

Incorporating epigenetic mechanisms to advance fetal programming theories

ELISABETH CONRADT, DANIEL E. ADKINS, SHEILA E. CROWELL, K. LEE RABY, LISA M. DIAMOND, AND
BRUCE ELLIS
University of Utah

Abstract

Decades of fetal programming research indicates that we may be able to map the origins of many physical, psychological, and medical variations and morbidities before the birth of the child. While great strides have been made in identifying associations between prenatal insults, such as undernutrition or psychosocial stress, and negative developmental outcomes, far less is known about how adaptive responses to adversity regulate the developing phenotype to match stressful conditions. As the application of epigenetic methods to human behavior has exploded in the last decade, research has begun to shed light on the role of epigenetic mechanisms in explaining how prenatal conditions shape later susceptibilities to mental and physical health problems. In this review, we describe and attempt to integrate two dominant fetal programming models: the cumulative stress model (a disease-focused approach) and the match–mismatch model (an evolutionary–developmental approach). In conjunction with biological sensitivity to context theory, we employ these two models to generate new hypotheses regarding epigenetic mechanisms through which prenatal and postnatal experiences program child stress reactivity and, in turn, promote development of adaptive versus maladaptive phenotypic outcomes. We conclude by outlining priority questions and future directions for the fetal programming field.

A key concern among developmental psychopathologists is to identify origins of risk for psychopathology. Decades of fetal programming research has revealed that many forms of problem behavior have roots before the child is born in the form of biobehavioral susceptibility to mental illness (Glover, 2011; O'Connor, Monk, & Fitelson, 2014). To date, the empirical literature is quite clear: prenatal exposures to certain forms of stress or maternal psychopathology plant seeds that place children at risk for problem behavior, above and beyond the quality of the child's postnatal environment (O'Connor, Heron, Golding, Glover, & ALSPAC Study Team, 2003). For example, prenatal exposure to maternal mood disorders may account for 10%–15% of the variance in children's behavior problems, above and beyond concurrent levels of maternal mood symptoms (Glover, 2015). However, this field of research lacks theory-driven, mechanistic explanations for the processes involved in the fetal programming of

both adaptive and maladaptive behavior. In other words, the empirical evidence largely describes prenatal exposure–behavior associations but does not explain them.

There are two dominant theoretical frameworks describing fetal programming processes, which, following Nederhof and Schmidt (2012), we term the *cumulative stress model* and the *match–mismatch model*. These relatively new theoretical frameworks seek to articulate developmental processes by which prenatal psychosocial conditions may shape postnatal outcomes. However, they fall short in two ways. First, they are largely devoid of hypothesized biological mechanisms by which prenatal programming of risk for psychopathology may occur. Second, they both stem from the metabolically (nutrition) focused fetal programming literature (e.g., Barker, 2002; Bateson, Gluckman, & Hanson, 2014; Gluckman, Buklijas, & Hanson, 2016). Although this literature is relatively advanced in terms of both theory and data, the links between this literature and fetal programming on the basis of psychosocial stress are unclear. A third model, biological sensitivity to context (BSC; Boyce & Ellis, 2005), offers a novel perspective on the function of biobehavioral reactivity to stress that, in conjunction with the cumulative stress and match–mismatch models, proposes a new set of hypotheses that we outline below.

A rather glaring gap in the fetal programming literature is the lack of integration between metabolically and psychosocially focused models, and especially the failure of psychosocially focused models to incorporate theoretical and empirical advances from the metabolic literature. A central goal of the current manuscript is to bridge this gap by building on extant

The first author (E.C.) and the senior author (B.E.) contributed equally to this manuscript. This manuscript was supported by the National Institute of Mental Health under Award Number R21MH109777 (to S.C. and E.C.), a Career Development Award from the National Institute on Drug Abuse 7K08DA038959-02 (to E.C.), and a grant from the University of Utah Consortium for Families and Health Research. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institute of Mental Health, the National Institute on Drug Abuse, or the National Institutes of Health.

Address correspondence and reprint requests to: Elisabeth Conradt, University of Utah, Department of Psychology, 380 South 1530 East BEHS 602, Salt Lake City, UT 84112; E-mail: Elisabeth.Conradt@psych.utah.edu.

frameworks to generate specific, testable hypotheses that take into account potential epigenetic mechanisms through which prenatal psychosocial stress may lead to increased susceptibility for psychopathology. We argue that a key biological mechanism mediating these effects is epigenetic regulation, and we describe research studies from the behavioral epigenetic and prenatal stress literatures that offer preliminary support for these hypotheses. We focus predominantly on the perinatal period given our emphasis on how prenatal processes may influence postnatal susceptibility for psychopathology.

Epigenetics

Epigenetics is the study of molecular processes occurring on and around the genome that regulate gene activity without changing the underlying DNA sequence (Bird, 2007). Thus, epigenetic changes modify the expression of specific genes without changing the sequence of an individual's genome. Epigenetics describes the microstructure of DNA and its associated proteins, which may be modified to induce upregulation or downregulation of specific genes. Epigenetic mechanisms include three primary, closely related processes: DNA methylation, chromatin remodeling, and histone modification. Through these processes, current and past stress exposures, including a parent's preconception exposures and childhood environmental effects, are "communicated" to the developing fetus. Although epigenetic changes can occur in response to one's own developmental experiences (as reviewed below), transmission of environmental signals across generations can occur via the passage of epigenetic marks through the germline (potentially allowing a parent's childhood experiences, such as hunger or trauma, to impact the developing fetus). Other transgenerational processes may include stress-induced programming of mitochondria and mitochondrial DNA in the cytoplasm of oocytes, and suboptimal reproductive tract environments that alter the structure of fetal organs (Aiken & Ozanne, 2014; Aiken, Tarry-Adkins, & Ozanne, 2016; Sharma, 2017).

We will draw from the behavioral epigenetic literature to describe exemplar studies that illustrate key hypotheses stemming from the cumulative stress and match–mismatch models that are described below. We focus specifically on DNA methylation of genes associated with the child's neuroendocrine system as a mechanism that may be especially relevant for psychosocial stressors. This is one possible route through which DNA methylation could exert an effect on children's postnatal outcomes and is consistent with cumulative stress models that have been used to describe the development of psychopathology.

Cumulative Stress Model

The *cumulative stress model* instantiates a disease-focused approach. Its central assumption is that developmental exposures to stress cumulatively add up to cause disruptions of

brain structure and function, resulting in dysregulation of physiological mediators "that are the precursors of later impairments in learning and behavior as well as the roots of chronic, stress-related physical and mental illness" (Shonkoff et al., 2012, p. e236).

The developing phenotype emerges via complex gene–environment interactions across the life span. The environment provides essential building blocks for development, such as oxygen, water, and amino acids in foods. Environmental factors also afford cues reflecting fluctuating conditions, such as changes in photoperiod or population density, which influence development and behavior. Operating through both genomic and nongenomic mechanisms, environmental factors "activate, inhibit, modulate, and coordinate developmental events and physiological processes" (West-Eberhard, 2003, p. 110). Stress exposures (i.e., environmental events signaling threats to survival or well-being) produce a set of complex, highly orchestrated responses within the neural circuitry of the brain and peripheral neuroendocrine pathways regulating metabolic, immunologic, and other physiological functions (reviewed in Boyce & Ellis, 2005).

Developmental pathways typically evolve canalization properties (e.g., biochemical buffering mechanisms) that confer robustness against accidents and other abrupt environmental insults. However, the accumulation of such events over time can negatively affect development, resulting in deviations from the target phenotype. A key assumption of cumulative stress models is that accidents and other environmental insults constrain, rather than adaptively calibrate, the developing phenotype. Cumulative stress models (e.g., Evans, Li, & Whipple, 2013; Seifer et al., 1996) assume that there is an optimal pattern of development, and that the more stressors (prenatally and postnatally) that children are exposed to, the more their developmental competencies will be compromised (i.e., deviate from the optimum; Belsky, Schlomer, & Ellis, 2012). Cumulative stress models have commonly been framed in terms of diathesis stress, where exposures to childhood adversities (prenatally and postnatally) interact with personal vulnerabilities to potentiate psychopathology. In the diathesis stress framework, certain children are vulnerable or resilient because of personal characteristics (e.g., heightened biobehavioral reactivity to stress, low-activity monoamine oxidase A [MAOA] allele) that moderate cumulative stress exposures (Belsky & Pluess, 2009).

From this cumulative stress perspective, high levels of prenatal stress exposure in utero contribute to excess fetal cortisol exposure, which, in turn, increases risk for poor gestational health outcomes such as preterm birth, low birth weight, and increased neonatal stress reactivity (Lester, Conradt, & Marsit, 2013; Monk, Spicer, & Champagne, 2012; Sandman, Glynn, & Davis, 2016). This process is conceptualized as fetal programming for adult disease and risk for psychopathology. We define *fetal programming* as the process by which exposures in the intrauterine environment can cause changes in structure and function of the fetal brain, organs, stress response systems, and behavior that occur in part via

epigenetic mechanisms (Glover, 2011). Of note, important programming effects such as alterations of fetal stress response systems occur prenatally (Jensen Peña, Monk, & Champagne, 2012; Sandman, Davis, Buss, & Glynn, 2012); less attention has been paid to the epigenetic basis of postnatal programming.

Most published behavioral epigenetic studies have been guided by and support the cumulative stress model (for reviews, see Cao-Lei et al., 2017; Turecki & Meaney, 2016). However, no study that we know of has reported prospective associations between prenatal stress, epigenetic processes, and problem behavior, though many have found relations between prenatal stress and epigenetic mechanisms, or epigenetic mechanisms and problem behavior. For example, Monk et al. (2016) reported that higher levels of perceived stress during pregnancy were associated with greater DNA methylation of *HSD11 β 2* and *FKBP5*, which are both involved in hypothalamic–pituitary–adrenal (HPA) axis functioning, and DNA methylation of these genes was associated with reduced fetal coupling, an index of fetal neurodevelopment.

Another exemplar study concerns the epigenetic regulation of the glucocorticoid receptor gene (*NR3c1*), history of maltreatment, and psychopathology (Parade et al., 2016). These authors found that in a sample of maltreated preschoolers ages 3–5, higher levels of early adversity were associated with greater methylation of exons 1_D and 1_F of *NR3c1*. Greater methylation of these regions of *NR3c1* was in turn related to higher levels of internalizing problems (but not externalizing problems). Furthermore, mediational analyses indicated there was a significant indirect effect from maltreatment to *NR3c1* methylation to children's internalizing problems. While no indicators of prenatal stress were available, it is likely that maltreated preschoolers were also exposed to high levels of prenatal stress. The results of both studies support the hypothesis that greater stress exposure leads to increased risk for problem behavior in part via DNA methylation of genes involved in neuroendocrine functioning, which aligns with the cumulative stress model. It is unclear whether there may be adaptive effects of stress in regulating development. Careful analysis, supported in part by the match–mismatch model, suggests that early stress exposures may result in developmental trade-offs with both negative and positive phenotypic effects (reviewed in Ellis, Bianchi, Griskevicius, & Frankenhuis, 2017; Ellis & Del Giudice, 2014).

Match–Mismatch Model

The match–mismatch model extends the cumulative stress model by applying evolutionary models to explain how a developing fetus adapts in anticipation of the postnatal environment as a result of cues experienced prenatally (e.g., Gluckman, Hanson, & Spencer, 2005; Kuzawa & Quinn, 2009; Nederhof & Schmidt, 2012). During fetal development and infancy, important features of the environment are communicated to the child via the placenta and lactation in the form of

nutrients, metabolites, hormones, growth factors, and immune factors that reflect the mother's current and past experiences (Bateson et al., 2014; Kuzawa & Quinn, 2009). For example, high levels of stress hormone exposure provide probabilistic information about the postnatal environment, specifically that it will likely also be harsh or unpredictable (e.g., Del Giudice, Ellis, & Shirtcliff, 2011). Beyond these molecular signals from the mother, relevant features of the extrauterine environment are detected and encoded directly through the child's ongoing experiences. These multilayered environmental signals reflect the continuity of development: the organized phenotype is initially provided by the parents in the form of a zygote, which at conception already encapsulates a rich array of genomic, epigenomic, and environmental information, and which then changes over ontogeny in response to ongoing genomic and environmental influences (West-Eberhard, 2003).

In most species, single “best” strategies for survival and reproduction (i.e., optimal developmental pathways) are unlikely to evolve. This is because the “best” strategy varies as a function of the physical, economic, and social parameters of one's environment (Crawford & Anderson, 1989), and thus a strategy that promotes success in some environmental contexts may lead to failure in others. This context specificity provides the evolutionary basis for adaptive developmental plasticity (i.e., development of different phenotypes that promote fitness under different conditions), which is critically important for enabling organisms to adapt to stress. Dangerous and stressful childhoods have always been part of the human experience. Almost half of children in hunter-gatherer societies (the best model for human demographics before the agricultural revolution) die before reaching adulthood (Volk & Atkinson, 2013). Thus, from an evolutionary–developmental perspective, stressful rearing conditions, even if those conditions engender sustained stress responses that must be maintained over time, should not so much impair physiological systems as direct or regulate them toward set points and reactivity patterns that promote survival and reproductive fitness under stressful conditions (Ellis, Boyce, Belsky, Bakermans-Kranenburg, & van IJzendoorn, 2011; Ellis et al., 2012). A central assumption of the match–mismatch model is that such calibration of physiological systems can result in a mismatched phenotype (one that is poorly prepared for its environment) when early rearing conditions fail to adequately predict the future.

