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Identification of the amino-acetonitrile derivative monepantel (AAD 1566) as a new anthelmintic drug development candidate

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Abstract Anthelmintic resistance has become a global phenomenon in gastro-intestinal nematodes of farm animals, including multi-drug resistance against the three major classes of anthelmintics. There is an urgent need for an anthelmintic with a new mode of action. The recently discovered amino-acetonitrile derivatives (AADs) offer a new class of synthetic chemicals with anthelmintic activity. The evaluation of AADs was pursued applying in vitro assays and efficacy and tolerability studies in rodents, sheep, and cattle. Amongst various suitable compounds, AAD 1566 eliminated many tested pathogenic nematode species, both at larval and adult stages, at a dose of 2.5 mg/kg bodyweight in sheep and 5.0 mg/kg bodyweight in cattle. The same doses were sufficient to cure animals infected with resistant or multi-drug-resistant nematode isolates. These findings, complemented by the good tolerability and low toxicity to mammals, suggest that AAD 1566, monepantel, would be a suitable anthelmintic drug development candidate.

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Introduction

Today, chemotherapy of nematode infections in livestock mainly relies on three classes of broad-spectrum anthelmintics, the benzimidazoles (BZs), imidothiazoles (LEV), and macrocyclic lactones (MLs). From these, the latest class introduced to the market were the MLs in 1981 (Chabala et al. 1980). The frequent use of these anthelmintics over many years has inevitably led to the development of drug resistance to each class in parasitic nematodes. While reports of anthelmintic resistance in cattle nematodes started to accumulate in recent years (Coles et al. 2001, Familton et al. 2001, Mejia et al. 2003, Anziani et al. 2004, Gasbarre and Smith 2004, Waghorn et al. 2006a), resistance in sheep nematodes is more established and continues to increase in prevalence and severity in most sheep-farming regions of the world (Coles et al. 1991, Echevarria et al. 1996, van Wyk et al. 1997, Coles 1998, Jackson and Coop 2000, Love et al. 2003, Kaplan 2004, Waghorn et al. 2006b, Traversa et al. 2007). The appearance of multi-drug-resistant nematode isolates is threatening successful chemotherapy of infected sheep in many countries (Yue et al. 2003, Kaminsky 2003, Besier and Love 2003, Wrigley et al. 2006). This has serious consequences and in some cases has even resulted in flock closure (Sargison et al. 2005, Blake and Coles 2007). Integrated parasite management approaches have recently been considered important in parasite control (Waller 1999) but so far have not shown sufficient effect without using suitable supportive anthelmintic therapy (Besier 2007). However, no new anthelmintic class has been introduced to the market within the last 28 years, with the exception of the cyclodepsipeptides, which to date are only licensed for use in cats to control nematodes. Besier (2007) describes the situation for sheep farmers as "sitting on a hypothetical nematode-control time bomb with a shortening fuse". Clearly, there is an urgent need for new anthelmintics for livestock, which are able to overcome resistance to the currently available drugs. Most recently, the amino-acetonitrile derivatives (AADs) were discovered to express anthelmintic activity (Kaminsky et al. 2008). AADs are a class of low molecular mass compounds bearing different aryloxy and aroyl moieties on an amino-acetonitrile core (Ducray et al. 2008). They offer a class of novel synthetic compounds with high activity against all gastro-intestinal nematodes, including resistant isolates (Kaminsky et al. 2008), an essential prerequisite for any potential new anthelmintic drug. To identify a suitable drug development candidate amongst the whole class of AADs, in vitro assays and efficacy and tolerability studies in rodents, sheep, and cattle were pursued.

