REVIEW

Immunoinformatics and Vaccine Development: An Overview

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has saved several lives, reduced treatment costs and raised the quality of animal and human lives. Current traditional vaccines came empirically with either vague or completely no knowledge of how they modulate our immune system. Even at the face of potential vaccine design advance, immune-related concerns (as seen with specific vulnerable populations, cases of emerging/reemerging infectious disease, pathogens with complex lifecycle and antigenic variability, need for personalized vaccinations, and concerns for vaccines' immunological safety -specifically vaccine likelihood to trigger non-antigen-specific responses that may cause autoimmunity and vaccine allergy) are being raised. And these concerns have driven immunologists toward research for a better approach to vaccine design that will consider these challenges. Currently, immunoinformatics has paved the way for a better understanding of some infectious disease pathogenesis, diagnosis, immune system response and computational vaccinology. The importance of this immunoinformatics in the study of infectious diseases is diverse in terms of computational approaches used, but is united by common qualities related to host-pathogen relationship. Bioinformatics methods are also used to assign functions to uncharacterized genes which can be targeted as a candidate in vaccine design and can be a better approach toward the inclusion of women that are pregnant into vaccine trials and programs. The essence of this review is to give insight into the need to focus on novel computational, experimental and computation-driven experimental approaches for studying of host-pathogen interactions and thus making a case for its use in vaccine development.

Abstract: The use of vaccines have resulted in a remarkable improvement in global health. It

Keywords: immunoinformatics, computational vaccinology, vaccine design, emerging infections, immune system; vaccinology

Introduction

Vaccination has been undeniably very helpful in promoting a healthy global population. It has severally saved lives, reduced healthcare costs and raised man's quality of life. 1 It greatly reduces disease burden, disability and death. However, newly emerging and reemerging infectious diseases (ERID), infectious agents with complex lifecycle and antigenic variability and the need for personalized vaccination present additional challenges in vaccine development.^{2,3}

For many pathogens (especially the emerging and those with antigenic variability), their genomes are known but their immune correlates of protection remain unclear.^{1,4} Some of these reasons are why vaccine development for ERID and multi-lifecycle pathogenic diseases is a tall order.

Serendipitous discoveries in immunology coupled with knowledge of bioinformatics tools for epitope predictions have resulted in the emergence of new pattern of

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vaccine design.^{5,6} The art and science of efficient and comprehensive information extraction and analysis of data deposited in relevant databases is now increasingly essential in researches related to immunology. Even with this capacity (efficient information extraction), some challenges in the application of bioinformatics in immunology include structure and/or function analysis and immune process analyses as concern the immune interaction specificity. Fortunately, although researches in immunology are experimentally costly and very intensive, colossal amounts of data are usually generated. Such data can only be analyzed with high precision and speed using bioinformatics tools. For instance, genome sequencing as well as in vitro T-cell confirmation is done in few months as opposed to years using the conventional vaccine design.8 Also, computational immunological methods drastically reduce both time and labor needs in epitopes screening.^{5,9} With computational immunology techniques, it is possible to discover vaccine candidate epitopes simply by scanning the protein sequences in a pathogen of interest.⁵ Many of these proteins are yet to be isolated or at least cloned. Being pathogens specific and unique, they present ready candidates in vaccine construct.

This review describes the need to use immunoinformatics-based techniques to unveil vital determinants of immunity made available in the genome sequence database and design vaccines. Also, this review gives insight into the need to focus on novel computational, experimental and computation-driven experimental approaches for studying of host-pathogen interactions and thus making a case for its use in vaccine development. This review will further show the need for new approaches for effective drugs or vaccine design so as to combat the antigenic variability of some pathogens.

Vaccine Immunology and Design

The process of generating vaccine-induced immunity is somewhat challenging in immunology. Current conventional vaccines came empirically when there were vague or no knowledge of vaccine immune system activation. A lot of research 10–12 has been geared toward the understanding of this challenge, but the complexity of it requires a different dimension of approach. An approach that must accommodate many factors affecting vaccine development like pathogen antigenic variability, the emergence of infectious disease, human genetic variation is the goal of immunoinformatics [Figure 1].

Activation of the immune system involves, among many processes, induction of the immune memory. The strength of this induction determines the efficacy of a vaccine. Hence, vaccine efficacy in the long run is influenced by the determinants of immunological memory stimulation, persisting antibodies and kind and type of immune memory cells induced.¹⁴

The primary vaccine-mediated immunological effectors (Table 1) are mainly the antibodies (from B lymphocytes/cell)^{15,16} and sometimes CD8+ and CD4+ T cells. These antibodies bind specifically to a particular kind of toxin or

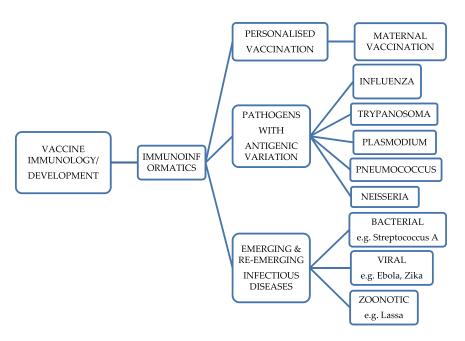


Figure 1 Schematic illustration of the cases stemming the need for immunoinformatics vaccine development approach.

Table I Functions of the Immunological Memories

Immune Cells	Immunological Memories (Effectors of the Immune Response)	Mechanistic Functions
B cells produced (humoral immunity)	Antibodies play vital roles in the control (including prevention) and complete removal or destruction of both extracellular and intracellular pathogens as well as in response to vaccination.	- activate the complement cascade - neutralize the replicating power of viruses (viral infectivity) - prevent the diffusion of toxins and/or bind to their enzymatic active sites (comprising of the binding site and the catalytic site) - induction of the macrophages and neutrophils for the purpose of clearing extracellular bacteria
T-cell produced (cellular immunity)	T cells of the CD4+ class. Clears the pathogens residing within and outside the cells	- produces several interleukins and supports B-cell stimulation and differentiation (Th2 cells response) - produces Th1 cell responses (produces interferon-γ, tumor necrotic factor-α/-β and Interleukins-2 and 3 and supports the proliferation and differentiation of CD8+T cells, B cells, and macrophages - activate the B cells, cytotoxic T cells and other cells not involved in the immune system
	T cells of the CD8+ T class. Clear the pathogens residing in the cells	- induces the release of antimicrobial cytokine for the purpose of killing microbial infected cells - kills infected cells directly by releasing proteolytic enzymes

pathogen. Vaccines and most antigens evoke humoral as well as cell-mediated immune responses.¹⁷ Vaccines that mediate immune responses through these systems (B and T cell responses) are said to be more effective. Although B cells are regarded as the primary vaccine immune effectors, T cells induce immune memory cells and high-affinity antibodies. Studies in reverse vaccinology and immunomics had also proved T cells as prime immune effectors following the discovery of novel vaccine targets with EpiMatrix. 18-20 This change of immune target has led to successful advances in vaccine design.

Even at the face of potential vaccine design advances, immune-related questions are now focused on specific vulnerable populations such as the young, elderly and immunocompromised.^{21,22} These concerns have propelled a better understanding of the efficacy of current vaccines on this vulnerable population and have also paved way for the application of new approaches that can put into consideration the differences of population and better targets that can generate optimum immune induction^{23–25} with the exception of type II T-cell-independent (TI-2) antigens (i.e., polysaccharide antigens).

Antigens that could provoke the B lymphocytes as well as the T lymphocyte responses stimulate the germinal centers causing antigen-specific highly efficient B-cell multiplication and eventual differentiation into antibodyforming plasma cells and memory B cells. All existing protein and DNA antigens induce immunological memory B cells unlike type II T-cell-independent (TI-2) antigens (i.e., polysaccharide antigens). These polysaccharide antigens do not generate memory B cells but can induce longlasting humoral immunity even when recall responses are lacking.²⁶ Vaccine efficacy may be short term²⁷ if only the B cells are activated.

The traditional approach for developing vaccines for infectious disease threats has shifted to include other vaccine design techniques like cloning and expressing major surface antigens²⁸ although this frequently resulted in the formulation of vaccines with poor immunogenicity, requiring strong adjuvantation.²⁹ This approach is particularly likely to be less specific for pathogens with complex lifecycles (e.g., parasites) or very high mutability (e.g., RNA viruses). These pathogens do not depend on one route for their virulence of pathogenesis in human and thus to alter this process, increasing the specificity of the vaccine should be the aim and not just the effectiveness as seen in the current conventional vaccines. 28,29

Vaccines for several neglected tropical diseases are in various stages of development, 30 thanks to mega drug companies that have continued to demonstrate a willingness to invest money in the research and development as regards to diseases plaguing the developing nations. 16,30-32 It is very pertinent to invest in researches that have an interest in vaccine specificity on the pathogen antigens than totally

on the efficacy. Fortunately, with global research efforts, the genomes for several pathogens causing neglected tropical diseases and several emerging pathogens are becoming available. Computational vaccinology may now be applied to screen these genomes for possible vaccine target. With these tools, many proteins of virulence interest can be sequenced and the most essential gene of interest modeled for a potential vaccine candidate specific for that pathogen. Immunoinformatics is the way forward in the identification of vaccine candidates for these tropical ERID, for pathogens with varying antigens and for individualized therapy.

Immunoinformatics and Infectious Disease

Immunology studies produce data in colossal quantities. Also, with proteomics and genomics projects, extensive screening of pathogens and/or pathogen–host interaction, it has become increasingly necessary to store, manage and analyze these data, hence the birth of immunoinformatics. Immunoinformatics deals with computational techniques and resources used to study the immune functions. Statistical, computational, mathematical and biological knowledge and tools are applied in immunoinformatics in order to accurately and specifically store, and analyze data concerning the immune system and its functions.

To handle evidence diversity, immunoinformatics uses tools that cut across several aspects of bioinformatics such as creation and management of databases, ^{34,35} use and definition of both structural and functional signatures and the formation and application of predictive tools. ^{35–37} These strategies can synergize toward a better understanding of the immune system of both man and animals and fight

against some less predictable pathogenesis. The complex nature of vertebrates' immune system, the variable nature of pathogens and environmental antigens coupled with the multi-regulatory pathways show that colossal quantities of data will be needed to unveil how the human immune systems work. Conventionally, much cannot be achieved based on the complexity of the immune system and the virulent antigen but with the application of computational vaccinology, researches on vaccine design have been made easier, accurate and specific. Applying immunoinformatics in disease study (Table 2) requires the knowledge of disease pathogenesis, the immune system dynamics, and computational vaccinology, painstaking searches of the database, sequence comparison, structural modeling as well as motif analysis. 35,38 These methods can go a long way in analyzing the pathogenesis of a disease and identification of vaccine candidates.