Developmental plasticity involves developmental change from one form to another. Consistent with our more specific definition of fetal programming, when such plasticity results in “durable biological change in the structure or function of a tissue, organ, or biological system” (Kuzawa & Quinn, 2009, p. 132), it is commonly described as developmental programming. The occurrence of developmental programming, which is ubiquitous in the animal world (see DeWitt & Scheiner, 2004; Pigliucci, 2001; West-Eberhard, 2003, for extensive reviews), is uncontroversial. For example, it is widely recognized that harsh developmental conditions, such as exposure

to a suboptimal intrauterine environment, can induce durable biological changes in the phenotype (Sandman et al., 2016). Rather, contention exists when trying to determine whether exposures to physical and psychosocial stressors simply constrain development, as assumed in the cumulative stress model, or guide it in an adaptive manner.

Developmental mismatch is a potential cost of developmental programming. It is a risk incurred by an organism when it employs early information (e.g., early life stress) to shape later developmental trajectories. Mismatch occurs when environmental cues encountered early in development have limited validity, and thus adaptive responses fail to correctly predict future environmental conditions. Selection can favor developmental programming, even if the potential costs are high, as long as the average benefits outweigh the average costs over evolutionary history (for extended discussion, see Ellis & Del Giudice, 2014; Frankenhuis & Del Giudice, 2012).

Metabolically Focused Fetal Programming Models

The costs and benefits of developmental plasticity means that the fetus potentially, but not always, benefits from having the ability to change structure and function in response to cues from the intrauterine environment (Sandman et al., 2016; Wadhwa et al., 2002). A variety of metabolically focused fetal programming models have been developed to explain early developmental adaptations to nutritional conditions. In some of these models (e.g., the *predictive adaptive response* hypothesis, Gluckman & Hanson, 2005; Gluckman et al., 2005; the *thrifty phenotype* hypothesis, Hales & Barker, 1992; Hales & Barker, 2001), the external environment, as experienced through fetal and infant nutrition, provides the key ecological cues to which the developing child adapts. In other metabolically focused models (e.g., the *maternal capital* hypothesis, Wells, 2003; Wells & Johnstone, 2017; the *intergenerational inertia* hypothesis, Kuzawa, 2005, 2008), “the information processed by offspring during placental nutrition and lactation derives from the maternal phenotype, rather than directly from the external environment” (Wells & Johnstone, 2017, p. 24). In these latter models, the maternal phenotype (e.g., maternal birth weight, body size, and nutritional status at conception) provides a smoothed, cumulative signal of nutritional conditions experienced by the mother, and even the mother’s mother, during development. This “smoothed signal” presumably affords a more reliable basis for predicting future conditions (and thus for selecting an appropriate developmental trajectory).

In turn, according to some metabolically focused fetal programming models (e.g., the thrifty phenotype hypothesis and the maternal capital hypothesis), the fetus receives cues prenatally regarding the expected quality of the early postnatal environment (especially conditions during infancy), and the fetus potentially employs these cues to calibrate physiological systems and organs to match this temporally proximal context. For example, according to the thrifty phenotype hypothesis, fetal malnutrition sets in motion mechanisms of fetal

nutritional thrift (e.g., insulin resistance), which results in greater resources being allocated to critical structures necessary for early survival (especially the brain) at the expense of less critical organs (such as the kidneys). In contrast, other metabolic fetal programming models (e.g., the predictive adaptive response hypothesis and the intergenerational inertia hypothesis) adopt a more long-term view. These models conceptualize cues about the state of the environment received during fetal developmental as forecasting the likely adult environment in which reproduction will occur. For example, according to the predictive adaptive response hypothesis, exposures to famine conditions during fetal or infant development adapts the individual’s physiology (e.g., insulin resistance), morphology (e.g., small size and central adiposity), and behavior (e.g., low activity level and higher set points for satiety) for persisting famine conditions in adulthood.

Each of these different metabolically focused fetal programming models can be conceptualized in a match–mismatch framework (as opposed to cumulative stress). Each model presumes that the developing organism detects and encodes cues to current/past environmental conditions and uses these cues as a basis for calibrating developmental trajectories (in either the short or the long term) to match those conditions. Although such matching depends on the stability of the environment over developmental time (e.g., Nettle, Frankenhuis, & Rickard, 2013; Rickard, Frankenhuis, & Nettle, 2014; Sheriff & Love, 2013), the degree of similarity between early/past conditions and later conditions that is necessary for matching to occur (e.g., for a nutritionally thrifty phenotype to remain adaptive) is an empirical question. When the past fails to adequately predict the future, the resulting mismatch can result in a phenotype that is poorly prepared for its environment (and is thus at risk for poor cardiometabolic outcomes; e.g., Gluckman & Hanson, 2005; Gluckman et al., 2005; Hales & Barker, 2001).

Psychosocially Focused Fetal Programming Models

The metabolically focused fetal programming models afford a framework for considering the programming effects of prenatal *psychosocial stress*. Psychosocially focused fetal programming models are currently at an early stage of development. To date, these models most closely parallel the thrifty phenotype hypothesis, insofar as they focus on maternal cues to psychosocial stress experienced during pregnancy and their effects on behavioral development in the postnatal period (infancy to early childhood).

Support for psychosocially focused fetal programming models has accrued in studies of human infants exposed to varying forms of prenatal stress. Infants exposed to high levels of stress prenatally who are then born prematurely may be more likely to experience high levels of stress postnatally, reflecting a “match” between their prenatal and postnatal environments. It may even benefit the fetus to be born prematurely rather than risk fetal demise by remaining in an intrauterine environment characterized by high levels of

stress hormone exposure (Glynn, Davis, & Sandman, 2013; Wadhwa et al., 2002). For instance, Sandman and colleagues (Sandman et al., 1999; Sandman & Davis, 2012) found that high levels of prenatal maternal stress is related to increased risk for a shortened gestational length and, in extreme cases, preterm birth, via high levels of corticotropin-releasing hormone exposure. Corticotropin-releasing hormone is produced by the hypothalamus and is part of the HPA axis, the end product being cortisol. Children exposed to stress prenatally and who are born earlier may exhibit a more reactive phenotype (DiPietro, Novak, Costigan, Atella, & Reusing, 2006). We review anthropological research below that describes how prenatal stress may produce a certain phenotype that supports survival in high-stress postnatal conditions (Scheper-Hughes, 1985).

A mismatch between a prenatal environment characterized by high stress and a low-stress postnatal environment may confer risk for cognitive and motor delays (Sandman, Davis, Buss, et al., 2012). The only study that we know of to explicitly test the match–mismatch model and underlying mechanisms in the human developmental literature was performed by Sandman, Davis, and Glynn (2012). They examined whether infants raised in prenatal and postnatal environments concordant for maternal depression (either exposed to maternal depression prenatally and postnatally or not exposed to maternal depression prenatally and postnatally) had better cognitive and motor outcomes compared to infants raised in environments discordant for maternal depression (either exposed to maternal depression prenatally but not postnatally or exposed to maternal depression postnatally but not prenatally). In support of the match–mismatch model, the results indicated that infants raised in environments concordant for maternal depression, even those exposed to maternal depression prenatally and postnatally, had higher cognitive and motor outcomes at 12 months compared to infants with discordant maternal depression exposure. These findings are inconsistent with the cumulative stress model because infants who experienced maternal depression during prenatal and postnatal periods did not differ from those infants whose mothers did not experience maternal depression. Maternal prenatal and postnatal basal plasma cortisol was tested as a possible mechanism for these findings, but maternal cortisol was not associated with maternal depression. It is therefore unclear what the probable mechanisms are, though one key mechanism is likely related to programming processes occurring in utero.

BSC Model

Fetal programming models focusing on cumulative stress and match–mismatch can potentially be extended by explicitly taking into account BSC theory (Boyce & Ellis, 2005; Bush & Boyce, 2014; Ellis, Essex, & Boyce, 2005). The BSC model rests on three important assumptions that we have used to generate our hypotheses below. First, individual differences in the magnitude of biological stress responses

function to regulate openness or susceptibility to environmental influences, ranging from harmful to protective. Second, early life experiences adaptively calibrate stress response systems to match developmental contexts. Third, patterns of stress reactivity may be altered during sensitive periods of development depending on environmental inputs; more biologically susceptible children may be the best candidates for reprogramming during sensitive periods. Here we elaborate on these assumptions.

First, the BSC model proposes that heightened psychobiologic reactivity to stress moderates the effects of early environmental exposures on physical and mental health outcomes in a bivalent manner, with more reactive children displaying increased sensitivity to both positive and negative environmental influences. Boyce and Ellis (2005) argued that these bivalent effects necessitated reconceptualizing stress reactivity more broadly as *biological sensitivity to context*, which they defined as neurobiological susceptibility to both cost-inflicting and benefit-conferring features of the environment. For example, Obradović, Bush, Stamperdahl, Adler, and Boyce (2010) found that kindergartners with high respiratory sinus arrhythmia reactivity exhibited the lowest levels of externalizing problems and more prosocial behaviors if raised in low adversity environments but high externalizing symptoms and low prosocial behaviors in high adversity contexts. An implication of BSC theory is that calibration of stress response systems, which we argue below has an epigenetic basis, regulates differential susceptibility to environmental influences.

Second, the BSC model proposes that, early in life, and likely even in utero, the fetus receives cues about the quality of the postnatal environment, akin to a weather forecast (Boyce & Ellis, 2005; see also Del Giudice et al., 2011). These cues, according to the theory, lead to the upregulation of stress response systems in both highly positive *and* highly adverse environments, resulting in a U-shaped curvilinear relation between such environmental exposures and heightened stress reactivity. A “sensitive” reactivity pattern is predicted to develop in safe, low-stress environments. High stress reactivity in this context is hypothesized to enhance social learning and engagement with the external world. By contrast, a “vigilant” reactivity pattern is predicted to develop in harsh, stressful environments. High reactivity in that context is hypothesized to enable people to cope more effectively with dangers and unpredictable threats. Finally, “buffered” patterns (low to moderate stress reactivity) are predicted to develop preferentially in conditions of moderate environmental stress, where they strike a balance between costs and benefits of reactivity. A reasonable empirical literature has supported the emergence of these two high-stress reactivity profiles in comparison to the buffered profile (reviewed in Ellis, Del Giudice, & Shirtcliff, 2017).

These patterns of stress reactivity are not fixed, however, and may be altered during sensitive periods of development. It is possible that exposure to harsh early environments could lead to the upregulation of stress response systems (as per the

vigilant pattern), but subsequent positive caregiving experiences could lead to the dampening of these stress response systems (Weaver et al., 2004). This reprogramming of the stress response system may be more likely in children who show high BSC (including sensitive and vigilant patterns). These changes in stress response system functioning could have epigenetic underpinnings, a point which we elaborate on below when outlining our hypotheses.

Using Epigenetic Methods to Uncover Mechanisms Underlying Prenatal and Postnatal Programming

Prenatal and postnatal programming models have been evaluated in rodent studies, with efforts to uncover mechanisms explaining how stress dysregulates developmental (as per the cumulative stress model) and how matched (compared to mismatched) environments may support adaptive phenotypic development (as per the match–mismatch model). A large rodent literature focusing on rats and mice has extensively examined the effects of both prenatal and postnatal stress exposures on development (reviewed in Howell, Neigh, & Sanchez, 2016). Much of this work focuses on observing or manipulating the powerful dam–pup relationship (e.g., maternal separation). As with humans, there is large natural variation in the quality of maternal investment in rats and mice. This variation has been most extensively studied in terms of the frequency of licking and grooming (LG; including both body and anogenital licking) and the amount of arched back nursing (ABN; where the mother assumes a crouching posture that enhances suckling). Because levels of LG and ABN are correlated across dams, researchers have been able to operationalize individual differences in maternal investment in terms of frequency of LG-ABN.

Although these individual differences are partially heritable, cross-fostering studies demonstrated that a component of variance in quality of maternal care is transmitted across generations through epigenetic mechanisms. The medial preoptic area (MPOA) of the hypothalamus is rich in estrogen receptors (ER), which substantially influence maternal behavior (via estrogen-induced oxytocin receptor binding). ER α expression in the MPOA and associated oxytocin receptor levels are increased in adult females who experienced high versus low levels of LG-ABN from their mothers. As a result of these changes in gene expression, the daughters of high LG-ABN mothers show increased LG-ABN during lactation when they are adults.

However, experiencing substantial stress during pregnancy completely reverses the intergenerational transmission of high LG-ABN: lactating mothers who have a developmental history of high LG-ABN only display a pattern of high LG-ABN themselves when their pregnancies occur under relatively safe, stable conditions. When exposed to substantial gestational stress (e.g., 1 hr of restraint stress per day for 10 days), these formerly high LG-ABN mothers shift to become low LG-ABN mothers; moreover, this new pattern of low LG-ABN persists across (at least) two subsequent

litters (after cessation of the original gestational stressor) without any further gestational stress exposures (Champagne & Meaney, 2006; Smith, Seckl, Evans, Costall, & Smythe, 2004). Finally, at a mechanistic level, gestational stress exposures resulted in reduced levels of oxytocin receptor binding in the MPOA. In total, gestational stress effectively remodeled the methylation patterns involved in maternal behavior. This contextual sensitivity of the epigenome may help explain the well-documented sensitivity of quality of parenting in rodents to variations in ecological context (e.g., predation, food availability, and social competition; Beery & Francis, 2011; Champagne, 2008). In total, high levels of LG-ABN were stable only under conditions of low stress exposure for females when they were pups and low stress exposure when the females were pregnant themselves (i.e., when early life and subsequent environments while pregnant were *matched* with respect to low stress exposure). The possibility of reprogramming prevented mismatch.

