



HHS Public Access

Author manuscript

Nat Rev Neurosci. Author manuscript; available in PMC 2018 August 16.

Published in final edited form as:

Nat Rev Neurosci. 2018 February 16; 19(3): 123–137. doi:10.1038/nrn.2018.1.

Imaging structural and functional brain development in early childhood

John H. Gilmore¹, Rebecca Knickmeyer Santelli¹, and Wei Gao²

¹Department of Psychiatry, CB# 7160, University of North Carolina School of Medicine, Chapel Hill, North Carolina 27599-7160, USA

²Biomedical Imaging Research Institute, Department of Biomedical Sciences and Imaging, Cedars-Sinai Medical Center, 116 N. Robertson Blvd. Ste. 400. Los Angeles, California 90048 USA

Abstract

In humans, the period from term birth to ~2 years of age is characterized by rapid and dynamic brain development and plays an important role in cognitive development and risk for disorders such as autism and schizophrenia. Recent imaging studies have begun to delineate the growth trajectories of brain structure and function in the first years after birth and their relationship to cognition and risk for neuropsychiatric disorders. This Review discusses the development of grey and white matter, structural and functional networks, as well as genetic and environmental influences on early childhood brain development. We also discuss initial evidence regarding the usefulness of early imaging biomarkers for predicting cognitive outcomes and risk for neuropsychiatric disorders.

Introduction

Early childhood — especially the period between term birth and about ~2 years of age — is increasingly recognized as being very important for establishing cognitive abilities and behaviours that last a lifetime¹, as well as for risk for neuropsychiatric disorders such as autism and schizophrenia². In spite of its importance, little is known about structural and functional brain development during this critical period, in part owing to difficulties with image acquisition and image analysis in very young children. Recently, however, imaging studies in early childhood have begun to illuminate the extremely rapid pace of postnatal structural and functional brain development^{3–6}.

Correspondence to J.H.G. john_gilmore@med.unc.edu.

Author contributions

The authors all researched data for the article, provided a substantial contribution to discussion of the content, wrote the article and reviewed and edited the manuscript before submission.

Competing interest statement

The authors declare no competing interests.

Further information

FNIH Baby Connectome Project: <https://fnih.org/what-we-do/current-research-programs/baby-connectome>
ERC Developing Human Connectome Project: <http://www.developingconnectome.org/>

These studies, along with more recent postmortem histological and gene-expression studies^{7, 8} (Box 1), suggest that the basic structural and functional framework of the human brain is in place by the second year of life and perhaps even earlier. Brain development after age 2 years is characterized mainly by reorganization, ‘fine-tuning’, plasticity and remodelling of the major circuits and networks that are already established. Despite the challenges associated with imaging young brains (Box 2), these studies are also beginning to provide the basis for a better understanding of how the different components of structural and functional development relate to one another in this period of rapid growth⁹. They suggest a model in which white-matter tracts and networks are largely in place at birth, and resting-state functional networks develop rapidly after birth, along with rapid myelination and maturation of existing white-matter connections. Cortical and subcortical grey matter also undergoes robust growth in the first year of life¹⁰, although cortical grey-matter networks seem to mature much later in childhood, perhaps driven by coordinated activity across the cortex from functional networks that arise much earlier in development.

Box 1

Human neurodevelopment at the cellular level

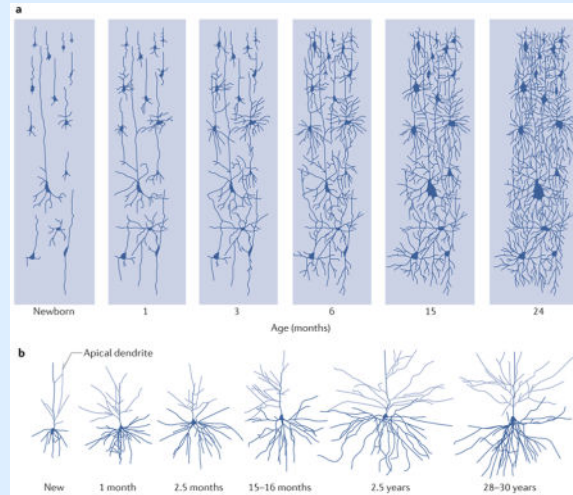
There are very few postmortem histologic studies of early childhood cortical development; however, the studies that do exist are consistent with imaging studies and provide insights into the neurobiological process that underlie change observed in imaging studies.

Grey matter

The bulk of neurons in the human brain are generated prenatally²⁰⁴. However, recent studies find some neurogenesis occurs after birth: one study showed that neurons continue to arise and migrate from the subventricular zone of the anterior lateral ventricles to the olfactory bulb and ventromedial prefrontal cortex in the first 18 months of postnatal life²⁰⁵. Another study found widespread migration of new neurons, mainly inhibitory interneurons, to the cingulate cortex and superior frontal gyrus in the first 5 months after birth²⁰⁶. These studies suggest that neurogenesis in early childhood contributes to grey-matter expansion in at least some areas of the frontal lobe.

The complexity of cortical neurons increases rapidly in the first years after birth, mirroring imaging findings of rapid cortical thickness growth and surface area expansion. Conel’s classic survey of cortical neuron development in children found that the overall complexity of cortical neurons increases quickly after birth, peaking at 2–4 years²⁰⁷ (see schematic in part a of the figure). Synapse numbers increase rapidly after birth and decrease in adolescence, with synapse numbers peaking in the auditory and visual sensory areas before those in the prefrontal cortex²⁰⁸, consistent with imaging findings that sensorimotor regions mature before higher-order association areas¹⁰. There are layer-specific differences in the developmental trajectories of dendritic complexity in the prefrontal cortex. For example, at birth layer V neurons have more complex basal dendritic trees which integrate synaptic inputs compared to layer IIIC neurons; layer V neurons reach maximum complexity at 16 to 30 months after birth, before layer IIIC²⁰⁹

(see layer III development in **b** of the figure²⁰⁹). Synaptic spine density in prefrontal cortex also peaks by 2–5 years and falls in adolescence⁷.



White matter

Postmortem studies of white-matter myelination in the human brain find that myelination begins prenatally and is present in many regions at the time of birth, especially primary sensory and motor pathways, including the optic radiations and internal capsule^{210, 211}. Postnatally, myelination follows a general pattern of sensory before motor regions, projection areas before associative areas, and posterior before anterior regions²¹¹. In the human hippocampus, myelination begins at 20 weeks gestational age, and by age 2 years, many regions have attained adult levels of myelination — although some areas, such as the hilus of the dentate, continue to myelinate throughout childhood into adulthood^{212–214}. In the neocortex, mature myelination patterns are attained only in adulthood, reflecting a protracted maturation of this brain region in humans compared with that in non-human primates²¹⁵. Oligodendrocytes are responsible for producing myelin, and the number of oligodendrocytes in cerebral white matter increases rapidly and linearly after birth, from approximately 7 billion at birth to 28 billion at age 3 years — a rate of 600 million per month²¹⁶. In the corpus callosum, oligodendrocyte numbers increase until age 5 years and are stable thereafter; a study of nuclear bomb test-derived ¹⁴C suggest that there is little turnover of oligodendrocytes after age 5 years²¹⁷.

Part **b** of the figure is adapted with permission from REF. ²⁰⁹.

Box 2

Challenges of structural imaging in early childhood

Studies of early childhood brain development have been limited by difficulties with image acquisition and analysis. Young children have a limited ability to cooperate with scanning procedures that require lying still and minimizing motion and are often scanned in a natural sleep, which is difficult to initiate and maintain in a noisy scanner environment. Moreover, the short image-acquisition protocols necessary in young

children tend to produce less-than-ideal tissue contrasts and image quality compared to those used in studies of adults. Image-analysis protocols developed for the adult brain often fail in infant brain owing to the latter's smaller size, reduced tissue contrast between grey and white matter, and large changes in size and tissue contrasts with age. For example, the relative intensities of grey and white matter in T1- and T2-weighted imaging change significantly over the first year of life and are typically very similar between 4 to 8 months of age; this makes distinguishing grey and white matter difficult. . This changing contrast also presents challenges for determining cortical thickness, which depends on a clear boundary between cortical grey matter and underlying white matter.²⁸ Fortunately, improved image-acquisition technology, such as parallel imaging with multi-channel head coils, multi-band techniques that allow excitation of multiple slices, and scanners with higher gradients now provide improved resolution and signal in the shorter timeframes required for imaging infants. These technical advances, combined with innovative image-analysis programs that provide improved brain tissue segmentation and parcellation, make imaging studies of young children much more feasible and informative.

It is increasingly recognized that motion in the scanner can significantly influence measures of cortical thickness and surface area^{27, 218}, as well as DTI parameters (Box 2)²¹⁹ and resting-state networks (Box 3)^{220,219}. Motion tends to reduce estimates of grey-matter volume and cortical thickness²²¹ and can influence observed trajectories of cortical thickness maturation as well²⁷. Thus, care must be taken to assess and correct for motion, especially in childhood, when motion can be more of an issue, or when comparing groups of subjects when one group may be more likely to experience motion in the scanner. There are several approaches to detecting motion during scanning, and for prospective and retrospective motion correction; prospective motion correction typically requires multiple extra images, increasing the scan time^{222, 223}. It is also important to recognize that imaging measures are indirect measures of brain structure and function and are subject to the effect of many potential confounds, such as hydration, blood lipid levels and cortisol levels²²⁴.

Box 3

Techniques used to study white-matter development in infants

Various imaging approaches are used to study white matter at different developmental stages. Early studies tracked white-matter maturation and myelination using standard T1- and T2-weighted images and T1 and T2 relaxation times — the time water magnetization returns to normal after changes in the magnetic field^{5, 225}. Diffusion-weighted imaging is widely used to study white-matter composition and development and takes advantage of the motion of water in response to changes in the magnetic field²²⁶. Water motion is constrained by the complex structure of axons and myelin, allowing the visualization of major fibre tracts and the analysis of the microstructure of the fibres, typically expressed as axial diffusivity (AD), radial diffusivity (RD), and fractional anisotropy (FA).

More recent advances in diffusion-weighted imaging attempt to provide more information about the microstructure of white matter, including the presence of fibres that cross one another, using biophysical models; these require increased numbers of diffusion values and directions using approaches such as high angular resolution diffusion-weighted imaging (HARDI) and multi-shell diffusion-weighted MRI^{5, 223}. These approaches typically require longer acquisition times than standard sequences and are therefore a challenge in young children, although time-efficient modifications of these approaches have begun to be used^{227–231}. Another imaging approach used to study white matter development in young children is that of an acquisition-time-efficient multicomponent relaxometry — mcDESPOT^{231,232} (multicomponent driven equilibrium single pulse observation of T1 and T2), which determines the myelin water fraction. This method assesses white-matter myelination more directly than does traditional diffusion-weighted imaging, but is dependent on acquisition protocol and modelling assumptions^{5,233,5, 234}. The patterns of white-matter maturation described by diffusion tensor imaging and myelin water fraction studies are consistent with one another and with previous postmortem studies of myelination (Box 1). Finally, the magnetization transfer ratio, or the ratio between free and bound water, has been used to assess white-matter microstructure and myelination²³⁵, but has not been used extensively in young children⁵.

Each of these imaging modalities assess components of white-matter microstructure, such as axon thickness, oligodendrocyte proliferation and development, myelin deposition and axon retraction to various degrees, but each approach provides at best indirect assessments of the various components of white-matter microstructure. Moreover, diffusion-weighted imaging has many limitations and sources of error at each step of the analysis pipeline that must be considered when interpreting results^{236, 237}. Motion is also an important issue for white-matter imaging. Scanners with higher-strength gradients can considerably shorten acquisition times and decrease motion-related artefacts²³⁸. For diffusion-weighted imaging, motion can be corrected during acquisition or in post-processing in a variety of ways, although standards have not been established (see discussion in REF. ²³⁹).

In this Review, we describe recent imaging studies of early childhood and place them in the context of previous studies of brain development that typically begin at age 6 years. We review studies of grey- and white-matter development, as well as studies of functional and structural network development. Although the current state of knowledge about genetic and environmental influences on early childhood brain development and the usefulness of imaging biomarkers for understanding cognitive development and risk for psychiatric and neurodevelopmental disorders does not yet allow definitive conclusions in young children, here we cover early efforts to address these issues. This Review will not address preterm imaging studies, either of premature infants or of *in utero* prenatal brain development in detail.^{11, 12}

Structural development

Prior knowledge about structural brain development has been based mainly on older postmortem studies that are typically limited by sample size and cross sectional design. While imaging studies cannot provide information at a cellular level, they have significantly improved our understanding of early childhood structural brain development in humans and complement postmortem studies (Box 1).

Global tissue volumes and subcortical structures

Overall growth of the brain is very rapid in the first years of life (Fig. 1, 2). Brain volume is about 35% of adult volume 2–3 weeks after birth after birth¹³, doubles from term size in the first year of life and increases an additional 15% in the second to about 80% of adult size³. After age 2, there is a more gradual increase in volume, consistent with previous postmortem studies of brain weight^{14,15}. Grey and white matter have different growth trajectories soon after birth, with rapid development of grey-matter volume compared to white-matter volume, which increases much more gradually^{15,3}. From birth to age 1, cortical grey-matter volume increases 108–149%, whereas white matter increases by about 11%; from age 1 to age 2 years, grey matter increases 14–19%, whereas white matter increases by 19%³. Postnatal growth of global tissue volume and subcortical structures is most rapid in the first 3 months after birth¹⁶. After age 2, total grey-matter volume shows minimal absolute increases through childhood and decreases in adolescence, whereas white-matter volumes steadily increase through early adulthood, peaking around age 30.^{17,18}

The growth rates of subcortical grey-matter structures are similar to cortical grey-matter growth rates, with the amygdala, thalamus, caudate, putamen, and pallidum growing about 105% in the first year and roughly 15% in the second. The hippocampus grows somewhat slower (about 84%) than other subcortical structures in year one¹⁰. Lateral ventricle volume increases by over 100% in year 1, but subsequently decreases by 7–24% in the second year of life^{3, 10, 19}.