In order to help understand complex pathogenrelated processes, computational models were developed for viral^{46,47} bacterial,⁴⁸ parasitic⁴⁹ and fungal pathogens.⁵⁰

The bioinformatics tools (Table 3) are used to identify possible epitopes for vaccine formulation. Each tool can screen protein sequences and identify aggregates of MHC binding and supertype motifs for possible use in epitope-based vaccine development and for use among human populations with genetic variability.

There are several databases (Table 4) that can provide a wide range of information for all forms of immunological studies. Generated data from the studies are further organized and stored in the databases (Table 4) to provide a means for the development and advance in

Table 2 The In	nportance of Bi	oinformatics i	n the Resea	rch on In	fectious Diseases

Importance	Applications		
Surveillance of infectious disease	Microbial genotyping is used to either confirm or refute epidemiological links with potential environmental sources.	39	
Determining the various strains of pathogens in circulation	The proteins used by variants of pathogens can be predicted and mutated for better analysis. Even the genes that code for the proteins can be manipulated in silico in order to predictive a better targeting		
Diagnostic microbiology	Bio-surveillance focused text-mining tools and microbial profiling are used to detect infectious disease outbreak	42	
Databases for Pathogens	Array of data on pathogens can help in their genome study and their virulence toward development of vaccine candidate		
Vaccinology	Bioinformatics have helped in the advance of DNA and Epitope-based vaccines both in silico and as a preliminary study for the in vivo validation study	44,45	

Table 3 Bioinformatics Tools for MHC Cluster Binding and Super-Type Motifs and Protein Sequences

Bioinformatics Tools	Applications/Description	Refs
EpiMatrix	is an in-silico product of EpiVax developed for predicting and identifying the immunogenicity of therapeutic eins and epitopes. It is also used to re-design proteins and in designing T-cell vaccine	
Conservatrix	las been applied in comparing strings from different strains of same pathogens and for pathogens identification. Configuration of Conservatrix allows for amino acid replacement at unusual positions. Highly conserved T-cell pitopes in variable genomes such as some viruses are amenable to the algorithm	
ClustiMer	Potential T-cell epitopes usually aggregate in specific immunogenic consensus sequence (ICS) regions as clusters of 9–25 amino acids with 4–40 binding motifs instead of randomly distribute throughout protein sequences. In combination with EpiMatrix, the ClustiMer algorithm may be used to identify those peptides with EpiMatrix immunogenicity cluster scores $\geq +10$. Such peptides are usually immunogenic and tend to make a promising vaccine candidate.	1,53,54
BlastiMer	ing BlastiMer program, one may also choose to automatically BLAST "putative epitopes against the human sequence cabase at GenBank". BLASTing screens off those epitopes with possible autoimmunity and cross reactivity questions d locates the epitopes that can safely be used in developing human or animal vaccine. BlastiMer can also BLASTs quences against PDB, SwissProt, PIR, PRF and non-redundant GenBank CDS translations.	
Vaccine CAD	This algorithm evaluates junctional epitopes for possible immunogenicity and inserts "spacers and breakers into the design of any string-of-beads construct".	1,58,59
NERVE	Predicts the best vaccine candidates starting from the flat file proteome of a prokaryotic pathogen. It's a fully automated reverse vaccinology system, developed to predict best VCs from bacteria proteomes and to manage and show data by user-friendly output.	
Jenner-Predict	Predicts PVCs from proteomes of bacterial pathogens. The web server targets host-pathogen interactions and pathogenesis by considering known functional domains of protein classes such as adhesin, virulence, invasion, porin, flagellin etc	
Vaxign	Is a vaccine target prediction and analysis system based on the principle of reverse vaccinology? Two programs exist in Vaxign: 1) Vaxign Query and 2)Dynamic Vaxign Analysis	
VaxiJen	Is the first server for alignment-independent prediction of protective antigens. It was developed to allow antigen classification solely based on the physicochemical properties of proteins without recourse to sequence alignment.	
VacSol	A high throughput in silico pipeline to predict potential therapeutic targets in prokaryotic pathogens using subtractive RV.	
PanRV	Pangenome-reverse vaccinology approach for identifications of potential vaccine candidates in microbial pangenome. It comprises of four functional modules including i) Pangenome Estimation Module(PEM) ii) Reverse Vaccinology Module(RVM) iii) Functional Annotation Module(FAM) and iv) Antibiotic Resistance Association Module(ARM).	65

immunological research. A tour on these databases will actually stimulate some interest in the vaccinology of emerging and re-surging disorders attributable to pathogen including cancer.

Case of Emerging and Re-Surging Disorders/Diseases Attributable to **Pathogens**

Emerging infections (EIs) include infections that are entirely new in a population or that may have existed before in the population but are now gaining rapid and continued spread and/or wide geographical range.

Re-emerging or re-surging infections represent the infections that were previously of historical relevance but are now quickly becoming relevant because of either increasing incidence or increasing geographical and/or human host range while emerging infections represent the infections that were not originally observed in man. 66 Several factors such as human behavioral changes, environmental changes, and host/intermediate factors, animal-human switching and microbial genetic changes all affect infectious disease emergence and spread.⁶⁷ These factors interact to promote the evolution of pathogens into new ecosystems, infect, spread and thrive in their new hosts.⁶⁸ The overall consequences of these are continued

Table 4 List of Some Immunological Databases

	Immunological databases	
IMGT	It contains well over 32, 000 entries of Immunoglobulins and T cells Receptor sequences from both human and non-human vertebrate species. A lot of information on the human leukocyte antigen is also provided.	http://www.imgt.org/IMGTindex/databases.php
HIV Molecular Immunology	It contains an annotated pull of HIV-1 CD4 ⁺ and CD8 ⁺ epitopes and antibody binding sites.	https://www.hiv.lanl.gov/content/ immunology/
ExactAntigen	Employed for the search of monoclonal antibodies utilised for therapeutic, commercial and academic purposes.	http://www.labome.com/index. html
EPIMHC	This the major curated database for MH ligands and Epitopes for tailor-made Computational Vaccinology	http://imed.med.ucm.es/epimhc/
JenPen (AntiJen)	This database contains quantitative binding data for peptides that bind to MH1, MH2 and TAP molecules. Also provides quantitative binding data for TAP, TCR-MHC complexes and MHC-ligand interactions	http://www.ddg-pharmfac.net/ antijen/AntiJen/antijenhomepage. htm
SNPBinder	A known database of predicted antigenic peptides and minor histocompatibility antigens (mHAgs)	http://www.imgt.org/about/immu noinformatics.php
SYFPEITHI	This is a collection of MH ligands and peptide motifs	http://www.syfpeithi.de/bin/ MHCServer.dll/ EpitopePrediction.htm
Bcipep	This a repertoire containing immunant dominant B cell epitopes	http://crdd.osdd.net/raghava/ bcipep/
kabat database immunoglobulin	It contains sequences of important immunological proteins of including Ig, TcR, MHC molecules, etc	http://immuno.bme.nwu.edu OR http://www.ftp.ebi.ac.uk/pub/data base/kabat/
EpiVaxb	Contains information on promiscuous and conserved epitopes of class I and II	http://www.epivax.comb
IEDB Binding, MHC Class I	Uses 3 different methods to predict class I-peptide binding	http://www.immuneepitope.org/ analyze/html/mhc_binding.html
IEDB Binding, MHC Class II	Prediction of class II-peptide binding	http://www.immuneepitope.org/ tools/matrix/iedb_input
MotifScan	Summary and location of anchor motifs	http://www.hiv.lanl.gov/content/ immunology/motif_scan
MHC Haplotype Project	The haplotype of MHC-linked-diseases, showing full genomic sequences, ancestral relationships and vital variations (SNPs and DIPs).	http://www.sanger.ac.uk/HGP/ Chr6/MHC
HCV Immunology	CD8b and CD4b T cell proteome and HCV epitopes maps	http://hcv.lanl.gov/immuno/

infectious disease emergence and re-emergence, epidemics and public health challenges. Emerging infections and multi-antibiotic-resistant strains of pathogenic bacteria usually surge from one geographic location from where it spreads to other places due to immigration of people. Most emerging infections originate from a specific population and can spread to a new population or become selectively advantaged that it can lead to the

emergence of new strains of the pathogen.^{67,70,71} Also, there could be microbial traffic, in which case, an infectious agent transfers from animals to humans or spreads from isolated groups to new populations.^{67,71,72}

Several factors, including ecology, are known to be associated with infectious disease outbreaks. These factors bring man into close contact with a natural disease reservoir/host.⁷⁰ With an increasing world population and

poor infection control, the emergence of infection and increased microbial populations are sure. The human growth population will only increase the spread of the infection across populations. The information provided in Table 5 is the list of remerging infections and current emerging diseases put forward during the WHO 2018 annual review.⁷³ The review noted that these infections, if not well controlled, can cause disease outbreaks, bioterrorism and similar occurrences requiring urgent public health attention and that with the dearth of efficacious medicines or vaccines, there is a compelling demand for continuous as well as accelerated research and development in those areas.

Advances in Genomics, proteomics, immunomics, vaccinomics and nanotechnology are being continually exploited in diagnostic, therapeutic and in rational drug and vaccine development. These advances have also served in the control of the afore-mentioned emergences. ^{74,75} The knowledge of the emerging pathogen's genome, protein make-up, pathogen-immune system interactions and researching the possible therapeutics will go a long way in directing the optimum path to containing the infection

spread and controlling potential re-emergence or emergence in a different population. Approaches in direct and computer-based structural determinations, ⁷⁶ protein–protein interactions predicting, and bioinformatics tools now exist and are used in modern-day development of drug and biologics. ⁷⁷

Vaccine development has been sped up through the advance in the knowledge of the immune system of man. Researches in the traditional targets of vaccines (adaptive immune response) and the less specific and fast-acting innate immune responses have been clear evidences for this advance. 78–80 As our understanding of the intercourse between innate and adaptive immunity increases, reasons and opportunities for more effective vaccine adjuvants will open up. This can be a step forward in solving a critical world's health challenge per population. Following the conventional approach of vaccine design, much cannot be achieved but when the knowledge of immunoinformatics is applied, population safety and disease control can be achieved through pathogen's genome sequencing leading to optimum new vaccine design or development of a novel vaccine for the infection.

