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# ICTV Virus Taxonomy Profile: Partitiviridae

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# ICTV Virus Taxonomy Profile: Partitiviridae

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#### Abstract

The Partitiviridae is a family of small, isometric, non-enveloped viruses with bisegmented double-stranded (ds) RNA genomes of 3-4.8 kbp. The two genome segments are individually encapsidated. The family has five genera, with characteristic hosts for members of each genus: either plants or fungi for genera Alphapartitivirus and Betapartitivirus, fungi for genus Gammapartitivirus, plants for genus Deltapartitivirus and protozoa for genus Cryspovirus. Partitiviruses are transmitted intracellularly via seeds (plants), oocysts (protozoa) or hyphal anastomosis, cell division and sporogenesis (fungi); there are no known natural vectors. This is a summary of the International Committee on Taxonomy of Viruses (ICTV) Report on the taxonomy of the Partitiviridae, which is available at www.ictv.global/report/partitiviridae.

#### Table 1. Characteristics of the family Partitiviridae

Typical member:	Atkinsonella hypoxylon virus, 2H (RNA1, L39125; RNA2, L39126), species Atkinsonella hypoxylon virus, genus Betapartitivirus
Genome	3-4.8 kbp of linear bisegmented dsRNA
Virion	Isometric, non-enveloped, 25–43 nm in diameter; dsRNA1 and dsRNA2 are separately encapsidated
Replication	Cytoplasmic. Genomic RNA acts as a template for mRNA synthesis within the virus particle; transcription occurs by a semiconservative mechanism
Translation	From monocistronic positive-sense transcripts of both genomic dsRNAs
Host range	Plants, fungi and protozoa
Taxonomy	Five genera, including >40 species, and 15 species unassigned to a genus

## VIRION

Virus particles are isometric, non-enveloped, and 25-43 nm in diameter (Table 1, Fig. 1a, b). Each capsid is composed of 120 copies of a single protein arranged as 60 dimers with T=1icosahedral symmetry [1]. Dimeric surface protrusions are frequently observed on viral capsids. One or two molecules of RNA-dependent RNA polymerase (RdRP) are packaged inside each particle [2].

## REPLICATION

Each dsRNA is monocistronic. The RdRP is believed to function as both a transcriptase and a replicase and catalyzes in vitro end-to-end transcription of each dsRNA to produce mRNA by a semi-conservative mechanism. Virions accumulate in the cytoplasm.

## GENOME

Members of all five genera possess two essential genome segments, dsRNA1 and dsRNA2, each containing one large ORF on the positive-strand RNA molecule (Fig. 2). The smaller of the two dsRNA genome segments usually encodes the coat protein (CP) and the larger usually encodes the virion-associated RNA polymerase. The linear dsRNA segments are separately encapsidated. Additional dsRNA segments (satellite or defective) may also be present.

## TAXONOMY

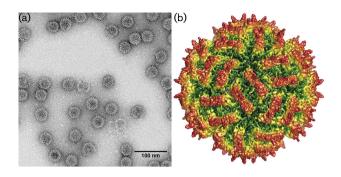
## Alphapartitivirus

Members of the genus Alphapartitivirus infect either plants, or ascomycetous or basidiomycetous fungi. The two

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Keywords: Partitiviridae; ICTV; taxonomy; Alphapartitivirus; Betapartitivirus; Deltapartitivirus; Gammapartitivirus; Cryspovirus. Abbreviations: CP, coat protein; RdRP, RNA-dependent RNA polymerase.



**Fig. 1.** (a) Transmission electron micrograph of negatively-stained purified particles of Penicillium stoloniferum virus S, a representative member of the genus *Gammapartitivirus*. (b) Cryo-EM reconstructions of Penicillium stoloniferum virus S at 0.45 nm resolution, and rendered with radial colour mapping.

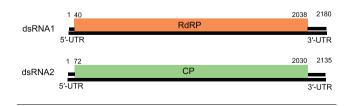
essential dsRNA genome segments are individually about 1.9–2.0 kbp (dsRNA1) and 1.7–1.9 kbp (dsRNA2), typically containing a poly(A) tract near the plus-strand 3'-terminus. There is a single major CP with predicted Mr of 51–57 kDa. Plant alphapartitiviruses cause persistent infections, whereas some fungal alphapartitiviruses cause host effects, such as hypovirulence or a reduced growth rate [3, 4].

