced Plants. Cambridge, U.K.: Cambridge
 University Press.
- 1033 Nicotra AB, Atkin OK, Bonser SP, Davidson AM, Finnegan EJ, Mathesius U, Poot P, Purugganan MD,
- 1034 Richards CL, Valladares F, and van Kleunen M. 2010. Plant phenotypic plasticity in a changing
 1035 climate. *Trends in Plant Science* 15: 684-692.
- Nowak MD, Russo G, Schlapbach R, Huu CN, Lenhard M, and Conti E. 2015. The draft genome of
 Primula veris yields insights into the molecular basis of heterostyly. *Genome Biology* 16: 12 doi:
 10.1186/s13059-014-0567-z
- 1039 **Ohno S. 1970.** *Evolution by Gene Duplication*. Springer-Verlag. New York.
- 1040 Osborne OG, Batstone TE, Hiscock SJ, and Filatov DA. 2013. Rapid speciation with gene flow
- following the formation of Mt. Etna. *Genome Biology and Evolution* 5: 1704-1715. doi:
 10.1093/gbe/evt127
- 1043 Otto SP. 2007. The evolutionary consequences of polyploidy. *Cell* 131: 452-462. doi:
- 1044 10.1016/j.cell.2007.10.022
- 1045 Otto SP, and Whitton J. 2000. Polyploid incidence and evolution. Annual Review of Genetics 34: 401-
- 1046 437. 10.1146/annurev.genet.34.1.401
- 1047 Paun O, Bateman RM, Fay MF, Luna JA, Moat J, Hedren M, and Chase MW. 2011. Altered gene
- 1048 expression and ecological divergence in sibling allopolyploids of *Dactylorhiza* (Orchidaceae).
- 1049 BMC Evolutionary Biology **11**: 113. doi: 10.1186/1471-2148-11-113

1050 **Preston CD, and Croft JM. 1998.** Britain's changing aquatic flora. *British Wildlife* **10**: 18-28.

- Preston CD, Pearman DA, and Dines TD. 2002. New Atlas of the British and Irish Flora. Oxford, UK:
 Oxford University Press.
- Preston CD, and Pearman DA. 2015. Plant hybrids in the wild: evidence from biological recording.
 Biological Journal of the Linnean Society. 115: 555-572.
- 1055 Ramsey J, Bradshaw HD, and Schemske DW. 2003. Components of reproductive isolation between
 1056 the monkeyflowers *Mimulus lewisii* and *M. cardinalis* (Phrymaceae). *Evolution* 57: 1520-1534.
- 1057 Ramsey J, and Ramsey TS. 2014. Ecological studies of polyploidy in the 100 years following its
- discovery. *Philosophical Transactions of the Royal Society of London B: Biological Sciences* 369:
 20130352. doi: 10.1098/rstb.2013.0352
- 1060 **Ramsey J, and Schemske DW. 1998.** Pathways, mechanisms, and rates of polyploid formation in

1061 flowering plants. *Annual Review of Ecology and Systematics* **29**: 467-501.

- Rich TCG, Houston L, Robertson A, and Proctor MCF. 2010. Whitebeams, Rowans and Service Trees
 of Britain and Ireland. A monograph of British and Irish Sorbus L. London: Botanical Society of
 the British Isles.
- 1065 Rich TCG, and Robertson A. 2015. Sorbus L. In: Hybrid Flora of the British Isles. Stace CA, Preston CD,
 1066 and Pearman DA (eds). Bristol, U.K.: Botanical Society of Britain and Ireland.
- 1067 Rieseberg LH. 1991. Homoploid reticulate evolution in *Helianthus* (Asteraceae). Evidence from
 1068 ribosomal genes. *American Journal of Botany* 78: 1218-1237. doi: 10.2307/2444926.
- 1069 Rieseberg LH. 1997. Hybrid origins of plant species. *Annual Review of Ecology and Systematics* 28:
 1070 359-389. 10.1146/annurev.ecolsys.28.1.359
- 1071 Rieseberg LH. 1995. The role of hybridization in evolution. Old wine in new skins. *American Journal* 1072 of Botany 82: 944-953. doi: 10.2307/2445981
- 1073 Rieseberg LH. 2001. Chromosomal rearrangements and speciation. *Trends in Ecology & Evolution* 16:
 1074 351-358. doi: 10.1016/S0169-5347(01)02187-5
- 1075 Rieseberg LH. 2006. Hybrid speciation in wild sunflowers. Annals of the Missouri Botanical Garden
 1076 93: 34-48.
- 1077 Rieseberg LH, Archer MA, and Wayne RK. 1999. Transgressive segregation, adaptation and
 1078 speciation. *Heredity* 83: 363-372.
- 1079 **Rieseberg LH, and Blackman BK. 2010.** Speciation genes in plants. *Annals of Botany* **106**: 439-455.
- 1080 doi: 10.1093/aob/mcq126.
- 1081 Rieseberg LH, and Brunsfeld SJ. 1992. Molecular evidence and plant introgression. Pages 151-176 In:
 1082 Soltis PS, Soltis DE, and Doyle JJ. *Molecular systematics of plants*. New York, U.S.A.: Springer,
 1083 151-176.

