Burton P. Drayer, MD

# Imaging of the Aging Brain Part I. Normal Findings<sup>1</sup>

A thorough knowledge of the normal changes that occur in the brain with age is critical before abnormal findings are analyzed. Magnetic resonance (MR) imaging improves the ability to distinguish normal and abnormal findings in the brain. The major changes that may occur in elderly individuals without neurologic deficits include enlargement of the ventricles, cortical sulci, and vermian subarachnoid spaces; multifocal areas of hyperintensity in the white matter and basal ganglia; a progressive prominence of hypointensity on T2-weighted images of the putamen, almost equal to that of the globus pallidus; an increase in the oxygen extraction ratio with normal or mildly decreased neuron metabolism; arteriosclerosis in large and small arteries and amyloid angiopathy in leptomeningeal cortical vessels; and decreased dopamine receptor binding in the corpus striatum. Since approximately half of the elderly population exhibits only negligible brain alterations, MR imaging may facilitate the distinction between usual (no neurologic dysfunction) and successful (no brain or vascular changes) aging.

Index terms: Aging • Arteriosclerosis, 17.721 • Brain, atrophy, 10.83 • Brain, MR studies, 10.1214 • Iron • State-of-Art reviews

Radiology 1988; 166:785-796

DVANCES in sophisticated and A sensitive imaging techniques and the expanding population of the elderly necessitate an understanding of normal and pathologic neurologic findings in the elderly. At this time, approximately 21% of the population in the United States is over 55 years old. By 2020, it is estimated that this segment of the population will exceed 30% (1). Elderly people harbor a far greater percentage of neurologic disease per capita than young people. The diagnosis of disease in elderly patients is often complicated because alterations in brain structure and function may occur normally. There is a surprising lack of clinical, radiologic, and pathologic information regarding the normal aging process in humans. Magnetic resonance (MR) imaging should result in a dramatic expansion of our understanding of aging due to its in vivo neuropathologic imaging capabilities and the ability to perform repeat studies over time.

There are various pitfalls that must be recognized when analyzing elderly people who are healthy or diseased (2). Studies may accentuate results from very healthy individuals because those with underlying illnesses will have died (survivor effect). Population heterogeneity is also greater in the elderly, due partially to an increased incidence of other nonneurologic disease. Socioeconomic status, environment, education, nutrition, and exercise may affect the manner in which an individual ages. Rowe and Kahn (3) suggest that normal human aging may be subdivided into usual aging (no overt neurologic symptoms) and successful aging (minimal physiologic loss even when compared with younger individuals). In usual aging, individuals may exhibit abnormalities on glucose tolerance tests (abnormal carbohydrate metabolism), arteriosclerosis (after 50 years of age, only 50% of

brains are free of atherosclerotic arterial changes), systolic hypertension, declining renal and immune function, decreased sensory input (visual and hearing loss), declining crystallized (verbal) and fluid (inductive reasoning and spatial orientation) intelligence, and progressive slowness in movement (4). These underlying alterations may increase the occurrence of various central nervous system (CNS) disorders (e.g., cerebral infarction) in the elderly. It is an enticing theory that successful aging may be enhanced by modification of diet, exercise, and social and intellectual stimulation—all of which may prevent or delay the onset of arteriosclerosis, hypertension, carbohydrate intolerance, or cognitive dysfunction (3).

#### NORMAL AGING

The analysis of MR images of normal brain requires a thorough understanding of the normal and pathologic alterations that occur in the elderly. Most studies of normal aging describe findings from pathologic or imaging studies from individuals who do not have overt neurologic dysfunction. They generally do not include a comprehensive analysis of vascular risk factors (e.g., hypertension, diabetes mellitus, myocardial infarction, arrhythmias), neuropsychologic tests, and extrapyramidal function. The possibility thus exists-and requires further testingthat pathologic differences may be manifest in those individuals with usual aging as compared with those with successful aging.

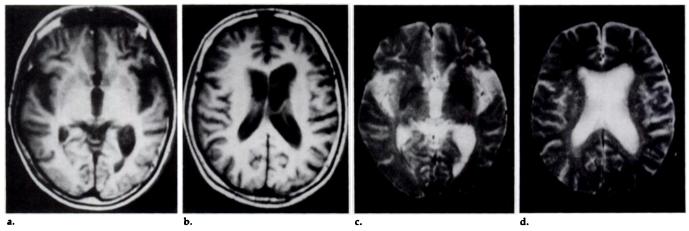
#### **Gray Matter and CSF Spaces**

A mild-to-moderate progressive enlargement of the ventricles, cortical sulci, and pericerebellar subarachnoid spaces may occur with aging (2, 5-9). In an autopsy study of 28 previ-

<sup>&</sup>lt;sup>1</sup> From the Barrow Neurological Institute, 350 W. Thomas Rd., Phoenix, AZ 85013. Received October 16, 1987; revision requested November 20; revision received November 30; accepted December 8. Address reprint requests to the author.

<sup>&</sup>lt;sup>©</sup> RSNA, 1988

See also the article by Drayer (pp. 797–806) in this issue.



**Figure 1.** MR images of the brain of an asymptomatic, 86-year-old person. (a, b) T1-weighted MR images (repetition time [TR] 500 msec, echo time [TE] 20 msec, spin echo [SE 500/20]). Effects of age include widening of the third ventricle, left lateral ventricle (vs. right), left circular sulcus (vs. right), vermian subarachnoid spaces, and anterior interhemispheric fissure. (c, d) T2-weighted MR images (SE 2,000/100). In addition to the above findings, subtle white matter hyperintensities are noted, suggesting arteriolar/hypoperfusion changes due to age.

ously hospitalized patients (age range, 65-92 years; mean, 75 years) without neurologic symptoms, Tomlinson et al. (10) found that 13 patients did not have cortical atrophy and 17 had normal or only mild ventricular enlargement. Cortical atrophy was most prominent in the frontal and parietal parasagittal regions in the study, and moderate ventricular enlargement was associated with infarction of the basal ganglia in six of 11 patients. The brain weights in these 28 patients varied from 1,170 g to 1,430 g (mean, 1,320 g) in men and from 1,080 g to 1,390 g (mean, 1,213 g) in women. Senile neuritic plaques (dense amyloid core surrounded by neurites, astrocytic processes, amyloid, adipose tissue, and microglia containing iron), neurofibrillary degeneration (tangles of twisted tubules and helically wound fibrils), and granulovacuolar degeneration (vacuoles in cytoplasm of hippocampal pyramidal cells) were found in significant numbers (though far less than that in Alzheimer disease) in five patients.

Various authors have reported a selective loss of neurons with age (10– 18). These changes were most prominent in the superior frontal and temporal gyri, precentral gyrus, corpus striatum, hippocampus, thalamus, amygdaloid body, inferior olive, and Purkinje cells and dentate nucleus of the cerebellum. There is a progressive intraneuronal accumulation of lipofuscin, melanin, and ceroid with age—similar findings were noted in healthy individuals and those with dementia (2, 13, 19-21). Lipofuscin is most prominent in the cranial and spinal motor nuclei, red nucleus, thalamus, globus pallidus, inferior

olive, and dentate nucleus of the cerebellum. A decrease in dendritic branching and possible loss of neuronal synapses in the temporal, frontal, and limbic regions of the cerebrum also characterize normal aging (22). Widening of the cortical sulci may be related to cortical and subcortical gray matter versus white matter loss. Miller et al. (23) reported that the ratio of gray matter to white matter was 1.28 at age 20, 1.13 at age 50, and 1.55 at age 100, which suggests that white matter atrophy exceeds that of gray matter with age.

Numerous and extensive studies have been performed with computed tomography (CT) to analyze the limits of normalcy in healthy, elderly individuals (8, 24-34). The methods that were used included visual ratings by experienced observers, measurements of a variety of ventricle-tobrain indices (e.g., Evans, frontal horn, bicaudate, cella media, third ventricle-Sylvian fissure), and volumetric pixel counting. Most CT studies indicate that a progressive enlargement of the ventricles and cortical sulci ("physiologic atrophy") is characteristic of the normal aging brain (Fig. 1). An analysis of 500 healthy patients by Nagata et al. (32), who used pixel counting and linear ventricle-to-brain measurements, confirms that the CSF-to-brain ratio (CSF volume/cranial volume) remains constant from 10 to 50 years of age, followed by a highly variable, progressive dilation of the CSF spaces with increasing age. This study corroborates the findings of Yamaura et al. (35) (228 healthy adults) and Schwartz et al. (36) (30 healthy men) who suggest physiologic atrophy (CSF-space enlargement) begins in the 5th decade. Other large CT studies that describe the CSF spaces in healthy adults conclude that dilatation may not become apparent until the 6th or 7th decade. Jacoby et al. (31), who studied 50 healthy subjects (ten men, 40 women) aged 62-88 years (mean, 73 years), found no significant alterations in neuropsychologic tests for memory and orientation or in Evans ratio (maximum width of frontal horns of lateral ventricles to the maximum diameter of the internal skull) but defined definite progressive alterations in the CSF spaces as determined with a visual rating scale and planimetry.

