SOILS, SEC 3 • REMEDIATION AND MANAGEMENT OF CONTAMINATED OR DEGRADED LANDS • RESEARCH ARTICLE

Immobilization and phytotoxicity reduction of heavy metals in serpentine soil using biochar

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Abstract

Purpose Serpentine soils derived from ultramafic rocks release elevated concentrations of toxic heavy metals into the environment. Hence, crop plants cultivated in or adjacent to serpentine soil may experience reduced growth due to phytotoxicity as well as accumulate toxic heavy metals in edible tissues. We investigated the potential of biochar (BC), a waste byproduct of bioenergy industry in Sri Lanka, as a soil amendment to immobilize Ni, Cr, and Mn in serpentine soil and minimize their phytotoxicity.

Materials and methods The BC used in this study was a waste byproduct obtained from a Dendro bioenergy industry in Sri Lanka. This BC was produced by pyrolyzing *Gliricidia sepium* biomass at 900 °C in a closed reactor. A pot experiment was conducted using tomato plants (*Lycopersicon esculentum* L.) by adding 1, 2.5, and 5 % (w/w) BC applications to evaluate the bioavailability and uptake of metals in serpentine soil. Sequential extractions were utilized to evaluate the effects of BC on bioavailable concentrations of Ni, Cr, and Mn as well as different metal fractionations in BCamended and BC-unamended soil. Postharvest soil in each

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Unit for Environmental Sciences and Management, North-West University, Private Bag X6001, Potchefstroom 2520, South Africa pot was subjected to a microbial analysis to evaluate the total bacterial and fungal count in BC-amended and BC-unamended serpentine soil.

Results and discussion Tomato plants grown in 5 % BCamended soil showed approximately 40-fold higher biomass than that of BC-unamended soil, whereas highly favorable microbial growth was observed in the 2.5 % BC-amended soil. Bioaccumulation of Cr, Ni, and Mn decreased by 93– 97 % in tomato plants grown in 5 % BC-amended soil compared to the BC-unamended soil. Sequentially extracted metals in the exchangeable fraction revealed that the bioavailabile concentrations of Cr, Ni, and Mn decreased by 99, 61, and 42 %, respectively, in the 5 % BC-amended soil. *Conclusions* Results suggested that the addition of BC to serpentine soil as a soil amendment immobilizes Cr, Ni, and Mn in serpentine soil and reduces metal-induced toxicities in tomato plants.

Keywords Bioavailability · Chemisorption · Metal immobilization · Sequential extraction · Serpentine

1 Introduction

Heavy metal contamination of soil due to anthropogenic and non-anthropogenic activities is a widespread environmental problem with serious consequences for agricultural crop productivity and human health (Neilson and Rajakaruna 2014). Ultramafic rocks and related soils and sediments are nonanthropogenic sources of metal contamination (Rajakaruna and Baker 2004). Serpentine soils, derived from the weathering of ultramafic rocks, may release elevated concentrations of heavy metals such as nickel (Ni), chromium (Cr), manganese (Mn), and cobalt (Co) into the environment (Rajapaksha et al. 2012; Vithanage et al. 2014a). In parts of northwestern Spain (Fernández et al. 1999), Canada (Baugé et al. 2013), Philippines (Susaya et al. 2010), and Japan (Kayama et al. 2002), where serpentine-associated soils are often subjected to agriculture and forestry, soils are intensively managed to make them amenable for plant growth.

Although these practices have allowed the growth of some plants, those growing on serpentine soils can accumulate high concentrations of toxic metals such as Cr, Ni, and Mn in their edible parts (Fernández et al. 1999; Susaya et al. 2010). Hence, the cultivation of crop plants in areas within and adjacent to serpentine outcrops and other heavy metalenriched sites may be of particular concern due to both phytotoxicity and metal accumulation (Neilson and Rajakaruna 2014). The prolonged consumption of metal-accumulating plants such as Zea mays L. (Almaroai et al. 2014), Allium sativum L. (Jiang et al. 2001), and Brassica napus L. (Houben et al. 2013a) may pose serious health risks when their consumption leads to concentrations above the toxicity threshold (Anderson et al. 2005), even for micronutrient metals such as Cu, Mn, and Zn. Therefore, the restoration of such heavy metal rich soils using novel and economically feasible technologies is an urgent necessity before they are used in agriculture (Neilson and Rajakaruna 2014; Rajakaruna et al. 2006).

Because of the drawbacks of conventional soil remediation technologies such as soil replacement, solidification, electrokinetic extraction, and washing strategies (Mulligan et al. 2001; Sruthy and Jayalekshmi 2014), a considerable interest has been expressed in the use of a variety of biowastes such as woodchips, peanut shells, chicken manure, cow bone, eggshell, and poultry manure to remediate heavy metalcontaminated soils (Almaroai et al. 2014; Usman et al. 2013). Recently, carbon-rich amendments such as biochar (BC) have been used as an alternative and economically viable strategy to immobilize bioavailable toxic metals and reduce the phytotoxicity of heavy metals in contaminated soils (Almaroai et al. 2014; Houben et al. 2013a; Houben et al. 2013b).

