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In vivo rescue of alveolar macrophages from SP-A knockout mice with exogenous SP-A nearly restores a wild type intracellular proteome; actin involvement

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Abstract

Background: Mice lacking surfactant protein-A (SP-A^{-/-}; knockout; KO) exhibit increased vulnerability to infection and injury. Although many bronchoalveolar lavage (BAL) protein differences between KO and wild-type (WT) are rapidly reversed in KO after infection, their clinical course is still compromised. We studied the impact of SP-A on the alveolar macrophage (AM) proteome under basal conditions. Male SP-A KO mice were SP-A-treated (5 micrograms/mouse) and sacrificed in 6 or 18 hr. The AM proteomes of KO, SP-A-treated KO, and WT mice were studied by 2D-DIGE coupled with MALDI-ToF/ToF and AM actin distribution was examined by phalloidon staining.

Results: We observed: a) significant differences from KO in WT or exogenous SP-A-treated in 45 of 76 identified proteins (both increases and decreases). These included actin-related/cytoskeletal proteins (involved in motility, phagocytosis, endocytosis), proteins of intracellular signaling, cell differentiation/regulation, regulation of inflammation, protease/chaperone function, and proteins related to Nrf2-mediated oxidative stress response pathway; b) SP-A-induced changes causing the AM proteome of the KO to resemble that of WT; and c) that SP-A treatment altered cell size and F-actin distribution.

Conclusions: These differences are likely to enhance AM function. The observations show for the first time that acute *in vivo* SP-A treatment of KO mice, under basal or unstimulated conditions, affects the expression of multiple AM proteins, alters F-actin distribution, and can restore much of the WT phenotype. We postulate that the SP-A-mediated expression profile of the AM places it in a state of "readiness" to successfully conduct its innate immune functions and ensure lung health.

Introduction

SP-A, a multi-functional protein, is known to play an important role in host defense. SP-A is a collectin, or collagenous lectin, that can recognize pathogen-associated molecular patterns (PAMP). The recognition and binding of PAMP is complex and may involve binding sites in addition to the C-type carbohydrate recognition domain. Although the direct interaction with pathogens constitutes one aspect of its host defense function, SP-A also plays a role in the clearance of particulate matter,

allergens, and debris from the alveolar surface [1-5]. SP-A appears to have a regulatory role on the alveolar macrophage by influencing the expression of a number of cytokines, including TNF- α , IL-1 β , and others [6-16], and cell surface molecules, such as CD11b (CR3), TLR2 and TLR4, the mannose receptor, scavenger receptor A, and CD14 [17-21]. Moreover, SP-A can help regulate redox balance [22-26], enhance bacterial phagocytosis by alveolar macrophages [27-30], contribute to bacterial killing [31-33], affect the development of dendritic cells [34], and provide an interface between innate and adaptive immunity [35]. Despite this diverse array of functions, many gaps remain in our knowledge of how SP-A influences lung host defense and the cell types it affects, especially under basal or unstimulated conditions.

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SP-A^{-/-} (knockout; KO) mice exhibit increased vulnerability to infection and injury. This has been illustrated with mouse models of pneumonia with organisms including *Klebsiella pneumoniae*, *Streptococcus pneumoniae*, *Pseudomonas aeruginosa*, *Pneumocystis carinii*, respiratory syncytial virus, and others [28,36-41]. Although the increased susceptibility was initially thought to be a consequence of the absence of the stimulatory effect of SP-A on phagocytosis, recent studies suggest a more complex picture. We have recently shown that in the absence of SP-A, baseline levels of many host defense molecules in bronchoalveolar lavage (BAL) samples [26,42] differ significantly (including both increases and decreases) from those in WT mice. However, although many of these differences in the SP-A KO mice are rapidly compensated for during infection and reach levels comparable to those of WT mice, the clinical course, and survival in particular [28] of the KO mice remains less favorable compared to that of the WT mice [27]. This may indicate that along with known direct effects of SP-A on phagocytosis and bacterial killing, there may be other direct and indirect effects of SP-A that may be instrumental in determining the clinical course and that these effects cannot occur in the absence of SP-A.

A likely source of these host defense deficits in the SP-A KO mouse is the alveolar macrophage, the primary effector cell for innate immunity in the lung. Although macrophages, which are derived from blood monocytes, are found throughout the body, their phenotype may vary depending on their environment. Alveolar macrophages exhibit a unique phenotype [43] that is clearly influenced by the presence of SP-A [17-21,27,28], but the extent of this influence is not entirely known. Moreover, virtually nothing is known about the *in vivo* effect of SP-A on the alveolar macrophage proteome under basal or unstimulated conditions, with regards to whether and which groups of proteins SP-A may affect under such conditions. However, based on recent *in vitro* studies, in response to LPS [44] or in extrapulmonary tissues [45], it appears that the integrity of the cytoskeleton is required in order for SP-A-mediated processes to occur.

In this study we tested the hypothesis that SP-A exerts a significant impact on the pattern of protein expression by the alveolar macrophage under basal or unstimulated conditions to make the alveolar macrophage ready and competent to subsequently mount a successful innate host defense response in response to various injurious agents. Towards this end we employed two-dimensional difference gel electrophoresis (2D-DIGE) to study *in vivo* the changes in the macrophage proteome in response to SP-A treatment of SP-A KO mice, the functional groups the changed proteins belong to, and

whether the SP-A-induced changes render the proteome from alveolar macrophages of the KO mice more like that of wild type (WT) mice. SP-A KO mice treated with SP-A and sacrificed at different times post-treatment, as well as WT mice on the same C57BL/6 genetic background were used in this study. The data were subjected to various analytical methods to assess, in an unbiased fashion, the similarities and differences in the proteomes of our experimental groups, as well as, by considering the function of the changed proteins, to obtain insight into underlying mechanisms and the SP-A-mediated functions of the alveolar macrophage. Based on the findings a “proof of principle” experiment was carried out where changes were measured in the actin cytoskeleton.

Results

BAL cells

The cells from all BAL samples were subjected to total and differential cell counts to exclude any mice with underlying infectious or inflammatory processes. No evidence of inflammation or infection was seen in any of the mice. The total cell counts did not differ significantly from one another and all BAL samples consisted of > 95% macrophages (data not shown).

Protein content was the same in all samples (data not shown) and identical amounts of protein (20 µg) were loaded on all gels.

2-D DIGE results

Overview

Following automatic spot detection and manual editing, 791 protein spots were defined and matched across 2D-DIGE gels from all samples and then subjected to further analysis. A reference gel is shown in Figure 1. Details of 2D-DIGE processing and data analysis are provided in Additional Files 1 and 2. When experimental groups were compared, ANOVA ($p < 0.05$) revealed significant differences in 234 spots, and these were subjected to additional analyses (Figures 2, 3, 4 and 5). The initial analysis was done using the individual spots data. All 791 spots were harvested from a preparative gel and we were able to identify 76 distinct proteins by MALDI-ToF/ToF with confidence intervals > 95% and ProteinPilot scores of > 61. The identified proteins were made up of 191 spots including “nearest neighbor” IDs and accounted for 24.1% of the detectable protein spots and 70.2% of the stained protein on 2D-DIGE gels. On the reference gel in Figure 1 the identified proteins with 191 spots circled are shown. The identified proteins were entered into the Ingenuity Pathways Analysis program to gain further insight into biological processes that might be affected (Table 1). Some of these identified proteins consisted of multiple spots and statistical