Cortical thickness, surface area and gyrification

Structural imaging of cortical development typically assesses two major components of cortical volume — cortical thickness and surface area— as well as indexes of cortical folding (gyrification). In contrast to the rather gradual changes in cortical thickness and surface area observed in later childhood and adolescence²⁰, these measures grow dramatically in the first 2 years of postnatal life.

Imaging studies of infants and older children^{21, 20, 22} have revealed that cortical thickness probably peaks in the range of 1–2 years and declines thereafter, whereas surface area continues to expand well into late childhood or early adolescence. The limited postmortem studies of cortical neuron development in humans are consistent with the temporal and regional patterns of grey-matter growth derived from imaging studies (Box 1).

Overall, the major patterns of gyrification of the newborn brain is very similar to the adult brain. The gyrification index (the amount of cortex within a sulcal fold), increases in the first 2 years of life in parallel with overall brain growth²³. Cortical surface area expands by 76%

from birth in year 1 and another 22% in year 2; by age 2 years, the mean surface area is about 69% of adult values^{4,23}. Surface area expansion is also regionally heterogeneous across the brain: the lateral frontal, lateral parietal and occipital cortices grow relatively rapidly, whereas regions in the orbital frontal, insula and occipital cortices tend to grow more slowly (Fig. 1). This pattern is generally consistent with an imaging study that compared surface area between infants and adults²⁴. Surface area expands linearly from age 3 until about 8–12 years and then declines gradually^{20, 25}.

Average cortical thickness increases by an average of 31% in year 1 and an additional 4.3% in the second year; by age 2, the mean cortical thickness is about 97% of the values observed in adults⁴. Regional heterogeneity of cortical thickness observed in adults is well established at birth and persists through age 2 years into adulthood^{4,26}. Regions of faster growth of cortical thickness after birth include speech and language regions [Heschel's gyrus, Rolandic operulum], the insula and cingulate cortex as well as some higher association areas. The primary and secondary sensory cortices tend to grow more slowly. Whereas some studies have suggested that cortical thickness increases from age 6 to 10 years and begins to thin after 10–12 years^{21,20}, most studies find that cortical thickness decreases linearly after age 4–5 years^{25,27,28}. Indeed, a recent study reported that cortical thickness might gradually decrease even between ages 1 and 6 years²².

White matter

White-matter volume increases through evolution²⁹ as longer white-matter fibres are needed to connect neurons in larger brains and the volume of white matter increases faster than the volume of grey matter^{30,29}. Diffusion imaging studies find that major white matter tracts such as the corpus callosum, superior and inferior longitudinal fasciculus, arcuate, and cingulum are present at the time of birth, indicating that most of the 'wiring' of the brain is established during prenatal brain development³¹ (Fig. 1). As noted above, white-matter volume growth is more slow and protracted compared to grey matter volume (Figs 1, 2).

In contrast to volume growth, the maturation of white-matter regions and tracts has been revealed by quantitative diffusion tensor imaging (DTI) studies to be similar to that observed for grey matter regions, with rapid rates of change in the first year of life, and a more gradual increase thereafter (Box 3; reviewed elsewhere⁵). With postnatal development, measures of white-matter microstructure and maturation, including axial diffusivity [G] (AD) and radial diffusivity [G] (RD), decrease, whereas fractional anisotropy [G] (FA) increases, thought to reflect the increasing organization of the already established white matter tracts, including myelination^{5,32}. For example, FA increases 9–44% in the first year, with most tracts increasing by more than 25%; in the second year, FA increases 5–9% over levels at age 1 year. Using multicomponent relaxometry, an alternative approach to the study of white matter (Box 3), myelination in the first year of life was shown to begin in the cerebellum, pons and internal capsule, and to proceed in a 'back-to-front' pattern, from the splenium of the corpus callosum and optic radiations (3–4 months), to the occipital and parietal lobes (4–6 months), and anteriorly to the frontal and temporal lobes (6–8 months)³³. Diffusion values change much more gradually through later childhood and adolescence, with AD and RD decreasing and FA increasing.^{34,35}; similarly, myelination

increases more gradually after the second year^{36,37}. The diffusion properties of white-matter tracts are highly correlated with each other at birth, but this correlation decreases with development as tracts mature and differentiate³⁸.

Networks

Beyond individual white-matter tracts and regional cortical thickness and surface area, there is also a growing interest in characterizing the growth of structural and functional networks during early brain development.

In the adult brain, graph theory-based approaches have been increasingly applied to quantify different information-transfer properties of structural and functional whole-brain networks³⁹. Among these properties, indices of local efficiency and global efficiency characterize the ease of information flow at the local neighborhood and global system levels, respectively; systems that possess both high local and global efficiencies are designated as having ‘small-world’ properties⁴⁰. Moreover, there is also interest in detecting the ‘hubs’ of the brain that serve as connecting centres between different parts of the system⁴⁰. Finally, the human brain also exhibits a ‘rich club’ phenomenon, in that network hubs also tend to be better-connected to each other⁴¹.

Leveraging these conceptual and methodological advances, there has been a tremendous progress in understanding brain-network development during the first years of life^{42–44}. Although the structural and functional connectomes of the brain have been linked to cognitive function and disease, little is known about the biology that underlies them. Recently, it has been found that rich-club regions of the functional connectome organize functional networks through state-dependent oscillatory activity⁴⁵, and that neuronal network behaviour can be used to predict neuron function in *Caenorhabditis elegans*⁴⁶, providing interesting first clues about the neurobiology of large scale networks.

Structural networks

White-matter networks in adults are characterized by global and local efficiency, small-world and modular structures that have a ‘rich-club’ of modules that are highly connected, and structural alterations in neuropsychiatric disease. Recent studies indicate that most of these white-matter network attributes are present at, and even before, birth (reviewed elsewhere⁴⁷). Overall, the white-matter connectome at birth is highly organized and includes hub regions and connections that are similar to those observed in adults^{48,49}. White-matter networks at birth, and even in premature infants at 30 weeks gestational age⁵⁰, have a rich club property similar to that in adults. Network maturation in the first 2 years after birth generally consists of increased network efficiency and network integration [G], decreased network segregation [G], and some changes in the modularity^{48,51}, with most major hubs and modules in place by age 2 years⁵².

Cortical grey-matter structural networks consist of regions of highly correlated variation in of grey-matter volume or cortical thickness^{53,54}. These structural covariance networks [G] (SCNs) are heritable⁵⁵, altered in neuropsychiatric disease⁵⁶ and have been related to intelligence⁵⁷. In adults, SCNs have small-world and modular properties; these SCNs

overlap with white-matter and functional connectivity networks to some extent, although the biological foundation for these SCNs is not clear^{53, 54}. Childhood development of SCNs is characterized by increasing global efficiency, decreasing local efficiency and changes in modularity. For example, at a network level, SCNs of grey-matter volume and cortical thickness exhibit small-world properties and modular organization at birth⁵⁸ and 3 years⁵⁹. Primary sensorimotor SCNs are well developed by birth, and higher-order association SCNs mature and become increasingly distributed, involving more regions from age 5 to 18 years.⁶⁰ Longitudinal studies indicate that networks of similarities in regional maturation rates of cortical thickness are similar to SCNs, indicating that, in between 9 and 22 years, structural covariance is the result of coordinated maturation⁵⁴. By contrast, in the first 2 years of life, SCNs and maturational networks [G] are not clearly related, suggesting that the fine tuning of cortical SCNs, especially high-order SCNs, occurs later in childhood⁶¹. More research is needed to fully understand how SCNs are related to white-matter and functional networks.

Functional networks

The development of the function of the human brain begins prenatally^{62, 63}. Electroencephalography (EEG) studies have documented intermittent bursts of electrical activity and synchronized oscillatory activity in premature infants^{64–66}. This early activity is believed to be crucial for the emergence and fine tuning of primary functional circuits during fetal development when sensory experience is limited the developmental period⁶². This patterned electrical activity in the brain is mostly elicited by spontaneous activity in sensory organs, such as spontaneous retinal waves^{67, 68}, spontaneous cochlear activity⁶⁹ and muscle twitching⁷⁰.

Non-invasive neuroimaging techniques such as functional MRI (fMRI)^{71, 72} and functional near-infrared spectroscopy (fNIRS)⁷³ provide functional information that is complementary to EEG (Box 4), and enable the study of the postnatal development of brain's functional architecture. For example, task-based fMRI and fNIRS studies using passive stimuli have revealed adult-like positive blood-oxygen-level-dependent (BOLD) signals in response to sensorimotor^{74, 75}, visual⁷⁶, and auditory stimuli^{77, 78} in human newborns and infants. fNIRS studies have documented the emergence and gradual improvement of various other functional responses during spatiotemporal processing⁷⁹, object recognition⁸⁰, learning⁸¹ and social processing⁸², among other processes, during the first year of life. Nevertheless, non-invasive brain imaging of awake infants remains challenging owing to their motion and their inability to perform certain tasks, which limit such studies to a restricted set of brain functions. Moreover, the neurovascular coupling mechanism is known to be immature during this stage⁸³ since key components enabling such coupling, including neurons, astrocytes, pericytes, and the brain's vasculature, are still actively developing during infancy. The best approaches to model infant BOLD-based activation detection is still an active area of research⁷⁴ (Box 4).

Box 4**Techniques used to study functional development**

There are several approaches to the study of brain function in adults are begin applied to young children. The advantages and limitations of each approach are outlined below.

Electroencephalogram (EEG)

EEG measures the electrical field generated by the postsynaptic activity of a synchronously activated palisade of neurons through electrodes placed on scalp²⁴⁰. EEG is mainly sensitive to extracellular volume currents.

The advantages of EEG are:

- Direct measure of neuronal activity
- High temporal resolution (milliseconds)
- Low cost and portable
- Relatively higher tolerance of head movement, thus easier for awake infants

The limitations are:

- Poor spatial resolution (~10 centimeter)
- Low depth penetration (restricted to cortical activity)
- Relatively long preparation time

Magnetoencephalography (MEG)

MEG measures the magnetic field generated by the postsynaptic electronic currents of a synchronously activated population of neurons through magnetometers²⁴¹. MEG is mainly sensitive to intracellular currents.

Advantages include:

- Direct measures of neuronal activity
- High temporal resolution (milliseconds)
- Relatively higher spatial resolution compared with EEG

Limitations include:

- Poor spatial resolution (centimeteres)
- Low depth penetration (restricted to cortical activity)
- High cost and non-portable (infant applications scarce).

Functional near-infrared spectroscopy (fNIRS)

fNIRS measures changes in the local concentrations of oxyhaemoglobin and deoxyhaemoglobin associated with neuronal activity, using near-infrared light.

Advantages of fNIRS include:

- Higher temporal resolution (10s of milliseconds)
- Low cost and portable; relatively higher tolerance of head movement thus easier for awake infants.

Limitations include

- Indirect measure of neuronal activity — the neurovascular coupling mechanism is partially understood
- Poor spatial resolution (centimetres)
- Low depth penetration (restricted to cortical activity)
- Relatively long preparation time

Task-based functional magnetic resonance imaging (fMRI)

fMRI measures changes in the local concentrations of oxyhaemoglobin and deoxyhaemoglobin (that is, the blood-oxygen-level-dependent (BOLD) signal) associated with neuronal activity through sensitivity to local changes in magnetic susceptibility.

Advantages include:

- High spatial resolution (millimeters)
- Deep brain structure coverage
- Sufficient temporal resolution for haemodynamic responses measurement (seconds to sub-seconds, depending on hardware and sequence)

Limitations include:

- Indirect measure of neuronal activity — the neurovascular coupling mechanism is partially understood
- High cost and non-portable
- Large acoustic noise (requiring special ear protection for infants)
- Difficult for awake infants owing to low tolerance of motion (most studies applied in sleeping infants with passive stimuli).
- One common concern for both task-fMRI and fNIRS studies is the immature neurovascular coupling mechanism or ‘haemodynamic response function’ (HRF) in neonates and infants. Although studies have shown adult-like positive BOLD responses to sensorimotor, visual, and auditory stimuli in human newborn and infants^{77, 78}, some studies in infants report that peak BOLD responses are temporally delayed or smaller^{74, 75} and some negative BOLD responses have also been observed^{242–244}, indicating the problem is more complex. The best ways to detect BOLD signal using either task-based fMRI or fNIRS remains an active area of research but age-specific HRFs should at least be considered in human newborn and infant studies

Resting-state fMRI (rsfMRI)

rsfMRI measures the temporal synchronization of spontaneous BOLD fluctuations in the absence of explicit tasks.