Table 5 Current Emerging and Re-Surging Infectious Disease

Infectious Disease	Endemic Population	Contributing Factors	Current Treatment
Lassa fever	West Africa	Urbanization favoring rodent host, increasing exposure	Ribavirin (no specific vaccine)
Streptococcus A (Invasive necrotizing)	Global	Uncertain	Antibiotic
Ebola	Central & West Africa	Unknown (In Europe, importation of monkeys)	No proven therapy (Experimental vaccine)
Variant Creutzfeldt-Jakob disease (cattle)	UK, France, Spain	Changes in rendering processes	No specific therapy
Shigellae sonnei Infection		Consumption of iceberg lettuce	
SARS	Southern China, Canada	Animal-to-animal transmission	No specific therapy
2009 HINI Influenza	Global attack	Droplets of unprotected cough or sneeze	Antiviral therapy (No specific vaccine)
Hantavirus pulmonary syndrome	USA	Rodent infestation	No specific therapy
MERS-CoV	Saudi Arabia,	Human-to-human, dromedaries	No specific therapy
Human T lymphocyte Virus I (HTLV-I)	Japan, Central & South Africa, USA	Human-to-human, Sexual contact	Experimental vaccines
Human immunodeficiency virus- 2 (HIV-2)	Africa	Sooty mangabey monkey	No specific therapy
Human herpes virus-6 (HHV-6)	USA, UK, Japan, Taiwan	Shedding of viral particle into saliva	No specific therapy

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Pathogens with Variable Antigens

Antigenic variability is an important mechanism pathogens use to evade their host immunity. The surface proteins of pathogens are normally variable. This assists them to escape recognition by the immune system. A successful infectious agent presents to the host immune system information that differs from that of its virulence. Pathogenic organisms have organized systems of escaping destruction by the immune system of their hosts. For instance, Toxoplasma invades and appropriates the host cells thereby circumventing phagocytosis and then spread within their host to establish infection.⁸¹ Vertebrates on their own are endowed with immune system robust enough to efficiently and effectively surmount the non-self-attacks. Yet the more the host's immune system elaborates, the better the organisms in their evasion of immune effector cells.

Antigenic variation refers to a pathogen's ability to modify its surface proteins such that it can circumvent the host's immunological attacks. It involves several mechanisms including the varying of surface protein's phase, shifting and drifting of surface protein antigens and/or any other form of alteration of antigenic protein.⁸² Antigenic variation plays significant roles in the pathogenicity of microorganisms by evasion of the host immune responses and establishment of re-infection. When a pathogen alters its surface antigens, it can evade the host's adaptive immunity and so reestablishes infection. The immune system may battle to generate new immunoglobulins against the new antigen. Certain bacteria like Neisseria gonorrhea, Neisseria meningitides, Mycoplasma and species of the genus Streptococcus show antigenic diversity.83 In eukaryotic pathogens, antigenic variation is shown by Trypanosoma brucei and Plasmodium falciparum. 81,84

Another vital cause of antigenic variation in bacteria is horizontal gene transfer (more important than point mutation) through plasmid acquisition and transduction via bacteriophages. Virulence genes are normally acquired by non-virulent organisms via these routes. Once this occurs, the new bacteria may quickly get established and cause fresh epidemic outbreak.

Neisseria

Species of the genus *Neisseria* are champions in the rapid change of surface antigens amongst bacterial pathogens. Pathogenic forms exhibit an amount of phenotypic

variability not found in the commensal species. The pathogenic forms are implicated in STD and meningitis. They employ amazing varieties of antigenic variability mechanisms.

- They can recombine their pilin genes in a similar manner that eukaryotes recombine their own genes, such that they can express variable surface protein.
- Some cell-surface proteins and enzymes synthesizing bacterial cell-surface carbohydrates are expressed in a variety of ways. This is as a result of replication slipping or slippage errors and repairs of simple tandem nucleotide repeats involving either the di-, or tri- or tetra-nucleotides.
- Neisseria is able to take up and incorporate environmental DNA into its genomes.^{83,87}
- Again, the chances of Neisseria acquiring new mutations by replication errors are high compared to other bacteria like E. coli.⁸⁸

These are why an effective vaccine against *Neisseria* infections is not yet developed. *Neisseria* may be considered as an extreme example. However, many other bacterial pathogens like *Streptococcus* and *Mycoplasma* in promoting their antigenic variation tend to utilize one or more of these techniques. Additionally, there are reports that DNA-related defects have a much greater association with bacterial pathogen from symptomatic patients than samples of the same bacterial species isolated from environmental sources. ^{87–89}

Pneumococcus

Streptococcus pneumoniae, Gram-positive cocci bacteria that cause otitis media, bacteremia and pneumonia, are a public health concern, causing morbidity and mortality in adults and children. Two forms of vaccines (polysaccharide and conjugate vaccines) are currently marketed for the prevention of pneumococcal infections. While the polysaccharide vaccines are for vaccination in the adult population, conjugate vaccines have an added immunogenic non-pneumococcal protein conjugated to the pneumococcal polysaccharides for enhanced immunogenicity in children. It is not yet known that these vaccines can evoke complete immunity against the infection.

A polysaccharide capsule is a major virulence factor in the bacteria. Several of these capsule types have been identified, and these form the basis of pathogen's antigenic serotyping. 91,92 Current pneumococcal vaccines are

combinations of various capsular (polysaccharide) antigens from the serotypes most common in a particular population. Currently, over 100 different serotypes are known but are not all covered in the available vaccines. 92 The discovery of a common antigen(s) will produce an effective vaccine. Knowledge of the genome of the organism and the different strains has led to a possible advance in driving the pneumococcal potential vaccine search through a different approach. And this consideration will help solve a lot of concerns about the current vaccines. With this knowledge, many methods are been tried to determine whether they can be a source of effective vaccine design that can accommodate all the serotypes of the organism. Search for antigen that is common to all the serotypes can be achieved with the knowledge of Genomics and immunoinformatics. introduction of genomic and computational technologies has given new directions in the study of bacterial pathogenesis and vaccine design. 93,94

Plasmodium

Plasmodium falciparum undergoes two life cycles: one in humans and the other in mosquitoes. The human host's erythrocytes and hepatocytes usually display modified parasite proteins called Plasmodium falciparum erythrocyte membrane protein 1 (PfEMP1) and Plasmodium falciparum hepatocyte membrane protein 1 (PfHMP1), respectively. These proteins function to assist the parasite to evade destruction by the host immune systems. 95,96 The PfEMP1 proteins were identified as the prime ligands responsible for cytoadherence and resetting. 97 They cause the infected RBCs of host tissues to sequester thus helping the parasite to circumvent clearance by the host's spleen. 98 The membrane proteins also shield infected host cells from destruction by the spleen by adhering to the endothelium. Luckily, if the PfEMP1 protein is expressed for a long while; it comes under attack by the naturally acquired immunity. 98,99 In defence to this, the parasite has expanded the var genes coding for PfEMP1 such that the genes can exist as a polymorphic family of as much as over 50 members in every genomic haploid. Antigenic switches work well here in that members of the polymorphic family (also called antigenic-variant-protein family) can be interchanged and cannot express their proteins at the same time. In this way, only one particular protein at the surface of the infected RBC is expressed at any given time. 97,100

When studying antigenic-variant-protein families, it is pertinent to understand if grouping them into single-family results in any meaningful antigenic activity. Studies have tried to understand the "languages" of the antigenic variant of PfEMP1 proteins. ^{97,101,102} They sought to know the PfEMP1 proteins binding properties or search to understand the correlation between motifs and infection severity.

The varDB database is a repository for protein sequences involved in antigenic variation and their associated functionalities. Antigenic variant data obtained from several pathogens may be regrouped into a unified database. This will enable researches from several multicopy gene families to be accessed and compared swiftly in a single moment. Updated varDB database contains close to 10,000 DNA sequences, several protein translations, tens of infectious diseases and pathogens with their gene families. With a novel sequencing-based approach, PacBio, the different PfEMP1 proteins can be sequenced and the related sequences used as potential vaccine targets.

Trypanosoma

For many pathogens, antigenic variability occurs during the infection pathogenesis and is to enable them to escape destruction by the host antibodies. For instance, some eukaryotic parasites take to genetic assortment and rearrangement thereby changing their surface antigens. A ready example is seen in Trypanosoma brucei, the causative organism for sleeping sickness. Trypanosoma brucei replicates in the bloodstream (outside the cells) of their host, but at maturity, it crosses the blood-brain barrier to cause several fatal complications. During replication in the bloodstream, the parasites are subjected to humoral as well as cellular immune responses. It evades the host defenses by encasing itself in homogeneous coat of glycoprotein called the variant surface glycoprotein (VSG). 106,107 Though at initial invasion, the protein coat tends to protect the microbe from the immune system but on constant exposure, the coat will be identified as a foreign matter, and at this point the immune effectors can launch an attack against it. In a particular Trypanosoma brucei, there are diversities of the VSG protein being coded by more than a thousand genes in the parasite's genome. Unfortunately for the host, the expression of these genes is mutually exclusive. Expressed VSG gene is normally due to genetic reassortments causing new alleles to be copied into the sites of expression. Some trypanosomes with these abnormal VSG genes evade humoral immunity and multiply thereby causing re-infection and chronic recurring infections. 107

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Influenza Virus

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Influenza is a viral infectious disease due to infection by any of the three types of RNA viruses, namely influenza Types A, B and C. Current vaccines contain double Type A and single Type B strains and induce strong antibody responses to neuraminidase and the surface glycoprotein hemagglutinin. These vaccines, however, cannot effectively protect against newly emerging viruses with antigenic shift and drift. ^{108,109}

Antigenic drift results in changes in the antigenic site (a minor change) while antigenic shift results in a new virus subtype. Hemagglutinin and neuraminidase are the two enzymes dictating the antigenic properties of the viruses. While inside its host, defined host proteases break the peptide bonds in the hemagglutinin molecule to form hemagglutinin 1 and 2 subunits. Virulence tendencies are decreased when the amino acids at the cleavage sites are lipophobic, the virus exhibits high virulence tendencies. The surface glycoprotein can be regarded as antigen and hence can serve as a target for the immune system which if sequenced, using the new immunoinformatics approach and a common site for the varying proteins identified, a potent vaccine can be developed which can accommodate the antigenic drift/shifts of the virus.

Influenza viruses are able to thrive for a long while in a given human population. The virus has a high mutation rate such that a once effective vaccine can easily lose efficacy. Antigenic variability is only one of the evidences of phenotypic variation in the biology of the Influenza virus.

Multiepitope Vaccines

The use of immunoinformatics in vaccine development has been accelerated toward the design of a multiepitope vaccine construct which has and will fully address the challenges faced with pathogens with mutagenic antigens. Previous vaccines developed by conventional approaches consist of several proteins or a whole pathogen. This constitutes unwarranted antigenic load and increases the chances of inducing allergy. The use of peptide-based vaccines surmounts these challenges. The vaccines are made from short peptide fragments capable of eliciting highly specific immune responses, precision targeting and multiepitope constructs, in the case of varying antigenic peptides, which has been made feasible with the advancements in the field of computational biology. 113–116

Vaccines for pathogens with immune escape potentials can basically be constructed by using most, if not all, of their

immunogenic peptides 116,117 because such vaccines prove to be better than single-epitope and classical vaccines. Multiepitope vaccines enjoy the following advantages over single-epitope and classical vaccines: a) they are an assemblage of several epitopes obtained from distinct protein targets/antigens of an intended infection; b) the multiple T-cell receptors (TCRs) in the vaccine recipient can easily recognize vaccines with multiple HLA epitopes; c); they can be easily adjuvanted to improve their immunogenicity; d) they can activate antibody-mediated and cell-mediated immunological responses because of their overlapping helper T lymphocytes (HTL), CD8+ T-cell and B-cell epitopes; and e) unwanted protein antigens are excluded in such construct thereby reducing the chances of untoward effects and/ or immune responses likely to cause disease(s). 118-123 Thus producing a vaccine with these qualities can provide chances of combating most infections such as Streptococcus pneumoniae and HIV infections.