Materials and methods

Nematode isolates The origin, history, and drug sensitivity status of nematode isolates used are summarized in Table 1. Most of the susceptible isolates were continuously passaged in sheep or cattle for many years at the Novartis Centre de Recherche Santé Animale (Switzerland). In addition, four drug-resistant isolates were used. The *Haemonchus contortus* Howick isolate (van Wyk et al. 1997) was received from Dr. J. van Wyk, Onderstepoort Veterinary Institute. The Howick isolate is multi-drug-resistant to BZs, MLs, and LEV (van Wyk et al. 1997, Ghisi et al. 2007). The Australian *H. contortus* Mox/Fen isolate was kindly provided by G. Hutchinson, New South Wales Department of Primary Industries. This isolate is resistant to at least the MLs and BZs (Love et al. 2003). The *H. contortus* Courtion and *Trichostrongylus colubriformis* Villarey were both isolated in

Table 1 Origin and drug sensitivity status of nematode isolates in sheep

2004 from farms near the Novartis Centre de Recherche Santé Animale in the Canton Fribourg, Switzerland. *H. contortus* Courtion is resistant to BZs, and *T. colubriformis* Villarey is resistant to BZs and LEV (Kaminsky et al. 2008).

In vitro assays The in vitro larval development assay was performed as described by Gill et al. (1995), with minor modifications. Briefly, freshly harvested and cleaned nematode eggs were used to seed a suitably formatted 96-well plate containing the test substances to be evaluated for anthelmintic activity. Each compound was tested by serial dilution in order to determine the effective concentration (EC₁₀₀), which eliminates all nematodes. The test compounds were embedded in an agar-based nutritive medium, allowing the full development of eggs through to third stage larvae (L3). The plates were incubated for 6 days at 28°C and 80% relative humidity. Nematodicidal activity was expressed as percent reduced development of L3 stages. The AADs were tested against drug susceptible and BZ-resistant *H. contortus*.

Efficacy testing in Meriones unguiculatus Evaluation in gerbils was performed according to Conder et al. (1990, 1991), with minor modifications. Gerbils were artificially infected by gavage with ca. 2,500 sheathed L3 larvae each of *T. colubriformis* and *H. contortus*, 6 and 5 days before treatment, respectively. Treatment with AAD compounds was performed orally by gavage or subcutaneously by injection. Control animals received a placebo formulation. Three days after treatment, gerbils were euthanized and dissected to recover *H. contortus* and *T. colubriformis*. After incubation of the organs for 2 h at 37°C, worms (L4 stage) were collected in water, stained with iodine, and counted using a stereomicroscope. Efficacy was expressed as a percentage reduction in worm numbers in comparison

Species	Origin	Isolated in or maintained since	Drug resistance to
Haemonchus contortus susc	South Africa	1984	susc
H. contortus Howick	South Africa, Howick	1997	BZs, LEV, MLs
H. contortus Courtion	Switzerland, Courtion	2004	BZs
H. contortus Mox/Fen	Australia	2003	BZs, MLs
Trichostrongylus colubriformis susc	UK, Weybridge	1989	susc
T. colubriformis Villarey	Switzerland, Villarey	2004	BZs, LEV
T. axei	UK, Weybridge	1989	susc
Teladorsagia circumcincta	UK, Glasgow	1988	susc
Cooperia curticei	UK, Weybridge	1991	susc
Nematodirus spathiger	UK, Weybridge	1989	susc
Chabertia ovina	Germany, Hannover	1993	susc
Ostertagia ostertagi	UK, Glasgow	1999	susc
C. oncophora	UK, Glasgow	1999	susc

BZs Benzimidazoles, LEV imidazothiozoles (levamisole), MLs macrocyclic lactones, susc susceptible

with the geometric mean of worms collected from the control group using Abbott's formula.

Efficacy studies in sheep and cattle Sheep and cattle efficacy studies were performed according to the guidelines for evaluating the efficacy of anthelmintics in ruminants (bovine, ovine, caprine) by the World Association for the Advancement of Veterinary Parasitology (Wood et al. 1995).

Ames test The study was performed as plate incorporation test and as pre-incubation test according to the OECD (1997a) guideline for the testing of chemicals 471 using the *Salmonella typhimurium* strains TA 1535, TA 97A, TA 98, TA 100, and TA 102. Both independent tests were performed with and without liver microsomal activation (S9 mix). Each AAD concentration, including the positive and negative controls, was tested in triplicate.

