

Identification and Characterization of Toxigenic *Fusaria* Associated with Sorghum Grain Mold Complex in India

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Abstract *Fusarium* species are dominant within the sorghum grain mold complex. Some species of *Fusarium* involved in grain mold complex produce mycotoxins, such as fumonisins. An attempt was made to identify *Fusarium* spp. associated with grain mold complex in major sorghum-growing areas in India through AFLP-based grouping of the isolates and to further confirm the species by sequencing part of α -Elongation factor gene and comparing the sequences with that available in the NCBI database. The dendrogram generated from the AFLP data clustered the isolates into 5 groups. Five species of *Fusarium*—*F. proliferatum*, *F. thapsinum*, *F. equiseti*, *F. andiyazi* and *F. sacchari* were identified based on sequence similarity of α -Elongation factor gene of the test isolates with those in the NCBI database. *Fusarium thapsinum* was identified as predominant species in *Fusarium*—grain mold complex in India and *F. proliferatum* as highly toxigenic for fumonisins production. Analysis of molecular variance (AMOVA) revealed 54% of the variation in the AFLP patterns of 63 isolates was due to the differences between *Fusarium* species, and 46% was due to differences between the strains within a species.

Keywords AFLP · α -Elongation factor · *Fusarium* · Mycotoxin · Fumonisin

Introduction

Grain mold is a major biotic constraint to production, marketing and utilization of grain sorghum. The term “grain mold” is used to describe the diseased appearance of sorghum grains resulting from infection by one or more pathogenic/saprophytic fungi. The disease is particularly important on improved, short- and medium-duration sorghum cultivars that mature during the rainy season in humid, tropical and subtropical climates. Several fungal species of the genera *Fusarium*, *Curvularia*, *Alternaria*, *Phoma*, *Bipolaris* and *Colletotrichum* have been reported to be associated with grain mold [1]. Of these, *Fusarium* spp. are dominant within the grain mold complex. Some species of *Fusarium* (*F. andiyazi*, *F. proliferatum*, *F. thapsinum* and *F. sacchari*) involved in grain mold complex produce mycotoxins, such as fumonisins, moniliformin, trichothecenes and fusaproliferin [2]. The secondary invading *Aspergillus* spp. may produce aflatoxins in the contaminated grains as well. These toxins reduce the quality of grains as food/feed sources as well as the value of grains in the market. These toxins are associated with a variety of human and animal health problems

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including acute toxicity, increased incidence of cancer, inhibition of normal growth and development, immunity suppression and increased disease susceptibility, increased risks of birth defects and reduced nutritional quality of resulting grain. Risk analysis studies have indicated that when these molds affect staple food grains such as sorghum and maize, the significance to human and animal health is considerable because their quantum of consumption is higher than that of other foods. The first report of fumonisins toxicity to human and poultry came in 1998 from the Deccan plateau of India [3]. An outbreak of food poisoning characterized by abdominal pain and diarrhea, attributed to the ingestion of fumonisins-contaminated maize and sorghum, had been reported from several villages in India [4]. The disease was observed only in adults who were consuming the molded grains. The affected people mostly belonged to lower socio-economic groups, such as marginal farmers and landless agricultural laborers who did not have access to other staples. Similarly, an outbreak of trichothecene mycotoxicosis associated with the consumption of mold-damaged wheat had been reported from Kashmir Valley in India [5]. To reduce levels of fumonisin in grain, efforts have been made to identify sources of maize with increased resistance to fungal infection and fumonisin contamination [6]. Although mycotoxins have been detected from various food grains infected with *Fusarium*, information regarding the profile of toxin-producing species of *Fusarium* involved in sorghum grain mold complex in India is not yet available. Since several species of *Fusarium* have been reported to be associated with this disease, it is essential to identify more precisely the toxigenic species of *Fusarium* and genetic resistance in sorghum against potential toxigenic strains.

