

RESEARCH ARTICLE

Hydrolysis of phytate and formation of inositol phosphate isomers without or with supplemented phytases in different segments of the digestive tract of broilers

Ellen Zeller¹, Margit Schollenberger¹, Imke Kühn² and Markus Rodehutschord^{1*}

¹Institut für Tierernährung, Universität Hohenheim, 70599 Stuttgart, Germany

²AB Vista Feed Ingredients, 64293 Darmstadt, Germany

(Received 27 March 2014 – Final revision received 31 July 2014 – Accepted 3 November 2014)

Journal of Nutritional Science (2015), vol. 4, e1, page 1 of 12

doi:10.1017/jns.2014.62

Abstract

The objective was to characterise degradation of *myo*-inositol 1,2,3,4,5,6-hexakis (dihydrogen phosphate) (InsP₆) and formation of inositol phosphate (InsP) isomers in different segments of the broiler digestive tract. Influence of an *Aspergillus niger* (PhyA) and two *Escherichia coli*-derived (PhyE1 and PhyE2) phytases was also investigated. A total of 600 16-d-old broilers were allocated to forty floor pens (ten pens per treatment). Low-P (5.2 g/kg DM) maize-soyabean meal-based diets were fed without (basal diet; BD) or with a phytase added. On day 25, digesta from different digestive tract segments were pooled per segment on a pen-basis, freeze-dried and analysed for P, InsP isomers and the marker TiO₂. InsP₆ degradation until the lower ileum (74 %) in BD-fed birds showed a high potential of broilers and their gut microbiota to hydrolyse InsP₆ in low-P diets. Different InsP patterns in different gut segments suggested the involvement of phosphatases of different origin. Supplemented phytases increased InsP₆ hydrolysis in the crop ($P < 0.01$) but not in the lower ileum. Measurements in the crop and proventriculus/gizzard confirmed published *in vitro* degradation pathways of 3- and 6-phytases for the first time. In the intestinal segments, specifically formed InsP₄₋₅ isomers of supplemented phytases were still present, indicating further activity of these enzymes. *Myo*-inositol tetrakisphosphate (InsP₄) accumulation differed between PhyE1 and PhyE2 compared with PhyA in the anterior segments of the gut ($P < 0.01$). Thus, the hydrolytic cleavage of the first phosphate group is not the only limiting step in phytate degradation in broilers.

Key words: Inositol phosphate isomers: Phytate hydrolysis: Phytases: Broilers

Phytate represents the primary storage form of P in plant seeds. It is defined as any salt of phytic acid (*myo*-inositol 1,2,3,4,5,6-hexakis (dihydrogen phosphate) or InsP₆). The utilisation of InsP₆-P depends on InsP₆ hydrolysis because P absorption occurs mainly as orthophosphate⁽¹⁾. InsP₆-hydrolysing enzymes such as phytases (*myo*-inositol hexakisphosphate phosphohydrolases) catalyse the hydrolytic cleavage of InsP₆ and its salts via several phosphorylated intermediary products (*myo*-inositol pentakis-, tetrakis-, tris-, bis- and monophosphate) down to *myo*-inositol. The International Union of Pure and Applied Chemistry/

International Union of Biochemistry differentiates among three types of phytases: 3-phytases (EC 3.1.3.8), 4-/6-phytases (EC 3.1.3.26) and 5-phytases (EC 3.1.3.72), a classification that refers to the initiating position on the inositol ring during *in vitro* InsP₆ dephosphorylation. The 6-phytases usually originate from plants and initiate hydrolysis at the D-4 (L-6) position of InsP₆⁽²⁾; 3-phytases are usually of microbial origin (starting hydrolysis at the D-3 (L-1) position), such as the fungal *Aspergillus niger* phytase⁽³⁻⁵⁾ or the bacterial *Pseudomonas* phytase⁽²⁾. However, *Escherichia coli* phytase as an exception was characterised as a 6-phytase (starting hydrolysis at the D-6

Abbreviations: BD, basal diet; InsP, inositol phosphate; InsP₃, *myo*-inositol trisphosphate; InsP₄, *myo*-inositol tetrakisphosphate; InsP₅, *myo*-inositol pentakisphosphate; InsP₆, *myo*-inositol 1,2,3,4,5,6-hexakis (dihydrogen phosphate); PhyA, *Aspergillus*-derived phytase Finase[®] P; PhyE1, *Escherichia coli*-derived phytase Quantum[®]; PhyE2, *E. coli*-derived phytase Quantum[®] Blue; tP, total P.

* Corresponding author: M. Rodehutschord, email inst450@uni-hohenheim.de



(1-4) position)^(6,7). We are not aware of any study that has investigated whether this *in vitro*-based classification is reflected also in the pathways of InsP₆ degradation in the more complex and variable environment of the digestive tract of broilers.

InsP₆-P has long been assumed to be poorly used by avian species because of the lack of sufficient endogenous InsP₆-hydrolysing enzymes and the denaturation of intrinsic plant phytases in the stomach and during feed manufacture. In broilers, although the activity of endogenous mucosal phytase in the small intestine has been described⁽⁸⁻¹⁰⁾, its contribution to InsP₆ hydrolysis has been considered almost negligible. Although some more recent studies have indicated that InsP₆-P is highly available for broilers⁽¹¹⁾, the origin of phytase activity in the digestive tract is controversial. Some authors have suggested that endogenous mucosal phytase of the small intestine is very capable of high InsP₆ hydrolysis^(12,13). Others hypothesise that InsP₆ is hydrolysed by phytases produced by micro-organisms present in the small intestine and particularly in the caeca^(14,15).

It is not known which positional inositol phosphate (InsP) isomers are formed by different phytases in the digestive tract of avian species. Some studies have described different positional isomers of InsP₆ degradation products in several segments of the digestive tract in pigs⁽¹⁶⁻¹⁸⁾, but the variations in digestive tract physiology and anatomy between pigs and birds (for example, passage rate, digesta viscosity, presence of a crop, variable pH values) caution against assuming that findings in pigs will be similar to that in broilers. One consequence of the uncertainties regarding the availability of InsP₆-P is that commercial poultry diets are supplemented with mineral P sources. This costly supplementation increases the P concentration in excreta, which may contribute to environmental problems such as eutrophication of surface waters and exhaustion of global raw phosphate resources⁽¹⁹⁾. An understanding of the rate of degradation of InsP₆ to the different positional InsP isomers along the digestive tract would enable a more precise alignment of the feed composition to the birds' P requirements and thus increase the likelihood of averting a 'potential planet phosphate crisis'⁽²⁰⁾. Phytases of different origin, varying in their properties, such as pH optimum, proteolytic stability and kinetic efficiency, may differ in effectiveness with transit as the conditions along the digestive tract change. The *in vitro*-determined pH optimum of most phytase supplements is particularly aligned to the conditions of the anterior segments of the digestive tract⁽²¹⁾.

The first objective of the present study therefore was to characterise InsP₆ hydrolysis and formation of InsP isomers in different segments of the digestive tract of broilers. The second objective was to investigate the InsP₆ degradation pattern of different phytase additives and their effectiveness in releasing phosphate in broilers and to compare the findings with known *in vitro* properties. In the absence of supplementary phytase, InsP₆ was hypothesised to be mainly hydrolysed in the posterior intestinal segments of the digestive tract. In contrast, the different phytase supplements were expected to result in greater overall rates of InsP₆ hydrolysis and to elicit different InsP patterns in the anterior segments.

Materials and methods

Experimental diets

The basal diet (BD) was calculated to contain adequate levels of all nutrients according to the recommendations of the Gesellschaft für Ernährungsphysiologie (Society for Nutrition Physiology)⁽²²⁾ with the exception of Ca and P. It was mainly based on maize and solvent-extracted soyabean meal (Table 1). Ingredients were chosen to obtain low concentrations of total P (tP), high proportions of InsP₆-P in tP and low intrinsic phytase activity. Concentrations of Ca and tP were calculated to be 7.9 and 5.0 g/kg of DM, respectively, and these levels were confirmed by analyses (Table 1). Titanium dioxide (TiO₂) was included at a rate of 5 g/kg as the indigestible marker, and the intended Ti concentration was confirmed by analysis.

Diets were prepared in the certified feed mill facilities of Hohenheim University's Agricultural Experiment Station. The BD was mixed in one lot and divided into four equal parts. One part remained without phytase supplementation (BD). The other parts were supplemented with three different phytase-containing products at an intended activity of 500 U/kg of diet. The supplemented phytase products were a commercial *A. niger*-derived 3-phytase (PhyA; Finase[®] P, EC 3.1.3.8; AB Vista) and two *E. coli*-derived thermotolerant 6-phytases (PhyE1 (Quantum[®]) and PhyE2 (Quantum[®] Blue); EC 3.1.3.26; AB Vista). To ensure adequate mixing of each phytase, premixes of each product were prepared by mixing with a small amount of the BD before addition to the treatment diet. Diets were pelleted through a 3-mm die without using steam. The temperature of pellets measured immediately after release from the press ranged between 57°C and 69°C. Representative samples of the diets were taken for analyses of phytase activity, proximate nutrients, DM, Ca, tP, Ti and InsP isomers. The samples were pulverised using a vibrating cup mill (type 6-TOPF; Siebtechnik GmbH) and stored at 4°C until further handling. The experimental diets contained similar concentrations of *myo*-inositol pentakisphosphate (InsP₅) and InsP₆ (Table 1). The InsP₆-P was 57 % of tP in the diets on average and that of InsP₅-P was 3 % of tP. Lower InsP isomers were not detected in the diets. The phytase activity of the BD was below the limit of detection, but phytase activities of the supplemented diets ranged between 399 and 467 U/kg of diet.

Animals and management

The study was conducted in the Agricultural Experiment Station of Hohenheim University, location Lindenhöfe in Eningen (Germany). It was approved by the Animal Welfare Commissioner of the University in accordance with the German Welfare Legislation. Birds underwent routine vaccination against coccidiosis, Newcastle disease and infectious bursal disease on 3, 10 and 14 d of age, respectively.