Chromosome aberration test The study was performed according to OECD (1997c) guideline for the testing of chemicals 473 in order to evaluate any clastogenic potential of a compound in vitro. This was done by determining the effect of the test item on the frequency of chromosomal aberrations in cultured human peripheral blood lymphocytes in the presence and absence of rat liver S9.

Micronucleus test The test was performed according to OECD (1997b) guideline for the testing of chemicals 474. Administration of the compound was via gavage. Two repeated doses of 2,000 mg/kg bodyweight each, 24 h apart, were administered to five male and five female mice per group. Mice were killed 24 h after the second dose. Bone marrow was collected from the femoral bone. The ratio of polychromatic/normochromatic erythrocytes (PCE/NCE) was determined by counting at least 1,000 erythrocytes, and the micronuclei were counted in at least 2,000 polychromatic erythrocytes.

Tolerability tests Six sheep of the brown Alp breed weighing between 30–50 kg were used. Sheep were treated on day 0 according to individual bodyweights assessed 5 days before compound administration. The AADs 85 and 96 were administered orally by syringe to the back of the throat of two sheep each at a dose of 50 mg racemate/kg. A placebo formulation was administered at a similar volume to the two sheep in the control group. After treatment, the animals were observed at least once a day for the remainder of the study regarding their general health and behavior. In addition, bodyweights, food consumption, clinical chemistry, hematology, heart rate, and body temperature were evaluated up to day 14 after dosing.

A second study was undertaken to examine the tolerability of the active enantiomer AAD 1566. Six Suffolk female sheep (aged 11 months) were allocated to three groups of two sheep each, one of which served as an untreated control. On day 0, two sheep were treated with AAD 1566 at 50 mg/kg, which was two times the dose used in the previous study. Seven days later, two other sheep were treated with the same formulation at 75 mg/kg. Following their successful treatment, the sheep treated with 50 mg/kg were re-treated at 100 mg/kg on day 14. Similarly, the sheep treated previously with 75 mg/kg were re-treated on day 21 at 125 mg/kg. The following measurements were taken throughout the study: visual observations including veterinary examinations, daily food intake, bodyweights, clinical blood chemistry and hematology, and body temperature.

Results

The AADs evaluated herein were derived from an optimization program (Ducray et al. 2008) and showed in vitro EC_{100} values between 0.01 and 0.04 µg/mL (individual data not shown). The in vitro susceptibility was similar for *H. contortus* and *T. colubriformis* and was independent of the sensitivity status of the nematode isolate.

When tested in gerbils, all AADs evaluated herein had an efficacy of 85-100% at an oral dose of 1 mg/kg against H. contortus susc and T. colubriformis susc (Table 2). Differences occurred at a dose of 0.32 mg/kg, when most AADs were not active against T. colubriformis susc (except AAD 96). The efficacy of AADs after subcutaneous application of 1 mg/kg was only slightly lower if not similar to that observed after oral application (Table 2). After separation of the racemic mixture of AAD 96, the two resulting enantiomers were tested against drug-resistant nematode isolates (Table 3). The racemic mixture AAD 96 showed 84-100% efficacy at 1 mg/kg when applied orally or subcutaneously. At the same doses, the enantiomer AAD 1566 showed a higher activity (99-100%) while the other enantiomer AAD 96i did not show any activity (Table 3). Furthermore, the AAD 1566 and the enantiomer AAD 85a, the active enantiomer of AAD 85, were both effective against the multi-drug-resistant H. contortus Howick isolate in gerbils (Table 4). While a dose of 10 mg/kg albendazole, 0.2 mg/kg ivermectin, or 10 mg/kg levamisole cured all gerbils infected with H. contortus susc, none of the gerbils infected with the H. contortus Howick was cured. On the contrary, the activity of AAD 85a and AAD 1566 at an oral dose of 0.5 mg/kg was similar against both the Howick isolate and the susceptible isolate (Table 4).