Materials and Methods

Identification of *Fusarium* spp. Through DNA Markers

Six hundred and seventy-two isolates of *Fusarium* collected from naturally molded sorghum grains from five locations (Akola, Parbhani, Patancheru, Palem and Surat) in India during 2003–2005 were inoculated on potato dextrose agar (PDA) plates. Morphological/cultural characters of these 672 isolates were studied

on PDA and banana leaf agar [7]. From these, 63 isolates from different regions representing different species of *Fusarium* (initial identification based on morphological/cultural characters) were selected to study genetic relatedness/diversity through amplified fragment length polymorphism (AFLP) and sequence-based identification, and their fumonisins production levels. Three- to four-day-old mycelia of the *Fusarium* isolates grown in potato dextrose broth were harvested by filtration and washed in sterile deionized water. The mycelia were ground using liquid nitrogen, and genomic DNA of the test isolates was extracted as per Kim et al. [8].

AFLP Analysis

AFLPs were generated based on selective amplification of DNA restriction fragments [9]. Five *EcoR1-Mse1* AFLP[®] primer combinations (Etg + Mcag, Eaa + Mctt, Eac + Mcag, Etc + Mctt and Etg + Mcag) were examined in the 63 isolates. The analysis was carried out using the commercial kit (Life Technologies, USA) following the manufacturer's protocols with slight modifications. Primary template DNA was prepared in a one-step restriction–ligation reaction. Fungal genomic DNA (400 ng) was digested with *EcoR1* and *Mse1* at 37°C for 2 h and heated at 70°C for 15 min to inactivate the enzyme. The DNA fragments were ligated to *EcoR1* and *Mse1* adapters at 20°C for 2 h. After terminating the reaction, the ligation mixture was diluted 10-fold with TE buffer and the fragments were pre-amplified in a thermal cycler (MJ Research, USA) using a temperature cycle of 94°C for 30 s, 56°C for 1 min and 72°C for 1 min, in a total of 30 cycles. Five *EcoR1-Mse1* AFLP primer combinations, each with two-base extension in *EcoR1* and three-base extension in *Mse1* primer, were examined in the 63 isolates. The *EcoR1* primer was labeled with [γ -³²P]-ATP (3,000 Ci/mmol), and the selective amplification was carried out according to the manufacturer's protocol. After selective amplification, the PCR products in 3 μ l sub-samples were separated by electrophoresis on 6% denaturing polyacrylamide DNA sequencing gel containing 7.5 M urea. Autoradiograms were obtained using Kodak X-Omat film.

AFLP profiles of 63 isolates were used to construct a binary matrix. Each band was scored as present (1) or absent (0) across the isolates. The data were then analyzed using Numerical Taxonomy System

Version 2.2 (NTSYSpc). The proximity matrix was computed using Dice similarity coefficient, and a dendrogram was constructed by unweighted pair group method of arithmetic averages (UPGMA) using the SAHN (sequential agglomerative hierarchical nested) cluster analysis module [10]. Analysis of molecular variance (AMOVA) version 1.55 was used to estimate variance components for the AFLP patterns and to partition the total variance into ‘within species’ and ‘among species’ [11]. To see the significance of variance components, 1,000 permutations were used.

Phylogenetic Analyses

DNA sequence variation was assessed by sequencing part of α -*Elongation factor* (EF-1 α) using the gene-specific primer pair using BigDye Terminator cycle sequencing kit on ABI3130XL (Applied Biosystems, CA, USA). EF1 (5'-ATGGGTAAGGARGACAAGA C-3') and EF2 (5'-GGARGTACCAGTSATCATGT T-3') primers [12] were used to amplify ~650-bp fragment, a part of EF-1 α gene of *Fusarium*. PCR was performed in a 25 μ l final reaction mixture containing 2.5 μ l 10 \times buffer, 1 U of XT-Taq, 0.5 μ l dNTP mix (2.5 mM each), 2 μ l template DNA (20–50 ng) and 1 μ l each of both forward and reverse primers (10 nM). DNA amplification was performed in a PCR thermocycler (Gene Amp PCR system 9700, Perkin–Elmer, USA) using an initial 5 min denaturation at 95°C followed by 34 cycles of 1 min denaturation at 95°C, 50 s annealing at 50°C and 1 min extension at 72°C followed by a final extension of 10 min at 72°C. The PCR amplification products were checked on 1.5% agarose gel electrophoresis. The EF-1 α PCR amplified products were sequenced using specific primers. Sequences of the EF1-PCR fragments were searched against those in the NCBI database using BLAST [13] for species identification. The sequences were aligned through CLUSTAL W. The dissimilarity index was calculated from the aligned sequences of different isolates using ‘Simple matching’ option in DARwin, and a weighted neighbor-joining tree was constructed.

