

Incidence and prognosis of *c-KIT* and *FLT3* mutations in core binding factor (CBF) acute myeloid leukaemias

RORY S. CARE,¹ PETER J. M. VALK,² ANNE C. GOODEVE,¹ FAISEL M. ABU-DUHIER,¹ WENDY M. C. GEERTSMA-KLEINEKOORT,² GIU A. WILSON,¹ MAMDOOH A. GARI,¹ IAN R. PEAKE,¹ BOB LÖWENBERG² AND JOHN T. REILLY¹ ¹Academic Unit of Haematology, Division of Genomic Medicine, Royal Hallamshire Hospital, Sheffield, UK, and ²Department of Haematology, Erasmus University Medical Centre, Rotterdam, the Netherlands

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Summary. DNA from 110 adult *de novo* acute myeloid leukaemia (AML) patients exhibiting either *inv(16)* ($n = 63$) or *t(8;21)* ($n = 47$) was screened for mutations in the *c-KIT* (exon 8 and Asp816) and *FLT3* (ITD and Asp835) genes. *c-KIT* exon 8 mutations were found in 15/63 (23.8%) *inv(16)* patients and 1/47 (2.1%) *t(8;21)* patients. *c-KIT* Asp816 mutations were present in 5/63 (7.9%) *inv(16)* AML and 5/47 (10.6%) *t(8;21)* AML. *FLT3* muta-

tions were identified in five patients (7.9%) with *inv(16)* and three patients (5.6%) with *t(8;21)* AML. All mutations were mutually exclusive; 40% of *inv(16)* AML patients possessed either a *c-KIT* or *FLT3* mutation. *c-KIT* exon 8 mutations were shown to be a significant factor adversely affecting relapse rate.

Keywords: *c-KIT*, *FLT3*, AML, CBF, *inv(16)*.

The core binding factor (CBF) leukaemias, *inv(16)(p13q22)* and *t(8;21)(q22;q22)*, express the fusion genes *CBFβ/MYH11* and *AML1/ETO* respectively. The mechanism by which *AML1* and *CBFβ* disruption induces leukaemogenesis is unclear, although fusion gene expression is not sufficient to cause leukaemia. For example, transgenic mice expressing *AML1/ETO* do not develop leukaemia, unless treated with the mutagenic agent N-ethyl-N-nitrosourea (ENU) (Yuan *et al.*, 2001). Similarly, mice transgenic for the chimaeric gene *CBFβ/MYH11* exhibit a myeloid maturation block but require additional mutations for the development of leukaemia (Castilla *et al.*, 1999). This suggests that *AML1/ETO* and *CBFβ/MYH11* may dictate the disease phenotype, but that additional 'hits' are required for leukaemic transformation.

Recently, Gilliland (2001) proposed that acute myeloid leukaemia (AML) results from two classes of mutation, a class I mutation that confers a proliferative signal [e.g. a RTK (receptor tyrosine kinase) or RAS mutation], and a class II mutation that impairs haematopoietic differentiation, such as the CBF fusion genes. RTK class III mutations have been linked to a number of haematological malignancies. For example, *FLT3* internal tandem duplication (ITD) and Asp 835 mutations occur in 30% of AML patients and

confer a poor prognosis, while *c-KIT* Asp816 mutations are associated with mast cell disease (see Reilly, 2002). In support of the 'two-hit' hypothesis, *FLT3* Asp835 and *c-KIT* Asp816 activating loop mutations have been reported in patients with CBF AML (Beghini *et al.*, 2000; Kottaridis *et al.*, 2001), while a strong association between *c-KIT* exon 8 mutations and *inv(16)* AML has been reported (Gari *et al.*, 1999).

We report that 40% of patients with AML and *inv(16)* possessed a class I mutation, supporting Gilliland's pathogenetic model, and that *c-KIT* exon 8 mutations are associated with an increased relapse rate in AML and *inv(16)*.

MATERIALS AND METHODS

Patient DNA samples and mutation detection. RNA and genomic DNA were obtained from the bone marrow or blood of 110 patients with CBF AML; 63 patients with *inv(16)(p13q22)* and 47 patients with *t(8;21)* (see Table 1A for clinical details). Patients were treated in the UK Medical Research Council (MRC) AML 10 and 12 trials ($n = 65$) and the Dutch–Belgian Haematology–Oncology Group HOVON Trials ($n = 45$). Median follow-up was 46 months. Amplification of the *FLT3* ITD and Asp835 mutations were performed as described by Abu-Duhier *et al.* (2000, 2001). *c-KIT* exon 8 mutations were analysed according to Gari *et al.* (1999) while *c-KIT* 816 mutations

Correspondence: Dr J. T. Reilly, Academic Unit of Haematology, M floor, Royal Hallamshire Hospital, Sheffield, S10 2JF, UK. E-mail: j.t.reilly@sheffield.ac.uk

Table 1.

(A) Clinical details of the 110 CBF AML patients studied.

	inv(16)	t(8;21)	Total
Patients (n)	63	47	110
Males (n)	36	13	68
Females (n)	27	16	42
Mean age (years)	43.9	39.6	42
Age range (years)	15–74	16–83	15–83

(B) Results of *c-KIT* and *FLT3* mutational analysis of the 110 CBF AML patients studied.

	inv(16)	t(8;21)	Total
Patients (n)	63	47	110
<i>c-KIT</i> 419	23.8% (15)	2.1% (1)	18.8% (16)
<i>c-KIT</i> 816	7.9% (5)	10.6% (5)	9.1% (10)
<i>FLT3</i> ITD	3.2% (2)	4.2% (2)	3.6% (4)
<i>FLT3</i> 3835	4.8% (3)	2.1% (1)	3.6% (4)
Total percentage	39.7% (25)	19.1% (9)	30.9% (34)

Data are given as the percentage (number of patients).

were analysed by the amplification of exon 17 using 1.1× Readymix PCR master mix containing 1.5 mmol/l MgCl₂ (ABgene, Surrey, UK) and 50 nm of each primer: E17F, 5'-TCCATCACCGGTACCTCCTA; and E17R, 5'-CACCACAGTGAGTGCAGTTG. cDNA, prepared from 50 ng of RNA using random hexamer priming, was also used for mutation detection in some patients.

