

2017

# Implementation of objective PASC-derived taxon demarcation criteria for official classification of filoviruses

Gaya K. Amarasinghe

*Washington University School of Medicine in St. Louis*

et al

Follow this and additional works at: [https://digitalcommons.wustl.edu/open\\_access\\_pubs](https://digitalcommons.wustl.edu/open_access_pubs)

---

## Recommended Citation

Amarasinghe, Gaya K. and et al, "Implementation of objective PASC-derived taxon demarcation criteria for official classification of filoviruses." *Viruses*.9,5. 106. (2017).

[https://digitalcommons.wustl.edu/open\\_access\\_pubs/5841](https://digitalcommons.wustl.edu/open_access_pubs/5841)

This Open Access Publication is brought to you for free and open access by Digital Commons@Becker. It has been accepted for inclusion in Open Access Publications by an authorized administrator of Digital Commons@Becker. For more information, please contact [engeszer@wustl.edu](mailto:engeszer@wustl.edu).

Letter

# Implementation of Objective PASC-Derived Taxon Demarcation Criteria for Official Classification of Filoviruses

Yīmíng Bào <sup>1</sup>, Gaya K. Amarasinghe <sup>2,†</sup>, Christopher F. Basler <sup>3,†</sup>, Sina Bavari <sup>4,†</sup>, Alexander Bukreyev <sup>5,†</sup>, Kartik Chandran <sup>6,†</sup>, Olga Dolnik <sup>7,†</sup>, John M. Dye <sup>4,†</sup>, Hideki Ebihara <sup>8,†</sup>, Pierre Formenty <sup>9,†</sup>, Roger Hewson <sup>10,†</sup>, Gary P. Kobinger <sup>11,†</sup>, Eric M. Leroy <sup>12,†</sup>, Elke Mühlberger <sup>13,†</sup>, Sergey V. Netesov <sup>14,†</sup>, Jean L. Patterson <sup>15,†</sup>, Janusz T. Paweska <sup>16,†</sup>, Sophie J. Smither <sup>17,†</sup>, Ayato Takada <sup>18,†</sup>, Jonathan S. Towner <sup>19,†</sup>, Viktor E. Volchkov <sup>20,†</sup>, Victoria Wahl-Jensen <sup>21,†</sup> and Jens H. Kuhn <sup>22,\*,†</sup>

<sup>1</sup> BIG Data Center, Beijing Institute of Genomics, Chinese Academy of Sciences, Beijing 100101, China; baoym@big.ac.cn

<sup>2</sup> Department of Pathology and Immunology, Washington University School of Medicine, St. Louis, MO 63110, USA; GAmarasinghe@path.wustl.edu

<sup>3</sup> Center for Microbial Pathogenesis, Institute for Biomedical Sciences, Georgia State University, Atlanta, GA 30302-3965, USA; cbasler@gsu.edu

<sup>4</sup> United States Army Medical Research Institute of Infectious Diseases, Fort Detrick, Frederick, MD 21702-5011, USA; sina.bavari.civ@mail.mil (S.B.); john.m.dye1.civ@mail.mil (J.M.D.)

<sup>5</sup> Department of Pathology, The University of Texas Medical Branch, Galveston, TX 77555-0144, USA; alexander.bukreyev@utmb.edu

<sup>6</sup> Department of Microbiology and Immunology, Albert Einstein College of Medicine, Bronx, New York, NY 10461, USA; kartik.chandran@einstein.yu.edu

<sup>7</sup> Institute of Virology, Philipps University Marburg, 35032 Marburg, Germany; Dolnik@staff.uni-marburg.de

<sup>8</sup> Department of Molecular Medicine, Mayo Clinic, Rochester, MN 55905, USA; Ebihara.Hideki@mayo.edu

<sup>9</sup> World Health Organization, 1211 Geneva, Switzerland; formentyp@who.int

<sup>10</sup> Public Health England, Porton Down, Wiltshire, Salisbury SP4 0JG, UK; Roger.Hewson@phe.gov.uk