BC is a carbon-rich product that is produced by pyrolyzing biowaste materials. It is capable of improving physical, chemical, and biological properties in soils due to its high organic carbon content (Almaroai et al. 2014). The application of BC as a soil amendment leads to increases in soil fertility and enhances plant growth by supplying and retaining essential nutrients while improving soil physical and biological properties (Houben et al. 2013b). The positive effects of BC on the immobilization of bioavailable heavy metals in contaminated soils have been investigated under greenhouse conditions (Houben et al. 2013a; Houben et al. 2013b; Park et al. 2011; Uchimiya et al. 2011). A commercial grade BC produced from Miscanthus straw has been successfully applied to contaminated soils for investigating the bioavailability of Cd, Pb, and Zn and the production of rapeseed (B. napus L.) biomass (Houben et al. 2013b). This study found that the addition of BC as a soil amendment reduces the bioavailability of Cd. Pb. and Zn and provides a viable alternative and a cost-effective strategy to promote revegetation and restoration of heavy metal-contaminated lands as well as the growth of agricultural crops. Arsenic (As) concentrations in soil, soil pore water, and plant tissues were evaluated in a pot experiment following the transplantation of tomato (Solanum lycopersicum L.) plants to a heavily As-contaminated mine soil amended with an orchard prune residue BC (Beesley et al. 2013). Biochar significantly increased As concentrations in soil pore water, while root and shoot concentrations were significantly reduced compared to the control without BC. Fruit As concentrations were very low, indicating minimal toxicity and transfer risk. In a study conducted by Park et al. (2011), application of BC derived from green waste to shooting range and spiked soils significantly immobilized and reduced the phytoavailability of Cd, Pb, and Cu in Indian mustard (Brassica juncea (L.) Czern.) plants. Moreover, a recent study demonstrated that bur cucumber-derived BC was effective in reducing the mobility of sulfamethazine (SMZ) in agricultural soils (Vithanage et al. 2014b). Similar to the benefits, the limitations and draw backs of the use of BC for the remediation of contaminated soils are also of particular concern. Although in short-term experiments of months to a few years, BC could be seen to enhance plant growth and soil nutrient status, neither the quantitative variability in response nor the durability of the effects is specified. Moreover, the addition of BC to soil in excessive levels may cause detrimental impacts on soil structure and pose serious consequences for the growth of crop plants. Hence, the potential limitations and drawbacks of the use of BC may be encountered at large-scale and long-term deployment in field.

The presence of excessive metal content in soils surrounding disturbed serpentine and other naturally metal-enriched systems may have serious consequences for groundwater and agricultural productivity; however, not many attempts have been made to remediate such sites for effective cultivation. Remediation of sites adjacent to metal-enriched settings can provide much-needed land for the cultivation of plants to use as food or animal feed. Hence, this study investigated the potential of waste woody BC from a bioenergy plant as a soil amendment to immobilize bioavailable toxic metals and reduce the phytotoxicity of heavy metals. Specifically, we examined whether this woody BC is capable of effectively immobilizing and reducing the phytotoxicity of Ni, Cr, and Mn in serpentine soil. We assessed how the application of different rates of BC to serpentine soil can influence the immobilization of Ni, Cr, and Mn found in serpentine soil and thereby reduce phytotoxicity in tomato plants (Lycopersicon esculentum L; Solanaceae) grown under greenhouse conditions. Tomato plants were selected for this study as this species has received much attention due to its cultivation as a crop plant in areas adjacent to metal-enriched serpentine outcrops in Sri Lanka.

2 Materials and methods

2.1 Serpentine soil

Serpentine soil was collected from the top 0–15 cm of soil from the Wasgamuwa area (Yudhaganawa serpentine outcrop), Sri Lanka (Rajapaksha et al. 2012; Vithanage et al. 2014a) for use in this study. Soil samples were air-dried and mechanically sieved to <2 mm of particle size prior to use in the laboratory and greenhouse experiments. Basic properties of this serpentine soil are summarized in Table 1.

2.2 Biochar

The BC used in this study was a waste byproduct from a bioenergy industry (Dendro) at Thiruppane, Anuradhapura District, Sri Lanka. This BC was produced by pyrolyzing the woody biomass (BM), *Gliricidia sepium* (Jacq.) Steud. (Fabaceae) in a closed reactor. The end temperature of this process was recorded as 900 °C at which *Gliricidia* wood is gasified for the generation of electricity. The BC obtained from this power plant was air-dried and ground in a blender and sieved to <1 mm of particle size prior to use in the experiments.

2.3 Biochar characterization

All chemicals used were analytical grade and purchased from Fluka (Switzerland) or Sigma (USA). This woody BC was tested for pH, electrical conductivity (EC), ash, moisture, volatile matter content, total organic carbon (TOC), cation exchange capacity (CEC), elemental composition, surface area, and Fourier transform infrared (FTIR) spectra. pH and EC in BC were measured in 1:5 suspensions of BC-to-water

Table 1 Total and exchangeable metal concentrations of serpentine soil

Parameter	Value
Total metal digestion (mg kg ⁻¹)	
Ni	6,567±3.08
Cr	14,880±6.14
Mn	2,609±2.83
Exchangeable Ni (mg kg ^{-1})	
With distilled water	$7.4{\pm}0.07$
With 0.01 CaCl ₂	211.9±0.16
Exchangeable Cr (mg kg ^{-1})	
With distilled water	ND
With 0.01 CaCl ₂	12.5 ± 0.03
Exchangeable Mn (mg kg^{-1})	
With distilled water	$0.79 {\pm} 0.05$
With 0.01 CaCl ₂	11.2±0.14

using a digital pH meter (702SM Titrino, Metrohm) and an electrical conductivity meter (Orion 5 star meter, Thermo Scientific), respectively. Exchangeable Ca²⁺, Mg²⁺, Na⁺, and K⁺ were extracted via the ammonium acetate procedure (Summer and Andersen 1996) and concentrations of metals were determined using an atomic absorption spectrophotometer (AAS, GBC933, Australia). The organic matter content was determined following the Walkley-Black method (Mebius 1960). Total N and other elements including C, O, H, and S were analyzed using a CHN analyzer (Vario MAX CN, elementar, Hanau, Germany). The surface area of BC was determined following the BET method (Peterson et al. 2012). The FTIR spectra of vacuum dried sample pellets, prepared with fused-KBr, were obtained with a resolution of 1 cm^{-1} between 4,000 and 400 cm^{-1} (Nicolet 6700, USA). The spectra were analyzed using OMNIC version 8.0 software, and the spectral data of Gliricidia BM and its BC derived at 900 °C are shown in Fig. 1. Moisture was determined by calculating the weight loss after heating the BC at 105 °C for 24 h to a constant weight. Mobile matter (analogous to volatile matter), which reflects the non-carbonized portion in BC, was determined as the weight loss after heating in a covered crucible at 450 °C for 30 min (Ahmad et al. 2014). Ash content was also measured as the residue remaining after heating at 700 °C in an open-top crucible. The portion of the BC not ashed, referred to as resident matter (analogous to fixed matter), was calculated by the difference in moisture, ash, and mobile matter. Each sample was analyzed in triplicate. Table 2 summarizes the data obtained from proximate and ultimate analyses for this high temperature-derived BC.