The CT studies correlate closely with pathologic reviews of CSF-space expansion with normal aging. Hubbard and Anderson (37) described individuals with ventricular enlargement after 60 years of age. Morel and Wildi (38) studied brains that were formalin fixed and found a progressive increase in ventricular size from 55 to 99 years of age. Tomlinson et al. (10) found enlargement of the cortical sulci or ventricles in approximately half of the autopsy studies of nondemented people over 65 years old. Both CT and postmortem studies highlight the heterogeneity of CSFspace size in the elderly population, with approximately 30%-50% within the range of normal for young adults (8). Although most authors have focused on the cortical sulci, the pericerebellar subarachnoid (especially superior vermian) spaces also dilate in the elderly (7, 26, 39).

Even though enlargement of the CSF spaces during aging is generally diffuse, there are specific locations in which dilatation and asymmetries are best delineated (8, 40) (Fig. 1, Table 1). There is regression of the median nuclei of the thalamus after 50 years of age (14), which explains the early demonstration of third ventricular enlargement (8, 25, 41). There is generally only mild enlargement of the temporal horns of the lateral ventricles with aging (10, 42). The left lateral ventricle is normally larger than the right (8). Widening of the superficial cortical sulci is often seen first in the frontal and parietal parasagittal regions (8, 10, 43). The anterior interhemispheric fissure and the cerebellar vermis also progressively widen with age (8, 18, 24, 26, 33). Enlargement of the cortical sulci in the central, precentral, postcentral, and superior frontal gyri occurs later and may be related to loss of white matter (8, 23, 44, 45). Because of involutionary changes in the temporal lobes with aging, the anterior end of the circular sulcus (Sylvian fissure) may become prominent in the 5th decade (8). At all times of life, the left circular sulcus is larger than the right; this finding should not be mistaken for adjacent ischemic changes (40).

### **Cerebral White Matter**

A variety of neuropathologic, CT, and MR imaging studies suggest that 30%-80% of elderly individuals without neurologic deficits have focal abnormalities in the cerebral white matter (10, 46-55). These alterations are usually demonstrated by MR imaging as small, focal (sometimes confluent) areas of increased signal intensity (SI) on T2-weighted images that are often found scattered throughout the deep cerebral white matter (especially in the frontal and parietooccipital areas), basal ganglia (notably globus pallidus and putamen), and capping the lateral ventricular margins. These signal hyperintensities, particularly when small and patchy, have been facetiously called "unidentified bright objects" or white matter/basal ganglia, that is, "subcortical hyperintensities" (Fig. 2). Because of the high prevalence of this MR finding in the elderly (47, 56-59) and an ongoing discussion on the nature of vascular dementia (52, 56, 60, 61), a certain amount of confusion exists concerning the pathologic substrate of subcortical hyperintensities. The issue of white matter and basal ganglia changes in normal aging is further confounded by limitations of normal postmortem studies,

Summary of MR Imaging: Normal and Pathologic Aging

Table 1

	Atrophy in	E		Subcortical T2	2	Basal Ganglia	slia	Arterial	erial		Brain M	Brain Metabolism
	Lor spaces	Ces	1	Hyperntensities	nes	Hypointensity	sity -	Nark	Narrowing			
Disorders	Ventricles	Sulci	White Matter	Basal Ganglia	Pons/ Thalamus	GP/RN/SN/DN	Putamen	Atheroma Infarction	Occlusion	Amyloid Angiopathy	CMR02	4 Dopamine
Normal Adults ≤50 vr	•	•	-	0	0	e	1	•	0	0	0	0
Successful aging Usual aging	-14	24	74	-4	01	ოო	1	77	0-	77	01	77
Demenna Alzheimer Pick‡	00	44	ოო	00	<b></b>	ოო	77		<b></b>	NK NK	20	99
Binswanger NPH <sup>5</sup>	) <del>- 4</del> 1 - <del>4</del> 1	' n n	40	4-	۰ <del>۵</del> –	ο Υ	2 3	ю <del>–</del>	÷.	žž	100	20
Wernicke Bradykinesia/	ñ	ñ	7	7	ę	£	7	-	1	NK	6	7
rigidity Parkinson	7	4	ť	2	1	4	ę	1	1	£	2	ŵ
Multiple system abnormalities	2	4	ę	2	1	4	4+	1	1	NK	2	4+
Progressive supranu- clear palsy Hypothyroidism	00	<b>4</b> W	е <b>-</b> 1	12	07	4 4+	3 4+	<del>,</del>		NK	20	<b>4</b> 4 4
ocal deficit Infarction Hematoma <sup>1</sup>	00	ωN	60 <del>4</del> 4	<b>₩</b> 4	€0 <b>4</b>	ლ <b>ლ</b>	2-3 2	40	<b>4</b> 4	45 4	40	<b>N</b> N

GP = globus pallidus, RN = red nucleus, SN = substantia nigra, DN = dentate nucleus. Abnormal gyral hypointensity in half, particularly in parietal cortex. Predominantly frontemporal atrophic changes. NPH = normal pressure hydrocephalus. Hyperintensity in subacute hematoma (hypointensity if acute or chronic).

Han

such as inclusion of patients with chronic systemic, cardiac, psychiatric, and neurologic diseases; incomplete premorbid information on cognitive function and vascular risk factors; and absence of comprehensive analyses of incidental alterations in the deep white matter or basal ganglia.

Subcortical hyperintensities are distributed in long, noncollateralizing, perforating vessels, such as medullary and lenticulostriate arteries. The common denominator of all such lesions is the loss of a focal area of brain parenchyma with increased tissue water, resulting in an increased SI on the MR image. After studying 240 consecutive MR imaging studies and concluding that patchy, subcortical foci of increased SI correlated best with ischemic cerebrovascular disease, hypertension, and aging, Awad et al. (57, 58) compared MR images with neuropathologic studies on eight postmortem brains. They found that the hyperintensities on MR images involved the periventricular white matter, optic radiations, basal ganglia, and centrum semiovale in a decreasing order of frequency and were associated with a spectrum of histologic changes. The most common histologic change was arteriolar ectasia with enlargement of surrounding perivascular spaces that reflected atrophy of the brain tissue around blood vessels. This change resulted in an "extensive network of tunnels filled with extracellular water." This condition was named état criblé (sievelike) by Durand-Fardel (62) in 1843; its association with hypertension and aging and confusion with, at times, coexisting multiple subcortical infarctions (état lacunaire) are of greatest importance (63-65). Additional associated findings that were less commonly found by Awad et al. (57, 58) and had the same hyperintense appearance were myelin pallor and lacunar infarction with associated arteriosclerosis of perforating arterioles. Degeneration of myelinated axons and gliosis was limited to subependymal (immediate periventricular) areas and surrounding areas of a small infarction.

Kirkpatrick and Hayman (59) performed a postmortem neuropathologic analysis of brains from 15 healthy (52-72 years old) subjects who had a high frequency (ten of 15) of systemic cancer. The researchers found small white matter lesions in 12 of the 15 autopsy examinations. The most common findings (eight subjects) were atrophy of axons and myelin with associated gliosis; tortuous, sclerotic, and thickened vessels; and increased extracellular water (i.e., atrophic perivascular demyelination). They found vascular malformations in four subjects (three telangiectasia, one capillary angioma). Awad et al. (57) also found, at autopsy, a small telangiectasia in two of eight brains and a diverticulum of the lateral ventricle that extended into the adjacent white matter in three brains. These researchers (57-59) postulate that hypertension may predispose to the atrophic perivascular demyelination—suggesting that arteriolar thickening and sclerosis result in a loss of the normal nutritive function of the arteriole (66); chronic, low-grade vascular insufficiency; and atrophic perivascular demyelination or myelin pallor rather than frank infarction.

It is difficult to analyze the significance of subcortical hyperintensities without an understanding of the theories concerning white matter abnormalities and their proposed relationships to multiinfarct and Binswanger dementia (Fig. 3). A review of this literature makes one increasingly aware that limitations in analysis of unidentified bright objects on MR images are a direct correlate of neuropathologic uncertainties (10, 52, 55, 67, 68). Vascular dementia secondary to multiple subcortical infarctions and diffuse myelin pallor, sparing of the subcortical arcuate fibers, and clinical hypertension was initially reported by Binswanger in 1894 (69). Multiple articles have provided refinements, theoretical considerations, and alterations in nomenclature (70-73) (e.g., subcortical arteriosclerotic encephalopathy). Hachinski et al. (74) popularized the term "multiinfarct dementia" for patients with large areas of cortical and subcortical infarction, decreased cerebral blood flow, and a clinical picture that differed from that of Alzheimer disease (75).