Detection of Fumonisin B₁ (FB₁)

All the single-spore cultures of test isolates were inoculated into Erlenmeyer flasks containing 50 g sterilized sorghum grains. The flasks were incubated for 4 weeks at 25°C for fumonisin production. The

Fusarium-colonized sorghum grains were removed from the flasks and made into powder using blender, and 20 g sub-sample was drawn for toxin extraction. Each 20 g sample was extracted with 100 ml of solvent containing 70 ml methanol + 30 ml water + 0.5 g KCl and blended for 2 min. The extracts were transferred to conical flask and shaken for 30 min at 300 rpm before filtering through Whatman No. 4. Then, the extracts were diluted 1:10 with 0.2% bovine serum albumin prepared in PBS–Tween, pH 7.4 [14].

All the methanol extracts were subjected to indirect competitive enzyme-linked immunosorbent assay (cELISA) as per method described by Yu and Chu [15] and Waliar et al. [16] with little modifications. In general, ELISA plates (Maxi-sorp, Nunc A/S, DK-4000, Roskilde, Denmark) were coated with FB₁–BSA conjugate prepared in carbonate buffer at a concentration of 200 ng/ml and dispensed 150 μ l/well. At each step, plates were incubated at 37°C for 1 h followed by three washes with PBS–Tween. Then, the plates were treated with 0.2% bovine serum albumin (BSA) prepared in PBS–Tween (PBST–BSA), followed by competition step in which standard or samples, and specific antibodies were added in the plate. The FB₁ concentrations of the standards starting from 100.00, 50.00, 25.00, 12.50, 6.25, 3.12, 1.56 and 0.78 ng/ml was used, and each concentration was duplicated in two wells. Similarly, duplicates of each test sample in two wells were maintained. To the diluted sample extract or standards, 50 μ l of FB₁ antiserum diluted to 1:5,000 in 0.2% PBST–BSA was added. Then, alkaline phosphatase-labeled goat antirabbit IgG conjugate diluted to 1:4,000 in PBST–BSA was added to the plate, followed by substrate *p*-nitrophenyl phosphate prepared in 10% diethanolamine. After 30-min incubation at room temperature, plates were read at 405 nm in an ELISA reader (Multiskan plus Labsystems). Regression curve was drawn using Log₁₀ values of toxin concentration plotted on the Y-axis and optical density values plotted on the X-axis, and using the regression equation, FB₁ concentration was determined.

Results

Genetic Variation

A high level of polymorphism was observed among isolates following selective amplification with 5 AFLP

primer combinations (Etg + Mcag, Eaa + Mctt, Eac + Mcag, Etc + Mctt and Etg + Mcag). A total of 473 bands were amplified, of which 470 were polymorphic. The dendrogram generated from the AFLP data revealed genetic diversity among the isolates by clustering them into 5 groups. Of the 63 isolates, 31 were clustered in one major group (group IV) with ~70% genetic similarity among isolates (Fig. 1). The remaining 32 isolates were clustered in 4 major groups, and many sub-groups within main clusters indicating the involvement of many species of *Fusarium* in the grain mold complex. Group I clustered 12 isolates, group II 3, group III 10 and group V 7 isolates.