The surface area of this woody BC was 714 m² g⁻¹ (Table 2) which is extremely high compared to the surface area of soybean stover BC (420.3 m² g⁻¹), and peanut shell BC (448.2 m² g⁻¹) produced at 700 °C (Ahmad et al. 2012). Elemental analyses revealed that the low molar O/C ratio is due to high pyrolytic temperature as the surfaces of hightemperature BC become less hydrophilic (Ahmad et al. 2012). Atomic ratios H/C and [(O+N)/C] are recognized as indices for aromaticity and polarity of BC, respectively (Ahmad et al. 2013; Chen et al. 2008). Lower values of both H/C and the polarity index [(O+N)/C] ratios of the BC indicated that the high temperature-derived BC are highly carbonized, exhibiting a highly aromatic structure (Ahmad et al. 2013). The reduction of O/C and H/C ratios is mainly due to dehydratation, decarboxylation, and decarbonylation processes during pyrolysis at higher temperatures (Sun et al. 2012). Furthermore, the concentrations of N and S were quite low, implying that even if they were to be burnt as the feedstock for BC production, they may give off only low rates of nitrogen oxide and sulfur oxide to the environment.

In the FTIR spectra (Fig. 1), the broad peak observed at $3,356 \text{ cm}^{-1}$ in BM corresponds to the stretching vibration of





-OH group, indicating the presence of bonded water (Ahmad et al. 2014; Chen et al. 2008). The band intensity is markedly reduced in BC due to the elimination of water molecules and other volatile functional groups as a result of higher temperature. The peaks at 2,919, 1,463, and 1,375 cm^{-1} in BM are assigned mainly to CH₂ stretching vibration, suggesting the presence of long-linear aliphatic groups in BM which are completely eliminated in BC (Chen et al. 2008). This suggests the removal of nonpolar functional groups during pyrolysis. The band at 1.635 cm^{-1} corresponds to the lignin content of BM, and its disappearance in BC suggests the loss of the original plant structure of Gliricidia (Chen et al. 2008). Peaks at 1,736 and 1,510 cm⁻¹ in BM are assigned to C=O stretching in the aromatic ring and C=C ring stretching vibration of lignin, respectively (Chen et al. 2008). The peak of 875 cm^{-1} in BC is assigned to the aromatic CH out-of-plane deformation condensing smaller aromatic units into larger sheets (Ahmad et al. 2014; Chen et al. 2008). The band at 1,264 cm⁻¹ indicates the aromatic CO- and phenolic-OH stretching, and it is completely eliminated in BC (Chen et al. 2008). The bands due to aliphatic C–O–C and alcohol–OH (1,161–1,034 cm⁻¹) in BM indicate oxygenated functional groups (Chen et al. 2008). Appearance of new peaks at 1,431 and 1,054 cm⁻¹ in BC are assigned to CO_3^{-2} and =S=O (sulfoxides), respectively. The overall interpretation of the spectra suggested the dehydration of cellulosic and ligneous contents in the BM and an increase in the condensation of aromatic units in the BC produced at a high temperature.

2.4 Pot experiment

Untreated soil (control) and soil treated with three rates of BC applications were used in the experiment. Soil was amended by mixing dry BC with a mass fraction of 1, 2.5, and 5 % (w/w). Amended soils were thoroughly homogenized in large plastic containers prior to use. Plastic pots (12.0 cm diameter, 9.0 cm height) were filled with ~250 g of BC-amended and BC-unamended soil and 100 g each of washed sand (to

Proximate an	alysis							
рН		Moisture (%)		Mobile matter (%)		Ash (%)		Resident matter (%)
$10.10 {\pm} 0.07$		$6.50{\pm}0.05$		$9.90{\pm}0.03$		$70 {\pm} 0.01$		63.±0.18
Ultimate anal	lysis							
C (%) 50±0.12	H (%) 1±0.05	O (%) 44±0.02	N (%) 0.5±0.01	S (%) 0.1±0.03	Molar H/C 0.02±0.001	Molar O/C 0.88±0.002	Molar [(O+N)/C] 0.89±0.002	BET Surface area/m ² g ⁻¹ 714.00 \pm 0.85

Table 2 Proximate and ultimate analyses data for the BC derived at 900 °C

prevent soil compaction). The pots were then placed in a dark room for the soil mixture to equilibrate over 2 weeks. Each treatment was performed in triplicate. After the equilibration period, the pots were transferred to an outdoor greenhouse and arranged in a randomized manner. In each pot, 25 seeds of tomato were sown and plants were grown for 9 weeks from June to August 2013. All pots were irrigated with 30 cm³ of tap water three times per week.

2.5 Plant tissue analysis

From each treatment, three whole plants per pot were harvested at six successive intervals from fourth to ninth week from germination. Whole plants were used as it was difficult to obtain enough biomass for metal analyses by separating shoots and roots. Harvested plants were thoroughly washed with tap water and rinsed with deionized water. After air drying in a forced draft oven at 60 °C for 48 h and subsequent cooling in a desiccator, the dry weight of each plant was measured. The total amount of Cr, Ni, and Mn in plant tissue was analyzed by AAS after digestion with 10 cm³ of concentrated HNO₃ acid in a closed-vessel temperature-controlled microwave digester system (Milestone ETHOS PLUS labstation with HRP-1000/10S high pressure segmented rotor).

2.6 Soil analysis

2.6.1 Sequential extraction

Sequential extractions involved the selective extraction of trace metals from soil solid fractions, providing detailed information on the availability of heavy metals among the different geochemical phases in soil (Paz-Ferreiro et al. 2013). Ideally, the extractants are chosen to selectively target a specific soil compartment with minimal dissolution of nontargeted phases. The sequential extraction procedure was carried out on both control and postharvest soils following the procedure described by Armienta et al. (1996). A mass of 1 g of soil (dry weight) was used for the initial extraction. A total of five replicate sequential extraction analyses were completed on the four soil treatments. The concentrations of Ni, Cr, and Mn were measured in the effluent after each extraction using AAS. Below is a list of the extraction procedures performed on the soil amendments.