### Betapartitivirus

Members of the genus *Betapartitivirus* infect either plants, or ascomycetous or basidiomycetous fungi. The two essential dsRNA genome segments are about 2.2–2.4 kbp (dsRNA1) and 2.1–2.4 kbp (dsRNA2), typically containing a poly(A) tract near the plus-strand 3'-terminus. There is a single major CP with predicted Mr of 71–77 kDa. Plant betapartitiviruses cause persistent infections [5, 6]. Some fungal betapartitiviruses cause reduced host virulence and changes in colony morphology [7].

#### Gammapartitivirus

All known members of the genus *Gammapartitivirus* infect ascomycetous fungi. The two essential dsRNA segments are about 1.6–1.8 kbp (dsRNA1) and 1.4–1.6 kbp (dsRNA2). There is a single major CP with predicted Mr of 44–47 kDa. Most gammapartitiviruses seem to induce latent infections. Aspergillus fumigatus partitivirus 1, a related, unclassified virus, has been associated with host effects.



**Fig. 2.** Atkinsonella hypoxylon virus [10], an isolate of the type species of the genus *Betapartitivirus*, has a bipartite genome consisting of dsRNA1 and dsRNA2.

#### Deltapartitivirus

All known members of the genus *Deltapartitivirus* induce persistent infections in plants [8]. They are transmitted by ovule and pollen to the seed embryo. The two essential dsRNA segments are individually 1.6-1.7 kbp (dsRNA1) and 1.4-1.6 kbp (dsRNA2). There is a single major CP with predicted *Mr* of 38–49 kDa.

## Cryspovirus

Members of the genus *Cryspovirus* infect apicomplexan protozoa of the genus *Cryptosporidium* [9]. The viral genome comprises two dsRNA segments, which are individually 1.5 and 1.8 kbp. There is a single major CP with predicted *Mr* of 37 kDa. Virions are disseminated within *Cryptosporidium* oocysts. Infections of the *Cryptosporidium* host cells appear to be latent.

## RESOURCES

Full ICTV Online (10th) Report: www.ictv.global/report/partitiviridae.

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#### Conflicts of interest

The authors declare that there are no conflicts of interest.

#### References

- Nibert ML, Tang J, Xie J, Collier AM, Ghabrial SA et al. 3D structures of fungal partitiviruses. Adv Virus Res 2013;86:59–85.
- Nibert ML, Ghabrial SA, Maiss E, Lesker T, Vainio EJ et al. Taxonomic reorganization of family *Partitiviridae* and other recent progress in partitivirus research. *Virus Res* 2014;188: 128–141.
- Chiba S, Lin YH, Kondo H, Kanematsu S, Suzuki N. Effects of defective interfering RNA on symptom induction by, and replication of, a novel partitivirus from a phytopathogenic fungus, *Rosellinia necatrix*. J Virol 2013;87:2330–2341.
- Vainio EJ, Hantula J. Taxonomy, biogeography and importance of Heterobasidion viruses. Virus Res 2016;219:2–10.
- Lesker T, Rabenstein F, Maiss E. Molecular characterization of five betacryptoviruses infecting four clover species and dill. Arch Virol 2013;158:1943–1952.
- 6. Roossinck MJ. Lifestyles of plant viruses. *Philos Trans R Soc Lond B Biol Sci* 2010;365:1899–1905.
- Xiao X, Cheng J, Tang J, Fu Y, Jiang D et al. A novel partitivirus that confers hypovirulence on plant pathogenic fungi. J Virol 2014; 88:10120–10133.
- Sabanadzovic S, Valverde RA. Properties and detection of two cryptoviruses from pepper (*Capsicum annuum*). Virus Genes 2011; 43:307–312.
- Nibert ML, Woods KM, Upton SJ, Ghabrial SA. Cryspovirus: a new genus of protozoan viruses in the family Partitiviridae. Arch Virol 2009;154:1959–1965.
- Oh CS, Hillman BI. Genome organization of a partitivirus from the filamentous ascomycete Atkinsonella hypoxylon. J Gen Virol 1995; 76:1461–1470.