Immunoinformatics can be employed in the docking of single and multiepitope vaccines and subsequently to predict their properties (physicochemical, allergenic and antigenic). This approach has seen the use of diverse tools and database in the analysis of ligands with their targets and has greatly helped to predict the binding score of antigenic peptides with the immune proteins like HLA. Peptides and HLA allele modeling can be done by the 3D structural designing of the epitopes using PEPFOLD3 (an online server), 124 retrieving from Protein Data Bank (PDB) the x-ray crystallographic structure of human population most occurring HLA alleles (HLA-DRB1 01:01, 15:01 and HLA-A 02:01) followed by filtration of previously bound ligands.

The following is a step-wise detail on how to construct a single or multiepitope-based vaccine and its property prediction;

- Molecular Docking Analysis: to determine the interaction pattern of the screened out epitopes with the
 HLA alleles by employing ClusPro v.2 (a protein-protein docking web server). This server performs
 this task by energy minimization, calculation of
 both the binding energy scores of the docked complex and electrostatic/shape complementarity.
- Target-Protein Comparative Modeling and associated Structure Validation: the sequence of the amino acid in the target protein (e.g., TLR-9) can be retrieved from UniProt and the tertiary structure with Raptor-X and I tasser (online comparative modeling tools). The server constructs and creates a 3D model

(mathematical representation) of the target protein using hierarchical algorithms. 118,125

- Designing of Final Single/Multiepitope Vaccine Construct: in the case of multiepitope-based vaccine; the different epitopes can be linked to each other using AAY and GPGPG linkers and to the adjuvant using EAAAK linker.
- Predicting the antigenicity and allergenicity together with other physicochemical information: allergenicity of the final vaccine construct can be analyzed by Vaxijen v.2 tool, antigenicity of the vaccine by the use of AlgPred server and the physicochemical properties (isoelectric point, half-life, solubility, molecular weight, aliphatic index, and average hydropathicity) with the ProtParam server.
- Structure modeling, Refinement and Validation of the final Vaccine-these can be investigated using the SOPMA server/Raptor-x, Galaxy Refine Server and ProSA-web, respectively. This process is being followed by molecular docking and molecular dynamics of receptor and the epitope vaccine using ClusPro and Molecular Dynamic Simulation (Desmond)¹²⁶ respectively.

Case of Personalized Vaccination

Personalized vaccine refers to vaccines "targeted" toward an optimized outcome. Immunogenicity is maximized while either the risk of vaccine failure or reactogenicity and side effect is minimized. Personalized are developed in the following cases;

The Individual Level

Vaccines are developed to take care of haplotype and polymorphism knowing that they can retard the formation of a protective immune response or become pointers to the risk of an adverse vaccine reaction.

The Gender Level

This is needed when it is clear that females produce a higher antibody titre against a particular vaccine antigen than do their male counterparts.

The Racial/Ethnic Level

Where it is clear that a particular human race or ethnic group has a higher or lower immune response to a particular vaccine antigen.

The Subpopulation Level

Personalized vaccines arise due to known complex interactions between host environmental, genetic and some other factors that may be influencing the vaccine immune responses. The associations between the immune response gene polymorphisms and variations in immune responses to a particular gene must be pine-pointed when it is clear that a particular drug either suppresses or augments the transcription of an immune response gene. This could help in designing vaccines or vaccine adjuvants that can circumvent restrictions due to immunological differences arising from varying genetic compositions. The some other particular drug either suppresses or augments the transcription of an immune response gene.

Personalized vaccines stem from our understanding of how, within the human leukocyte antigen (HLA) system - also referred to as the major histocompatibility complex (MHC), the T cells are able to recognize peptides of pathogenic origin. 131,132 HLA molecules enjoy the double advantages of having stable polymorphisms and being fully characterized. 133 These advantages make good candidates for personalized vaccine design. HLA polymorphism, although stable, is complex. For instance, more than 12,000 alleles of HLA class I molecules and greater than 4000 class II molecules have been identified among human populations. 133,134 HLA class I and II molecules have heterodimeric character comprising of α and β chains, three highly variable extracellular domains ($\alpha 1$, $\alpha 2$, and α3) and then transmembrane and intracytoplasmic domains that are less variable. 133,135 HLA genes contain eight exons. Exon 1 is responsible for producing the leader peptide; exons 2,3,4 produce α1, α2, and α3 extracellular domains, respectively, for MHC class 1 or α 1, β 1 and α 3, respectively, for MHC class II; exon 5 encodes transmembrane anchor; exons 6 and 7 encodes the cytoplasmic tail while exon 8 encodes the 3'-untranslated region. 135 Most of the several forms associated with the class I and II genes are seen in α -1 and α -2 (as known as class I) and in α -1 and β -1 (as known as class II) domains. ¹³³ MHC I and II bind and present the peptide to T cells.

T cell responses to viral pathogens differ from one patient to the other, basically because of the expression of differing HLA (MHC) alleles which determine the several viral amino acid sequence brought to the T cells to read. ^{136,137} It is most likely that during an infection, diverse epitopes are usually presented to the T cells to read owing to the several forms of HLA alleles and also because each human person expresses six HLA Class I and six HLA Class II alleles. ¹³⁸ Now, antibody-binding sites in a given

HLA (MHC) molecule are mostly prediction-servers predetermined on the basis of particular binding motifs and the anchor residues. These residues refer to known amino acids located at defined locations in the peptide chain and which are peculiar to each MHC molecule. Prediction-server database of peptide motifs and/or of MHC ligands may be obtained from web-based and/or from prediction-servers dedicated to NetMHC family. 143,144

In another example, sequence analysis of Lassa fever virus (the LASV) and other viruses' immunoproteomic was used to identify the best immunogenic protein predicting T-cell as well as B-cell epitopes and also target sequence and binding sites. 145,146 The SSNLYKGVY peptide sequence at AA41-49 of glycoprotein 1 (produced by the L segment) was the best candidate epitope for the induction of humoral as well as the cell-mediated immunity for Lassa fever vaccine construct. 17 HLA-I and 16 HLA-II molecules have been proven in sizable African populations and their combination with the SSNLYKGVY peptide sequence may prove useful in such Lassa fever virus endemic areas. 145 This approach will strongly improve individualized vaccination and help combat emerging infections. The HLA region is suspected to contribute, to a large extent, to genetic propensity to infections and differences in vaccine expected immune responses. 132,147

Studies show that females exhibit stronger immune responses to immunization compared to males. ^{148,149} There are differential antibody responses to rubella and measles viral protein between males and females and that both hormonal and genetic difference may be influencing the immune responses. ^{148,150,151}

Practical issues may stand in the way of achieving this new development (personalized vaccinology). Having to use different vaccines for different persons based because of personal genetic composition requires more time and labor during the vaccination process. Also, screening for individual factors for targeted vaccination can significantly increase vaccination cost. But with all these challenges, personalized vaccination is the new age approach in achieving an optimum immunization that takes into consideration the individual immune differences in a particular population and it is a new dawn for vaccine development.

Personalized vaccine development is strongly improved by vaccinomics. The field of vaccinomics looks at how immune response gene polymorphisms affect the cellmediated, humoral and innate immune responses to vaccine antigens at population and specific individual levels. "Vaccinomics" encompasses both immunogenomics and immunogenetics as it concerns immune responses to vaccine antigen. The fields of personalized vaccinology and vaccinomics were the products of Phase I of the international HapMap and that of the Human Genome Project. Also, modern molecular assay techniques permitting high-throughput detection of variations at gene level, in particular linkage disequilibrium maps and single nucleotide polymorphism (SNP), played significant roles in the development of personalized vaccinology and vaccinomics. It has also been shown that polymorphisms at vital immune response genes can bring about differing immune responses to bio-pharmaceutical products including vaccines. ^{152–154}

Newer, accurate, cheap and reproducible sequencing technologies; validated databases containing genotype-phenotype data; statistical and bioinformatics tools are needed in order to analyze and interpret data that will help and improve vaccine adverse and immune response quantifiability and predictability. The information will enhance clinical practice and accelerate rational and directed vaccine development.

The Pregnant Women

Safe vaccines are a critical requirement for any immunization program. Conventional vaccination has been an approach targeted at all groups and individuals but has failed toward the enrolment of pregnant women into vaccination programs because of presumed fetal and maternal harms. Evidence on the safety of vaccination in pregnancy is very small because pregnant women were usually excluded from participating in vaccine trials. Pregnancy can alter the maternal as well as fetal immunological responses. It is pertinent to explore research opportunities presented in advanced vaccine designs such as immunoinformatics (multiepitope vaccine docking) by studying human immune system functions and responses specific to pregnant women and their unborn children.

According to a report¹⁶¹ from the Dominican Republic of Congo, during the 2016–2017 Zika virus outbreaks, over a thousand pregnant women were suspected of being infected with the virus and a sizable number were at their first trimester. The report further stated that fetal loss was approximately one-tenth of the pregnancies and that there were up to 3 cases of fetus with head circumferences smaller than normal. The widespread morbidity during the epidemic showed that Zika virus infection adversely affects pregnancy outcome. ^{160,161}

Currently, there is no proof that pregnancy predisposes to Ebola virus infections in comparison with the non-

pregnant population, but there is some evidence suggesting pregnancy to worsen the disease prognosis including fetal loss. Also, evidence showed that the virus can pass the placental barriers to establish infection in the unborn child. Designing, implementing and enrolling pregnant women as well as perspective pregnant women into vaccine trials and programs is imperative in order to protect this group and ensure good vaccine uptake by them during infection outbreaks and epidemics. 157,163

These recommendations will give an informed decision to be investigated using the immunoinformatic tools to determine the immunogenic responses worthy of safe vaccine development for the pregnant women and perspective pregnant women group.