- 1084 **Rieseberg LH, and Carney SE. 1998.** Plant hybridization. *New Phytologist* **140**: 599-624.
- 1085 Rieseberg LH, and Ellstrand NC. 1993. What can molecular and morphological markers tell us about
 1086 plant hybridization. *Critical Reviews in Plant Sciences* 12: 213-241. doi: 10.1080/713608045
- 1087 **Rieseberg LH, Sinervo B, Linder CR, Ungerer MC, and Arias DM. 1996.** Role of gene interactions in
- hybrid speciation: Evidence from ancient and experimental hybrids. *Science* 272: 741-745. doi:
 10.1126/science.272.5262.741
- 1090 Rieseberg LH, and Wendel JF. 1993. Introgression and its Consequences in Plants. In: Harrison R, ed.
 1091 *Hybrid Zones and the Evolutionary Process*. New York: Oxford University Press. 70-109.
- 1092 Rieseberg LH, and Willis JH. 2007. Plant speciation. Science 317: 910-914.
- 1093 **Roberts RH. 1964.** *Mimulus* hybrids in Britain. *Watsonia* **6**: 70-75.

1094 Robertson A, Rich TCG, Allen AM, Houston L, Roberts C, Bridle JR, Harris SA, and Hiscock, SJ. 2010.

- 1095 Hybridization and polyploidy as drivers of continuing evolution and speciation in *Sorbus* L.
- 1096 (Rosaceae). *Molecular Ecology* **19**: 1675-1690.
- 1097 **Rosser EM. 1955.** A new British species of *Senecio*. *Watsonia* **3**: 228-232.
- Ruhsam M, Hollingsworth P, and Ennos R. 2011. Early evolution in a hybrid swarm between
 outcrossing and selfing lineages in *Geum. Heredity* 107: 246-255.
- Ruhsam M, Hollingsworth PM, and Ennos RA. 2013. Patterns of mating, generation of diversity, and
 fitness of offspring in a *Geum* hybrid swarm. *Evolution* 67: 2728-2740.
- Schumer M, Rosenthal GG, and Andolfatto P. 2014. How common is homoploid hybrid speciation?
 Evolution 68: 1553-1560. doi: 10.1111/evo.12399
- Scopece G, Musacchio A, Widmer A, and Cozzolino S. 2007. Patterns of reproductive isolation in
 Mediterranean deceptive orchids. *Evolution* 61: 2623-2642.
- 1106 Scott RJ, Tratt JL, and Bolbol A. 2013. Seed development in interploidy hybrids. In Chen ZJ and
- 1107 Birchler JA, eds. *Polyploid and Hybrid Genomics*. Oxford, UK: John Wiley & Sons. 271-290.
- 1108 Sherry RA, Zhou X, Gu S, Arnone JA, 3rd, Schimel DS, Verburg PS, Wallace LL, and Luo Y. 2007.
- Divergence of reproductive phenology under climate warming. *Proceedings of the National Academy of Sciences, U. S. A.* **104**: 198-202.
- Silvertown J. 2011. Demons in Eden: The Paradox of Plant Diversity. Chicago, U.S.A.: University of
 Chicago Press.
- 1113 Soltis DE, Soltis PS, Pires JC, Kovarik A, Tate JA, and Mavrodiev E. 2004. Recent and recurrent
- 1114 polyploidy in *Tragopogon* (Asteraceae): Cytogenetic, genomic and genetic comparisons.

1115 Biological Journal of the Linnean Society **82**: 485-501.

Soltis PS, and Soltis DE. 2009. The role of hybridization in plant speciation. *Annual Review of Plant Biology* 60: 561-588.