One theory suggests that the brain substance in the distribution of the most distal branches of the brain arteries-the cerebral white matter and basal ganglia—is most susceptible to a reduction in blood flow; that is, the deep centrum ovale is a watershed zone (71, 76). Studies in baboons and dogs have shown that a progressive reduction in blood pressure can result in absent blood flow in the centrum ovale (with infarction) while the cerebral cortex is still being perfused (77, 78). The subcortical arcuate fibers are spared because the blood supply for this region is from the cor-

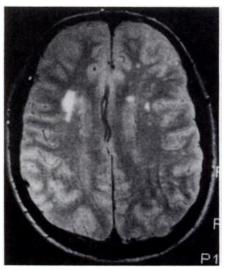


Figure 2. MR image shows subcortical hyperintensities. Intermediate-weighted MR images (SE 2,500/40) are often optimal for delineating white matter hyperintensities, particularly when they are periventricular as seen in this MR image of the brain of an asymptomatic 71-year-old patient with hypertension. These lesions may represent état criblé, atrophic perivascular demyelination, gliosis, myelin pallor, demyelination, or infarction.

tical (or from the cortical Duvernoy type-5 and medullary arterioles) rather than exclusively from the deep medullary supply (79-82). Hypoperfusion in humans-whether due to carotid artery occlusion, hypoxia, or hypotension-often results in cerebral infarction that involves the deep white matter in a distribution similar to Binswanger dementia with sparing of the cortical surface due to a leptomeningeal collateral arterial supply. Ginsburg et al. (83, 85) further correlated the extent of the white matter abnormality with the degree of systolic hypotension and metabolic acidosis rather than with the amount of hypoxia.

Another theory suggests that the long, perforating medullary arteries that supply the centrum semiovale are particularly sensitive to the effects of hypertension (76). Arteriolar narrowing, loss of vasoregulation, and chronic ischemia may result in atrophic perivascular demyelination, myelin pallor, gliosis, and/or infarction (56, 71, 76). Feigin et al. (85, 86) postulated that cerebral edema played a key role in white matter disease leading to secondary myelin pallor and thick-walled, hyalinized arterioles. Congestion and stasis within the deep venous system secondary to obstruction or right-sided heart failure may also cause damage to the deep white matter. Van den Bergh and vander Eecken (87) found that

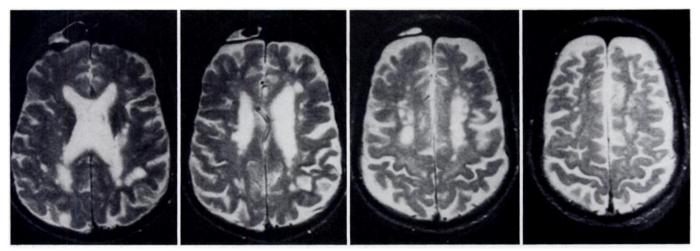


Figure 3. MR images show microangiopathic leukoencephalopathy. T2-weighted MR images (SE 2,500/80) demonstrate multiple, at times confluent, areas of increased SI throughout the deep white matter with sparing of the arcuate fibers in the brain of a 73-year-old patient with dementia. Although the marked prominence of the cortical sulci may be related to white matter destruction, the dementia and atrophy in this case could also be due to a primary degenerative dementia (Alzheimer disease) with arteriolar/hypoperfusion abnormalities that was suspected clinically. This case highlights the complex problems of determining a precise cause for a signal hyperintensity (i.e., infarction vs. myelin pallor vs. gliosis vs. atrophic perivascular demyelination) and distinguishing a vascular versus a degenerative cause for dementia.

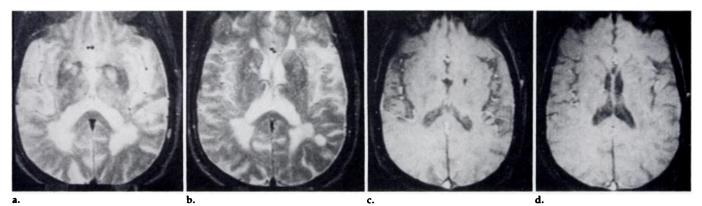


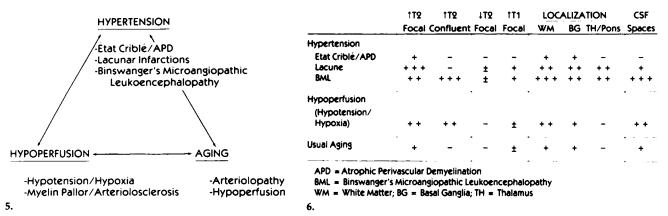
Figure 4. Microangiopathic leukoencephalopathy with dementia and hemorrhage. (a, b) T2-weighted MR images (SE 2,500/80). Extensive, confluent hyperintensities in the cerebral white matter (particularly adjacent to the occipital horns), internal capsules, basal ganglia, and thalamus likely represent infarction secondary to hypertensive arteriolar disease; however, myelin pallor, atrophic perivascular demyelination, and/or état criblé may also be present. (c, d) Gradient-echo T1-weighted MR images (SE 300/12, flip angle 60°). Small hypointensity in the left genu of the internal capsule-globus pallidus region is consistent with a slit hemorrhage residual of hypertensive hematoma, confirming hypertensive vascular (arteriolar) disease.

the cerebral cortex and underlying arcuate fibers drained via the superficial venous system, while the cerebral white matter was drained by the deep venous system.

Although many of the reported MR and neuropathologic studies (46, 47, 52, 57, 59) suggest that hypertension is an important accompanying feature with subcortical hyperintensities (Fig. 4), many healthy individuals with white matter alterations do not have hypertension. In a recent study, Fazekas et al. (48) found subcortical hyperintensities in the majority of patients who were either healthy (control subjects) or suffering from Alzheimer disease but found no relationship of this finding to hypertension or other vascular risk factors. They did, however, describe

a correlation between multiinfarct dementia with prominent hyperintensities in the white matter and basal ganglia and a history of hypertension. Autopsy studies of 97 patients (20 with Alzheimer disease, 28 with senile dementia Alzheimer-type, 23 with multiinfarct dementia, 16 nondemented, normotensive patients 70-100 years old, and ten nondemented, normotensive patients 49-69 years old) by Brun and Englund (56) found definite white matter abnormalities in 11 of 20 (four moderate or severe) patients with presenile Alzheimer disease, 19 of 28 (six moderate or severe) with senile dementia of the Alzheimer-type, 23 of 23 with multiinfarct dementia, zero of ten nondemented, normotensive patients aged 49-69 years, and two of 16 nondemented, normotensive patients aged 70-100 years. Although 32 of 48 patients with Alzheimer dementia or disease had a history of cardiovascular disease and/or hypotension, only one of 48 was hypertensive and none had nephrosclerosis. This suggests that brain hypoperfusion and hypotension were more important than hypertension as precursors to white matter damage and that white matter alterations were common in Alzheimer disease at all ages (56).

The white matter changes described by Brun and Englund (56) consisted of myelin pallor (rather than frank infarction); fibrohyaline arteriosclerosis with no staining with Congo red; partial loss of axons, myelin sheaths, and oligodendroglia; and mild reactive astrocytosis—these



Figures 5, 6. (5) Diagram shows hypoperfusion/arteriolopathy spectrum. (6) Diagram shows the MR imaging characteristics of arteriolar disease.

changes are similar to those seen with Binswanger disease, except that the myelin loss and asymmetric scattered infarctions in the white matter and basal ganglia were less common. The cerebral white matter changes were predominantly symmetric, extended from the periventricular region outward, spared the subcortical arcuate fibers, and were most prominent in the frontal and parietal lobes. In a series of 40 patients with Alzheimer disease who underwent MR imaging, 21 had subcortical hyperintensities of variable extent; these 21 had a far higher frequency of hypertension-extensive information concerning hypotensive episodes was not obtained (personal observations).

It is increasingly apparent that various theories regarding the origin of white matter and basal ganglia hyperintensities may all be at least partially true. An attempt has been made to synthesize these MR imaging and pathologic observations into a single, unifying hypothesis (Figs. 5, 6). A common denominator of subcortical hyperintensities, particularly in the asymptomatic elderly population, is brain hypoperfusion and arteriolar disease. The hypoperfusion occurs in the distribution of the long, noncollateralizing, perforating vessels that supply the periventricular and deep cerebral white matter (sparing the arcuate fibers) and basal ganglia. The most common causes of hypoperfusion are episodes of hypotension, hypoxia secondary to cardiac or carotid artery disease, hypertension, and/or aging. The entire centrum semiovale is a watershed zone supplied by the most distal intraparenchymal penetrating arterioles. Finally, other chronic processes that involve the white matter (including multiple sclerosis, acute disseminated encephalomyelitis, traumatic injury) and diseases such as systemic lupus erythematosis may mimic the leukoencephalopathic alterations seen with the hypoperfusion arteriolar diseases. The absence of white matter changes in an elderly individual may be an important hallmark of successful aging.