Maternal immunization offers palpable benefits in several ways. Some vaccines are primarily administered to shield these pregnant women from morbid conditions and/or death including fetal death. Pregnant women stand the risk of being exposed to virulent pathogens and may be at a higher risk of morbidity and/or mortality when compared to the general population. 166

Conclusion and Future Research Directions

There has been an explosion of new immunological data (Table 4) due to an increase in research to understand the immune system pathway in infectious disease pathogenesis and the application of the knowledge of bioinformatics has led to a better exposition of the immune system importance through immunoinformatics. The knowledge of immune system and the cost-effective, specific and effective approach like immunoinformatics, the concerns for emerging and re-surging diseases caused by pathogenic organisms, antigenic variability/complex lifecycle of pathogens and the need of personalized vaccination can be combated on a molecular level.

The future of immunological research is sharpened by the ability to make discoveries in biologics (e.g., vaccines) more effectively and efficiently. This will mean reduction and better targeting of wet laboratory experiments and will only be possible if wet laboratory experimentation is combined with bioinformatics techniques.

Limitation of Immunoinformatics Predictions in Vaccine Design

• Immunoinformatics depends on experimental science (wet laboratory) to produce raw data for analysis.

The predictions are not formal proofs of any concepts. They do not replace the traditional experimental research methods of actually testing hypotheses.

 The quality of immunoinformatics predictions depends on the quality of data and the sophistication of the algorithms being used. Sequence data from high-throughput analysis often contain errors. If the sequences are wrong, or annotations incorrect, the results from the downstream analysis are misleading as well.¹⁶⁷

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Availability of Data

All the needed data are included in this manuscript.

Author Contributions

All authors contributed to data analysis, drafting or revising the article, gave final approval of the version to be published, and agree to be accountable for all aspects of the work.

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References

- Terry FE, Moise L, Martin RF, et al. Time for T? Immunoinformatics addresses vaccine design for neglected tropical and emerging infectious diseases. Expert Rev Vaccines. 2015;14(1):21–35. doi:10.1586/ 14760584.2015.955478
- Poland GA, Whitaker JA, Poland CM, Ovsyannikova IG, Kennedy RB. Vaccinology in the third millennium: scientific and social challenges. *Curr Opin Virol*. 2016;17:116–125. doi:10.1016/j. coviro.2016.03.003
- Servín-Blanco R, Zamora-Alvarado R, Gevorkian G, Manoutcharian K. Antigenic variability: obstacles on the road to vaccines against traditionally difficult targets. *Hum Vaccin Immunother*. 2016;12(10):2640–2648. doi:10.1080/21645515.2016.11 91718
- Tomaras GD, Plotkin SA. Complex immune correlates of protection in HIV-1 vaccine efficacy trials. *Immunol Rev.* 2017;275(1):245–261. doi:10.1111/jmr.12514
- Urrutia-Baca VH, Gomez-Flores R, De La Garza-ramos MA, Tamez-Guerra P, Lucio-Sauceda DG, Rodríguez-Padilla MC. Immunoinformatics approach to design a novel epitope-based oral vaccine against *Helicobacter pylori*. *J Comput Biol*. 2019;(2019 (May):23. doi:10.1089/cmb.2019.0062
- Ali SA, Almofti YA, Abd-Elrahman KA. Immunoinformatics approach for multiepitopes vaccine prediction against glycoprotein B of avian infectious laryngotracheitis virus. Adv Bioinformatics. 2019;2019(1270485):1–23. doi:10.1155/2019/1270485

 Manzoni C, Kia DA, Vandrovcova J, et al. Genome, transcriptome and proteome: the rise of omics data and their integration in biomedical sciences. *Brief Bioinform*. 2018;19(2):286–302. doi:10.1093/bib/bbw114

- Peng M, Mo Y, Wang Y, et al. Neoantigen vaccine: an emerging tumor immunotherapy. *Mol Cancer*. 2019;18(1):128. doi:10.1186/ s12943-019-1055-6
- Kaliamurthi S, Selvaraj G, Kaushik AC, Gu KR, Wei DQ. Designing of CD8⁺ and CD8⁺-overlapped CD4⁺ epitope vaccine by targeting late and early proteins of human papillomavirus. *Biologics*. 2018;12:107–125. doi:10.2147/BTT.S177901.
- Kim HI, Ha NY, Kim G, et al. Immunization with a recombinant antigen composed of conserved blocks from TSA56 provides broad genotype protection against scrub typhus. *Emerg Microbes Infect*. 2019;8(1):946–958. doi:10.1080/22221751.2019.1632676
- Richner JM, Jagger BW, Shan C, et al. Vaccine mediated protection against zika virus-induced congenital disease. *Cell.* 2017;170 (2):273–283. e12. doi:10.1016/j.cell.2017.06.040
- Goh YS, McGuire D, Rénia L. Vaccination with sporozoites: models and correlates of protection. *Front Immunol*. 2019;10:1227. doi:10.3389/fimmu.2019.01227
- Wallis J, Shenton DP, Carlisle RC. Novel approaches for the design, delivery and administration of vaccine technologies. *Clin Exp Immunol*. 2019;196(2):189–204. doi:10.1111/cei.13287
- Sarkander J, Hojyo S, Tokoyoda K. Vaccination to gain humoral immune memory. Clin Transl Immunol. 2016;5(12):e120. doi:10.1038/cti.2016.81
- Giacomet V, Masetti M, Nannini P, et al. Humoral and cell-mediated immune responses after a booster dose of HBV vaccine in HIV-infected children, adolescents and young adults. *PLoS One*. 2018;13(2):e0192638. doi:10.1371/journal.pone.0192638
- Oli AN, Agu RU, Ihekwereme CP, Esimone CO. An evaluation of the cold chain technology in South-East, Nigeria using Immunogenicity study on the measles vaccines. *Pan Afr Med J.* 2017;27(Suppl 3):28. doi:10.11604/pamj.supp.2017.27.3.11491
- Munang'andu HM, Mutoloki S, Evensen Ø. A review of the immunological mechanisms following mucosal vaccination of finfish. Front Immunol. 2015;6:427. doi:10.3389/fimmu.2015.00427
- Rappuoli R, Bottomley MJ, D'Oro U, Finco O, De Gregorio E. Reverse vaccinology 2.0: human immunology instructs vaccine antigen design. *J Exp Med.* 2016;213(4):469–481. doi:10.1084/ jem.20151960
- Davies DH, Duffy P, Bodmer J-L, Felgner PL, Doolan DL. Large screen approaches to identify novel malaria vaccine candidates. *Vaccine*. 2015;33(52):7496–7505. doi:10.1016/j.vaccine.2015.09.059
- Vilela Rodrigues TC, Jaiswal AK, de Sarom A, et al. Reverse vaccinology and subtractive genomics reveal new therapeutic targets against *Mycoplasma pneumoniae*: a causative agent of pneumonia. *R Soc Open Sci.* 2019;6(7):190907. doi:10.1098/rsos.190907
- Simon AK, Hollander GA, McMichael A. Evolution of the immune system in humans from infancy to old age. *Proc Biol Sci.* 2015;282 (1821):20143085. doi:10.1098/rspb.2014.3085
- Nikolich-Žugich J. The twilight of immunity: emerging concepts in aging of the immune system. *Nat Immunol*. 2018;19:10–19. doi:10.1038/s41590-017-0006-x
- Berical AC, Harris D, Dela Cruz CS, Possick JD. Pneumococcal vaccination strategies. An update and perspective. *Ann Am Thorac* Soc. 2016;13(6):933–944. doi:10.1513/AnnalsATS.201511-778FR;
- Voss G, Casimiro D, Neyrolles O, et al. Progress and challenges in TB vaccine development. F1000Res. 2018;7:199. doi:10.12688/ f1000research.13588.1
- Weinberger B. Vaccines for the elderly: current use and future challenges. *Immun Ageing*. 2018;15:3. doi:10.1186/s12979-017-0107-2
- Palm AE, Henry C. Remembrance of things past: long-term B cell memory after infection and vaccination. Front Immunol. 2019;10:1787. doi:10.3389/fimmu.2019.01787

 Ali A, Jafri RZ, Messonnier N, et al. Global practices of meningococcal vaccine use and impact on invasive disease. *Pathog Glob Health*. 2014;108(1):11–20. doi:10.1179/2047773214Y.0000000126

- Rauch S, Jasny E, Schmidt KE, Petsch B. New vaccine technologies to combat outbreak situations. *Front Immunol*. 2018;9:1963. doi:10.3389/fimmu.2018.01963
- Burton DR. What are the most powerful immunogen design vaccine strategies? Reverse vaccinology 2.0 shows great promise. *Cold Spring Harb Perspect Biol.* 2017;9(11):a030262. doi:10.1101/cshperspect.a030262
- Paul A. The new malaria vaccine program for African children is promising but still quite limited. Quartz Africa. 2019April 28.
 Available from: https://qz.com/africa/1606511/malaria-vaccine-by-gsk-for-african-children-is-still-too-limited/. Accessed 26 Sept 2019.
- Ashley T. Merck's Ebola vaccine helps combat deadly outbreak in the Congo as the virus spreads. BIOTECH and PHARMA. 2019.
 Available from: https://www.cnbc.com/2019/07/09/mercks-ebola-vaccine-helps-combat-deadly-outbreak-in-the-congo.html.
 Accessed 2019.
- Hotez PJ, Dumonteil E, Heffernan MJ, Bottazzi ME. Innovation for the 'bottom 100 million': eliminating neglected tropical diseases in the Americas. Adv Exp Med Biol. 2013;764:1–12.
- Nii-Trebi NI. Emerging and neglected infectious diseases: insights, advances, and challenges. *Biomed Res Int.* 2017;2017:5245021. doi:10.1155/2017/5245021
- Pritam M, Singh G, Swaroop S, Singh AK, Singh SP. Exploitation of reverse vaccinology and immunoinformatics as promising platform for genome-wide screening of new effective vaccine candidates against *Plasmodium falciparum*. *BMC Bioinformatics*. 2019;19:468. doi:10.1186/s12859-018-2482-x
- Hegde NR, Gauthami S, Sampath Kumar HM, Bayry J. The use of databases, data mining and immunoinformatics in vaccinology: where are we? *Expert Opin Drug Discov.* 2018;13(2):117–130. doi:10.1080/17460441.2018.1413088
- Raeven R, van Riet E, Meiring HD, Metz B, Kersten G. Systems vaccinology and big data in the vaccine development chain. *Immunology*. 2019;156(1):33–46. doi:10.1111/imm.13012
- Mills CL, Beuning PJ, Ondrechen MJ. Biochemical functional predictions for protein structures of unknown or uncertain function. *Comput Struct Biotechnol J.* 2015;13:182–191. doi:10.10 16/j.csbj.2015.02.003
- Michalik M, Djahanshiri B, Leo JC, Linke D. Reverse vaccinology: the pathway from genomes and epitope predictions to tailored recombinant vaccines. In: Thomas S, editor. *Vaccine Design. Methods in Molecular Biology.* Vol. 1403. New York, NY: Humana Press; 2016: 87–106. doi:10.1007/978-1-4939-3387-7 4
- Guthrie JL, Kong C, Roth D, et al. Universal genotyping for tuberculosis prevention programs: a 5-year comparison with on-request genotyping. *J Clin Microbiol*. 2018;56(5):e01778–17. doi:10.1128/JCM.01778-17
- Collineau L, Boerlin P, Carson CA, et al. Integrating Whole-genome sequencing data into quantitative risk assessment of foodborne antimicrobial resistance: a review of opportunities and challenges. Front Microbiol. 2019;10:1107. doi:10.3389/ fmicb.2019.01107
- Xia X. Bioinformatics and drug discovery. Curr Top Med Chem. 2017;17(15):1709–1726. doi:10.2174/1568026617666161116143440
- Christaki E. New technologies in predicting, preventing and controlling emerging infectious diseases. *Virulence*. 2015;6(6):55 8–565. doi:10.1080/21505594.2015.1040975
- Mottram L, Chakraborty S, Cox E, Fleckenstein J. How genomics can be used to understand host susceptibility to enteric infection, aiding in the development of vaccines and immunotherapeutic interventions. *Vaccine*. 2019;37(34):4805–4810. doi:10.1016/j. vaccine.2019.01.016