Statistical analysis. Survival curves were determined by the Kaplan–Meier method (Kaplan & Meier, 1958) and compared as described by Mantel and Haenszel (1959). Overall survival was measured from diagnosis to death or last follow-up, with March 2002 as the final analysis date. Disease-free survival was measured from achievement of complete remission to date of first relapse, death or last follow-up.

RESULTS

Incidence of c-KIT and FLT3 mutations

Of 63 AML patients with inv(16), 23.8% possessed a *c-KIT* exon 8 mutation (Table 1B) that involved Asp419 in 93% of cases. Deletions varied from 6 to 13 bp and insertions from 1 to 15 bp. A single patient, who retained the Asp419 *c-KIT* residue, had only an insertion. Five patients (7.9%) possessed a *c-KIT* Asp816 mutation. *FLT3* ITD and Asp835 mutations were only found in 3.2% and 4.8% of patients respectively. Mutations were mutually exclusive, so that overall 39.7% of patients with AML and inv(16) possessed a RTK mutation. Nine mutations were detected in the 47 patients with t(8;21): *c-KIT* Asp816 (*n* = 5), *c-KIT* exon 8 (*n* = 1), *FLT3* ITD (*n* = 2) and *FLT3* Asp835 (*n* = 1) (Table 1B). Overall, 19.1% of patients with t(8;21) possessed a RTK mutation.

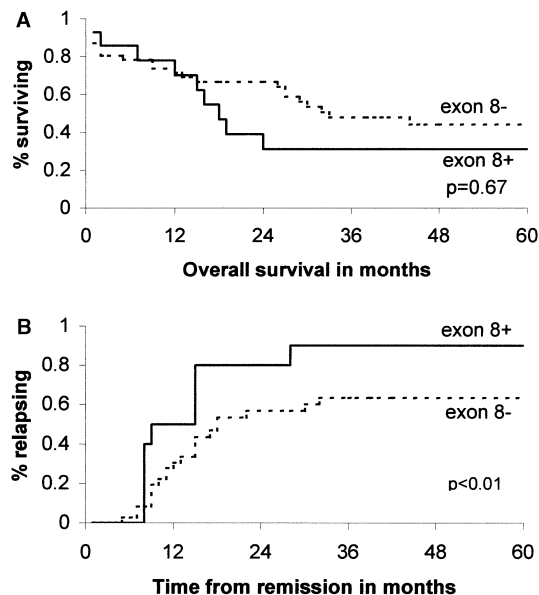


Fig 1. Kaplan–Meier plots showing overall survival (A) and relapse (B) of inv(16) patients with (exon8⁺) or without (exon8⁻) an insertion/deletion in *c-KIT* exon 8 involving Asp419.

Prognostic impact of c-KIT and FLT3 mutations

Overall survival of patients with AML and t(8;21) (*n* = 47) was 61% [95% confidence interval (CI) 51.8–70.1] at 5 years compared with 41% (95% CI 35.7–46.3) for patients with AML and inv(16) (*n* = 63). Kaplan–Meier plots revealed *c-KIT* exon 8 mutations to be a significant factor adversely affecting relapse rate (*P* < 0.01), but not overall survival (*P* = 0.67). (Fig 1A and B).

DISCUSSION

Recently, an intriguing association between *c-KIT* mutations and CBF AML has been documented. Beghini *et al* (1998), for example, reported an Asp816Tyr activating mutation in an AML-M2 with t(8;21), and later strengthened this link by documenting Asp816 mutations in 4/9 patients with t(8;21) and 2/6 patients with inv(16) (Beghini *et al*, 2000). In addition, Gari *et al* (1999) reported *c-KIT* exon 8 mutations with consistent loss of Asp419 in a third of patients with AML-M4Eo and inv(16).

The present study has extended these findings and indicated that 25% of AML and inv(16) patients exhibit a *c-KIT* exon 8 mutation. These data, together with the five additional patients exhibiting an Asp816 mutation, showed that a third of AML patients with inv(16) had a *c-KIT* mutation. *c-KIT* mutations were less common in patients with t(8;21) (12.7%) and were not found in over a hundred patients with CBF-negative AML (results not shown). Although AML patients with inv(16) or t(8;21) generally have a better outcome following chemotherapy, there is still a significant proportion of patients who will relapse following an initial complete remission. From the analysis presented here, it appears that inv(16) AML with *c-KIT*

exon 8 mutations are associated with a greater probability of relapse following complete remission. Thus, the molecular characterization of inv(16) AML, a heterogeneous category of AML in terms of prognosis, may allow the identification of a subset with higher risk disease. Interestingly, RTK mutations appeared to be mutually exclusive, so that 40% of inv(16) patients possessed either a *c-KIT* or *FLT3* mutation. We speculate that the remaining 60% of inv(16) patients should have at least one additional class I mutation.

In conclusion, current evidence suggests that AML results from the collaboration of at least two classes of mutation: class I mutations that confer a proliferative and/or survival advantage and class II mutations that impair differentiation. We believe that our finding of a high frequency of RTK mutations in CBF AML, especially patients expressing *CBFβ/MYH11*, supports this model.

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