Advantages of this approach are:

- The same advantages as task-based fMRI, plus easier experimental setup and low subject-compliance requirement (that is, no task stimuli, can be conducted during natural or sedation-induced sleeping states)
- Full exploration of all functional networks or circuits without limiting to a certain task

As the newest technology for infant study, rsfMRI has extra limitations on top of those shared with task-based fMRI that deserve further consideration and research. These include:

- The neuronal mechanisms underlying the ultra-slow BOLD-signal fluctuations (that is, those occurring at <0.1 Hz, the main frequency band investigated in rsfMRI studies) remains only partially understood (although slow spontaneous modulations in firing rate and gamma-band local field potential have been proposed as potential candidates)
- The likely differences in functional organization⁷⁵ between the brains of infants and adults call for infant-specific functional atlases²⁴⁵ for better definition of regions of interests and results interpretation
- Motion artefacts and physiological confounds affect the estimation of rsfMRI correlations more than detection of task-based fMRI activation, thus require more research for optimal strategies. Although extensive work has been done in adults^{220, 246}, infant-specific optimal strategies deserve more research

The advent of resting-state functional connectivity fMRI⁸⁴, which can probe the functional organization of the whole brain in sleeping infants⁸⁵, has opened a new era for the study of early brain functional development owing to its less complex experimental setup compared to task-based assessments and full coverage of different functional circuits (Box 2). Since the first publication of such studies in sedated premature babies at term age⁸⁶ and naturally sleeping newborns^{87, 88}, an exciting body of work has been accumulating that documents the sequential, coordinated and hierarchical development of functional brain networks^{85, 86, 89–91} (Fig. 3). Specifically, primary functional networks, including the sensorimotor, visual, and auditory networks, develop first and show adult-like topologies in premature babies at term age^{86, 89, 92, 93} and in full-term newborns^{6, 90, 91}. Progressive maturation of these primary networks has been shown in human fetuses⁹⁴ and in premature infants before term age^{88, 89, 89}, corroborating with reports of synchronized oscillatory electrical activity in primary cortices during prenatal development⁶². Thalamus-based relay of information has long been known to be crucial for the development of cortical networks⁹⁵, and indeed, topographic thalamic connections with each of the primary cortical networks has been documented in premature babies at term age⁹⁶ and in full-term newborns⁹⁷, and undergo postnatal refinement and expansion⁹⁷. Particularly, both structural and functional thalamocortical connectivity have been shown to predict later general

cognition and working memory^{97, 98}. White-matter development, particularly myelination, probably interacts with and facilitates the experience-dependent postnatal development of spatially distributed higher-order functional networks. Among them, the default-mode network^{99, 100} shows the earliest synchronization of its constituent regions and achieves an adult-like core structure [G] by around 6 months of age⁹¹, consistent with the emergence of self-awareness, which is thought to depend on the default-mode network, at this age¹⁰¹. The language network, in adults is lateralized and encompasses the inferior frontal gyrus (IFG) and the superior temporal gyrus (STG); these regions begin to exhibit asymmetry in functional connectivity with the homotopic region on the contralateral hemisphere at about age 2 years¹⁰². In general, when defined either based on hypothesis-driven seed-based functional connectivity analysis⁸⁴ (i.e., assessing synchronization with a particular seed, defined based on prior knowledge of the corresponding network in adults) or data-driven independent component analysis¹⁰³ (i.e., data-drivenly defining temporally synchronized but spatially “independent” functional networks without relying on prior knowledge), the default-mode network^{99, 104}, the dorsal attention network [G]^{105, 106} and the salience network [G]¹⁰⁷ establish distributed network-like topologies by 2 years of age^{90, 91} earlier than the executive control networks^{91, 97, 107}, consistent with the protracted development of executive functions¹⁰⁸.

Beyond individual networks, the study of inter-network relationships reveals that functional network development is coordinated¹⁰⁹. For example, preliminary manifestations of sensorimotor-auditory integration through convergent connectivity within the sylvian-parietal-temporal area^{110, 111} has been observed in neonates. Moreover, the inverse correlation between the dorsal attention network and the default-mode network, interpreted by most as a sign of ‘competition’ between the processing of external and internal events in the mature brain¹⁰⁶, also emerges during the first year of life¹¹².

The hierarchical nature of the brain’s functional organization has long been recognized^{113, 114} and step-wise analysis [G] of functional connectivity emerges as a novel technique that could probe this hierarchy¹¹⁵. Notably, sensory networks have been demonstrated to exchange information through two-step inter-regional functional connectivity to the default-mode network and lateral frontoparietal, limbic and basal ganglia regions in subsequent steps^{111, 115}, mimicking the hierarchy proposed by the theory of predictive encoding [G]^{116, 117}. Infants gradually develop this hierarchy in a region- and step-dependent manner, displaying many of the same regions in the top hierarchy (i.e., lateral frontoparietal, limbic and basal ganglia regions) at 1 year of age¹¹¹, suggesting that the brain’s functional hierarchy also emerges during infancy. When evaluating the whole brain as an integrated system, the newborn brain already demonstrates ‘small-world’ functional network properties^{93, 118}, although whole-brain efficiency continues to improve during postnatal development^{118, 119}.

Overall, these studies have greatly enriched the understanding of how functional networks develop in the human brain. Genetically determined early synaptic connections enable the first appearance of synchronized oscillatory electrical activity in the prenatal brain^{62, 63}. Through activity-dependent but largely experience-independent development, primary functional networks are formed before birth. Transitioning from the later part of gestation to

postnatal development, interactions between the experience-dependent processes¹²⁰ and gene expression further modify or refine the primary functional circuits for more efficient signal processing and action generation. Enabled by and rooted in these primary functions, new social, cognitive, and emotional experiences parallel the emergence, growth and fine tuning of corresponding higher-order functional circuits.

Influences of genes and environment

Sex differences

There is a large literature on sex differences in brain structure and function in adults and older children^{121,122}. Studies focused on infancy and early childhood are more limited, but indicate that many of these sex differences are present at birth. Male neonate brains are about 6% larger than female brains at birth, and even at this stage there are already several areas that evidence local sexual dimorphism¹²³. For example, the medial temporal cortex and rolandic operculum are larger in males, whereas the dorsolateral prefrontal, motor, and visual cortices are larger in females¹²³. A more recent study with a larger sample size and more stringent statistical thresholds replicated the finding of increased medial temporal cortex in males and also indicated that female neonates have greater grey-matter volumes around the temporal-parietal junction, a crucial brain region for social cognition¹²⁴. On average, males exhibit a higher number of neocortical neurons than females, and this difference is a likely contributor to the larger intracranial volume observed in males¹²⁵ and may be important for the observed regional differences as well. If sex differences in ICV reflect differences in neocortical neuron number, this suggests an origin in the second trimester, when neurogenesis is at its peak and male fetal testosterone levels are high.

Postnatally, brain volume increases more rapidly in male infants than in female infants, leading to a widening separation in brain sizes^{16, 126}. These differences are probably driven by differences in surface area expansion, as the maturational trajectories of global and regional cortical thickness are highly similar in males and females^{4, 20, 27}. Males also show increased global gyrfication at 2 years of age compared with females (adjusting for total brain volume), but not at 0 or 1 year of age; by contrast, local or regional gyrfication index is similar between males and females at each age¹²⁷.

DTI studies find minimal sex differences in neonates^{128, 129} and in the maturation pattern of major white-matter fibre bundles with time³². By contrast, between 3 and 60 months, females exhibit a higher rate of myelination compared with males in the genu of the corpus callosum, in left frontal and left temporal white matter, and in the right optic radiation¹³⁰. Nevertheless, studies in adults suggest that males have higher FA, higher AD, and lower RD in widespread white-matter regions^{131–135}. These sex differences may therefore emerge in adolescence, as FA increases more rapidly in males than females across this time period, driven by reduction in RD. However, it is important to note that some studies have shown more focal differences in the thalamus, corpus callosum, cingulum, and superior cerebellar peduncles^{136, 137}, and some studies also show regions of increased FA in adult females compared with males, including the corpus callosum and fronto-occipital fasciculus^{137, 138}.

Developmental patterns of resting-state functional connectivity are remarkably similar between males and females, with the exception that interconnectivity between the two lateralized fronto-parietal networks increases at a greater rate in males⁹⁰. In adults, sex differences in amygdala functional connectivity have been reported with females showing stronger positive connectivity between the amygdala and areas associated with face monitoring and discrimination (middle temporal gyrus and inferior frontal gyrus), sensory processing (postcentral gyrus), and emotional processing (hippocampus) and males showing greater connectivity between the amygdala and areas involved in the acquisition of conditioned fear, extinction learning and extinction memory^{139, 140}, consistent with observations during later childhood and adolescent development¹⁴¹.

Sexual differentiation of the brain during the prenatal, perinatal and early postnatal period probably reflects a dynamic interplay of many mechanisms — both biological (for example, prenatal and neonatal hormone production and direct sex chromosome effects) and experiential (for example, resulting from parental expectations and interactive behaviour, exposure to physical hazards or culturally influenced lifestyle differences). Detailed longitudinal studies carried out through collaborations between the biological and social spheres are needed to disentangle the complex dynamics of sex-related patterns in brain development. Ultimately, a better understanding of the pathways leading to sexually dimorphic brain development may help explain sex differences in vulnerability to various psychiatric disorders and open up possibilities for sex-tailored interventions and therapeutics.

Heritability

Twin studies in adults and older children find that the genetic contribution to variance of global brain-tissue volumes is high, with heritability [**G**] estimates typically greater than 80% for global grey and white matter^{142, 143}. Cortical thickness and surface area also have considerable genetic influences that vary by cortical region, but interestingly share little shared genetic variance between them, indicating that these two aspects of cortical volume are regulated by different sets of genetic influences^{144, 145}. Studies have indicated that genetic factors account for 85% variance in neonatal global white-matter volume and 56% variance in neonatal global grey-matter volume¹⁴⁶. Together with the adult data, these results suggest that the heritability of white-matter volume is stable over the course of postnatal brain development and that the heritability of grey-matter volume seems to increase during the period of rapid grey-matter growth from birth to later childhood. Diffusion-tensor imaging studies also indicate that white-matter microstructure, as represented by FA, is generally highly heritable in adults, with variability across different white-matter tracts^{147, 148}. By comparison, the heritability of tract-based FA in neonates^{149, 150} and in 9- and 12-year-olds¹⁵¹ is relatively low, suggesting that the heritability of white-matter microstructure increases over the course of childhood; however, longitudinal studies are lacking.

Studies in adults indicate that structural networks of white matter and resting-state functional networks each have considerable genetic components⁴³. For example, the path length [**G**] and clustering coefficients [**G**] of adult white-matter networks have heritabilities

of 57% and 68% respectively¹⁵², whereas the heritability of default resting-state network connectivity is 42%¹⁵³. One twin study in children aged 5–18 years¹⁵⁴ found that 60% of the genetic variability of cortical thickness was shared across different regions of the cortex, with SCNs of shared genetic variance in fronto-parietal and occipital regions. In 12-year-olds, the path length of resting-state networks has a heritability of 42%¹⁵⁵. Regions of significant heritability of functional connectivity are evident at birth and expand by age 2 years¹⁰⁹, suggesting that genetic influences on resting-state networks are present as networks are established in the first years after birth.

Genetic and transcriptomic studies

The morphometry and functional organization of the infant brain arises, in part, through the precise spatiotemporal regulation of gene expression¹⁵⁶. In particular, prenatal cortical development is characterized by robust regional differences in gene expression that are probably crucial for establishing area-specific subcortical-cortical and cortico-cortical projections⁸. By contrast, infancy and early childhood are characterized by relatively minimal differences in gene expression across cortical areas, as more ‘general’ neuronal and glial differentiation transcription programmes are initiated⁸. These developmental programmes are mediated by transcription factors that bind to specific genomic sequences (such as *cis*-regulatory elements), as well as transcriptional cofactors, chromatin regulators, epigenetic modifications, RNA-binding proteins and non-coding RNAs¹⁵⁷. Genomic variants that influence these processes are expected to have an important role in generating individual differences in brain structure and function.

Genetic mutations are associated with various brain abnormalities that are evident on MRI (see REF. ¹⁵⁸ for review). Common variants in psychiatric risk genes are associated with individual differences in brain tissue volumes in neonates¹⁷¹. For example, of particular interest, neonates homozygous for the rs821616 serine allele of *DISC1* (which encodes disrupted-in schizophrenia 1) exhibit numerous large areas of reduced grey matter in the frontal lobes. Furthermore, neonates homozygous for the rs4680 valine allele of *COMT* (which encodes catechol-O-methyltransferase) exhibit reduced grey matter in the temporal cortex and hippocampus, mirroring findings in adults. Perhaps even more surprisingly, neonates carrying the $\epsilon 4$ allele of *APOE* (which encodes apolipoprotein E), a major susceptibility allele for late-onset Alzheimer disease, have reduced volumes in temporal cortex, highly similar to those reported in the elderly. In another study of *APOE*, 2- to 25-month-old infants who were $\epsilon 4$ carriers showed lower myelin water fraction in the precuneus, the posterior and middle cingulate, the lateral temporal cortex and medial occipitotemporal regions — areas preferentially affected by Alzheimer disease — and greater myelin water fraction in extensive frontal regions compared with noncarriers¹⁷². Although few in number, these studies indicate that risk genes may influence the earliest stages of human brain development, highlight the important role of the prenatal period for future psychiatric and neurological risk and support the idea that Alzheimer disease could be, in part, a developmental disorder.

Infant genotype may also moderate the impact of environmental variables on brain development. This was demonstrated in a study of the effects of antenatal maternal anxiety

and infant *COMT* genotypes (Val158Met, rs737865 and rs165599) on neonatal cortical thickness¹⁵⁹. Neither maternal anxiety nor infant *COMT* genotype were related to regional cortical thickness individually; however, interaction effects between maternal anxiety and infant genotype were observed in several regions, including the right ventromedial prefrontal cortex, an area involved in the regulation of anxiety and mood. Here, maternal anxiety decreased cortical thickness in neonates with two copies of the Met *COMT* allele but increased cortical thickness in neonates with two copies of the Val allele. *BDNF* genotype may also regulate the sensitivity of the methylome [G] to maternal anxiety, with differential effects on amygdala and hippocampal volume¹⁶⁰. Finally, it has also recently been reported that the common variant rs17203281 in *DLG4* (also known as *PSD95*, which encodes postsynaptic density protein 95) is associated with significant differences white-matter microstructure (as indexed by FA) in preterm infants. In addition to being a marker of post-synaptic density, *DLG4* is also expressed by microglia, and the authors speculate that the rs17203281 variant could affect responses to neuroinflammation in children who are born preterm¹⁶¹.