<sup>11</sup> Department of Microbiology, Immunology & Infectious Diseases, Université Laval, Quebec City, QC G1V 0A6, Canada; gary.kobinger@crchudequebec.ulaval.ca

<sup>12</sup> Centre International de Recherches Médicales de Franceville, Institut de Recherche pour le Développement, BP 769 Franceville, Gabon; eric.leroy@ird.fr

<sup>13</sup> Department of Microbiology and National Emerging Infectious Diseases Laboratories, Boston University School of Medicine, Boston, MA 02118, USA; muehlber@bu.edu

<sup>14</sup> Novosibirsk State University, Novosibirsk, Novosibirsk Oblast, Russia 630090; netesov.s@nsu.ru

<sup>15</sup> Department of Virology and Immunology, Texas Biomedical Research Institute, San Antonio, TX 78245-0549, USA; jpatters@TxBiomed.org

<sup>16</sup> Center for Emerging and Zoonotic Diseases, National Institute for Communicable Diseases of the National Health Laboratory Service, Sandringham-Johannesburg 2131, Gauteng, South Africa; januszp@nicd.ac.za

<sup>17</sup> Chemical, Biological and Radiological Division, Defence Science and Technology Laboratory, Porton Down, Salisbury, Wiltshire SP4 0JQ, UK; SJSMITHER@mail.dstl.gov.uk

<sup>18</sup> Division of Global Epidemiology, Hokkaido University Research Center for Zoonosis Control, Sapporo 001-0020, Japan; atakada@czc.hokudai.ac.jp

<sup>19</sup> Viral Special Pathogens Branch, Division of High-Consequence Pathogens Pathology, National Center for Emerging and Zoonotic Infectious Diseases, Centers for Disease Control and Prevention, Atlanta, GA 30329-4027, USA; jit8@cdc.gov

<sup>20</sup> Molecular Basis of Viral Pathogenicity, CIRI, INSERM U1111-CNRS UMR5308, Université de Lyon, Université Claude Bernard Lyon 1, Ecole Normale Supérieure de Lyon, Lyon 69007, France; viktor.volchkov@inserm.fr

<sup>21</sup> National Biodefense Analysis and Countermeasures Center, Fort Detrick, Frederick, MD 21702, USA; victoria.jensen@nbacc.dhs.gov

<sup>22</sup> Integrated Research Facility at Fort Detrick, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Frederick, MD 21702, USA

\* Correspondence: kuhnjens@mail.nih.gov; Tel.: +1-301-631-7245

† The members of the International Committee on Taxonomy of Viruses (ICTV) *Filoviridae* Study Group.

Academic Editor: Eric O. Freed

Received: 26 April 2017; Accepted: 2 May 2017; Published: 11 May 2017

**Abstract:** The mononegaviral family *Filoviridae* has eight members assigned to three genera and seven species. Until now, genus and species demarcation were based on arbitrarily chosen filovirus genome sequence divergence values ( $\approx 50\%$  for genera,  $\approx 30\%$  for species) and arbitrarily chosen phenotypic virus or virion characteristics. Here we report filovirus genome sequence-based taxon demarcation criteria using the publicly accessible PAirwise Sequencing Comparison (PASC) tool of the US National Center for Biotechnology Information (Bethesda, MD, USA). Comparison of all available filovirus genomes in GenBank using PASC revealed optimal genus demarcation at the 55–58% sequence diversity threshold range for genera and at the 23–36% sequence diversity threshold range for species. Because these thresholds do not change the current official filovirus classification, these values are now implemented as filovirus taxon demarcation criteria that may solely be used for filovirus classification in case additional data are absent. A near-complete, coding-complete, or complete filovirus genome sequence will now be required to allow official classification of any novel “filovirus.” Classification of filoviruses into existing taxa or determining the need for novel taxa is now straightforward and could even become automated using a presented algorithm/flowchart rooted in RefSeq (type) sequences.

**Keywords:** cuevavirus; Ebola; ebolavirus; *Filoviridae*; filovirus; marburgvirus; *Mononegavirales*; virus taxonomy; virus classification; ICTV

## 1. Introduction

The family *Filoviridae*, one of eight families in the order *Mononegavirales* [1], has eight members assigned to seven species included in three genera (Table 1) [2–4].