This ecologically sensitive developmental programming of LG-ABN is central to developmental plasticity because, through epigenetic mechanisms, LG-ABN calibrates variation in physiological and behavioral reactivity to stress and reproductive strategies to match developmental conditions (reviewed in Cameron et al., 2005; Ellis, Jackson, & Boyce, 2006). Heightened biobehavioral reactivity to stress and sexual precocity may promote vigilance for environmental dangers and early opportunities for mating and reproduction. Operating mechanistically through its effects on methylation of glucocorticoid receptors in hippocampal neurons, low LG-ABN upregulates autonomic and adrenocortical stress reactivity in pups, resulting in higher rates of fear-induced behavior, increased burying behavior in response to threats, stronger startle reflexes, and decreased open-field exploration (reviewed in Cameron et al., 2005; Meaney, 2010). Maternal LG-ABN also affects variation in the sexual development and reproductive behavior of offspring through epigenetic regulation of ER α expression in the anteroventral periventricular nucleus of the hypothalamus. Operating through increases in ER α expression in this cell cluster, low LG-ABN biases development of female pups toward earlier onset of puberty, higher sexual proceptivity toward novel males, increased lordosis in response to male mounts, and sharply higher rates of pregnancy following mating sessions (Cameron, Del Corpo, et al., 2008; Cameron, Shahrokh, et al., 2008; Sakhai, Kriegsfeld, & Francis, 2011). In total, at multiple functionally related levels, low maternal LG-ABN biases offspring development toward “faster” life history strategies that, theoretically, are matched to harsh ecological contexts.

Life history theory and research specifies relations between childhood exposures to stress and individual differences in the development of *life history strategies* (Belsky, Steinberg, & Draper, 1991; Chisholm, 1999; Ellis, Figueredo, Brumbach, & Schlomer, 2009; Nettle, Coall, & Dickins, 2011), which encompass integrated suites of traits that vary along a dimension of slow versus fast (Ellis et al., 2009; Kaplan, & Gangestad, 2005). In terms of behavioral processes,

individuals pursuing a slower life history strategy tend to have longer time horizons, tolerate more delay of gratification, show more aversion to risk, display better self-regulation and behavioral control, start sex and reproduction at relatively late ages, and devote more effort to parenting. By contrast, individuals pursuing a faster life history strategy tend to have shorter time horizons, discount future rewards and losses, engage in more risky and aggressive behavior, begin sex and reproduction at relatively early ages, and allocate less effort to parenting (Belsky et al., 1991; Ellis et al., 2012; Figueredo et al., 2006, 2014). Conceptualizing these individual differences as life history strategies provides a framework for explaining the coherent, functional phenotypic outcomes of developmental programming.

The development of faster life history strategies under conditions of early life adversity reflects resource allocation trade-offs that “make the best of a bad job” (by specializing skills and behavioral strategies to match high-adversity contexts), even though “the best” may constitute a high-risk strategy with substantial costs (Ellis et al., 2011, 2017), including stress-related physical and mental health problems (as per the cumulative stress model). Such costs may be minimized, however, in matching environmental contexts. For example, low maternal LG-ABN alters pups’ stress physiology and brain morphology in ways that seem harmful (i.e., higher corticosterone levels, shorter dendritic branch lengths, and lower spine density in hippocampal neurons) and even impair performance on tests of spatial learning and memory (e.g., object recognition tests and the Morris water maze) under low-stress conditions (reviewed in Bagot et al., 2009). However, when such rats are tested under high-stress conditions, they showed increased hippocampal long-term potentiation (a cellular process underlying learning and memory) and increased memory performance on a hippocampal-dependent contextual fear-conditioning task. By contrast, rats that experienced high LG-ABN, while advantaged in learning and memory tasks performed under low-stress conditions, performed relatively poorly when tested in stressful contexts (Bagot et al., 2009; Champagne et al., 2008). Thus, rats that experienced low levels of LG-ABN performed poorly in cognitive tasks under low-stress (mismatched environment), but not under high-stress (matched environment) conditions. In total, the rat data provide intriguing support for the match–mismatch model and, potentially, a rich source for hypothesis generation regarding human development, to which we now turn.

Fetal Programming Models: A Need for Testable Hypotheses That Incorporate Epigenetic Mechanisms

The match–mismatch model could be considered a first-generation fetal programming theory given the empirical support from the nutrition literature. This hypothesis was born from a series of studies known collectively as the Dutch Hunger Winter. During the autumn of 1944–May 1945, the Dutch government ordered a transportation strike against the Nazis because the government believed the Netherlands would soon be

liberated, and in retaliation the Nazis blockaded the western region of the Netherlands (Smith, 1947). Tragically these towns were not liberated until May 1945 and so during the Winter of 1944–1945 no food was transported to the cities of Rotterdam and the Hague. As a result of this strike the inhabitants, including pregnant women, were restricted to as little as 1,145 calories per day (Smith, 1947). The recommended caloric intake for a healthy pregnant woman ranges from 1,800 calories per day in the first trimester to 2,400 calories per day in the third trimester (West, Hark, & Catalano, 2017), so these women experienced severe caloric restriction during pregnancy. Detailed records of this event as well as the birth weights of the babies born were taken and so it was possible to link birth weight with metabolic outcomes approximately 60 years later (Lumey, Stein, Kahn, & Romijn, 2009). Lumey et al. (2009) found that adult females exposed prenatally to the Dutch Hunger Winter had elevated LDL cholesterol and triglycerides compared to unexposed peers. Individuals exposed early in gestation (but not in middle or late gestation) were significantly more likely to have coronary heart disease (Roseboom et al., 2000), and individuals exposed during the third trimester were more likely to have a lower head circumference and birth weight (Stein, Zybert, van de Bor, & Lumey, 2004).

The mechanisms driving these effects are likely due in part to epigenetics; specifically DNA methylation of *IGF2*, a maternally imprinted gene related to human growth and development (Heijmans et al., 2008). In other words, severe undernutrition may have resulted in less DNA methylation of *IGF2*, a “cue” to the fetus that food would likely be scarce postnatally and to compensate with a reduction in glucose metabolism and insulin secretion in order to conserve as many calories as possible (Gluckman et al., 2016). Once the famine was lifted, however, these children were raised in a “mismatched” environment consisting of plentiful food. However, their glucose metabolism did not change, and so reductions in metabolism combined with abundant food resulted in poor metabolic outcomes in adulthood (Gluckman et al., 2016; Lumey et al., 2009; Roseboom et al., 2000). As per the thrifty phenotype hypothesis (Hales & Barker, 1992, 2001), a mismatched prenatal and postnatal environment consisting of exposure to poor prenatal but adequate postnatal conditions resulted in increased likelihood of metabolic disease. In addition, consistent with the predictive adaptive response hypothesis (Gluckman & Hanson, 2005; Gluckman et al., 2005), women who were exposed in utero to the Dutch Hunger Winter started reproducing at a younger age, had more offspring, more twins, and were less likely to remain childless than their peers who were not exposed in utero (Painter et al., 2008). In total, prenatal exposure to the Dutch Hunger Winter resulted in fetal programming for a faster life history strategy.

The psychological literature provides sparse empirical evidence outlining specific mechanisms underlying the match–mismatch phenomenon as it applies to psychosocial stress and the development of behavior problems in children. Here we attempt to advance the psychosocially focused fetal programming field by generating four testable hypotheses that

incorporate epigenetic mechanisms of action. These hypotheses are guided by the cumulative stress, match–mismatch, and BSC models and, in some cases, offer competing predictions designed to discriminate between these three perspectives.

For each of these hypotheses, it is important that peripheral tissues be sampled for epigenetic analysis prenatally and postnatally. To date, there are no published studies examining DNA methylation prenatally (e.g., in placenta samples) and DNA methylation postnatally (e.g., in blood or buccal cells). We therefore identify a behavioral epigenetic study that has tested at least one part of each hypothesis and describe how future research could be conducted to more fully test the complete hypothesis. There is a large literature theorizing how programming processes may occur postpartum that we do not intend to summarize here (Del Giudice et al., 2011). We provide a short description of each hypothesis along with a corresponding behavioral epigenetic study in Table 1. We focus on DNA methylation because it is the most widely used epigenetic mechanism in the behavioral epigenetic literature.

Hypothesis 1. Consistent stress exposures that are low both prenatally and postnatally promote DNA methylation patterns related to low to moderate stress reactivity (as per the cumulative stress model and the buffered pattern in the BSC model). Postnatal development in a highly supportive and protected environment may shift development toward greater stress reactivity (as per the BSC conceptualization of a sensitive pattern). Consistently low stress exposures prenatally and postnatally should promote the development of slow life history strategies that are matched to safe, stable contexts (as per the match–mismatch model) and low risk for psychopathology (as per both the cumulative stress and match–mismatch models).

The cumulative stress and match–mismatch models both predict that young children in this matched environment are at the lowest risk for developing psychopathology given that their prenatal and postnatal environments are consistent and characterized by low stress conditions (Nederhof & Schmidt, 2012). Children in this group are typically “control” children in the stress literature as they are more likely to be raised in stable, nurturing homes. There is some divergence between

the two models with respect to the DNA methylation profiles of these children. On the one hand, consistent with cumulative stress models, low stress both prenatally and postnatally may result in DNA methylation profiles that are associated with low stress reactivity (Filiberto et al., 2011; Lester, Conradt, & Marsit, 2014). On the other hand, a particularly supportive prenatal environment characterized by low stress exposure and adequate nutrition may lead to the upregulation of stress response systems via DNA methylation of genes regulating these systems, predisposing infants to a more physiologically reactive phenotype (as per the BSC sensitive profile; Boyce & Ellis, 2005; Del Giudice et al., 2011). According to the BSC model, this heightened stress responsiveness increases susceptibility to the development-enhancing features of what would presumably be a stable, supportive postnatal environment. In this context, heightened stress reactivity has been linked to socially desirable outcomes in young children such as good health (Boyce et al., 1995), higher levels of academic achievement, school competence, and prosocial behaviors (Obradovic, Bush et al., 2010), and lower levels of problem behavior (Conradt, Measelle, & Ablow, 2013).

While the behavioral epigenetic literature is small, there is some indication that young children exposed to low-stress prenatal and postnatal environments tend to have DNA methylation levels that suggest lowered stress reactivity. For example, Murgatroyd, Quinn, Sharp, Pickles, and Hill (2015) found that infants exposed to low levels of maternal depressive symptoms prenatally and postnatally had the lowest levels of DNA methylation of *NR3c1* relative to infants exposed to higher levels of maternal depressive systems prenatally and postnatally. This study provides some initial support for the hypothesis that consistent low stress exposure is actually related to DNA methylation associated with low stress reactivity. Nonetheless, over the first 7–24 months of life, children with typically sensitive mothers who experience increasing levels of maternal sensitivity tend to experience increases in basal cortisol (Berry et al., 2017). Thus, calibration toward higher activation of stress systems may occur postnatally.

Hypothesis 2. Consistent stress exposures that are high both prenatally and postnatally promote DNA methylation of genes associated with upregulation of the stress response

Table 1. Behavioral epigenetic studies advancing the match–mismatch model

	Low postnatal stress	High postnatal stress
Low prenatal stress	N/A	Infants exposed to low prenatal but high postnatal maternal depression have elevated levels of <i>NR3c1</i> methylation compared to infants exposed to concordant high levels of maternal depression (Murgatroyd et al., 2015)
High prenatal stress	Infants whose mothers are depressed but sensitive exhibit DNA methylation levels of <i>NR3c1</i> similar to infants whose mothers are neither depressed nor insensitive (Conradt et al., 2016)	DNA methylation of <i>NR3c1</i> mediates the effect of early life stress on internalizing symptoms in maltreated preschoolers (Parade et al., 2016)

systems early in life (the BSC vigilant profile). This stress-adapted responsivity profile should promote the development of faster life history strategies that are matched to higher adversity contexts (as per the match–mismatch model) and elevated risk for psychopathology (as per the cumulative stress model).

Hypothesis 2 concerns a matching of environmental conditions (high stress exposure) prenatally and postnatally, which according to the match–mismatch model, should support the development of phenotypes (e.g., faster life history strategies) that are adaptive in high-adversity contexts. We argue here that consistent high prenatal and postnatal stress exposure may be related to physiological changes via epigenetic mechanisms that promote fitness in these high-stress conditions.

Adaptations to harsh environments involve trade-offs that can have detrimental effects and undermine fitness, relative to outcomes achieved in well-resourced, safe environments. Despite evidence that children adapt to harsh environments (reviewed in Ellis, Bianchi, et al., 2017; Ellis et al., 2009), such adaptations have costs that can jeopardize child health and survival (see Mulvihill, 2005; Shonkoff, Boyce, & McEwen, 2009). Although developmental adaptations to stress may enable children to “make the best of a bad job,” this does not imply that stress is good for children or should be accepted as an inevitable fact of life.

