

## ERRATUM

In the article "Improved Methods for the Isolation and Study of the C<sub>18</sub>, C<sub>20</sub> and C<sub>22</sub> Monoethylenic Fatty Acid Isomers of Biological Samples: Hg Adducts, HPLC, AgNO<sub>3</sub>-TLC/FID, and Ozonolysis" by J-L. Sebedio, T.E. Farquharson and R.G. Ackman (*Lipids* 17:469-475, 1982), a section was inadvertently omitted from the text. On p. 474, the second column, after line 19, the following text should be inserted:

"... mixture of *cis* 18:1Δ9 and *trans* 18:1Δ9 was submitted to an HPLC analysis using a solvent mixture of MeOH/H<sub>2</sub>O (90:10) at 0.7 ml/min. No difference was observed between the actual isomer weight percentage and the chart area percentage given by the HPLC analysis, indicating no difference in the detector response factor for the *cis* and *trans* 18:1Δ9 isomers. The quantitation of *trans* fatty acids by the 2 other methods (AgNO<sub>3</sub>-TLC/GLC on SILAR-

7CP; AgNO<sub>3</sub>-Iatroscan) was not influenced by the chain lengths. These two methods can therefore be applied to any of the common chain lengths (C<sub>16</sub>-C<sub>22</sub>). An advantage of the AgNO<sub>3</sub>-Iatroscan method over GLC methods is the small sample size (10 μg) and the short time needed for an analysis (Fig. 3A) (24). An advantage of the HPLC quantitation using a refractive index detector is the possibility of recovering the sample after analysis. The main disadvantage with HPLC is the lack of sensitivity of the refractive index detector.

The approach to the study of monoethylenic fatty acids, based on the HPLC fractionation of the monoenoic fraction recovered from the methoxy-bromomercuri-adducts, is especially useful for biological samples and/or also for partially hydrogenated oils containing significant amounts of interfering conjugated diethylenic fatty acids. Moreover, although this method will be more time-consuming than the method using preparative GLC as a technique . . ."