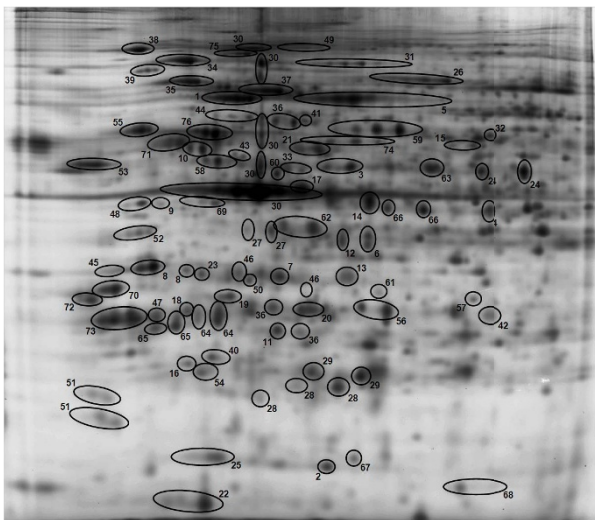


Figure 1 Reference gel image of mouse alveolar macrophage proteins identified by 2D-DIGE. Some of the proteins may include multiple spots reflecting different isoforms, fragments, or multimers (see also Additional File 3). 1, 65-kDa macrophage protein; 2, Actin related protein 2/3 complex, subunit 5; 3, Actin-related protein 3; 4, Actr2 protein; 5, Alpha-fetoprotein; 6, Annexin A2; 7, Annexin A4; 8, Anxa5 protein; 9, Arsa arsenite transporter, ATP-binding, homolog 1; 10, Atp5b protein; 11, Calpain, small subunit 1; 12, Capping protein (actin filament) muscle Z-line, alpha 2; 13, Capping protein (actin filament) muscle Z-line, beta isoform a; 14, Cathepsin D precursor; 15, Chaperonin subunit 2 (beta); 16, Chia protein; 17, Chitinase 3-like 3 precursor; 18, Chitinase-related protein MCRP; 19, Chloride intracellular channel 1; 20, Chloride intracellular channel 4; 21, CNDP dipeptidase 2; 22, Coactosin-like 1; 23, EF hand domain containing 2; 24, Eno1 protein (Alpha-enolase); 25, Eukaryotic translation initiation factor 5A; 26, Ezrin; 27, F-actin capping protein alpha-1 subunit; 28, Ferritin heavy chain 1; 29, Ferritin light chain 1; 30, Gamma-actin; 31, Gelsolin precursor; 32, Glucose-6-phosphate dehydrogenase X-linked; 33, Guanine deaminase; 34, Heat shock protein 1, beta; 35, Heat shock protein 5 precursor; 36, Heat shock protein 65; 37, Heat shock protein 8; 38, Heat shock protein 90, beta (Grp94), member 1; 39, Hematopoietic cell specific Lyn substrate 1; 40, Heme-binding protein; 41, Heterogeneous nuclear ribonucleoprotein K; 42, High mobility group 1 protein; 43, Hnrfp protein; 44, Kappa-B motif-binding phosphoprotein; 45, Keratin complex 2, basic, gene 8; 46, Keratin type II; 47, Krt13 protein; 48, Laminin receptor; 49, Major vault protein (MVP); 50, Microtubule-associated protein, RP/EB family, member 1; 51, Myosin light chain, regulatory B-like; 52, Nucleophosmin 1; 53, p50b; 54, Peroxiredoxin 2; 55, Prolyl 4-hydroxylase, beta polypeptide precursor; 56, Proteasome (prosome, macropain) 28 subunit, alpha; 57, Proteasome alpha 1 subunit; 58, Protein disulfide isomerase associated 6; 59, Protein disulfide-isomerase A3 precursor; 60, Protein synthesis initiation factor 4A; 61, Purine nucleoside phosphorylase; 62, Put. beta-actin (aa 27-375); 63, Rab GDP dissociation inhibitor beta; 64, Rho GDP dissociation inhibitor (GDI) alpha; 65, Rho, GDP dissociation inhibitor (GDI) beta; 66, Serine (or cysteine) proteinase inhibitor, clade B, member 1a; 67, Stathmin; 68, Superoxide dismutase 1, soluble; 69, Superoxide dismutase 1, soluble; 70, Tropomyosin 3, gamma; 71, Tubulin, beta 5; 72, Tyrosine 3/tryptophan 5-monooxygenase activation protein, epsilon polypeptide; 73, Tyrosine 3-monooxygenase/tryptophan 5-monooxygenase activation protein, beta polypeptide; 74, Vacuolar adenosine triphosphatase subunit B; 75, Valosin-containing protein; 76, Vimentin.

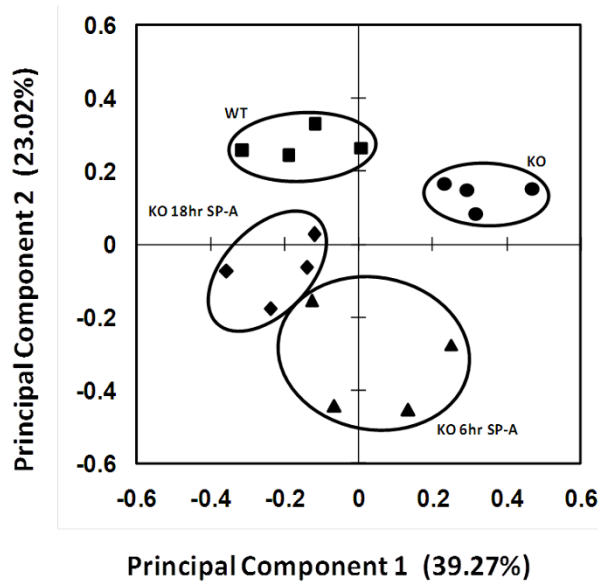
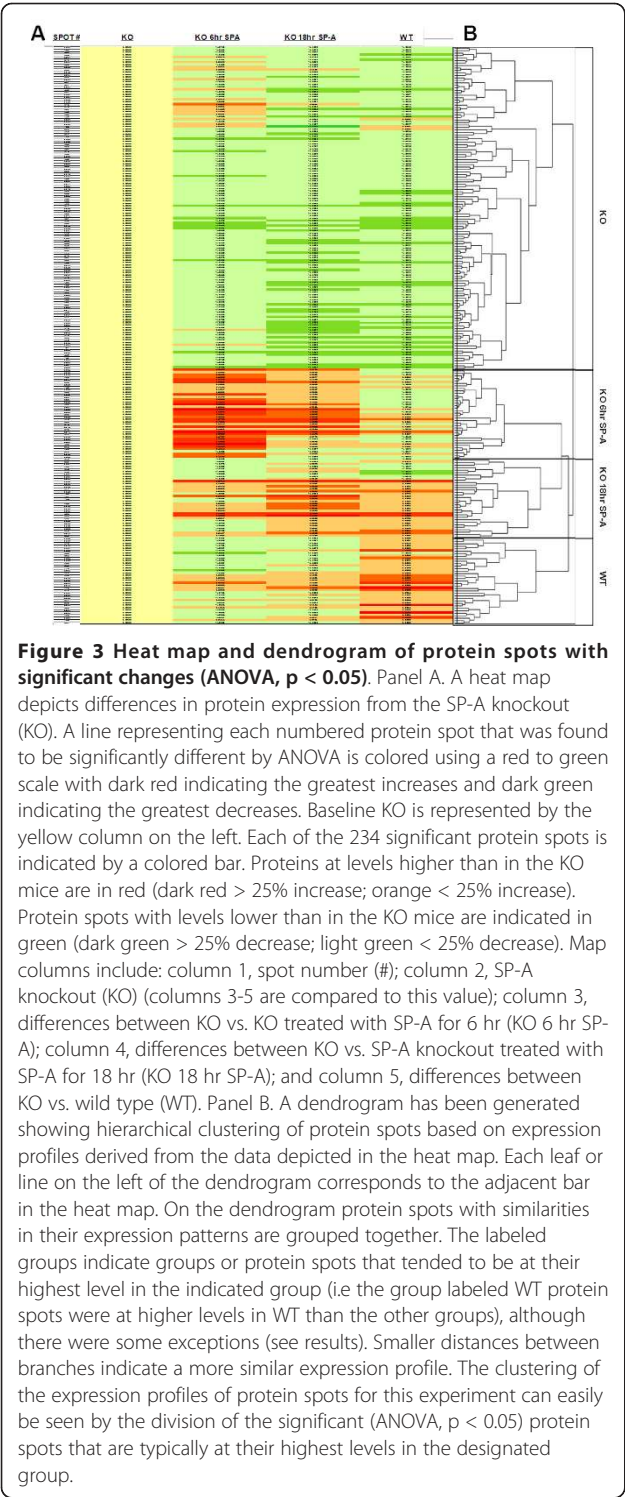


Figure 2 Principal component analysis. A plot of the principal component analysis for the 234 significant (ANOVA, $p < 0.05$) protein spots is shown. The markers represent the weighted average for the first two principal components for the 234 proteins for each individual in each of the groups: SP-A knockout (KO) (●), SP-A knockout treated with SP-A for 6 hr (KO 6 hr SP-A) (▲), SP-A knockout treated with SP-A for 18 hr (KO 18 hr SP-A) (◆), and wild type (WT) (■).

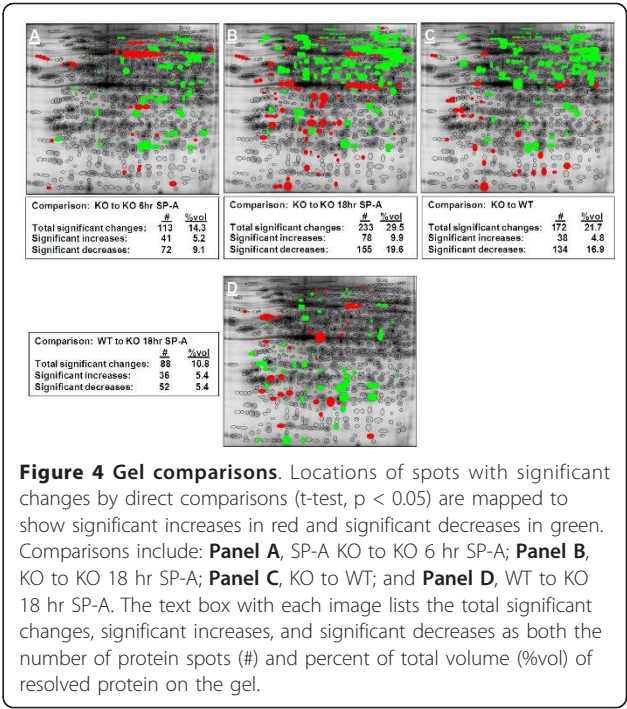
analysis was done on the combined normalized volume of all of the spots constituting a specific protein in each of our functional groups (Tables 2, 3, 4, 5, and 6). As mentioned above, some of the individual proteins are represented by a single spot and others may be one of multiple spots making up isoforms of a specific protein. Pairwise statistical comparisons between treatment groups were then done for both the individual spots and identified proteins and the results are summarized in Figure 5. Assignment of all identified proteins into subgroups is provided in Additional File 3 and comparisons for all identified proteins are listed in Additional File 4.

Principal Component Analysis

When principal component analysis (PCA) was performed on the significantly different spots, the markers for the four gels (each marker represents the gel from one individual animal within an experimental group) were grouped fairly tightly, and each of the experimental groups was well separated from one another (Figure 2). The first principal component accounted for 39.27% of the variance and the second accounted for 23.02%. In each group (KO mice, WT mice, and KO mice treated with SP-A for 18 hr), the four individual points are all rather tightly grouped indicating relatively small variance within each group. The KO mice treated with SP-A for 6 hr are not grouped as tightly indicating greater



variance among individual samples as these mice begin responding to SP-A treatment. The relative positions of each group are also informative. A clear separation between WT and KO mice is evident based on principal component 1. However, the KO mice treated with SP-A



for 6 hr are quite different from the KO baseline mice and the WT baseline mice. By contrast, at 18 hr after SP-A treatment the mice, based on principal component 1, are in closer proximity to the WT baseline mice than the KO baseline group indicating their overall greater

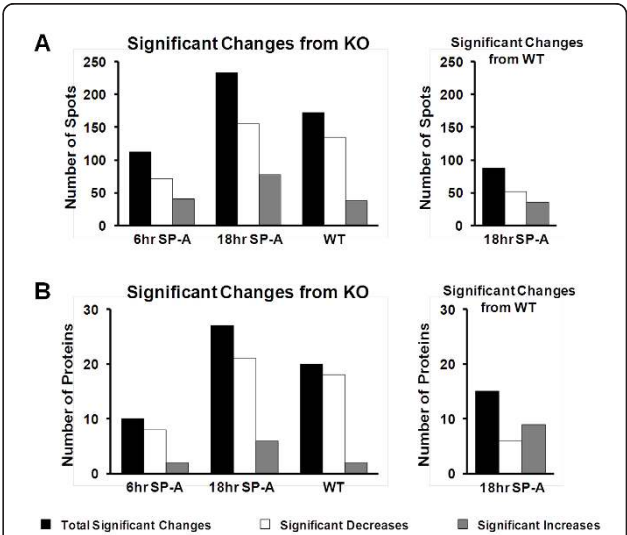


Table 1 p values for pathways identified by Ingenuity Pathways Analysis.