### **Basal Ganglia**

Nonheme brain iron is normally found within oligodendroglia and astrocytes with smaller amounts in neurons and myelinated axons. Approximately half the cellular iron is in the mitochondria and microsomes, 10%-15% in the nuclei, and 40% in a soluble fraction presumably representing ferritin (88). Histochemical, histopathologic, and MR imaging studies have determined that maximum iron concentration in normal adults is found in the globus pallidus, red nucleus, pars reticulata of the substantia nigra, and dentate nucleus of the cerebellum (88-94) (Figs. 7, 8). Intracellular brain iron is probably stored in two metabolically active compartments-ferritin and free iron (95-98). Iron plays an important role in oxidative phosphorylation, dopamine synthesis (99, 100) and turnover (cofactor in monoamine oxidase reaction), and hydroxyl free radical formation (101). Iron is present in all animal systems with the localization quite similar in rats, baboons, and humans (102, 103). The mechanism and site of transport across the bloodbrain barrier (BBB) is poorly understood because the greatest transferrin receptor density correlates poorly with the highest iron distribution (104). The concentration of brain iron is independent of body stores, even in hemochromatosis. Iron is best seen on T2-weighted and gradient-echo MR images as a hypointensity due to field heterogeneity and magnetic susceptibility (T2) effects (90, 105,

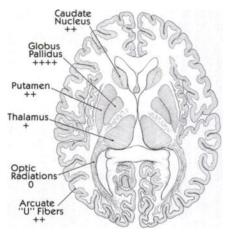


Figure 7. Diagram shows the localization of iron in normal brain. ++++=>20 mgFe/100 g, +++=15-20 mg Fe/100 g, ++=10-15 mg Fe/100 g, +=5-10 mg Fe/100 g, 0=0-5 mg Fe/100 g.

### 106) (Fig. 9).

There is a preferential progressive increase of iron in the corpus striatum (caudate and putamen) with aging so that the iron concentration normally may be equal to that in the globus pallidus by the 8th decade (88, 90). This increased accumulation of iron with aging may be related to a combination of factors including decreased oxidative phosphorylation, declining oligodendroglial function, decreased dopamine production and turnover, abnormal BBB permeability, or accelerated hydroxyl free radical formation with lipid membrane peroxidation. Aging is not only associated with increased iron in the brain tissue but also with an increased concentration of iron in the walls of blood vessels (vascular ferrugination). In addition to the normal increases of iron in the corpus striatum of the elderly, there is smudging and an increased indistinctness of the iron in the dentate nucleus (90), and a mild increase of iron in the oc-

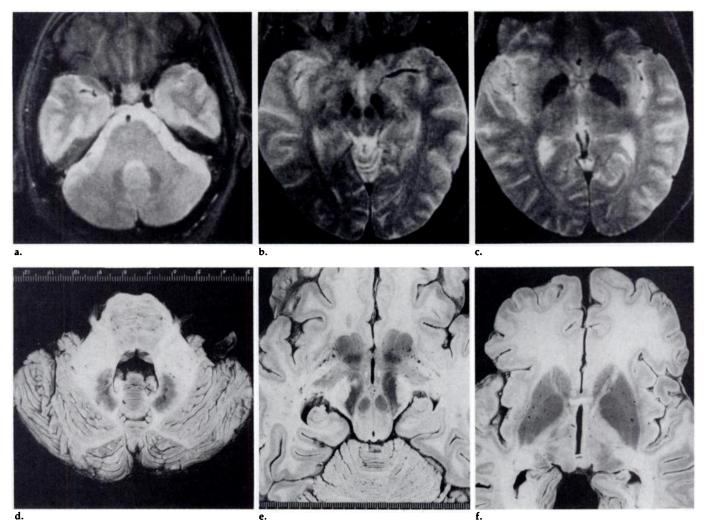


Figure 8. Normal brain iron in asymptomatic, middle-aged adults. (a-c) T2-weighted MR images (SE 2,500/80). There is normally a decreased SI in the dentate nucleus (a), red nucleus and pars reticulata of substantia nigra (b), and globus pallidus (c), correlating with sites of maximum iron distribution. (d-f) Anatomic sections stained for hemosiderin with Perls reaction confirm the iron localization in the dentate nucleus (d), red nucleus and pars reticulata of globus pallidus (f). A higher iron concentration than that in the thalamus or white matter is apparent in the caudate nucleus and putamen.

cipital and motor cortices that roughly parallels lipofuscin in neurons and neuroglia (107). MR studies that show decreased signal intensity on T2-weighted images and Perls reaction, which shows increased blueness, for demonstration of ferric iron in postmortem brains of elderly patients (90) correlate fully with prior histochemical and histopathologic findings of a progressive increase of iron concentration in the corpus striatum with aging in individuals who do not have neurologic deficits (88) (Fig. 10). An abnormal accumulation of iron has been described in parkinsonism (91, 108-110) and Alzheimer disease (107, 111, 112).

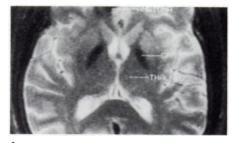
The formation of hydroxyl free radicals requires iron ions and oxygen free radicals and results in membrane lipid peroxidation, aldehyde production, and accumulation of lipofuscin in brain neurons (21, 101, 113). Neurons in the thalamus, lateral geniculate body, and dentate nucleus of the cerebellum are particularly rich in lipofuscin. Oxidative mechanisms are important in the formation of lipofuscin, which accumulates in postmitotic cells, such as brain neurons. The aging brain is quite susceptible to oxidative damage (114-117) because it contains a high concentration of unsaturated lipids, it uses over 20% of the body oxygen, and it has low concentrations of antioxidant enzymes (e.g., superoxide dismutase catalase, glutathione peroxidase) and vitamin E. Iron in the brain helps regulate the dopamine receptor, dopamine synthesis, and monoamine oxidase activity (90, 99, 100); nigrostriatal dopaminergic function is the only neurotransmitter system that declines with normal aging (118-120). Floyd et al. (117) described a direct correlation of total iron content in the brain (ferritin plus mobile iron) and brain peroxidation. To initiate membrane lipid peroxidation, iron must be moved from

ferritin into a mobile form (e.g., iron nucleotide complex). The correlation of iron concentration and peroxidation may not hold true for the corpus striatum, presumably because iron dominantly ligates with dopamine and prevents the participation of mobile iron in hydroxyl free radical formation and membrane peroxidation (117). The high concentration of iron in the basal ganglia makes these structures quite susceptible to oxidative injury when the dopamine activity decreases and may help explain the decline in mobility (bradykinesia, tremor) that occurs with normal and accelerated (Parkinson disease) aging. There is also a loss of neurons in the putamen of elderly patients (121).

#### Vascular

Evidence of arteriosclerosis is seen in the brain vasculature in 50% of patients over 50 years of age (122). In a series of 994 consecutive autopsy examinations, Jorgensen and Torvik (123) found that 320 patients had ischemic cerebrovascular disease. Of these 320 patients, symptoms of infarction were absent in 124 cases. Fisher (61, 64) found neither a history of stroke nor evidence of neurologic deficits in 88 of 114 cases of a single lacunar infarction, even though a history of hypertension was common. Cerebrovascular disease is seven times more common in patients with hypertension, and approxmately 25% of elderly patients who have had cerebrovascular accidents had hypertension (124, 125). Cardiac disease ranked first (even before hypertension) as the major risk factor for cerebrovascular disease in the elderly (124, 125). Although atrial fibrillation and other cardiac arrhythmias with resultant bradycardia are the most important risk factors for cerebrovascular accidents (126, 167), congestive heart failure and coronary heart disease may also play a significant role.

Cerebral amyloid angiopathy is a common finding in elderly individuals both with and without neurologic abnormalities. Esiri and Wilcock (128) described amyloid changes in the leptomeningeal, intracortical, and other small arteries in 37 (11 extensive) of 45 autopsy cases of Alzheimer disease, 14 (three extensive) of 41 cases with other degenerative or cerebrovascular dementias, ten (three extensive) of 32 cases with cerebrovascular disease and no dementia, and 34% (none extensive) of nondemented individuals either with or without associated, nonvascular disease. In the non-Alzheimer group as a whole, amyloid angiopathy was found in 33% of patients and did not increase with age from 60 to 102 years. In mild cases, only the tunica media of small vessels is involved while the full vessel wall may be involved in the more severe cases. Esiri and Wilcock (128) also reported more extensive leptomeningeal vessel involvement at the depths of sulci, sparing of subcortical and deep white matter as well as lenticulostriate vessels, equal involvement of each cerebral lobe, and only minimal amyloid changes in the hippocampus. Vinters and Gilbert (129) reported similar findings in 84 consecutive autopsy examinations: a patchy asymmetric amyloid angiopathy that affected 46% of the brains of patients over 70 years old, involvement of small- and medium-sized cortical and leptomeningeal vessels with sparing of the



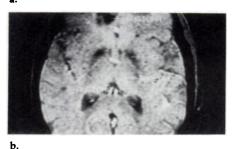


Figure 9. MR images show ferritin and hemosiderin T2 hypointensity. The signal hypointensity in the globus pallidus (GP) due to ferritin is prominent on (a) T2-weighted (SE 2,500/80) and (b) T1-weighted gradientecho (SE 300/12, flip angle 60°) MR images confirming that visualization is due to magnetic susceptibility. A cavernous hemangioma is also hypointense due to hemosiderinladen macrophages.

hippocampus and cerebral white matter, and no correlation with amyloid outside the CNS. This high frequency of cerebral amyloid angiopathy in all aging brains and the greater extent and higher frequency of Alzheimer disease has been confirmed by many investigators (130– 133).