α -Elongation Factor Sequence–Based Identification

The sequences of EF-1 α gene from each of the 63 *Fusarium* isolates (query) were compared to those from various known species of *Fusarium* (subject) in the NCBI database. Five species of *Fusarium*—*F. proliferatum*, *F. thapsinum*, *F. equiseti*, *F. andiyazi* and *F. sacchari* were identified based on sequence similarity. Neighbor-joining tree constructed using α -Elongation factor gene sequence of test isolates clustered the isolates of same species in the distinct groups (Fig. 2).

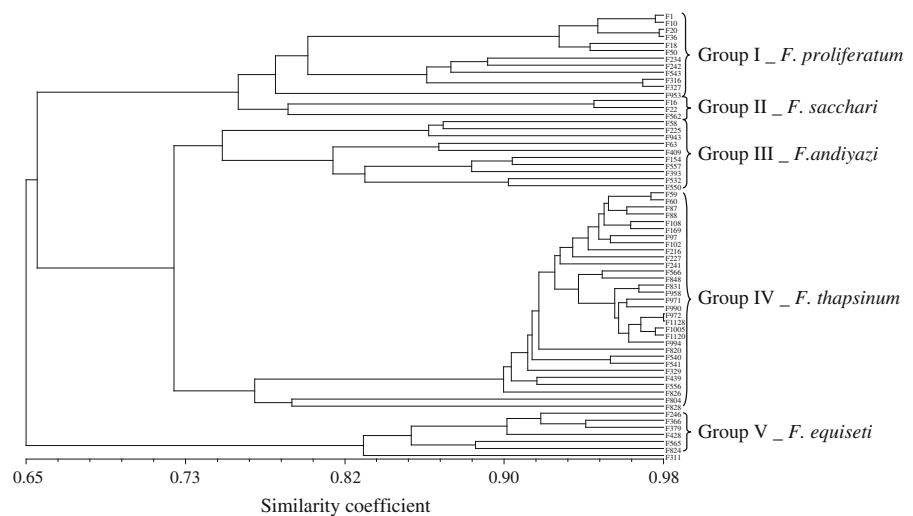
Analysis of Molecular Variance

Analysis of molecular variance (AMOVA) revealed 54% of the variance in the AFLP patterns of 63 isolates was due to the differences between different species of *Fusarium*, and 46% was due to differences between the isolates within a species (Table 1). The variance component exhibited almost 50% variation because of genetic variation between and within *Fusarium* species which indicate large variation between strains/isolates of the same species.

Identification of Toxigenic *Fusarium* spp.

Strains of *F. proliferatum* were identified as highly toxigenic for fumonisin production followed by *F. andiyazi* (Fig. 3). Isolate F 242 (*F. proliferatum*) was the highest fumonisin (B₁)-producing strain (476,539 $\mu\text{g kg}^{-1}$ seed). Ten of the 12 *F. proliferatum* isolates produced high levels (>28,000 $\mu\text{g kg}^{-1}$ seed) of fumonisins, whereas two isolates F316 and F 953 produced less (<300 $\mu\text{g kg}^{-1}$ seed) fumonisin. Two isolates of *F. andiyazi* F 409 and F 943 also produced high levels (>100,000 $\mu\text{g kg}^{-1}$ seed) of fumonisin. Strains of *F. thapsinum*, *F. equiseti* and *F. sacchari* were non-fumonisin producers; however, one isolate of *F. thapsinum* (F88) produced good amount of fumonisins. One isolate of *F. sacchari* (F 22) and two isolates of *F. thapsinum*

Fig. 1 Dendrogram depicting genetic relatedness among 63 isolates of *Fusarium* based on similarity coefficient calculated from AFLP fingerprints