A total of 600 unsexed Ross 308 broilers aged 1 d were obtained from a commercial hatchery (Brüterei Süd GmbH & Co.) and randomly allocated to forty floor pens (approximately 1.5 m × 1.5 m) bedded with wood shavings. Each pen had fifteen birds. The room temperature was 34 and 32°C on days 1 and 2, respectively. Thereafter, the temperature was reduced in steps of 0.5°C per d, reaching 20°C on day 25. Artificial lighting

**Table 1.** Ingredient composition and analysed characteristics of the experimental diets

	BD	PhyA	PhyE1	PhyE2
Ingredients (g/kg as fed)				
Maize	553			
Solvent-extracted soyabean meal (48 % crude protein)	400			
Soyabean oil	20			
Limestone	13			
Sodium chloride	1			
Choline chloride	2			
Sodium bicarbonate	3			
Mineral mix*	1			
Vitamin mix†	2			
Titanium dioxide	5			
Analysed characteristics of the diets				
DM (g/kg)	895	896	896	895
Crude ash (g/kg)	53	52	53	51
Crude protein (g/kg)	230	231	232	226
Diethyl ether extract (g/kg)	54	53	52	53
Crude fibre (g/kg)	23	21	24	22
Metabolisable energy (calculated) (MJ/kg)	12.5	12.5	12.5	12.5
Phytase activity (U/kg)‡	<50	399§	467	442
Ca (g/kg DM)	7.5	7.6	7.4	7.3
Total P (g/kg DM)	5.2	5.3	5.2	5.2
InsP ₆ -P (g/kg DM)	3.0	3.0	3.0	3.0
Ins(1,2,3,4,6)P ₅ (nmol/g DM)	ND	<LOQ	<LOQ	<LOQ
Ins(1,2,3,4,5)P ₅ (nmol/g DM)	200	200	300	300
Ins(1,2,4,5,6)P ₅ (nmol/g DM)	600	600	600	600
Ins(1,3,4,5,6)P ₅ (nmol/g DM)	ND	ND	ND	ND
InsP ₆ (nmol/g DM)	15 900	16 000	16 200	16 100

BD, basal diet; PhyA, BD supplemented with *Aspergillus niger* 3-phytase, Finase® P; PhyE1, BD supplemented with *Escherichia coli* 6-phytase, Quantum®; PhyE2, BD supplemented with *E. coli* 6-phytase, Quantum® Blue; InsP₆, myo-inositol 1,2,3,4,5,6-hexakis (dihydrogen phosphate); InsP₅, myo-inositol pentakisphosphate; ND, not detected; LOQ, limit of quantification.

* P-free: Mineral mix (Gelamin SG 1 Geflügel, GFT MBH), provided per kg of complete diet: 15 mg Cu; 1.6 mg I; 90 mg Fe; 120 mg Mn; 80 mg Zn; 0.5 mg Se; 0.6 mg Co.

† Vitamin mix (Raiffeisen Kraftfutterwerke Süd GmbH), provided per kg of complete diet: 3.6 mg retinol; 75 µg cholecalciferol; 30 mg α-tocopherol; 2.4 mg menadione; 3 mg thiamin; 6 mg riboflavin; 6 mg pyridoxine; 0.03 mg cyanocobalamin; 50 mg nicotinic acid; 14 mg pantothenic acid; 0.1 mg biotin; 1 mg pteroyl(mono)glutamic acid.

‡ Determined at pH 4.5 and 60°C.

§ Determined at pH 5.0 and 37°C.

was provided with an intensity of 10 lux. During the first 2 d, the animal house was illuminated continuously. A lighting regimen of 18 h light and 6 h dark was applied from day 3 onwards. Feed and tap water were available for *ad libitum* consumption. Until day 15, the animals were fed a commercial starter diet containing 1.10 % Ca, 0.55 % tP, 22.0 % crude protein, 6.6 % diethyl ether extract and 12.5 MJ metabolisable energy/kg. On day 16, the birds were weighed, and ten pens of fifteen birds were assigned to each of the four dietary treatments and distributed in a completely randomised block design.

Sampling and analytical methods

At 25 d of age, the animals were asphyxiated by CO₂ exposure and weighed. To standardise feed intake before sampling and thus retention time of feed in the crop, birds were deprived of

feed for 1 h. The feed troughs were then moved back into the pens 1 h before the birds were killed, on an individual-pen basis to ensure the same time schedule for all replicates. The first samples were taken 1 h after the beginning of the light period. Samples from five parts of the digestive tract (crop, proventriculus and gizzard (pooled), duodenum and jejunum (pooled), the terminal part of the ileum (defined as the posterior two-thirds of the section between Meckel's diverticulum and 2 cm anterior to the ileo-caeco-colonic junction^(2,3)) and the caeca) were taken. After opening of the abdominal cavity, the total digestive tract was removed except the crop. Digesta of the intestinal segments were gently flushed out with double-distilled water whereas the segments of the anterior digestive tract (crop, proventriculus and gizzard) were cut open and purged. The samples were pooled for all birds from one pen separately for each segment, immediately frozen at -18°C, freeze-dried (type Delta 1-24; Martin Christ Gefriertrocknungsanlagen GmbH) and pulverised as explained for the diets. The ground samples were stored at 4°C until analysis.

Concentrations of proximate nutrients were determined according to the official methods in Germany (Verband Deutscher Landwirtschaftlicher Untersuchungs- und Forschungsanstalten; VDLUFA)⁽²⁴⁾. Feed samples were analysed for DM and crude ash (method 3.1), crude protein (method 4.1.1), diethyl ether extract (method 5.1.1) and crude fibre (method 6.1.1). The concentrations of Ca, tP and Ti in diet and digesta samples were determined by a modification of the method of Boguhn *et al.*⁽²⁵⁾. In brief, 20 ml of sulfuric acid (18 mol/l) and 2.5 ml of nitric acid (14 mol/l) were added to 0.4 g of sample. Solutions were heated from 100 to 200°C for 30 min in a block digestion system equipped with a system to trap nitrous gases (Behr K 20 L; Behr Labor-Technik GmbH). After cooling to 100°C, 2.5 ml of the nitric acid were added. Following heating from 225 up to 300°C for 75 min and subsequent cooling to room temperature, the solutions were filled with double-distilled water to a volume of 500 ml and filtered. The Ca, P and Ti concentrations of the solutions were measured using an inductively coupled plasma optical emission spectrometer (VISTA PRO; Varian Inc.) at specific wavelengths for each element according to Shastak *et al.*⁽²⁶⁾.

For the analysis of InsP isomers in diet and digesta samples, 1.0 g of the sample was extracted for 30 min with 10 ml of a solution containing 0.2 M-EDTA and 0.1 M-sodium fluoride (pH = 10) as phytase inhibitor using a rotary shaker. The samples were centrifuged at 12 000 g for 15 min and the supernatant fraction was removed and preserved on ice. The residue was re-suspended in 5 ml of the EDTA-sodium fluoride solution and extracted again for 30 min. The supernatant fractions of the two extraction steps were then combined. A quantity of 1 ml of the pooled supernatant fraction was centrifuged at 14 000 g for 15 min and 0.5 ml of the resulting supernatant fraction were filtered through a 0.2 µm cellulose acetate filter (VWR) into a Microcon filter (cut-off 30 kDa) device (Millipore) and centrifuged again at 14 000 g for 30 min. Throughout the whole extraction procedure, the samples were kept below 6°C. The procedure for caecal samples was slightly different: glass beads (diameter 0.6 mm) were added before extraction. To obtain a clear supernatant fraction for the caecal



matrix, the extracts were centrifuged for 30 min at 12 000 g and 6°C. Filtrates were analysed by high-performance ion chromatography and UV detection at 290 nm after post-column derivatisation using an ICS-3000 system (Dionex). InsP with different degrees of phosphorylation (InsP₃₋₆) and their positional isomers were separated, without enantiomer differentiation, on a Carbo Pac PA 200 column and corresponding guard column. Fe(NO₃)₃ solution (1 g/l, Fe(NO₃)₃·9H₂O, product no. 103883; Merck KGaA) in HClO₄ (20 g/l, product no. 100518; Merck KGaA) was used as reagent for derivatisation according to Philippy & Bland⁽²⁷⁾. The elution order of InsP isomers was established using commercial standards if available. InsP₅ isomer standards were purchased from Sirius Fine Chemicals. Seven out of nine *myo*-inositol tetrakisphosphate (InsP₄) and nine out of twelve *myo*-inositol trisphosphate (InsP₃) isomer standards were available from Santa Cruz Biotechnology. One detected peak out of the group of InsP₄ isomers could not be attributed but was presumed to be Ins(1,2,4,6)P₄ by comparison with the elution order of Chen & Li⁽²⁸⁾, who used similar chromatographical conditions. A clear identification of the InsP₃ isomers present was not possible. However, a peak was detected, corresponding in its retention time to the retention time of Ins(1,3,4)P₃, Ins(1,4,6)P₃, Ins(1,2,6)P₃, Ins(1,4,5)P₃ and Ins(2,4,5)P₃ (out of the available standards), which all coeluted under the conditions used. InsP₂ and InsP₁ could not be analysed with this method. InsP₆ was used for quantification, and correction factors for differences in detector response for InsP₃₋₅ were used according to Skoglund *et al.*⁽²⁹⁾. The limit of detection was defined for a signal:noise ratio of 3:1 and was 0.1 µmol/g of DM for InsP₃₋₄ isomers and 0.05 µmol/g of DM for InsP₅ isomers and InsP₆. The limit of quantification was defined for a signal:noise ratio of 6:1. A mean for an InsP isomer was calculated only if the isomer was detected in at least five out of the ten samples of one treatment. If the detected value was below the limit of quantification in five or more samples, this was noted as less than the limit of quantification in the tables, and means were not calculated. All samples were analysed in duplicate. The InsP concentration is reported on a DM basis.