There is a close relationship between amyloid angiopathy and intracerebral hematoma (134-137) (Fig. 11). These hematomas generally occur in elderly, often demented (with histologic abnormalities that resemble Alzheimer disease) individuals. They are characterized by cortical or immediate subcortical localization, direct extension into the adjacent subarachnoid space, multiplicity of sites, and amyloid replacement of the tunica media in small- and mediumsized arteries. There is a close similarity in the immunologic staining of the core of vascular and senile plaque amyloid, which share a common antigen with neurofibrillary tangles. Vascular amyloid may reflect an abnormality in the nerve terminals that innervate the leptomeningeal and cortical blood vessels. In a familial form of cerebral amyloid angiopathy found in younger individuals, a gamma trace-protein deposition has been reported (137).



Figure 10. Anatomic section represents normal aging. A Perls reaction for hemosiderin in the brain of a 74-year-old man (no premortem neurologic abnormalities) shows excess staining in the putamen and caudate (almost equal to the globus pallidus), which may occur normally with aging.

## **Brain Metabolism**

Controversy exists concerning whether regional cerebral blood flow, the cerebral metabolic rate for oxygen, or the cerebral metabolic rate for glucose declines with age in healthy individuals (2, 138-144). These discrepancies may occur due to a variety of problems that plague metabolic imaging studies, particularly in the aged: Auditory and visual stimuli will affect regional cerebral blood flow, and vision and hearing are often physiologically impaired in the elderly (138, 145-149); precise anatomic localization is difficult with positron emission tomography (PET) techniques that result in partial volume averaging of gray (approximately 80 mL/100g/min) and white (approximately 20 mL/100g/min) matter flow (148, 150-156); arteriosclerotic cerebrovascular disease is present, even if minimal, in approximately 50% of individuals over 50 years old, possibly resulting in decreased regional cerebral blood flow (2, 4, 122, 138, 144, 157); and cerebral atrophy and ventricular enlargement associated with aging result in less tissue per unit volume and thus may cause the false impression of decreased regional cerebral blood flow and cerebral metabolic rates for oxygen and glucose, when actually the intrinsic resting cellular metabolism of the tissue per unit weight is normal (2, 138, 151, 156).

To account for some of these factors, some researchers have suggested that cerebral blood flow and metabolic rate for oxygen do not vary with age (138, 139, 148, 157). Frackowiak and Gibbs (154) report that regional cerebral blood flow and

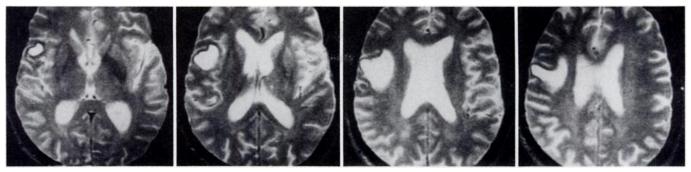


Figure 11. Lobar intracerebral hematoma: amyloid angiopathy. T2-weighted MR images (SE 2,500/80). A prominent mass of hyperintensity (extracellular deoxyhemoglobin) surrounded by a rim of hypointensity (macrophages laden with hemosiderin) in the brain of a 75-year-old patient with mild dementia and acute onset of a focal neurologic deficit is indicative of subacute lobar hematoma.

oxygen extraction decrease with age, while the cerebral metabolic rate for oxygen is normal. Smith (138) concludes that vascular disease is an important contribution to senescence, may accelerate the aging process, and may account for reported declines in cerebral blood flow and metabolism.

Kuhl et al. (158), using F-18 deoxyglucose and PET imaging, suggested that healthy, elderly individuals may have a decrease in the cerebral metabolic rate for glucose. Declines have also been noted in the dominant electroencephalogram rhythm (50% have slowing, particularly over the left anterior temporal region [159-161]), reaction times in psychomotor tests (2), and fluid (reasoning and spatial orientation) intelligence (148). A reduction in the cerebral metabolic rate for glucose was found in the auditory system, visual system, globus pallidus, and corpus striatum in middle-aged and elderly Sprague-Dawley rats (162). The decline of glucose utilization in the visual and auditory cortex, however, may reflect degenerative alterations in the retina and cochlea (138, 145-148). Slowed movements (parkinsonian features) and deterioration of the dopamine system may be either the cause or the result of decreased glucose utilization in the corpus striatum and globus pallidus. Duara et al. (148) found that age did not affect the cerebral metabolic rate of glucose when patients 21-83 years old were studied during sensory (visual and auditory) deprivation; this suggests that declines in brain metabolism with aging may reflect decreased sensory input (138, 163).

Age-related alterations have been pronounced in only the nigrostriatal dopaminergic system (119, 120, 164). Wong et al. (118) studied 44 healthy volunteers with PET and carbon-11labeled 3-N-methylspiperone, which preferentially binds to the D2 dopamine receptor. They found a progres-

sive decline in specific binding to the D2 dopamine receptor in the corpus striatum with increasing age; this decrease was less prominent in women. Explanations for decreased receptor binding include a decline in the number of D2 dopamine receptors, a decrease in the number and size of cell bodies in the substantia nigra (pars compacta) and putamen, and/or a decline in the concentration of the synthetic enzyme tyrosine hydroxylase in the corpus striatum and nucleus accumbens. Although the muscarinic cholinergic system is thought to play an important role in memory functions (165), there is no consensus on whether choline acetyltransferase or the density of cholinergic receptors decrease with age (120, 165-167).

#### References

- 1. Ansberry C. Day-care centers for elderly spring up as alternative to costly nursing homes. Wall Street Journal, December 8, 1986, p. 29.
- 2. Creasey H, Rapoport SI. The aging human brain. Ann Neurol 1985; 17:2-10.
- Rowe JW, Kahn RL. Human aging: usual and successful. Science 1987; 237:143– 149.
- O'Brien MD, Mallett BL. Cerebral cortex perfusion rates in dementia. J Neurol Neurosurg Psychiatry 1970; 33:497–500.
- Dekaban AS, Sadowsky D. Changes in brain weights during the span of human life: relation of brain weights to body heights and body weights. Ann Neurol 1978; 4:345-356.
- Davis PJM, Wright EA. A new method for measuring cranial cavity volume and its application to the assessment of cerebral atrophy at autopsy. Neuropathol Appl Neurobiol 1977; 3:341-358.
- Roessmann U, Ho KC, Straumfjord JV, et al. The weight of the infratentorial portion of the adult brain and analysis of the infratentorial/whole brain weight ratio. J Neuropathol Exp Neurol 1982; 41:536– 547.
- LeMay M. Radiologic changes of the aging brain and skull. AJR 1984; 143:383– 389.
- 9. Gomori JM, Steiner I, Melamed E, et al. The assessment of changes in brain vol-

ume using combined linear measurements: a CT-scan study. Neuroradiology 1984; 26:21-24.

- Tomlinson BE, Blessed G, Roth M. Observations on the brains of nondemented old people. J Neurol Sci 1968; 7:331-356.
- Brody H. Organization of the cerebral cortex. III. A study of aging in the human cerebral cortex. J Comp Neurol 1955; 102:511-556.
- Anderson JM, Hubbard BM, Coghill GR, et al. The effect of advanced old age on the neurone content of the cerebral cortex: observations with an automatic image analyser point counting method. J Neurol Sci 1983; 58:235-246.
- Brody H, Vijayashankar N. Anatomical changes in the nervous system. In: Fince CE, Hayflick L, eds. Handbook of the biology of aging. New York: Van Nostrand, 1977; 241-261.
- Yakovlev PL. Morphological criteria of growth and maturation of the nervous system in man. Ment Retard 1961; 39:3-46.
- Terry RD. Some biological aspects of the aging brain. Mech Ageing Dev 1980; 14:191-201.
- Peress NS, Kane WC, Aronson SM. Central nervous system findings in a tenth decade autopsy population. Prog Brain Res 1973; 40:473-483.
- Miquel J, Johnson JE Jr, Cervos-Navarro J. Comparison of CNS aging in humans and experimental animals. In: Cervos-Navarro J, Sarkander HI, eds. Brain aging: neuropathology and neuropharmacology. Aging. Vol. 21. New York: Raven, 1983; 231-258.
- Huag H. Quantitative investigation of the human cerebral cortex. Gerontology 1981; 27:105-111.
- Mann DNA, Yates PO. Lipoprotein pigments: their relationship to ageing in the human nervous system. I. The lipofuscin content of nerve cells. Brain 1974; 97:481-488.
- Mann DMA, Yates PO. The effects of ageing on the pigmented nerve cells of the human locus caeruleus and substantia nigra. Acta Neuropathol (Berl) 1979; 47:93-97.
- Barden H. The histochemical relationship of neuromelanin and lipofuscin. J Neuropathol Exp Neurol 1969; 28:419– 441.
- 22. Huttenlocher PR. Synaptic density in human frontal cortex: developmental changes and effects of aging. Brain Res 1979; 163:195-205.
- 23. Miller AKH, Alston RL, Corsellis JAN. Variation with age in the volumes of grey and white matter in the cerebral

hemispheres of man: measurements with an image analyser. Neuropathol Appl Neurobiol 1980; 6:119–132.