IPA Top Canonical Pathways	p-value
Regulation of Actin-based Motility by Rho	6.85E-08
RhoA Signaling	3.98E-07
Actin Cytoskeleton Signaling	3.35E-06
Fcy Receptor-mediated Phagocytosis in Macrophages and Monocytes	3.36E-06
Clathrin-mediated Endocytosis Signaling	6.11E-05
NRF2-mediated Oxidative Stress Response	1.09E-4

similarity to the WT animals following the SP-A treatment.

Heat diagram

A dynamic picture of the changes in expression is afforded by the heat diagram in Figure 3A. The relative expression levels depicted by the heat diagram provide the basis for defining the expression patterns that are summarized by the dendrogram (Figure 3B). In this diagram the KO baseline mice (1st column in yellow) provide the reference values to which the other groups are compared. Each of the 234 significant protein spots is indicated by a colored bar. Proteins at levels higher than in the KO mice are in red (dark red > 25% increase; orange < 25% increase). Protein spots with levels lower than in the KO mice are indicated in green (dark green > 25% decrease; light green < 25% decrease). It is readily apparent that both increases and decreases occur in all of the experimental groups compared to KO mice and, based on the preponderance of the green color in the diagram, most of the proteins are at higher levels in the KO baseline mice. However, a variety of expression patterns can be distinguished. For example, near the top of the diagram there are more than a dozen orange bars in the 6 hr group, indicating an increase over KO baseline levels. In the 18 hr and WT columns most of these have become green, indicating that the increases were transient and that most of the proteins have fallen to levels below those seen in KO baseline mice. In the bottom half of the diagram, there are a number of orange and dark red bars at 6 hr that remain red in the 18 hr SP-A group and in the WT group, indicating a persistent increase in the levels of these proteins after SP-A treatment of KO mice that resembles the WT levels. A number of other patterns are present representing increases and decreases and/or transient and sustained changes. The heat map shows a shift of changes with time after SP-A treatment towards the WT phenotype.

Cluster analysis

In Figure 3B the arrangement of the clusters produced by hierarchical clustering of the data depicted in the heat diagram (Figure 3A) is portrayed with a dendrogram in which the 234 significant spots are grouped

according to patterns of protein expression as levels of each spot are compared in the various experimental groups. With this arrangement the changes between groups can clearly be seen along the axis of the dendrogram, where protein spots are bracketed together and labeled with the group name where they are typically being expressed at the highest levels. For example, in the WT portion of the dendrogram, each leaf or each line (which represents one protein spot) on the axis is a protein that is typically (but not always) found at higher levels in the WT group compared to any of the other groups. The expression of more than half of the significant spots was highest in the untreated KO mice. The 6 hr SP-A, 18 hr SP-A, and WT groups all have approximately equal numbers of spots reaching their highest levels in each group. There were two major branches in the dendrogram. One represents the untreated KO mice and the other represents the two SP-A-treated KO groups and the WT mice. Within that second branch, the two SP-A-treated KO groups had a common sub-branch and were separated from the WT mice. The overall dendrogram analysis indicates that the KO and WT mice are on different branches and clearly separated from one another, whereas the treated mice share a common branch with the WT and appear to be closer to them than the KO mice. Although, as stated above, spots are typically at their highest abundance in the named category (KO 6 hr SP-A, WT, etc.), there are exceptions. An example of this is seen in the heat map at the top of the KO 6 hr SP-A column where there are a number of spots that are at their most abundant level, as indicated by the orange and red bars. It is also important to note that the KO division of the dendrogram has two major branches. At the top of the dendrogram, the smaller first branch appears to represent many protein spots that are at higher levels in the 6 hr SP-A-treated group, but then apparently (at 18 hr, green bars in heat map) drop to levels below those of the KO mice, suggesting a transient response at 6 hr (as discussed above). These spots could potentially be viewed as more related to the major 6 hr SP-A cluster. The much larger remainder of the KO branch contains multiple subdivisions, but all represent protein spots that are typically at lower levels in the SP-A-treated groups and the WT than in the KO mice.

Gel maps of protein changes

Figure 4 shows the reference gel with significant differences in protein spots between groups colored either green for decreases or red for increases. The significant differences are noted as the number of spots (out of a total of 791 spots) that changed significantly, and as the percentage of total volume of protein on the gel that are represented by the significant spots. Total volume is determined by adding the densitometric values of all of

Table 2 Changes in actin-related/cytoskeletal (ARC) proteins (as compared to KO baseline).

Gel No.	Protein Name	KO 6 hr SP-A	KO 18 hr SP-A	WT
1	65-kDa macrophage protein	↓	↓*	↓
2	Actin related protein 2/3 complex, subunit 5	↓	UN	↑
3	Actin-related protein 3	↓*	↓*	↓
4	Actr2 protein	↓	↓	↓
6	Annexin A2	↓*	↓*	↓*
11	Calpain, small subunit 1	↓	↓*	↓
12	Capping protein (actin filament) muscle Z-line, alpha 2 (CapZ alpha-2)	↓	↑	↑
13	Capping protein (actin filament) muscle Z-line, beta isoform a (CapZ beta)	↓*	↓	↓
15	Chaperonin subunit 2 (beta)	↓	↓*	↓*
19	Chloride intracellular channel 1	↓	↓*	↓*
20	Chloride intracellular channel 4 (mitochondrial)	↑	↑*	↑
22	Coactosin-like 1	↑	↑*	↑
24	Eno1 protein (Alpha-enolase)	↑	↓	↓
25	Eukaryotic translation initiation factor 5A	↑	↑	↑
26	Ezrin	↓	↓*	↓*
27	F-actin capping protein alpha-1 subunit (CapZ alpha-1)	↑	↑	↑
30	Gamma-actin	↓	↓*	↓*
31	Gelsolin precursor	↑	↓	↑
33	Guanine deaminase	↓	↓	↓
34	Heat shock protein 1, beta	↑	↓	↑
39	Hematopoietic cell specific Lyn substrate 1	UN	↑	↑
45	Keratin complex 2, basic, gene 8	↓	↑	↑*
46	Keratin type II	↓	↓*	↓
47	Krt13 protein	↑	↑*	↓
49	Major vault protein (MVP)	↓*	↓*	↓*
50	Microtubule-associated protein, RP/EB family, member 1	↓	↓*	↓
51	Myosin light chain, regulatory B-like	↑	↑	↑
53	p50b; Leukocyte-specific protein 1 (LSP1)	↓	↓	↓
62	Put. beta-actin (aa 27-375)	↑	↑*	↑
63	Rab GDP dissociation inhibitor beta	↓	↓*	↓*
64	Rho GDP dissociation inhibitor (GDI) alpha	↑	↓	↓
65	Rho, GDP dissociation inhibitor (GDI) beta	↑	↑	↑
67	Stathmin	↑	↑	↑*
69	Tropomodulin 3	↑	↑	↑
70	Tropomyosin 3, gamma	↓	↓	↑
71	Tubulin, beta 5	↑	↓	↑
75	Valosin-containing protein	↓	↓*	↓*
76	Vimentin	↑	↓	↓*
Total changes		21↓, 16↑, 1UN	24↓, 13↑, 1UN	21↓, 17↑
Total significant changes		4↓*	13↓*, 4↑*	9↓*, 2↑*

Comparison of mean normalized volumes (see Additional File 4) for proteins from KO mice to KO 6 hr SP-A, KO 18 hr SP-A, and wild-type mice. Increased (↑), decreased (↓), unchanged (UN), determined to be significant ($p < 0.05$) by t-test (*).

the expressed proteins resolved on the gel. Panels A through C of Figure 4 show changes as compared to the KO baseline control animals. In Panel D the protein spots from WT mice are compared to those of the KO mice 18 hr after SP-A treatment.

Comparison of the KO baseline mice to the KO 6 hr animals (Panel A) revealed 113 significant changes (41 increases and 72 decreases) accounting for 14.3% (5.2%

increases and 9.1% decreases) of the resolved protein. The largest number of significant changes is seen when KO baseline mice are compared to KO 18 hr. There are 233 significant changes with approximately 2/3 being proteins that are at higher levels in the KO mice (Panel B). The 233 protein spots make up 29.5% of the resolved protein on the gel. Of these, 78 are proteins expressed at increased levels and 155 are proteins expressed at

Table 3 Changes in regulation of inflammation (ROI) proteins (as compared to KO baseline).