**Table 1.** Official filovirus taxonomy endorsed by the 2015–2017 International Committee on Taxonomy of Viruses (ICTV) *Filoviridae* Study Group and accepted by the ICTV.

Current Taxonomy and Nomenclature
Order <i>Mononegavirales</i>
Family <i>Filoviridae</i>
Genus <i>Marburgvirus</i>
Species <i>Marburg Marburgvirus</i>
Virus 1: Marburg virus (MARV)
Virus 2: Ravn virus (RAVV)
Genus <i>Ebolavirus</i>
Species <i>Bundibugyo ebolavirus</i>
Virus: Bundibugyo virus (BDBV)
Species <i>Reston ebolavirus</i>
Virus: Reston virus (RESTV)
Species <i>Sudan ebolavirus</i>
Virus: Sudan virus (SUDV)
Species <i>Tai Forest ebolavirus</i>
Virus: Tai Forest virus (TAFV)
Species <i>Zaire ebolavirus</i>
Virus: Ebola virus (EBOV)
Genus <i>Cuevavirus</i>
Species <i>Lloviu cuevavirus</i>
Virus: Lloviu virus (LLOV)

Traditionally, the eight currently recognized filoviruses have been classified using phenotypic characteristics of virions and/or partial filovirus genome sequences [5–7]. Sequence-based filovirus taxon demarcation criteria (nucleotide and amino acid sequence identity values and/or phylogenies) were officially introduced as additional demarcation criteria in 2000 [8] and further refined thereafter [9]. Yet, true filovirus genome sequence-based taxon demarcation was only introduced in 2011. At that time, the International Committee on Taxonomy of Viruses (ICTV) *Filoviridae* Study Group decided arbitrarily that marburgvirus genomes differ from ebolavirus genomes by  $\geq 50\%$  and that ebolavirus species are differentiated on the basis of glycoprotein (*GP*) gene sequence differences ( $\geq 30\%$ ) or genome sequence differences ( $\geq 30\%$ ) [3]. These values were used to develop a decision algorithm/flowchart for filovirus taxon assignment that could guide filovirus classification [10]. In 2012, two pairwise sequence comparison methods, PAirwise Sequence Comparison (PASC) and DivErsity pArTitioning by hieRarchical Clustering (DEmARC), confirmed that the then official filovirus taxonomy (identical to the current one shown in Table 1) is justified, but that the 50% and 30% values ought to be adjusted objectively based on the PASC and/or DEmARC results [11,12]. Both analyses were based on the available  $\approx 50$  near-complete, coding-complete or complete filovirus genomes (see [13,14] for nomenclature) in the US National Center for Biotechnology Information (NCBI, Bethesda, MD, USA) GenBank database. Yet, at the time it was unclear whether the ICTV would accept classification of viruses based on sequence analysis alone.

In 2017, the ICTV members reached a consensus together with other experts that “the development of a robust framework for sequence-based virus taxonomy is indispensable for the comprehensive characterization of the global virome” [15]. Under proper oversight by, for instance, ICTV Study Groups, virus classification criteria can now be based on measurable objective criteria inferable only from viral genome sequence data. Thus, using automatic classification algorithms is possible.

The number of GenBank-deposited near-complete, coding-complete, and complete filovirus genome sequences has increased substantially in recent years (from the  $\approx 50$  in 2012 to  $\approx 1400$  at the time of writing in 2017). We analyzed these sequences using PASC, a method that can be easily used by any scientist using an open-access software platform [16–18]. We created inferred objective filovirus taxon demarcation criteria and updated the algorithm/flowchart for filovirus taxon assignment using the recently decided type filovirus sequences (NCBI RefSeq database sequences) [10] as starting points.