All AADs tested were active against the larval (L4) stage of most nematode species in sheep at an oral dose of 5 mg active enantiomer/kg (Table 5). AAD 85 and AAD 96 were 92–100% effective against all species tested

		Efficacy in %					
		Per oral Subcutaneous					aneous
		1 m	g/kg	0.32 1	ng/kg	1 mg/kg	
Structure	AAD	Нс	Тс	Hc	Тс	Нс	Тс
NC H CF3 O CF3	79	95	98	nd	nd	91	97
$C_{F_3}^{CN} \rightarrow C_{F_3}^{H} \rightarrow C_{F_3}^{N}$	85	95	85	70	0	97	75
NC CF3 NC CF3	93	97	91	0	0	72	0
NC H S CF3	94	90	98	23	0	92	86
	95	97	87	62	0	90	72
CN H CF_3 NC O CF_3	96	100	100	92	68	98	86

Table 2 Efficacy of AADs in gerbils (n=2) against the susceptible isolates H. contortus susc (South Africa; Hc) and T. colubriformis susc (UK; Tc)

The doses are for the racemic mixture of each AAD containing equal amounts of active and inactive enantiomers. *nd* Not done

at a dose of 2.5 mg active enantiomer/kg. At lower doses, the efficacy decreased for Cooperia curticei and Nematodirus spathiger to below 90%. The efficacy of AAD 1566, the active enantiomer of the racemic mixture AAD 96, against all species tested (L4 stages) was above 96% at an oral dose of 2.5 mg/kg (Table 5). The same dose was sufficient to cure all sheep infected with adult stages of either the multi-drug-resistant H. contortus Howick or the H. contortus Australian Mox/Fen isolate (Table 6). These isolates were not eliminated from sheep when treated with 0.2 mg/kg ivermectin or moxidectin, respectively, and neither were eliminated with a combination of ivermectin plus albendazole plus levamisole nor with a combination of moxidectin plus fenbendazole plus levamisole at the recommended doses, respectively. AAD 96 cured infected sheep with each of the two isolates at a dose down to 1.25 mg active enantiomer/kg (Table 6).

AAD 96 was also tested against adult stages of various nematode species. Sheep infected with *H. contortus*,

Teladorsagia circumcincta, T. axei, T. colubriformis, C. curticei, N. spathiger, and Chabertia ovina were treated 29 days after infection, by which time the infections had matured so that all sheep had high nematode eggs per gram feces (epg). Three sheep each were treated with 5.0, 2.5, and 1.25 mg racemate/kg corresponding to 2.5, 1.25, and 0.63 mg active enantiomer/kg bodyweight. In all treated sheep, the epg decreased substantially within the first 2 days (Fig. 1). The epg of sheep treated with 2.5 and 1.25 mg active enantiomer/kg remained negative until day 20 posttreatment. Two sheep treated with 0.63 mg active enantiomer/kg remained negative, while one sheep, the epg became positive at very low level (maximum of 100 epg) compared to the untreated control sheep (Fig. 1). Efficacy based on worm counts after necropsy were 99-100% for all species in sheep treated with 2.5 mg active enantiomer/kg and 96-100% for most species treated with 1.25 mg/kg. In the latter group, which had a negative epg, the efficacy for N. spathiger was 87% and 79% for C. ovina.