Because of differences in the extractability of the phytases used, the phytase activity in the diets was determined under product-specific conditions and expressed as U/kg for all diets. Determination of phytase activity was assayed according to the internal, validated methods of the supplier (BD + PhyA: assay at pH 5.5 and 37°C; BD, BD + PhyE1 and BD + PhyE2: assay at pH 4.5 and 60°C). Both assays were run by Enzyme Services & Consultancy.

Calculations and statistical analysis

Body-weight gain, feed consumption and feed:gain ratio were determined on a pen basis for the period between days 16 and 25. InsP₆ hydrolysis and P net absorption in the digestive tract (*y*) were calculated for each pen based on the ratio of InsP₆ or P and Ti according to the generally accepted equation:

$$y(\%) = 100 - 100 \times \left(\frac{\text{Ti in the diet (g/kg DM)}}{\text{Ti in the digesta (g/kg DM)}} \right) \times \left(\frac{\text{InsP}_6 \text{ or P in the digesta (g/kg DM)}}{\text{InsP}_6 \text{ or P in the diet (g/kg DM)}} \right)$$

InsP₆ hydrolysis was calculated for the crop, duodenum/jejunum, lower ileum and caeca. It was not calculated for the proventriculus/gizzard since this segment clearly contained particles of different sizes, which presumably were of variable retention times and thus were not accurately represented by the marker⁽³⁰⁾. P net absorption was calculated in the duodenum/jejunum and lower ileum.

The percentage of InsP₃, \sum InsP₄ or \sum InsP₅ in \sum InsP₃₋₅ was calculated for each treatment and segment to investigate the rapidity of InsP₆ hydrolysis and the extent to which intermediary products with different degrees of phosphorylation were formed. Because InsP₁₋₂ isomers were not determined, \sum InsP₃₋₅ was calculated, representing the sum of identified InsP₆ hydrolysis products.

Untransformed data are expressed as means with their standard error or the pooled standard error of the mean. Statistical analysis was performed using the MIXED procedure of the software package SAS for Windows (version 9.1.3; SAS Institute Inc.). Before statistical analysis, data that showed non-normal residuals or heterogeneity of variance were log- or square root-transformed. For data expressed as percentages the arc-sine transformation was used. The following statistical model was chosen: $y_{ij} = \mu + \tau_i + \tau_j + e_{ij}$, where y_{ij} is the *i*th measurement in the *j*th treatment, μ is the overall mean, τ_i is the *i*th block (random), τ_j is the effect of the *j*th treatment (fixed) and e_{ij} is the error term. Statistical significance was evaluated by a one-way ANOVA. Mean separation was computed using Fisher's protected least significant difference test ($P \leq 0.05$) only if the overall *F* test was significant ($P \leq 0.05$).

Results

The initial body weight was on average 531 g and was similar between treatments ($P > 0.05$). During the 9-d assay period, average body-weight gain, feed consumption and feed:gain ratio were not significantly different between treatments (Table 2).

Table 2. Body-weight (BW) gain, feed consumption (FC) and feed:gain (F:G) ratio of broiler chickens between the ages of 16 and 25 d (Mean values and pooled standard errors; ten pens per treatment with fifteen birds per pen)

	BD	PhyA	PhyE1	PhyE2	Pooled SEM
BW gain (g/d)*	59	58	60	56	1.4
FC (g/d)*	95	94	97	92	1.9
F:G ratio (g/g)*	1.61	1.61	1.62	1.63	0.023

BD, basal diet; PhyA, BD supplemented with *Aspergillus niger* 3-phytase, Finase[®] P; PhyE1, BD supplemented with *Escherichia coli* 6-phytase, Quantum[®]; PhyE2, BD supplemented with *E. coli* 6-phytase, Quantum[®] Blue.

* The overall *F* test was not significant.



Table 3. Myo-inositol 1,2,3,4,5,6-hexakis (dihydrogen phosphate) hydrolysis (%) in different segments of the digestive tract of broiler chickens (Mean values and pooled standard errors; ten pens per treatment with fifteen birds per pen)

	BD	PhyA	PhyE1	PhyE2	Pooled SEM
Crop	9 ^d	64 ^a	31 ^c	44 ^b	4.5
Duodenum/jejunum	59	63	65	68	2.8
Lower ileum	74	74	79	82	3.8
Caeca	91 ^b	93 ^b	95 ^a	96 ^a	0.7

BD, basal diet; PhyA, BD supplemented with *Aspergillus niger* 3-phytase, Finase[®] P; PhyE1, BD supplemented with *Escherichia coli* 6-phytase, Quantum[®]; PhyE2, BD supplemented with *E. coli* 6-phytase, Quantum[®] Blue.

^{a-d} Mean values in a row with unlike superscript letters were significantly different ($P \leq 0.05$; Fisher's protected least significant difference test). Mean separation was only computed if the overall *F* test was significant.

Myo-inositol 1,2,3,4,5,6-hexakis (dihydrogen phosphate) hydrolysis

In birds fed the BD, low InsP₆ hydrolysis was measured in the crop (9 %; Table 3). Average InsP₆ hydrolysis in this treatment was 59, 74 and 91 % until the duodenum/jejunum, the lower ileum and the caeca, respectively.

Supplementation of phytase had a significant effect on InsP₆ hydrolysis in the crop ($P < 0.01$) but not in the duodenum/jejunum and lower ileum. In the crop, supplemented phytases significantly increased InsP₆ hydrolysis, and the effect by PhyA (64 %) was significantly higher than by the other phytase treatments (PhyE1: 31 %; PhyE2: 44 %). Until the duodenum/jejunum and the lower ileum, the difference in InsP₆ hydrolysis between treatments became lower and statistical difference between treatments disappeared. The average InsP₆ hydrolysis was 64 % (duodenum/jejunum) and 77 % (lower ileum). InsP₆ hydrolysis up to the caeca was significantly higher with PhyE1 (95 %) and PhyE2 (96 %) compared with the BD (91 %) and PhyA (93 %) ($P < 0.01$).

Net absorption of phosphorus

In birds fed the BD, a P net absorption of 34 and 57 % was measured until the duodenum/jejunum and lower ileum (Table 4). Supplementation of phytases caused a significant increase in P net absorption until the duodenum/jejunum (PhyA: 38 %; PhyE1: 38 %; PhyE2: 39 %) ($P = 0.04$). P net absorption until the lower ileum tended to be higher with PhyE1 (60 %) and was significantly higher with PhyE2 (64 %) compared with PhyA (56 %) and BD (57 %) ($P = 0.03$).

Appearance of inositol phosphate isomers

In the crop digesta of birds fed the BD, an average concentration of 638 and 388 nmol/g DM was detected for Ins

(1,2,4,5,6)P₅ and Ins(1,2,3,4,5)P₅ and lower concentrations were measured for Ins(1,2,3,4,6)P₅ and Ins(1,3,4,5,6)P₅ (Table 5 and Fig. 1(A)). The only detectable inositol tetrakisphosphate Ins(1,2,5,6)P₄ was found in low concentration (141 nmol/g DM), and inositol trisphosphates were not found in the crop in this treatment. About 90 % of Σ InsP₃₋₅ in the crop was present as InsP₅ when the BD was fed. In the proventriculus/gizzard, Ins(1,2,4,5,6)P₅ and Ins(1,2,3,4,5)P₅ were the only detectable lower InsP (Table 6), and their concentrations were lower than in the crop. In the duodenum/jejunum, the InsP pattern again was more diverse than in the proventriculus/gizzard and different from the crop (Table 7).

The predominant InsP₅ isomer changed in the intestinal segments (Fig. 1 (B) and (C)). Ins(1,2,3,4,5)P₅ was the predominant InsP₅ isomer in the duodenum/jejunum and the subsequent intestinal segments, accompanied by Ins(1,2,3,4,6)P₅ and Ins(1,2,4,5,6)P₅ (Tables 7–9). In the duodenum/jejunum, high concentrations of Ins(1,2,3,4)P₄ (355 nmol/g DM) were noted and it remained the predominant InsP₄ isomer in subsequent segments in the BD treatment. In the lower ileum, the same pattern of InsP₅ isomers as in the duodenum/jejunum was found whereas within the InsP₄ isomers, Ins(1,2,4,6)P₄ also appeared (Table 8). The pattern of the InsP₅ isomers in the caeca of birds fed the BD was similar to that of the lower ileum, except that Ins(1,3,4,5,6)P₅ appeared in relatively low concentrations (75 nmol/g DM) (Table 9). High concentrations of Ins(1,2,3,4)P₄ (750 nmol/g DM) were detected in the caeca whereas Ins(1,2,4,6)P₄ was not found and InsP₃ and traces of Ins(1,2,5,6)P₄ appeared.

In the crop digesta of all phytase-containing treatments, the InsP₅ pattern was less broad compared with the BD. For the PhyA treatment, the percentage of Ins(1,2,4,5,6)P₅ in Σ InsP₅ was significantly higher ($P < 0.01$) whereas the percentage of Ins(1,2,3,4,5)P₅ in Σ InsP₅ was significantly lower ($P < 0.01$) compared with the three other treatments (Fig. 1 (A)). In

Table 4. Net absorption of phosphorus (%) in segments of the small intestine of broiler chickens (Mean values and pooled standard errors; ten pens per treatment with fifteen birds per pen)

	BD	PhyA	PhyE1	PhyE2	Pooled SEM
Duodenum/jejunum	34 ^b	38 ^a	38 ^a	39 ^a	1.2
Lower ileum	57 ^b	56 ^b	60 ^{a,b}	64 ^a	2.1

BD, basal diet; PhyA, BD supplemented with *Aspergillus niger* 3-phytase, Finase[®] P; PhyE1, BD supplemented with *Escherichia coli* 6-phytase, Quantum[®]; PhyE2, BD supplemented with *E. coli* 6-phytase, Quantum[®] Blue.

^{a,b} Mean values in a row with unlike superscript letters were significantly different ($P \leq 0.05$; Fisher's protected least significant difference test). Mean separation was only computed if the overall *F* test was significant.