Gel No.	Protein Name	KO 6 hr SP-A	KO 18 hr SP-A	WT
5	Alpha-fetoprotein	↑*	↑*	↑
6	Annexin A2	↓*	↓*	↓*
7	Alpha-fetoprotein	↓	↓	↓
16	Chia protein	↓	↑	↓
17	Chitinase 3-like 3 precursor (Ym1)	↑	↓*	↓*
18	Chitinase-related protein MCRP	↑	↑	↓
24	Eno1 protein (Alpha-enolase)	↑	↓	↓
25	Eukaryotic translation initiation factor 5A	↑	↑	↑
34	Heat shock protein 1, beta (HSP90AB1)	↑	↓	↓
35	Heat shock protein 5 precursor (GRP78)	↑	↓*	↓
36	Heat shock protein 65 (HSP60)	↓	↓*	↓*
37	Heat shock protein 8 (HSC70, HSC71)	↑	↑	↓
38	Heat shock protein 90, beta (Grp94), member 1	↓	↓*	↓*
39	Hematopoietic cell specific Lyn substrate 1	UN	↑	↑
40	Heme-binding protein	↓	↓	↓*
42	High mobility group 1 protein	↑	↓	↑
53	p50b; Leukocyte-specific protein 1 (LSP1)	↓	↓	↓
72	Tyrosine 3/tryptophan 5-monooxygenase activation protein, ε	↑	↓	↓
73	Tyrosine 3-monooxygenase/tryptophan 5-monooxygenase activation protein, β	↑	↓	↓
76	Vimentin	↑	↓	↓*
Total changes		7↓, 12↑, 1UN	14↓, 6↑	16↓, 4↑
Total significant changes		1↓*, 1↑*	5↓*, 1↑*	6↓*

Comparison of mean normalized volumes (see Additional File 4) for proteins from KO mice to KO 6 hr SP-A, KO 18 hr SP-A, and wild-type mice. Increased (↑), decreased (↓), unchanged (UN), determined to be significant ($p < 0.05$) by t-test (*).

Table 4 Changes in protease balance/chaperone function (PBCF) proteins (as compared to KO baseline).

Gel No.	Protein Name	KO 6 hr SP-A	KO 18 hr SP-A	WT
11	Calpain, small subunit 1	↓	↓*	↓
14	Cathepsin D precursor	↓	↓	↓
15	Chaperonin subunit 2 (beta)	↓	↓*	↓*
19	Chloride intracellular channel 1	↓	↓*	↓*
21	CNDP dipeptidase 2	↓*	↓*	↓
22	Coactosin-like 1	↑	↑*	↑
24	Eno1 protein (Alpha-enolase)	↑	↓	↓
34	Heat shock protein 1, beta (HSP90AB1)	↑	↓	↓
35	Heat shock protein 5 precursor (GRP78)	↑	↓*	↓
36	Heat shock protein 65 (HSP60)	↓	↓*	↓*
37	Heat shock protein 8 (HSC70, HSC71)	↑	↑	↓
38	Heat shock protein 90, beta (Grp94), member 1	↓	↓*	↓*
55	Prolyl 4-hydroxylase, beta polypeptide precursor	↑*	↑	↓*
56	Proteasome (prosome, macropain) 28 subunit, alpha	↓	↑	↑
57	Proteasome alpha 1 subunit	↓	↓	↓
58	Protein disulfide isomerase associated 6	↓	↓	↓
59	Protein disulfide-isomerase A3 precursor	↓	↓	↓
66	Serine (or cysteine) proteinase inhibitor, clade B, member 1a	↓	↑	↑
75	Valosin-containing protein	↓	↓*	↓*
Total changes		13↓, 6↑	14↓, 5↑	16↓, 3↑
Total significant changes		2↓*	8↓*, 1↑*	6↓*

Comparison of mean normalized volumes (see Additional File 4) for proteins from KO mice to KO 6 hr SP-A, KO 18 hr SP-A, and wild-type mice. Increased (↑), decreased (↓), unchanged (UN), determined to be significant ($p < 0.05$) by t-test (*).

Table 5 Changes in regulatory/differentiative processes (RDP) proteins (as compared to KO baseline).

Gel No.	Protein Name	KO 6 hr SP-A	KO 18 hr SP-A	WT
25	Eukaryotic translation initiation factor 5A	↑	↑	↑
41	Heterogeneous nuclear ribonucleoprotein K	↓*	↓*	↓*
42	High mobility group 1 protein	↑	↓	↑
43	Hnrpf protein	↓	↓	↓
44	Kappa-B motif-binding phosphoprotein	↓	↓	↓
52	Nucleophosmin 1	↓	↓	↓
53	p50b	↓	↓	↓
60	Protein synthesis initiation factor 4A	↑	↓	↓*
Total changes		5↓, 3↑	7↓, 1↑	6↓, 2↑
Total significant changes		1↓*	1↓*	2↓*

Comparison of mean normalized volumes (see Additional File 4) for proteins from KO mice to KO 6hr SP-A, KO 18hr SP-A, and wild-type mice. Increased (↑), decreased (↓), unchanged (UN), determined to be significant ($p < 0.05$) by t-test (*).

decreased levels in the KO 18 hr mice compared to the KO baseline mice. These proteins make up 9.9% and 19.6% of the resolved protein, respectively. When the KO to KO 18 hr gel and the KO to WT gel are compared, the pattern of changes is similar, suggesting that many of the changes in protein expression resulting from the exogenous SP-A treatment are restoring protein levels to those approximating the levels characterizing the WT phenotype. This trend can be further seen when the WT is compared to the KO 18 hr (Panel D).

The number of significant changes is markedly reduced, confirming the similarity of these two groups. In this comparison there are only 88 protein spots that differ significantly, making up 10.8% of total resolved protein. Panels A and D each depict a lower number of changes compared to Panels B and C, indicating that the protein expression profiles, in terms of the magnitude and number of changing spots, are closer to KO (after 6 hr of SP-A treatment) and WT (after 18 hr of SP-A treatment) profiles, respectively. This can be better

Table 6 Changes in Nrf2-regulated (NRF) proteins (as compared to KO baseline).

Gel No.	Protein Name	KO 6 hr SP-A	KO 18 hr SP-A	WT
1	65-kDa macrophage protein	↓	↓*	↓
14	Cathepsin D precursor	↓	↓	↓
20	Chloride intracellular channel 4 (mitochondrial)	↑	↑*	↑
28	Ferritin heavy chain 1	↓	↑*	↓
29	Ferritin light chain 1	↓*	↓	↓
30	Gamma-actin	↓	↓*	↓*
31	Gelsolin precursor	↑	↓	↑
32	Glucose-6-phosphate dehydrogenase X-linked	↓	↓*	↓*
34	Heat shock protein 1, beta (HSP90AB1)	↑	↓	↓
35	Heat shock protein 5 precursor (GRP78)	↑	↓*	↓
38	Heat shock protein 90, beta (Grp94), member 1	↓	↓*	↓*
45	Keratin complex 2, basic, gene 8	↓	↑	↑*
47	Krt13 protein	↑	↑*	↓
54	Peroxisome oxidin 2	↓	UN	↑
57	Proteasome alpha 1 subunit	↓	↓	↓
60	Protein synthesis initiation factor 4A	↑	↓	↓*
62	Put. beta-actin (aa 27-375)	↑	↑*	↑
68	Superoxide dismutase 1, soluble	↑	↑	↓
71	Tubulin, beta 5	↑	↓	↑
75	Valosin-containing protein	↓	↓*	↓*
76	Vimentin	↑	↓	↓*
Total changes		11↓, 10↑	14↓, 6↑, 1UN	15↓, 6↑
Total significant changes		1↓*	5↓*, 4↑*	6↓*, 1↑*

Comparison of mean normalized volumes (see Additional File 4) for proteins from KO mice to KO 6 hr SP-A, KO 18 hr SP-A, and wild-type mice. Increased (↑), decreased (↓), unchanged (UN), determined to be significant ($p < 0.05$) by t-test (*).

appreciated in Figure 5, Panel A, where the total number of spots changes as well as the numbers of spots increased and decreased in each experimental group compared to KO or WT are graphed. A similar conclusion is reached in Panel B of Figure 5 where the same comparisons are made using the 76 identified proteins rather than the 791 total protein spots.

In Additional File 4 the mean normalized volumes (\pm SD) are given for all 76 identified proteins and significant differences for comparisons for each experimental group between groups that are statistically significant are indicated.

Categorization of identified proteins

We used different approaches as done previously [26,42,46] with other types of samples (BAL proteins, plasma proteins) to assess the function of the set of 76 identified proteins and their biological relevance to macrophage function. Valuable insight was provided by analyzing the data with the Ingenuity Pathways Analysis (IPA) program. These analyses identified: 1) regulation of actin-based motility; 2) Rho A signaling; 3) Actin cytoskeletal signaling; 4) Fc γ receptor-mediated phagocytosis; 5) clathrin-mediated endocytosis signaling; and 6) the NRF2-mediated oxidative stress response (Table 1). However, as often occurs with IPA, PANTHER, and similar programs, available information is limited because many published functions/relationships of specific proteins have not yet been included and an organ/tissue-specific context is frequently lacking. Another frequent challenge is that functions are sometimes subdivided into so many subgroups that the overall significance is diminished. For these reasons, we also employed a manual curation approach, emphasizing findings from the literature related to the lung and macrophages, where available. This approach, as did IPA, strongly implicated motility, phagocytosis, actin signaling, and endocytosis (which we retained as a single functional group of "actin-related/cytoskeletal proteins" rather than dividing it into 4 subgroups) as the most involved cellular processes and included 37 of the 76 proteins identified in our study (Table 2). Given the role of the alveolar macrophage as a mobile phagocyte, this was anticipated and the identification of a substantial subset of identified proteins involved in these processes indicates that SP-A plays a pivotal role in these macrophage functions.