Table 3 Efficacy of the racemic mixture AAD 96 and the two enantiomers, AAD 1566 and AAD 96i, against the drug-resistant isolates *H. contortus* Courtion (HcC) and *T. colubriformis* Villarey (TcV) in gerbils (n=2)

	Efficacy (%)						
		Per	Subcutaneous				
	1 m	g/kg	0.32 r	ng/kg	1 mg/kg		
Compound	HcC	TcV	HcC	TcV	HcC	TcV	
AAD 96	100	95	76	17	100	84	
AAD 1566 $\downarrow^{CN}_{CF_3} \circ \uparrow^{(S)}_{NC} \rightarrow \downarrow^{S} \circ_{CF_3}$	100	100	100	88	100	99	
AAD 96i $\downarrow^{CN}_{CF_3} \circ \uparrow^{(P)}_{NC} \downarrow^{S} \circ^{CF_3}_{CF_3}$	0	0	6	0	15	0	

Four AADs were tested for efficacy in cattle. AAD 96 cured cattle infected with larval stages of *Ostertagia ostertagi* and *C. oncophora* at an oral dose of 7.5 mg active enantiomer/kg and had an efficacy of 98–100% at a dose of 3.8 mg/kg. AAD 1566 eliminated all adult *O. ostertagi* and *C. oncophora* when applied topically at a dose of 5 mg/kg (Table 7).

The most efficacious AADs were tested for mutagenic, clastogenic, or aneugenic potential using the Ames test, the in vitro chromosome aberration test with human cells, and the mouse micronucleus test.

Ames test AAD 1566 precipitated on the test plates at 5,000 µg/plate (plate incorporation) or at \geq 200 µg/plate (pre-incubation). No evidence of bacteriotoxicity was

 Table 4
 Efficacy of the active enantiomers AAD 85a and AAD 1566

 after oral application against multi-drug-resistant *H. contortus* (Howick) in gerbils

Dose (mg/kg)	Efficacy (%) against <i>H. contortus</i> susc	Efficacy (%) against <i>H. contortus</i> Howick
10	100	39
0.2	100	35
10	100	51
0.5	97	97
0.5	100	100
	Dose (mg/kg) 10 0.2 10 0.5 0.5	Dose (mg/kg) Efficacy (%) against H. contortus susc 10 100 0.2 100 10 100 0.5 97 0.5 100

Each dose was tested with six to ten *Meriones unguiculatus* per group, except the ivermectin group with 0.2 mg/kg (n=4)

detected up to the highest concentrations tested. Treatment with AAD 1566 did not increase the revertant numbers of any of the bacterial tester strains used. Thus, AAD 1566 did not show evidence of a mutagenic potential under the experimental conditions used in this bacterial reverse mutation assay. Similar results were obtained in screening tests with AADs 96, 85, and 95 (data not shown).

Chromosome aberration test AAD 1566 produced a concentration-dependent decrease in the mitotic index. Based on the mitotic index data, the following concentrations of AAD 1566 were selected for analysis in the chromosomal aberration assay: without metabolic activation, 25.8, 33.4, 43.1 µg/mL (20-h continuous treatment) and 57.2, 81.8, 97.8 µg/mL (3-h treatment, 17-h recovery); with metabolic activation, 77.5, 100.1, 129.3 µg/mL (3-h treatment, 17-h recovery) and 81.8, 117.0, 139.9 µg/mL (3-h treatment, 17-h recovery). In the absence, as well as in the presence of metabolic activation, the aberration frequencies of cultures treated with AAD 1566 were all within the historical control range. No increase in the frequencies of polyploid cells was observed in the absence or presence of rat liver S9. In conclusion, AAD 1566 did not show any clastogenic potential under the test conditions used in the chromosomal aberration test with human peripheral blood lymphocytes.

Micronucleus test There were no clinical signs and no mortality of mice for AAD 1566. No cytotoxicity was

AAD	Dose (mg/kg) of the active enantiomer	Efficacy (%)						
		Нс	T circ	Та	Tc	Cc	Ns	Со
79	5.00	100	100	96	100	75	100	100
85	5.00	100	100	98.5	99.5	92	99.5	100
	2.50	100	100	99	100	96	99	100
	1.25	100	98	79	100	88	33	90
93	5.00	76	68	73	100	65	50	100
94	5.00	100	100	99.5	100	95.5	100	99
	2.50	100	100	99	100	97	90	97
95	5.00	100	100	99	100	97.5	100	100
	2.50	100	100	97	100	97	76	100
96	5.00	100	100	100	100	96	100	100
	2.50	100	100	100	100	92	100	100
	1.25	100	100	96	100	80	46	95
1566	3.75	100	100	99	100	98	99	100
	2.50	100	100	99	100	98	96	100