More recently, common variants associated with neonatal brain structure have been identified using a genome-wide association study (GWAS) approach¹⁶². In this study, an intronic single-nucleotide polymorphism (SNP) in *IGFBP7* (rs114518130) was found to be significantly associated with grey-matter volume (volume increases with increasing dose of the G allele). Intriguingly, this locus is within 100kb of *REST*, a master negative regulator of neurogenesis¹⁶³. An intronic SNP in *WWOX* (rs10514437) fell just short of genome-wide significance for white matter. Mutations in *WWOX* are associated with autosomal recessive spinocerebellar ataxia 12, mental retardation, microcephaly and hypomyelination¹⁶⁴, among other neurological phenotypes. This is the first study of its kind¹⁶² and independent replication is crucial, especially for the *IGFBP7* intronic SNP, which was imputed, has a low minor allele frequency, and was not present in the largest racial/ethnic group in the study. A separate study¹⁶⁵ also used genome-wide data and pathway- and network-based approaches to investigate whether common genetic variation influences white-matter microstructure in preterm infants. The results indicated a possible role for peroxisome proliferator-activated receptor (PPAR) signaling in white matter development in preterm infants; however, this study might best be considered a proof-of-concept study, given its very small sample size.

Socioeconomic factors and stress

Recent studies indicate that socio-economic status (SES) can influence the structure of the developing brain^{166, 167}. In older children and adolescents, lower family income and lower parental education have been associated with reduced volumes of cortical grey matter, the hippocampus, and the amygdala¹⁶⁸ as well as reduced cortical surface area and cortical thickness^{169, 170}. Interestingly, white-matter volume does not seem to be affected by SES^{169, 171}, whereas white-matter microstructure can be¹⁷². The influence of SES on brain structure seems to become greater with age in early childhood¹⁷¹, with only marginal associations between parental education and brain volumes at birth¹²⁴. Household income and the level of maternal education are each correlated with within-network connectivity of the default resting-state network at 6 months of age⁹¹.

Studies have begun to elucidate the influence of stress, depression and anxiety during pregnancy on infant brain development. For example, high maternal cortisol levels during pregnancy are associated with greater amygdala volumes in female 7-year-olds¹⁷³. Maternal depression during pregnancy is associated with reduced cortical thickness and white-matter diffusivity in children aged 2.5–5 years¹⁷⁴, and altered microstructure and functional connectivity of the amygdala in 6-month-olds^{175, 176}. Maternal depression and use of selective serotonin-uptake inhibitors during pregnancy is associated with reduced FA across many white-matter tracts in neonates¹⁷⁷. Maternal anxiety is associated with slower hippocampal growth between birth and 6 months of age¹⁷⁸, and in 6- to 12-month-olds, interparental conflict is associated with stronger resting-state connectivity among hubs of the default network, posterior cingulate cortex and anterior medial prefrontal cortex, as well as between the posterior cingulate cortex and the amygdala¹⁷⁹. Thus, early exposure to stress may have an impact on early structural and functional brain development; longitudinal studies are needed to determine how these influences are related to later development and risk.

Prenatal exposure to alcohol or other drugs of abuse also have important effects on developing brain structure; however imaging studies of these effects on the brains of children younger than 5 years old are rare^{180, 181}. Recent studies have revealed substantial alterations of resting-state networks due to prenatal drug exposure^{182, 183}. Specifically, prenatal cocaine exposure is associated with aberrant functional connectivity between the amygdala and the medial prefrontal cortex¹⁸³ and between the thalamus and frontal cortex¹⁸², whereas prenatal marijuana exposure is associated with altered striatal and insular functional connectivity¹⁸⁴.

Predicting risk and cognitive function

The driving force behind many imaging studies of early childhood is the long-term goal of identifying early imaging biomarkers of later cognitive function, behaviour or risk for psychiatric or neurodevelopmental disorders¹⁸⁵. Most studies to date have focused mainly on premature infants. For example, studies of premature infants have generally found that abnormalities of grey-matter and white-matter volume and of white-matter diffusivity are associated with poor neurodevelopmental outcomes^{11, 186}. Some more recent studies have begun to explore other risk groups.

There are emerging studies of infants at risk for psychiatric illness. For example, male neonates with high genetic risk for schizophrenia have increased grey-matter volumes compared with controls¹⁸⁷. Children who are at risk of autism have higher FA values at 6 months and a slower increase of FA after 6 months, resulting in lower FA values at 24 months¹⁸⁸; they also exhibit a hyper-expansion of cortical surface area between 6 and 12 months¹⁸⁹. Stronger amygdala functional connectivity to the anterior insula and ventral striatum in neonates is associated with higher levels of fear in 6-month-olds¹⁹⁰, which may be a precursor of later anxiety disorders. In addition, lower connectivity between the thalamus and the salience network in 1-year-olds is associated with poorer working memory performance and lower cognitive development at 2 years of age⁹⁷. Thus, thalamus–salience network connectivity might have potential as a candidate biomarker for differences in

cognitive performance, a core component in various developmental and psychiatric disorders.

Studies using cranial ultrasound at 6 weeks of age find that a smaller corpus callosum length is related to parental reports of childrens' executive function at age 4 years¹⁹¹, whereas smaller subcortical volumes are associated with higher internalizing behaviour [G] scores at 18 and 36 months¹⁹². Moreover, a recent study found that white-matter networks at birth may be predictive of internalizing and externalizing behaviour [G] at 2 and 4 years of age¹⁹³. These studies suggest that early imaging biomarkers for risk for later behavioral abnormalities may exist, although the associations between imaging markers and outcomes are generally not strong. The search for imaging biomarkers for brain disorders is fraught with difficulty related to the complexity and distribution of cognitive processing in the brain, the generalization of findings from heterogeneous clinical samples, and small effect sizes that have limited clinical utility^{194, 195}.

There have been a few studies of structure-cognitive function relationships in early childhood, with a general theme that structure-cognitive function relationships in early childhood are age dependent¹⁹⁶. Cortical surface morphology, as determined by deformation of cortical surfaces (a measure of grey-matter density), is associated with cognitive development in the first 2 years of life, and this association varies with age¹⁹⁷. In addition, white-matter microstructure properties measured using DTI are moderately associated with working memory performance in 1-year-olds¹⁹⁸. Studies using myelin water fraction have found small but statistically significant associations between myelination trajectories in several regions including bilateral frontal and temporal cortex and language development from 3 months to 4 years¹⁹⁹, as well as age-dependent relationships between myelination and general cognitive ability from 3 months to 5 years²⁰⁰. Factors that describe common variation of FA, AD or RD across groups of white matter tracts are weakly related to cognitive development in an age-specific way; for example, a common factor of AD across 12 white matter tracts at birth is associated with cognitive development at age 1 year.³⁸

Conclusions and future directions

Imaging studies in early childhood have begun to characterize structural and functional brain development in this important but still understudied period. Studies to date have found that, by birth, major white-matter tracts are in place and white-matter structural networks and sensorimotor resting-state functional networks are well developed. The first year of life is a period of robust grey-matter growth, rapid myelination and maturation of microstructure of existing white-matter tracts, as well as development of higher-order resting-state functional networks. By age 2 years, the fundamental structural and functional architecture of the brain seems to be in place, and the brain maturation that occurs in later childhood is much slower.

Much work is yet to be done and major challenges exist (Boxes 2–4). Early studies find only modest relationships between structural and functional biomarkers and cognition and behaviour. Machine-learning based prediction frameworks are beginning to be applied to infant imaging^{201–203} and are needed to better predict developmental outcomes from early imaging parameters, in context of genetic, environmental and birth-related variables. It is

unlikely that a single or even a few imaging parameters will have sufficient power for accurate prediction. ‘Big data’ projects in which data are collected from large samples during early childhood (for example, in the US Foundation for the National Institutes of Health (FNIH) Baby Connectome Project and the European Research Council’s Developing Human Connectome Project; see Further Information) will be required to capture the imaging, genetic and environmental variables that contribute a small percentage of variance.

Imaging studies in early childhood offer the promise of understanding the early origins of, and genetic and environmental influences affecting, individual differences in cognition, behaviour and risk for neuropsychiatric disease, and these studies are beginning to fulfil these aims. Moreover, these studies may be able to identify risk biomarkers that are present long before cognitive and clinical abnormalities arise and ultimately inform the timing and nature of early interventions to modify suboptimal developmental trajectories, improve outcome and mitigate risk.

Acknowledgments

The authors thank members of the University of North Carolina Early Brain Development Lab for their help with the manuscript and figures, and M. Styner and D. Rubinow for their thoughtful reading and comments. This work is supported by the US National Institutes of Health: MH070890, HD053000, and MH111944 to J.H.G.; NS088975, DA043171, and DA036645 to W.G.; and MH092335 and MH104330 to R.K.S.

Glossary terms

Axial diffusivity (AD)

Water diffusion along the principal axis of a fibre, thought to be dependent on fibre compactness and microstructure.

Radial diffusivity (RD)

Water diffusion perpendicular to the main axis of the fibre, sensitive to myelination.

Fractional anisotropy (FA)

A summary measure of microstructure that takes into account axial diffusivity and radial diffusivity.

Network integration

The overall capacity of the network to be interconnected and exchange information.

Network segregation

The degree to which parts of a network form localized clusters of nodes or modules of connections.

Structural covariance networks

Regions of cortical grey matter with correlated variance in cortical thickness.

Maturational networks

Regions of cortical grey matter with correlated changes in cortical thickness over time.

Core structure

The collection of key hub regions that possess the most connections that bring the whole network together.

Dorsal attention network

The network involved with the “top-down” or voluntary focusing of attention; includes the intraparietal sulcus, frontal eye fields and middle temporal regions.

Saliency network

The network involved with the selection of relevant stimuli; includes the insula, anterior cingulate cortex and amygdala.

Step-wise analysis

A technique that attempts to identify intermediate connector regions and multi-step links between two brain regions that do not show directly correlated functional activity.

Predictive encoding

Theoretical framework in which higher-level cortices continuously generate predictions about the environment based on learned input regularities, to minimize errors between lower-level inputs and predictions.

Heritability

The proportion of variance in a trait or measure that is due to genetic variation.

Path length

A measure of efficiency in a network, the average number of edges along the shortest paths connecting all pairs of nodes within a network.

Clustering coefficient

A measure of the proportion of existing links between one node’s neighbors divided by the total number of links for a fully connected neighborhood. It reflects how densely one node and its neighbors are locally connected.

Methylome

The pattern of DNA methylation within a genome.

Internalizing behavior

Problem behaviours that are typically directed inward, such as anxiety, depression, social withdrawal and somatic symptoms.

Externalizing behavior

Problem behaviours that are directed outward toward others, such as physical aggression, defiance, hyperactivity, bullying and theft.

References

1. Nelson CA 3rd, et al. Cognitive recovery in socially deprived young children: the Bucharest Early Intervention Project. *Science*. 2007; 318:1937–40. [PubMed: 18096809]

2. National Advisory Mental Health Council Workgroup on Neurodevelopment. Transformative Neurodevelopmental Research in Mental Illness. 2008. (http://www.nimh.nih.gov/about/advisory-boards-and-groups/namhc/neurodevelopment_workgroup_report.pdf)
3. Knickmeyer RC, et al. A structural MRI study of human brain development from birth to 2 years. *J Neurosci*. 2008; 28:12176–82. The first comprehensive study of structural brain development in the first 2 years of life demonstrating the rapid growth of cortical gray matter in the first year of life. [PubMed: 19020011]
4. Lyall AE, et al. Dynamic Development of Regional Cortical Thickness and Surface Area in Early Childhood. *Cereb Cortex*. 2015; 25:2204–12. [PubMed: 24591525]
5. Dubois J, et al. The early development of brain white matter: a review of imaging studies in fetuses, newborns and infants. *Neuroscience*. 2014a; 276:48–71. A comprehensive review of the use of imaging to study white matter in early brain development. [PubMed: 24378955]
6. Gao W, et al. Evidence on the emergence of the brain's default network from 2-week-old to 2-year-old healthy pediatric subjects. *Proc Natl Acad Sci U S A*. 2009a; 106:6790–5. [PubMed: 19351894]
7. Petanjek Z, et al. Extraordinary neoteny of synaptic spines in the human prefrontal cortex. *Proc Natl Acad Sci U S A*. 2011; 108:13281–6. [PubMed: 21788513]
8. Pletikos M, et al. Temporal specification and bilaterality of human neocortical topographic gene expression. *Neuron*. 2014; 81:321–32. [PubMed: 24373884]
9. Geng X, et al. Structural and Maturational Covariance in Early Childhood Brain Development. *Cereb Cortex*. 2017; 27:1795–1807. [PubMed: 26874184]
10. Gilmore JH, et al. Longitudinal development of cortical and subcortical gray matter from birth to 2 years. *Cereb Cortex*. 2012; 22:2478–85. [PubMed: 22109543]
11. Anderson PJ, Cheong JL, Thompson DK. The predictive validity of neonatal MRI for neurodevelopmental outcome in very preterm children. *Semin Perinatol*. 2015; 39:147–58. [PubMed: 25724792]
12. Jakab A, et al. Fetal Cerebral Magnetic Resonance Imaging Beyond Morphology. *Semin Ultrasound CT MR*. 2015; 36:465–75. [PubMed: 26614130]
13. Gilmore JH, et al. Regional gray matter growth, sexual dimorphism, and cerebral asymmetry in the neonatal brain. *J Neurosci*. 2007; 27:1255–60. [PubMed: 17287499]
14. Courchesne E, et al. Normal brain development and aging: quantitative analysis at in vivo MR imaging in healthy volunteers. *Radiology*. 2000; 216:672–82. [PubMed: 10966694]
15. Matsuzawa J, et al. Age-related volumetric changes of brain gray and white matter in healthy infants and children. *Cereb Cortex*. 2001; 11:335–42. [PubMed: 11278196]
16. Holland D, et al. Structural growth trajectories and rates of change in the first 3 months of infant brain development. *JAMA Neurol*. 2014; 71:1266–74. [PubMed: 25111045]
17. Groeschel S, Vollmer B, King MD, Connelly A. Developmental changes in cerebral grey and white matter volume from infancy to adulthood. *Int J Dev Neurosci*. 2010; 28:481–9. [PubMed: 20600789]
18. Mills KL, et al. Structural brain development between childhood and adulthood: Convergence across four longitudinal samples. *Neuroimage*. 2016; 141:273–81. [PubMed: 27453157]
19. Bompard L, et al. Multivariate longitudinal shape analysis of human lateral ventricles during the first twenty-four months of life. *PLoS One*. 2014; 9:e108306. [PubMed: 25265017]
20. Raznahan A, et al. How does your cortex grow? *J Neurosci*. 2011b; 31:7174–7. [PubMed: 21562281]
21. Shaw P, et al. Neurodevelopmental trajectories of the human cerebral cortex. *J Neurosci*. 2008; 28:3586–94. [PubMed: 18385317]
22. Remer J, et al. Quantifying Cortical Development in Typically Developing Toddlers and Young Children, 1–6 Years of Age. *NeuroImage*. 2017 Apr 5. In Press.
23. Li G, et al. Mapping region-specific longitudinal cortical surface expansion from birth to 2 years of age. *Cereb Cortex*. 2013; 23:2724–33. [PubMed: 22923087]
24. Hill J, et al. Similar patterns of cortical expansion during human development and evolution. *Proc Natl Acad Sci U S A*. 2010a; 107:13135–40. [PubMed: 20624964]