- (i) *Exchangeable*: Soil was reacted at room temperature for 1 h with 20 cm³ of magnesium chloride solution (1 M MgCl₂, pH 7.0) with continuous agitation.
- (ii) Bound to carbonates: Residue from (i) was leached at room temperature for 2 h with 20 cm³ of 1 M sodium acetate (NaOAc) adjusted to pH 5.0 with acetic acid (HOAc) and with continuous agitation.

- (iii) Bound to Fe–Mn oxide: Residue from (ii) was treated with 20 cm³ of 0.04 M hydroxylamine hydrochloride (NH₂OH–HCl) in 25 % (ν/ν) HOAc heated at 90 °C with slow continuous agitation for 2 h.
- (iv) Bound to organic matter: Residue from (iii) was treated with 3 mL of 0.02 M HNO₃ and 5 cm³ of 30 % H₂O₂ adjusted to pH 2 with HNO₃, heated to 85 °C for 2 h with occasional agitation. A 3 cm³ aliquot of 30 % H₂O₂ (pH 2 with HNO₃) was added, and the sample was heated again to 85 °C for 3 h with intermittent agitation. After cooling, 5 cm³ of 3.2 M NH₄OAc in 20 % (ν/ν) HNO₃ was added and the sample was diluted to 20 cm³ and agitated continuously.
- (v) Residual: Residue from (iv) was treated with a mixture of 10 cm³ concentrated HF and 2 cm³ concentrated HClO₄ and heated to near dryness. It was then treated with 1 cm³ of HClO₄⁺, 10 cm³ of HF and heated again to near dryness; 1 cm³ HClO₄ was added, heated until the appearance of white fumes, and finally dissolved with 12 N HC1 and diluted to 25 cm³ with deionized water.

Between each successive extraction listed above [(ii) to (v)], the sample was centrifuged at 3,500 rpm for 15 min. Additionally, the supernatant was filtered using 0.45- μ m filter paper prior to AAS analysis.

2.7 Microbial analysis

The postharvest soil in each pot was subjected to a microbial analysis to evaluate the effects of BC on the total microbial count in BC-amended and BC-unamended serpentine soil. A dilution series was prepared from 10^{-1} to 10^{-5} , and each dilution was plated in nutrient agar (NA) and potato dextrose agar (PDA) media to determine the number of bacteria and fungi in soil, respectively. The NA and PDA plates were incubated at 37 °C in 24 and 72 h, respectively. Finally, the number of colony forming units (CFU) was counted (Onipe and Adebayo 2011).

2.8 Statistical analyses

Statistical analyses were carried out to compare how different BC application rates were influencing the growth of plants and the accumulation of metals in plant tissues by using a one-way analysis of variance (ANOVA) followed by Fisher's test (p<0.05) for multiple comparisons. Mean separation procedure (least significant different test) and group comparison-contrast were used after performing the ANOVA for randomized complete block designs (RCBD). All statistical analyses were carried out using Statistical software package (SAS 9.1).

3 Results and discussion

3.1 Influence of biochar amendments on changes in soil pH, EC, TOC, and CEC

The pH, EC, TOC, and CEC of serpentine soil increased significantly with the increasing concentration of BC amendment (Table 3). The increase in soil pH after the addition of BC is due to the alkali nature of the BC. The dissolution of oxides, hydroxides, and carbonates of alkaline substances, decarboxylation of organic anions, and the ammonification of the soil provide nutrients for plant growth (Houben et al. 2013b) as well as the increase in soil pH may minimize the bioavailability of heavy metals in soil (Gall and Rajakaruna 2013; Neilson and Rajakaruna 2014). The TOC also increased significantly with the application of BC due to the addition of high carbon into the soil. The content of TOC in 5 % BC-amended soil was twice as much as in the BC-unamended soil (Table 3).

The addition of BC significantly increased the soil CEC. This increase of soil CEC in the presence of BC is corroborated by earlier findings (Houben et al. 2013b; Karami et al. 2011). A significant increase in available nutrients was observed with increasing BC application rates (Table 3). Calcium increased the most after the addition of BC to serpentine soil. Given that Ca is often considered to be the most limiting nutrient for plant growth in serpentine soil (O'Dell and Rajakaruna 2011; Rajakaruna et al. 2012), this observed increase in response to BC application is worthy of note. The available amounts of K, Ca, and Mg in the 5 % BC amendment were multiplied by 1.2-2.0-fold compared to the BC-unamended soil. The increase in available nutrient content with increasing BC application rates results from free bases such as $K^{\scriptscriptstyle +},\ Ca^{2+},\ and\ Mg^{2+}$ present in BC releasing into the soil solution, thereby increasing the pH of the soil and providing readily available nutrients for plant growth (Houben et al. 2013a).

3.2 Effects of biochar on the growth of plants

Tomato plants grown in BC-unamended serpentine soil displayed low biomass production compared to the plants that had received 1, 2.5, and 5 % BC application rates (Fig. 2). Three weeks after sowing, signs of metal toxicity in the aboveground parts of tomato plants were apparent in the BC-unamended serpentine soil, and after the first harvest, plants were not able to survive any longer in the BC-unamended soil. The common diagnostic symptoms of heavy metal phytotoxicity such as leaf chlorosis, necrosis, leaf epinasty, and growth retardation were displayed in plants that grew in the BC-unamended soil. Such symptoms are usual for tomato plants exposed to heavy metal stress, and metal toxicity symptoms have been previously documented (Moral et al. 1995).