Table 5. Concentrations of different inositol phosphate (InsP) isomers (nmol/g DM) in the crop digesta (Mean values and pooled standard errors; ten pens per treatment with fifteen birds per pen)

	BD	PhyA	PhyE1	PhyE2	Pooled SEM
InsP ₃ *	ND	4268 ^a	783 ^b	845 ^b	214
Proportion of InsP ₃ in Σ InsP ₃₋₅		0.69 ^a	0.15 ^b	0.13 ^b	0.018
Ins(1,2,4,6)P ₄	ND	ND	ND	ND	
Ins(1,2,3,4)P ₄	ND	ND	170	ND	20.8
Ins(1,2,5,6)P ₄	141 ^c	922 ^c	2708 ^b	4505 ^a	421
Proportion of Σ InsP ₄ in Σ InsP ₃₋₅	0.10 ^c	0.14 ^c	0.57 ^b	0.74 ^a	0.032
Ins(1,2,3,4,6)P ₅	116 ^a	ND	65 ^b	ND	5.1
Ins(1,2,3,4,5)P ₅	388 ^{a,b}	101 ^c	475 ^a	355 ^b	23.0
Ins(1,2,4,5,6)P ₅	638 ^b	886 ^a	371 ^c	241 ^d	46.2
Ins(1,3,4,5,6)P ₅	51	ND	ND	ND	8.6
Proportion of Σ InsP ₅ in Σ InsP ₃₋₅	0.90 ^a	0.16 ^c	0.28 ^b	0.13 ^c	0.037
InsP ₆	14 491 ^a	5790 ^d	10 922 ^b	8670 ^c	720

BD, basal diet; PhyA, BD supplemented with *Aspergillus niger* 3-phytase, Finase[®] P; PhyE1, BD supplemented with *Escherichia coli* 6-phytase, Quantum[®]; PhyE2, BD supplemented with *E. coli* 6-phytase, Quantum[®] Blue; InsP₃, myo-inositol trisphosphate; ND, not detected (the InsP isomer was not detectable in the majority of samples); InsP₄, myo-inositol tetrakisphosphate; InsP₅, myo-inositol pentakisphosphate; InsP₆, myo-inositol 1,2,3,4,5,6-hexakis (dihydrogen phosphate).

^{a-d} Mean values in a row with unlike superscript letters were significantly different ($P \leq 0.05$; Fisher's protected least significant difference test). Mean separation was only computed if the overall *F* test was significant.

* At least one out of the following InsP₃ isomers: Ins(1,4,5)P₃, Ins(1,2,6)P₃, Ins(2,4,5)P₃, Ins(1,3,4)P₃, Ins(1,4,6)P₃.

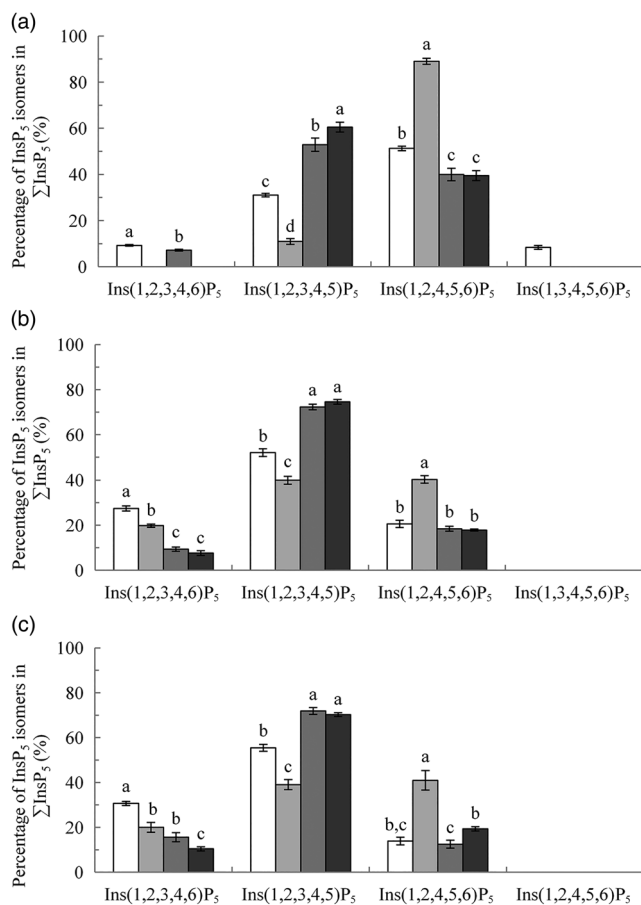


Fig. 1. Myo-inositol pentakisphosphate (InsP₅) isomers in the crop (A), duodenum/jejunum (B) and ileum (C), expressed as a percentage of Σ InsP₅. Values are means, with their standard errors represented by vertical bars. ^{a-d} Values with unlike letters within an InsP₅ isomer were significantly different ($P \leq 0.05$; Fisher's protected least significant difference test). Mean separation was only computed if the overall *F* test was significant. □, Basal diet (BD); ▨, BD supplemented with *Aspergillus niger* 3-phytase, Finase[®] P (PhyA); ▩, BD supplemented with *Escherichia coli* 6-phytase, Quantum[®] (PhyE1); ▪, BD supplemented with *E. coli* 6-phytase, Quantum[®] Blue (PhyE2).

contrast, for PhyE1 and PhyE2, the percentage of Ins(1,2,3,4,5)P₅ in Σ InsP₅ was significantly higher, but the percentage of Ins(1,2,4,5,6)P₅ in Σ InsP₅ was significantly lower compared with the two other treatments. Ins(1,2,5,6)P₄ and InsP₃ also occurred in the crop in high concentrations for the phytase-containing treatments (Table 5). Ins(1,2,5,6)P₄ appeared for the PhyE1 and PhyE2 treatment in significantly higher concentrations compared with the BD and PhyA ($P < 0.01$) and as the only inositol tetrakisphosphate formed by PhyA. InsP₃ appeared in significantly higher concentrations for PhyA than PhyE1 and PhyE2 ($P < 0.01$). The proportion of Σ InsP₄ in Σ InsP₃₋₅ was significantly higher for PhyE2 (74 %) than PhyE1 (57 %) and PhyA (14 %) ($P < 0.01$), and that of InsP₃ in Σ InsP₃₋₅ was significantly higher for PhyA (69 %) compared with PhyE1 (15 %) and PhyE2 (13 %) ($P < 0.01$) in the crop. In the proventriculus/gizzard, the InsP₅ pattern of the phytase-containing treatments was similar to that of the crop except Ins(1,2,3,4,6)P₅ was not seen in the proventriculus/gizzard (Table 6). Ins(1,2,5,6)P₄ and InsP₃ were detected in significantly lower concentrations for PhyA compared with PhyE1 and PhyE2 ($P < 0.01$ for both). A higher proportion of Σ InsP₅ in Σ InsP₃₋₅ and a lower proportion of Σ InsP₄ in Σ InsP₃₋₅ were identified for PhyA compared with PhyE1 and PhyE2 ($P < 0.01$).

In the segments of the small intestine, almost the same InsP isomers were detected in all treatments, except Ins(1,2,5,6)P₄, which occurred only in the phytase treatments (Tables 7 and 8). In the duodenum/jejunum, Ins(1,2,3,4,6)P₅ appeared for the first time for PhyA and PhyE2 and again appeared for PhyE1. Ins(1,2,3,4)P₄ was the predominating InsP₄ isomer for all treatments. The concentrations of Ins(1,2,3,4,5)P₅ in the duodenum/jejunum were similar for the BD and the PhyA, but significantly higher for the PhyE1 and PhyE2 treatments ($P < 0.01$) (Table 7). Ins(1,2,4,5,6)P₅ was detected in significantly higher concentrations for the PhyA compared with the other treatments ($P < 0.01$). For all phytase treatments, a significantly lower proportion of Ins(1,2,3,4,6)P₅ in Σ InsP₅ was determined



Table 6. Concentrations of different inositol phosphate (InsP) isomers (nmol/g DM) in the proventriculus/gizzard digesta (Mean values and pooled standard errors; ten pens per treatment with fifteen birds per pen)

	BD	PhyA	PhyE1	PhyE2	Pooled SEM
InsP ₃ *	ND	105 ^c	348 ^b	517 ^a	54.2
Proportion of InsP ₃ in ΣInsP ₃₋₅		0.10	0.09	0.11	0.012
Ins(1,2,4,6)P ₄	ND	ND	ND	ND	
Ins(1,2,3,4)P ₄	ND	ND	<LOQ	ND	
Ins(1,2,5,6)P ₄	ND	254 ^c	2457 ^b	3537 ^a	211
Proportion of ΣInsP ₄ in ΣInsP ₃₋₅		0.22 ^c	0.67 ^b	0.79 ^a	0.017
Ins(1,2,3,4,6)P ₅	ND	ND	ND	ND	
Ins(1,2,3,4,5)P ₅	92 ^c	107 ^c	659 ^a	295 ^b	30.6
Ins(1,2,4,5,6)P ₅	188 ^b	618 ^a	155 ^b	80 ^c	28.0
Ins(1,3,4,5,6)P ₅	ND	ND	ND	ND	
Proportion of ΣInsP ₅ in ΣInsP ₃₋₅	1.00 ^a	0.68 ^b	0.24 ^c	0.09 ^d	0.020
InsP ₆	6877 ^a	4968 ^b	2219 ^c	739 ^d	196

BD, basal diet; PhyA, BD supplemented with *Aspergillus niger* 3-phytase, Finase[®] P; PhyE1, BD supplemented with *Escherichia coli* 6-phytase, Quantum[®]; PhyE2, BD supplemented with *E. coli* 6-phytase, Quantum[®] Blue; InsP₃, *myo*-inositol trisphosphate; ND, not detected (the InsP isomer was not detectable in the majority of samples); InsP₄, *myo*-inositol tetrakisphosphate; LOQ, limit of quantification (the InsP isomer was not quantifiable in the majority of samples); InsP₅, *myo*-inositol pentakisphosphate; InsP₆, *myo*-inositol 1,2,3,4,5,6-hexakis (dihydrogen phosphate).