25. Brown TT, et al. Neuroanatomical assessment of biological maturity. *Curr Biol.* 2012; 22:1693–8. [PubMed: 22902750]
26. Li G, Lin W, Gilmore JH, Shen D. Spatial Patterns, Longitudinal Development, and Hemispheric Asymmetries of Cortical Thickness in Infants from Birth to 2 Years of Age. *J Neurosci.* 2015; 35:9150–62. [PubMed: 26085637]
27. Ducharme S, et al. Trajectories of cortical thickness maturation in normal brain development--The importance of quality control procedures. *Neuroimage.* 2016; 125:267–79. [PubMed: 26463175]
28. Walhovd KB, Fjell AM, Giedd J, Dale AM, Brown TT. Through Thick and Thin: a Need to Reconcile Contradictory Results on Trajectories in Human Cortical Development. *Cereb Cortex.* 2017; 27:1472–1481. [PubMed: 28365755]
29. Schoenemann PT, Sheehan MJ, Glotzer LD. Prefrontal white matter volume is disproportionately larger in humans than in other primates. *Nat Neurosci.* 2005; 8:242–52. [PubMed: 15665874]
30. Zhang K, Sejnowski TJ. A universal scaling law between gray matter and white matter of cerebral cortex. *Proc Natl Acad Sci U S A.* 2000; 97:5621–6. [PubMed: 10792049]
31. Qiu A, Mori S, Miller MI. Diffusion tensor imaging for understanding brain development in early life. *Annu Rev Psychol.* 2015c; 66:853–76. [PubMed: 25559117]
32. Geng X, et al. Quantitative tract-based white matter development from birth to age 2years. *Neuroimage.* 2012a; 61:542–57. [PubMed: 22510254]
33. Deoni SC, et al. Mapping infant brain myelination with magnetic resonance imaging. *J Neurosci.* 2011; 31:784–91. The initial imaging study of myelin development in the human infant using the myelin water fraction approach, a more direct assessment of myelin compared to standard diffusion weighted imaging. [PubMed: 21228187]
34. Faria AV, et al. Atlas-based analysis of neurodevelopment from infancy to adulthood using diffusion tensor imaging and applications for automated abnormality detection. *Neuroimage.* 2010; 52:415–28. [PubMed: 20420929]
35. Krogsrud SK, et al. Changes in white matter microstructure in the developing brain--A longitudinal diffusion tensor imaging study of children from 4 to 11 years of age. *Neuroimage.* 2016; 124:473–86. [PubMed: 26375208]
36. Dean DC 3rd, et al. Modeling healthy male white matter and myelin development: 3 through 60 months of age. *Neuroimage.* 2014; 84:742–52. [PubMed: 24095814]
37. Dean DC 3rd, et al. Characterizing longitudinal white matter development during early childhood. *Brain Struct Funct.* 2015; 220:1921–33. [PubMed: 24710623]
38. Lee SJ, et al. Common and heritable components of white matter microstructure predict cognitive function at 1 and 2 y. *Proc Natl Acad Sci U S A.* 2017; 114:148–153. [PubMed: 27994134]
39. Bullmore E, Sporns O. Complex brain networks: graph theoretical analysis of structural and functional systems. *Nat Rev Neurosci.* 2009; 10:186–98. [PubMed: 19190637]
40. Rubinov M, Sporns O. Complex network measures of brain connectivity: uses and interpretations. *Neuroimage.* 2010; 52:1059–69. [PubMed: 19819337]
41. Bullmore E, Sporns O. The economy of brain network organization. *Nat Rev Neurosci.* 2012; 13:336–49. [PubMed: 22498897]
42. Khundrakpam BS, Lewis JD, Zhao L, Chouinard-Decorte F, Evans AC. Brain connectivity in normally developing children and adolescents. *Neuroimage.* 2016; 134:192–203. [PubMed: 27054487]
43. Richmond S, Johnson KA, Seal ML, Allen NB, Whittle S. Development of brain networks and relevance of environmental and genetic factors: A systematic review. *Neurosci Biobehav Rev.* 2016; 71:215–239. [PubMed: 27590832]
44. Cao M, Huang H, He Y. Developmental Connectomics from Infancy through Early Childhood. *Trends Neurosci.* 2017; 40:494–506. [PubMed: 28684174]
45. Senden M, Reuter N, van den Heuvel MP, Goebel R, Deco G. Cortical rich club regions can organize state-dependent functional network formation by engaging in oscillatory behavior. *Neuroimage.* 2017; 146:561–574. [PubMed: 27989843]
46. Yan G, et al. Network control principles predict neuron function in the *Caenorhabditis elegans* connectome. *Nature.* 2017; 550:519–523. [PubMed: 29045391]

47. Cao M, Huang H, Peng Y, Dong Q, He Y. Toward Developmental Connectomics of the Human Brain. *Front Neuroanat.* 2016; 10:25. [PubMed: 27064378]
48. Yap PT, et al. Development trends of white matter connectivity in the first years of life. *PLoS One.* 2011; 6:e24678. [PubMed: 21966364]
49. van den Heuvel MP, et al. The Neonatal Connectome During Preterm Brain Development. *Cereb Cortex.* 2015; 25:3000–13. [PubMed: 24833018]
50. Ball G, et al. Rich-club organization of the newborn human brain. *Proc Natl Acad Sci U S A.* 2014; 111:7456–61. An early study of the white matter connectome in preterm and term infants demonstrating that major hubs are already present at 30 weeks gestational age. [PubMed: 24799693]
51. Huang H, et al. Development of human brain structural networks through infancy and childhood. *Cereb Cortex.* 2015; 25:1389–404. [PubMed: 24335033]
52. Hagmann P, et al. White matter maturation reshapes structural connectivity in the late developing human brain. *Proc Natl Acad Sci U S A.* 2010; 107:19067–72. [PubMed: 20956328]
53. Evans AC. Networks of anatomical covariance. *Neuroimage.* 2013; 80:489–504. [PubMed: 23711536]
54. Alexander-Bloch A, Raznahan A, Bullmore E, Giedd J. The convergence of maturational change and structural covariance in human cortical networks. *J Neurosci.* 2013a; 33:2889–99. [PubMed: 23407947]
55. Schmitt JE, et al. Identification of genetically mediated cortical networks: a multivariate study of pediatric twins and siblings. *Cereb Cortex.* 2008; 18:1737–47. [PubMed: 18234689]
56. Alexander-Bloch A, Giedd JN, Bullmore E. Imaging structural co-variance between human brain regions. *Nat Rev Neurosci.* 2013b; 14:322–36. [PubMed: 23531697]
57. Khundrakpam BS, et al. Imaging structural covariance in the development of intelligence. *Neuroimage.* 2017; 144:227–240. [PubMed: 27554529]
58. Fan Y, et al. Brain anatomical networks in early human brain development. *Neuroimage.* 2011; 54:1862–71. [PubMed: 20650319]
59. Nie J, Li G, Shen D. Development of cortical anatomical properties from early childhood to early adulthood. *Neuroimage.* 2013; 76:216–24. [PubMed: 23523806]
60. Zielinski BA, Gennatas ED, Zhou J, Seeley WW. Network-level structural covariance in the developing brain. *Proc Natl Acad Sci U S A.* 2010; 107:18191–6. [PubMed: 20921389]
61. Khundrakpam BS, et al. Developmental changes in organization of structural brain networks. *Cereb Cortex.* 2013; 23:2072–85. [PubMed: 22784607]
62. Khazipov R, Luhmann HJ. Early patterns of electrical activity in the developing cerebral cortex of humans and rodents. *Trends Neurosci.* 2006; 29:414–418. [PubMed: 16713634]
63. Dehaene-Lambertz G, Spelke ES. The Infancy of the Human Brain. *Neuron.* 2015; 88:93–109. [PubMed: 26447575]
64. Dreyfus-Brisac C, Larroche JC. Discontinuous electroencephalograms in the premature newborn and at term. Electro-anatomo-clinical correlations. *Rev Electroencephalogr Neurophysiol Clin.* 1971; 1:95–9. [PubMed: 5173705]
65. Anderson CM, Torres F, Faoro A. The EEG of the early premature. *Electroencephalogr Clin Neurophysiol.* 1985; 60:95–105. [PubMed: 2578372]
66. Arichi T, et al. Localization of spontaneous bursting neuronal activity in the preterm human brain with simultaneous EEG-fMRI. *Elife.* 2017; 6
67. Hanganu IL, Ben-Ari Y, Khazipov R. Retinal waves trigger spindle bursts in the neonatal rat visual cortex. *J Neurosci.* 2006; 26:6728–36. [PubMed: 16793880]
68. Ackman JB, Burbridge TJ, Crair MC. Retinal waves coordinate patterned activity throughout the developing visual system. *Nature.* 2012; 490:219–25. [PubMed: 23060192]
69. Tritsch NX, Yi E, Gale JE, Glowatzki E, Bergles DE. The origin of spontaneous activity in the developing auditory system. *Nature.* 2007; 450:50–5. [PubMed: 17972875]
70. Blumberg MS. Developing sensorimotor systems in our sleep. *Current directions in psychological science.* 2015; 24:32–37. [PubMed: 25937709]

71. Kwong KK, et al. Dynamic magnetic resonance imaging of human brain activity during primary sensory stimulation. *Proc Natl Acad Sci U S A.* 1992; 89:5675–9. [PubMed: 1608978]
72. Ogawa S, et al. Intrinsic signal changes accompanying sensory stimulation: functional brain mapping with magnetic resonance imaging. *Proc Natl Acad Sci U S A.* 1992; 89:5951–5. [PubMed: 1631079]
73. Jobsis FF. Noninvasive, infrared monitoring of cerebral and myocardial oxygen sufficiency and circulatory parameters. *Science.* 1977; 198:1264–7. [PubMed: 929199]
74. Arichi T, et al. Development of BOLD signal hemodynamic responses in the human brain. *Neuroimage.* 2012; 63:663–73. [PubMed: 22776460]
75. Allievi AG, et al. Maturation of Sensori-Motor Functional Responses in the Preterm Brain. *Cereb Cortex.* 2016; 26:402–413. [PubMed: 26491066]
76. Karen T, et al. Hemodynamic response to visual stimulation in newborn infants using functional near-infrared spectroscopy. *Hum Brain Mapp.* 2008; 29:453–60. [PubMed: 17525986]
77. Dehaene-Lambertz G, Dehaene S, Hertz-Pannier L. Functional neuroimaging of speech perception in infants. *Science.* 2002; 298:2013–5. An important early task-based functional imaging study that demonstrated that adult language areas are already active in 2–3 month olds. [PubMed: 12471265]
78. Dehaene S, Sergent C, Changeux JP. A neuronal network model linking subjective reports and objective physiological data during conscious perception. *Proc Natl Acad Sci U S A.* 2003; 100:8520–5. [PubMed: 12829797]
79. Wilcox T, Haslup JA, Boas DA. Dissociation of processing of featural and spatiotemporal information in the infant cortex. *Neuroimage.* 2010; 53:1256–63. [PubMed: 20603218]
80. Wilcox T, Stubbs J, Hirshkowitz A, Boas D. Object processing and functional organization of the infant cortex. *NeuroImage.* 2012; 62:1833–1840. [PubMed: 22634218]
81. Nakano T, Watanabe H, Homae F, Taga G. Prefrontal cortical involvement in young infants' analysis of novelty. *Cereb Cortex.* 2009; 19:455–63. [PubMed: 18544555]
82. Grossmann T, Johnson MH. Selective prefrontal cortex responses to joint attention in early infancy. *Biol Lett.* 2010; 6:540–3. [PubMed: 20106861]
83. Kozberg M, Hillman E. Neurovascular coupling and energy metabolism in the developing brain. *Prog Brain Res.* 2016; 225:213–42. [PubMed: 27130418]
84. Biswal B, Yetkin FZ, Haughton VM, Hyde JS. Functional connectivity in the motor cortex of resting human brain using echo-planar MRI. *Magn Reson Med.* 1995; 34:537–41. [PubMed: 8524021]
85. Gao W, Lin W, Grewen K, Gilmore JH. Functional Connectivity of the Infant Human Brain: Plastic and Modifiable. *Neuroscientist.* 2016
86. Fransson P, et al. Resting-state networks in the infant brain. *Proc Natl Acad Sci U S A.* 2007; 104:15531–6. This study was the first to delineate resting-state functional networks in human infants utilizing a cohort of sedated premature infants. [PubMed: 17878310]
87. Lin W, et al. Functional connectivity MR imaging reveals cortical functional connectivity in the developing brain. *American Journal of Neuroradiology.* 2008; 29:1883–1889. [PubMed: 18784212]
88. Liu WC, Flax JF, Guise KG, Sukul V, Benasich AA. Functional connectivity of the sensorimotor area in naturally sleeping infants. *Brain Res.* 2008; 1223:42–9. [PubMed: 18599026]
89. Smyser CD, et al. Longitudinal analysis of neural network development in preterm infants. *Cereb Cortex.* 2010; 20:2852–62. [PubMed: 20237243]
90. Gao W, Alcauter S, Smith JK, Gilmore JH, Lin W. Development of human brain cortical network architecture during infancy. *Brain Struct Funct.* 2015b; 220:1173–86. [PubMed: 24469153]
91. Gao W, et al. Functional Network Development During the First Year: Relative Sequence and Socioeconomic Correlations. *Cereb Cortex.* 2015a; 25:2919–28. This paper was the first to delineate the developmental sequence of nine major resting state networks during the first year of life in a cohort of full-term infants. [PubMed: 24812084]
92. Doria V, et al. Emergence of resting state networks in the preterm human brain. *Proc Natl Acad Sci U S A.* 2010; 107:20015–20. [PubMed: 21041625]