Figure 3 depicts the variation of dry biomass of tomato plants grown in BC-amended and BC-unamended serpentine soil across successive harvests. There was a significant increase in the biomass of tomato plants with increasing BC concentrations across consecutive harvests (Table 4). The biomass of tomatoes grown in BC-unamended serpentine soil was reduced by 97 % in the second harvest compared to the plants that had received 5 % BC application. This biomass reduction could be due to the bioaccumulation of heavy metals and the resulting phytotoxicity inhibiting the growth of plants. Overall, 5 % BC application in serpentine soil resulted in increasing the biomass of tomato plants by 40fold compared to the BC-unamended soil. Such an increase in biomass production of tomato plants after addition of BC to serpentine soil could be due to both its fertilizing effect and the immobilization of heavy metals in serpentine soil. However, the plant tissues analyses data (Fig. 5) demonstrated that the addition of BC to serpentine soil had mostly contributed to the promotion of plant growth while enhancing the fertilizing effects. Moreover, these findings agree with those of recent studies (Houben et al. 2013a) confirming higher plant productivity when BC is applied, likely resulting from the immobilization of heavy metals in contaminated soil.

Table 3	Effects of different BC	application rates of	n changes in pH,	EC, TOC, and C	CEC of serpentine soil
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		Control	1 % B+S	2.5 % B+S	5 % B+S
pН		5.5±0.06d	6.3±0.08c	6.5±0.03b	7.6±0.01a
EC	$\mu S \ cm^{-1}$	120.1±0.12a	71.4±0.08d	94.9±0.15c	$100.1 \pm 0.17b$
TOC	mg g^{-1}	115.0±0.05d	129.5±0.11c	$148.4 {\pm} 0.18b$	224.3±0.15a
CEC	$cmol kg^{-1}$	3.4±0.08d	3.7±0.01c	4.0±0.07b	4.3±0.03a
Available K	mg kg^{-1}	650.0±0.17d	777.7±0.08c	1,044.3±0.14b	1,294.3±0.09a
Available Ca	mg kg^{-1}	2,527.5±0.06d	3,721.9±0.16a	3,633.0±0.07b	3,521.9±0.19c
Available Mg	mg kg^{-1}	899.3±0.11d	1,037.8±0.15c	1,215.7±0.08a	1,087.8±0.16b

Control—BC-unamended soil, 1 % B+S—1 % BC-amended soil, 2.5 % B+S—2.5 % BC-amended soil, 5 % B+S—5 % BC-amended soil. For each parameter, column means with the *same letter* do not differ significantly at the 5 % level according to the Fisher's multiple-comparison test



Fig. 2 Representative image showing differences in tomato plant growth 4 weeks after sowing. From left to right: BC-unamended soil, 1 % BC-amended soil, 2.5 % BC-amended soil, and 5 % BC-amended soil. Only one of the three replicates is shown for each treatment

3.3 Effects of biochar on microbial growth

The influence of BC as a soil amendment on changes in microbial quantity in serpentine soil is also of particular concern. No considerable difference was observed for the amount of bacteria and fungi between 1 % BC application and control. The maximum number of CFU was found in 2.5 % BC application, whereas 5 % BC showed a reduction in CFU (Fig. 4). This positive effect may be attributed to a general improvement in physical and chemical characteristics of the 2.5 % BC application. The pores in BC may provide a suitable habitat for microorganisms by protecting them from predation and drying while providing nutrients (Janice and Rillig 2009). Several recent studies have shown that BC is capable of stimulating the activity of agriculturally important soil microorganisms (Pietikäinen et al. 2000). Even though the increment of BC application increases the quality of soil

physicochemical properties (Janice and Rillig 2009), infinite increase in BC application would not be beneficial for the biological activities in soil. Hence, a reduction in the CFU was observed in 5 % BC application. This could be due to the changes in soil structural properties and the adsorption of extracellular enzymes and inorganic nutrients to BC surface (Janice and Rillig 2009). Moreover, high cation exchange capacity and high density of functional groups in the 5 % BC amendment may enhance the binding capacity of important macronutrients such as nitrogen and phosphorus, negatively impacting microbial growth (Gomez et al. 2014).

3.4 Effects of biochar on uptake of heavy metals in Tomato plants

The effects of BC on bioaccumulation of Cr, Ni, and Mn in tomato plants grown in BC-amended and BC-unamended



	Treatment	Biomass per plant/mg	Total metal concentration (mg kg ⁻¹)			
			Ni	Cr	Mn	
1st harvest	Control	1.7±0.6c	1,646.2±73.8a	832.9±21.4a	258.0±20.3a	
	1 % B+S	23.6±1.1b	293.8±20.4b	88.7±9.7b	185.6±12.8b	
	2.5 % B+S	30.3±6.3b	238.6±21.0b	39.6±22.7c	99.3±19.8c	
	5 % B+S	46.0±7.2a	131.4±26.0c	10.8±0.9c	16.7±4.3d	
2nd harvest	Control	1.9±0.6c	1,898.8±58.0a	3,087.3±157.0a	813.0±82.6a	
	1 % B+S	29.5±4.7b	323.3±33.0b	337.7±56.3b	267.0±30.4b	
	2.5 % B+S	35.4±9.0b	243.1±16.4c	204.0±2.4bc	155.7±21.0c	
	5 % B+S	56.3±9.4a	132.4±14.6d	88.3±12.0c	58.3±9.7d	
3rd harvest	1 % B+S	34.5±7.2b	306.7±15.6a	549.0±43.9a	162.6±22.2a	
	2.5 % B+S	45.7±12.5b	203.3±21.3b	235.3±51.1b	130.7±5.0b	
	5 % B+S	67.0±6.8a	137.7±24.4c	121.0±43.0c	80.3±3.9c	
4th harvest	1 % B+S	51.4±9.4b	354.3±33.6a	169.3±19.8a	172.3±12.0a	
	2.5 % B+S	61.0±9.5ab	257.3±31.4b	103.7±10.1b	140.6±3.7b	
	5 % B+S	69.7±7.2a	134.0±12.0c	54.7±8.0c	73.3±17.1c	
5th harvest	1 % B+S	61.7±5.6b	386.0±26.2a	304.0±60.2a	203.3±23.1a	
	2.5 % B+S	69.2±3.2ab	268.7±41.1b	256.7±25.1a	181.3±16.1a	
	5 % B+S	78.0±8.9a	139.9±11.2c	81.0±9.8b	96.6±6.1b	
6th harvest	1 %+S	70.4±4.5b	382.2±9.4a	347.0±21.0a	184.6±2.0a	
	2.5 % B+S	75.7±1.4ab	255.5±26.3b	232.3±31.4b	115.6±13.8b	
	5 % B+S	83.6±8.4a	142.7±5.4c	106.0±17.9c	70.0±7.5c	

