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Identification of peptide sequences that potentially trigger HLA-A2.1-restricted cytotoxic T lymphocytes*

We used the human processing defective cell line 174CEM.T2 (T2) to identify potential cytotoxic T lymphocyte (CTL) epitopes of human proteins. Exogenously added peptides can increase the number of properly folded HLA-A2.1 molecules on the cell surface of T2 cells, as shown by immunofluorescence measurements using the mouse monoclonal antibody BB7.2 (anti-HLA-A2.1) and fluorescein isothiocyanate-labeled goat anti-mouse F(ab')₂ antibody. The peptides were selected on the basis of a computer score derived from the recently described HLA-A2.1 specific motif. Analysis of the influenza matrix protein showed that 15 out of 35 high-scoring peptides up-regulate the expression of HLA-A2.1 molecules on the T2 cell surface. The combination of the computer scoring program and an immunofluorescence-based peptide binding assay allows rapid detection of potential CTL target peptides.

1 Introduction

The MHC gene products associate with peptides and are recognized by cytotoxic T lymphocytes (CTL) [1]. Class I molecules are membrane heterodimers composed of a 45-kDa heavy (H) chain noncovalently associated with an invariant light chain, β_2 -microglobulin (β_2m). The H chain has three extracellular domains: $\alpha 1$, $\alpha 2$ and $\alpha 3$. The amino-terminal $\alpha 1$ and $\alpha 2$ domains interact with each other to form a peptide-binding groove, while the $\alpha 3$ domain adjacent to the plasma membrane directly contacts fern [2]. The H chains associate with β_2m shortly after synthesis in the endoplasmic reticulum (ER) [3]. Assembly of the H chain with β_2m and peptide is required for conformational stability and presentation at the cell surface of peptides [4-7]. Proteins present in the cytosol appear to be cleaved into fragments of 8-11 amino acids [8, 9]. The class I molecules are, on their way to the cell surface, loaded with peptide in or between the ER and the Golgi complex [10-13].

Recent studies have identified mouse and human cell lines, designated RMA-S [4], 721.174 [14] and 174CEM.T2 (T2) [15], which produce abundant cytoplasmic H chains and β_2m , but present low amounts of surface MHC molecules. These so-called processing-defective cell lines are unable to present endogenously synthesized peptides to CTL [4, 16]. The processing defect in the T2 cell line is due to a homozygous deletion of the MHC class II region located on chromosome 6 including the TAP1 and TAP2 (transporters associated with antigen processing) genes which encode the transporter proteins [17]. The HLA-A2.1 molecules are the

only HLA molecules present, at low density, on the cell surface of the T2 cell line and are at least partially occupied by peptides derived from the signal peptide domains of normal cellular proteins (IP-30, calreticulin) [18-20].

The 721.174 cells present exogenously added peptides which can bind to HLA-A2.1 at relatively high levels despite the relatively low levels of steady state expression of cell surface HLA-A2.1 molecules [21]. We used the T2 line, a cell line with the same processing defect as 721.174 cells, to identify peptide binding to HLA-A2.1 by measuring the HLA-A2.1 cell surface expression. The peptides used in our tests were selected using a computer scoring program derived from the HLA-A2.1 peptide motif ([22];●). We chose to study the influenza matrix protein because a previously described influenza matrix-specific HLA-A2.1-restricted CTL clone recognizes a defined influenza matrix epitope [23, 24] which we expected to detect in our test system.

2 Materials and methods

2.1 Scoring system

Sequencing of peptides eluted from purified HLA-A2.1 molecules revealed an HLA-A2.1 allele-specific peptide motif [20, 22]. It includes peptides with a length of nine amino acids with two anchor residues, a leucine, isoleucine or methionine at position 2 and a hydrophobic aliphatic residue at position 9, the C-terminal residue. The term anchor residue was used for the positions revealing a strong signal for only one amino acid or a few amino acids with closely related side chains. Furthermore, "strong and weak" amino acids are described in the different positions. We developed our own computer scoring program deduced from the HLA-A2.1 peptide motif. We scored six points for an anchor residue, four points for a strong and two points for a weak residue. The score for a given peptide is obtained

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by multiplication of the scores for each amino acid position. In this way the maximum score for a peptide of a length of nine amino acids is 36 864 points (6^2 points \times 4^3 points \times 2^4 points). Peptides lacking anchor residues at position 2 and at the C-terminal end were discarded. Peptides longer than nine amino acids can also bind to MHC molecules depending on the sequence [25]. Therefore, the computer program also scores for peptides with a second anchor residue at position 10 or 11.