High maternal prenatal stress should be associated with elevated fetal exposure to corticotropin-releasing hormone and cortisol, which provides the child with cues that the postnatal environment is also likely to be stressful. A large literature across multiple laboratories suggests that infants exposed to high levels of prenatal stress may be born with a more reactive phenotype (Davis et al., 2007; Gutteling et al., 2005; Huizink, Robles De Medina, Mulder, Visser, & Buitelaar, 2002; Montirosso et al., 2016). This type of biobehavioral reactivity in infancy may be adaptive in a stressful early postnatal environment. For instance, infants exposed to at least moderate levels of prenatal stress in some cases exhibit enhanced cognitive and motor outcomes (DiPietro et al., 2006; Sandman, Davis, & Glynn, 2012). Children reared in stressful prenatal and early postnatal environments may be more likely to survive if they exhibit greater stress reactivity and are more active. For example, in a Brazilian shantytown in which families live in abject poverty and where mothers on average lose 3.5 out of 9.5 children, infants who are described as temperamentally “wild” and who are “fighters” are more likely to survive compared to calm infants (Scheper-Hughes, 1985). In this research, mothers with passive infants were more likely to withdraw from child rearing, which is undoubtedly due in part to the impact of infant illness on infant behavior (i.e., in theory, illness → passive infant behavior → maternal withdrawal). While the specific direction of effects is unclear, this study challenges the dominant notion of what features of temperament may lead to more competent developmental outcomes in our Western society. Given that infant mortality is greatest from birth to 6 months of life, it may be that in

particularly harsh environments it is *adaptive* to be more reactive. High infant emotional reactivity (difficult temperament and negative emotionality) is also a reliable indicator of differential susceptibility to parental influences, for better and for worse (Slagt, Dubas, Deković, & van Aken, 2016).

In the behavioral epigenetic literature, the study by Parade et al. (2016) reviewed above partially supports Hypothesis 2, given that high postnatal stress exposure was related to DNA methylation of *NR3c1* exons 1_D and 1_F and greater internalizing behavior. The study by Murgatroyd et al. (2015) also supports this hypothesis, as these researchers found that the highest levels of DNA methylation of *NR3c1* were associated with exposure to high levels of prenatal and postnatal maternal depressive symptoms. High levels of DNA methylation of *NR3c1* may be related to greater neuroendocrine reactivity (Conradt et al., 2015; Oberlander et al., 2008) and a more reactive phenotype in infancy (Conradt et al., 2015). This reactive phenotype may elicit more attention from caregivers (DiPietro, Ghera, & Costigan, 2008; Scheper-Hughes, 1985) and a greater likelihood that the infants’ needs for food and comfort are being met. This phenotype is generally not considered desirable in Western society. For example, a reactive, vigilant phenotype may not be valued in our traditional school settings where children are required to sit still and pay attention. Thus, in Western societies, this hypothesis linking concordant stressful prenatal and postnatal environments to adaptive outcomes may only be supported when considering outcomes in high-adversity contexts.

According to cumulative stress models, under prolonged high-stress conditions, increased activation of stress response systems may ultimately lead to increased risk for psychopathology via “wear and tear” of these systems (Juster, Russell, Almeida, & Picard, 2016; McEwen, 1998). Therefore, while the prenatal and postnatal environment may be matched in the form of high stress exposure, over time children exposed to chronically high levels of stress are likely to experience physiological deterioration (i.e., allostatic load), particularly in the form of a blunted neuroendocrine response to stress and increased risk for psychopathology (Fisher, Gunnar, Dozier, Bruce, & Pears, 2006).

Hypothesis 3. Consistent with both the cumulative stress and match–mismatch models, high prenatal stress exposures promote DNA methylation of placental genes related to the upregulation of stress response systems and high infant stress reactivity (as per the vigilant profile). Because heightened stress reactivity increases susceptibility to environmental influence, children displaying the vigilant pattern who grow up in safe, stable postnatal caregiving environments should recalibrate, resulting in postnatal epigenetic modifications that support lower stress reactivity and a slower life history strategy (and thus lower risk for psychopathology).

Hypothesis 3 is an example of a “mismatch” in the prenatal and postnatal environment. High prenatal stress exposure may calibrate susceptibilities to the postnatal environment

and prepare the fetus for a high-stress postnatal environment (Glover, 2015; Pluess & Belsky, 2009). The resulting vigilant reactivity pattern should support development of a faster life history strategy (Del Giudice et al., 2011) and elevated risk for psychopathology. However, if the postnatal environment is favorable, and if the caregiver can provide sensitive, responsive care for a presumably high reactive infant, then this child may be at lowered risk for psychopathology later in life. This hypothesis is supported by the differential susceptibility literature indicating that the combination of high infant reactivity with sensitive care is related to lower levels of problem behavior (for reviews, see Belsky & Pluess, 2009; Rabinowitz & Drabick, 2017; Slagt et al., 2016). For example, in a sample of toddlers all reared in poverty, Conradt et al. (2013) found evidence for a physiological susceptibility factor, high baseline respiratory sinus arrhythmia, that was associated with the lowest levels of problem behavior in toddlers, but only if the toddlers were raised by supportive, positive caregivers. High prenatal stress exposure, such as exposure to poverty, may therefore lead to physiological changes in stress response systems that predispose children to take advantage of a particularly desirable postnatal environment characterized by low stress exposure.

This hypothesis is also supported by the stress buffering literature. Sensitive caregivers may buffer the child to the effects of stress, resulting in more adaptive developmental outcomes (Hostinar, Sullivan, & Gunnar, 2014). For example, children who are securely attached but who must separate from the mother while in daycare exhibit lower levels of the stress hormone cortisol compared to insecurely attached children attending daycare (Ahnert, Gunnar, Lamb, & Barthel, 2004; Bergman, Sarker, Glover, & O'Connor, 2010).

In the behavioral epigenetic literature incorporating stress research, there is no complete test of this hypothesis given the dearth of research on studies of DNA methylation both prenatally and postnatally. However, the buffering hypothesis was partially supported by evidence indicating that the association between postnatal maternal depression on DNA methylation of *NR3c1* exon 1_F depended on maternal sensitivity during a face-to-face interaction with 5-month-old infants (Conradt et al., 2016). Five-month-old infants exposed to maternal depressive symptoms and maternal insensitivity had the expected high levels of DNA methylation of *NR3c1* exon 1_F. However, infants who were exposed to maternal depression, but whose mothers were sensitive, had DNA methylation levels similar to infants with no exposure to maternal depression or insensitivity (Conradt et al., 2016). These findings partially support Hypothesis 3 given that a postnatal environment characterized by high levels of maternal depression and greater sensitivity was related to lower levels of DNA methylation of *NR3c1* exon 1_F and thus may have partially buffered the child to the effects of maternal depression. Still, a more complete test of this hypothesis is needed that includes prenatal placental methylation.

In the first series of studies that we know of to assess DNA methylation at more than one time point, Parent et al. (2017)

found that maltreated preschoolers exhibited significantly lower levels of DNA methylation of *NR3c1* 6 months after state agency involvement. In this same sample, Parade et al. found that maltreated children who received more services exhibited increases in *FKBP5* methylation in one particular region, which may be related to better regulation of the neuroendocrine stress response system. Thus, high stress exposure in the form of child maltreatment may be associated with epigenetic mechanisms that upregulate genes regulating the stress response system (*NR3c1* and *FKBP5*), while “intervention” in the form of agency involvement and service utilization could then lead to the dampening of these stress response systems again via epigenetic mechanisms. These epigenetic studies support Hypothesis 3 and provide important evidence that DNA methylation is malleable to early intervention.

Hypothesis 4. Consistent with both the cumulative stress and the match–mismatch models, low prenatal stress exposures promote DNA methylation of placental genes associated with downregulation of stress response systems and low to moderate infant stress reactivity (as per the buffered reactivity pattern). Subsequent exposure to a harsh, unpredictable postnatal caregiving environment should recalibrate the buffered pattern, resulting in postnatal epigenetic modifications that upregulate infant stress reactivity (promoting a vigilant pattern) and support development of a faster life history strategy (and thus higher risk for psychopathology)

Hypothesis 4 also concerns a mismatch between the prenatal and postnatal environments. According to the match–mismatch hypothesis, children who develop under these conditions are at increased risk for psychopathology because their prenatal environment programmed them for a low-stress postnatal caregiving environment. These children may have difficulty with biobehavioral regulation if their stress response system is not programmed to adaptively respond to a high-stress postnatal environment. In other words, these children’s physiological systems may be poorly prepared to deal with environmental threats and uncertainty, which could undermine social, cognitive, and health outcomes in high-adversity contexts. It is possible that a recalibration of the stress response system may occur in early childhood that could support alternative forms of biobehavioral self-regulation (Del Giudice et al., 2011).

A close test of this hypothesis comes from the work of Murgatroyd et al. (2015), who examined the effects of prenatal and postnatal depression exposure on DNA methylation of *NR3c1* exon 1_F in infants. Infants who were exposed to low levels of prenatal maternal depression but high levels of postnatal maternal depression exhibited greater methylation of *NR3c1* exon 1_F compared to infants exposed to concordant prenatal and postnatal depression. Remarkably, this effect was reversed by maternal stroking of the infant during the first few weeks of life, lending support to Hypothesis 4. These results suggest that postnatal programming processes could

prepare the infant for a stressful postnatal environment given that infants in the low prenatal, high postnatal stress group exhibited greater methylation of *NR3c1*. However, postnatal signals “communicated” to the child by maternal touch reversed the effects, potentially communicating to the child that the postnatal environment will not be a stressful one.

Key Issues for the Field

In an attempt to advance theory and research on fetal programming, we have discussed the cumulative stress model (which instantiates a disease-focused approach) and the match–mismatch model (which instantiates an evolutionary–developmental approach based in life history theory) and tried to integrate and extend these models in a BSC framework. Some may argue that a common vocabulary is warranted, because the conceptual redundancies may limit future work rather than advance the fetal programming field. We agree, and we argue here for the importance of these theories in generating testable hypotheses that can be put at risk through empirical research. If a theory is all encompassing, then it is not generative. We therefore had as a goal to articulate four testable hypotheses that could be put at risk via epigenetic data collected at multiple time points. Here we review priority questions and future directions for the field that are relevant to these hypotheses.

Moving beyond cumulative stress models to study coherent, functional adaptations to stress

There are two key limitations of the disease-focused, cumulative stress approach. First, by emphasizing the pathways leading directly from adversity to dysfunction, the cumulative stress perspective misses the coherent, functional biobehavioral changes that occur in response to stress over time (Ellis & Del Giudice, 2014). We need to characterize these functional developmental changes to understand dysfunctional outcomes including the developmental of psychopathology. These changes not only promote adaptation to temporally proximal contexts (such as using prenatal stress as the basis for calibrating postnatal stress physiology) but also shape longer term developmental trajectories to match expected future conditions (as per the match–mismatch model). Shifting to this kind of a match–mismatch perspective would move psychosocially focused fetal programming research toward investigating integrated suites of traits (the coherent, functional responses to stress that form the basis of life history strategies) that develop over childhood, adolescence, and early adulthood. The developmental programming of life history strategies is critical to explaining and predicting the development of psychopathology (see Ellis & Del Giudice, 2014). This shift toward a life history framework would bring the psychosocially focused literature into better alignment with the metabolically focused fetal programming literature and the rich epigenetic literature on rodents.

Second, evolutionary–developmental perspectives, such as the match–mismatch model, emphasize the importance

of studying *positive* phenotypic outcomes, such as stress-adapted social and cognitive abilities, as well as the usual psychopathology outcomes, because prenatal and postnatal stress exposures can have positive, adaptive effects (Ellis et al., 2017). Stress exposure can result in improved cognition (e.g., attention, perception, detection, learning, and memory) and emotion detection in children who are asked to perform tasks that are ecologically relevant to them, over and above their peers who grew up under relatively safe, stable conditions (Ellis et al., 2017; Frankenhuis & de Weerth, 2013). For example, Raver, Blair, Garrett-Peters, and Family Life Project Key Investigators (2015) found that exposure to interparental verbal aggression in early childhood was related to enhanced accuracy at detection of emotions at age 6, though impaired emotion detection was detected among young children exposed to more physical aggression. Although stress-adapted abilities have been well documented, especially in the animal literature, they may be only observable if tested in ecologically valid contexts (Ellis et al., 2017; Sandman, Davis, & Glynn, 2012).

What information crosses the placenta and is sensed or incorporated by the fetus?