^{a-d} Mean values in a row with unlike superscript letters were significantly different ($P \leq 0.05$; Fisher's protected least significant difference test). Mean separation was only computed if the overall F test was significant.

* At least one out of the following InsP₃ isomers: Ins(1,4,5)P₃, Ins(1,2,6)P₃, Ins(2,4,5)P₃, Ins(1,3,4)P₃, Ins(1,4,6)P₃.

compared with the BD in the duodenum/jejunum (Fig. 1 (B)) and lower ileum (Fig. 1 (C)) ($P < 0.01$). In the lower ileum, the significant difference in Ins(1,2,4,5,6)P₅ concentrations persisted ($P < 0.01$) whereas the difference in Ins(1,2,3,4,5)P₅ concentrations lost significance (Table 8). InsP₃ was not detectable in any of the treatments in the small intestine.

In the caeca, the differences in InsP patterns between the phytase-containing diets and the BD were less distinct than in other sections. Concentrations of Ins(1,2,3,4,6)P₅ and Ins(1,2,3,4)P₄ were lower ($P < 0.01$) and concentrations of InsP₃ ($P = 0.03$) and Ins(1,2,5,6)P₄ ($P = 0.04$) partially higher when phytases were supplemented, especially for PhyE2 (Table 9). The specific InsP isomers of the supplemented phytases were still present in the caeca. The predominating InsP₅ isomer was Ins(1,2,3,4,5)P₅ for all phytase treatments. For the PhyA treatment, significantly higher concentrations of Ins

(1,2,4,5,6)P₅ were measured compared with the other treatments ($P < 0.01$).

Discussion

Myo-inositol 1,2,3,4,5,6-hexakis (dihydrogen phosphate) hydrolysis and net absorption of phosphorus

In agreement with previous studies that used low-Ca and low-P diets⁽¹¹⁻¹³⁾ we found a high rate of InsP₆ hydrolysis (76 %) and P net absorption (57 %) in the lower ileum. This raises the question of the origin of phytase responsible for this hydrolysis. We found that without supplemented phytase, the majority of InsP₆ hydrolysis occurred by the end of the duodenum/jejunum, but hydrolysis still continued in the ileum and caeca. In the small intestine of broilers, the greatest endogenous mucosa phytase activity was found in the

Table 7. Concentrations of different inositol phosphate (InsP) isomers (nmol/g DM) in the duodenal/jejunal digesta (Mean values and pooled standard errors; ten pens per treatment with fifteen birds per pen)

	BD	PhyA	PhyE1	PhyE2	Pooled SEM
InsP ₃ *	ND	ND	ND	ND	
Ins(1,2,4,6)P ₄	ND	ND	ND	ND	
Ins(1,2,3,4)P ₄	355	<LOQ	303	265	75.1
Ins(1,2,5,6)P ₄	ND	<LOQ	123 ^b	234 ^a	44.9
Proportion of ΣInsP ₄ in ΣInsP ₃₋₅	0.28		0.26	0.27	0.025
Ins(1,2,3,4,6)P ₅	207 ^a	184 ^a	110 ^b	99 ^b	16.7
Ins(1,2,3,4,5)P ₅	399 ^b	376 ^b	864 ^a	957 ^a	111
Ins(1,2,4,5,6)P ₅	148 ^c	379 ^a	214 ^{b,c}	232 ^b	31.7
Ins(1,3,4,5,6)P ₅	ND	ND	ND	ND	
Proportion of ΣInsP ₅ in ΣInsP ₃₋₅	0.72 ^b	1.00 ^a	0.74 ^b	0.73 ^b	0.022
InsP ₆	13 392	12 664	11 812	10 701	1076

BD, basal diet; PhyA, BD supplemented with *Aspergillus niger* 3-phytase, Finase[®] P; PhyE1, BD supplemented with *Escherichia coli* 6-phytase, Quantum[®]; PhyE2, BD supplemented with *E. coli* 6-phytase, Quantum[®] Blue; InsP₃, *myo*-inositol trisphosphate; ND, not detected (the InsP isomer was not detectable in the majority of samples); InsP₄, *myo*-inositol tetrakisphosphate; LOQ, limit of quantification (the InsP isomer was not quantifiable in the majority of samples); InsP₅, *myo*-inositol pentakisphosphate; InsP₆, *myo*-inositol 1,2,3,4,5,6-hexakis (dihydrogen phosphate).

^{a,b,c} Mean values in a row with unlike superscript letters were significantly different ($P \leq 0.05$; Fisher's protected least significant difference test). Mean separation was only computed if the overall F test was significant.

* At least one out of the following InsP₃ isomers: Ins(1,4,5)P₃, Ins(1,2,6)P₃, Ins(2,4,5)P₃, Ins(1,3,4)P₃, Ins(1,4,6)P₃.



Table 8. Concentrations of different inositol phosphate (InsP) isomers (nmol/g DM) in the digesta of the lower ileum (Mean values and pooled standard errors; ten pens per treatment with fifteen birds per pen)

	BD	PhyA	PhyE1	PhyE2	Pooled SEM
InsP ₃ *	ND	ND	ND	ND	
Ins(1,2,4,6)P ₄	121	ND	<LOQ	<LOQ	30.5
Ins(1,2,3,4)P ₄	309	248	282	216	69.3
Ins(1,2,5,6)P ₄	ND	275	177	192	61.4
Proportion of Σ InsP ₄ in Σ InsP ₃₋₅	0.34	0.31	0.31	0.31	0.036
Ins(1,2,3,4,6)P ₅	231 ^a	209 ^a	149 ^{a,b}	106 ^b	36.0
Ins(1,2,3,4,5)P ₅	422	410	776	735	156
Ins(1,2,4,5,6)P ₅	110 ^b	463 ^a	163 ^b	195 ^b	51.5
Ins(1,3,4,5,6)P ₅	ND	ND	ND	ND	
Proportion of Σ InsP ₅ in Σ InsP ₃₋₅	0.66	0.69	0.69	0.69	0.036
InsP ₆	11 575	12 348	9965	8913	2089

BD, basal diet; PhyA, BD supplemented with *Aspergillus niger* 3-phytase, Finase® P; PhyE1, BD supplemented with *Escherichia coli* 6-phytase, Quantum®; PhyE2, BD supplemented with *E. coli* 6-phytase, Quantum® Blue; InsP₃, *myo*-inositol trisphosphate; ND, not detected (the InsP isomer was not detectable in the majority of samples); InsP₄, *myo*-inositol tetrakisphosphate; LOQ, limit of quantification (the InsP isomer was not quantifiable in the majority of samples); InsP₅, *myo*-inositol pentakisphosphate; InsP₆, *myo*-inositol 1,2,3,4,5,6-hexakis (dihydrogen phosphate).

^{a,b} Mean values in a row with unlike superscript letters were significantly different ($P \leq 0.05$; Fisher's protected least significant difference test). Mean separation was only computed if the overall *F* test was significant.

* At least one out of the following InsP₃ isomers: Ins(1,4,5)P₃, Ins(1,2,6)P₃, Ins(2,4,5)P₃, Ins(1,3,4)P₃, Ins(1,4,6)P₃.

duodenum^(8,10). Phytate-degrading activity has also been reported for different lactic acid bacteria isolated from chicken intestine⁽³¹⁾. Thus, intestinal InsP₆ hydrolysis was the result of a combination of endogenous and microbiota phytase, with as-yet-unknown contributions from each source. When phytase activity in different segments of the digestive tract was compared, the highest activity was found in the caeca⁽³²⁾ and Kerr *et al.*⁽¹⁴⁾ detected higher concentrations of InsP₆ in the caeca of gnotobiotic compared with conventional broilers. In line with this observation, caecal InsP₆ hydrolysis determined in the present study was greater than 90%. It should be noted that retrograde movement of digesta and micro-organisms has been described for all segments of the digestive tract in broilers⁽³³⁾ and it cannot be ruled out that this affected concentrations of InsP isomers, P and Ti anterior to the caeca.

With supplemented phytase, the crop and the proventriculus/gizzard were the main sites of InsP₆ hydrolysis in the present

study. The differences found between the supplemented phytases in these segments might be related to differences in enzyme kinetics, pH or resistance against gastrointestinal proteases. A higher temperature optimum was reported for *E. coli* compared with *Aspergillus* phytases^(34,35). At approximate body temperature (42°C), *Aspergillus* phytases show an activity of 85% of the maximum whereas the activity of some *E. coli* phytases is reduced to 60% of the *in vitro* analysed maximum⁽³⁴⁾. Furthermore, *E. coli* phytases are more resistant than *Aspergillus* phytases against pepsin and pancreatin and show a higher activity at pH 3, which is close to the pH in the proventriculus/gizzard⁽³⁵⁻³⁷⁾. This might explain why the differences between phytases noted in the crop disappeared in the duodenum/jejunum. Moreover, a residual activity of 93 and 60% has been found for an *E. coli* and an *Aspergillus* phytase, respectively, after incubation in digesta of the proventriculus⁽³⁵⁾.