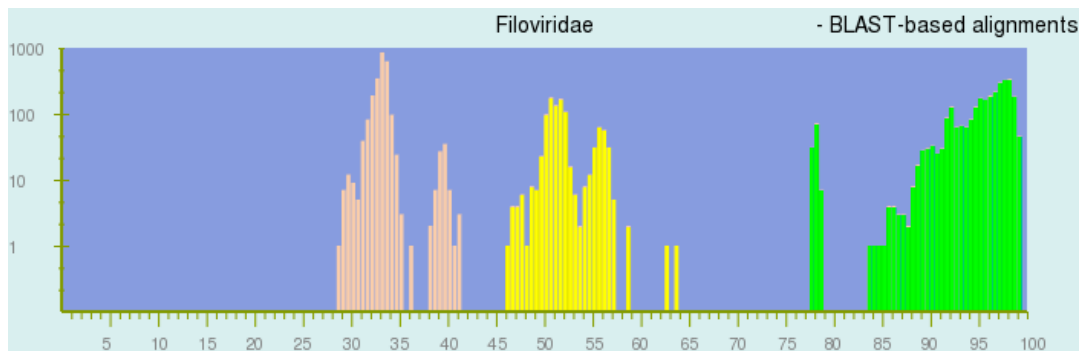
## 2. Materials and Methods

All 1404 near-complete, coding-complete, or complete filovirus genomes available from GenBank (NCBI, Bethesda, MD, USA) on 04/16/2017 were downloaded from the NCBI viral genomes resource [19]. Redundant filovirus genome sequences (here defined as sequences with PASC identities  $>99.5\%$ ) were removed, leaving 112 filovirus genome sequences for further analysis [20]. PASC analysis was performed with those 112 genome sequences as previously described [18] using the open-access PASC tool (NCBI). The new taxon demarcation algorithm/flowchart was developed based on the previously developed chart presented in [10] using type filoviruses [4] and type filovirus genome sequences (RefSeq, NCBI) [10].

## 3. Results

PASC analysis of 112 filovirus near-complete, coding-complete, or complete genome sequences revealed clear clustering into three higher ranks (genera), with two of those genera including single species and one genus including five species (visualized in Figure 1).

Unblinding of input sequences revealed the three genera and seven species to correspond to those already established and depicted in Table 1, raising confidence in PASC as a method to adequately recreate current knowledge on filovirus diversity. However, the analysis indicated an ideal genus demarcation threshold range of 55–58% sequence divergence rather than the currently used 50% threshold and an ideal species demarcation threshold range of 23–36% rather than the currently used 30% threshold.