 Table 4
 Comparison of the biomass of tomato plants and accumulated concentrations of heavy metals in plant tissues due to the influence of different BC application rates

Control—BC-unamended soil, 1 % B+S—1 % BC-amended soil, 2.5 % B+S—2.5 % BC-amended soil, 5 % B+S—5 % BC-amended soil. For each harvest, column means with the *same letter* do not differ significantly at the 5 % level according to the Fisher's multiple-comparison test

serpentine soils across six successive harvests are depicted in Fig. 5. Consecutive harvesting was conducted on monitoring metal ions to assess the variation of accumulated

concentrations of heavy metals in plant tissues with respect to growing time. All BC applications significantly reduced the uptake and bioaccumulation of Cr, Ni, and Mn across all

Fig. 4 Total number of bacteria and fungi per gram of soil in different BC application rates. For each harvest, means topped with the *same letter* do not differ significantly at the 5 % level according to the Fisher's multiple-comparison test



Fig. 5 Accumulated concentrations of a Ni, b Mn, and c Cr in tomato plants grown on BC-amended and BC-unamended serpentine soil across six consecutive harvests. For each harvest, means topped with the *same letter* do not differ significantly at the 5 % level according to the Fisher's multiple-comparison test



harvests (Table 4), and maximum accumulated concentrations of metals were found in tomato plants grown in BCunamended soil. Compared to the control, 1 % BC treatment reduced the accumulation of Cr, Ni, and Mn by 83, 89, and 67 %, respectively. In the presence of 2.5 % BC treatment, the reduction percentage increased up to 87, 93, and 81 %, whereas in the presence of 5 % BC treatment, it increased, reaching up to 93, 97, and 92 %, respectively. Overall, the bioaccumulation of Cr, Ni, and Mn in tomato plants grown in BCunamended soil was 14-35-fold higher than that of 5 % BCamended soil. Because of such high bioaccumulation of Cr, Ni, and Mn, tomato plants grown in BC-unamended serpentine soil were not able to survive the duration of the experiment due to metal-induced phytotoxicity. Meanwhile, the accumulation of these metals in tomato plants with consecutive harvesting is also of particular concern in order to assess the variation of bioaccumulation as a function of growing time. Although the accumulation of Ni in plants grown in BC-unamended soil in the second harvest was almost same as the first harvest, the accumulated concentrations of Cr and Mn were 3-4-fold higher than that of in the first harvest. Whereas, the bioaccumulation of these metals in plants that grew in BCamended soil was not significant (p>0.05) with consecutive harvests. Hence, this suggests that the bioaccumulation of Cr and Mn in tomato plants tends to increase with time, and Cr and Mn could have the most detrimental effect on growth of tomato plants in BC-unamended soil.

The permissible limit values of Ni and Cr in plants recommended by WHO are 10 and 1.3 mg/kg, respectively. Accumulation of excessive amounts of Ni and Cr is believed to interfere with Fe uptake and metabolism causing chlorosis and necrosis in plants (Ghani 2011; Khalid and Tinsley 1980). Therefore, the bioaccumulation of such high concentrations of Cr and Ni resulted in decreasing biomass and the occurrence of visible signs of metal toxicity in the aboveground parts of tomato plants grown in BC-unamended soil. Ghani (2011) found that the increase in the concentration of Cr in soil from 10-40 mg/kg is accompanied by the alteration of nutrient uptake in Brassica juncea shoots, thereby causing chlorosis. In our study, Mn was relatively less concentrated in plant tissues compared to other metals, suggesting that both Ni and Cr were the toxic elements that were predominantly responsible for the death of tomato plants grown in BCunamended soil.

The reduction of accumulated concentrations in plants that had received different BC application rates was due to greater immobilization of Cr, Ni, and Mn in serpentine soil via sorption and adsorption mechanisms. The capability of retention of these metals in mesopores of the BC could be increased with BC application rates. Therefore, the reduction of toxic levels of these heavy metals in the presence of BC could be attributed to improving the patterns of essential nutrient uptake by tomato plants, resulting in high biomass production. Hence, plant tissue analyses suggested that the higher biomass production in the presence of 5 % BC application was accompanied by the enhancement of soil fertility as well as the reduction in plant uptake of metal ions.

3.5 Bioavailability of heavy metals

Environmental risks associated with the presence of heavy metals in soils are mainly dependent on the bioavailability of metals (Houben et al. 2013a). The exchangeable fraction in sequential extractions is the fraction of the total amount of metals that can either be bioavailable or made available for uptake by plants. The MgCl₂ extractable concentrations of Cr, Ni, and Mn in BC-amended and BC-unamended serpentine soils are depicted in Fig. 6. In sequential extractions, the exchangeable metal concentrations in different BC amendments showed a considerable decrease with increasing BC application rates. The pattern of individual MgCl₂ extractable metal concentrations was thus Ni>Mn>Cr and hence. Ni showed the highest bioavailability in both BC-amended and BC-unamended soil. Compared to the control soil, 1 % BC application decreased the bioavailable concentrations of Cr. Ni, and Mn concentrations by 95, 14, and 19 %, respectively. In the presence of 2.5 % BC application, the reduction percentage increased reaching up to 96, 36, and 21 %, while in the presence of 5 % BC application, it reached 99, 61, and 42 % for Cr, Ni, and Mn, respectively. Hence, the bioavailability of Cr, Ni, and Mn appears to have been greatly influenced by the BC application rates with BC readily immobilizing metals in serpentine soil.