Table 9. Concentrations of different inositol phosphate (InsP) isomers (nmol/g DM) in the caecal digesta (Mean values and pooled standard errors; ten pens per treatment with fifteen birds per pen)

	BD	PhyA	PhyE1	PhyE2	Pooled SEM
InsP ₃ *	217 ^b	275 ^b	355 ^{a,b}	470 ^a	59.4
Proportion of InsP ₃ in Σ InsP ₃₋₅	0.09 ^b	0.13 ^b	0.19 ^b	0.30 ^a	0.020
Ins(1,2,4,6)P ₄	ND	ND	ND	ND	
Ins(1,2,3,4)P ₄	750 ^a	484 ^b	422 ^b	284 ^b	86.1
Ins(1,2,5,6)P ₄	<LOQ	309 ^a	190 ^b	318 ^a	38.9
Proportion of Σ InsP ₄ in Σ InsP ₃₋₅	0.34 ^b	0.41 ^a	0.36 ^b	0.39 ^{a,b}	0.015
Ins(1,2,3,4,6)P ₅	365 ^a	186 ^b	88 ^b	ND	43.8
Ins(1,2,3,4,5)P ₅	578 ^a	324 ^b	436 ^{a,b}	308 ^b	63.7
Ins(1,2,4,5,6)P ₅	119 ^b	302 ^a	96 ^b	82 ^b	18.8
Ins(1,3,4,5,6)P ₅	75	85	86	85	7.5
Proportion of Σ InsP ₅ in Σ InsP ₃₋₅	0.57 ^a	0.47 ^b	0.45 ^b	0.31 ^c	0.023
InsP ₆	6751 ^a	4494 ^b	3648 ^{b,c}	2255 ^c	544

BD, basal diet; PhyA, BD supplemented with *Aspergillus niger* 3-phytase, Finase® P; PhyE1, BD supplemented with *Escherichia coli* 6-phytase, Quantum®; PhyE2, BD supplemented with *E. coli* 6-phytase, Quantum® Blue; InsP₃, *myo*-inositol trisphosphate; InsP₄, *myo*-inositol tetrakisphosphate; ND, not detected (the InsP isomer was not detectable in the majority of samples); LOQ, limit of quantification (the InsP isomer was not quantifiable in the majority of samples); InsP₅, *myo*-inositol pentakisphosphate; InsP₆, *myo*-inositol 1,2,3,4,5,6-hexakis (dihydrogen phosphate).

^{a,b,c} Mean values in a row with unlike superscript letters were significantly different ($P \leq 0.05$; Fisher's protected least significant difference test). Mean separation was only computed if the overall *F* test was significant.

* At least one out of the following InsP₃ isomers: Ins(1,4,5)P₃, Ins(1,2,6)P₃, Ins(2,4,5)P₃, Ins(1,3,4)P₃, Ins(1,4,6)P₃.



Appearance of inositol phosphate isomers: basal diet

Past experiments in poultry have focused on the analysis of InsP_6 and rarely considered clarification of the location of InsP_6 hydrolysis^(14,32,38). A few studies have investigated the sum of InsP_5 , InsP_4 and InsP_3 isomers in digesta samples of poultry, without differentiation among positional isomers^(39–41). The authors are not aware of any published study investigating positional InsP isomers in the digestive tract of broilers, which led us to making this one of our objectives.

When the BD was fed, the dominating InsP_5 isomers found in the crop were $\text{Ins}(1,2,4,5,6)\text{P}_5$ and $\text{Ins}(1,2,3,4,5)\text{P}_5$. $\text{Ins}(1,2,3,4,6)\text{P}_5$ and $\text{Ins}(1,3,4,5,6)\text{P}_5$ that occurred in concentrations close to the limit of detection may have originated from the diet (Table 1). The changing pattern of InsP_5 isomers (Fig. 1 (A)) in the crop compared with the diet affirm InsP_6 hydrolysis in the crop, as does the additional occurrence of InsP_4 . The appearance of $\text{Ins}(1,2,4,5,6)\text{P}_5$ might have been caused by residual intrinsic soyabean phytase, which is a 3-phytase, that withstood exposure to heat in the desolventiser-toaster and was below the limit of detection in the feed. However, this could also have been caused by microbial phytases because 3-phytases are primarily found in fungi (*A. niger*, *A. terreus*, *A. fumigatus*, *Neurospora crassa*), yeasts (*Saccharomyces castellii*, *Saccharomyces cerevisiae*) and bacteria (*Selenomonas ruminantium*, *Selenomonas lactificifex*, *Megasphaera elsdenii*, *Klebsiella terrigena*, *Pantoea agglomerans*, *Pseudomonas syringae*, *Bacillus subtilis*, *Bacillus amyloliquefaciens*). Phytases of plant origin are primarily classified as 4-/6-phytases but 6-phytases were also found for specific bacteria such as *E. coli*, *Peniophora lycii* and *Bifidobacterium pseudocatenulatum*^(7,42,43). Therefore, it remains open whether in the present study $\text{Ins}(1,2,3,4,5)\text{P}_5$ was formed by plant or microbiota phytase. $\text{Ins}(1,2,5,6)\text{P}_4$ (perhaps co-eluted with $\text{Ins}(2,3,4,5)\text{P}_4$) measured in the crop could have been formed by 3- or 6-phytases^(5,7). A formation by other phosphatases which further degraded $\text{Ins}(1,2,4,5,6)\text{P}_5$ or $\text{Ins}(1,2,3,4,5)\text{P}_5$ to $\text{Ins}(1,2,5,6)\text{P}_4$ also is possible.

The fact that there was so little InsP_5 in the proventriculus/gizzard suggests that when no phytase is added there is very little InsP_6 hydrolysis in that acid environment. Some remaining intrinsic plant phytase may rapidly be inactivated at low pH values and in the presence of pepsin and pancreatin^(44–47). The lack of InsP_{3-4} isomers and the dominance of InsP_5 isomers in the proventriculus/gizzard further indicate that InsP_5 hydrolysis is less rapidly advanced. However, InsP and their complexes are soluble under acidic conditions and mineral chelates of lower InsP s are more soluble. Hence fast breakdown of InsP_{3-4} also could have happened in the proventriculus/gizzard.

Following passage through the acid phase of the proventriculus/gizzard, a higher accessibility of phytate in the posterior segments can be assumed. In the duodenum/jejunum, a greater range of InsP isomers compared with those found in the stomach was found for the BD treatment, showing intense hydrolysis of InsP_6 in this segment. This increase probably was the result of substrate-induced InsP_6 hydrolysis by microbiota or endogenous mucosa phytase. Phytate-induced phytase

production was reported for several bacteria^(48–50). The results of Schlemmer *et al.*⁽¹⁶⁾ showed substrate dependence of microbiota phytase activity in the colon of pigs. For mucosal phytase, increased activity with dietary phytic acid was also reported in rats⁽⁵¹⁾. In addition to an alteration of InsP_6 hydrolysis and diversity of InsP isomers, the InsP pattern changed between the anterior and intestinal segments of the digestive tract (Fig. 1). This change in InsP pattern suggests the involvement of phosphatases of different origin in different segments, with 3- and 6-phytases dominating in the crop and 6- and 5-phytases dominating in the intestinal segments. $\text{Ins}(1,2,3,4,5)\text{P}_5$ can be formed by bacterial 6-phytase of the intestinal microbiota, for example, an *E. coli* 6-phytase. $\text{Ins}(1,2,3,4)\text{P}_4$ first appeared in the duodenum/jejunum and was the dominating InsP_4 isomer in all intestinal segments when the BD was fed. It might have been a hydrolysis product of a 5-phytase because the majority of phytases continue dephosphorylation adjacent to a free hydroxyl group. $\text{Ins}(1,2,3,4)\text{P}_4$ (perhaps co-eluted with $\text{Ins}(1,2,3,6)\text{P}_4$) was also characterised as a hydrolysis product of a 5-phytase in lily pollen and *Selenomonas ruminantium* subsp. *lactilytica*^(52,53). However, as $\text{Ins}(1,2,3,4)\text{P}_4$ was also detected as a minor hydrolysis product of specific 6-phytases, it principally could have been formed by both 5- and 6-phytases. In addition, the involvement of other phosphatases which further degraded InsP_5 cannot be ruled out. Because 5-phytase is described only for *Selenomonas ruminantium* subsp. *lactilytica*⁽⁵³⁾, lily pollen and *Bifidobacterium pseudocatenulatum*⁽⁴³⁾, its origin in the intestine of broilers was unexpected. Human gut-isolated *Bifidobacterium pseudocatenulatum* initiates InsP_6 hydrolysis at the C-6 and C-5 position of the *myo*-inositol ring and proceeds via $\text{Ins}(1,2,3,4)\text{P}_4$ ⁽⁴³⁾, but the authors are not aware of any study that found this species of *Bifidobacterium* in broilers.

For the caeca, the broad pattern of InsP isomers is an indication of a highly diverse microbial population likewise producing several phytate-degrading enzymes. From bacteria occurring in the chicken digestive tract, phytate-degrading activity has been described for *Lactobacillus* spp.⁽³¹⁾, *Enterobacter* spp.⁽⁵⁴⁾, *E. coli*⁽⁷⁾, *Klebsiella pneumoniae*⁽⁵⁵⁾, *Bacillus* spp.⁽⁵⁶⁾, *Bifidobacterium* spp.⁽⁵⁷⁾ and *Pseudomonas aeruginosa*⁽⁵⁸⁾. Differences between the InsP pattern of the intestinal segments might be caused by the differing microbial community composition as described by Lu *et al.*⁽⁵⁹⁾ and coupled with the variations in activity of the endogenous phytase. $\text{Ins}(1,3,4,5,6)\text{P}_5$, which appeared in the caeca, indicates the activity of a phosphatase in the caeca that initiates hydrolysis at the C-2 position of the inositol ring. If this was not undegraded $\text{Ins}(1,3,4,5,6)\text{P}_5$ from the feed then this finding contradicts the general assumption that phytate-degrading enzymes are unable to cleave the axial phosphate group of the *myo*-inositol ring.