- Barron SA, Jacobs L, Kinkel WR. Change in size of normal lateral ventricles during aging determined by computed tomography. Neurology 1976; 26:1101-1113.
- Brinkman SD, Sarwar M, Levin HS, et al. Quantitative indexes of computed tomography in dementia and normal aging. Radiology 1981; 138:89–92.
- Cala LA, Thickbroom GW, Black JL, et al. Brain density and cerebrospinal fluid space size: CT of normal volunteers. Am J Neuroradiol 1981; 2:41-47.
- Gado M, Hughes CP, Danziger W, et al. Volumetric measurements of the cerebrospinal fluid spaces in demented subjects and controls. Radiology 1982; 144:535-538.
- Laffey PA, Peyster RG, Nathan R, et al. Computed tomography in aging: results in a normal elderly population. Neuroradiology 1984; 26:273-278.
- Haug G. Age and sex dependence of the size of normal ventricles on computed tomography. Neuroradiology 1977; 14:201-204.
- Hughes CO, Gado M. Computed tomography and aging of the brain. Radiology 1981; 139:291-396.
- Jacoby RJ, Levy R, Dawson JM. Computed tomography in the elderly. I. The normal population. Br J Psychiatry 1980; 136:249-255.
- 32. Nagata K, Basugi N, Fukushima T, et al. A quantitative study of physiological cerebral atrophy with aging: a statistical analysis of the normal range. Neuroradiology 1987; 29:327–332.
- Gyldensted C. Measurements of the normal ventricular system and hemispheric sulci of 100 adults with computed tomography. Neuroradiology 1977; 14:183-192.
- Damasio H, Eslinger P, Damasio AR, et al. Quantitative computed tomographic analysis in the diagnosis of dementia. Arch Neurol 1983; 40:715-719.
- Yamaura H, Ito M, Kubota K, et al. Brain atrophy during aging: a quantitative study with computed tomography. J Geront Aging 1980; 35:492-498.
   Schwartz M, Creasey H, Grady CL, et al.
- Schwartz M, Creasey H, Grady CL, et al. Computed tomographic analysis of brain morphometrics in 30 healthy men, aged 21 to 81 years. Ann Neurol 1985; 17(2):146-157.
- Hubbard BM, Anderson JM. Age, senile dementia, and ventricular enlargement. J Neurol Neurosurg Psychiatry 1981; 44:631-635.
- Morel J, Wildi E. General and cellular pathochemistry of senile and presenile alterations of the brain. In: Proceedings of the First International Congress of Neuropathology. Vol. 2. Rome, 1952; 347–374.
- Koller WC, Glatt SL, Fox JH, et al. Cerebellar atrophy: relationship to aging and cerebral atrophy. Neurology 1981; 31:1486-1488.
- 40. LeMay M. Morphological cerebral asymmetries of modern man, fossil man, and nonhuman primate. Ann NY Acad Sci 1976; 280:349-366.
- 41. Borgersen D. Width of the third ventricle. Acta Radiol (Stockh) 1966; 4:645-661.
- Messert B, Wannamaker BB, Dudley AW Jr. Reevaluation of the size of the lateral ventricles of the brain: postmortem study of an adult population. Neurology 1972; 22:941-951.

- Hooper MW, Vogel FS. The limbic system in Alzheimer's disease. Am J Pathol 1976; 85:1-13.
- Kido DK, Lemay M, Levinson AW, et al. Computed tomographic localization of the precentral gyrus. Radiology 1980; 135:373-377.
- Valentine AR, Moseley IF, Kendall BE. White matter abnormality in cerebral atrophy: clinicoradiological correlations. J Neurol Neurosurg Psychiatry 1980; 43:139-142.
- 46. Bradley WG, Waluch V, Brant-Zawadzki M, et al. Patchy, periventricular white matter lesions in the elderly: a common observation during NMR imaging. Noninvasive Med Imaging 1984; 1(1):35-41.
- Brant-Zawadzki M, Fein G, Van Dyke C, et al. MR imaging of the aging brain: patchy white-matter lesions and dementia. AJNR 1985; 6:675–682.
- Fazekas F, Chawluk JB, Alavi A, et al. MR signal abnormalities at 1.5 T in Alzheimer's dementia and normal aging. AJNR 1987; 8:421-426.
- George AE, de Leon MJ, Gentes CI, et al. Leukoencephalopathy in normal and pathologic aging. I. CT of brain lucencies. AJNR 1986; 7:561-566.
- George AE, de Leon MJ, Kalnin A, et al. Leukoencephalopathy in normal and pathologic aging. II. MRI of brain lucencies. AJNR 1986; 7:567–570.
- 51. Goto K, Ishii N, Fukasawa H. Diffuse white-matter disease in the geriatric population. Radiology 1981; 141:687-695.
- Tomlinson BE, Blessed G, Roth M. Observations on the brains of demented old people. J Neurol Sci 1970; 11:205-242.
- Zatz LM, Jernigan TL, Ahumada AJ Jr. White matter changes in cerebral computed tomography related to aging. J Comput Assist Tomogr 1982; 6:19-23.
- Gerard G, Weisberg L. MRI periventricular lesions in adults. Neurology 1986; 36:998-1001.
- Blessed G, Tomlinson BE, Roth M. The association between quantitative measures of dementia and of senile change in the cerebral grey matter of elderly subjects. Br J Psychiatry 1968; 114:797-811.
- Brun A, Englund E. A white matter disorder in dementia of the Alzheimer type: a pathoanatomical study. Ann Neurol 1986; 19:253-262.
- Awad IA, Johnson PC, Spetzler RF, et al. Incidental subcortical lesions identified on magnetic resonance imaging in the elderly. II. Postmortem pathological correlations. Stroke 1986; 17(6):1090-1097.
- Awad IA, Spetzler RF, Hodak JA, et al. Incidental subcortical lesions identified on magnetic resonance imaging in the elderly. I. Correlation with age and cerebrovascular risk factors. Stroke 1986; 17(6):1084-1089.
- Kirkpatrick JB, Heyman LA. White-matter lesions in MR imaging of clinically healthy brains of elderly subjects: possible pathologic basis. Radiology 1987; 162:509-511.
- 60. Brust JCM. Vascular dementia: still overdiagnosed. Stroke 1983; 140:298-300.
- 61. Fisher CM. Lacunar strokes and infarcts: a review. Neurology 1982; 32:871-876.
- 62. Durand-Fardel M. Traite du ramollissement du cerveau. Paris: Bailliere, 1843.
- Escourolle R, Poirier J. Other cerebrovascular lesions of ischemic nature. In: Escourolle R, Poirier J, eds. Manual of basic neuropathology. Philadelphia: Saunders, 1978; 101–103.
- 64. Fisher CM. Lacunes: small, deep cere-

bral infarcts. Neurology 1965; 15:774-784.