Table 5 Efficacy of AADs against larval (L4)stages of various nematode species in sheep after oral application

All AADs, except the AAD 1566, were tested as racemic mixtures containing half of the active enantiomer; doses in the table have been calculated for the active enantiomer. AAD 1566 was tested as the purified active enantiomer. Each dose was tested in two to five infected sheep. *Hc H. contortus, T circ Teladorsagia circumcincta, Ta T. axei, Tc T. colubriformis, Cc C. curticei, Ns N. spathiger, Co C. ovina*

evidenced by the PCE/NCE ratio. There was no biologically relevant increase of the number of micronucleated PCEs indicating a lack of mutagenic potential.

Tolerability in sheep AAD 85 and AAD 96 did not reveal any side effects in the treated animals when administered at a dose of 50 mg/kg bodyweight. No deviations from normal values were observed in these sheep, and they stayed in good health during the entire study. AAD 85 and AAD 96 appear to be well tolerated by sheep after oral administration of a dose of 50 mg/kg bodyweight. Similarly, no relevant differences were observed or measured in any animals during the second tolerability study where AAD 1566 was evaluated. There was a transient loss in appetite (several hours) in the sheep treated at 125 mg/kg (50 time the probable minimum dose rate).

Drug/compound	Dose (mg/kg)	Worm counts				
		H. contortus Howick	H. contortus Mox/Fen Australia			
Ivermectin	0.2	977	nd			
Combination of		238	nd			
Ivermectin+	0.2					
Albendazole+	3.8					
Levamisole	7.5					
AAD 96	2.5	0	nd			
AAD 96	1.3	0	nd			
Moxidectin	0.2	nd	407			
Combination of		nd	508			
Moxidectin+	0.2					
Fenbendazole+	5.0					
Levamisole	7.5					
AAD 96	1.3	nd	0			
AAD 1566	2.5	nd	0			

Table 6 Efficacy of AAD 96 and AAD 1566 against adult stages of resistant H. contortus in sheep

Each dose was tested in two to three sheep. The sheep treated with the combination of drugs were used as the internal control for the sheep treated with the AADs. The dose of AAD 96 has been calculated for the active enantiomer (AAD 1566), which makes 50% of the racemic mixture AAD 96. *nd* Not determined



Fig. 1 Efficacy of AAD 96 against adult trichostrongyles in sheep determined by egg count reduction (mean of three sheep per group); treatment at day 0

Discussion

The discovery of the AADs as a new class of synthetic compounds with anthelmintic efficacy is only the first step to finally provide veterinarians and livestock farmers with an alternative for the control of drug-resistant gastrointestinal nematodes. As a next step, at least one AAD needs to be selected to enter a drug development program. Ideally, a development candidate combines best efficacy against all gastro-intestinal nematodes, including the resistant isolates with low toxicity and best tolerability. We followed the approach described by Prichard and Geary (2008) to evaluate previously described (Kaminsky et al. 2008) and herein described follow-up AADs for their potential as drug development candidates. The in vitro EC_{100} values of the AADs evaluated herein were higher than those of some previously tested AADs (Ducray et al. 2008). However, it was not possible to differentiate the AADs described herein on the basis of their in vitro activities since they were all in the same range. Differences in anthelmintic activity were however observed in gerbils. Here, the AADs 85, 95, and 96 expressed higher efficacy at both oral dosing of 0.32 mg/kg