2.2 Peptide binding study

Peptides were synthesized by solid-phase strategies on an automated multiple peptide synthesizer (Abimed AMS 422) using Fmoc chemistry. The purity of the peptides was determined by reverse-phase HPLC. Peptides were dissolved in DMSO (final DMSO concentration 0.25%) and diluted in 0.9% NaCl to a peptide concentration of 2 ng/ml and stored at -20°C . The T2 cell line, a gift from Dr. P. Cresswell (Dept. of Immunology, Yale University, New Haven, CT), was cultured in Iscove's modified Dulbecco's medium (IMDM) (Biochrom KG, Seromed Berlin, FRG) with 2 mM glutamine, 100 IU/ml penicillin, 100 $\mu\text{g}/\text{ml}$ kanamycin and 10% fetal calf serum (FCS, Hyclone Laboratories Inc. Logan, UT). The T2 cells were washed twice in serum-free culture medium. Cells (80 000) in 40 μl serum-free medium were put into a V-bottom 96-well plate together with 10 μl of the peptide dilution at different concentrations or 10 μl 0.9% NaCl, and incubated overnight at 37°C , 5% CO_2 in humidified air. The cells were washed once with cold (4°C) "PBA" (0.9% NaCl, 0.5% BSA, 0.02% NaN_3) and stained by indirect immunofluorescence. The first antibody BB7.2 (HLA-A2.1-specific monoclonal antibody) was added in a saturating amount and incubated for 30 min at 4°C . After two washing steps with cold PBA, FITC-labeled F(ab')₂ fragments of goat anti-mouse IgG (Tago Inc. Burlingame, CA) in a dilution of 1:40 were added as the second antibody and incubated for another 30 min at 4°C . The cells were washed once and fluorescence was measured at 488 nm on a FACScan flow cytometer (Becton Dickinson, Franklin Lakes, NJ).

The fluorescence ratio (FR) was calculated with the formula: mean fluorescence experimental sample / mean fluorescence background.

Human $\beta_2\text{m}$ (Sigma, St. Louis, MO) was dissolved in serum-free IMDM and stored at -20°C at a concentration of 1 mg/ml. Human $\beta_2\text{m}$ was used during the incubation period of the T2 cells with the peptide.

2.3 Competition cytotoxicity assay

The influenza matrix-specific HLA-A2.1-restricted CTL clone was a generous gift from Dr. H. Spits (DNAX, Palo Alto, CA). The CTL clone was grown on HLA-A2.1-positive EBV-transformed B cell lines irradiated with 30 Gy in RPMI (Gibco Paisley, Scotland) with 2 mM glutamine, 100 IU/ml penicillin, 100 $\mu\text{g}/\text{ml}$ kanamycin and 10% FCS (Hyclone Laboratories). In the competition cytotoxicity assay 2000 ^{51}Cr -labeled T2 cells (25 μl complete IMDM medium/well) were incubated with 5 μl of the influenza CTL epitope (0.5 $\mu\text{g}/\text{ml}$, final concentration) in the pre-

sence or absence of 5 μl of one of the competitor synthetic peptides (20 $\mu\text{g}/\text{ml}$, final concentration) in a U-bottom 96-well plate. After 20 min of incubation the influenza-specific CTL clone was added in a volume of 25 μl of complete IMDM medium at an effector to target ratio of 5:1 for an additional 4 h. The percentage ^{51}Cr release was calculated with the formula: [(cpm experimental well - background ^{51}Cr release) / (maximum ^{51}Cr release - background ^{51}Cr release)] \times 100.

The background was always less than 25% of maximal release. The percentage blocking was calculated with the formula: [1 - (% ^{51}Cr release of target cells incubated with epitope and competitor peptide / % ^{51}Cr release of target cells incubated with epitope)] \times 100.

3 Results

3.1 Scoring system

Fig. 1 illustrates the score distribution for peptides of the influenza matrix protein using a computer scoring program, the highest score was 1152 points. Peptides with anchor residues at position 2 and the C-terminal end were selected for synthesis (Tables 1 and 2). Eighteen peptides chosen randomly *i. e.* not according to the selection rules, were used as control peptides (Table 3). The influenza matrix epitope recognized by a HLA-A2.1-restricted CTL is one of the selected peptides with a high score (seq. no. 58-66; 576 points, Table 1). The computer scoring program revealed only 35 peptides (8% of the total number, Table 1) with a relatively high score of 144 points or higher, which makes it a useful first step in finding potential CTL epitopes. Peptides with anchor residues at position 2 and the C-terminal end with a score lower than 144 points are depicted in Table 2.