 Zheng J, Lin X, Wang X, et al. In silico analysis of epitope-based vaccine candidates against hepatitis B virus polymerase protein. Viruses. 2017;9(5):112. doi:10.3390/v9050112

- Dadar M, Chakraborty S, Dhama K, et al. Advances in designing and developing vaccines, drugs and therapeutic approaches to counter human papilloma virus. Front Immunol. 2018;9:2478. doi:10.3389/fimmu.2018.02478
- Qiu X, Duvvuri VR, Bahl J. Computational approaches and challenges to developing universal influenza vaccines. *Vaccines* (*Basel*). 2019;7(2):45. doi:10.3390/vaccines7020045.
- 47. Imle A, Kumberger P, Schnellbächer ND, et al. Experimental and computational analyses reveal that environmental restrictions shape HIV-1 spread in 3D cultures. *Nat Commun.* 2019;10(1):2144. doi:10.1038/s41467-019-09879-3
- Miller SE, Bell CS, McClain MS, Cover TL, Giorgio TD. Dynamic computational model of symptomatic bacteremia to inform bacterial separation treatment requirements. *PLoS One*. 2016;11(9): e0163167. doi:10.1371/journal.pone.0163167
- Fonseca LL, Joyner CJ, Galinski MR, Voit EO. A model of *Plasmodium vivax* concealment based on *Plasmodium cynomolgi* infections in *Macaca mulatta*. *Malar J*. 2017;16(1):375. doi:10.1186/s12936-017-2008-4
- 50. Utesch T, de Miguel Catalina A, Schattenberg C, et al. A computational modeling approach predicts interaction of the antifungal protein AFP from *Aspergillus giganteus* with fungal membranes via its γ-core motif. *mSphere*. 2018;3(5):e00377–18. doi:10.1128/mSphere.00377-18
- Soria-Guerra RE, Nieto-Gomez R, Govea-Alonso DO, Rosales-Mendoza S. An overview of bioinformatics tools for epitope prediction: implications on vaccine development. *J Biomed Inform*. 2015;53:405–414. doi:10.1016/j.jbi.2014.11.003
- Martin W, Sbai H, De Groot AS. Bioinformatics tools for identifying class I-restricted epitopes. *Methods*. 2003;29(3):289–298. doi:10.1016/S1046-2023(02)00351-1
- Moise L, McMurry JA, Buus S, Frey S, Martin WD, De Groot AS. In silico-accelerated identification of conserved and immunogenic variola/vaccinia T-cell epitopes. *Vaccine*. 2009;27(46):6471–6479. doi:10.1016/j.vaccine.2009.06.018
- 54. Pisitkun T, Hoffert JD, Saeed F, Knepper MA. NHLBI-AbDesigner: an online tool for design of peptide-directed antibodies. *Am J Physiol*. 2012;302(1):C154–C164. doi:10.1152/ajpcell.00325.2011
- 55. Moise L, Gutierrez A, Kibria F, et al. iVAX: an integrated toolkit for the selection and optimization of antigens and the design of epitope-driven vaccines. *Hum Vaccin Immunother*. 2015;11 (9):2312–2321. doi:10.1080/21645515.2015.1061159
- De Groot AS. iVAX EpiVax, Inc; 2013. Available from: www. epivax.com/wp-content//EpiVax_AAI_AAPS_iVAX_FINAL_ 29Apr13.pdf. Accessed 29 Dec, 2017.
- De Groot AS, Bosma A, Chinai N, et al. From genome to vaccine: in silico predictions, ex vivo verification. *Vaccine*. 2015;19 (31):4385–4395. doi:10.1016/S0264-410X(01)00145-1
- Schubert B, Kohlbacher O. Designing string-of-beads vaccines with optimal spacers. *Genome Med.* 2016;8:9. doi:10.1186/s13073-016-0263-6
- De Groot Anne S, Luisa M, Bishop Elizabeth A, et al. HIV vaccine development by computer assisted design: the GAIA vaccine. Vaccine. 2005;18:2136–48. doi:10.1016/j.vaccine.2005.01.097
- Vivona S, Filippo B, Francesco F. NERVE: new Enhanced reverse vaccinology environment. *BMC Biotechnol*. 2006;6:35. doi:10.1186/ 1472-6750-6-35
- Jaiswal V, Chanumolu SK, Gupta A, Chauhan RS, Rout C. Jennerpredict server: prediction of protein vaccine candidates (PVCs) in bacteria based on host-pathogen interactions. *BMC Bioinformatics*. 2013;14:211. doi:10.1186/1471-2105-14-211

- 62. Xiang Z, He Y. Genome-wide prediction of vaccine target of human herpes simplex viruses using Vaxign RV. BMC Bioinformatics. 2013;14(Suppl4):S2(8March 2013). doi:10.1186/ 1471-2105-14-S4-S2
- Doytchinova IA, Flower DR. VaxiJen: a server for prediction of protective antigens, tumour antigens and subunit vaccines. *BMC Bioinformatics*. 2007;8:4. doi:10.1186/1471-2105-8-4
- 64. Rizwan M, Naz A, Ahmad J, et al. VacSol: a high throughput in silico pipeline to predict potential therapeutic targets in prokaryotic pathogens using subtractive reverse vaccinology. BMC Bioinformatics. 2017;18(1):106. doi:10.1186/s12859-017-1540-0
- Naz K, Naz A, Ashraf ST, et al. PanRV: pangenome-reverse vaccinology approach for identifications of potential vaccine candidates in microbial pangenome. *BMC Bioinformatics*. 2019;20(1):123. doi:10.1186/s12859-019-2713-9
- 66. National Institutes of Health (US); Biological Sciences Curriculum Study. NIH Curriculum Supplement Series [Internet]. Understanding Emerging and Re-Emerging Infectious Diseases. Bethesda (MD: National Institutes of Health (US); 2007. Available from: https://www. ncbi.nlm.nih.gov/books/NBK20370/. Accessed February 10, 2020.
- Lindahl JF, Grace D. The consequences of human actions on risks for infectious diseases: a review. *Infect Ecol Epidemiol*. 2015;5:30048. doi:10.3402/iee.v5.30048
- 68. Institute of Medicine (US) Forum on Microbial Threats. Microbial evolution and co-adaptation: a tribute to the life and scientific legacies of joshua lederberg: workshop summary. Washington (DC): National Academies Press (US); 2009.: doi: 10.17226/12586
- Mhondoro M, Ndlovu N, Bangure D, et al. Trends in antimicrobial resistance of bacterial pathogens in Harare, Zimbabwe, 2012–2017: a secondary dataset analysis. *BMC Infect Dis.* 2019;19(1):746. doi:10.1186/s12879-019-4295-6
- National Institutes of Health (US); Biological Sciences Curriculum Study. NIH Curriculum Supplement Series [Internet]. Understanding Emerging and Re-Emerging Infectious Diseases. Bethesda (MD): National Institutes of Health (US); 2007. Available from https://www.ncbi.nlm.nih.gov/books/NBK20370/. Accessed 19 November 2019.
- Leventhal GE, Hill AL, Nowak MA, Bonhoeffer S. Evolution and emergence of infectious diseases in theoretical and real-world networks. *Nat Commun.* 2015;6:6101. doi:10.1038/ncomms7101
- Oli AN, Itumo CJ, Okam PC, et al. Carbapenem-resistant enterobacteriaceae posing a dilemma in effective healthcare delivery. *Antibiotics (Basel)*. 2019;8(4). doi:10.3390/antibiotics8040156
- 73. WHO Research and Development Blueprint. A research and development blueprint for action to prevent epidemics, 2018 annual review of diseases prioritized under the research and development blueprint. Available from: http://www.who.int/csr/research-and-development /en/. Accessed February 10, 2020.Accessed February 5, 2019
- Singh L, Kruger HG, Maguire GEM, Govender T, Parboosing R. The role of nanotechnology in the treatment of viral infections. Ther Adv Infect Dis. 2017;4(4):105–131. doi:10.1177/204993611 7713593
- Adamczyk-Poplawska M, Markowicz S, Jagusztyn-Krynicka EK. Proteomics for development of vaccine. *J Proteomics*. 2011;74 (12):2596–2616. doi:10.1016/j.jprot.2011.01.019
- Sliwoski G, Kothiwale S, Meiler J, Lowe EW Jr. Computational methods in drug discovery. *Pharmacol Rev.* 2013;66(1):334–395. doi:10.1124/pr.112.007336
- Nourani E, Khunjush F, Durmuş S. Computational approaches for prediction of pathogen-host protein-protein interactions. *Front Microbiol.* 2015;6:94. doi:10.3389/fmicb.2015.00094
- Furman D, Davis MM. New approaches to understanding the immune response to vaccination and infection. *Vaccine*. 2015;33 (40):5271–5281. doi:10.1016/j.vaccine.2015.06.117

 Paust S, Senman B, von Andrian UH. Adaptive immune responses mediated by natural killer cells. *Immunol Rev.* 2010;235 (1):286–296. doi:10.1111/j.0105-2896.2010.00906.x