- 65. Ferrand J. Essai sur phemiplegie des vieillards: les lacunes de desintegration cerebrale. Paris: Thesis, 1902.
- 66. Cervos-Navarro J, Rozas IJ. The arteriole as a site of metabolic exchange. Adv Neurol 1978; 20:17-20.
- 67. Liston EH, La Rue A. Clinical differentiation of primary degenerative and multiinfarct dementia: a critical review of the evidence. I. Clinical studies. Biol Psychiatry 1983; 18(12):1451-1465.
- Liston EH, La Rue A. Clinical differentiation of primary degenerative and multiinfarct dementia: a critical review of the evidence. II. Pathological studies. Biol Psychiatry 1983; 18(12):1467-1483.
- Binswanger O. Die abgrenzung der algemeinen progressiven paralyse (referat, erstattet auf der jahresversammlung des vereins deutscher irrenarzte zu dresden am 20 Sept. 1894). Berl Klin Wochenschr 1894; 31:1103-1105, 1137-1139, 1180-1186.
- Caplan LR, Schoene WC. Clinical features of subcortical arteriosclerosis encephalopathy (Binswanger disease). Neurology 1978; 28:1206-1215.
- DeReuck J, Crevits L, DeCoster W, et al. Pathogenesis of Binswanger chronic progressive subcortical encephalopathy. Neurology 1980; 30:920-928.
- Kinkel WR, Jacobs L, Polachini I. Subcortical arteriosclerotic encephalopathy (Binswanger's disease): computed tomographic, nuclear magnetic resonance, and clinical correlations. Arch Neurol 1985; 42:951–959.
- Olzewski J. Subcortical arteriosclerotic encephalopathy: review of the literature on the so-called Binswanger's disease and presentation of two cases. World Neurol 1962; 3:359-375.
- Hachinski VC, Lassen NA, Marshall J. Multi-infarct dementia: a cause of mental deterioration in the elderly. Lancet 1974; 2:207-210.
- Rosen WG, Terry RD, Fuld PA, et al. Pathological verification of ischemic score in differentiation of dementias. Ann Neurol 1980; 7:486-488.
- Burger PC, Burch JG, Kunze U. Subcortical arteriosclerotic encephalopathy (Binswanger's disease): a vascular etiology of dementia. Stroke 1976; 7:626-631.
- Lewis AJ, Zingg W. Experimental brain damage in dogs due to systemic, induced hypotension and head-up tilt for short periods. Angiology 1966; 17:800-818.
- 78. Symon L, Pasztor E, Dorsch NWC, et al. Physiological responses of local areas of the cerebral circulation in experimental primates determined by the method of hydrogen clearance. Stroke 1973; 4:632– 642.
- De Reuck J. The cortico-subcortical arterial angioarchitecture in the human brain. Acta Neurol Belg 1972; 72:323-329.
- De Reuck J. The human periventricular arterial blood supply and the anatomy of cerebral infarctions. Eur Neurol 1971; 5:321-334.
- 81. De Reuck J, vander Eecken H. The arterial angioarchitecture in lacunar state. Acta Neurol Belg 1976; 76:142-149.
- Takashima S, Armstrong D, Becker LE. Subcortical leukomalacia: relation to development of the cerebral sulcus and its vascular supply. Arch Neurol 1978; 35:470-472.
- Ginsberg MD, Myers RE, McDonagh BF. Experimental carbon monoxide encephalopathy in the primate. II. Clinical as-

pects, neuropathology and physiological correlation. Arch Neurol 1974; 30:209-216.

- Ginsberg MD, Hedley-White TE, Richardson EP. Hypoxic-ischemic leukoencephalopathy in man. Arch Neurol 1976; 33:5-14.
- Feigin I, Popoff N. Neuropathological changes late in cerebral edema: the rela- tionship to trauma, hypertensive disease, and Binswanger's encephalopathy. J Neur-opathol Exp Neurol 1963; 22:500-511.
- Feigin I, Budzilovich C, Weinberg S, et al. Degeneration of white matter in hypoxia, acidosis, and edema. J Neuropath Exp Neurol 1973; 32:125–143.
- Van den Bergh R, vander Eecken H. Anatomy and embroyology of the cerebral circulation. Prog Brain Res 1968; 30:1-25.
- Hallgren B, Sourander P. The effect of age on the non-haemin iron in the human brain. J Neurochem 1958; 3:41-51.
- 89. Cumings JN. The copper and "inorganic" iron content of human tissues. Biochem J 1935; 29:480-486.
- 90. Drayer B, Burger P, Darwin R, et al. Magnetic resonance imaging of brain iron. AJNR 1986; 7:373-380.
- Drayer B, Olanow W, Burger P, et al. Parkinson plus syndrome: diagnosis using high field MR imaging of brain iron. Radiology 1986; 159(2):493-498.
- Diezel PB. Iron in the brain: a chemical and histochemical examination. In: Waelsch H, ed. Biochemistry of the developing nervous system. New York: Academic Press, 1954; 145–152.
- Spatz H. Uber den eisennachweis im gehirn, besonders in zentren des extrapyramidal-motorischen systems. Zentralbl Gesamte Neurol Psychiatr 1922; 77:261-390.
- 94. Hock A, Demmel U, Schicha H, et al. Trace element concentration in human brain: activation analysis of cobalt, iron, rubidium, selenium, zinc, chromium, silver, cesium, antimony, and scandium. Brain 1975; 98:49–64.
- 95. Swaiman KF, Machen VL. Iron uptake by cortical neurons. Ann Neurol 1984; 16:66–70.
- Rafaelson OJ, Kofod B. Iron. In: Lajtha A, ed. Handbook of neurochemistry. New York: Plenum, 1969; 261-271.
- 97. Richter GW. The iron-loaded cell-the cytopathology of iron storage: a review. Am J Pathol 1978, 91:361-404.
- Mulligan M, Linder M. The size of small molecular weight iron pools in rat tissues. In: Saltman P, Hagenauer J, eds. The biochemistry and physiology of iron. New York: Elsevier Biomedical, 1982; 313–314.
- 99. Youdim MBH, Green AR, Bloomfield MR, et al. The effects of iron deficiency on brain biogenic monoamine biochemistry and function in rats. Neuropharmacology 1980; 19:259-267.
- Sourkes TL. Transition elements and the nervous system. In: Pollitt E, Leibel RL, eds. Iron deficiency: brain biochemistry and behavior. New York: Raven, 1982; 1-29.
- 101. Park BE, Netsky MG, Betsill W Jr. Pathogenesis of pigment and spheroid formation in Hallervorden-Spatz syndrome and related disorders: peroxidation as a common mechanism. Neurology 1975: 25:1171-1178
- ogy 1975; 25:1171-1178. 102. Hill JM, Switzer RC. The regional distribution and cellular localization of iron in the rat brain. Neuroscience 1984; 3(11):595-603.

- Francois C, Nguyen-Legros J, Percheron G. Topographical and cytological localization of iron in rat and monkey brains. Brain Res 1981; 215:317–322.
- 104. Jefferies WA, Brandon MR, Hunt SV, et al. Transferrin receptor on endothelium of brain capillaries. Nature 1984; 312:162–163.
- 105. Koenig SH, Baglin CM, Brown RD III. Magnetic field dependence of solvent proton relaxation in aqueous solutions of Fe<sup>3+</sup> complexes. Magn Reson Med 1985; 2:283-288.
- Brittenham GM, Farrell DE, Harris JW, et al. Magnetic-susceptibility measurements of human iron stores. N Engl J Med 1982; 307:1671-1675.
- Hallgren B, Sourander P. The non-haemin iron in the cerebral cortex in Alzheimer's disease. J Neurochem 1960; 5:307-310.
- 108. Earle KM. Studies of Parkinson's disease, including x-ray fluorescent spectroscopy of formalin fixed brain tissue. J Neuropath Exp Neurol 1968; 27:1-14.
- 109. Borit A, Rubinstein LJ, Urich H. The striatonigral degenerations: putaminal pigments and nosology. Brain 1975; 98:101-112.
- Tygstrup I, Norholm T. Neuropathological findings in 12 patients operated for parkinsonism. Acta Neurol Scand 1963; 4(suppl.):188-195.
- 111. Goodman L. Alzheimer's disease: a clinico-pathologic analysis of twenty-three cases with a theory on pathogenesis. J Nerv Ment Dis 1953; 117:97–130.
- 112. Drayer BP. Neurometabolic applications of magnetic resonance. In: American College of Radiology categorical course on magnetic resonance (syllabus). Bethesda, Md.: ACR, 1985; 185-211.
- 113. Gutteridge JMC, Westermarck T, Santavuori P. Iron and oxygen radicals in tissue damage: implications for the neuronal ceroid lipofuscinoses. Acta Neurol Scand 1983; 68:365–370.
- 114. Harman D. Aging: a theory based on free radical and radiation chemistry. J Gerontol 1956; 11:298-300.
- Sylvia AL, Rosenthal M. Effects of age on brain oxidative metabolism in vivo. Brain Res 1979; 165:235-248.
- 116. Siesjo BK, Rehncrona S, Smith D. Neuronal cell damage in the brain: possible involvement of oxidative mechanisms. Acta Physiol Scand 1980; 492(suppl.): 121-128.
- 117. Floyd RA, Zaleska MM, Harmon HJ. Possible involvement of iron and oxygen free radicals in aspects of aging in brain. In: Armstrong D, Sohal RS, Cutler RG, Slater TS, eds. Free radicals in molecular biology, aging, and disease. New York: Raven, 1984; 143-161.
- Wong DF, Wagner HN Jr, Dannals RF, et al. Effects of age on dopamine and serotonin receptors measured by positron tomography in the living human brain. Science 1984; 226:1393–1396.
- 119. Carlsson A, Adolfsson R, Aquilonius SM, et al. Biogenic amines in human brain in normal aging, senile dementia, and chronic alcoholism. In: Goldstein M, Calne DB, Lieberman A, Thorner MO, eds. Ergot compounds and brain function: neuroendocrine and neuropsychiatric aspects. Advances in biochemical psychopharmacology, vol. 23. New York: Raven, 1980; 295-304.
- 120. McGeer F, McGeer PL. Neurotransmitter metabolism in the aging brain. In: Terry RD, Gershon S, eds. Neurobiology of aging. Aging, vol. 3. New York: Ra-

ven, 1976; 389-403.