3.2 Peptide binding assay

Out of 35 high-scoring influenza matrix peptides, including the nonamer 58-66, 15 up-regulated the HLA-A2.1 mole-

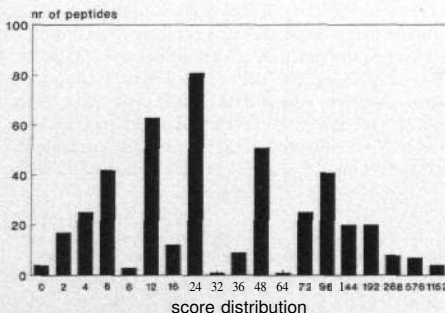


Figure 1. The score distribution for peptides of the influenza matrix protein using the computer scoring program. The maximum score for a peptide is 36 864 points (6^2 points \times 4^3 points \times 2^4 points). Peptides with an anchor residue at position 2 and the C-terminal end were used in our experiments together with the 18 control peptides.

Table 1. The Influenza matrix peptides with a high score^{a)}

Seq. nr.	score	peptides	F.R.	% blocking
14	1152	I I P S G P L K A	1.00	12
14	1152	I I P S G P L K A E I	0.95	
41	1152	V L M E W L K T R P I	2.79 **	67
3	576	L L T E V E T Y V V	2.69 *	86
3	576	L L T E V E T Y V V L	3.11 **	50
51	576	I L S P L T K G I	2.73 *	80
51	576	I L S P L T K G I L	1.56 *	63
54	576	P L T K G I L G F V	0.98	13
58	576	G I L G F V F T L	2.92 **	0
58	576	G I L G F V F T L T V	2.14 **	
98	576	K L Y R K L K R E I	0.96	8
106	576	E I T F F H G A K E I	1.05	
2	288	S L L T E V E T Y V V	2.38 **	80
2	288	S L L T E V E T Y V V L	3.51 **	63
50	288	P I L S P L T K G I	1.66 *	
50	288	P I L S P L T K G I L	0.98	
134	288	R M G A V T T E V	2.12 **	64
134	288	R M G A V T T E V A	0.93	
218	288	R I G T H P S S S A	1.00	
11	144	V L D I I P S G P L	1.20	11
19	144	P L K A E I A Q R L	1.06	10
23	144	E I A Q R L E D V	1.02	
23	144	E I A Q R L E D V F A	1.01	
59	144	I L G F V F T L T V V	1.87 *	2
102	144	K L K R E I T F F H G A	0.97	0
130	144	L I Y N R M G A V	1.24	
145	144	G L V C A T C E Q I A	1.83 **	56
145	144	G L V C A T C E Q I	1.04	18
164	144	Q M V T T T N P L	3.31 **	
164	144	Q M V T T T N P L I	1.98 *	
172	144	L I R H E N R M V	0.85	
172	144	L I R H E N R M V L	0.90	
172	144	L I R H E N R M V L A	1.08	
191	144	Q M A G S S E Q A	0.98	9
228	144	G L K N D L E N L	1.03	

a) The sequence number (Seq. nr.) of the first amino acid is shown. The peptides are ranked according to their computer score. The fluorescence ratio (final peptide concentration in the binding assay was 100 µg/ml). % blocking of lysis using an E/T of 5:1. T2 was the target cell line. The asterisk indicates the binding peptides. For conditions of blocking see Sect. 2.3.

Table 2. The Influenza matrix peptides with a low score^{a)}

Seq. nr.	score	peptides	F.R.
13	72	D I I P S G P L K A	1.06
42	72	L M E W L K T R P I	1.05
42	72	L M E W L K T R P I L	1.04
45	72	W I K T R P I L S P L	1.08
114	72	E I S L S Y S A G A	1.05
114	72	E I S L S Y S A G A L	1.04
116	72	S L S Y S A G A L	1.06
116	72	S L S Y S A G A L A	1.05
123	72	A L A S C M G L I	1.26
127	72	C H G L I Y N R M	1.14
127	72	C M G L I Y N R M G A	1.17
129	72	G L I Y N R H G A	1.03
129	72	G L I Y K R H G A V	1.02
171	72	P L I R H E N R M	1.05
171	72	P L I R H E N R M V	0.95
171	72	P L I R H E N R M V L	0.96
236	72	N L Q A Y Q K R H	1.01
236	72	N L Q A Y Q K S H G V	0.98
38	36	D L E V L M E W L	1.11
65	36	T L T V P S E R G L	1.08
73	36	G L Q R R R F V Q N A	1.05
178	36	R M V L A S T A K A	1.05
211	36	Q M V Q A M I R I	1.11
180	24	V L A S T A K A M	1.08

a) See footnote to Table 1.