- Dittmer U, Sutter K, Kassiotis G, et al. Friend retrovirus studies reveal complex interactions between intrinsic, innate and adaptive immunity. FEMS Microbiol Rev. 2019;43(5):435–456. doi:10.1093/ femsre/fuz012
- Delgado Betancourt E, Hamid B, Fabian BT, Klotz C, Hartmann S, Seeber F. From entry to early dissemination-*Toxoplasma* gondii's initial encounter with its host. *Front Cell Infect Microbiol*. 2019;9:46. doi:10.3389/fcimb.2019.00046
- Rash A, Morton R, Woodward A, et al. Evolution and divergence of H3N8 equine influenza viruses circulating in the United Kingdom from 2013 to 2015. *Pathogens*. 2017;6(1):6. doi:10.33 90/pathogens6010006
- Harrison OB, Schoen C, Retchless AC, et al. Neisseria genomics: current status and future perspectives. Pathog Dis. 2017;75(6): ftx060. doi:10.1093/femspd/ftx060
- Recker M, Buckee CO, Serazin A, et al. Antigenic variation in Plasmodium falciparum malaria involves a highly structured switching pattern. *PLoS Pathog*. 2011;7(3):e1001306. doi:10.13 71/journal.ppat.1001306
- Ella R, Steven SH. Neisseria gonorrhoeae MutS affects pilin antigenic variation through mismatch correction and not by pilE guanine quartet binding. J Bacteriol. 2015;197. 1828–1838. doi:10.11 28/JB.02594-14.
- Palmer GH, Bankhead T, Seifert HS. Antigenic variation in bacterial pathogens. *Microbiology Spectrum*. 2016;4(1). doi:10.1128/microbiolspec.VMBF-0005-2015
- Ibáñez de Aldecoa AL, Zafra O, González-Pastor JE. Mechanisms and regulation of extracellular DNA release and its biological roles in microbial communities. *Front Microbiol*. 2017;8:1390. doi:10. 3389/fmicb.2017.01390
- Cehovin A, Lewis SB. Mobile genetic elements in *Neisseria gonor-rhoeae*: movement for change. *Pathog Dis.* 2017;75(6):ftx071. doi:10.1093/femspd/ftx071
- Diallo K, MacLennan J, Harrison OB, et al. Genomic characterization of novel Neisseria species. *Sci Rep.* 2019;9(1):13742. doi:10. 1038/s41598-019-50203-2
- Henriques-Normark B, Tuomanen EI. The pneumococcus: epidemiology, microbiology, and pathogenesis. *Cold Spring Harb Perspect Med.* 2013;3(7):a010215. doi:10.1101/cshperspect. a010215
- Morais V, Dee V, Suárez N. Purification of capsular polysaccharides of *Streptococcus pneumoniae*: traditional and new methods. *Front Bioeng Biotechnol*. 2018;6:145. doi:10.3389/fbioe.2018.00
- KA G, Gilbert GL, JY S, et al. Pneumococcal capsules and their types: past, present, and future. *Clin Microbiol Rev.* 2015;28 (3):871–899. doi:10.1128/CMR.00024-15
- Quainoo S, Coolen JPM, van Hijum SAFT, et al. Whole-genome sequencing of bacterial pathogens: the future of nosocomial outbreak analysis. *Clin Microbiol Rev.* 2017;30(4):1015–1063. doi:10. 1128/CMR.00016-17
- Mboowa G, Sserwadda I, Amujal M, Namatovu N. Human genomic loci important in common infectious diseases: role of high-throughput sequencing and genome-wide association studies. Can J Infect Dis Med Microbiol. 2018;2018:1875217. doi:10.1155/2018/1875217
- Belachew Esmael B. Immune response and evasion mechanisms of Plasmodium falciparum parasites. J Immunol Res. 2018; Article ID 6529681:6. doi:10.1155/2018/6529681
- Sampaio NG, Eriksson EM, Schofield L. Plasmodium falciparum PfEMP1 modulates monocyte/macrophage transcription factor activation and cytokine and chemokine responses. Infect Immun. 2017;86(1):e00447–17. doi:10.1128/IAI.00447-17

 Hermand P, Cicéron L, Pionneau C, Vaquero C, Combadière C, Deterre P. Plasmodium falciparum proteins involved in cytoadherence of infected erythrocytes to chemokine CX3CL1. *Sci Rep.* 2016;6:33786. doi:10.1038/srep33786

- Khoury DS, Cromer D, Best SE, et al. Effect of mature blood-stage Plasmodium parasite sequestration on pathogen biomass in mathematical and in vivo models of malaria. *Infect Immun*. 2014;82 (1):212–220. doi:10.1128/IAI.00705-13
- Bachmann A, Bruske E, Krumkamp R, et al. Controlled human malaria infection with *Plasmodium falciparum* demonstrates impact of naturally acquired immunity on virulence gene expression. *PLoS Pathog*. 2019;15(7):e1007906. doi:10.1371/journal.ppat.1007906
- 100. Jeppesen A, Ditlev SB, Soroka V, et al. Multiple *Plasmodium falciparum* erythrocyte membrane protein 1 variants per genome can bind IgM via Its Fc fragment Fcμ. *Infect Immun*. 2015;83 (10):3972–3981. doi:10.1128/IAI.00337-15
- 101. Abdi AI, Hodgson SH, Muthui MK, et al. Plasmodium falciparum malaria parasite var gene expression is modified by host antibodies: longitudinal evidence from controlled infections of Kenyan adults with varying natural exposure. *BMC Infect Dis.* 2017;17(1):585. doi:10.1186/s12879-017-2686-0
- 102. Mira-Martínez S, van Schuppen E, Amambua-Ngwa A, et al. Expression of the Plasmodium falciparum Clonally variant clag3 genes in human infections. *J Infect Dis.* 2017;215(6):938–945. doi:10.1093/infdis/jix053
- 103. Hayes CN, Diez D, Joannin N, et al. varDB: a pathogen-specific sequence database of protein families involved in antigenic variation. *Bioinformatics*. 2008;24(21):2564–2565. doi:10.1093/ bioinformatics/btn477
- 104. Gangnard S, Chêne A, Dechavanne S, et al. VAR2CSA binding phenotype has ancient origin and arose before *Plasmodium fal*ciparum crossed to humans: implications in placental malaria vaccine design. Sci Rep. 2019;9(1):16978. doi:10.1038/s41598-019-53334-8
- 105. Benavente ED, Oresegun DR, de Sessions PF, et al. Global genetic diversity of var2csa in *Plasmodium falciparum* with implications for malaria in pregnancy and vaccine development. *Sci Rep.* 2018;8 (1):15429. doi:10.1038/s41598-018-33767-3
- 106. Schwede A, Macleod OJ, MacGregor P, Carrington M. How does the VSG coat of bloodstream form African trypanosomes interact with external proteins? *PLoS Pathog*. 2015;11(12):e1005259. doi:10.1371/journal.ppat.1005259
- Pinger J, Chowdhury S, Papavasiliou FN. Variant surface glycoprotein density defines an immune evasion threshold for African trypanosomes undergoing antigenic variation. *Nat Commun.* 2017;8 (1):828. doi:10.1038/s41467-017-00959-w
- 108. Sautto GA, Kirchenbaum GA, Ross TM. Towards a universal influenza vaccine: different approaches for one goal. *Virol J.* 2018;15(1):17. doi:10.1186/s12985-017-0918-y
- 109. Kumar B, Asha K, Khanna M, Ronsard L, Meseko CA, Sanicas M. The emerging influenza virus threat: status and new prospects for its therapy and control. *Arch Virol*. 2018;163(4):831–844. doi:10. 1007/s00705-018-3708-y
- 110. Hensley SE, Das SR, Gibbs JS, et al. Influenza A virus hemagglutinin antibody escape promotes neuraminidase antigenic variation and drug resistance. *PLoS One*. 2011;6(2):e15190. doi:10.1371/journal.pone.0015190
- 111. Singh RK, Dhama K, Karthik K, et al. A comprehensive review on equine influenza virus: etiology, epidemiology, pathobiology, advances in developing diagnostics, vaccines, and control strategies. Front Microbiol. 2018;9:1941. doi:10.3389/fmicb.2018. 01941
- Kumar A, Meldgaard TS, Bertholet S. Novel platforms for the development of a universal influenza vaccine. Front Immunol. 2018;9(600). doi:10.3389/fimmu.2018.00600

113. Faisal AM, Imtiaz SH, Zerin T, Rahman T, Shekhar HU. Computer aided epitope design as a peptide vaccine component against Lassa virus. *Bioinformation*. 2017;13(12):417–429. doi:10.6026/9732063 0013417

- 114. Singh SP, Srivastava D, Mishra BN. Genome-wide identification of novel vaccine candidates for *Plasmodium falciparum* malaria using integrative bioinformatics approaches. *3 Biotech*. 2017;7(5):318. doi:10.1007/s13205-017-0947-7
- 115. Shey RA, Ghogomu SM, Esoh KK, et al. In-silico design of a multi-epitope vaccine candidate against onchocerciasis and related filarial diseases. *Sci Rep.* 2019;9(1):4409. doi:10.1038/s41598-019-40833-x
- Ernst JD. Antigenic variation and immune escape in the MTBC.
 Adv Exp Med Biol. 2017;1019:171–190. doi:10.1007/978-3-319-64371-7
- 117. Oany AR, Pervin T, Mia M, et al. Vaccinomics approach for designing potential peptide vaccine by targeting *Shigella* spp. Serine Protease autotransporter subfamily protein SigA. *J Immunol Res*. 2017;2017 (6412353):1–14. doi:10.1155/2017/6412353
- 118. Chauhan V, Rungta T, Goyal K, Singh MP. Designing a multi-epitope based vaccine to combat Kaposi Sarcoma utilizing immunoinformatics approach. *Sci Rep.* 2019;9(1):2517. doi:10.10 38/s41598-019-39299-8
- 119. He R, Yang X, Liu C, et al. Efficient control of chronic LCMV infection by a CD4 T cell epitope-based heterologous prime-boost vaccination in a murine model. *Cell Mol Immunol*. 2018;15 (9):815–826. doi:10.1038/cmi.2017.3
- 120. Lu C, Meng S, Jin Y, et al. A novel multi-epitope vaccine from MMSA-1 and DKK1 for multiple myeloma immunotherapy. Br J Haematol. 2017;178(3):413–426. doi:10.1111/bjh.14686
- Jiang P, Cai Y, Chen J, et al. Evaluation of tandem *Chlamydia trachomatis* MOMP multi-epitopes vaccine in BALB/c mice model. Vaccine. 2017;35(23):3096–3103. doi:10.1016/j.vaccine.2017.04.031
- 122. Saadi M, Karkhah A, Nouri HR. Development of a multi-epitope peptide vaccine inducing robust T cell responses against brucellosis using immunoinformatics based approaches. *Infect Genet Evol.* 2017;51::227–234. doi:10.1016/j.meegid.2017.04.009
- 123. Lu IN, Farinelle S, Sausy A, Muller CP. Identification of a CD4 T-cell epitope in the hemagglutinin stalk domain of pandemic H1N1 influenza virus and its antigen-driven TCR usage signature in BALB/c mice. Cell Mol Immunol. 2017;14(6):511–520. doi:10.1038/cmi.2016.20
- 124. Lamiable A, Thévenet P, Rey J, Vavrusa M, Derreumaux P, Tufféry P. PEP-FOLD3: faster de novo structure prediction for linear peptides in solution and in complex. *Nucleic Acids Res*. 2016;44(W1):W449–54. doi:10.1093/nar/gkw329
- Haas J, Roth S, Arnold K, et al. The Protein Model Portal—a comprehensive resource for protein structure and model information. *Database (Oxford)*. 2013;2013:bat031. doi:10.1093/database/bat031
- 126. Khatoon N, Pandey RK, Prajapati VK. Exploring Leishmania secretory proteins to design B and T cell multi-epitope subunit vaccine using immunoinformatics approach. Sci Rep. 2017;7 (1):8285. doi:10.1038/s41598-017-08842-w
- Atsaves V, Leventaki V, Rassidakis GZ, Claret FX. AP-1 transcription factors as regulators of immune responses in cancer. *Cancers (Basel)*. 2019;11(7):1037. doi:10.3390/cancers11071037
- 128. Poland GA, Kennedy RB, Ovsyannikova IG. Vaccinomics and personalized vaccinology: is science leading us toward a new path of directed vaccine development and discovery? *PLoS Pathog.* 2011;7(12):e1002344. doi:10.1371/journal.ppat.1002344
- 129. Piasecka B, Duffy D, Urrutia A, et al., Milieu Intérieur Consortium. Distinctive roles of age, sex, and genetics in shaping transcriptional variation of human immune responses to microbial challenges. *Proc Natl Acad Sci U S A*. 2018;115(3):E488–E497. doi:10.1073/ pnas.1714765115