- 1118 Soltis DE, Visger CJ, and Soltis PS. 2014a. The polyploidy revolution then...and now: Stebbins
- 1119 revisited. *American Journal of Botany*. **101**: 1057-1078. doi: 10.3732/ajb.1400178
- 1120 Soltis PS, Liu X, Marchant DB, Visger CJ, and Soltis DE. 2014b. Polyploidy and novelty: Gottlieb's
- 1121 legacy. *Philosophical Transactions of the Royal Society of London. B: Biological Sciences.* **369**:

1122 20130351. doi: 10.1098/rstb.2013.0351

- 1123 Stace CA. 2010. New Flora of the British Isles. Cambridge: Cambridge University Press.
- 1124 **Stace CA, and Crawley MJ. 2015.** *Alien Plants.* London, U.K.: William Collins.
- Stace CA, Preston CD, and Pearman DA. 2015. *Hybrid flora of the British Isles*. Bristol, U.K.: Botanical
 Society of Britain and Ireland.
- 1127 Stathos A, and Fishman L. 2014. Chromosomal rearrangements directly cause underdominant F1
- 1128 pollen sterility in *Mimulus lewisii–Mimulus cardinalis* hybrids. *Evolution* **68**: 3109-3119.
- 1129 **Stebbins GL. 1950.** Variation and Evolution in Plants. New York: Columbia University Press.
- 1130 **Stebbins GL. 1958.** The inviability, weakness, and sterility of interspecific hybrids. *Advances in*
- 1131 *Genetics Incorporating Molecular Genetic Medicine* 9: 147-215. doi: 10.1016/S00651132 2660(08)60162-5.
- Stebbins GL. 1959. The role of hybridization in evolution. *Proceedings of the American Philosophical Society* 103: 231-251.
- 1135 **Stebbins GL. 1971.** *Chromosomal Evolution in Higher Plants.* London: Edward Arnold.
- Straub SC, Parks M, Weitemier K, Fishbein M, Cronn RC, and Liston A. 2012. Navigating the tip of
 the genomic iceberg: Next-generation sequencing for plant systematics. *American Journal of*
- 1138 Botany. 99: 349-364. doi: 10.3732/ajb.1100335
- Strong DR, and Ayres DR. 2013. Ecological and evolutionary misadventures of *Spartina*. *Annual Review of Ecology, Evolution, and Systematics* 44: 389-410.
- 1141 **Taylor E, Boughman J, Groenenboom M, Sniatynski M, Schluter D, and Gow J. 2006.** Speciation in
- 1142 reverse: morphological and genetic evidence of the collapse of a three-spined stickleback

1143 (*Gasterosteus aculeatus*) species pair. *Molecular Ecology* **15**: 343-355.

- Taylor SA, Larson EL, and Harrison RG. 2015. Hybrid zones: windows on climate change. *Trends in Ecology & Evolution* 30: 198-406. doi: 10.1016/j.tree.2015.04.010
- 1146 **Taylor SJ, Arnold M, and Martin NH. 2009.** The genetic architecture of reproductive isolation in
- 1147 Louisiana irises: hybrid fitness in nature. *Evolution* **63**: 2581-2594.
- 1148 **Theodoridis S, Randin C, Broennimann O, Patsiou T, and Conti E. 2013.** Divergent and narrower
- 1149 climatic niches characterize polyploid species of European primroses in *Primula* sect. *Aleuritia*.
- 1150 *J. Biogeogr.* **40**: 1278-1289.

- Thomas CD. 2015. Rapid acceleration of plant speciation during the Anthropocene. *Trends in Ecology & Evolution* 30: 448-455.
- 1153 **Twyford AD, and Ennos RA. 2012.** Next-generation hybridization and introgression. *Heredity* **108**:

1154 179-189. 10.1038/hdy.2011.68.