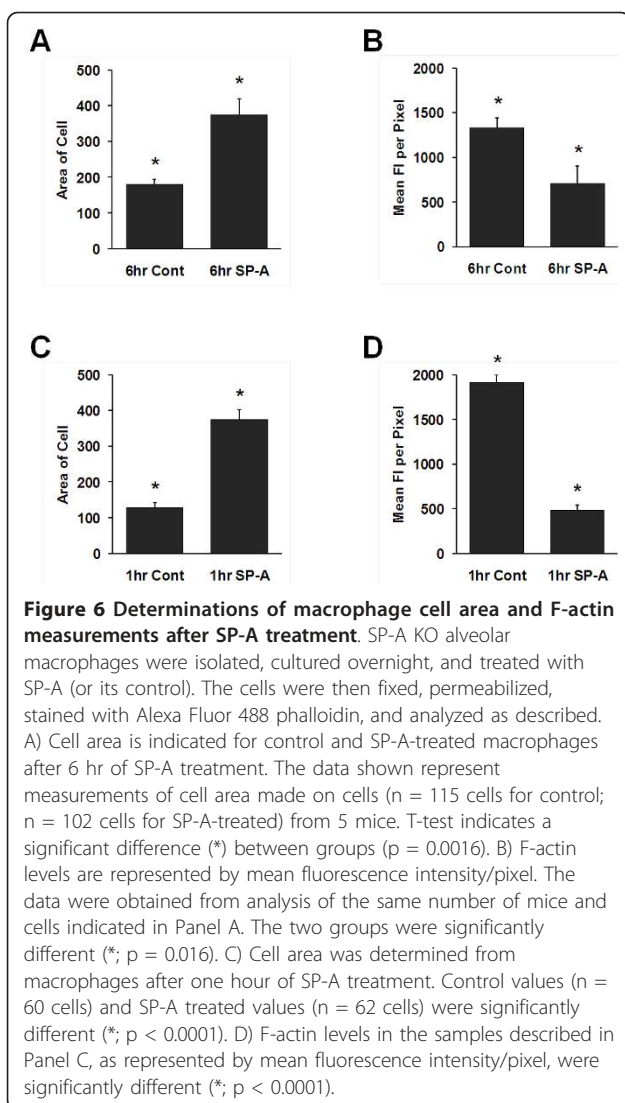
Other major processes implicated by our protein list included regulation of inflammation (19 proteins) (Table 3), protease balance/chaperone function (18 proteins) (Table 4), regulatory/differentiative processes (8 proteins) (Table 5), and Nrf2-regulated proteins (21 proteins) (Table 6). All proteins and the functional groups they were assigned to are listed in Additional File 3 [47-115]. These functional groups also represent

important facets of alveolar macrophage biology and thereby constitute a valuable tool in assessing macrophage function in the presence or absence of SP-A. Protease/chaperone function may be very important in the repair of damage to lung tissue and proteins potentially resulting from exposure to noxious material or pathogens and other danger signals. Similarly, the regulation of inflammatory processes is extremely important for innate immune processes, with dysregulation of inflammation playing a central role in many pulmonary disease processes. Finally, the profound differences between circulating blood monocytes and the alveolar macrophage [43,116,117] and between macrophages that have undergone different modes of activation [116,118] indicate the presence of an active regulatory mechanism directing the differentiation of macrophages from monocytes and their activation in various directions.

In addition to the functional categories described above, 21 of the identified proteins (~15-20) have been reported to be regulated by Nrf2. All of these have been reported to either contain antioxidant response elements (ARE) in their genes, or have been demonstrated to be Nrf2-regulated. We did not include proteins that may interact with Nrf2, but are not regulated by it. Nrf2, and its inhibitor, Keap1, are instrumental in regulating proteins that contain an ARE in their promoter regions and have been shown to regulate many antioxidant proteins, a number of cytoprotective enzymes (including Phase 2 detoxifying enzymes), as well as a number of heat shock proteins/chaperones. These proteins help maintain redox balance in the lung by limiting the effects of exogenous and endogenous reactive oxidant species. A number of the identified proteins fall into these categories. In some cases (ferritin light and heavy chain, glucose-6-phosphate dehydrogenase, peroxiredoxin, superoxide dismutase, valosin-containing protein, *et al.*) the presence of an ARE in their genes has been documented, confirming this potential regulatory mechanism. However, many other proteins that we identified have functions such as those listed above that are typically viewed as Nrf2-regulated. Of significant relevance, Nrf2/Keap1 activity can be regulated by interaction with actin filaments [119], adding another dimension to the biological relevance of the changes occurring in the many actin-related/cytoskeletal proteins we identified in the present study.

Cell area and F-actin measurements after SP-A treatment

Given the relatively large number of actin-related/cytoskeleton proteins that changed in response to SP-A, as "proof of principle" we measured cell size and levels of F-actin. In macrophages that were treated with SP-A for 6 hr and stained with Alexa Fluor 488 phalloidin we examined cell area (Figure 6A) and the mean



fluorescence intensity/pixel (Figure 6B) as a measure of the amount of F-actin/cell. Data were acquired from > 100 cells obtained from 5 mice. Analysis of cell size revealed that control macrophages from SP-A KO mice were significantly ($p = 0.0016$) smaller than macrophages from the same mice that were treated *in vitro* with SP-A for 6 hr. The mean fluorescence intensity/pixel in cells after SP-A treatment for 6 hr was significantly ($p = 0.016$) reduced to approximately 1/2 of that seen in the control cells.

Macrophages treated with SP-A for 1 hr and their controls (Figure 6C, D), the cell size and the mean fluorescence intensity/pixel exhibited similar changes to those seen at 6 hr. In fact, at the 1 hr time point (a time point used in several studies in which we studied phagocytosis as a biological readout [27,29]) cell size increased three-fold after SP-A treatment and the mean fluorescence intensity/pixel decreased almost four-fold. Both

differences were highly significant ($p < 0.0001$; $p < 0.0001$).

Discussion

The absence of SP-A in KO mice has been shown to increase susceptibility to infection, compromise bacterial clearance and survival from pneumonia, and adversely affect the response to various other types of lung injury [26,28,33,36-42,120]. However, virtually nothing is known about the impact of SP-A on the alveolar macrophage proteome under basal or unstimulated conditions. We employed a discovery proteomics approach to investigate, *in vivo*, the impact of SP-A on global protein expression by the alveolar macrophage as a way of gaining insight into molecules and mechanisms affected by SP-A under basal conditions. We thus studied and compared the intracellular proteome of the alveolar macrophage from WT mice and SP-A KO mice, and SP-A KO mice "rescued" with exogenous SP-A and sacrificed at 6 hr, a time point we have previously shown to be sufficient for the same dose of exogenous SP-A to restore tubular myelin formation in KO mice [121], and 18 hr after treatment. In the present study under basal conditions we observed: 1) differences in individual protein content between macrophages from KO and WT mice; 2) that a single *in vivo* treatment of KO mice with SP-A exerted a significant impact on the expression of proteins by alveolar macrophages compared to untreated KO mice; 3) as the time after SP-A treatment of KO mice increased, the proteomic profile more closely approximated that of WT mice; 4) a diverse set of SP-A-dependent functions in the alveolar macrophage, some of which have not been identified *in vivo* previously; including proteins involved in the regulation of the cytoskeleton; and 5) changes in F-actin and cell size of the alveolar macrophage in response to SP-A.

In previous studies of BAL proteome analysis, SP-A was found, under both resting conditions and after various perturbations (infection, ozone exposure), to have a major impact on BAL protein expression, particularly with respect to proteins involved in host defense or the regulation of redox balance [26,42]. Proteins secreted by the alveolar macrophage, and by other lung cells (particularly epithelial cells), together with proteins from plasma, are the principal sources of BAL protein. The observations made here indicate that the alveolar macrophage may also be a significant contributor to the observed BAL changes. Furthermore, the observed alveolar macrophage proteome changes in response to SP-A may contribute to the SP-A-dependent differences in the phagocytic function of these cells, which may in turn contribute to differences in disease susceptibility. In fact, it has been shown that both the phagocytic activity of the alveolar macrophages from SP-A KO

mice [28], and the survival of SP-A KO mice were compromised compared to WT mice following infection with *K. pneumoniae* in the presence or absence of ozone-induced oxidative stress [27,28]. The present findings, as assessed by the multitude of molecules and pathways affected by SP-A, support the notion that the alveolar macrophage from the KO mice is not biologically "ready" to successfully combat injurious agents, and hence the poorer clinical outcome shown in studies of the SP-A KO mice.

The SP-A dose and time points, as noted above, were based on a study where using the same conditions we were able to restore tubular myelin to the lungs of KO mice [121]. Moreover, based on measurements of SP-A levels in the BAL of lungs of C57BL/6 mice, the dose used was within the physiological range. A similar exogenous SP-A treatment has been reported [3,122], although a number of other studies have used much higher doses [20,123-125] that are clearly supraphysiological.

The "restoration" of the global pattern of protein synthesis in KO macrophages to that resembling WT macrophages, when exogenous SP-A was given, was remarkable. In 59 out of the 76 (78%) identified proteins the change seen, compared to KO at 18 hr after exogenous SP-A, was consistent with the differences between KO and WT. In other words, when WT levels were lower than KO, the 18 hr SP-A value was lower than KO and the opposite trend was apparent when WT levels were greater than KO. However, this transition from KO towards WT in response to SP-A treatment was complex, because levels of some proteins (~1/3) increased and others (~2/3) decreased in WT macrophages. This raised the possibility that with different proteins SP-A may influence protein synthesis, metabolism, and/or secretion by macrophages to achieve the intracellular levels appropriate for baseline levels in the macrophage.

Changes in the proteome of the treated versus untreated mice could be the result of several different regulatory mechanisms. SP-A may act directly on the alveolar macrophage, through different receptors to which it is known to bind [13,126-128], to directly influence the levels of expression of specific proteins. A second potential mechanism would involve indirect effects of SP-A on macrophage protein expression that result from the direct influence of SP-A on other cells (such as the type II cell), causing the synthesis and release of mediators that, in turn, regulate the production of specific proteins by the alveolar macrophages in a paracrine fashion. A third possibility is that SP-A exerts its influence on the alveolar macrophage proteome by regulating the release of regulatory molecules from one set of alveolar macrophages that act on themselves or another

set of alveolar macrophages via either autocrine or paracrine mechanisms, respectively. Regardless of whether one or more of these possibilities contribute to the proteome changes observed after SP-A treatment, what is clear is that SP-A, either via its direct or indirect effects, has a major impact on the protein expression pattern of the alveolar macrophage. This impact includes significant changes in more than half of the identified proteins.