Table 7 Efficacy of AADs in cattle

and subcutaneous dosing of 1 mg/kg than other AADs tested (Table 2; Kaminsky et al. 2008, Ducray et al. 2008). Further differences occurred in efficacy studies in sheep. All AADs eliminated H. contortus and T. circumcincta at a dose of 5 mg/kg except AAD 93 (Table 5). At a dose of 2.5 mg/kg, AADs 85, 94, 95, 96, and 1566 (the active enantiomer in the racemic mixture of AAD 96) were >90% effective against C. curticei, but only AADs 85, 96, and 1566 were >90% effective against N. spathiger. A further comparison in cattle revealed that AAD 96 with its active enantiomer AAD 1566 was the most effective compound curing cattle infected with larval stages of O. ostertagi and C. oncophora at an oral dose of 3.8 mg/kg or adult stages at a topical dose of 5 mg/kg (Table 7). Efficacy studies in gerbils confirmed that only one enantiomer (AAD 1566) of the racemic mixture (AAD 96) expresses anthelmintic activity (Table 3).

Following the observation that AAD 1566 was one of the most effective AADs, it also met the other requirements for a drug development candidate. Essential for any new anthelmintic drug beyond the broad-spectrum efficacy will be the efficacy against drug-resistant, particularly multidrug-resistant nematodes. Both of the two most-suitable AADs, AAD 85a (active enantiomer in the racemic mixture of AAD 85) and AAD 1566, eliminated susceptible and multi-drug-resistant H. contortus in gerbils at the same dose (Table 4). These results were later confirmed in sheep. A dose of 2.5 mg AAD 1566/kg, which was required for broad-spectrum efficacy in sheep, was sufficient to cure sheep infected with each of two multi-drug-resistant H. contortus isolates (Table 6). None of the infected sheep were cured with either triple drug combination. The adult stages of the multi-drug-resistant H. contortus were eliminated in sheep with only half (1.3 mg active enantiomer of AAD 96/kg) of the broad-spectrum dose. The efficacy against multi-drug-resistant nematodes confirms the previous observations of other AADs (Kaminsky

AAD	Dose (mg/kg)	Application route	Stage	Efficacy (%) against		
				O. ostertagi	C. oncophora	
85	9.5	Oral	L4	80	100	
94	9.5	Oral	L4	100	100	
96	9.5	Oral	L4	100	100	
	7.5	Oral	L4	100	100	
	5.0	Oral	L4	99.5	100	
	3.8	Oral	L4	98	100	
1566	4.0	Topical	L4	93	100	
	3.0	Topical	L4	91	99	
	10.0	Topical	Adult	100	100	
	5.0	Topical	Adult	100	100	

AADs were given in all oral applications as racemic mixtures of the two enantiomers. Doses in the table have been calculated for the active enantiomer. AAD 1566 given topical was tested as the purified active enantiomer

et al. 2008) and appears to be a common feature of this chemical class. The resistance-breaking ability indicates a new mode of action.

In addition to the evaluation of efficacy, basic toxicity and tolerability tests were conducted for selected AADs. All AADs appear to be of low toxicity to mammals as demonstrated by the results of the Ames, chromosome aberration, and micronucleus tests. Furthermore, for AAD 85a and 96, at doses ten times higher than the curative dose of 2.5 mg/kg for all tested gastro-intestinal nematodes species in sheep, no side effects were observed. There was only a transient loss of appetite at a 50 times higher than curative dose in sheep of AAD 1566. The observed good tolerability of the AADs in this study confirms the same observation with previously tested AADs (Kaminsky et al. 2008). The most likely reason for the selective action is the nematode-specific molecular target of AADs, which is not present in mammals. AADs interfere with a unique ACR-23 nicotinic acetylcholine receptor subunit (Kaminsky et al. 2008), which belongs to the DEG-3 nematode-specific subfamily (Mongan et al. 2002).

The broad-spectrum efficacy of AAD 1566, monepantel, and the efficacy against resistant and multi-drug-resistant nematode isolates due to a new mode of action, complemented by the low toxicity to mammals and the excellent tolerability in sheep due to a nematode-specific target, render this AAD an anthelmintic drug development candidate. It remains subject to further evaluation if monepantel will become a commercially available anthelmintic product.

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