How and when stress exposures occur, for both baby and mother, is a key issue for future research. The term “prenatal stress” is amorphous and, within the behavioral epigenetic literature, is typically conceptualized broadly. For example, stress is defined by a wide range of exposures, including maternal psychopathology or distress, fetal exposure to high levels of maternal corticotropin releasing hormone (Glynn et al., 2013; Wadhwa et al., 2002), and prenatal smoking exposure (Knopik, Maccani, Francazio, & McGeary, 2012). However, the metabolically focused fetal programming literature suggests that the emphasis of psychosocially focused fetal programming research on “prenatal stress” may be misguided. Consider the case of birth weight. As reviewed by Kuzawa (2008), maternal metabolism and physiology maintains a fairly constant flow of nutritional resources to the fetus, regardless of how much the mother eats while she is pregnant. Accordingly, maternal dietary intake during gestation has relatively little impact on birth weight. During the Dutch Hunger Winter, for example, babies born during the famine (and thus exposed during late gestations when caloric deprivation has the largest effect on birth weight) were <.5 lbs. (about 200 g) lighter than babies who were not exposed to the famine (Smith, 1947). Rather than maternal nutrition during pregnancy, it is the mother’s phenotypic condition prior to pregnancy that more strongly predicts child birth weight (Kuzawa, 2008), as per the intergenerational inertia and maternal capital hypotheses. Key indicators of mother’s phenotypic condition in relation to her child’s birth weight include mother’s childhood leg length (more than her adult leg length), the mother’s birth weight (independent of gestational age and the mother’s adult size), and the mother’s nutritional status at the time of conception. These data suggest that the

hormones and metabolites that cross the placenta and are sensed or incorporated by the fetus substantially reflect the mother's developmental history, even in the womb. This highlights the need for psychosocially focused studies of fetal programming to expand their focus beyond "prenatal stress" to more broadly assess the mother's condition and history.

To date, most of the behavioral epigenetic literature has focused on examining the impact of conditions during pregnancy on methylation of a few candidate genes that are involved in the regulation of the HPA axis (Conradt, 2016). This may be a problem because the literature examining links between prenatal stress and cortisol exposure is equivocal, with some studies finding associations between constructs such as pregnancy-specific anxiety and diurnal cortisol in pregnant women (Obel et al., 2005) while others find null results between stress during pregnancy and cortisol in amniotic fluid (Glover, Bergman, Sarkar, & O'Connor, 2009). As noted by Beijers, Buitelaar, and de Weerth (2014), there is a clear need to examine how prenatal stress may become biologically embedded via additional mechanisms such as immune system functioning and through the microbiome. This focus could inform future epigenetic research examining DNA methylation of genes involved in other stress response systems or the immune system.

Incorporating both prenatal and postnatal environmental exposures into developmental programming models

The application of fetal programming concepts to the study of prenatal psychosocial stress represents a relatively new area of research. To date, much of the work on this topic has focused on describing whether various forms of prenatal stress have *unique* consequences for children's postnatal outcomes. These efforts have been fruitful. For example, meta-analyses indicate that maternal depression or anxiety during the prenatal period are associated with children's birth outcomes (Ding et al., 2014; Grote et al., 2010), and growing evidence suggests that these prenatal risks also are uniquely associated with children's behavior problems (O'Connor et al., 2003). That said, a limitation of this research approach is that postnatal experiences have often been conceptualized simply as a potential confound, and few studies examine differences in behavioral outcomes depending on the match (vs. mismatch) of the prenatal and postnatal environments. As we have emphasized throughout this paper, we believe that a critical next step for this field will be testing theoretical models that emphasize understanding children's adaptations to stressful environments (or the lack thereof) during both the prenatal and the postnatal periods. There is a clear need for prospective longitudinal studies that collect epigenetic information beginning prenatally. Studies that collect psychosocial stress data along with blood or buccal cells from newborns at both birth and a later date would be well situated to evaluate the degree to which children's epigenetic profiles are initially programmed by prenatal experiences and are capable of changing in response to children's postnatal environments.

It will also be important to consider the *consistency, stability, and predictability* of prenatal stress exposures, given recent work with animals that suggests that cues experienced prenatally (rather than postnatally) help the offspring to adapt to the postnatal environment (Berghänel, Heistermann, Schülke, & Ostner, 2016). In research with the Assamese macaque (*Macaca assamensis*), high prenatal maternal stress exposure was related to less prenatal food availability and accelerated offspring growth, decelerated motor development, and poorer immune system functioning (Berghänel et al., 2016), independent of the postnatal environment, which was characterized as highly unstable. This work highlights the importance of studying the stability of the prenatal and postnatal environments at multiple times during prenatal and postnatal life, and to incorporate those data into assessments of how early life stress can impact social competence and mental and physical health.

Implementing causally informative designs

There also are a number of innovative research designs that can be leveraged to test the programming hypotheses advanced here, with many of these studies identifying links between prenatal stress exposure and epigenetic regulation of the immune system. A common feature of the designs is their ability to uncouple prenatal and postnatal experiences, which is important because many psychosocial stressors (including socioeconomic disadvantage, stressful life events, and maternal emotional dysregulation) are relatively stable across this period of time. Quasi-experiments, in which individuals' exposure to a stressful life event is unplanned, seemingly independent of the usual confounding variables (such as socioeconomic status), and circumscribed to a finite period of time, are a powerful tool for evaluating the implications of prenatal stress for individuals' development (Rutter, 2007). For example, the previously mentioned studies of individuals born around the time of the Dutch Hunger Famine represent a highly influential example of a quasi-experimental approach to examining the prenatal origins of physical health outcomes (Tobi et al., 2009). Similarly, Project Ice Storm was initiated soon after a series of ice storms that left millions of individuals in Canada without power for up to 6 weeks during January 1998 (King, Dancause, Turcotte-Tremblay, Veru, & Laplante, 2012). Longitudinal follow-ups with the children who were born shortly after the storm have identified various behavioral outcomes that appear to have been impacted by children's in utero exposure to the storm's effects (King et al., 2012). Moreover, pregnant mothers' experiences of stress and subjective appraisals during the storms are associated with children's epigenetic profiles in genes associated with the immune system 13 years later (Cao-Lei et al., 2014, 2015). These longitudinal studies, which integrate information on prenatal exposures with epigenetic mechanisms, could be used to test whether (mis)matches in prenatal and postnatal environments are related to differences in epigenetic profiles.

Studying children adopted at birth offers opportunities for evaluating unique and interactive influences of prenatal and postnatal experiences and include research designs that are ideal for testing mechanisms related to the match–mismatch model. On average, children placed for adoption at birth experience a shift from a high- to a low-stress environment. For example, within the Early Growth and Development Study, a study of over 550 adopted children in the United States, the birth mothers who placed their children for adoption were, on average, 24 years old, 70% were not married, and only 4% had received a 4-year college degree. In contrast, the adoptive mothers had an average age of 37 years, 91% were married, and 64% had received a 4-year college degree (Leve et al., 2007). In addition to mean-level changes in the degree of exposure to stress, children's prenatal and postnatal experiences are independent because children's birth parents provide the prenatal environment but adoptive parents provide the postnatal environment. Finally, there is sufficient variation in the prenatal and postnatal environments to test the ideas about the significance of (mis)matches in psychosocial stress across these periods for children's behavioral outcomes, stress physiology, and epigenetic profiles (Laurent et al., 2013).

One of the most powerful strategies for evaluating causal processes is through the use of experimental designs. Although it is not ethical to assign pregnant mothers to high-stress conditions, interventions can be used to reduce stress for some high-risk pregnant mothers. For example, one way of testing hypotheses about the interplay of prenatal and postnatal experiences would be to randomly assign a third of a sample of distressed pregnant mothers to receive a control intervention, assign another third to receive intervention immediately after birth, and assign the final third of the sample to receive the intervention prenatally. In this way, the mothers who received the control intervention would correspond to the high prenatal stress and high postnatal stress group described in Table 1. The mothers who received the intervention during the postnatal period would correspond to the high prenatal stress but low postnatal stress group. The mothers who received the intervention during the prenatal period would likely correspond to the low prenatal stress and low postnatal stress group because the benefits of the intervention may persist into the postnatal period. Comparing the outcomes, including epigenetic changes, for the children of these three groups of mothers would allow for strong inferences about the potentially causal effects of fetal programming.

Identifying functional epigenetic pathways

Another challenge is to improve epigenetic research through the exploration of functional epigenetic pathways, as opposed to narrowly focused candidate epigenetic approaches. Such pathway approaches have only recently emerged in the epigenetics literature, but build upon decades of genetics research, including both critical advances and missteps in the field. Movement toward polygenic pathway methods has been mo-

tivated by evidence that individual epi/genetic locus effect sizes are generally small, making their individual detection problematic, particularly in highly dimensional (e.g., genome-wide) multiple testing scenarios (Gratten, Wray, Keller, & Visscher, 2014; International Schizophrenia Consortium et al., 2009). Thus, researchers are increasingly investigating networks of functionally related genes clustered into biological systems/ontologies (e.g., the serotonergic system; Adkins et al., 2012). Relatedly, for complex phenotypes, such as depression, statistical geneticists are increasingly applying *polygenic risk score* approaches (de Moor et al., 2015; International Schizophrenia Consortium et al., 2009). These methods require a priori information regarding the strength of association between the assayed epi/genetic loci and phenotype, which is derived from previous large sample studies or meta-analyses (i.e., training data). These training data association estimates are then used as weights for the calculation of polygenic risk scores in a second, independent genomic data set (i.e., testing data; Wray et al., 2014). Independent replication of the polygenic scores in the testing data then serves to both validate and further refine the polygenic risk score for future studies. This approach holds great, largely unrealized, promise for epigenetic studies as well. Rather than looking at methylation patterns in one or two genes, a functional epigenetic pathways approach will hold more promise for testing the theories outlined here.

A related challenge is to expand current research by exploring epigenetic adaptations across multiple biological systems. For example, rather than looking only at networks within a single system (e.g., cortisol-related genes), it is important to examine the effects of stress exposure on genes involved in monoamine neurotransmitter functioning (i.e., serotonin, dopamine, and norepinephrine), immune systems, insulin and weight regulation networks, inflammatory processes, and so on (Beauchaine, Neuhaus, Zalewski, Crowell, & Potapova, 2011; McDade et al., 2017). These epigenetic changes must also be contextualized alongside other neurodevelopmental processes that are affected by adversity, such as neural migration, neurogenesis, myelination, and apoptosis among other stress-sensitive biological adaptations (Perry, 2008). Because many of these changes are difficult to study in human tissues, it remains essential to link findings within the animal literature to human research on the development of psychopathology within high-risk samples.

Moderation

Finally, it is important to examine moderating influences on the association between early exposure to adversity and both adaptive and maladaptive outcomes. As reviewed above, the BSC model focusing on heightened stress responsivity as a marker of differential susceptibility to environmental influence has advanced work in this area. Whereas high stress reactivity was once considered a vulnerability factor that increased risk for psychopathology, it is now widely considered a susceptibility factor that increases sensitivity

to both risk-promoting and development-enhancing environmental conditions (Boyce & Ellis, 2005). Biological sex is another essential moderating variable that remains understudied due to sample sizes that are underpowered to detect sex differences. Nonetheless, there is some literature to suggest that boys may be more vulnerable to the effects of prenatal stress. In one influential paper, Mueller and Bale (2008) found that male mice exposed to stress early in gestation showed maladaptive stress reactivity, anhedonia, and heightened sensitivity to selective serotonin reuptake inhibitor (i.e., antidepressant) treatment. The hypothesized mechanism for these differences was sex-specific placenta responsiveness leading to epigenetic changes across multiple genes in males but not females. In another higher prenatal stress was associated with greater DNA methylation of *NR3c1* and greater infant fearfulness, but only in girls (Ostlund et al., 2016).

Conclusion

Our goal was to advance the field of developmental programming by building on first-generation fetal programming theories (the cumulative stress and match–mismatch models) to outline a series of four testable hypotheses that highlight epigenetic mechanisms through which prenatal and postnatal experiences may program child stress reactivity and, in turn, risk for psychopathology. Our pursuit of this goal was facilitated by two well-developed literatures: the metabolically focused fetal programming literature in humans and the epigenetically focused developmental programming literature in rodents. These two literatures are relatively advanced in terms

of theory and data, with strong foundations in life history theory. By contrast, the psychosocially focused fetal programming literature is relatively undeveloped and not strongly linked to these other literatures. Herein we tried to forge relevant connections by using the metabolically focused fetal programming literature and the epigenetically focused rodent literature (in conjunction with first-generation fetal programming theories) to articulate (a) new psychosocially based fetal programming hypotheses and (b) priority questions and future directions for the field.

Our hypotheses necessarily oversimplified stress as a construct in order to highlight epigenetic processes (for excellent reviews on developmental sequelae of adversity, see Doyle & Cicchetti, 2017; McCrory, De Brito, & Viding, 2010). These hypotheses can and should be revised depending on the empirical evidence available. For example, our hypotheses may only be supported in studies of individuals who are highly susceptible to their environments (Nederhof & Schmidt, 2012). Fetal programming may also only be apparent when studying those who are highly susceptible *and* who are assessed during a stage characterized by a high degree of neural plasticity (e.g., prenatal and early postnatal development). To evaluate these hypotheses, additional research should be conducted that incorporates epigenetic assessments prenatally and postnatally, which will allow for tests of epigenetic changes across development and different tissue types. Only then can we come closer to building a more complete picture of how prenatal conditions do or do not calibrate stress response systems and, in turn, adaptive and maladaptive development.