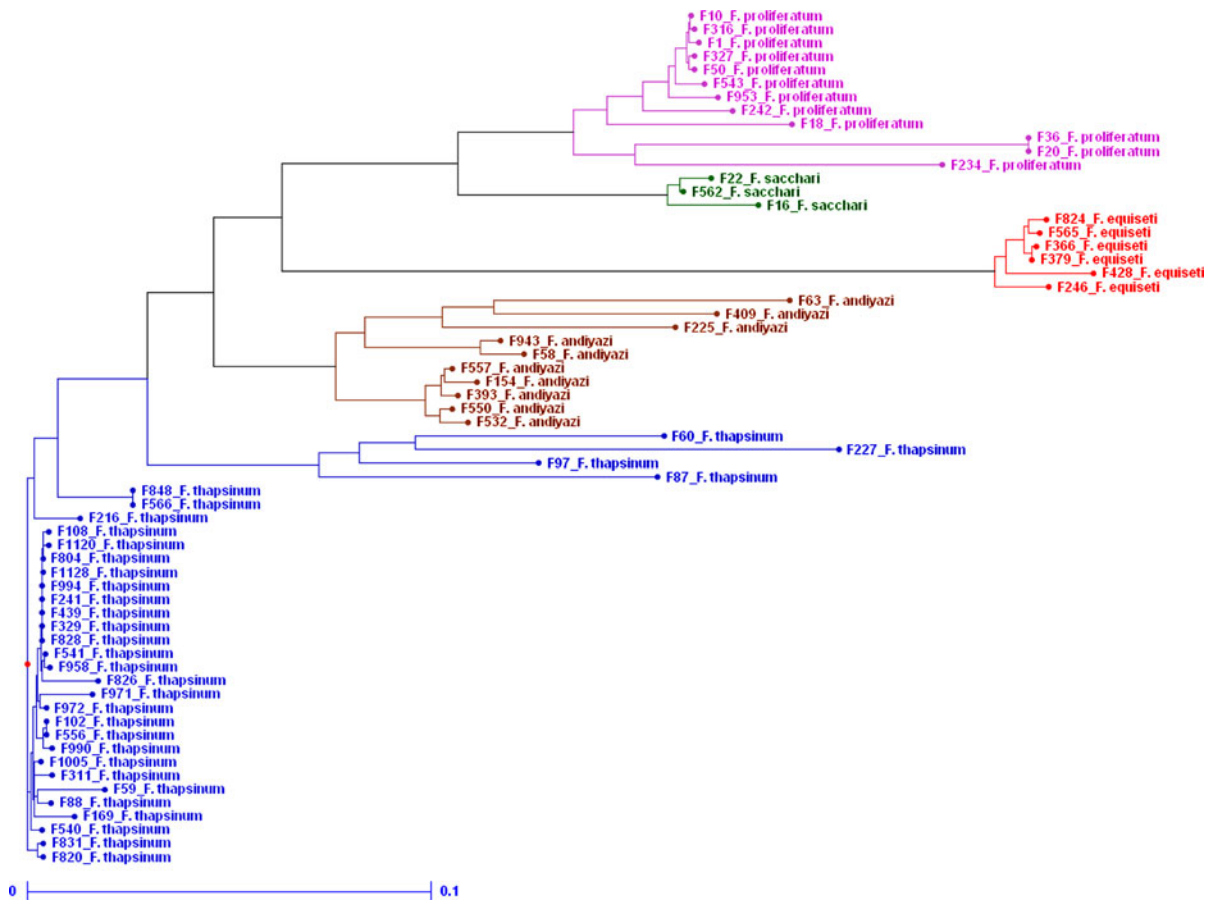


Fig. 2 Neighbor-joining consensus tree for translation elongation factor-1 α (EF-1 α) sequences of *Fusarium* spp.

Table 1 AMOVA for partitioning AFLP variation within and between *Fusarium* species

Source of variation	<i>df</i>	Sum of squares	Variance component	Total (%)
Among species	4	1829.35	40.97 Va	54.41
Within species	58	1990.71	34.32 Vb	45.59
Total	62	3820.06		

Fixation index (*F_{st}*) = 0.544

(F 241 and F 329) produced about 1,500 μg fumonisin kg^{-1} seed.

Prevalence of *Fusarium* spp. in Major Sorghum-Growing Areas in India

Based on α -Elongation factor sequence similarity, 63 *Fusarium* isolates were identified as five species of *Fusarium*—*F. proliferatum*, *F. thapsinum*, *F. equiseti*,

F. andiyazi and *F. saccharii*. Frequency of occurrence of these species varied across locations (Table 2). *Fusarium thapsinum* was identified as predominant (32 of the 63 isolates tested) species in *Fusarium*—grain mold complex. *F. thapsinum*, *F. proliferatum* and *F. andiyazi* were present at all the five locations, whereas *F. equiseti* was not detected at Patancheru and Surat, and *F. saccharii* was present only at Patancheru and Palem.