An important indirect factor in terms of SP-A regulation of gene expression becomes apparent when we consider one of our previous studies of the BAL proteome of KO mice in which we postulated increased oxidative stress in the absence of SP-A [26]. In the current study, this would likely be manifested by changes in the expression of Nrf2-regulated proteins. Indeed, the Nrf2-mediated oxidative stress response pathway was identified as a significant pathway in the present study. Consistent with this notion are the observations made here where nearly 1/3 of the identified proteins are Nrf2-regulated, and are expressed at higher levels in KO than in WT mice. In the WT and with SP-A treatment, levels of expression of many of these proteins are significantly reduced, suggesting a lessening of oxidative stress. Glucose-6-phosphate dehydrogenase, a regulator of oxidative stress [129] stands out as an example of this. This enzyme is expressed at significantly higher levels in KO than in WT mice and the 18 hr SP-A treatment reduces it to levels significantly lower than KO baseline.

The most common phenotypic trait that has been examined with the SP-A KO mice is increased susceptibility to pneumonia. The simplest explanation for this phenotype is that host defense against specific pathogens is compromised in the absence of SP-A and its opsonic function. Indeed, replacement of SP-A in these models has afforded some restoration of normal host defense function by apparently increasing pathogen clearance [39,130]. Additional studies in which exogenous SP-A has been administered to SP-A KO mice have shown a diversity of functions or regulatory mechanisms being restored. These include SP-A effects on the regulation of allergic or inflammatory responses [3,120,122,131,132] and other host defense molecules [20,125], as well as the ability of exogenous SP-A to restore tubular myelin, an extracellular form of surfactant [121]. This diversity of effects, along with our BAL proteomic studies [26,42], indicate that SP-A exerts a pleiotropic effect in lung biology and/or health. The observations from the present study not only provide support for an SP-A pleiotropic effect on lung biology, but identify many protein molecules and pathways affected by the interaction of SP-A with the alveolar macrophage. Together these put forward the notion that the interaction of SP-A with the alveolar macrophage

under basal conditions is critical, in terms of preparing and maintaining the alveolar macrophage in a state of “readiness” so that it can potentially successfully combat injurious agents. Furthermore, based on the findings of the present study where SP-A exhibits a broad and varied impact on the alveolar macrophage proteome and previous studies of the BAL proteome where the baseline expression profile of SP-A KO and WT mice differed significantly [26,42], we postulate that these differences are key to underlying mechanisms that are responsible for the less favorable clinical outcome of the SP-A KO mice with pneumonia [28,39,40,131] or bleomycin-induced lung injury [120] where SP-A may be dysfunctional due to its being oxidized. Thus, the collective evidence of the role of SP-A in the first line defense of the alveolar surface depicts a diverse array of SP-A-mediated functions that help maintain lung health, or attenuate injury when the lung is challenged with various injurious agents, irritants, pathogens, and toxins present in the external environment. Given the constant exposure of the distal lung surface to all these external harmful agents, it is important that mechanisms be in place to remove these potentially damaging agents and maintain normal lung structure. The alveolar macrophage and SP-A, both important components of innate immunity, appear to play key roles in insuring that this occurs.

SP-A has previously been shown to exert regulatory effects on the production of several macrophage proteins [7,14,16-21,125]. The present study shows for the first time that an acute *in vivo* treatment with SP-A under unstimulated, baseline conditions affected the levels of significantly more and diverse macrophage proteins than previously known. These include proteins involved in macrophage motility, phagocytosis, neutralization of reactive oxidant species and other toxins, removal of damaged tissue/proteins from the alveolus, and other functions. These effects begin to be evident by as early as 6 hr after exogenous SP-A treatment. By 18 hr after intratracheal SP-A treatment, the alveolar macrophage proteome of the KO mouse becomes strikingly similar to that of WT animals that have been continually and naturally exposed to SP-A. The diversity of changed proteins broadly indicates that SP-A, under basal conditions, plays a major role in alveolar macrophage “health” in terms of its potential ability to move about and its “readiness” for pathogen elimination and tissue repair.

The responses identified exhibited several consistent patterns of protein expression. In one pattern the KO mice and WT mice differed significantly from one another (both increases and decreases) and intermediate responses proceeding in the same direction were observed at the two SP-A replacement time points. This

pattern was observed in 17 of the 20 proteins in which KO and WT differed significantly (see Additional File 4). The obvious conclusion from this subset of responses is that lack of continually available SP-A in the KO mice alters protein expression and that acute administration of exogenous SP-A leads toward a restoration of the WT phenotype. Of the total of 76 proteins identified in the present study, 37 were proteins with functions broadly related to actin and the cytoskeleton, and of these 37 proteins, 11 differed significantly between KO and WT and followed the pattern described above where SP-A treatment changes expression levels toward those seen in WT. Many other proteins in the actin-related/cytoskeletal group ($n = 21$ of 37) exhibited the same trend without achieving statistical significance. In most of the 32 responses made up by these two groups (significant, $n = 11$ + not significant but with a similar trend, $n = 21$), the time points after SP-A replacement showed a logical progression from KO to WT, indicating a trend toward restoration of the WT phenotype by exogenous SP-A treatment. The inclusion of so many of the significantly changing proteins ($n = 11$) in the actin/cytoskeleton-related protein group out of 20 significant changes between WT and KO indicates that SP-A has a profound influence on actin-related/cytoskeletal processes such as macrophage motility and phagocytosis, potentially explaining the susceptibility of the KO mice to injury or infection. *In vitro* studies have shown that an intact actin cytoskeleton, as assessed by the use of cytochalasin D, an actin depolymerizing agent, is required for certain SP-A-mediated processes [44] and in extrapulmonary tissues SP-A regulates F-actin filament organization [45]. Although given the importance of actin in cell motility, phagocytosis, and endocytosis, it is not surprising that changes in these proteins do occur, but the extent to which SP-A affects the content of so many actin/cytoskeletal-related proteins *in vivo* is remarkable. Significant changes in protein content with a similar pattern were also seen in several proteins involved in the inflammatory regulation ($n = 4$) and protease/chaperone ($n = 5$) functional groups.

The SP-A-dependent changes in actin-related/cytoskeletal proteins may also be the nexus for the differences in the other functional groups. As mentioned earlier, many of the changing proteins are either known to be regulated by Nrf2 or belong to functional classes known to be Nrf2-regulated [133]. Keap 1, the repressor of Nrf2, has been shown to regulate Nrf2 activity through its binding to the actin cytoskeleton [119]. Similarly, a role for the actin cytoskeleton has been described for NF- κ B [134], a mechanism via which SP-A exerts some of its functions [44,135,136]. These two transcriptional regulators in addition to each interacting with the actin

cytoskeleton, respond in concert to some stimuli in various cells or tissues and a number of genes regulated by both of them [137]. However, the interaction between NF- κ B and Nrf2 is complex and may vary in different systems or under different circumstances [138].

In addition, a number of proteins ($n = 13$) differed significantly between KO mice and mice that had received SP-A 18 hr earlier, even though no significant differences were observed between KO and WT mice. For those proteins however, in most cases the trend in the WT usually resembled that of the effect seen 18 hr after SP-A. This subset of responses supports a scenario where a robust acute response to SP-A is observed, with some of the proteins exhibiting an overcompensation of expression. For example, a number of protein spots (Figure 5) are increased at 6 hr but decreased at 18 hr after treatment, as in the WT, although they may not have not reached significance yet, indicating that some protein changes need a longer exposure to SP-A and/or multiple doses of SP-A to reach the WT phenotype. These responses included 6 proteins from the actin group and 3 from the protease/chaperone group, indicating that the acute SP-A treatment (like the continuous presence of SP-A in WT) has a significant potential to contribute to alterations in macrophage function.

Although statistical techniques such as gene set enrichment analysis or hypergeometric distribution calculations have been used with microarray data to determine whether changes in subsets of genes differ significantly from a reference group or the overall population in a specific group, these cannot be used here. This is because of the relatively small data set examined by gel-based proteomics and the comparatively large numbers of proteins in our subsets that make up a substantial percentage of our overall population or reference group. For example 50% of the identified proteins in our reference group were in our actin-related group and 28% were included in the Nrf2-related proteins, thereby violating the assumption that the analyzed groups be independent of one another and precluding the use of such methods in this study [139,140].

The large number of actin-related proteins with altered levels of expression after SP-A treatment led us to speculate that we would be able to validate the proteomic findings with a preliminary examination of parameters related to actin cytoskeletal dynamics. In fact, macrophages from SP-A KO mice treated with SP-A *in vitro* were significantly larger than comparable control cells and had lower amounts of F-actin, both at the 6 hr time point used for this proteomic study (although total gamma-actin levels were unchanged at this time point) and at 1 hr, a time point where we have previously demonstrated an SP-A-dependent enhancement of phagocytosis by alveolar macrophages [29]. As SP-A

stimulation increased cell size, redistribution of the cortical actin over a larger area likely contributes to the decrease in the actin intensity per pixel. We speculate that decreases in F-actin reflect cells that are more actively remodeling their cytoskeletons, resulting in cells of increased size. The larger size may indicate the availability of adequate amounts of excess membrane for phagocytic or endocytic events to occur and the changes in F-actin distribution likely reflect more dynamic actin filaments with altered rates of actin polymerization/depolymerization and branching. Thus, in the absence of SP-A, macrophages appear to be small and “stiff”, limiting their motility and phagocytic ability. However, when SP-A is made available, in as little as one hour a more dynamic actin cytoskeleton allows for enhanced motility and ability to successfully locate and clear particles and pathogens. Our previously published observations on *in vivo* comparisons of SP-A KO and WT mice in a model of pneumonia induced by *Klebsiella pneumoniae* [28] and on phagocytosis by alveolar macrophages *in vitro* support this scenario [29].

This study was designed to allow us to study restoration of the WT phenotype from KO alveolar macrophages after giving a single dose of SP-A and studying its effect 6 hr and 18 hr later. Remarkably, the dose and sampling times chosen restored levels of many of the proteins we studied. It is likely that additional doses and time points will further restore the phenotype by regulating the expression of additional proteins with different kinetics of translation and secretion. It is also highly likely that SP-A regulation by continually present SP-A (as in the WT state) or by multiple doses of SP-A will differ somewhat from the pattern resulting from a single acute dose, because in those cases a steady-state equilibrium of the SP-A effects on the alveolar macrophage proteome may be attained. Moreover, we had previously proposed that SP-A “primes” the alveolar macrophage [141] to prepare it to effectively respond to infectious or toxic agents. The proteomic data presented here support a role for SP-A under baseline conditions to drive an ill-prepared “dysfunctional” alveolar macrophage from an SP-A KO mouse towards readiness and effective host defense as afforded by WT alveolar macrophages. Moreover, these data demonstrate the potential for the therapeutic use of a single dose of exogenous SP-A to counter challenges due to injury or infection.