References

- Adkins, D. E., Khachane, A. N., McClay, J. L., Åberg, K., Bukszár, J., Sullivan, P. F., & van den Oord, E. J. C. G. (2012). SNP-based analysis of neuroactive ligand–receptor interaction pathways implicates PGE2 as a novel mediator of antipsychotic treatment response: Data from the CATIE study. *Schizophrenia Research*, *135*, 200–201. doi:10.1016/j.schres.2011.11.002
- Ahnert, L., Gunnar, M. R., Lamb, M. E., & Barthel, M. (2004). Transition to child care: Associations with infant–mother attachment, infant negative emotion, and cortisol elevations. *Child Development*, *75*, 639–650. doi:10.1111/j.1467-8624.2004.00698.x
- Aiken, C. E., & Ozanne, S. E. (2014). Transgenerational developmental programming. *Human Reproduction Update*, *20*, 63–75. doi:10.1093/humupd/dmt043
- Aiken, C. E., Tarry-Adkins, J. L., & Ozanne, S. E. (2016). Transgenerational effects of maternal diet on metabolic and reproductive ageing. *Mammalian Genome*, *27*, 430–439. doi:10.1007/s00335-016-9631-1
- Bagot, R. C., van Hasselt, F. N., Champagne, D. L., Meaney, M. J., Krugers, H. J., & Joëls, M. (2009). Maternal care determines rapid effects of stress mediators on synaptic plasticity in adult rat hippocampal dentate gyrus. *Neurobiology of Learning and Memory*, *92*, 292–300. doi:10.1016/j.nlm.2009.03.004
- Barker, D. (2002). Fetal programming of coronary heart disease. *Trends in Endocrinology and Metabolism*, *13*, 364–368. doi:10.1016/S1043-2760(02)00689-6
- Bateson, P., Gluckman, P., & Hanson, M. (2014). The biology of developmental plasticity and the Predictive Adaptive Response hypothesis: Developmental plasticity and the PAR response. *Journal of Physiology*, *592*, 2357–2368. doi:10.1113/jphysiol.2014.271460
- Beauchaine, T. P., Neuhaus, E., Zalewski, M., Crowell, S. E., & Potapova, N. (2011). The effects of allostatic load on neural systems subserving motivation, mood regulation, and social affiliation. *Development and Psychopathology*, *23*, 975–999. doi:10.1017/S0954579411000459
- Beery, A. K., & Francis, D. D. (2011). Adaptive significance of natural variations in maternal care in rats: A translational perspective. *Neuroscience and Biobehavioral Reviews*, *35*, 1552–1561. doi:10.1016/j.neubiorev.2011.03.012
- Beijers, R., Buitelaar, J. K., & de Weerth, C. (2014). Mechanisms underlying the effects of prenatal psychosocial stress on child outcomes: Beyond the HPA axis. *European Journal of Child and Adolescent Psychiatry*, *23*, 943–956. doi:10.1007/s00787-014-0566-3
- Belsky, J., & Pluess, M. (2009). Beyond diathesis stress: Differential susceptibility to environmental influences. *Psychological Bulletin*, *135*, 885–908. doi:10.1037/a0017376
- Belsky, J., Schlomer, G. L., & Ellis, B. J. (2012). Beyond cumulative risk: Distinguishing harshness and unpredictability as determinants of parenting and early life history strategy. *Developmental Psychology*, *48*, 662–673. doi:10.1037/a0024454
- Belsky, J., Steinberg, L., & Draper, P. (1991). Childhood experience, interpersonal development, and reproductive strategy: An evolutionary theory of socialization. *Child Development*, *62*, 647–670. doi:10.1111/j.1467-8624.1991.tb01558.x
- Berghanel, A., Heistermann, M., Schulke, O., & Ostner, O. (2016). Prenatal stress effects in a wild, long-lived primate: predictive adaptive responses in an unpredictable environment. *Proceedings of the Royal Academy of Sciences B*, *283*, 1–9. doi:10.1098/rspb.2016.1304
- Bergman, K., Sarkar, P., Glover, V., & O'Connor, T. G. (2010). Maternal prenatal cortisol and infant cognitive development: Moderation by infant–mother attachment. *Biological Psychiatry*, *67*, 1026–1032. doi:10.1016/j.biopsych.2010.01.002

- Berry, D., Blair, C., Willoughby, M., Granger, D. A., Mills-Koonce, W. R., & Family Life Project Key Investigators. (2017). Maternal sensitivity and adrenocortical functioning across infancy and toddlerhood: Physiological adaptation to context? *Development and Psychopathology*, *29*, 303–317. doi:10.1017/S0954579416000158
- Bird, A. (2007). Perceptions of epigenetics. *Nature*, *447*, 396–398. doi:10.1038/nature05913
- Boyce, W. T., & Ellis, B. J. (2005). Biological sensitivity to context: I. An evolutionary–developmental theory of the origins and functions of stress reactivity. *Development and Psychopathology*, *17*, 271–301. doi:10.1017/S0954579405050145
- Boyce, W. T., Chesney, M., Alkon, A., Tschann, J. M., Adams, S., & Cheserman, B. (1995). Psychobiologic reactivity to stress and childhood respiratory illnesses: Results of two prospective studies. *Psychosomatic Medicine*, *57*, 411–422.
- Bush, N. R., & Boyce, W. T. (2014). The contributions of early experience to biological development and sensitivity to context. In M. Lewis & K. D. Rudolph (Eds.), *Handbook of developmental psychopathology* (pp. 287–309). Boston: Springer.
- Cameron, N. M., Champagne, F. A., Parent, C., Fish, E. W., Ozaki-Kuroda, K., & Meaney, M. J. (2005). The programming of individual differences in defensive responses and reproductive strategies in the rat through variations in maternal care. *Neuroscience and Biobehavioral Reviews*, *29*, 843–865.
- Cameron, N., Del Corpo, A., Diorio, J., McAllister, K., Sharma, S., & Meaney, M. J. (2008). Maternal programming of sexual behavior and hypothalamic–pituitary–gonadal function in the female rat. *PLOS ONE*, *3*, e2210. doi:10.1371/journal.pone.0002210
- Cameron, N. M., Shahrokh, D., Del Corpo, A., Dhir, S. K., Szyf, M., Champagne, F. A., & Meaney, M. J. (2008). Epigenetic programming of phenotypic variations in reproductive strategies in the rat through maternal care. *Journal of Neuroendocrinology*, *20*, 795–801. doi:10.1111/j.1365-2826.2008.01725.x
- Cao-Lei, L., de Rooij, S. R., King, S., Matthews, S. G., Metz, G. A. S., Roseboom, T. J., & Boyce, W. T. (2017). Prenatal stress and epigenetics. *Neuroscience and Biobehavioral Reviews*. Advance online publication. doi:10.1016/j.neubiorev.2017.05.016
- Cao-Lei, L., Elgbeili, G., Massart, R., Laplante, D. P., Szyf, M., & King, S. (2015). Pregnant women's cognitive appraisal of a natural disaster affects DNA methylation in their children 13 years later: Project Ice Storm. *Translational Psychiatry*, *5*, e15. doi:10.1038/tp.2015.13
- Cao-Lei, L., Massart, R., Suderman, M. J., Machnes, Z., Elgbeili, G., Laplante, D. P., . . . King, S. (2014). DNA methylation signatures triggered by prenatal maternal stress exposure to a natural disaster: Project Ice Storm. *PLOS ONE*, *9*, e107653. doi:10.1371/journal.pone.0107653
- Champagne, D. L., Bagot, R. C., van Hasselt, F., Ramakers, G., Meaney, M. J., De Kloet, E. R., . . . Krugers, H. (2008). Maternal care and hippocampal plasticity: Evidence for experience-dependent structural plasticity, altered synaptic functioning, and differential responsiveness to glucocorticoids and stress. *Journal of Neuroscience*, *28*, 6037–6045.
- Champagne, F. A. (2008). Epigenetic mechanisms and the transgenerational effects of maternal care. *Frontiers in Neuroendocrinology*, *29*, 386–397.
- Champagne, F. A. (2009). Nurturing nature: Social experiences and the brain. *Journal of Neuroendocrinology*, *21*, 867–868. doi:10.1111/j.1365-2826.2009.01901.x
- Chisholm, J. S. (1999). *Death, hope and sex: Steps to an evolutionary ecology of mind and morality*. Cambridge: Cambridge University Press.
- Conradt, E., Fei, M., LaGasse, L., Tronick, E., Guerin, D., Gorman, D., . . . Lester, B. M. (2015). Prenatal predictors of infant self-regulation: The contributions of placental DNA methylation of NR3C1 and neuroendocrine activity. *Frontiers in Behavioral Neuroscience*, *9*. doi:10.3389/fnbeh.2015.00130
- Conradt, E., Hawes, K., Guerin, D., Armstrong, D. A., Marsit, C. J., Tronick, E., & Lester, B. M. (2016). The contributions of maternal sensitivity and maternal depressive symptoms to epigenetic processes and neuroendocrine functioning. *Child Development*, *87*, 73–85. doi:10.1111/cdev.12483
- Conradt, E., Measelle, J., & Ablow, J. C. (2013). Poverty, problem behavior, and promise: Differential susceptibility among infants reared in poverty. *Psychological Science*, *24*, 235–242. doi:10.1177/0956797612457381
- Crawford, C. B., & Anderson, J. L. (1989). Sociobiology: An environmentalist discipline? *American Psychologist*, *44*, 1449–1459. doi:10.1037/0003-066X.44.12.1449
- Davis, E. P., Glynn, L. M., Schetter, C. D., Hobel, C., Chicx-Demet, A., & Sandman, C. A. (2007). Prenatal exposure to maternal depression and cortisol influences infant temperament. *Journal of the American Academy of Child & Adolescent Psychiatry*, *46*, 737–746. doi:10.1097/chi.0b013e318047b775
- Del Giudice, M., Ellis, B. J., & Shirtcliff, E. A. (2011). The adaptive calibration model of stress reactivity. *Neuroscience and Biobehavioral Reviews*, *35*, 1562–1592. doi:10.1016/j.neubiorev.2010.11.007
- de Moor, M. H. M., van den Berg, S. M., Verweij, K. J. H., Krueger, R. F., Luciano, M., Arias Vasquez, A., . . . Boomsma, D. I. (2015). Meta-analysis of genome-wide association studies for neuroticism, and the polygenic association with major depressive disorder. *JAMA Psychiatry*, *72*, 642. doi:10.1001/jamapsychiatry.2015.0554
- DeWitt, T. J., & Scheiner, S. M. (2004). *Phenotypic plasticity: Functional and conceptual approaches*. Oxford: Oxford University Press.
- Ding, X.-X., Wu, Y.-L., Xu, S.-J., Zhu, R.-P., Jia, X.-M., Zhang, S.-F., . . . Tao, F.-B. (2014). Maternal anxiety during pregnancy and adverse birth outcomes: A systematic review and meta-analysis of prospective cohort studies. *Journal of Affective Disorders*, *159*, 103–110. doi:10.1016/j.jad.2014.02.027
- DiPietro, J. A., Ghera, M. M., & Costigan, K. A. (2008). Prenatal origins of temperamental reactivity in early infancy. *Early Human Development*, *84*, 569–575. doi:10.1016/j.earlhumdev.2008.01.004
- DiPietro, J. A., Novak, M. F. S. X., Costigan, K. A., Atella, L. D., & Reusing, S. P. (2006). Maternal psychological distress during pregnancy in relation to child development at age two. *Child Development*, *77*, 573–587. doi:10.1111/j.1467-8624.2006.00891.x
- Doyle, C., & Cicchetti, D. (2017). From the cradle to the grave: The effect of adverse caregiving environments on attachment and relationships throughout the lifespan. *Clinical Psychology: Science and Practice*, *24*, 203–217. doi:10.1111/cpsp.12192
- Ellis, B. J., Bianchi, J., Griskevicius, V., & Frankenhuis, W. E. (2017). Beyond risk and protective factors: An adaptation-based approach to resilience. *Perspectives on Psychological Science*, *12*, 561–587. doi:10.1177/1745691617693054
- Ellis, B. J., Boyce, W. T., Belsky, J., Bakermans-Kranenburg, M. J., & van IJzendoorn, M. H. (2011). Differential susceptibility to the environment: An evolutionary–neurodevelopmental theory. *Development and Psychopathology*, *23*, 7–28. doi:10.1017/S0954579410000611
- Ellis, B. J., & Del Giudice, M. (2014). Beyond allostatic load: Rethinking the role of stress in regulating human development. *Development and Psychopathology*, *26*, 1–20. doi:10.1017/S0954579413000849
- Ellis, B. J., Del Giudice, M., Dishion, T. J., Figueredo, A. J., Gray, P., Griskevicius, V., . . . Wilson, D. S. (2012). The evolutionary basis of risky adolescent behavior: Implications for science, policy, and practice. *Developmental Psychology*, *48*, 598–623. doi:10.1037/a0026220
- Ellis, B. J., Del Giudice, M., & Shirtcliff, E. A. (2017). The adaptive calibration model of stress reactivity: Concepts, findings, and implications for developmental psychopathology. In T. P. Beauchaine & S. P. Hinshaw (Eds.), *Child and adolescent psychopathology* (3rd ed., pp. 237–276). New York: Wiley.
- Ellis, B. J., Essex, M. J., & Boyce, W. T. (2005). Biological sensitivity to context: II. *Empirical explorations of an evolutionary–developmental theory*. *Development and Psychopathology*, *17*, 303–328. doi:10.1017/S0954579405050157
- Ellis, B. J., Figueredo, A. J., Brumbach, B. H., & Schlomer, G. L. (2009). Fundamental dimensions of environmental risk. *Human Nature*, *20*, 204–268. doi:10.1007/s12110-009-9063-7
- Ellis, B. J., Jackson, J. J., & Boyce, W. T. (2006). The stress response systems: Universality and adaptive individual differences. *Developmental Review*, *26*, 175–212.
- Evans, G. W., Li, D., & Whipple, S. S. (2013). Cumulative risk and child development. *Psychological Bulletin*, *139*, 1342–1396. doi:10.1037/a0031808
- Figueredo, A. J., Wolf, P. S. A., Olderbak, S. G., Gladden, P. R., Fernandes, H. B. F., Wenner, C., . . . Rushton, J. P. (2014). The psychometric assessment of human life history strategy: A meta-analytic construct validation. *Evolutionary Behavioral Sciences*, *8*, 148–185. doi:10.1037/h0099837
- Figueredo, A. J., Vasquez, G., Brumbach, B., Schneider, S., Sefcek, J., Tal, I., . . . Jacobs, W. (2006). Consilience and life history theory: From genes to brain to reproductive strategy. *Developmental Review*, *26*, 243–275. doi:10.1016/j.dr.2006.02.002
- Filiberto, A. C., Maccani, M. A., Koestler, D. C., Wilhelm-Benartzi, C., Avisar-Whiting, M., Banister, C. E., . . . Marsit, C. J. (2011). Birthweight is associated with DNA promoter methylation of the glucocorticoid receptor in human placenta. *Epigenetics*, *6*, 566–572. doi:10.4161/epi.6.5.15236