93. Fransson P, Aden U, Blennow M, Lagercrantz H. The functional architecture of the infant brain as revealed by resting-state fMRI. *Cereb Cortex*. 2011; 21:145–54. [PubMed: 20421249]
94. Thomason ME, et al. Cross-hemispheric functional connectivity in the human fetal brain. *Sci Transl Med*. 2013; 5:173ra24.
95. Jones, EG. *The thalamus*. Springer Science & Business Media; 2012.
96. Toulmin H, et al. Specialization and integration of functional thalamocortical connectivity in the human infant. *Proc Natl Acad Sci U S A*. 2015; 112:6485–90. [PubMed: 25941391]
97. Alcauter S, et al. Development of thalamocortical connectivity during infancy and its cognitive correlations. *J Neurosci*. 2014; 34:9067–75. [PubMed: 24990927]
98. Ball G, et al. Thalamocortical Connectivity Predicts Cognition in Children Born Preterm. *Cereb Cortex*. 2015; 25:4310–8. [PubMed: 25596587]
99. Raichle ME, et al. A default mode of brain function. *Proc Natl Acad Sci U S A*. 2001; 98:676–82. [PubMed: 11209064]
100. Gao W, et al. Temporal and spatial development of axonal maturation and myelination of white matter in the developing brain. *AJNR Am J Neuroradiol*. 2009b; 30:290–6. [PubMed: 19001533]
101. Amsterdam B. Mirror self-image reactions before age two. *Dev Psychobiol*. 1972; 5:297–305. [PubMed: 4679817]
102. Emerson RW, Gao W, Lin W. Longitudinal Study of the Emerging Functional Connectivity Asymmetry of Primary Language Regions during Infancy. *J Neurosci*. 2016; 36:10883–10892. [PubMed: 27798142]
103. Calhoun VD, Adali T, Pearlson GD, Pekar JJ. A method for making group inferences from functional MRI data using independent component analysis. *Hum Brain Mapp*. 2001; 14:140–51. [PubMed: 11559959]
104. Shulman GL, et al. Common Blood Flow Changes across Visual Tasks: II. Decreases in Cerebral Cortex. *J Cogn Neurosci*. 1997; 9:648–63. [PubMed: 23965122]
105. Corbetta M, Shulman GL. Control of goal-directed and stimulus-driven attention in the brain. *Nat Rev Neurosci*. 2002; 3:201–15. [PubMed: 11994752]
106. Fox MD, et al. The human brain is intrinsically organized into dynamic, anticorrelated functional networks. *Proc Natl Acad Sci U S A*. 2005; 102:9673–8. [PubMed: 15976020]
107. Seeley WW, et al. Dissociable intrinsic connectivity networks for salience processing and executive control. *J Neurosci*. 2007; 27:2349–56. [PubMed: 17329432]
108. Rothbart MK, Sheese BE, Rueda MR, Posner MI. Developing Mechanisms of Self-Regulation in Early Life. *Emot Rev*. 2011; 3:207–213. [PubMed: 21892360]
109. Gao W, et al. Intersubject variability of and genetic effects on the brain's functional connectivity during infancy. *J Neurosci*. 2014; 34:11288–96. [PubMed: 25143609]
110. Andersen RA. Multimodal integration for the representation of space in the posterior parietal cortex. *Philos Trans R Soc Lond B Biol Sci*. 1997; 352:1421–8. [PubMed: 9368930]
111. Pendl SL, et al. Emergence of a hierarchical brain during infancy reflected by stepwise functional connectivity. *Hum Brain Mapp*. 2017; 38:2666–2682. [PubMed: 28263011]
112. Gao W, et al. The synchronization within and interaction between the default and dorsal attention networks in early infancy. *Cereb Cortex*. 2013; 23:594–603. [PubMed: 22368080]
113. Damasio AR. Time-locked multiregional retroactivation: a systems-level proposal for the neural substrates of recall and recognition. *Cognition*. 1989; 33:25–62. [PubMed: 2691184]
114. Mesulam MM. From sensation to cognition. *Brain*. 1998; 121(Pt 6):1013–52. [PubMed: 9648540]
115. Sepulcre J, Sabuncu MR, Yeo TB, Liu H, Johnson KA. Stepwise connectivity of the modal cortex reveals the multimodal organization of the human brain. *J Neurosci*. 2012; 32:10649–61. [PubMed: 22855814]
116. Bastos AM, et al. Canonical microcircuits for predictive coding. *Neuron*. 2012; 76:695–711. [PubMed: 23177956]
117. Barrett LF, Simmons WK. Interoceptive predictions in the brain. *Nat Rev Neurosci*. 2015; 16:419–29. [PubMed: 26016744]
118. Gao W, et al. Temporal and spatial evolution of brain network topology during the first two years of life. *PLoS One*. 2011; 6:e25278. [PubMed: 21966479]

119. Supekar K, Musen M, Menon V. Development of large-scale functional brain networks in children. *PLoS Biol.* 2009; 7:e1000157. [PubMed: 19621066]
120. Feldman DE, Brecht M. Map plasticity in somatosensory cortex. *Science.* 2005; 310:810–5. [PubMed: 16272113]
121. Ruigrok AN, et al. A meta-analysis of sex differences in human brain structure. *Neurosci Biobehav Rev.* 2014; 39:34–50. [PubMed: 24374381]
122. Sacher J, Neumann J, Okon-Singer H, Gotowiec S, Villringer A. Sexual dimorphism in the human brain: evidence from neuroimaging. *Magn Reson Imaging.* 2013; 31:366–75. [PubMed: 22921939]
123. Knickmeyer RC, et al. Impact of sex and gonadal steroids on neonatal brain structure. *Cereb Cortex.* 2014a; 24:2721–31. [PubMed: 23689636]
124. Knickmeyer RC, et al. Impact of Demographic and Obstetric Factors on Infant Brain Volumes: A Population Neuroscience Study. *Cereb Cortex.* 2016
125. Pakkenberg B, Gundersen HJ. Neocortical neuron number in humans: effect of sex and age. *J Comp Neurol.* 1997; 384:312–20. [PubMed: 9215725]
126. Tanaka C, Matsui M, Uematsu A, Noguchi K, Miyawaki T. Developmental trajectories of the fronto-temporal lobes from infancy to early adulthood in healthy individuals. *Dev Neurosci.* 2012; 34:477–87. [PubMed: 23257954]
127. Li G, et al. Mapping longitudinal development of local cortical gyrification in infants from birth to 2 years of age. *J Neurosci.* 2014; 34:4228–38. [PubMed: 24647943]
128. Liu Y, et al. Gender differences in language and motor-related fibers in a population of healthy preterm neonates at term-equivalent age: a diffusion tensor and probabilistic tractography study. *AJNR Am J Neuroradiol.* 2011; 32:2011–6. [PubMed: 21940804]
129. Ratnarajah N, et al. Structural connectivity asymmetry in the neonatal brain. *Neuroimage.* 2013; 75:187–94. [PubMed: 23501049]
130. Deoni SC, Dean DC 3rd, O'Muircheartaigh J, Dirks H, Jerskey BA. Investigating white matter development in infancy and early childhood using myelin water fraction and relaxation time mapping. *Neuroimage.* 2012; 63:1038–53. [PubMed: 22884937]
131. Inano S, Takao H, Hayashi N, Abe O, Ohtomo K. Effects of age and gender on white matter integrity. *AJNR Am J Neuroradiol.* 2011; 32:2103–9. [PubMed: 21998104]
132. van Hemmen J, et al. Sex Differences in White Matter Microstructure in the Human Brain Predominantly Reflect Differences in Sex Hormone Exposure. *Cereb Cortex.* 2017; 27:2994–3001. [PubMed: 27226438]
133. Rametti G, et al. White matter microstructure in female to male transsexuals before cross-sex hormonal treatment. A diffusion tensor imaging study. *J Psychiatr Res.* 2011; 45:199–204. [PubMed: 20562024]
134. den Braber A, et al. Sex differences in gray and white matter structure in age-matched unrelated males and females and opposite-sex siblings. *International Journal of Psychological Research.* 2013; 6:7–21.
135. Takao H, Hayashi N, Ohtomo K. Sex dimorphism in the white matter: fractional anisotropy and brain size. *J Magn Reson Imaging.* 2014; 39:917–23. [PubMed: 24123288]
136. Menzler K, et al. Men and women are different: diffusion tensor imaging reveals sexual dimorphism in the microstructure of the thalamus, corpus callosum and cingulum. *Neuroimage.* 2011; 54:2557–62. [PubMed: 21087671]
137. Kanaan RA, et al. Gender influence on white matter microstructure: a tract-based spatial statistics analysis. *PLoS One.* 2014; 9:e91109. [PubMed: 24603769]
138. Chou KH, Cheng Y, Chen IY, Lin CP, Chu WC. Sex-linked white matter microstructure of the social and analytic brain. *Neuroimage.* 2011; 54:725–33. [PubMed: 20633662]
139. Kogler L, et al. Sex differences in the functional connectivity of the amygdalae in association with cortisol. *Neuroimage.* 2016; 134:410–23. [PubMed: 27039701]
140. Engman J, Linnman C, Van Dijk KR, Milad MR. Amygdala subnuclei resting-state functional connectivity sex and estrogen differences. *Psychoneuroendocrinology.* 2016; 63:34–42. [PubMed: 26406106]

141. Alarcon G, Cservenka A, Rudolph MD, Fair DA, Nagel BJ. Developmental sex differences in resting state functional connectivity of amygdala sub-regions. *Neuroimage*. 2015; 115:235–44. [PubMed: 25887261]
142. Jansen AG, Mous SE, White T, Posthuma D, Polderman TJ. What twin studies tell us about the heritability of brain development, morphology, and function: a review. *Neuropsychol Rev*. 2015; 25:27–46. [PubMed: 25672928]
143. Blokland GA, de Zubicaray GI, McMahon KL, Wright MJ. Genetic and environmental influences on neuroimaging phenotypes: a meta-analytical perspective on twin imaging studies. *Twin Res Hum Genet*. 2012; 15:351–71. [PubMed: 22856370]
144. Panizzon MS, et al. Distinct genetic influences on cortical surface area and cortical thickness. *Cereb Cortex*. 2009; 19:2728–35. [PubMed: 19299253]
145. Chen CH, et al. Genetic topography of brain morphology. *Proc Natl Acad Sci U S A*. 2013; 110:17089–94. [PubMed: 24082094]
146. Gilmore JH, et al. Genetic and environmental contributions to neonatal brain structure: A twin study. *Hum Brain Mapp*. 2010a; 31:1174–82. [PubMed: 20063301]
147. Kochunov P, et al. Heritability of fractional anisotropy in human white matter: a comparison of Human Connectome Project and ENIGMA-DTI data. *Neuroimage*. 2015; 111:300–11. [PubMed: 25747917]
148. Vuoksima E, et al. Heritability of white matter microstructure in late middle age: A twin study of tract-based fractional anisotropy and absolute diffusivity indices. *Hum Brain Mapp*. 2016
149. Geng X, et al. White matter heritability using diffusion tensor imaging in neonatal brains. *Twin Res Hum Genet*. 2012b; 15:336–50. [PubMed: 22856369]
150. Lee SJ, et al. Quantitative tract-based white matter heritability in twin neonates. *Neuroimage*. 2015; 111:123–35. [PubMed: 25700954]
151. Brouwer RM, et al. White matter development in early puberty: a longitudinal volumetric and diffusion tensor imaging twin study. *PLoS One*. 2012; 7:e32316. [PubMed: 22514599]
152. Bohlken MM, et al. Genes contributing to subcortical volumes and intellectual ability implicate the thalamus. *Hum Brain Mapp*. 2014; 35:2632–42. [PubMed: 24038793]
153. Glahn DC, et al. Genetic control over the resting brain. *Proc Natl Acad Sci U S A*. 2010; 107:1223–8. [PubMed: 20133824]
154. Schmitt JE, et al. The dynamic role of genetics on cortical patterning during childhood and adolescence. *Proc Natl Acad Sci U S A*. 2014; 111:6774–9. [PubMed: 24753564]
155. van den Heuvel MP, et al. Genetic control of functional brain network efficiency in children. *Eur Neuropsychopharmacol*. 2013; 23:19–23. [PubMed: 22819192]
156. Silbereis JC, Pochareddy S, Zhu Y, Li M, Sestan N. The Cellular and Molecular Landscapes of the Developing Human Central Nervous System. *Neuron*. 2016; 89:248–68. [PubMed: 26796689]
157. Shibata M, Gulden FO, Sestan N. From trans to cis: transcriptional regulatory networks in neocortical development. *Trends Genet*. 2015; 31:77–87. [PubMed: 25624274]
158. Poretti A, Boltshauser E, Huisman TA. Congenital brain abnormalities: an update on malformations of cortical development and infratentorial malformations. *Semin Neurol*. 2014; 34:239–48. [PubMed: 25192502]
159. Qiu A, et al. COMT haplotypes modulate associations of antenatal maternal anxiety and neonatal cortical morphology. *Am J Psychiatry*. 2015b; 172:163–72. [PubMed: 25320962]
160. Chen L, et al. Brain-derived neurotrophic factor (BDNF) Val66Met polymorphism influences the association of the methylome with maternal anxiety and neonatal brain volumes. *Dev Psychopathol*. 2015; 27:137–50. [PubMed: 25640836]
161. Krishnan ML, et al. Integrative genomics of microglia implicates DLG4 (PSD95) in the white matter development of preterm infants. *Nat Commun*. 2017; 8:428. [PubMed: 28874660]
162. Xia K, et al. Genome-wide association analysis identifies common variants influencing infant brain volumes. *Transl Psychiatry*. 2017; 7:e1188. [PubMed: 28763065]
163. Johnson DS, Mortazavi A, Myers RM, Wold B. Genome-wide mapping of in vivo protein-DNA interactions. *Science*. 2007; 316:1497–502. [PubMed: 17540862]