Conclusions

The observations in the present study show for the first time that an acute *in vivo* treatment with SP-A affects, under basal or unstimulated conditions, the expression of diverse groups of proteins in the alveolar macrophage. The group of actin-related/cytoskeletal proteins appeared to be a prominent one, indicating a significant

in vivo role of SP-A in alveolar macrophage motility and related functions. This was further substantiated by experiments showing significant changes in cell size and cortical F-actin distribution 1 and 6 hr after SP-A treatment. Furthermore, a significant number of the changed proteins are likely to be Nrf2-regulated indicating a role of SP-A in oxidative stress. We postulate that the SP-A-mediated expression profile of the alveolar macrophage places it in a state of “readiness” to successfully combat various injurious insults. Published comparison studies of SP-A KO and WT mice that showed a poorer outcome in KO in response to various injurious agents support the concept of “readiness.” Therefore, SP-A appears to be a critical factor for the “priming” and/or “readiness” of the alveolar macrophage to carry out its innate immune functions effectively and ensure lung health.

Materials and methods

Animals

This study was conducted using pathogen-free male WT and SP-A KO male mice on the C57BL/6 genetic background. WT mice were obtained from Jackson Laboratories (Bar Harbor, ME). Breeder pairs of SP-A KO mice had been obtained from Dr. Samuel Hawgood at the University of California, San Francisco and were propagated and raised under specific pathogen-free conditions in a barrier facility at the Penn State College of Medicine [142]. The SP-A KO mice and sentinel mice housed in the same room showed no evidence of respiratory pathogens. The Institutional Animal Care and Use Committee at the Penn State College of Medicine approved this study.

A total of 16, 25-34 g C57BL/6 WT and SP-A KO mice were used to complete the proteomics study. These were divided into four groups with 4 animals per group: 1) SP-A KO control (baseline) mice that did not receive any treatment; 2) SP-A KO mice that were treated with SP-A and sacrificed 6 hr after treatment; 3) SP-A KO mice that were treated with SP-A and sacrificed 18 hr after SP-A treatment; and 4) WT control (baseline) mice that did not receive any treatment.

SP-A preparation

SP-A was purified from the BAL fluid from normal human lungs obtained from organ donors. The protocol was approved by the Penn State College of Medicine Institutional Review Board. Donor lungs were lavaged with 0.9% saline and the lavage fluid collected and centrifuged at $150 \times g$ for 10 min at 4°C to obtain cell-free BAL. The SP-A was then purified by repeated precipitation with 5 mM calcium chloride. The purity of the preparation was checked by 1D-PAGE with silver stain and by Western blot and determined to be greater than 99 percent pure. LPS content of the SP-A

preparation was measured with the QCL-1000 Limulus Amebocyte Lysate assay (Lonza, Walkersville, MD). The LPS content of a 1 µg sample of SP-A was below the detectable limit of the assay (0.1 EU/ml) or < 500 fg/µg of SP-A.

Treatment of mice with SP-A

Animals were anesthetized with an intramuscular injection of a mixture of Ketamine HCl (Ketaject, Phoenix Pharmaceuticals Inc., St. Joseph, MO) and Xylazine (XYLA-JECT, Phoenix Pharmaceuticals Inc., St. Joseph, MO). Mice were suspended from their incisors and intrapharyngeally instilled with 5 µg of normal human SP-A in 50 µL of 0.9% sodium chloride containing 2 mM calcium chloride. Mice were closely watched to ensure that the entire dose was aspirated. We selected a dose of 5 µg of SP-A/mouse based on our SP-A determinations of total BAL SP-A from C57BL/6 mice (mean 3.2 µg; n = 9). We have previously demonstrated that using this dose of SP-A and a 6 hr time point, we were able to detect tubular myelin in the BAL of SP-A KO mice [121].

Collection of alveolar macrophages

Mice were anesthetized and subjected to BAL at intervals of 6 hr and 18 hr following treatment with SP-A. We chose 6 hr as our initial time point because we used it successfully in a previous study examining tubular myelin formation [121]. In addition, we postulated that this time interval would be sufficient for new protein synthesis to occur in response to SP-A treatment. The 18 hr time point was chosen to determine the longer term effects of a single dose of SP-A, that could potentially include the consequences of the indirect effects of SP-A. BAL fluid was obtained by instilling PBS, 1 mM EDTA into the lungs through a tracheal cannula using a volume equal to 80% of lung vital capacity ($5 \times$ with 0.5 ml of PBS, 1 mM EDTA) for a total of 2.5 ml. For each of the five washes, the fluid was instilled and withdrawn 3 times with chest massage during withdraw. The BAL fluid was then centrifuged at $150 \times g$ for 5 min at 4°C and the cell pellet washed with 1 mL of PBS, 1 mM EDTA. Total cell counts were performed using a hemocytometer and cytocentrifuge preparations done to obtain differential cell counts. The washed cells were then centrifuged again and the pellets frozen at -80°C for subsequent proteomic studies.

Preparation of samples for 2D-DIGE

Frozen macrophage pellets were lyophilized until complete dryness and resuspended in 25 µL of standard cell lysis buffer (30 mM TrisHCl, 2 M thiourea, 7 M urea, 4% CHAPS, pH 8.5). Protein determinations were done using the Bio-Rad Protein Assay (Bio-Rad, Hercules,

CA) and the concentration of protein was adjusted to 1 mg/ml for CyDye labeling.

2D-DIGE labeling (minimal labeling) and electrophoresis for 2D-DIGE

We have used this basic technique in previous studies for other types of protein samples [26,46,143] but a detailed account, including a number of modifications and refinements appears below. Information about the 2D-DIGE study is provided in a form that complies with the most recent version of Minimum Information About a Proteomics Experiment-Gel Electrophoresis (MIAPE-GE) standards currently under development (http://www.psdev.info/miape/MIAPE_GE_1_4.pdf) by the Human Proteome Organization Proteomics Standards Initiative (see Additional File 1). Two sets of samples from each group were prepared for minimal labeling with CyDyes. Samples from each group were randomly assigned to Cy3 or Cy5 to ensure no dye-based artifacts in quantitation. A 20 µg aliquot of macrophage proteins from each sample was labeled with either Cy3 or Cy5 (200 picomoles). A normalization pool was created by combining equal amounts of protein from every sample (16 samples) and an aliquot of the pool was labeled with Cy2 (200 picomoles/20 µg). Equal amounts (20 µg) of Cy3-labeled sample, Cy5-labeled sample, and Cy2-labeled pool samples were mixed and applied to each gel. An equal volume of 2X sample buffer (2 M thiourea, 7 M urea, 2% IPG buffer (pH 4-7) and 1.2% DeStreak reagent) was added to all samples to give a final volume of 140 µl. Twenty-four cm, pH 4-7 gradient Immobiline DryStrips (GE Healthcare) were rehydrated for 18 hr with 450 µl of rehydration buffer (DeStreak™ Rehydration Solution containing 0.5% IPG buffer (pH 4-7)). Proteins were subjected to isoelectric focusing on rehydrated strips using an Ettan IPGphor 3 cup loading manifold (GE Healthcare) following the manufacturer's instruction at 20°C and under mineral oil to prevent evaporation. Proteins were focused using the following voltages and times: 3 hr at 300 V (step and hold); 7 hr at 1000 V (gradient); 4 hr at 8000 V (gradient); 4 hr at 8000 V (step and hold). After isoelectric focusing the IEF strips were equilibrated in equilibration solution-1 (50 mM TrisHCl, 6 M urea, 30% glycerol, 2% sodium dodecyl sulphate (SDS), 0.5% dithiothreitol) and equilibration solution-2 (50 mM TrisHCl, 6 M urea, 30% glycerol, 2% SDS, 4.5% iodoacetamide) for 15 min, respectively, and then applied to 10% polyacrylamide gels (26 cm-w × 20 cm-h × 1 mm thick), sealed with 0.5% low melting point agarose containing bromophenol blue in a buffer of 1X Tris/glycine/SDS buffer (25 mM Tris, 192 mM glycine, 0.1% (W/V) SDS, pH 8.3). Proteins were then separated on the basis of molecular weight using the Ettan DALTtwelve system (GE

Healthcare) at 5 W/gel for 30 min and then at 15 W/gel at 20°C until the bromophenol blue dye front was 0.5 cm from the bottom of the gel (~4 hr).

For the preparative (picking) gel an aliquot of 450 µg of sample was diluted with an equal volume of 2X sample buffer (2 M thiourea, 7 M urea, 2% IPG buffer (pH 4-7) and 1.2% DeStreak reagent) and then brought up to a volume of 450 µl with rehydration buffer (DeStreak™ Rehydration Solution and 0.5% IPG buffer (pH 4-7)). Proteins were focused using the following voltages and times: 14 hr at 0 V (passive rehydration); 6 hr at 30 V (active rehydration); 3 hr at 300 V (step and hold); 3 hr at 600 V (gradient); 3 hr at 1000 V (gradient); 3 hr at 8000 V (gradient); 4 hr at 8000 V (step and hold). The strip was then equilibrated as described above and applied to a 10% polyacrylamide gel (26 cm-w × 20 cm-h × 1 mm thick). For the preparative picking gel a single plate for the gel plate sandwich was treated with Bind-Silane solution (80% ethanol, 0.02% glacial acetic acid, 0.001% Bind-Silane) and had reference marker stickers placed on it. After the completion of electrophoresis, the plate that had not been silane-treated was removed from the sandwich and the gel fixed overnight with 30% ethanol, 7.5% glacial acetic acid. The preparative picking gel was then stained with Deep Purple Total Protein Stain (GE, Healthcare) for 2 hr.