- 121. Bugiani O, Salvarani S, Perdelli F, et al. Nerve cell loss with aging in the putamen. Eur Neurol 1978; 17:286-291.
- 122. Moossy J. Cerebral atherosclerosis: intracranial and extracranial lesions. In: Minckler J, ed. Pathology of the nervous system. Vol. 2. New York: McGraw-Hill, 1971; 1423–1432.
- 123. Jorgensen L, Torvik A. Ischaemic cerebrovascular diseases in an autopsy series. I. Prevalence, location and predisposing factors in verified thromboembolic occlusions, and their significance in the pathogenesis of cerebral infarction. J Neurol Sci 1966; 3:490-509.
- Katzman R. Vascular disease and dementia. In: Yahr MD, ed. H. Houston Merrit memorial volume. New York: Raven, 1983; 153-176.
- 125. Librach G, Schadel M, Seltzer M, et al. Stroke: incidence and risk factors. Geriatrics 1977; 32:85–96.
- Shapiro W, Chawla N. Effects of pacing on cerebral blood flow and cardiac dynamics. Circ Suppl 1969; 39/40:184-187.
- 127. Sulg IA, Cronquist S, Schuller H, et al. The effect of intracardial pacemaker therapy on cerebral blood flow and electroencephalogram in patients with complete atrioventricular block. Circulation 1969; 39:487-494.
- Esiri MM, Wilcock GK. Cerebral amyloid angiopathy in dementia and old age. J Neurol Neurosurg Psychiatry 1986; 49:1221-1226.
- 129. Vinters HV, Gilbert JJ. Cerebral amyloid angiopathy: incidence and complications in the aging brain. II. The distribution of amyloid vascular changes. Stroke 1983; 14:924–928.
- Cosgrove GR, Leblanc R, Meagher-Villemure K, et al. Cerebral amyloid angiography. Neurology 1985; 35:625-631.
- 131. Mountjoy CQ, Tomlinson BE, Gibson PH. Amyloid and senile plaques and cerebral blood vessels: a semi-quantitative investigation of a possible relationship. J Neurol Sci 1982; 57:89–103.
- 132. Okazaki H, Reagan TJ, Campbell RJ. Clinicopathologic studies of primary cerebral amyloid angiopathy. Mayo Clin Proc 1979; 54:22-31.
- 133. Tomonaga M. Cerebral amyloid angiopathy in the elderly. J Ann Geriatr Soc 1981; 29:151-158.
- Ishii N, Nishihara Y, Horie A. Amyloid angiopathy and lobar cerebral haemorrhage. J Neurol Neurosurg Psychiatry 1984; 47:1203-1210.
- Jellinger K. Cerebrovascular amyloidosis with cerebral hemorrhage. J Neurol 1977; 214:195-206.
- Lee SS, Stemmerman GN. Congophilic angiopathy and cerebral hemorrhage. Arch Pathol Lab Med 1978; 102:317-321.
- 137. Gilbert JJ, Vinters HV. Amyloid angiopathy: incidence and complications in the aging brain. I. Cerebral hemorrhage. Stroke 1983; 14:915-923.
- Smith CB. Aging and changes in cerebral energy metabolism. Trends Neurosci 1984; 7:203-208.
- 139. Arnold KG. Cerebral blood flow in geriatrics: a review. Age Aging 1981; 10:5-9.
- 140. Kety SS. Human cerebral blood flow and oxygen consumption as related to aging. Res Publ Assoc Res Nerv Ment Dis 1956; 35:31-45.
- 141. Klee A. The relationship between clinical evaluation of mental deterioration, psychological test results, and the cerebral metabolic rate of oxygen. Acta Neurol Scand 1964; 40:337–345.

- 142. Melamed E, Lavy S, Bentin S, et al. Reduction in regional cerebral blood flow during normal aging in man. Stroke 1980; 11:31–36.
- 143. Shaw TG, Mortel KF, Meyer JS, et al. Cerebral blood flow changes in benign aging and cerebrovascular disease. Neurology 1984; 34:855-862.
- Sokoloff L. Cerebrovascular disease. Vol. XLI. Baltimore: Williams & Wilkins, 1966; 237-254.
- 145. Ordy JM, Brizzee KR. Functional and structural age differences in the visual system of man and nonhuman primate models. In: Ordy JM, Brizzee KR, eds. Sensory systems and communication in the elderly. Aging, vol. 10. New York: Raven, 1979; 13-50.
- 146. Ordy JM, Brizzee KR, Beavers T, et al. Age differences in the functional and structural organization of the auditory system in man. In: Ordy JM, Brizzee KR, eds: Sensory systems and communication in the elderly. Aging vol. 10. New York: Raven, 1979; 153-166.
- Devaney KO, Johnson HA. Neuron loss in the aging visual cortex of man. J Gerontol 1980; 35:836-841.
- 148. Duara R, Margolin RA, Robertson-Tchabo EA, et al. Cerebral glucose utilization as measured with positron emission tomography in 21 resting healthy men between the ages of 21 and 83 years. Brain 1983; 106:761-775.
- 149. Mazziotta JC, Phelps ME, Miller J, et al. Tomographic mapping of human cerebral metabolism: normal unstimulated state. Neurology 1981; 31:503–516.
- 150. Gur RC, Packer IK, Hungerbuhler JP, et al. Differences in the distribution of gray and white matter in human cerebral

hemispheres. Science 1980; 207:1226-1228.

- Grubb RL Jr, Raichle ME, Gado MH, et al. Cerebral blood flow, oxygen utilization, and blood volume in dementia. Neurology 1977; 27:905-910.
- 152. Huang SC, Phelps ME, Hoffman EJ, et al. Noninvasive determination of local cerebral metabolic rate of glucose in man. Am J Physiol 1980; 238:E69–82.
- 153. Frackowiak RSJ, Lenzi GL, Jones T, et al. Quantitative measurement of regional cerebral blood flow and oxygen metabolism in man using <sup>15</sup>O and positron emission tomography: theory, procedure and normal values. J Comput Assist Tomogr 1980; 4:727-736.
- 154. Frackowiak RSJ, Gibbs JM. The pathophysiology of Alzheimer's disease studied with positron emission tomography. In: Katzman R, ed. Biological aspects of Alzheimer's disease. Brandury report 15. Cold Spring Harbor, N.Y.: Cold Spring Harbor Laboratory, 1983; 317-324.
- 155. De Leon MJ, Ferris SH, George AE, et al. Positron emission tomographic studies of aging and Alzheimer disease. AJNR 1983; 4:568–571.
- Drayer BP. Functional applications of computed tomography of the central nervous system. AJNR 1981; 2:495-510.
- 157. Dastur DK, Lane MH, Hansen DB, et al. Effects of aging on cerebral circulation and metabolism in man. In: Birren JE, Butler RN, Greenhouse SW, et al., eds. Human aging: a biological and behavioral study. Washington, D.C.: Government Printing Office, 1963; 59-76.
- 158. Kuhl DE, Metter EJ, Riege WH, et al. Effects of human aging on patterns of local cerebral glucose utilization determined by the [<sup>18</sup>F] fluorodeoxyglucose method. J Cereb Blood Flow Metab 1982; 2:163–171.

- 159. Obrist WD. Problems of aging. In: Chatrian GE, Lairy GC, eds. Handbook of electroencephalography and clinical neurophysiology. Vol. 6. Amsterdam: Elsevier, 1976; 6A274-6A292.
- 160. NIA Task Force. Senility reconsidered. JAMA 1962; 244:259-263.
- 161. Stigsby B, Johannesson G, Ingvar DH. Regional EEG analysis and regional cerebral blood flow in Alzheimer's and Pick's disease. Electroencephalogr Clin Neurophysiol 1981; 51:537–547.
- 162. Smith CB, Goochee C, Rapoport SI, et al. Effects of ageing on local rates of cerebral glucose utilization in the rat. Brain 1980; 103:351–365.
- 163. Rapaport SI, Duara R, Horowitz B, et al. Brain aging in 40 healthy men: rCMR-glc and correlated functional activity in various regions in the resting state. J Cerebr Blood Flow Metab 1983; 3(suppl. 1):5484.
- Samorajski T. Central neurotransmitter substances and aging: a review. J Am Geriatr Soc 1977; 25:337–348.
- Bartus RT, Dean RL III, Beer B, et al. The cholinergic hypothesis of geriatric memory dysfunction. Science 1982; 217:408– 417.
- 166. Davies P, Verth AH. Regional distribution of muscarinic acetylcholine receptor in normal and Alzheimer's type dementia brains. Brain Res 1978; 138:385-392.
- 167. Perry EK, Blessed G, Tomlinson BE, et al. Neurochemical activities in human temporal lobe related to aging and Alzheimer-type changes. Neurobiol Aging 1981; 2:251–256.