cule cell surface expression on the T2 cell line, while none of the low-scoring or control peptides did so (cut off > 1.5) (Tables 1-3 column FR). The FACScan results shown in Tables 1-3 were highly reproducible. The FACScan procedure was an effective second step in finding potential CTL epitopes.

Table 3. The Influenza matrix peptides used as a control group^{a),b)}

Seq. nr.	peptides	F.R.	% blocking
20	L K A E I A Q R L	0.98	2
31	V F A G K N T D L	1.00	37
41	V L M E W L K T R	1.07	45
47	K T R P I L S P L	1.05	20
55	L T K G I L G F V	1.00	12
60	L G F V F T L T V	0.95	22
75	Q R R R R F V Q N A	0.99	1
88	G D P N N M D K A	0.97	15
91	N N M D K A V K L	1.05	36
95	K A V K L Y R K L	1.07	31
98	K L Y R K L K R E	0.95	4
99	L Y R K L K R E I	0.99	8
107	I T F H G A K E I	0.96	10
173	I R H E N R M V L	0.92	18
179	M V L A S T A K A	1.09	20
194	G S S E Q A A E A	0.97	2
201	E A M E V A S Q A	0.98	1-
232	D L L E N L Q A V	0.99	5

b) The peptides are ranked according to their sequence number.

To determine if peptides which up-regulate the HLA-A2.1 molecule actually bind to the HLA-A2.1 molecule we tested 11 binding peptides and 26 non-binding peptides for competition in a cytotoxicity assay. We expected those which up-regulate the HLA-A2.1 molecule on the T2 cell surface to block lysis of an HLA-A2.1-restricted CTL clone specific for the influenza matrix protein. The results in the last column of Tables 1 and 3 show that peptides which up-regulate HLA-A2.1 block the lysis of the CTL clone, except the peptides 58-66 and 59-68 which are recognized by the CTL clone (cut off $\geq 50\%$ blocking).

3.3 Effect of human β_2m

The detection threshold of HLA-A2.1 up-regulation lies at a concentration of 25 to 50 $\mu\text{g/ml}$ peptide without β_2m (Fig. 2). The detection level of HLA-A2.1 up-regulation is at a concentration of 6.25 to 12.5 $\mu\text{g/ml}$ peptide with β_2m (Fig. 2). The addition of free human β_2m increases the overall HLA-A2.1 cell surface expression compared to tests in which no β_2m was added (Fig. 2). The sensitivity of the peptide binding assay is not further enhanced by the addition of higher concentrations ($> 20 \mu\text{g/ml}$) of β_2m (data not shown). We conclude that human β_2m increases the sensitivity of our peptide binding assay without affecting background binding. The optimal final concentration of human β_2m is 15 $\mu\text{g/ml}$.

4 Discussion

Peptides which bind to HLA-A2.1, if immunoreactive, might be suitable for use as vaccine components to prevent the outgrowth of tumors or pathogens which express those peptides in the context of HLA-A2.1. A number of recent studies in mice have indicated the feasibility of this approach [26-28]. Preventive peptide vaccination using a T cell epitope of the lymphocytic choriomeningitis virus (LCMV)-NP can reduce the virus titer when mice are infected with LCMV [26]. Immunization of mice with a virus-specific CTL epitope derived from the Sendai virus protects them from a lethal infection [27]. Immunization with synthetic peptides derived from a tumor-inducing virus reduces the growth of these tumors [28].

The therapeutic value of CTL has been reviewed [29, 30]. Binding peptides might be used to induce a primary CTL response using the T2 cell line loaded with a binding peptide as an antigen-presenting cell. A similar approach has been reported for the mouse system [31]. One of the problems might be that *in vitro* primed CTL stimulated with peptides might not always recognize endogenously processed antigens, in contrast to their recognition of cells sensitized with the target antigen [32]. Harty and Bevan overcame this problem by *in vitro* restimulation of spleen cells of mice immunized with a synthetic peptide with cells transfected with the gene encoding the epitope [33]. Using this method, high-affinity CTL clones were generated, instead of CTL clones which only recognize peptide-loaded target cells.