- 1155 Ubsdell R. 1979. Studies on variation and evolution in *Centaurium erythraea* Rafn and *C. littorale* (D.
 1156 Turner) Gilmour in the British Isles, 3. Breeding systems, floral biology and general discussion.
 1157 Watsonia 12: 225-232.
- 1158 Vallejo-Marín M. 2012. *Mimulus peregrinus* (Phrymaceae): A new British allopolyploid species.
 1159 *PhytoKeys* 14: 1-14. 10.3897/phytokeys.14.3305.
- 1160 Vallejo-Marín M, and Lye GC. 2013. Hybridisation and genetic diversity in introduced *Mimulus*1161 (Phrymaceae). *Heredity* 110: 111-122.
- Vallejo-Marín M, Buggs RJ, Cooley AM, and Puzey JR. 2015. Speciation by genome duplication:
 Repeated origins and genomic composition of the recently formed allopolyploid species
- 1164 *Mimulus peregrinus. Evolution* **69**: 1487-1500.
- 1165 Vallejo-Marín M, Cooley AM, Qi MLY, Folmer M, McKain MR, and Puzey JR. *in press*. Strongly
 asymmetric hybridization barriers shape the origin of a new polyploid species and its hybrid
 ancestor. *American Journal of Botany*.
- Vallejo-Marín M, Dorken ME, and Barrett SCH. 2010. The ecological and evolutionary consequences
 of clonality for plant mating. *Annual Review of Ecology, Evolution, and Systematics* 41: 193-213.
- 1170 Vickery RK. 1959. Barriers to gene exchange within *Mimulus guttatus* (Scrophulariaceae). *Evolution*1171 13: 300-310.
- 1172 Vilà M, Weber E, and Antonio CMD. 2000. Conservation implications of invasion by plant
 1173 hybridization. *Biological Invasions* 2: 207-217.
- 1174 Walther G, Roques A, Hulme PE, Sykes MT, Pysek P, Kuehn I, Zobel M, Bacher S, Botta-Dukat Z,
- 1175 Bugmann H, Czucz B, Dauber J, Hickler T, Jarosik V, Kenis M, Klotz S, Minchin D, Moora M,
- 1176 Nentwig W, Ott J, Panov VE, Reineking B, Robinet C, Semenchenko V, Solarz W, Thuiller W,
- 1177 Vila M, Vohland K, and Settele J. 2009. Alien species in a warmer world: risks and
- 1178 opportunities. *Trends in Ecology and Evolution* **24**: 686-693. doi: 10.1016/j.tree.2009.06.008.
- Weir J, and Ingram R. 1980. Ray morphology and cytological investigations of *Senecio cambrensis* Rosser. *New Phytologist* 86: 237-241.
- 1181 Whitton J, Sears CJ, Baack EJ, and Otto SP. 2008. The dynamic nature of apomixis in the
- angiosperms. *International Journal of Plant Sciences* **169**: 169-182.
- Widmer A, Lexer C, and Cozzolino S. 2009. Evolution of reproductive isolation in plants. *Heredity* 102: 31-38.

- 1185 Wolf DE, Takebayashi N, and Rieseberg LH. 2001. Predicting the risk of extinction through
- 1186 hybridization. *Conservation Biology* **15**: 1039-1053. doi: 10.1046/j.1523-
- 1187 1739.2001.0150041039.x.
- 1188 Yakimowski SB, and Rieseberg LH. 2014. The role of homoploid hybridization in evolution: a century
- 1189 of studies synthesizing genetics and ecology. *American Journal of Botany* **101**: 1247-1258. doi:
- 1190 10.3732/ajb.1400201
- 1191 Yeo P. 1968. The evolutionary significance of the speciation of *Euphrasia* in Europe. *Evolution*: 736-
- 1192 747.

1193 Figure Legends

1194

Figure 1. Reproductive isolation barriers preventing the formation and establishment of hybrids.
 Reproductive isolation barriers are usually divided in pre-zygotic and post-zygotic. Notice that in
 plants, some barriers such as pollen precedence act after mating (pollination) but before zygote
 formation.

- 1199
- 1200 **Figure 2.** Meiosis in hybrids is adversely affected by differences in chromosome number and
- 1201 structure between parental species. The result will generally be gamete inviability, but, rarely,
- 1202 gametes with atypical chromosome numbers (aneuploid and unreduced gametes) may be produced
- 1203 and viable. The figure shows ways in which viable gametes could be produced in a hypothetical
- 1204 triploid hybrid (2n = 3x). Unpaired chromosomes during meiosis (in both univalents and
- 1205 multivalents) migrate to either cell pole at random, resulting in unbalanced gametes (top pathway).
- 1206 Most of these unbalanced gametes will be inviable, causing a very severe reduction in fertility. A
- small fraction of these gametes may by chance end up with the parental cytotype, and are thus
- 1208 viable. Another small fraction may be viable despite carrying new aneuploid combinations.
- 1209 Occasionally, gametes may be produced with the entire set of chromosomes present in the hybrid
- 1210 (unreduced gametes), which may be subsequently involved in polyploid formation.
- 1211
- 1212 Figure 3. Potential fate of viable hybrids that are able to persist over ecological or evolutionary
- 1213 timescales after their origin and initial establishment.