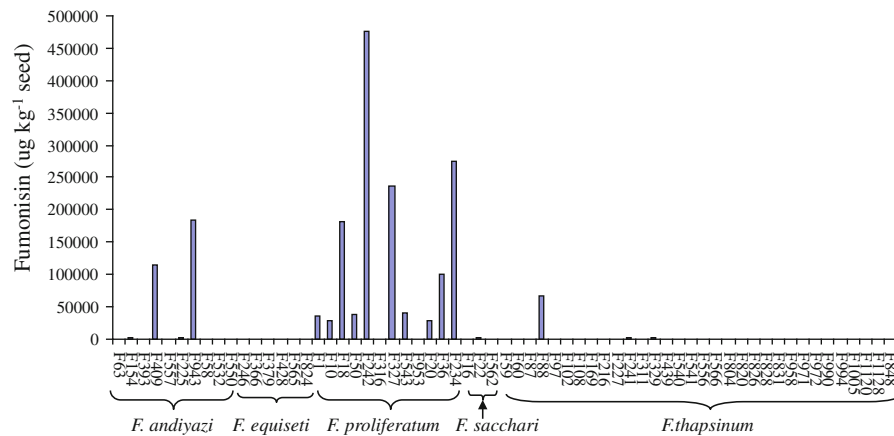


Fig. 3 Fumonisin ($\mu\text{g kg}^{-1}$ seed) produced by strains of *Fusarium* spp.

Table 2 Prevalence of different *Fusarium* spp. associated with sorghum grain mold complex in major sorghum growing areas in India

Location/state	Total samples	<i>Fusarium</i> spp.*				
		<i>F. andiyazi</i>	<i>F. proliferatum</i>	<i>F. equiseti</i>	<i>F. sacchari</i>	<i>F. thapsinum</i>
Patancheru/Andhra Pradesh	17	2	6	–	2	7
Palem/Andhra Pradesh	13	4	1	1	1	6
Surat/Gujarat	10	1	1	–	–	8
Parbhani/Maharashtra	10	2	2	3	–	3
Akola/Maharashtra	13	1	2	2	–	8
Total	63	10	12	6	3	32

*Species identified based on α -Elongation factor sequence similarity

Discussion

There is a general consensus that *Fusarium* growing on sorghum belongs mainly to *Gibberella fujikuroi* species complex, which comprises *Fusarium* species included in section *Liseola* [2]. Since several species of *Fusarium* have been reported to be associated with sorghum grain mold [17], and most of them are morphological similar, it is essential to identify species of *Fusarium* causing grain mold more precisely. Therefore, an attempt was made to identify *Fusarium* spp. associated with grain mold complex from five locations (Patancheru, Parbhani, Palem, Surat and Akola) representing major sorghum growing areas in India using AFLP-based grouping of the isolates and to further confirm the species by sequencing part of α -Elongation factor gene and

comparing the sequences with that available in NCBI database.

Dendrogram generated from AFLP data obtained from selective amplification with 5 primer combinations (Etg + Mcag, Eaa + Mctt, Eac + Mcag, Etc + Mctt and Etg + Mcag) clustered the isolates in 5 distinct groups. Since isolates of the same species of *Fusarium* have been reported to have >65% genetic similarity [2], it was expected that the isolates representing each group with more than 70% similarity to be the same species. The grouping of each species in distinct cluster was confirmed by α -Elongation factor sequence-based species identification. Neighbor-joining tree constructed using α -Elongation factor gene sequence of test isolates clustered the isolates of same species in the distinct groups. A very high level of correspondence was

observed between AFLP and α -Elongation factor sequence-based grouping with only one exception. Isolate F 311 was identified as *F. thapsinum*, but it clustered along with *F. equiseti* group in the AFLP generated dendrogram. AMOVA results showed that both within and between species variance was high indicating that strains variation also exist in *Fusarium* species associated with sorghum grain mold.