Human *fam* is required for the post-translational processing and transport of all HLA-A, -B and C gene products to the cell surface [34, 35]. The effect of *fcm* in our peptide

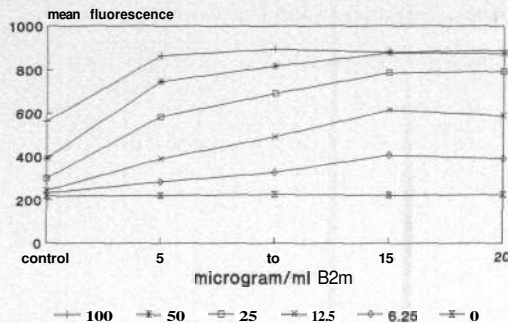


Figure 2. Effect of β_2m on the up-regulation of HLA-A2.1 molecules on the T2 cell surface. β_2m was used at four different concentrations (5-10-15-20 $\mu\text{g/ml}$). No β_2m was used in the control test. The peptide concentrations were 100, 50, 25, 12.5, 6.25, 0 $\mu\text{g/ml}$ (several binding peptides were used).

binding assay can be explained in different ways. First, the addition of human β_2m might increase the number of peptide binding sites on the cell surface, for example by rescue of free heavy chains from dissociation. In that case the HLA molecules remain available for peptide acquisition. Indeed, Boyd et al. showed that the number of high affinity binding sites increased when β_2m was used together with purified soluble analogues of MHC molecules [36]. Second, β_2m may selectively bind to the heavy chain which is already stabilized by a binding peptide [37]. Finally, Otten et al. reported that when the fern-heavy chain complex begins to dissociate it would become more accessible to peptide binding [38]. The number of HLA-A2.1 molecules on the cell surface of the T2 cell line does not increase when β_2m is used (maximum β_2m concentration 20 $\mu\text{g/ml}$) without a peptide or a non-binding peptide (see Fig. 2 at peptide concentration of 0 $\mu\text{g/ml}$).

One may expect that further refinements of the HLA-A2.1 peptide motif will be possible in the near future. The peptide motif does not take into account that flanking amino acids of the presented peptide might influence the processing and presentation of peptides [39]. Therefore, not all binding peptides are likely to be processed *in vivo*. On the other hand most naturally processed self peptides and viral epitopes conform to the motif.

Peptide sequences that potentially trigger HLA-A2.1-restricted CTL can be easily detected using our test system. Our computer scoring program in combination with the peptide binding assay can be used for other HLA molecules. For that purpose one requires a peptide motif, T2 cells transfected with the specific HLA-allele and a monoclonal antibody specific for that HLA molecule. The HLA-A2.1 molecule is present in 40% of the Western European Caucasoid population. Thus, in principle those individuals could benefit from treatment with tumor-specific CTL or tumor-associated peptides.