Appearance of inositol phosphate isomers: phytase treatments

The second objective of the present study was to investigate the InsP_6 degradation pattern of different phytase supplements and their effectiveness in releasing phosphate in different



segments of the digestive tract. For the PhyA treatment, Ins(1,2,4,5,6)P₅ was the predominant InsP₅ isomer whereas Ins(1,2,3,4,5)P₅ was predominant for the PhyE1 and PhyE2 treatments. This shows, for the first time, that in the crop of broilers the patterns are very similar to the *in vitro* pattern of hydrolysis of these 3- and 6-phytases, initiating InsP₆ hydrolysis at the D-3 (L-1) and D-6 (L-4) positions of the inositol ring^(5,7). Ins(1,2,5,6)P₄ (perhaps co-eluted with Ins(2,3,4,5)P₄) and InsP₃, both of which were present in high concentrations for the phytase treatments, are the two other main hydrolysis products of the three phytases. This pattern conforms with *in vitro* results that showed D-Ins(2,3,4,5)P₄ as a hydrolysis product of *E. coli* and D-Ins(1,2,5,6)P₄ of *Aspergillus* phytase^(5,7). The proportion of Σ InsP₄ in Σ InsP₃₋₅ was higher for PhyE1 and PhyE2 compared with PhyA whereas the proportion of InsP₃ in Σ InsP₃₋₅ was higher for PhyA in the crop. Accumulation of *myo*-inositol tris- and bisphosphates following hydrolysis by *A. niger* phytase has already been shown *in vitro*⁽⁶⁰⁾. In contrast, a fast progression from InsP₅ to InsP₄ is expected for *E. coli*, but InsP₄ accumulated. This result corresponds to *in vitro* findings in which InsP₄ accumulated during InsP₆ hydrolysis by *E. coli* phytase and was later slowly hydrolysed to InsP₃⁽⁶⁾. Because the lower-molecular-weight esters of InsPs have a lower mineral-binding strength than InsP₆ or InsP₅⁽⁶¹⁾, the solubility of these esters in the small intestine will be improved, allowing access to them by the endogenous phytase/phosphatases⁽⁶²⁾.

In the duodenum/jejunum and to some extent also in the ileum, the InsP₅ and InsP₄ isomers specifically formed by the respective supplemented phytases were present in higher concentrations compared with both the BD and the other phytase treatments. Thus, further activity of the enzymes can be assumed in these intestinal segments. However, the proportion of Ins(1,2,3,4,6)P₅ in Σ InsP₅ was significantly lower, and concentrations of Ins(1,2,3,4)P₄ tended to be lower for the phytase treatments compared with the BD treatment in the duodenum/jejunum and lower ileum. Supplementation of phytase tends to reduce lactic acid bacterial count and significantly reduce *E. coli* count in the ileal digesta of broilers⁽⁶³⁾, and, as mentioned earlier, both bacterial groups were suspected to be involved in phytate degradation. Aydin *et al.*⁽⁶³⁾ speculated that the decrease is related to a possible reduction in the quantity of substrate available to the intestinal microbiota. If there was a reduction of lactic acid bacteria (for example, *Bifidobacteria*) this might explain the decrease in 5-phytase activity for all phytase-containing treatments whereas a reduction of *E. coli* bacteria might explain the decrease in 6-phytase activity for PhyA. Furthermore, a reduction in intestinal mucosal phytase activity has been reported when chickens were supplemented with phytase⁽¹⁰⁾. Thus, a decrease in activity of endogenous mucosal phytase might also have contributed to different InsP pattern between the phytase and the BD treatments in the small intestine. In the lower ileum, the significant differences between treatments in Ins(1,2,4,5,6)P₅ concentration persisted, but the differences in Ins(1,2,3,4,5)P₅ concentration did not. An explanation could be that the activity of supplemented phytases was significantly reduced at this point due to increasing pH and/or proteolytic degradation.

Igbasan *et al.*⁽³⁵⁾ detected a residual activity of 60 and 55 % for an *Aspergillus* and 87 and 80 % for an *E. coli* phytase in the jejunal and ileal digesta, respectively. Intestinal phosphatases might continue to hydrolyse InsP₃₋₅ isomers formed by the supplemented PhyE1 and PhyE2, but InsP₅ isomers formed by PhyA seemed to be less degradable by intestinal phosphatases. In this regard, Yu *et al.*⁽⁶⁴⁾ showed that Ins(1,2,4,5,6)P₅ was a more potent aggregator of protein at low pH compared with Ins(1,2,3,4,5)P₅ which probably would reduce susceptibility of this isomer to phosphatase activity in the intestine.

Conclusions

We conclude that broilers and their microbiota have a high capacity to hydrolyse InsP₆ in the intestine. The differentiation between InsP₆ hydrolysis products of endogenous or microbiota phytases and their contribution to InsP₆ hydrolysis in different segments still requires experimental work. Phytase supplements are more effective in the anterior than in the intestinal segments of the digestive tract, supporting *in vitro* properties. The main InsP₆ degradation products of *Aspergillus* and *E. coli* phytases as determined from *in vitro* studies are also formed in the crop and proventriculus/gizzard of broilers. Differences in InsP₆ hydrolysis between PhyE1 and PhyE2 compared with PhyA existing in the crop disappeared until the ileum. InsP₄ accumulated in the crop when PhyE1 and PhyE2 were used. However, InsP₃ accumulated when PhyA was used. It became apparent that the hydrolytic cleavage of the first phosphate group is not the only limiting step in phytate degradation in broilers.

Acknowledgements

The present study was supported by the Ministerium für Wissenschaft, Forschung und Kunst Baden-Württemberg (through a doctoral fellowship for E. Z.).

We would like to thank J. Boguhn for her assistance in trial planning and implementation.

The authors' contributions are as follows: E. Z. designed and performed the experiments, conducted parts of the chemical analysis, analysed data and drafted the manuscript. M. S. established and optimised the high-performance ion chromatography method and contributed to data presentation and interpretation. I. K. contributed to experimental design and data interpretation. M. R. contributed to all stages of the work, especially in the design, critical reflection on and interpretation of results, and preparation of the manuscript. All authors were involved in the discussion of the results and in editing of the manuscript.

The authors declare that there are no conflicts of interest.

References

- Greiner R & Konietzky U (2006) Phytase for food application. *Food Technol Biotech* **44**, 125–140.
- Cosgrove DJ (1970) Inositol phosphate phosphatases of microbiological origin. Inositol phosphate intermediates in the dephosphorylation of the hexaphosphates of *myo*-inositol, scyllo-inositol, and