Gel imaging, image analysis, and statistics

Information about the acquisition and processing of data from the 2D-DIGE studies are provided in the form that complies with the most recent version of the guidelines established for Minimum Information about a Proteomics Experiment-Gel Informatics (MIAPE-GI) currently under development by the Human Proteome Organization Proteomics Standards Initiative <http://www.psdev.info/files/miape-gi-v1.pdf> (see Additional File 2). All two-dimensional gels were imaged on a Typhoon 9410 fluorescent imager (GE Healthcare) at a resolution of 100 µm. Photomultiplier tube voltages were individually set for each of the three colored lasers to ensure maximum, linear signals. The same voltages were used for all the gels. The DIGE gels were imaged at three different wavelengths (Cy2: 520 nm; Cy3: 580 nm; Cy5: 670 nm) and the Deep Purple Total Protein Stain-stained gels were imaged at 100 µm with a separate filter (610 nm).

Gel images were imported into the Progenesis SameSpots v4.0 program (Nonlinear Dynamics) for analysis and their quality assessed by the program. Image quality control with Progenesis SameSpots v4.0 involves checking images for bit depth, color, manipulation prior to analysis, proper file type, saturation, low dynamic range, and stretched contrast. A reference gel with minimum distortion and streaks was selected from the Cy2 gels. Gel alignment was conducted automatically and then

checked manually to ensure correct alignment. Spot detection and spot matching across all the gels were conducted automatically, and then spot matching was checked and manually edited to ensure correct matching. Data from all the spots included in analysis were transported to Progenesis PG240 module of the Progenesis SameSpots v4.0 software for further analysis. Statistical analysis was performed by ANOVA and by t-test to confirm the level of significance among various groups. Differences were considered statistically significant with $p < 0.05$. Principal components analysis (PCA) was performed and dendrograms plotted in the Statistics module of Progenesis SameSpots v4.0 for the further analysis of the patterns of protein expression and protein clustering among groups.

For identified proteins having multiple isoforms, the normalized volumes of all isoforms of a given protein were added together and statistical analysis was performed on the totals using Microsoft Excel.

Protein identification by mass spectrometry

We have used this procedure in previous studies for other types of protein samples [26,46,143] but a detailed account, including many modifications and refinements appears below. For identification of spots, all 791 protein spots from the experiment were picked from the picking gel using a robot-directed spot picker (Ettan Spot Picker, GE Healthcare). The picker head was calibrated using the reference stickers placed on the preparative picking gel and the gel was picked and gel plugs placed in bar-coded 96 well plates. All gel plugs were washed twice with 200 μ l of 200 mM ammonium bicarbonate, 40% acetonitrile for 30 min at 37°C, lyophilized to complete dryness and stored desiccated at 4°C until processing. Plugs selected for processing were rehydrated with 20 μ l of 0.02 μ g/ μ l trypsin (Trypsin, proteomics grade, Sigma, St. Louis, MO) for one hour at room temperature. The trypsin solution was then removed and replaced with 50 μ l of 40 mM ammonium bicarbonate, 9% acetonitrile and incubated overnight at 48°C. The overnight incubation solution was transferred to new 96-well extraction plates and the plugs extracted with 50 μ l of 0.1% trifluoroacetic acid (TFA) for 30 min at 37°C, followed by a second extraction with 50 μ l of dH_2O for 30 min at 37°C. All of the extracts for each plug were pooled and then dried by lyophilization. Dried extracts were then brought up in 200 μ l of dH_2O and dried by lyophilization for a series of three cycles to remove any remaining ammonium bicarbonate. The final dried extracted proteins/peptides were brought up in 20 μ l of 0.5% TFA and desalted and concentrated using C_{18} ZipTips (Millipore Corporation, Billerica, MA). Tips were wetted with 10 μ l of 100% acetonitrile and equilibrated with 10 μ l 0.1% TFA ($\text{pH} < 4$). Samples

were then drawn into ZipTip columns by aspirating for 10 cycles and then washed twice with 10 μ l 0.1% TFA. Peptides were then eluted from the column twice with 10 μ l of 50% acetonitrile, 0.1% TFA. The ZipTip procedure was performed four times for each extract to maximize peptide yield and the elutions were pooled, lyophilized and resuspended in 2 μ l of 50% Acetonitrile, 1% TFA for MALDI plate spotting.

Peptides were analyzed by MALDI-ToF/ToF mass spectrometry (5800 Proteomic Analyzer Applied Biosystems, Foster City, CA) in the Mass Spectrometry Core at the Penn State University College of Medicine. All of each ZipTip cleaned samples (1 μ l at a time) was applied onto a 384-well MALDI plate (Opti-TOF™ 384 Well Insert, Applied Biosystems) and then 0.7 μ l of 2 mg/ml ACH cinnamic acid in 60:40 (acetonitrile: water) was spotted on each well containing peptide. All 13-calibration wells on the MALDI plate were spotted with (1:12 diluted) 4700 calibrant. Autolytic trypsin peptides were also used to internally calibrate the spectra to an accuracy of 20 ppm. Using the GPS Explorer 3.0 software (Applied Biosystems), the MS and MS/MS data were submitted to the MASCOT search engine using the NCBI non-redundant database and mouse taxonomy for identification. The search parameters included: trypsin digestion with a maximum of three missed cleavages; fixed modifications, carbamidomethylation; variable modifications, carbamylation, acetylation, deamidation, oxidation; peptide mass tolerance, 0.15 Da. MASCOT confidence interval scores of $> 95\%$ combined with a ProteinPilot score of greater than 61 were considered as a positive protein identification. An image of the reference gel is shown in Figure 1 with all identified proteins circled and numbered. The PANTHER database and the scientific literature were used to provisionally assign molecular function and biological process to each identified protein. We then re-assigned the identified proteins to four broad functional classes including: a) actin-related/cytoskeletal proteins; b) proteins involved in protease balance/chaperone function; c) proteins involved in regulation of inflammation; and d) proteins involved in regulatory/differentiative processes. It should be noted that some proteins are in more than one functional group. This classification scheme was more inclusive than relying solely on the biological function classification provided by PANTHER and similar gene ontology databases. We also used the Ingenuity Pathway Analysis program (Ingenuity Systems, Redwood City, CA) to gain additional insight into the functional significance of the observed changes. Protein names, accession numbers, and the functional groups we assigned them to are listed in Additional File 3 together with a list of supporting references.

Collection, culture and *in vitro* SP-A treatment of mouse AMs for F-actin staining

AMs were collected from the BAL fluid of male KO mice as described earlier. AMs were washed 1X with AM medium (RPMI-Hepes modification containing 2 mM L-glutamine and 1:100 antibiotic antimycotic solution). AMs were then resuspended in AM medium at 1×10^5 /mL and 1 mL (10^5 AMs) plated to cover glasses (No. 1 circles, 18 mm, 0.15 mm thick) in 12-well tissue culture plates and allowed to adhere for 90 min at 37°C. The medium was then removed and replaced with 1 mL of fresh AM medium and the cells were cultured overnight at 37°C. AMs were treated with SP-A for either one or six hours by adding 5 μ L (5 μ g) of a 1 μ g/ μ L human SP-A stock solution in 10 mM Hepes (pH 7.5) directly to the 1 mL of medium from the overnight culture.

Alexa Fluor 488 phalloidin staining of F-Actin on AMs

After incubation with SP-A, AMs were washed 1X with PBS at 37°C, fixed with 3.7% paraformaldehyde in PBS for 10 min at 37°C and permeabilized with 0.5% Triton in PBS for 5 min. After permeabilization, cells were washed with PBS, incubated for 30 min in staining solution (PBS, 300 nM DAPI, 6.6 nM Alexa Fluor 488 phalloidin) and again washed with PBS 3X for 10 min. Cover glasses containing the AMs were mounted to slides using PBS containing 80% glycerol and 0.3% n-propyl gallate, and then sealed using nail polish.

Image acquisition and data analysis

AMs were visualized using a Nikon TE-2000 PFS fluorescent microscope with a 60X (1.4 NA) phase contrast lens with a 1.5 \times tube lens in place and the images captured using a Photometrics Coolsnap HQ2 digital camera (0.07 μ m/pixel) and saved as TIFF files. Exposure time (200 ms) was kept constant for all images acquired. Nikon NIS-Elements AR (version 3.0) software was used to do both the image acquisition and data analysis. AMs were analyzed by manually drawing a border for each cell as well as copying an area of equal size to a location immediately adjacent to each cell to be used for background subtraction. Data were then collected for area (cell size), mean fluorescence intensity per pixel of cell area, and the sum of the fluorescence intensity for all of the pixels within a cell. Statistical analysis was performed using Minitab and Excel.

Additional material

Additional file 1: MIAPE: Gel Electrophoresis. File containing Minimum Information About a Proteomics Experiment-Gel Electrophoresis in the format recommended by the Human Proteome Organization Proteomic Standards Initiative.

Additional file 2: MIAPE: Gel Informatics. File containing Minimum Information About a Proteomics Experiment-Gel Informatics in the format recommended by the Human Proteome Organization Proteomic Standards Initiative.

Additional file 3: Protein names and cross references to accession numbers and categories. File containing a table that has the gel numbers and names of all identified proteins. It contains both NCBI GI numbers and Swiss Prot Accession numbers for all proteins, as well as the designation for the functional group(s) to which each protein was assigned. Reference numbers listed in the table are from the reference list in the manuscript.

Additional file 4: Values for all identified alveolar macrophage proteins with note of significant changes. File containing a table that gives normalized volumes for all proteins for each individual group \pm SD and indicates comparisons between groups that were significantly different.

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Authors' contributions

DSP and JF designed the study. DSP interpreted data and prepared the manuscript. JF assisted with data interpretation, and participated in manuscript preparation. TMU treated mice, collected samples, ran gels, prepared mass spec samples, evaluated mass spec data, did preliminary analysis of proteomic data, and participated in the writing of the manuscript. OAQ prepared the cell samples for the microscopy experiments, collected images, and performed the image analyses. CMY participated in design of actin experiments and interpreted actin data. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

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