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5 References

- 1 Townsend, A. and Bodmer, H., *Annu. Rev. Immunol.* 1989, 7: 601.
- 2 Bjorkman, P. J., Saper, M. A., Samraoui, B., Bennet, W. S., Strominger, J. L. and Wiley, D. C., *Nature* 1987, 329: 506.
- 3 Dobberstein, B., Garoff, H., Warren, G. and Robinson, P. J., *Cell* 1979, 17: 759.
- 4 Townsend, A., Ohlen, C., Bastin, J., Ljunggren, H. G., Foster, L. and Karre, K., *Nature* 1989, 340: 443.
- 5 Arce-Gomez, B., Jones, E. A., Barnstable, C. J., Solomon, E. and Bodmer, W. E., *Tissue Antigens* 1978, 11: 96.
- 6 Ploegh, H. L., Orr, H. T. and Strominger, J. L., *Cell* 1981, 24: 287.
- 7 Seong, R. H., Clayberger, C. A., Krensky, A. M. and Parnes, J. R., *J. Exp. Med.* 1988, 167: 288.
- 8 Röttschke, O., Falk, K., Deres, K., Schild, H., Norda, M., Metzger, J., Jung, G. and Rammensee, H., *Nature* 1990, 345: 252.
- 9 Van Bleek, G. M. and Nathenson, S. G., *Proc. Natl. Acad. Sci. USA* 1991, 88: 11032.
- 10 Nuchtern, J. G., Bonifacio, J. S., Biddison, W. E. and Klausner, R. D., *Nature* 1989, 339: 223.
- 11 Yewdell, J. W. and Bennink, J. R., *Science* 1989, 244: 1072.
- 12 Yewdell, J. W. and Bennink, J. R., *Cell* 1990, 62: 203.
- 13 Kvist, S. and Hamann, U., *Nature* 1990, 348: 446.
- 14 DeMars, R., Chang, C. C., Shaw, S., Reitnauer, P. J. and Sondel, P. M., *Hum. Immunol.* 1984, 11: 77.
- 15 Salter, R. D. and Cresswell, P., *EMBO J.* 1986, 5: 943.
- 16 Hosken, N. A. and Bevan, M. J., *Science* 1990, 245: 367.
- 17 DeMars, R. and Spies, T., *Trends in Cell Biol.* 1992, 2: 81.
- 18 Henderson, R. A., Michel, H., Sakaguchi, K., Shabanowitz, J., Appella, E., Hunt, D. F. and Engelhard, V. H., *Science* 1992, 255: 1264.
- 19 Wei, M. L. and Cresswell, P., *Nature* 1992, 356: 443.
- 20 Hunt, D. F., Henderson, R. A., Shabanowitz, J., Sakaguchi, K., Michel, H., Sevilir, N., Cox, A. L., Appella, E. and Engelhard, V. H., *Science* 1992, 255: 1261.
- 21 Cerundolo, B., Alexander, J., Anderson, K., Lamb, C., Cresswell, P., McMichael, A., Gotch, F. and Townsend, A., *Nature* 1990, 345: 449.
- 22 Falk, K., Röttschke, O., Stevanovic, S., Jung, G. and Rammensee, H. G., *Nature* 1991, 351: 290.
- 23 Winter, G. and Fields, S., *Nucleic Acids Res.* 1980, 8: 1965.
- 24 Morrison, J., Elvin, J., Latron, F., Gotch, E., Moots, R., Strominger, J. L. and McMichael, A., *Eur. J. Immunol.* 1992, 22: 903.
- 25 Deres, K., Schumacher, T. N. M., Wiesmüller, K. H., Stevanovic, S., Greiner, G., Jung, G. and Ploegh, H. L., *Eur. J. Immunol.* 1992, 22: 1603.
- 26 Schulz, M., Zinkernagel, R. M. and Hengartner, H., *Proc. Natl. Acad. Sci. USA* 1991, 88: 991.
- 27 Kast, W. M., Roux, L., Curren, J., Blom, H. J. J., Voordouw, A. C. M., Kolakofsky, D. and Melief, C. J. M., *Proc. Natl. Acad. Sci. USA* 1991, 88: 2283.
- 28 Reinholdsson-Ljunggren, G., Ramqvist, T., Åhrlund-Richter, L. and Dalians, T., *Int. J. Cancer* 1992, 50: 142.
- 29 Kast, W. M. and Melief, C. J. M., *Immunol. Lett.* 1991, 30: 292.
- 30 Melief, C. J. M., *Adv. Cancer Res.* 1992, 58: 143.
- 31 De Bruijn, M. L. H., Schumacher, T. N. M., Nieland, J. D., Ploegh, H. L., Kast, W. M. and Melief, C. J. M., *Eur. J. Immunol.* 1991, 21: 2963.
- 32 Schild, H., Norda, M., Deres, K., Falk, K., Röttschke, O., Wiesmüller, K. H., Jung, G. and Rammensee, H. G., *J. Exp. Med.* 1991, 174: 1665.
- 33 Harty, J. T. and Bevan, M. J., *J. Exp. Med.* 1992, 175: 1531.
- 34 Neefjes, J. J. and Ploegh, H. L., *Eur. J. Immunol.* 1988, 18: 801.
- 35 Hochman, J. H., Shimizu, Y., DeMars, R. and Edidin, M., *J. Immunol.* 1988, 140: 2322.
- 36 Boyd, L. E., Kozlowski, S. and Margulies, D. H., *Proc. Natl. Acad. Sci. USA* 1992, 89: 2242.
- 37 Townsend, A., Elliot, T., Cerundolo, V., Foster, L., Barber, B. and Tse, A., *Cell* 1990, 62: 285.
- 38 Otten, G. R., Bikoff, E., Ribaldo, R. K., Koziowski, S., Margulies, D. H. and Germain, R. N., *J. Immunol.* 1992, 148: 3723.
- 39 Del Val, M., Schlicht, H. J., Ruppert, T., Reddehase, J. and Koszinowski, U. H., *Celt* 1991, 66: 1145.