- D-chiro-inositol by a bacterial (*Pseudomonas* sp.) phytase. *Aust J Biol Sci* **23**, 1207–1220.
3. Skoglund E, Näsi M & Sandberg AS (1998) Phytate hydrolysis in pigs fed a barley–rapeseed meal diet treated with *Aspergillus niger* phytase or steeped with whey. *Can J Anim Sci* **78**, 175–180.
 4. Dvořáková J, Kopecký J, Havlíček V, *et al.* (2000) Formation of *myo*-inositol phosphates by *Aspergillus niger* 3-phytase. *Folia Microbiol* **45**, 128–132.
 5. Greiner R, da Silva LG & Couri S (2009) Purification and characterization of an extracellular phytase from *Aspergillus niger* 11t53a9. *Braz J Microbiol* **40**, 795–807.
 6. Greiner R, Konietzny U & Jany KD (1993) Purification and characterization of two phytases from *Escherichia coli*. *Arch Biochem Biophys* **303**, 107–113.
 7. Greiner R, Carlsson NG & Alminger ML (2000) Stereospecificity of *myo*-inositol hexakisphosphate dephosphorylation by a phytate-degrading enzyme of *Escherichia coli*. *J Biotechnol* **84**, 53–62.
 8. Maenz DD & Classen HL (1998) Phytase activity in the small intestinal brush border membrane of the chicken. *Poultry Sci* **77**, 557–563.
 9. Applegate TJJ, Angel R & Classen HL (2003) Effect of dietary calcium, 25-hydroxycholecalciferol, or bird strain on small intestinal phytase activity in broiler chickens. *Poultry Sci* **82**, 1140–1148.
 10. Abudabos AM (2012) Intestinal phytase activity in chickens (*Gallus domesticus*). *Afr J Microbiol Res* **6**, 4932–4938.
 11. Leytem AB, Willing BP & Thacker PA (2008) Phytate utilization and phosphorus excretion by broiler chickens fed diets containing cereal grains varying in phytate and phytase content. *Anim Feed Sci Tech* **146**, 160–168.
 12. Tamim NM & Angel R (2003) Phytate phosphorus hydrolysis as influenced by dietary calcium and micro-mineral source in broiler diets. *J Agric Food Chem* **51**, 4687–4693.
 13. Tamim NM, Angel R & Christman M (2004) Influence of dietary calcium and phytase on phytate phosphorus hydrolysis in broiler chickens. *Poultry Sci* **83**, 1358–1367.
 14. Kerr MJ, Classen HL & Newkirk RW (2000) The effects of gastrointestinal tract micro-flora and dietary phytase on inositol hexaphosphate hydrolysis in the chicken. *Poultry Sci* **79**, Suppl. 1, 11 (abstract).
 15. Leytem AB, Thacker PA & Turner BL (2007) Phosphorus characterization in feces from broiler chicks fed low-phytate barley diets. *J Sci Food Agr* **87**, 1495–1501.
 16. Schlemmer U, Jany KD, Berk A, *et al.* (2001) Degradation of phytate in the gut of pigs –pathway of gastrointestinal inositol phosphate hydrolysis and enzymes involved. *Arch Anim Nutr* **55**, 225–280.
 17. Kemme PA, Schlemmer U, Mroz Z, *et al.* (2006) Monitoring the stepwise phytate degradation in the upper gastrointestinal tract of pigs. *J Sci Food Agr* **86**, 612–622.
 18. Pontoppidan K, Glitsøe V, Guggenbuhl P, *et al.* (2012) *In vitro* and *in vivo* degradation of *myo*-inositol hexakisphosphate by a phytase from *Citrobacter braakii*. *Arch Anim Nutr* **66**, 431–444.
 19. Rodehutsord M (2009) Approaches and challenges for evaluating phosphorus sources for poultry. In *Proceedings of the 17th European Symposium on Poultry Nutrition*, pp. 2–6 [WPSA UK Branch, editor]. Edinburgh, Scotland.
 20. Abelson PH (1999) A potential phosphate crisis. *Science* **283**, 2015.
 21. Campbell GL & Bedford MR (1992) Enzyme applications for monogastric feeds: a review. *Can J Anim Sci* **72**, 449–466.
 22. Gesellschaft für Ernährungsphysiologie (Society for Nutrition Physiology) (1999) *Empfehlungen zur Energie- und Nährstoffversorgung der Legehennen und Masthühner (Broiler)* (Recommendations for the Energy and Nutrient Supply of Laying Hens and Broilers), 1st ed. Frankfurt am Main: DLG Verlag.
 23. Rodehutsord M, Dieckmann A, Witzig M, *et al.* (2012) A note on sampling digesta from the ileum of broilers in phosphorus digestibility studies. *Poultry Sci* **91**, 965–971.
 24. Verband Deutscher Landwirtschaftlicher Untersuchungs- und Forschungsanstalten (Association of German Agricultural Testing and Research Institutes) (2006) *Handbuch der landwirtschaftlichen Versuchs- und Untersuchungsmethodik (VDLUFU-Methodenbuch)*, vol.3: *Die Chemische Untersuchung von Futtermitteln (Handbook of Agricultural Testing and Research Methodology (VDLUFU Method Book), vol 3: The Chemical Control of Feed)*, 1st ed., Darmstadt: VDLUFU-Verlag.
 25. Boguhn J, Baumgärtel T, Dieckmann A, *et al.* (2009) Determination of titanium dioxide supplements in different matrices using two methods involving photometer and inductively coupled plasma optical emission spectrometer measurements. *Arch Anim Nutr* **63**, 337–342.
 26. Shastak Y, Witzig M, Hartung K, *et al.* (2012) Comparison of retention and prececal digestibility measurements in evaluating mineral phosphorus sources in broilers. *Poultry Sci* **91**, 2201–2209.
 27. Phillippy BQ & Bland JM (1988) Gradient ion chromatography of inositol phosphates. *Anal Biochem* **175**, 162–166.
 28. Chen QC & Li BW (2003) Separation of phytic acid and other related inositol phosphates by high-performance ion chromatography and its applications. *J Chromatogr A* **1018**, 41–52.
 29. Skoglund E, Carlsson NG & Sandberg AS (1997) Determination of isomers of inositol mono- to hexaphosphates in selected foods and intestinal contents using high-performance ion chromatography. *J Agric Food Chem* **45**, 431–436.
 30. Svihus B, Hetland H, Choct M, *et al.* (2002) Passage rate through the anterior digestive tract of broiler chickens fed on diets with ground and whole wheat. *Br Poultry Sci* **43**, 662–668.
 31. Raghavendra P & Halami PM (2009) Screening, selection and characterization of phytic acid degrading lactic acid bacteria from chicken intestine. *Int J Food Microbiol* **133**, 129–134.
 32. Marounek M, Skřivan M, Rosero O, *et al.* (2010) Intestinal and total tract phytate digestibility and phytase activity in the digestive tract of hens fed a wheat–maize–soyabean diet. *J Anim Feed Sci* **19**, 430–439.
 33. Sacranie A, Svihus B, Denstadli V, *et al.* (2012) The effect of insoluble fiber and intermittent feeding on gizzard development, gut motility, and performance of broiler chickens. *Poultry Sci* **91**, 693–700.
 34. Brüning P (2009) *In-vitro*- und *in-vivo*-Untersuchungen zur Effizienz verschiedener mikrobieller Phytasen als Futterzusatzstoff (*In vitro*- and *in vivo*-studies on the efficiency of various microbial phytases as a feed additive). PhD Thesis, Free University Berlin.
 35. Igbasan FA, Männer K, Miksch G, *et al.* (2000) Comparative studies on the *in vitro* properties of phytases from various microbial origins. *Arch Anim Nutr* **53**, 353–373.
 36. Garrett JB, Kretz KA, O'Donoghue E, *et al.* (2004) Enhancing the thermal tolerance and gastric performance of a microbial phytase for use as a phosphate-mobilizing monogastric-feed supplement. *Appl Environ Microbiol* **70**, 3041–3046.
 37. Elkhail EAI, Männer K, Borriss R, *et al.* (2007) *In vitro* and *in vivo* characteristics of bacterial phytases and their efficacy in broiler chickens. *Br Poultry Sci* **48**, 64–70.
 38. Leytem AB, Widyaratne GP & Thacker PA (2008) Phosphorus utilization and characterization of ileal digesta and excreta from broiler chickens fed diets varying in cereal grain, phosphorus level, and phytase addition. *Poultry Sci* **87**, 2466–2476.
 39. Leslie MA (2006) Effect of phytase and glucanase, alone or in combination, on nutritive value of corn and soybean meal fed to broilers. PhD Thesis, Auburn University, AL.
 40. Leytem AB, Kwanyuen P, Plumstead PW, *et al.* (2008) Evaluation of phosphorus characterization in broiler ileal digesta, manure, and litter samples: ³¹P-NMR vs. HPLC. *J Environ Qual* **37**, 494–500.
 41. Elkhail EAI, Osman HAH & Simon O (2011) Phytic acid analysis by different bacterial phytases. *Aust J Basic Appl Sci* **5**, 2295–2302.
 42. Van der Kaay J & Van Haastert PJM (1995) Stereospecificity of inositol hexakisphosphate dephosphorylation by *Paramecium* phytase. *Biochem J* **312**, 907–910.
 43. Haros M, Carlsson NG, Almgren A, *et al.* (2009) Phytate degradation by human gut isolated *Bifidobacterium pseudocatenuatum* ATCC27919 and its probiotic potential. *Int J Food Microbiol* **135**, 7–14.



44. Greiner R, Konietzny U & Jany KD (1998) Purification and properties of a phytase from rye. *J Food Biochem* **22**, 143–161.
45. Phillippy BQ (1999) Susceptibility of wheat and *Aspergillus niger* phytases to inactivation by gastrointestinal enzymes. *J Agric Food Chem* **47**, 1385–1388.
46. Greiner R, Jany KD & Larsson Alminger M (2000) Identification and properties of *myo*-inositol hexakisphosphate phosphohydrolases (phytases) from barley (*Hordeum vulgare*). *J Cereal Sci* **31**, 127–139.
47. Greiner R, Muzquiz M, Burbano C, *et al.* (2001) Purification and characterization of a phytate-degrading enzyme from germinated faba beans (*Vicia faba* var. *Alameda*). *J Agric Food Chem* **49**, 2234–2240.
48. Greiner R, Haller E, Konietzny U, *et al.* (1997) Purification and characterization of a phytase from *Klebsiella terrigena*. *Arch Biochem Biophys* **341**, 201–206.
49. Kerovuo J, Lauraeus M, Nurminen P, *et al.* (1998) Isolation, characterization, molecular gene cloning, and sequencing of a novel phytase from *Bacillus subtilis*. *Appl Environ Microbiol* **64**, 2079–2085.
50. Lan GQ, Abdullah N, Jalaludin S, *et al.* (2002) Culture conditions influencing phytase production of *Mitsuokella jalaludinii*, a new bacterial species from the rumen of cattle. *J Appl Microbiol* **93**, 668–674.
51. Lopez HW, Vallery F, Levrat-Verny MA, *et al.* (2000) Dietary phytic acid and wheat bran enhance mucosal phytase activity in rat small intestine. *J Nutr* **130**, 2020–2025.
52. Barrientos L, Scott JJ & Murthy PPN (1994) Specificity of hydrolysis of phytic acid by alkaline phytase from lily pollen. *Plant Physiol* **106**, 1489–1495.
53. Puhl AA, Greiner R & Selinger LB (2008) A protein tyrosine phosphatase-like inositol polyphosphatase from *Selenomonas ruminantium* subsp. *lactilytica* has specificity for the 5-phosphate of *myo*-inositol hexakisphosphate. *Int J Biochem Cell B* **40**, 2053–2064.
54. Yoon SJ, Choi YJ, Min HK, *et al.* (1996) Isolation and identification of phytase-producing bacterium, *Enterobacter* sp. 4, and enzymatic properties of phytase enzyme. *Enzyme Microb Tech* **18**, 449–454.
55. Escobin-Mopera L, Ohtani M, Sekiguchi S, *et al.* (2012) Purification and characterization of phytase from *Klebsiella pneumoniae* 9-3B. *J Biosci Bioeng* **113**, 562–567.
56. Greiner R, Farouk A, Alminger ML *et al.* (2002) The pathway of dephosphorylation of *myo*-inositol hexakisphosphate by phytate-degrading enzymes of different *Bacillus* spp. *Can J Microbiol* **48**, 986–994.
57. Palacios MC, Haros M, Rosell CM, *et al.* (2008) Selection of phytate-degrading human bifidobacteria and application in whole wheat dough fermentation. *Food Microbiol* **25**, 169–176.
58. Sasirekha B, Bedashree T & Champa KL (2012) Statistical optimization of medium components for improved phytase production by *Pseudomonas aeruginosa*. *Int J Chem Tech Res* **4**, 891–895.
59. Lu J, Idris U, Harmon B *et al.* (2003) Diversity and succession of the intestinal bacterial community of the maturing broiler chicken. *Appl Environ Microbiol* **69**, 6816–6824.
60. Wyss M, Pasamontes L, Friedlein A, *et al.* (1999) Biophysical characterization of fungal phytases (*myo*-inositol hexakisphosphate phosphohydrolases): molecular size, glycosylation pattern, and engineering of proteolytic resistance. *Appl Environ Microbiol* **65**, 359–366.
61. Persson H, Türk M, Nyman M, *et al.* (1998) Binding of Cu^{2+} , Zn^{2+} , and Cd^{2+} to inositol tri-, tetra-, penta-, and hexaphosphates. *J Agric Food Chem* **46**, 3194–3200.
62. Cowieson AJ, Wilcock P & Bedford MR (2011) Super-dosing effects of phytase in poultry and other monogastrics. *World Poultry Sci J* **67**, 225–235.
63. Aydin A, Pekel AY, Issa G, *et al.* (2010) Effects of dietary copper, citric acid, and microbial phytase on digesta pH and ileal and carcass microbiota of broiler chickens fed a low available phosphorus diet. *J Appl Poultry Res* **19**, 422–431.
64. Yu S, Cowieson A, Gilbert C, *et al.* (2012) Interactions of phytate and *myo*-inositol phosphate esters (IP1–5) including IP5 isomers with dietary protein and iron and inhibition of pepsin. *J Anim Sci* **90**, 1824–1832.