Frequency of occurrence of different species also varied across different locations. Of the 63 isolates analyzed, 32 were identified as *F. thapsinum*, followed by *F. proliferatum* (12 isolates) and *F. andiyazi* (10 isolates) and were present at all the five locations from where the grain mold samples were collected, whereas *F. equiseti* (6 isolates) and *F. sacchari* (3 isolates) were present in small proportion. *Fusarium equiseti* was not detected at Patancheru and Surat, whereas *F. sacchari* was present only at Patancheru and Palem. *Fusarium thapsinum* has been reported to be common in sorghum grain mold [2]. Other species identified in this study, such as *F. proliferatum*, *F. andiyazi* and *F. sacchari* have been reported to be associated with sorghum grain mold [17]. At some locations, *F. andiyazi* may be the dominant *Fusarium* species on sorghum. However, it has not been searched for extensively, but probably has a much broader distribution with sorghum [18].

Members of *Fusarium* species included in section Liseola are generally known to produce fumonisins. However, fumonisin production levels vary among the species. Strains of *F. proliferatum* were identified as highly toxigenic for fumonisin production followed by those of *F. andiyazi*. Fumonisins are produced by *F. proliferatum*, often at high levels [19, 20], and the *FUM* gene cluster encoding for fumonisin biosynthesis has been sequenced and characterized in some detail [21]. *Fusarium thapsinum*, although identified most commonly associated with sorghum grain mold complex in India, it was atoxigenic for fumonisin production except one isolate F 88. *Fusarium thapsinum* has been reported to produce high levels of moniliformin but only traces of fumonisins [2, 22, 23].

Genetic resistance to ear rot and mycotoxin content has been found in European maize inoculated with *F. graminearum* and *F. verticillioides* [24]. Maize genetics and breeding studies have guided strategies to improve resistance to fumonisin accumulation [6]. In this study, we have been able to identify toxigenic strains of *F. proliferatum* (F 242 or

F 234), *F. andiyazi* (F 943) and *F. thapsinum* (F 88) which can be used in greenhouse for evaluating sorghum lines for resistance to toxigenic *Fusarium* associated with grain mold complex. Such resistance sources could be used in breeding fumonisin-tolerant sorghum varieties and hybrids.

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References

1. Thakur RP, Rao VP, Navi SS, Garud TB, Agarkar GD, Bharati B. Sorghum grain mold: variability in fungal complex. *Int Sorg Mill Newsl.* 2003;4:104–8.
2. Leslie JF, Zeller KA, Lamprecht SC, Rheeder JP, Marasas WFO. Toxicity, pathogenicity, and genetic differentiation of five species of *Fusarium* from sorghum and millet. *Phytopathology.* 2005;95:275–83.
3. Vasanthi S, Bhat RV. Mycotoxins in food occurrence, health and economic significance and food control measures. *Indian J Med Res.* 1998;108:212–22.
4. Bhat RV, Shetty PH, Amruth RP, Sudershan RV. A foodborne disease outbreak due to the consumption of moldy sorghum and maize containing fumonisins mycotoxins. *J Toxicol-Clin Toxic.* 1997;35:249–55.
5. Bhat RV, Beedu SR, Ramakrishna Y, Munshi KL. Outbreak of trichothecene mycotoxicosis associated with consumption of mould damaged wheat products in Kashmir valley, India. *Lancet.* 1989;7:35–7.
6. Eller MS, Holland JB, Payne GA. Breeding for improved resistance to fumonisin contamination in maize. *Toxin Rev.* 2008;27:371–89.
7. Navi SS, Girish AG, Thakur RP, Yang XB. Banana leaves as a substitute for carnation leaves in characterizing *Fusarium* spp. *Phytopathology.* 2006;96:S83.
8. Kim DH, Martyn RD, Magill CW. Restriction fragment length polymorphism groups and physical map of mitochondrial DNA from *Fusarium oxysporum* f. sp. *niveum*. *Phytopathology.* 1992;82:346–53.
9. Vos P, Hogers R, Bleeker M, van de Lee T, Homes M, Frijters A, Pot J, Peleman J, Kuiper M, Zabeau M. AFLP: a new technique for DNA fingerprinting. *Nucleic Acids Res.* 1995;23:4407–14.
10. Rohlf FJ. NTSYS-pc: Numerical taxonomy and multivariate analysis system version 2.2. Exeter Software: Setauket, New York; 2000.
11. Excoffier L, Smouse P, Quattro L. Analysis of molecular variance inferred from metric distances among DNA haplotypes: application to human mitochondrial restriction data. *Genetics.* 1992;131:479–91.
12. O'Donnell K, Kistler HC, Cigelnik E, Ploetz RC. Multiple evolutionary origins of the fungus causing Panama disease of banana: concordant evidence from nuclear and mitochondrial gene genealogies. *Proc Natl Acad Sci.* 1998;95:2044–9.