- Fisher, P. A., Gunnar, M. R., Dozier, M., Bruce, J., & Pears, K. C. (2006). Effects of therapeutic interventions for foster children on behavioral problems, caregiver attachment, and stress regulatory neural systems. *Annals of the New York Academy of Sciences*, 1094, 215–225. doi:10.1196/annals.1376.023
- Frankenhuis, W. E., & Del Giudice, M. (2012). When do adaptive developmental mechanisms yield maladaptive outcomes? *Developmental Psychology*, 48, 628–642. doi:10.1037/a0025629
- Frankenhuis, W. E., & de Weerth, C. (2013). Does early-life exposure to stress shape or impair cognition? *Current Directions in Psychological Science*, 22, 407–412. doi:10.1177/0963721413484324
- Glover, V. (2011). Annual Research Review: Prenatal stress and the origins of psychopathology: An evolutionary perspective: Prenatal stress and the origins of psychopathology. *Journal of Child Psychology and Psychiatry*, 52, 356–367. doi:10.1111/j.1469-7610.2011.02371.x
- Glover, V. (2015). Prenatal stress and its effects on the fetus and the child: Possible underlying biological mechanisms. In M. C. Antonelli (Ed.), *Perinatal programming of neurodevelopment* (pp. 269–283). New York: Springer.
- Glover, V., Bergman, K., Sarkar, P., & O'Connor, T. G. (2009). Association between maternal and amniotic fluid cortisol is moderated by maternal anxiety. *Psychoneuroendocrinology*, 34, 430–435. doi:10.1016/j.psyneuen.2008.10.005
- Gluckman, P. D., Buklijas, T., & Hanson, M. A. (2016). The developmental origins of health and disease (DOHaD) concept. In C. S. Rosenfeld (Ed.), *The epigenome and developmental origins of health and disease* (pp. 1–15). London: Elsevier.
- Gluckman, P. D., & Hanson, M. (2005). *The fetal matrix: Evolution, development, and disease*. New York: Cambridge University Press.
- Gluckman, P. D., Hanson, M. A., & Spencer, H. G. (2005). Predictive adaptive responses and human evolution. *Trends in Ecology & Evolution*, 20, 527–533. doi:10.1016/j.tree.2005.08.001
- Glynn, L. M., Davis, E. P., & Sandman, C. A. (2013). New insights into the role of perinatal HPA-axis dysregulation in postpartum depression. *Neuropeptides*, 47, 363–370. doi:10.1016/j.npep.2013.10.007
- Gratten, J., Wray, N. R., Keller, M. C., & Visscher, P. M. (2014). Large-scale genomics unveils the genetic architecture of psychiatric disorders. *Nature Neuroscience*, 17, 782–790. doi:10.1038/nn.3708
- Grote, N. K., Bridge, J. A., Gavin, A. R., Melville, J. L., Iyengar, S., & Katon, W. J. (2010). A meta-analysis of depression during pregnancy and the risk of preterm birth, low birth weight, and intrauterine growth restriction. *Archives of General Psychiatry*, 67, 1012. doi:10.1001/archgenpsychiatry.2010.111
- Gutting, B. M., de Weerth, C., Willemsen-Swinkels, S. H. N., Huizink, A. C., Mulder, E. J. H., Visser, G. H. A., & Buitelaar, J. K. (2005). The effects of prenatal stress on temperament and problem behavior of 27-month-old toddlers. *European Child & Adolescent Psychiatry*, 14, 41–51. doi:10.1007/s00787-005-0435-1
- Hales, C. N., & Barker, D. J. P. (1992). Type 2 (non-insulin-dependent) diabetes mellitus: The thrifty phenotype hypothesis. *Diabetologia*, 35, 595–601. doi:10.1007/BF00400248
- Hales, C. N., & Barker, D. J. (2001). The thrifty phenotype hypothesis. *British Medical Bulletin*, 60, 5–20.
- Heijmans, B. T., Tobi, E. W., Stein, A. D., Putter, H., Blauw, G. J., Susser, E. S., . . . Lumey, L. H. (2008). Persistent epigenetic differences associated with prenatal exposure to famine in humans. *Proceedings of the National Academy of Sciences*, 105, 17046–17049. doi:10.1073/pnas.0806560105
- Kaplan, H. S., & Lancaster, J. B. (2003). An evolutionary and ecological analysis of human fertility, mating patterns, and parental investment. In K. W. Wachter & R. A. Bulatao (Eds.), *Offspring: Human fertility behavior in biodemographic perspective* (pp. 170–223). Washington, DC: National Academies Press.
- Hostinar, C. E., Sullivan, R. M., & Gunnar, M. R. (2014). Psychobiological mechanisms underlying the social buffering of the hypothalamic–pituitary–adrenocortical axis: A review of animal models and human studies across development. *Psychological Bulletin*, 140, 256–282. doi:10.1037/a0032671
- Howell, B. R., Neigh, G. N., & Sanchez, M. (2016). Animal models of developmental psychopathology. In D. Cicchetti (Ed.), *Developmental psychopathology: Vol. 2. Developmental neuroscience* (3rd ed., pp. 166–201). New York: Wiley.
- Huizink, A. C., Robles De Medina, P. G., Mulder, E. J. H., Visser, G. H. A., & Buitelaar, J. K. (2002). Psychological measures of prenatal stress as predictors of infant temperament. *Journal of the American Academy of Child & Adolescent Psychiatry*, 41, 1078–1085. doi:10.1097/00004583-200209000-00008
- International Schizophrenia Consortium, Purcell, S. M., Wray, N. R., Stone, J. L., Visscher, P. M., O'Donovan, M. C., . . . Sklar, P. (2009). Common polygenic variation contributes to risk of schizophrenia and bipolar disorder. *Nature*, 460, 748–752. doi:10.1038/nature08185
- Jensen Peña, C., Monk, C., & Champagne, F. A. (2012). Epigenetic effects of prenatal stress on 11 β -hydroxysteroid dehydrogenase-2 in the placenta and fetal brain. *PLOS ONE*, 7, e39791. doi:10.1371/journal.pone.0039791
- Juster, R.-P., Russell, J. J., Almeida, D., & Picard, M. (2016). Allostatic load and comorbidities: A mitochondrial, epigenetic, and evolutionary perspective. *Development and Psychopathology*, 28(4, Pt. 1), 1117–1146. doi:10.1017/S0954579416000730
- Kaplan, H. S., & Gangestad, S. W. (2005). Life history theory and evolutionary psychology. In D. M. Buss (Ed.), *The handbook of evolutionary psychology* (pp. 68–95). Hoboken, NJ: Wiley.
- Kaplan, H. S., & Lancaster, J. B. (2003). An evolutionary and ecological analysis of human fertility, mating patterns, and parental investment. In K. W. Wachter & R. A. Bulatao (Eds.), *Offspring: Human fertility behavior in biodemographic perspective* (pp. 170–223). Washington, DC: National Academies Press.
- King, S., Dancause, K., Turcotte-Tremblay, A.-M., Veru, F., & Laplante, D. P. (2012). Using natural disasters to study the effects of prenatal maternal stress on child health and development: Natural disasters and prenatal maternal stress. *Birth Defects Research Part C: Embryo Today: Reviews*, 96, 273–288. doi:10.1002/bdrc.21026
- Knopik, V. S., Maccani, M. A., Francazio, S., & McGeary, J. E. (2012). The epigenetics of maternal cigarette smoking during pregnancy and effects on child development. *Development and Psychopathology*, 24, 1377–1390. doi:10.1017/S0954579412000776
- Kuzawa, C. W. (2005). Fetal origins of developmental plasticity: Are fetal cues reliable predictors of future nutritional environments? *American Journal of Human Biology*, 17, 5–21. doi:10.1002/ajhb.20091
- Kuzawa, C. W. (2008). The developmental origins of adult health: Intergenerational inertia in adaptation and disease. In W. Trevathan, E. O. Smith, & J. J. McKenna (Eds.), *Evolutionary medicine and health: New perspectives* (pp. 325–349). New York: Oxford University Press.
- Kuzawa, C. W., & Quinn, E. A. (2009). Developmental origins of adult function and health: Evolutionary hypotheses. *Annual Review of Anthropology*, 38, 131–147. doi:10.1146/annurev-anthro-091908-164350
- Laurent, H. K., Leve, L. D., Neiderhiser, J. M., Natsuaki, M. N., Shaw, D. S., Harold, G. T., & Reiss, D. (2013). Effects of prenatal and postnatal parent depressive symptoms on adopted child HPA regulation: Independent and moderated influences. *Developmental Psychology*, 49, 876–886. doi:10.1037/a0028800
- Lester, B. M., Conradt, E., & Marsit, C. J. (2013). Epigenetic basis for the development of depression in children. *Clinical Obstetrics and Gynecology*, 56, 556–565. doi:10.1097/GRF.0b013e318299d2a8
- Lester, B. M., Conradt, E., & Marsit, C. J. (2014). Are epigenetic changes in the intrauterine environment related to newborn neurobehavior? *Epigenomics*, 6, 175–178. doi:10.2217/epi.14.9
- Leve, L. D., Neiderhiser, J. M., Ge, X., Scaramella, L. V., Conger, R. D., Reid, J. B., . . . Reiss, D. (2007). The Early Growth and Development Study: A prospective adoption design. *Twin Research and Human Genetics*, 10, 84–95. doi:10.1375/twin.10.1.84
- Lumey, L., Stein, A. D., Kahn, H. S., & Romijn, J. (2009). Lipid profiles in middle-aged men and women after famine exposure during gestation: The Dutch Hunger Winter Families Study. *American Journal of Clinical Nutrition*, 89, 1737–1743. doi:10.3945/ajcn.2008.27038
- McCrary, E., De Brito, S. A., & Viding, E. (2010). Research Review: The neurobiology and genetics of maltreatment and adversity. *Journal of Child Psychology and Psychiatry*, 51, 1079–1095. doi:10.1111/j.1469-7610.2010.02271.x
- McDade, T. W., Ryan, C., Jones, M., MacIsaac, J., Morin, A., Meyer, J., . . . Kuzawa, C. (2017). Social and physical environments early in development predict DNA methylation of inflammatory genes in young adulthood. *Proceedings of the National Academy of Sciences*, 114, 7611–7616.
- McEwen, B. S. (1998). Stress, adaptation, and disease: Allostasis and allostatic load. *Annals of the New York Academy of Sciences*, 840, 33–44. doi:10.1111/j.1749-6632.1998.tb09546.x
- Meaney, M. J. (2010). Epigenetics and the biological definition of Gene \times Environment interactions. *Child Development*, 81, 41–79.
- Monk, C., Feng, T., Lee, S., Krupka, I., Champagne, F. A., & Tycko, B. (2016). Distress during pregnancy: Epigenetic regulation of placenta glucocorticoid-related genes and fetal neurobehavior. *American Journal of Psychiatry*, 173, 705–713. doi:10.1176/appi.ajp.2015.15091171