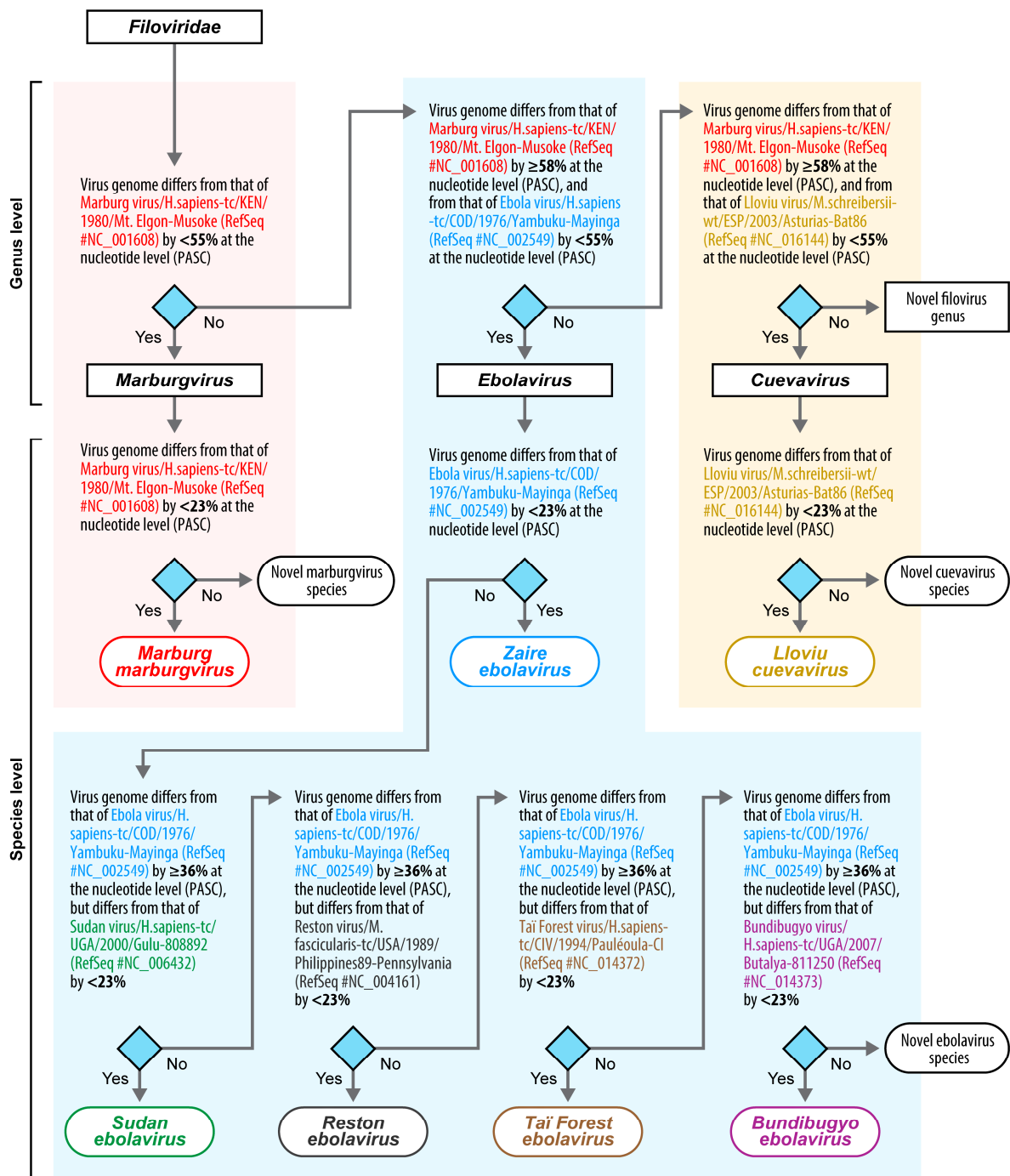


**Figure 1.** Screenshot of the US National Center for Biotechnology Information (NCBI) PAIRwise Sequence Comparison (PASC) tool result after comparing 112 distinct near-complete, coding-complete or complete filovirus genome sequences. Brown bars represent genome pairs assigned to (three) different genera; yellow bars represent genome pairs assigned to (seven) separate species; and green bars represent genome pairs assigned to the same species. BLAST: Basic Local Alignment Search Tool.

#### 4. Discussion

Using the new filovirus taxon demarcation criteria established here using PASC, the earliest discovered filovirus (Marburg virus; MARV) as the type virus for the family *Filoviridae* [4], the RefSeq MARV genome sequence as the MARV type sequence, and the remaining filovirus RefSeq genome sequences as additional anchor points, we created a filovirus classification decision matrix in form of an algorithm/flowchart (Figure 2). Using the NCBI PASC tool and Figure 2, any user can now quickly assess whether a novel filovirus sequence of interest represents a filovirus already classified in one of the established filovirus taxa or whether establishment of a new taxon/new taxa may be necessary. PASC requires at least near-complete or coding-complete genome input sequences. Therefore, the ICTV *Filoviridae* Study Group decided that moving forward, at least a coding-complete filovirus genome sequence will be minimally required for filovirus classification into novel filovirus taxa. Partial filovirus-like nucleic acids, for instance, those recently discovered in Chinese bats [21,22], may point towards the existence of novel filoviruses but will not suffice for official recognition of novel filoviruses or establishment of novel filovirus taxa. The Study Group recommends that such sequences be referred to as “filovirus-like sequences” and not as “filoviruses.” Likewise, a virus for which a partial filovirus-like sequence information exists ought to be referred to as a “putative filovirus” until at least coding-complete genome sequence information is available.