13. Altschul SF, Madden TL, Schaffer AA, Zhang JH, Zhang Z, Miller W, Lipman DJ. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Res.* 1997;25:3389–402.
14. Reddy SV, Mayi DK, Reddy MU, Devi KT, Reddy DVR. Aflatoxins B₁ in different grades of chillies (*Capsicum annum* L.) in India as determined by indirect competitive-ELISA. *Food Addit Contam.* 2001;18:553–8.
15. Yu FY, Chu FS. Production and characterization of antibodies against Fumonisin B₁. *J Food Protect.* 1996;59:992–7.
16. Waliyar F, Reddy SV, Kumar PL. Review of immunological methods for the quantification of aflatoxins in peanut and other foods. *Peanut Sci.* 2009;36:54–9.
17. Thakur RP, Reddy BVS, Indira S, Rao VP, Navi SS, Yang XB, Ramesh S. Sorghum Grain Mold. Information Bulletin No. 72. International Crops Research Institute for the Semi-Arid Tropics, Patancheru 502324, Andhra Pradesh, India; 2006.
18. Leslie JF, Summerell BA. The *Fusarium* laboratory manual. 1st ed. Ames, USA: Blackwell Publishing Professional; 2006. p. 388.
19. Bacon CW, Nelson PE. Fumonisin production in corn by toxigenic strains of *Fusarium moniliforme* and *Fusarium proliferatum*. *J Food Protect.* 1994;57:514–21.
20. Dantzer WR, Pomettoi ALI, Murphy PA. Fumonisin B₁ production by *Fusarium proliferatum* strain M5991 in a modified Myro liquid medium. *Nat Toxins.* 1996;4:168–73.
21. Proctor RH, Plattner RD, Brown DW, Seo JA, Lee YW. Discontinuous distribution of fumonisin biosynthetic genes in the *Gibberella fujikuroi* species complex. *Mycol Res.* 2004;108:815–22.
22. Bryden WL, Logrieco A, Abbas HK, Porter JK, Vesonder RF, Richard JL, Cole RJ. Other significant *Fusarium* mycotoxins. In: Summerell BA, Leslie JF, Backhouse D, Bryden WL, Burgess LW, editors. *Fusarium*: Paul E. Nelson Memorial Symposium. APS Press, St. Paul, Minnesota; 2001. pp. 360–392.
23. Leslie JF, Marasas WFO, Shephard GS, Sydenham EW, Stockenstrom S, Thiel PG. Duckling toxicity and the production of fumonisin and moniliformin by isolates in the A and F mating populations of *Gibberella fujikuroi*. *Appl Environ Microb.* 1996;62:1182–7.
24. Miedaner T, Löffler M, Bolduan C, Kessel B, Ouzunova M, Mirdita V, Melchinger AE. Genetic variation for resistance, mycotoxin content of European maize inoculated with *Fusarium graminearum*, *F. verticillioides*. *Cereal Res Commun.* 2008;36:45–8.