- Monk, C., Spicer, J., & Champagne, F. A. (2012). Linking prenatal maternal adversity to developmental outcomes in infants: The role of epigenetic pathways. *Development and Psychopathology*, *24*, 1361–1376. doi:10.1017/S0954579412000764
- Montirosso, R., Provenzi, L., Fumagalli, M., Sirgiiovanni, I., Giorda, R., Pozzoli, U., . . . Borgatti, R. (2016). Serotonin transporter gene (*SLC6A4*) methylation associates with neonatal intensive care unit stay and 3-month-old temperament in preterm infants. *Child Development*, *87*, 38–48. doi:10.1111/cdev.12492
- Mueller, B. R., & Bale, T. L. (2008). Sex-specific programming of offspring emotionality after stress early in pregnancy. *Journal of Neuroscience*, *28*, 9055–9065. doi:10.1523/JNEUROSCI.1424-08.2008
- Mulvihill, D. (2005). The health impact of childhood trauma: An interdisciplinary review, 1997–2003. *Issues in Comprehensive Pediatric Nursing*, *28*, 115–136. doi:10.1080/01460860590950890
- Murgatroyd, C., Quinn, J. P., Sharp, H. M., Pickles, A., & Hill, J. (2015). Effects of prenatal and postnatal depression, and maternal stroking, at the glucocorticoid receptor gene. *Translational Psychiatry*, *5*, e560. doi:10.1038/tp.2014.140
- Nederhof, E., & Schmidt, M. V. (2012). Mismatch or cumulative stress: Toward an integrated hypothesis of programming effects. *Physiology & Behavior*, *106*, 691–700. doi:10.1016/j.physbeh.2011.12.008
- Nettle, D., Coall, D. A., & Dickins, T. E. (2011). Early-life conditions and age at first pregnancy in British women. *Proceedings of the Royal Society B: Biological Sciences*, *278*, 1721–1727. doi:10.1098/rspb.2010.1726
- Nettle, D., Frankenhuus, W. E., & Rickard, I. J. (2013). The evolution of predictive adaptive responses in human life history. *Proceedings of the Royal Society B: Biological Sciences*, *280*, 20131343–20131343. doi:10.1098/rspb.2013.1343
- Obel, C., Hedegaard, M., Henriksen, T. B., Secher, N. J., Olsen, J., & Levine, S. (2005). Stress and salivary cortisol during pregnancy. *Psychoneuroendocrinology*, *30*, 647–656. doi:10.1016/j.psyneuen.2004.11.006
- Oberlander, T. F., Weinberg, J., Papsdorf, M., Grunau, R., Misri, S., & Devlin, A. M. (2008). Prenatal exposure to maternal depression, neonatal methylation of human glucocorticoid receptor gene (*NR3C1*) and infant cortisol stress responses. *Epigenetics*, *3*, 97–106. doi:10.4161/epi.3.2.6034
- Obradović, J., Bush, N. R., Stamperdahl, J., Adler, N. E., & Boyce, W. T. (2010). Biological sensitivity to context: The interactive effects of stress reactivity and family adversity on socioemotional behavior and school readiness. *Child Development*, *81*, 270–289. doi:10.1111/j.1467-8624.2009.01394.x
- O'Connor, T. G., Heron, J., Golding, J., Glover, V., & ALSPAC Study Team. (2003). Maternal antenatal anxiety and behavioural/emotional problems in children: A test of a programming hypothesis. *Journal of Child Psychology and Psychiatry, and Allied Disciplines*, *44*, 1025–1036.
- O'Connor, T. G., Monk, C., & Fitelson, E. M. (2014). Practitioner review: Maternal mood in pregnancy implications for child psychology and psychiatry. *Journal of Child Psychology and Psychiatry*, *55*, 99–111. doi:10.1111/jcpp.12153
- Ostlund, B. D., Conradt, E., Crowell, S. E., Tyrka, A. R., Marsit, C. J., & Lester, B. M. (2016). Prenatal stress, fearfulness, and the epigenome: Exploratory analysis of sex differences in DNA methylation of the glucocorticoid receptor gene. *Frontiers in Behavioral Neuroscience*, *10*. doi:10.3389/fnbeh.2016.00147
- Painter, R. C., Westendorp, R. G. J., de Rooij, S. R., Osmond, C., Barker, D. J. P., & Roseboom, T. J. (2008). Increased reproductive success of women after prenatal undernutrition. *Human Reproduction (Oxford, England)*, *23*, 2591–2595. doi:10.1093/humrep/den274
- Parade, S. H., Ridout, K. K., Seifer, R., Armstrong, D. A., Marsit, C. J., McWilliams, M. A., & Tyrka, A. R. (2016). Methylation of the glucocorticoid receptor gene promoter in preschoolers: Links with internalizing behavior problems. *Child Development*, *87*, 86–97. doi:10.1111/cdev.12484
- Parent, J., Parade, S., Laumann, L., Ridout, K. K., Yang, B., Marsit, C. J., . . . Tyrka, A. R. (2017). Dynamic stress-related epigenetic regulation of the glucocorticoid receptor gene promoter during early development: The role of child maltreatment. *Development and Psychopathology*. Advance online publication.
- Perry, B. D. (2008). Child maltreatment: A neurodevelopmental perspective on the role of trauma and neglect in psychopathology. In T. Beauchaine & S. P. Hinshaw (Eds.), *Child and adolescent psychopathology* (pp. 93–128). Hoboken, NJ: Wiley.
- Pigliucci, M. (2001). *Phenotypic plasticity: Beyond nature and nurture*. Baltimore, MD: Johns Hopkins University Press.
- Pluess, M., & Belsky, J. (2009). Differential susceptibility to rearing experience: The case of childcare. *Journal of Child Psychology and Psychiatry*, *50*, 396–404. doi:10.1111/j.1469-7610.2008.01992.x
- Rabinowitz, J. A., & Drabick, D. A. G. (2017). Do children fare for better and for worse? Associations among child features and parenting with child competence and symptoms. *Developmental Review*. Advance online publication. doi:10.1016/j.dr.2017.03.001
- Raver, C. C., Blair, C., Garrett-Peters, P., & Family Life Project Key Investigators. (2015). Poverty, household chaos, and interparental aggression predict children's ability to recognize and modulate negative emotions. *Development and Psychopathology*, *27*, 695–708. doi:10.1017/S0954579414000935
- Rickard, I. J., Frankenhuus, W. E., & Nettle, D. (2014). Why are childhood family factors associated with timing of maturation? A role for internal prediction. *Perspectives on Psychological Science*, *9*, 3–15. doi:10.1177/1745691613513467
- Roseboom, T. J., van der Meulen, J. H. P., Osmond, C., Barker, D. J. P., Ravelli, A. C. J., Schroeder-Tanka, J. M., . . . Bleker, O. P. (2000). Coronary heart disease after prenatal exposure to the Dutch famine, 1944–45. *Heart*, *84*, 595. doi:10.1136/heart.84.6.595
- Rutter, M. (2007). Proceeding from observed correlation to causal inference: The use of natural experiments. *Perspectives on Psychological Science*, *2*, 377–395. doi:10.1111/j.1745-6916.2007.00050.x
- Sakhai, S. A., Kriegsfeld, L. J., & Francis, D. D. (2011). Maternal programming of sexual attractivity in female Long Evans rats. *Psychoneuroendocrinology*, *36*, 1217–1225. doi:10.1016/j.psyneuen.2011.02.016
- Sandman, C. A., & Davis, E. P. (2012). Neurobehavioral risk is associated with gestational exposure to stress hormones. *Expert Review of Endocrinology & Metabolism*, *7*, 445–459. doi:10.1586/eem.12.33
- Sandman, C. A., Davis, E. P., Buss, C., & Glynn, L. M. (2012). Exposure to prenatal psychobiological stress exerts programming influences on the mother and her fetus. *Neuroendocrinology*, *95*, 8–21. doi:10.1159/000327017
- Sandman, C. A., Davis, E. P., & Glynn, L. M. (2012). Prescient human fetuses thrive. *Psychological Science*, *23*, 93–100. doi:10.1177/0956797611422073
- Sandman, C. A., Glynn, L. M., & Davis, E. P. (2016). Neurobehavioral consequences of fetal exposure to gestational stress. In N. Reissland & B. S. Kisilevsky (Eds.), *Fetal development* (pp. 229–265). Berlin: Springer.
- Sandman, C. A., Wadhwa, P., Glynn, L., Chicz-Demet, A., Porto, M., & Garite, T. J. (1999). Corticotrophin-releasing hormone and fetal responses in human pregnancy. *Annals of the New York Academy of Sciences*, *897*, 66–75. doi:10.1111/j.1749-6632.1999.tb07879.x
- Scheper-Hughes, N. (1985). Culture, scarcity, and maternal thinking: Maternal detachment and infant survival in a Brazilian shantytown. *Ethos*, *13*, 291–317. doi:10.1525/eth.1985.13.4.02a00010
- Seifer, R., Sameroff, A. J., Dickstein, S., Gitner, G., Miller, I., Rasmussen, S., & Hayden, L. C. (1996). Parental psychopathology, multiple contextual risks, and one-year outcomes in children. *Journal of Clinical Child Psychology*, *25*, 423–435. doi:10.1207/s15374424jccp2504_7
- Sharma, A. (2017). Transgenerational epigenetics: Integrating soma to germline communication with gametic inheritance. *Mechanisms of Aging and Development*, *163*, 15–22. doi:10.1016/j.mad.2016.12.015
- Sheriff, M. J., & Love, O. P. (2013). Determining the adaptive potential of maternal stress. *Ecology Letters*, *16*, 271–280. doi:10.1111/ele.12042
- Shonkoff, J. P., Boyce, W. T., & McEwen, B. S. (2009). Neuroscience, molecular biology, and the childhood roots of health disparities: Building a new framework for health promotion and disease prevention. *Journal of the American Medical Association*, *301*, 2252. doi:10.1001/jama.2009.754
- Shonkoff, J. P., & Garner, A. S., The Committee on Psychosocial Aspects of Child and Family Health, Committee on Early Childhood, Adoption, and Dependent Care, and Section on Developmental and Behavioral Pediatrics, Siegel B. S., Dobbins M. I., Earls M. F., . . . Wood D. L. (2012). The lifelong effects of early childhood adversity and toxic stress. *Pediatrics*, *129*, e232–e246. doi:10.1542/peds.2011-2663
- Slagt, M., Dubas, J. S., Deković, M., & van Aken, M. A. G. (2016). Differences in sensitivity to parenting depending on child temperament: A meta-analysis. *Psychological Bulletin*, *142*, 1068–1110. doi:10.1037/bul0000061

- Smith, C. A. (1947). Effects of maternal undernutrition upon the newborn infant in Holland (1944–1945). *Journal of Pediatrics*, *30*, 229–243. doi:10.1016/S0022-3476(47)80158-1
- Smith, J. W., Seckl, J. R., Evans, A. T., Costall, B., & Smythe, J. W. (2004). Gestational stress induces post-partum depression-like behaviour and alters maternal care in rats. *Psychoneuroendocrinology*, *29*, 227–244.
- Stein, A. D. (2004). Intrauterine famine exposure and body proportions at birth: The Dutch Hunger Winter. *International Journal of Epidemiology*, *33*, 831–836. doi:10.1093/ije/dyh083
- Stein, A. D., Zybert, P. A., van de Bor, M., & Lumey, L. H. (2004). Intrauterine famine exposure and body proportions at birth: The Dutch Hunger Winter. *International Journal of Epidemiology*, *33*, 831–836. doi:10.1093/ije/dyh083
- Tobi, E. W., Lumey, L. H., Talens, R. P., Kremer, D., Putter, H., Stein, A. D., . . . Heijmans, B. T. (2009). DNA methylation differences after exposure to prenatal famine are common and timing- and sex-specific. *Human Molecular Genetics*, *18*, 4046–4053. doi:10.1093/hmg/ddp353
- Turecki, G., & Meaney, M. J. (2016). Effects of the social environment and stress on glucocorticoid receptor gene methylation: A systematic review. *Biological Psychiatry*, *79*, 87–96. doi:10.1016/j.biopsych.2014.11.022
- Volk, A. A., & Atkinson, J. A. (2013). Infant and child death in the human environment of evolutionary adaptation. *Evolution and Human Behavior*, *34*, 182–192. doi:10.1016/j.evolhumbehav.2012.11.007
- Wadhwa, P. D., Glynn, L., Hobel, C. J., Garite, T. J., Porto, M., Chicz-De-Met, A., . . . Sandman, C. A. (2002). Behavioral perinatology: Biobehavioral processes in human fetal development. *Regulatory Peptides*, *108*, 149–157.
- Weaver, I. C. G., Cervoni, N., Champagne, F. A., D'Alessio, A. C., Sharma, S., Seckl, J. R., . . . Meaney, M. J. (2004). Epigenetic programming by maternal behavior. *Nature Neuroscience*, *7*, 847–854. doi:10.1038/nn1276
- Wells, J. C. K. (2003). The thrifty phenotype hypothesis: Thrifty offspring or thrifty mother? *Journal of Theoretical Biology*, *221*, 143–161. doi:10.1006/jtbi.2003.3183
- Wells, J. C. K., & Johnstone, R. A. (2017). Modeling developmental plasticity in human growth: Buffering the past or predicting the future? In G. Jasienska, D. S. Sherry, & D. J. Holmes (Eds.), *The arc of life* (pp. 21–39). New York: Springer.
- West, E. H., Hark, L., & Catalano, P. M. (2017). Nutrition during pregnancy. In S. G. Gabbe (Ed.), *Obstetrics: Normal and problem pregnancies* (7th ed.). Philadelphia, PA: Elsevier.
- West-Eberhard, M. J. (2003). *Developmental plasticity and evolution*. New York: Oxford University Press.
- Wray, N. R., Lee, S. H., Mehta, D., Vinkhuyzen, A. A. E., Dudbridge, F., & Middeldorp, C. M. (2014). Research Review: Polygenic methods and their application to psychiatric traits. *Journal of Child Psychology and Psychiatry*, *55*, 1068–1087. doi:10.1111/jcpp.12295