Table 1. Examples of the "Significant Eight" alien hybrid complexes of the British Flora recognised by Stace & Crawley (2015). Introduced taxa (neophytes) are indicated with *. Data compiled by Stace & Crawley (2015) and Stace *et al.* (2015).

Hybrid	Parents		Sexual Fertility	Clonal reproduction	Allopolyploid derivative
Fallopia x bohemica	F. japonica*	F. sachalinensis*	Low	Yes	
(2n = 66)	(2n = 88)	(2n = 44)			
Senecio x baxteri	S. vulgaris	S. squalidus*	Very low	No	<i>S. cambrensis</i> (2n = 60)
(2n = 30)	(2n = 40)	(2n = 20)			
Spartina x townsendii	S. maritima	S. alterniflora*	Very low	Yes	<i>S. anglica</i> (2n = 124)
(2n = 62)	(2n = 60)	(2n = 62)			
Mimulus x robertsii	M. guttatus*	M. luteus*	Very low	Yes	<i>M. peregrinus</i> (2n = 92)
(2n = 44-46)	(2n = 28)	(2n = 60-62)			
Calystegia x lucana	C. sepium	C. sylvatica*	High	Yes	
(2n = 22)	(2n = 22)	(2n = 22)			
Hyacinthoides x massartiana	H. non-scripta	H. hispanica*	High	Yes	
(2n = 16, 24)	(2n = 16, 24)	(2n = 16, 24)			
Rhododendron x superponticum	R. ponticum*	R. catawbiense*, R. maximum*,	High	Yes	
(2n = 26)	(2n = 26)	R. macrophyllum*			
		(2n = 26)			

Centaurea x gerstlaueri	C. nigra	C. jaceae*	High?	Yes (but limited	
(2n = 44)	(2n = 44)	(2n = 44)		lateral spread)	

Table 2. Reproductive mode and occurrence of 274 alien angiosperm hybrids and 1,590 non-hybrid aliens in the British and Irish flora. Data from Stace and Crawley (2015). Hectad = 10km x 10km square.

Dropagation Mode	# Hybrid alien taxa	Average # hectads per	# Non-hybrid alien	Average # hectads per non-
Propagation Mode	(proportion)	hybrid taxon (maximum)	taxa (proportion)	hybrid taxon (maximum)
Seeds only	48 (0.175)	180 (2,242)	559 (0.351)	224 (3,530)
Clonal only	55 (0.200)	157 (1,147)	122 (0.077)	212 (2,723)
Seeds and clonal	71 (0.259)	199 (2,419)	330 (0.207)	218 (2,535)
No reproduction in the British isles	100 (0.365)	32 (444)	579 (0.364)	37 (1,738)
Total	274	126 (2,419)	1,590	153 (3,530)



The genus *Sorbus* (Rosaceae) includes sexual diploid species and apomictic species/taxa with varying ploidy (usually 3x and 4x) that have arisen through hybridisation between sexual species and rare backcrosses with their apomictic allopolyploid derivatives (Rich & Robertson 2015). There are 31 Sorbus agamospecies native to the British Isles, 10 of which are reported in Stace et al., (2015). The parentage of these hybrids is often complex but molecular

data is helping to resolve their origins (Robertson *et al.*, 2010). 'Hotspots' for *Sorbus* diversity in the British Isles include: the Isle of Arran, Cheddar Gorge, the Wye Valley and the Avon Gorge (Rich & Robertson 2015) – the latter being by far the richest, possibly the richest in the world.