3.6 Sequential extractions of Ni, Mn, and Cr

Sequentially extracted concentrations of Ni, Mn, and Cr from different fractions in BC-amended and BC-unamended serpentine soil are depicted in Fig. 6. The highest amount of Cr and Ni in both BC-amended and BC-unamended serpentine soil was found to be in the residual fraction, whereas Mn was dominantly bound in the Fe-Mn oxide phase. Fe-Mn oxide fraction is a major source of Mn in serpentine soil (Rajapaksha et al. 2012; Vithanage et al. 2014a). The application of 5 % BC treatment to serpentine soil reduced the fraction of Cr, Ni, and Mn in Fe-Mn oxide phase by 67, 27, and 43 % respectively, compared to the BC-unamended soil. The residual fraction is associated with silicates as well as with other primary oxides such as spinels (Rajapaksha et al. 2012). Ni was dominantly bound in all phases in serpentine soil other than that in the Fe-Mn oxide fraction, while Mn was dominant in the Fe-Mn oxide fraction of both BC-amended and BC-unamended serpentine soil. Such a reduction of available Cr, Ni, and Mn found in different phases after addition of BC to serpentine soil could be due to diffusion and adsorption phenomena. Adsorption of these metals onto BC could be in part explained

Fig. 6 Effects of BC on **a** Ni, **b** Mn, and **c** Cr fractions in the phases of exchangeable, carbonate bound, Fe–Mn oxide bound, organic matter bound, and residual of serpentine soil. For each harvest, means topped with the *same letter* do not differ significantly at the 5 % level according to the Fisher's multiple-comparison test



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by its high surface area $(714 \text{ m}^2/\text{g})$ and pore size (40.8 Å). The diffusion rate of the metals is being governed by BC pore size, and mesopores are highly available for adsorbing metals due to the lack of many surface functional groups as indicated by the FTIR data. The higher attenuation of Cr in the presence of 5 % BC rate may be due to its smaller ionic radius (0.52 Å) which favored its diffusion compared to that of Ni (0.69 Å) and Mn (0.91 Å). However, the adsorption may also be limited since organic substances such as humic acids present in the soil can readily be attached to the BC surface, rending inner pores unavailable for metal diffusion and further adsorption. Hence, further studies should be undertaken for better understanding the metal immobilization mechanisms in BC-amended soils.

4 Conclusions

The present study was conducted to investigate the potential of woody BC, a waste byproduct of a bioenergy industry, as a soil amendment to immobilize bioavailable toxic metals and reduce the phytotoxicity of serpentine soil. Tomato plants grown in 5 % BC-amended serpentine soil increased dry biomass by 30-fold compared to the BC-unamended soil. However, the highest bacterial and fungal counts were found in the 2.5 % BC-amended soil. Bioaccumulation of Cr. Ni, and Mn in tomato plants grown in 5 % BC-amended soil also decreased by 93-97 % compared to the BC-unamended soil. Sequential extraction data demonstrated that the least exchangeable fraction of all the metals was in the 5 % BCamended serpentine soil, revealing that the reduction of exchangeable metal fractions is primarily due to the immobilization of Cr, Ni, and Mn, thereby reducing their bioavailability. Hence, the present study contributes further evidence that the release of Ni, Cr, and Mn from serpentine soil and their bioavailability and phytotoxicity can be reduced with the increase in the BC application. Furthermore, the present study shows that the application of BC to heavy metal-rich sites, including areas that were formerly used for metal extraction, may immobilize metal translocation and accumulation in plants, including in agricultural crops. Hence, BC as a byproduct from the bioenergy industry may potentially be used in the restoration of heavy metal-contaminated soil.

References

Ahmad M, Lee SS, Dou X, Mohan D, Sung J-K, Yang JE, Ok YS (2012) Effects of pyrolysis temperature on soybean stover-and peanut shellderived biochar properties and TCE adsorption in water. Bioresour Technol 118:536–544

- Ahmad M, Moon DH, Vithanage M, Koutsospyros A, Lee SS, Yang JE et al (2014) Production and use of biochar from buffalo-weed (*Ambrosia trifida* L.) for trichloroethylene removal from water. J Chem Technol Biotech 89:150–157
- Ahmad M, Rajapaksha AU, Lim JE, Zhang M, Bolan N, Mohan D et al (2013) Biochar as a sorbent for contaminant management in soil and water: a review. Chemosphere 99:19–33
- Almaroai YA, Usman AR, Ahmad M, Moon DH, Cho JS, Joo Y et al (2014) Effects of biochar, cow bone, and eggshell on Pb availability to maize in contaminated soil irrigated with saline water. Environ Earth Sci 71:1289–1296
- Anderson B, de Peyster A, Gad SC (2005) Encyclopedia of toxicology. Elsevier, Amsterdam
- Armienta M, Rodríguez R, Ceniceros N, Juarez F, Cruz O (1996) Distribution, origin and fate of chromium in soils in Guanajuato, Mexico. Environ Pollut 91:391–397
- Baugé SMY, Lavkulich LM, Schreier HE (2013) Phosphorus and trace metals in serpentine-affected soils of the Sumas Basin, British Columbia. Can J Soil Sci 93:359–367
- Beesley L, Marmiroli M, Pagano L, Pigoni V, Fellet G, Fresno T et al (2013) Biochar addition to an arsenic contaminated soil increases arsenic concentrations in the pore water but reduces uptake to tomato plants (*Solanum lycopersicum* L.). Sci Total Environ 454: 598–603
- Chen B, Zhou D, Zhu L (2008) Transitional adsorption and partition of nonpolar and polar aromatic contaminants by biochars of pine needles with different pyrolytic temperatures. Environ Sci Technol 42:5137–5143
- Fernández S, Seoane S, Merino A (1999) Plant heavy metal concentrations and soil biological properties in agricultural serpentine soils. Commun Soil Sci Plan 30:1867–1884
- Gall JE, Rajakaruna N (2013) The physiology, functional genomics, and applied ecology of heavy metal-tolerant Brassicaceae. In: Lang M (ed) Brassicaceae: Characterization, Functional Genomics and Health Benefits. Nova, New York, pp 121–148
- Ghani A (2011) Effect of chromium toxicity on growth, chlorophyll and some mineral nutrients of *Brassica juncea* L. Egypt Acad J Biol Sci 2:9–15
- Gomez JD, Denef K, Stewart CE, Zheng J & Cotrufo MF (2014) Biochar addition rate influences soil microbial abundance and activity in temperate soils. Eur J Soil Sci 65. doi:10.1111/ejss.12097
- Houben D, Evrard L, Sonnet P (2013a) Beneficial effects of biochar application to contaminated soils on the bioavailability of Cd, Pb and Zn and the biomass production of rapeseed (*Brassica napus* L.). Biomass Bioenerg 57:196–204
- Houben D, Evrard L, Sonnet P (2013b) Mobility, bioavailability and pHdependent leaching of cadmium, zinc and lead in a contaminated soil amended with biochar. Chemosphere 92:1450–1457
- Janice ET, Rillig MC (2009) Characteristics of biochar: biological properties. In: Lehmann J, Joseph S (eds) Biochar for environmental management: science and technology. Earthscan, London, pp 85– 102
- Jiang W, Liu D, Hou W (2001) Hyperaccumulation of cadmium by roots, bulbs and shoots of garlic (*Allium sativum* L.). Bioresour Technol 76:9–13
- Karami N, Clemente R, Moreno-Jiménez E, Lepp NW, Beesley L (2011) Efficiency of green waste compost and biochar soil amendments for reducing lead and copper mobility and uptake to ryegrass. J Hazard Mater 191:41–48
- Kayama M, Sasa K, Koike T (2002) Needle life span, photosynthetic rate and nutrient concentration of *Picea glehnii*, *P. jezoensis and P. abies* planted on serpentine soil in northern Japan. Tree Physiol 22:707–716
- Khalid BY, Tinsley J (1980) Some effects of nickel toxicity on rye grass. Plant Soil 55:139–144
- Mebius LJ (1960) A rapid method for the determination of organic carbon in soil. Anal Chimi Acta 22:120–124