Importantly, PASC analysis followed by use of the algorithm/flowchart (Figure 2) alone does not constitute official classification, and the Study Group sees PASC results as highly informative, but not binding. Thus, if the PASC algorithm/flowchart indicates the need for a novel filovirus genus and/or species to a user analyzing a particular sequence, the user should follow the official pathway for ICTV classification starting with submission of an official taxonomic proposal (TaxoProp [23]). The user is recommended to engage with the ICTV *Filoviridae* Study Group as early as possible during that process. The Study Group and ICTV will evaluate all available data on a particular putative filovirus (e.g., host information, disease phenotype, biophysical properties of virions) and make their decisions accordingly. Phylogenetic results obtained with methods more sophisticated than PASC are always desired and may ultimately overrule PASC results.



**Figure 2.** Algorithm/flow chart for filovirus classification based on genomics sequence information (modified from [10]) and PASC-derived sequence demarcation criteria. A putative filovirus genome of interest is compared to the type filovirus RefSeq genome sequence (i.e., that of Marburg virus/H.sapiens-tc/KEN/1980/Mt. Elgon-Musoke [10]) and then sequentially moved through the process until its proper placement in a species is revealed. If the sequence comparison reveals the need for the creation of a novel genus and/or species, official taxonomic proposals ought to be submitted to the ICTV.



**Acknowledgments:** We thank Laura Bollinger and Jiro Wada (U.S. National Institutes of Health and National Institute of Allergy and Infectious Diseases (NIH/NIAID) Integrated Research Facility at Fort Detrick, Frederick, MD, USA) for critically editing the manuscript and figure creation, respectively. The views and conclusions contained in this document are those of the authors and should not be interpreted as necessarily representing the official policies, either expressed or implied, of the US Department of the Army, the U.S. Department of Defense, the U.S. Department of Health and Human Services, the Department of Homeland Security (DHS) Science and Technology Directorate (S&T), or of the institutions and companies affiliated with the authors. In no event shall any of these entities have any responsibility or liability for any use, misuse, inability to use, or reliance upon the information contained herein. The U.S. departments do not endorse any products or commercial services mentioned in this publication. This work was supported in part through Battelle Memorial Institute's prime contract with the U.S. NIAID under Contract No. HHSN2722007000161. A subcontractor to Battelle Memorial Institute who performed this work is: J.H.K., an employee of Tunnell Government Services, Inc. This work was also supported in part by the 100 Talent Program of the Chinese Academy of Sciences (Y.B.). This work was also funded in part under Contract No. HSHQDC-15-C-00064 awarded by DHS S&T for the management and operation of the National Biodefense Analysis and Countermeasures Center (NBACC), a Federally Funded Research and Development Center (V.W.-J.).

**Author Contributions:** Y.B. and J.H.K. conceived and designed the experiments; Y.B. performed the experiments; all authors analyzed the data; J.H.K. wrote the paper.

**Conflicts of Interest:** The authors declare no conflict of interest. The founding sponsors had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, and in the decision to publish the results.