 Odimegwu DC, Onah AI, Ifudu F, Oli AN. Evaluation of levamisole as an adjuvant for typhoid fever vaccine formulation. *PhOL*. 2019;2:336–346.

- Blackwell JM, Jamieson SE, Burgner D. HLA and infectious diseases. Clin Microbiol Rev. 2009;22(2):370–85, Table of Contents. doi:10.1128/CMR.00048-08
- 132. Gfeller D, Bassani-Sternberg M. Predicting antigen presentation-what could we learn from a million peptides? *Front Immunol.* 2018;9:1716. doi:10.3389/fimmu.2018.01716
- Wang Y. Development of a human leukocyte antigen-based HIV vaccine. F1000Res. 2018;7:F1000Faculty Rev–874. doi:10.12688/ f1000research.13759.1
- 134. Buhler S, Sanchez-Mazas A. HLA DNA sequence variation among human populations: molecular signatures of demographic and selective events. *PLoS One*. 2011;6(2):e14643. doi:10.1371/journal.pone.0014643
- 135. Cruz-Tapias P, Castiblanco J, Anaya JM. Major histocompatibility complex: antigen processing and presentation. In: Anaya JM, Shoenfeld Y, Rojas-Villarraga A, et al. editors. Autoimmunity: From Bench to Bedside [Internet]. Bogota (Colombia): El Rosario University Press; 2013 Jul 18. Chapter 10 Available from: https://www.ncbi.nlm.nih.gov/books/NBK459467/. Accessed October 1, 2019.
- 136. Clemens EB, van de Sandt C, Wong SS, Wakim LM, Valkenburg SA. Harnessing the power of T cells: the promising hope for a universal influenza vaccine. *Vaccines (Basel)*. 2018;6 (2):18. doi:10.3390/vaccines6020018
- Auladell M, Jia X, Hensen L, et al. Recalling the future: immunological memory toward unpredictable influenza viruses. Front Immunol. 2019;10:1400. doi:10.3389/fimmu.2019.01400
- 138. Chandra H, Yadav JS. Human leukocyte antigen (HLA)-binding epitopes dataset for the newly identified T-cell antigens of Mycobacterium immunogenum. Data Brief. 2016;8:1069–1071. doi:10.1016/j.dib.2016.06.045
- 139. Sanchez-Trincado JL, Gomez-Perosanz M, Reche PA. Fundamentals and methods for T- and B-cell epitope prediction. J Immunol Res. 2017;2017:2680160. doi:10.1155/2017/2680160
- 140. Crux NB, Elahi S. Human Leukocyte Antigen (HLA) and immune regulation: how do classical and non-classical HLA alleles modulate immune response to human immunodeficiency virus and hepatitis C virus infections? Front Immunol. 2017;8:832. doi:10.33 89/fimmu.2017.00832
- 141. Molineros JE, Looger LL, Kim K, et al. Amino acid signatures of HLA Class-I and II molecules are strongly associated with SLE susceptibility and autoantibody production in Eastern Asians. *PLoS Genet*. 2019;15(4):e1008092. doi:10.1371/journal.pgen.1008092
- 142. Binkowski TA, Marino SR, Joachimiak A. Predicting HLA class I non-permissive amino acid residues substitutions. *PLoS One*. 2012;7(8):e41710. doi:10.1371/journal.pone.0041710
- 143. Lundegaard C, Lund O, Buus S, Nielsen M. Major histocompatibility complex class I binding predictions as a tool in epitope discovery. *Immunology*. 2010;130(3):309–318. doi:10.1111/j.1365-2567.2010.03300.x
- 144. Reche PA, Glutting JP, Reinherz EL. Prediction of MHC class I binding peptides using profile motifs. *Hum Immunol*. 2002;63 (9):701–709. doi:10.16/s0198-8859(02)00432-9
- 145. Hossain MU, Omar TM, Oany AR, et al. Design of peptide-based epitope vaccine and further binding site scrutiny led to groundswell in drug discovery against Lassa virus. 3 Biotech. 2018;8(2):81. doi:10.1007/s13205-018-1106-5
- 146. Adhikari UK, Tayebi M, Rahman MM. Immunoinformatics Approach for epitope-based peptide vaccine design and active site prediction against polyprotein of emerging oropouche virus. *J Immunol Res.* 2018;2018:6718083. doi:10.1155/2018/ 6718083

- 147. Kaifu T, Nakamura A. Polymorphisms of immunoglobulin receptors and the effects on clinical outcome in cancer immunotherapy and other immune diseases: a general review. *Int Immunol*. 2017;29 (7):319–325. doi:10.1093/intimm/dxx041
- 148. Fischinger S, Boudreau CM, Butler AL, Streeck H, Alter G. Sex differences in vaccine-induced humoral immunity. Semin Immunopathol. 2019;41(2):239–249. doi:10.1007/s00281-018-0726-5
- Voigt EA, Ovsyannikova IG, Kennedy RB, et al. Sex differences in older adults' immune responses to seasonal influenza vaccination. Front Immunol. 2019;10:180. doi:10.3389/fimmu.2019.00180
- 150. Fink AL, Engle K, Ursin RL, Tang WY, Klein SL. Biological sex affects vaccine efficacy and protection against influenza in mice. *Proc Natl Acad Sci U S A*. 2018;115(49):12477–12482. doi:10.10 73/pnas.1805268115
- 151. Mitchell LA, Zhang T, Tingle AJ. Differential antibody responses to rubella virus infection in males and females. *J Infect Dis*. 1992;166:1258–1265. doi:10.1093/infdis/166.6.1258
- 152. Majumder PP. Genomics of immune response to typhoid and cholera vaccines. *Philos Trans R Soc.* 2015;370(1671):20140142. doi:10.1098/rstb.2014.0142
- Brodin P, Davis MM. Human immune system variation. *Nat Rev Immunol*. 2017;17(1):21–29. doi:10.1038/nri.2016.125
- 154. Cotugno N, Ruggiero A, Santilli V, et al. OMIC Technologies and vaccine development: from the identification of vulnerable individuals to the formulation of invulnerable vaccines. *J Immunol Res*. 2019;2019:8732191. doi:10.1155/2019/8732191
- 155. Gunawardena S, Karunaweera ND. Advances in genetics and genomics: use and limitations in achieving malaria elimination goals. Pathog Glob Health. 2015;109(3):123–141. doi:10.1179/204777 3215Y.0000000015
- 156. Oli AN, Agu RU, Oli UC, Nwoye CU, Ejiofor OS, Esimone CO. Safety evaluation in mice of the childhood immunization vaccines from two south-eastern states of Nigeria. *Asian Pac J Trop Biomed*. 2015;5(2):132–137. doi:10.1016/S2221-1691(15)30157-X
- 157. Krubiner CB, Faden RR, Karron RA, et al. PREVENT working group. Pregnant women & vaccines against emerging epidemic threats: ethics guidance for preparedness, research, and response. *Vaccine*. 2019;S0264-410X(19):30045. doi:10.1016/j.vaccine.2019. 01.011

- Swamy GK, Heine RP. Vaccinations for pregnant women. Obstet Gynecol. 2015;125(1):212–226. doi:10.1097/AOG.00000000000 00581
- 159. Furuta M, Sin J, Ng ESW, Wang K. Efficacy and safety of pertussis vaccination for pregnant women a systematic review of randomised controlled trials and observational studies. *BMC Pregnancy Childbirth*. 2017;17(1):390. doi:10.1186/s12884-017-1559-2
- 160. Wilcox CR, Jones CE. Beyond passive immunity: is there priming of the fetal immune system following vaccination in pregnancy and what are the potential clinical implications? *Front Immunol*. 2018;9:1548. doi:10.3389/fimmu.2018.01548
- 161. Peña F, Pimentel R, Khosla S, Mehta SD, Brito MO. Zika Virus epidemic in pregnant women, dominican republic, 2016–2017. Emerg Infect Dis. 2019;25(2):247–255. doi:10.3201/eid2502.18 1054
- 162. Muehlenbachs A, de la Rosa Vázquez O, Bausch DG, et al. Ebola virus disease in pregnancy: clinical, histopathologic, and immuno-histochemical findings. *J Infect Dis.* 2017;215(1):64–69. doi:10.10 93/infdis/jiw206
- 163. The Ethics Working Group on ZIKV Research & Pregnancy. Pregnant Women & the Zika Virus Vaccine Research Agenda: Ethics Guidance on Priorities, Inclusion, and Evidence Generation. Baltimore, MD; 2017.
- 164. Omer SB. Maternal Immunization. N Engl J Med. 2017;376 (13):1256–1267. doi:10.1056/NEJMra1509044
- 165. Vojtek I, Dieussaert I, Doherty TM, et al. Maternal immunization: where are we now and how to move forward? *Ann Med.* 2018;50 (3):193–208. doi:10.1080/07853890.2017.1421320
- 166. Abramson JS, Mason E. Strengthening maternal immunisation to improve the health of mothers and infants. *Lancet*. 2016;388 (10059):2562–2564. doi:10.1016/S0140-6736(16)30882-0
- 167. Bahrami AA, Payandeh Z, Khalili S, Zakeri A, Bandehpour M. Immunoinformatics: in Silico approaches and computational design of a multi-epitope, immunogenic protein. Int Rev Immunol. 2019;38(6):307–322. doi:10.1080/08830185.2019.1657426

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