The Avon Gorge contains at least 21 Sorbus taxa, six of which are endemic and shown to have evolved within the Gorge, probably within the last 10,000 years since the last ice-age (Robertson et al., 2010; Ludwig et al., 2013). Studies using molecular markers revealed that this Sorbus diversification has been driven primarily by a series of interspecific hybridisations and backcrosses among closely related taxa, with each new genotype being fixed and perpetuated via apomixis (Robertson et al., 2010; Ludwig et al., 2013). Once established these new microspecies then occasionally participate in further rare hybridisation events leading to a complex pattern of ongoing reticulate evolution (Robertson et al. 2010; Ludwig et al., 2013). This occurs because of subtle variation ('leakiness') in the apomictic mating system that permits occasional sexual unions, usually involving pollen from the most common species, S. aria, which is diploid and sexual (outcrossing) and its close tetraploid relative S. porrigentiformis, apomictic. Pollen from these taxa is essential for apomictic seed production by the triploid apomictic hybrid taxa (e.g., S. bristoliensis and S. wilmottiana) because apomixis is pseudogamous (endosperm formation requires fertilization of the central cell by sperm) and triploids (unlike tetraploids) are selfincompatible (SI) (Ludwig et al., 2013; Ludwig & Hiscock in preparation). This unusual mating system therefore fuels the possibility of rare hybridisations being successful because interspecific pollination is required for apomictic seed production in the triploids, which in all but one microspecies tested, are genetically identical (indicating a single hybrid origin event) and therefore incompatible because they all share the same self-incompatibility (S) genotype (Ludwig et al., 2013). Populations of at least two undescribed microspecies have been identified together with numerous puzzling individuals, which do not fit known taxa (Tim Rich pers. comm.) indicating that these evolutionary processes are ongoing. The 2km long Avon Gorge which cuts through Carboniferous Limestone to a depth of 80m, displays a great diversity of habitats (including woodland, scrub, open rocks and quarries) which has probably facilitated and continues to facilitate the survival of newly divergent (perhaps pre-adapted) Sorbus taxa (Rich et al., 2010). Conservation strategies for the rare Sorbus taxa endemic to the Avon Gorge have taken these findings on board in revising management regimes such that all Sorbus taxa are considered so as to conserve the evolutionary process rather than just the rare ones.

Box 2. Monkeyflower hybrids: The birth of a new species

Mimulus has a long tradition in studies of hybridisation, reproductive isolation and speciation (Vickery 1959, Bradshaw *et al.*, 1995, Fishman & Willis 2001, Brandvain *et al.*, 2014). The spread of some *Mimulus* species beyond their native range thanks to human-assisted dispersal, has created new opportunities for hybridisation between previously isolated *Mimulus* taxa (Vallejo-Marín & Lye 2013).

The best studied case is the invasion of the British Isles by two related monkeyflower taxa: The North American diploid *M. guttatus*, and the South American tetraploid *M. luteus*. Both species were introduced into the British Isles in the early 19th century as botanical curiosities, and quickly became naturalised (Stace *et al.*, 2015). Hybridisation between these two taxa produce a highly sterile triploid, which is nevertheless capable of vegetative growth. The hybrid has been established in the wild since at least the 1870's, and it is currently the second most abundant *Mimulus* in the UK after *M. guttatus* being present in approximately 40% of extant populations (Vallejo-Marín & Lye 2013). The triploid hybrid *M. x robertsii*, has produced the new allopolyploid species *M*.



peregrinus, which has originated at least twice, independently, in Scotland (Vallejo-Marín 2012, Vallejo-Marín *et al.*, 2015). *M. peregrinus* produces both viable pollen and seeds, as well as retaining its capacity for vegetative reproduction. Multiple origins seem to be the rule rather than the exception for the formation of allopolyploids (Soltis *et al.*, 2014b), and this provides a natural system to investigate hybrid speciation in a replicated fashion.

Although the parentage of Mimulus hybrids and the origin of M. peregrinus has been recently documented using genome-wide analyses (Vallejo-Marín et al., 2015), a missing piece of the puzzle is to determine which taxon has served as the maternal parent and which as the paternal parent in hybrid formation. Inter-ploidy hybridisation often results in asymmetric reproductive barriers, and it is likely that hybridisation between *M. guttatus* and *M. luteus* does not work equally well in both directions (Roberts 1964). Determining the ancestry of inter-specific hybrids can be done by analysing uniparentally inherited genomes (e.g., chloroplast and mitochondria) (Rieseberg & Brunsfeld 1992, Twyford & Ennos 2012). However, genetic analysis of hybrid ancestry can be complicated by limited availability of species-diagnostic polymorphism in uniparentally-inherited genomes. Recent studies have pointed out how whole genomes of cytoplasmic organelles are recovered in both targeted and whole-genome sequence projects, as by-products of even low-depth sequencing efforts (genome skimming, Dodsworth 2015). This trove of genetic information can be used to identify the taxon that acted as the maternal parent of hybrids, even in the absence of closely related reference genomes (Bakker et al., 2016). Vallejo-Marín et al., (in press) applied a genome skimming approach to determine the ancestry of Mimulus hybrids in the British Isles. They used data obtained from both whole-genome and targeted sequencing projects, to rescue chloroplast and mitochondrial genomes of M. x robertsii and M. peregrinus and their parental taxa. They showed that hybrids have been produced unidirectionally, with M. guttatus as the maternal and M. luteus as the paternal parent. Genome skimming (Straub et al., 2012) holds great potential to investigate the ancestry of neo-hybrids, and we expect similar approaches to be increasingly exploited in the near future.