## References

1. Amarasinghe, G.K.; Bào, Y.; Basler, C.F.; Bavari, S.; Beer, M.; Bejerman, N.; Blasdel, K.R.; Bochnowski, A.; Briese, T.; Bukreyev, A.; et al. Taxonomy of the order *Mononegavirales*: Update 2017. *Arch. Virol.* **2017**, *162*. [[CrossRef](#)] [[PubMed](#)]
2. Bukreyev, A.A.; Chandran, K.; Dolnik, O.; Dye, J.M.; Ebihara, H.; Leroy, E.M.; Mühlberger, E.; Netesov, S.V.; Patterson, J.L.; Paweska, J.T.; et al. Discussions and decisions of the 2012–2014 International Committee on Taxonomy of Viruses (ICTV) *Filoviridae* Study Group, January 2012–June 2013. *Arch. Virol.* **2014**, *159*, 821–830. [[CrossRef](#)] [[PubMed](#)]
3. Kuhn, J.H.; Becker, S.; Ebihara, H.; Geisbert, T.W.; Jahrling, P.B.; Kawaoka, Y.; Netesov, S.V.; Nichol, S.T.; Peters, C.J.; Volchkov, V.E.; et al. Family *Filoviridae*. In *Virus Taxonomy—Ninth Report of the International Committee on Taxonomy of Viruses*; King, A.M.Q., Adams, M.J., Carstens, E.B., Lefkowitz, E.J., Eds.; Elsevier/Academic Press: London, UK, 2011; pp. 665–671.
4. Kuhn, J.H.; Becker, S.; Ebihara, H.; Geisbert, T.W.; Johnson, K.M.; Kawaoka, Y.; Lipkin, W.I.; Negredo, A.I.; Netesov, S.V.; Nichol, S.T.; et al. Proposal for a revised taxonomy of the family *Filoviridae*: Classification, names of taxa and viruses, and virus abbreviations. *Arch. Virol.* **2010**, *155*, 2083–2103. [[CrossRef](#)] [[PubMed](#)]
5. Kiley, M.P.; Bowen, E.T.W.; Eddy, G.A.; Isaäcson, M.; Johnson, K.M.; McCormick, J.B.; Murphy, F.A.; Pattyn, S.R.; Peters, D.; Prozesky, O.W.; et al. *Filoviridae*: A taxonomic home for Marburg and Ebola viruses? *Intervirology* **1982**, *18*, 24–32. [[CrossRef](#)] [[PubMed](#)]
6. Francki, R.I.B.; Fauquet, C.M.; Knudson, D.L.; Brown, F. *Classification and Nomenclature of Viruses—Fifth Report of the International Committee on Taxonomy of Viruses*; Springer-Verlag: Vienna, Austria, 1991; Volume 2.
7. Jahrling, P.B.; Kiley, M.P.; Klenk, H.-D.; Peters, C.J.; Sanchez, A.; Swanepoel, R. Family *Filoviridae*. In *Virus Taxonomy—Sixth Report of the International Committee on Taxonomy of Viruses*; Murphy, F.A., Fauquet, C.M., Bishop, D.H.L., Ghabrial, S.A., Jarvis, A.W., Martelli, G.P., Mayo, M.A., Summers, M.D., Eds.; Springer-Verlag: Vienna, Austria, 1995; Volume 10, pp. 289–292.
8. Netesov, S.V.; Feldmann, H.; Jahrling, P.B.; Klenk, H.-D.; Sanchez, A. Family *Filoviridae*. In *Virus Taxonomy—Seventh Report of the International Committee on Taxonomy of Viruses*; van Regenmortel, M.H.V., Fauquet, C.M., Bishop, D.H.L., Carstens, E.B., Estes, M.K., Lemon, S.M., Maniloff, J., Mayo, M.A., McGeoch, D.J., Pringle, C.R., Eds.; Academic Press: San Diego, CA, USA, 2000; pp. 539–548.
9. Feldmann, H.; Geisbert, T.W.; Jahrling, P.B.; Klenk, H.-D.; Netesov, S.V.; Peters, C.J.; Sanchez, A.; Swanepoel, R.; Volchkov, V.E. Family *Filoviridae*. In *Virus Taxonomy—Eighth Report of the International Committee on Taxonomy of Viruses*; Fauquet, C.M., Mayo, M.A., Maniloff, J., Desselberger, U., Ball, L.A., Eds.; Elsevier/Academic Press: San Diego, CA, USA, 2005; pp. 645–653.