164. Tabarki B, et al. Severe CNS involvement in WWOX mutations: Description of five new cases. *Am J Med Genet A*. 2015; 167A:3209–13. [PubMed: 26345274]
165. Krishnan ML, et al. Possible relationship between common genetic variation and white matter development in a pilot study of preterm infants. *Brain Behav*. 2016; 6:e00434. [PubMed: 27110435]
166. Brito NH, Noble KG. Socioeconomic status and structural brain development. *Front Neurosci*. 2014; 8:276. [PubMed: 25249931]
167. Farah MJ. The Neuroscience of Socioeconomic Status: Correlates, Causes, and Consequences. *Neuron*. 2017; 96:56–71. [PubMed: 28957676]
168. Luby J, et al. The effects of poverty on childhood brain development: the mediating effect of caregiving and stressful life events. *JAMA Pediatr*. 2013; 167:1135–42. [PubMed: 24165922]
169. Mackey AP, et al. Neuroanatomical correlates of the income-achievement gap. *Psychol Sci*. 2015; 26:925–33. [PubMed: 25896418]
170. Noble KG, et al. Family income, parental education and brain structure in children and adolescents. *Nat Neurosci*. 2015; 18:773–8. [PubMed: 25821911]
171. Hanson JL, et al. Family poverty affects the rate of human infant brain growth. *PLoS One*. 2013; 8:e80954. [PubMed: 24349025]
172. Ursache A, Noble KG. Pediatric Imaging N, Genetics S. Socioeconomic status, white matter, and executive function in children. *Brain Behav*. 2016; 6:e00531. [PubMed: 27781144]
173. Buss C, et al. Maternal cortisol over the course of pregnancy and subsequent child amygdala and hippocampus volumes and affective problems. *Proc Natl Acad Sci U S A*. 2012; 109:E1312–9. [PubMed: 22529357]
174. Lebel C, et al. Prepartum and Postpartum Maternal Depressive Symptoms Are Related to Children's Brain Structure in Preschool. *Biol Psychiatry*. 2016; 80:859–868. [PubMed: 26822800]
175. Rifkin-Graboi A, et al. Prenatal maternal depression associates with microstructure of right amygdala in neonates at birth. *Biol Psychiatry*. 2013; 74:837–44. [PubMed: 23968960]
176. Qiu A, et al. Prenatal maternal depression alters amygdala functional connectivity in 6-month-old infants. *Transl Psychiatry*. 2015a; 5:e508. [PubMed: 25689569]
177. Jha SC, et al. Antenatal depression, treatment with selective serotonin reuptake inhibitors, and neonatal brain structure: A propensity-matched cohort study. *Psychiatry Res*. 2016; 253:43–53.
178. Qiu A, et al. Maternal anxiety and infants' hippocampal development: timing matters. *Transl Psychiatry*. 2013; 3:e306. [PubMed: 24064710]
179. Graham AM, Pfeifer JH, Fisher PA, Carpenter S, Fair DA. Early life stress is associated with default system integrity and emotionality during infancy. *J Child Psychol Psychiatry*. 2015; 56:1212–22. [PubMed: 25809052]
180. Derauf C, Kekatpure M, Neyzi N, Lester B, Kosofsky B. Neuroimaging of children following prenatal drug exposure. *Semin Cell Dev Biol*. 2009; 20:441–54. [PubMed: 19560049]
181. Donald KA, et al. Neuroimaging effects of prenatal alcohol exposure on the developing human brain: a magnetic resonance imaging review. *Acta Neuropsychiatr*. 2015; 27:251–69. [PubMed: 25780875]
182. Salzwedel AP, Grewen KM, Goldman BD, Gao W. Thalamocortical functional connectivity and behavioral disruptions in neonates with prenatal cocaine exposure. *Neurotoxicol Teratol*. 2016; 56:16–25. [PubMed: 27242332]
183. Salzwedel AP, et al. Prenatal drug exposure affects neonatal brain functional connectivity. *J Neurosci*. 2015; 35:5860–9. [PubMed: 25855194]
184. Grewen K, Salzwedel AP, Gao W. Functional Connectivity Disruption in Neonates with Prenatal Marijuana Exposure. *Front Hum Neurosci*. 2015; 9:601. [PubMed: 26582983]
185. Gabrieli JD, Ghosh SS, Whitfield-Gabrieli S. Prediction as a humanitarian and pragmatic contribution from human cognitive neuroscience. *Neuron*. 2015; 85:11–26. [PubMed: 25569345]
186. Keunen K, et al. Brain Volumes at Term-Equivalent Age in Preterm Infants: Imaging Biomarkers for Neurodevelopmental Outcome through Early School Age. *J Pediatr*. 2016; 172:88–95. [PubMed: 26774198]

187. Gilmore JH, et al. Prenatal and neonatal brain structure and white matter maturation in children at high risk for schizophrenia. *Am J Psychiatry*. 2010b; 167:1083–91. [PubMed: 20516153]
188. Wolff JJ, et al. Differences in white matter fiber tract development present from 6 to 24 months in infants with autism. *Am J Psychiatry*. 2012; 169:589–600. [PubMed: 22362397]
189. Hazlett HC, et al. Early brain development in infants at high risk for autism spectrum disorder. *Nature*. 2017; 542:348–351. An important early example of how early imaging can be used to predict the development of autism. [PubMed: 28202961]
190. Graham AM, et al. Implications of newborn amygdala connectivity for fear and cognitive development at 6-months-of-age. *Dev Cogn Neurosci*. 2016; 18:12–25. [PubMed: 26499255]
191. Ghassabian A, et al. Infant brain structures, executive function, and attention deficit/hyperactivity problems at preschool age. A prospective study. *J Child Psychol Psychiatry*. 2013; 54:96–104. [PubMed: 22928649]
192. Herba CM, et al. Infant brain development and vulnerability to later internalizing difficulties: the Generation R study. *J Am Acad Child Adolesc Psychiatry*. 2010; 49:1053–63. [PubMed: 20855050]
193. Wee CY, et al. Neonatal neural networks predict children behavioral profiles later in life. *Hum Brain Mapp*. 2017; 38:1362–1373. [PubMed: 27862605]
194. Woo CW, Chang LJ, Lindquist MA, Wager TD. Building better biomarkers: brain models in translational neuroimaging. *Nat Neurosci*. 2017; 20:365–377. [PubMed: 28230847]
195. Reddan MC, Lindquist MA, Wager TD. Effect Size Estimation in Neuroimaging. *JAMA Psychiatry*. 2017; 74:207–208. [PubMed: 28099973]
196. Fjell AM, et al. Multimodal imaging of the self-regulating developing brain. *Proc Natl Acad Sci U S A*. 2012; 109:19620–5. [PubMed: 23150548]
197. Spann MN, Bansal R, Rosen TS, Peterson BS. Morphological features of the neonatal brain support development of subsequent cognitive, language, and motor abilities. *Hum Brain Mapp*. 2014; 35:4459–74. [PubMed: 24615961]
198. Short SJ, et al. Associations between white matter microstructure and infants' working memory. *Neuroimage*. 2013; 64:156–66. [PubMed: 22989623]
199. O'Muircheartaigh J, et al. White matter development and early cognition in babies and toddlers. *Hum Brain Mapp*. 2014; 35:4475–87. [PubMed: 24578096]
200. Deoni SC, et al. White matter maturation profiles through early childhood predict general cognitive ability. *Brain Struct Funct*. 2016; 221:1189–203. [PubMed: 25432771]
201. Emerson RW, et al. Functional neuroimaging of high-risk 6-month-old infants predicts a diagnosis of autism at 24 months of age. *Sci Transl Med*. 2017; 9
202. Smyser CD, et al. Prediction of brain maturity in infants using machine-learning algorithms. *Neuroimage*. 2016; 136:1–9. [PubMed: 27179605]
203. Ball G, et al. Machine-learning to characterise neonatal functional connectivity in the preterm brain. *Neuroimage*. 2016; 124:267–275. [PubMed: 26341027]
204. Bhardwaj RD, et al. Neocortical neurogenesis in humans is restricted to development. *Proc Natl Acad Sci U S A*. 2006; 103:12564–8. [PubMed: 16901981]
205. Sanai N, et al. Corridors of migrating neurons in the human brain and their decline during infancy. *Nature*. 2011; 478:382–6. [PubMed: 21964341]
206. Paredes MF, et al. Extensive migration of young neurons into the infant human frontal lobe. *Science*. 2016; 354
207. Conel, JL. *The Cortex of the Four-Year Child*. Harvard University Press; Cambridge, Massachusetts: 1963.
208. Huttenlocher PR, Dabholkar AS. Regional differences in synaptogenesis in human cerebral cortex. *J Comp Neurol*. 1997; 387:167–78. [PubMed: 9336221]
209. Petanjek Z, Judas M, Kostovic I, Uylings HB. Lifespan alterations of basal dendritic trees of pyramidal neurons in the human prefrontal cortex: a layer-specific pattern. *Cereb Cortex*. 2008; 18:915–29. [PubMed: 17652464]

210. Hasegawa M, et al. Development of myelination in the human fetal and infant cerebrum: A myelin basic protein immunohistochemical study. *Brain and Development*. 1992; 14:1–6. [PubMed: 1375444]
211. Kinney HC, Brody BA, Kloman AS, Gilles FH. Sequence of Central Nervous System Myelination in Human Infancy II. Patterns of Myelination in Autopsied Infants. *Journal of Neuropathology & Experimental Neurology*. 1988; 47:217–234. [PubMed: 3367155]
212. Abraham H, et al. Myelination in the human hippocampal formation from midgestation to adulthood. *Int J Dev Neurosci*. 2010; 28:401–10. [PubMed: 20417266]
213. Arnold SE, Trojanowski JQ. Human fetal hippocampal development: I. Cytoarchitecture, myeloarchitecture, and neuronal morphologic features. *Journal of Comparative Neurology*. 1996; 367:274–292. [PubMed: 8708010]
214. Benes FM, Turtle M, Khan Y, Farol P. Myelination of a key relay zone in the hippocampal formation occurs in the human brain during childhood, adolescence, and adulthood. *Arch Gen Psychiatry*. 1994; 51:477–84. [PubMed: 8192550]
215. Miller DJ, et al. Prolonged myelination in human neocortical evolution. *Proc Natl Acad Sci U S A*. 2012; 109:16480–5. [PubMed: 23012402]
216. Sigaard RK, Kjaer M, Pakkenberg B. Development of the Cell Population in the Brain White Matter of Young Children. *Cereb Cortex*. 2016; 26:89–95. [PubMed: 25122465]
217. Yeung MS, et al. Dynamics of oligodendrocyte generation and myelination in the human brain. *Cell*. 2014; 159:766–74. [PubMed: 25417154]
218. Alexander-Bloch A, et al. Subtle in-scanner motion biases automated measurement of brain anatomy from in vivo MRI. *Hum Brain Mapp*. 2016; 37:2385–97. [PubMed: 27004471]
219. Sairanen V, Kuusela L, Sipila O, Savolainen S, Vanhatalo S. A novel measure of reliability in Diffusion Tensor Imaging after data rejections due to subject motion. *Neuroimage*. 2017; 147:57–65. [PubMed: 27915115]
220. Power JD, Schlaggar BL, Petersen SE. Recent progress and outstanding issues in motion correction in resting state fMRI. *Neuroimage*. 2015; 105:536–51. [PubMed: 25462692]
221. Reuter M, et al. Head motion during MRI acquisition reduces gray matter volume and thickness estimates. *Neuroimage*. 2015; 107:107–15. [PubMed: 25498430]
222. Godenschweger F, et al. Motion correction in MRI of the brain. *Phys Med Biol*. 2016; 61:R32–56. [PubMed: 26864183]
223. Lerch JP, et al. Studying neuroanatomy using MRI. *Nat Neurosci*. 2017; 20:314–326. [PubMed: 28230838]
224. Weinberger DR, Radulescu E. Finding the Elusive Psychiatric "Lesion" With 21st-Century Neuroanatomy: A Note of Caution. *Am J Psychiatry*. 2016; 173:27–33. [PubMed: 26315983]
225. Paus T, et al. Maturation of white matter in the human brain: a review of magnetic resonance studies. *Brain Res Bull*. 2001; 54:255–66. [PubMed: 11287130]
226. Mukherjee P, Berman JI, Chung SW, Hess CP, Henry RG. Diffusion tensor MR imaging and fiber tractography: theoretic underpinnings. *AJNR Am J Neuroradiol*. 2008a; 29:632–41. [PubMed: 18339720]
227. Zhang H, Schneider T, Wheeler-Kingshott CA, Alexander DC. NODDI: practical in vivo neurite orientation dispersion and density imaging of the human brain. *Neuroimage*. 2012; 61:1000–16. [PubMed: 22484410]
228. Kunz N, et al. Assessing white matter microstructure of the newborn with multi-shell diffusion MRI and biophysical compartment models. *Neuroimage*. 2014; 96:288–99. [PubMed: 24680870]
229. Jelescu IO, et al. One diffusion acquisition and different white matter models: how does microstructure change in human early development based on WMTI and NODDI? *Neuroimage*. 2015; 107:242–56. [PubMed: 25498427]
230. Hutter J, et al. Time-efficient and flexible design of optimized multishell HARDI diffusion. *Magn Reson Med*. 2017
231. Dean DC 3rd, et al. Mapping White Matter Microstructure in the One Month Human Brain. *Sci Rep*. 2017; 7:9759. [PubMed: 28852074]