- Moral R, Pedreno JN, Gomez I, Mataix J (1995) Effects of chromium on the nutrient element content and morphology of tomato. J Plant Nutr 18:815–822
- Mulligan CN, Yong RN, Gibbs BF (2001) Remediation technologies for metal-contaminated soils and groundwater: an evaluation. Eng Geol 60:193–207
- Neilson S, Rajakaruna N (2014) Phytoremediation of agricultural soils: using plants to clean metal-contaminated arable lands. In: Ansari AA, Gill SS, Lanza GR (ed) Phytoremediation: management of environmental contaminants. Springer (in press)
- O'Dell RE, Rajakaruna N (2011) Intraspecific variation, adaptation, and evolution. In: Harrison SP, Rajakaruna N (eds) Serpentine: evolution and ecology in a model system. University of California Press, California, pp 97–137
- Onipe O, Adebayo A (2011) Bacteriological and mineral studies of road side soil samples in Ado-Ekiti metropolis, Nigeria. J Microbiol Biotechn Food Sci 1:247–266
- Park JH, Choppala GK, Bolan NS, Chung JW, Chuasavathi T (2011) Biochar reduces the bioavailability and phytotoxicity of heavy metals. Plant Soil 348:439–451
- Paz-Ferreiro J, Lu H, Fu S, Méndez A, Gascó G (2013) Use of phytoremediation and biochar to remediate heavy metal polluted soils: a review. Solid Earth Discus 5:2155–2179
- Peterson SC, Jackson MA, Kim S, Palmquist DE (2012) Increasing biochar surface area: optimization of ball milling parameters. Powder Technol 228:115–120
- Pietikäinen J, Kiikkilä O, Fritze H (2000) Charcoal as a habitat for microbes and its effect on the microbial community of the underlying humus. Oikos 89:231–242
- Rajakaruna N, Baker AJM (2004) Serpentinine: a model habitat for botanical research in Sri Lanka. Ceylon J Sci 32:1–19
- Rajakaruna N, Knudsen K, Fryday AM, O'Dell RE, Pope N, Olday FC, Woolhouse S (2012) Investigation of the importance of rock chemistry

for saxicolous lichen communities of the New Idria serpentinite mass, San Benito County, California, USA. Lichenologist 44:695–714

- Rajakaruna N, Tompkins KM, Pavicevic PG (2006) Phytoremediation: an affordable green technology for the clean-up of metalcontaminated sites in Sri Lanka. Ceylon J Sci 35:25–39
- Rajapaksha AU, Vithanage M, Oze C, Bandara W, Weerasooriya R (2012) Nickel and manganese release in serpentine soil from the Ussangoda Ultramafic Complex, Sri Lanka. Geoderma 189:1–9
- Sruthy OA, Jayalekshmi S (2014) Electrokinetic remediation of heavy metal contaminated soil. Int J Struct & Civil Engg 3
- Summer ME, Andersen CP (1996) Methods of soil analysis. Journal, Soil Science Society of America
- Sun K, Gao B, Ro KS, Novak JM, Wang Z, Herbert S, Xing B (2012) Assessment of herbicide sorption by biochars and organic matter associated with soil and sediment. Environ Pollut 163:167–173
- Susaya J, Kim K-H, Asio V, Chen Z-S, Navarrete I (2010) Quantifying nickel in soils and plants in an ultramafic area in Philippines. Environ Monit Assess 167:505–514
- Uchimiya M, Klasson KT, Wartelle LH, Lima IM (2011) Influence of soil properties on heavy metal sequestration by biochar amendment: 1. Copper sorption isotherms and the release of cations. Chemosphere 82:1431–1437
- Usman AR, Almaroai YA, Ahmad M, Vithanage M, Ok YS (2013) Toxicity of synthetic chelators and metal availability in poultry manure amended Cd, Pb and As contaminated agricultural soil. J Hazard Mater 262:1022–1030
- Vithanage M, Rajapaksha AU, Oze C, Rajakaruna N, Dissanayake C (2014a) Metal release from serpentine soils in Sri Lanka. Environ Monit Assess 186:3415–3429
- Vithanage M, Rajapaksha AU, Tang X, Thiele-Bruhn S, Kim KH, Lee S-E, Ok YS (2014b) Sorption and transport of sulfamethazine in agricultural soils amended with invasive-plant-derived biochar. J Environ Manage 141:95–103