10. Kuhn, J.H.; Andersen, K.G.; Bào, Y.; Bavari, S.; Becker, S.; Bennett, R.S.; Bergman, N.H.; Blinkova, O.; Bradfute, S.; Brister, J.R.; et al. Filovirus RefSeq entries: Evaluation and selection of filovirus type variants, type sequences, and names. *Viruses* **2014**, *6*, 3663–3682. [[CrossRef](#)] [[PubMed](#)]
11. Bao, Y.; Chetvernin, V.; Tatusova, T. PAirwise Sequence Comparison (PASC) and its application in the classification of filoviruses. *Viruses* **2012**, *4*, 1318–1327. [[CrossRef](#)] [[PubMed](#)]
12. Lauber, C.; Gorbalenya, A.E. Genetics-based classification of filoviruses calls for expanded sampling of genomic sequences. *Viruses* **2012**, *4*, 1425–1437. [[CrossRef](#)] [[PubMed](#)]
13. Ladner, J.T.; Beitzel, B.; Chain, P.S.G.; Davenport, M.G.; Donaldson, E.F.; Frieman, M.; Kugelman, J.R.; Kuhn, J.H.; O’Rear, J.; Sabeti, P.C.; et al. Standards for sequencing viral genomes in the era of high-throughput sequencing. *MBio* **2014**, *5*, e01360-14. [[CrossRef](#)] [[PubMed](#)]
14. Ladner, J.T.; Kuhn, J.H.; Palacios, G. Standard finishing categories for high-throughput sequencing of viral genomes. *Rev. Sci. Tech.* **2016**, *35*, 43–52. [[CrossRef](#)] [[PubMed](#)]
15. Simmonds, P.; Adams, M.J.; Benkő, M.; Breitbart, M.; Brister, J.R.; Carstens, E.B.; Davison, A.J.; Delwart, E.; Gorbalenya, A.E.; Harrach, B.Z.; et al. Consensus statement: Virus taxonomy in the age of metagenomics. *Nat. Rev. Microbiol.* **2017**, *15*, 161–168. [[CrossRef](#)] [[PubMed](#)]
16. Bao, Y.; Chetvernin, V.; Tatusova, T. Improvements to pairwise sequence comparison (PASC): A genome-based web tool for virus classification. *Arch. Virol.* **2014**, *159*, 3293–3304. [[CrossRef](#)] [[PubMed](#)]
17. Bao, Y.; Kapustin, Y.; Tatusova, T. Virus classification by PAirwise Sequence Comparison (PASC). In *Encyclopedia of Virology*, 3rd ed.; Mahy, B.W.J., van Regenmortel, M.H.V., Eds.; Elsevier: Oxford, UK, 2008; Volume 5, pp. 342–348.
18. Bào, Y.; Kuhn, J.H. Preliminary classification of novel hemorrhagic fever-causing viruses using sequence-based PAirwise Sequence Comparison (PASC) analysis. In *Hemorrhagic Fever Viruses: Methods and Protocols*; Salvato, M.S., Ed.; Humana Press: Totowa, NJ, USA, 2017; in press.
19. Brister, J.R.; Ako-Adjei, D.; Bao, Y.; Blinkova, O. NCBI viral genomes resource. *Nucleic Acids Res.* **2015**, *43*, D571–D577. [[CrossRef](#)] [[PubMed](#)]
20. National Center for Biotechnology Information. PASC—Filoviridae. List of Non-Redundant Sequences (Using BLAST-Based Alignments). 2017. Available online: <https://www.ncbi.nlm.nih.gov/sutils/pasc/viridty.cgi?textpage=main&action=gilist&id=333> (accessed on 9 May 2017).
21. He, B.; Feng, Y.; Zhang, H.; Xu, L.; Yang, W.; Zhang, Y.; Li, X.; Tu, C. Filovirus RNA in fruit bats, China. *Emerg. Infect. Dis.* **2015**, *21*, 1675–1677. [[CrossRef](#)] [[PubMed](#)]
22. Yang, X.-L.; Zhang, Y.-Z.; Jiang, R.-D.; Guo, H.; Zhang, W.; Li, B.; Wang, N.; Wang, L.; Waruhiu, C.; Zhou, J.-H.; et al. Genetically diverse filoviruses in *Rousettus* and *Eonycteris* spp. bats, China, 2009 and 2015. *Emerg. Infect. Dis.* **2017**, *23*, 482–486. [[CrossRef](#)] [[PubMed](#)]
23. International Committee on Taxonomy of Viruses. *Taxonomy Proposal Templates*. Available online: <https://talk.ictvonline.org/files/taxonomy-proposal-templates/> (accessed on 9 May 2017).