232. Deoni SC, Rutt BK, Arun T, Pierpaoli C, Jones DK. Gleaning multicomponent T1 and T2 information from steady-state imaging data. *Magn Reson Med*. 2008; 60:1372–87. [PubMed: 19025904]
233. Lankford CL, Does MD. On the inherent precision of mcDESPOT. *Magn Reson Med*. 2013; 69:127–36. [PubMed: 22411784]
234. Teixeira RP, Malik SJ, Hajnal JV. Joint system relaxometry (JSR) and Cramer-Rao lower bound optimization of sequence parameters: A framework for enhanced precision of DESPOT T1 and T2 estimation. *Magn Reson Med*. 2017
235. Wozniak JR, Lim KO. Advances in white matter imaging: a review of in vivo magnetic resonance methodologies and their applicability to the study of development and aging. *Neurosci Biobehav Rev*. 2006; 30:762–74. [PubMed: 16890990]
236. Jones DK, Cercignani M. Twenty-five pitfalls in the analysis of diffusion MRI data. *NMR Biomed*. 2010; 23:803–20. [PubMed: 20886566]
237. Jones DK, Knosche TR, Turner R. White matter integrity, fiber count, and other fallacies: the do's and don'ts of diffusion MRI. *Neuroimage*. 2013; 73:239–54. [PubMed: 22846632]
238. Mukherjee P, Chung SW, Berman JI, Hess CP, Henry RG. Diffusion tensor MR imaging and fiber tractography: technical considerations. *AJNR Am J Neuroradiol*. 2008b; 29:843–52. [PubMed: 18339719]
239. Dubois J, et al. Correction strategy for diffusion-weighted images corrupted with motion: application to the DTI evaluation of infants' white matter. *Magn Reson Imaging*. 2014b; 32:981–92. [PubMed: 24960369]
240. Berger H. Über das Elektrenkephalogramm des Menschen. *Archiv für Psychiatrie und Nervenkrankheiten*. 1929; 87:527–570.
241. Cohen D. Magnetoencephalography: detection of the brain's electrical activity with a superconducting magnetometer. *Science*. 1972; 175:664–6. [PubMed: 5009769]
242. Born P, Rostrup E, Leth H, Peitersen B, Lou HC. Change of visually induced cortical activation patterns during development. *Lancet*. 1996; 347:543. [PubMed: 8596290]
243. Born P, et al. Visual activation in infants and young children studied by functional magnetic resonance imaging. *Pediatr Res*. 1998; 44:578–83. [PubMed: 9773849]
244. Meek JH, et al. Regional hemodynamic responses to visual stimulation in awake infants. *Pediatr Res*. 1998; 43:840–3. [PubMed: 9621996]
245. Shi F, Salzwedel AP, Lin W, Gilmore JH, Gao W. Functional Brain Parcellations of the Infant Brain and the Associated Developmental Trends. *Cereb Cortex*. 2017:1–11.
246. Ciric R, et al. Benchmarking of participant-level confound regression strategies for the control of motion artifact in studies of functional connectivity. *Neuroimage*. 2017; 154:174–187. [PubMed: 28302591]
247. Bullins, J., Jha, SC., Knickmeyer, R., Gilmore, J. *Handbook of Preschool Mental Health: Development, Disorders, and Treatment*. Luby, JL., editor. The Guilford Press; New York: 2017. p. 73-97.
248. Smith SM, et al. Correspondence of the brain's functional architecture during activation and rest. *Proceedings of the National Academy of Sciences*. 2009; 106:13040–13045.

Key points

1. Adult patterns of regional cortical thickness appear to be present at birth, and cortical gray matter expands rapidly in the first year of life. Cortical thickness peaks between 1 and 2 years, while surface area expands through childhood.
2. White matter tracts are established before birth and postnatal myelination of these tracts matter occurs rapidly over the first 2 years of life. The white matter connectome is fairly mature at birth.
3. Sensorimotor resting state functional networks are present at birth, while higher order functional networks gradually emerge and develop over the first 2 years of life.
4. Studies have begun to explore genetic and environmental influences on early childhood brain development and the predictive value of early imaging biomarkers.
5. Future studies will need to better define normal and abnormal brain development in early childhood and determine if it is possible to identify early imaging biomarkers of later cognitive and behavioral outcomes.

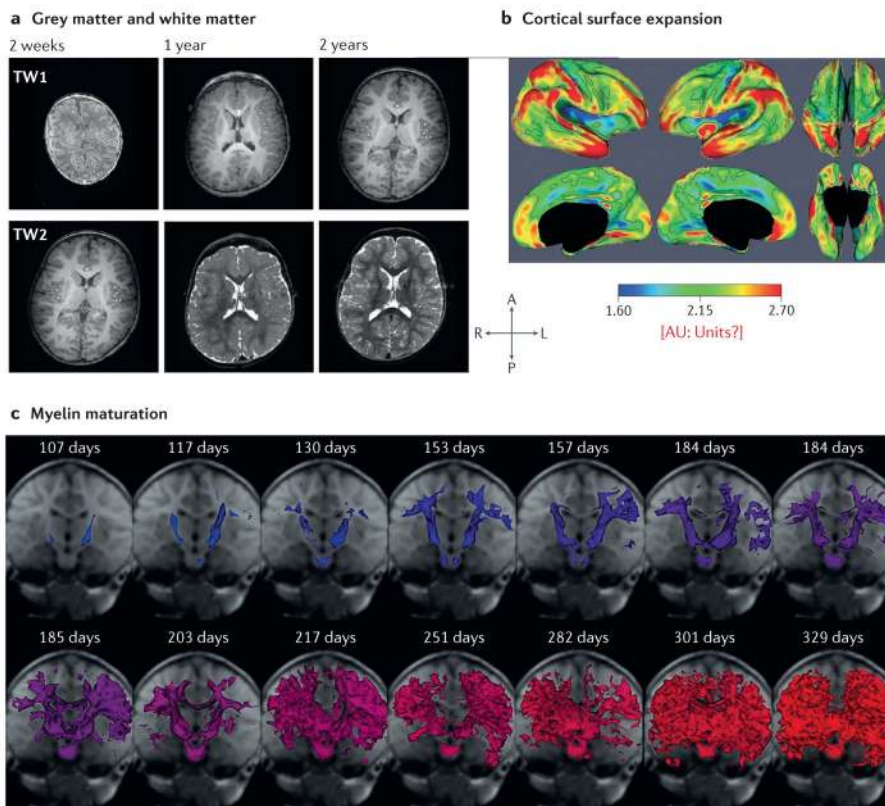


Figure 1. Structural brain development in early childhood

a | T1-weighted and T2-weighted images, birth to age 2 years. Note doubling of overall brain size between birth and 1 year with more gradual growth after age 1. In the neonate T1 scan, note that most white matter is not myelinated and therefore appears darker than cortical grey matter. Myelination proceeds rapidly in the first year of life; at 1 year and older, white matter assumes the typical white appearance seen in adults. This rapid change in tissue contrasts presents challenges for image analysis. Also note the relatively thin cortical grey-matter rim in the neonate T1 scan; by age 1, grey-matter thickness has increased significantly, reaching near-maximal thickness. White matter is more intense than grey matter at birth; this pattern is reversed by age 1 year. **b** | Regional expansion of cortical surface area from birth to 2 years derived from surface reconstructions of T2 (birth) and T1 (ages 1 and 2 years) scans, with greatest expansion in parietal, prefrontal and temporal regions. **c** | Myelin maturation in the first year of life imaged with mcDESPOt (see Box 2). Myelination begins in central white matter and spreads peripherally. Part **a** is adapted from REF.¹⁰. Part **b** is adapted from REF.²³. Part **c** is adapted from REF.³³.

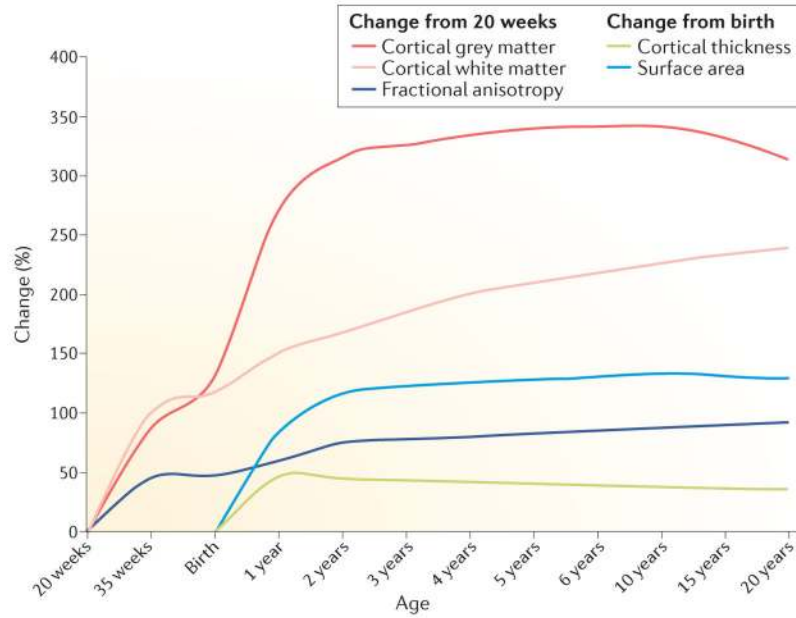


Figure 2. Estimated trajectories of brain structural parameters during development Relative growth trajectories (depicting percentage change from 20 weeks or neonate scan) in imaging studies of cortical grey and white matter, cortical thickness, surface area and fractional anisotropy (FA) from the prenatal or neonatal period to adulthood. After substantial prenatal development, there is robust postnatal growth of cortical grey-matter volumes in the first 2 years of life, and comparatively slower growth of cortical white-matter volume. FA, a measure of white-matter microstructure, also increases rapidly in the first year of life and much more gradually thereafter, consistent with trajectories of myelination. Cortical thickness peaks at 1–2 years of life and decreases gradually thereafter, whereas surface area develops rapidly in the first year of life and continues to expand thereafter, indicating that cortical volume growth observed after 1–2 years may be driven mainly by surface area expansion. Grey-matter volume and cortical thickness tend to decrease somewhat during adolescence (10–20 years), whereas white-matter volume and FA continue to increase through this period. Figure adapted from REF. ²⁴⁷.

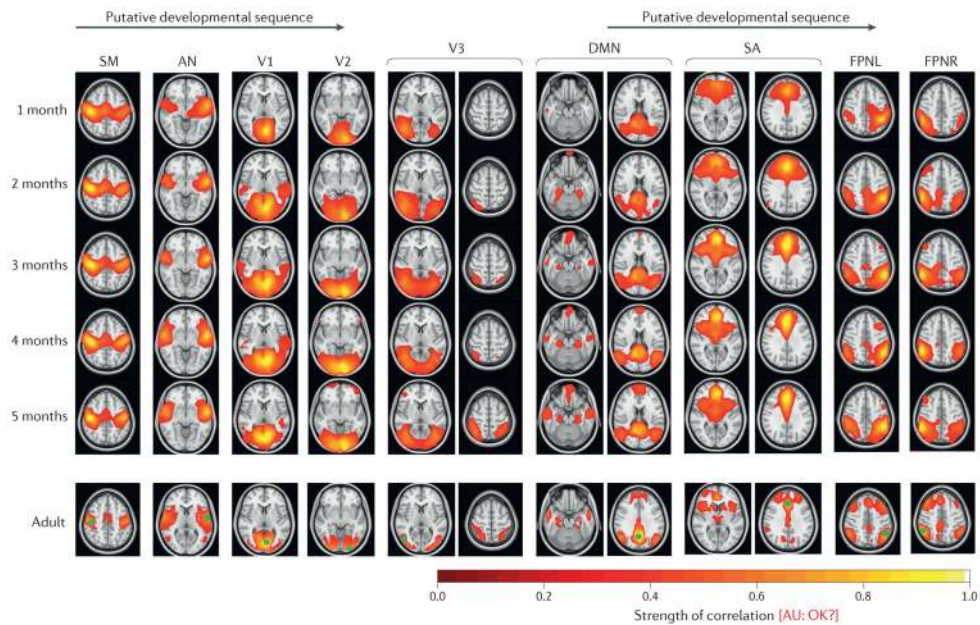


Figure 3. The development of resting-state functional networks during infancy

This grid of fMRI images depicts the growth of nine resting-state networks (RSNs) during the first year of life, with 3-month intervals between different time points, and in adulthood. Arrows at the top indicate the suggested developmental sequence of different RSNs based on the growth rates of relative degrees of similarity of their spatial topology with corresponding adult ones and the strength of within-network functional connectivity (for more details, see refs ^{90, 91}). Colour bar represents Pearson correlation strength between the BOLD signals of each brain voxel with the corresponding seed (green dots denoted in corresponding adult maps in the bottom row). The images are shown in radiological convention (that is, left side of brain on right side of image). All RSNs, including the two lateralized FPN networks (FPN L/R) were defined after Smith et al. ²⁴⁸ SM, sensorimotor network; AUD, auditory network; V1, visual 1 (medial visual) network; V2, visual 2 (occipital pole) network; V3, visual 3 (lateral visual) network; DMN, default-mode network; SA, salience network; FPN, frontoparietal executive control network. Figure adapted from REF. ⁹¹.