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Increased expression of bronchial epithelial transient receptor potential vanilloid 1 channels in severe asthma

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Abstract: Background: The airway epithelium is exposed to a range of physical and chemical irritants in the environment which are known to trigger asthma. Transient Receptor Potential (TRP) cation channels play a central role in sensory responses to noxious physical and chemical stimuli. Recent genetic evidence suggests an involvement of TRPV1, one member of the vanilloid subfamily of TRP channels, in the pathophysiology of asthma. The functional expression of TRPV1 on airway epithelium has yet to be elucidated.

Objective: In this study we examined the molecular, functional and immunohistochemical expression of TRPV1 in the asthmatic and healthy airway.

Methods: Bronchial biopsies and bronchial brushings were obtained from healthy volunteers (n=18), mild to moderate asthmatics (n=24) and refractory asthmatics (n=22). To investigate the functional role of TRPV1, cultured primary bronchial epithelial cells (PBEC) from mild asthmatics (n=4), non asthmatic coughers (n=4) and healthy subjects (n=4) were studied.

Results: Quantitative immunohistochemistry revealed significantly more TRPV1 expression in asthmatics compared to healthy subjects and with greatest expression in the refractory asthmatics (p=0.001). PCR and Western blotting analysis confirmed gene and protein expression of TRPV1 in cultured primary bronchial epithelial cells (PBEC). Patch-clamp electrophysiology directly confirmed functional TRPV1 expression in all three groups. In functional assays, the TRPV1 agonist capsaicin induced dose dependent interleukin-8 (IL-8) release which could be blocked by the antagonist capsazepine. Reduction of external pH from 7.4 to 6.4 activated a capsazepine-sensitive outwardly-rectifying membrane current.

Conclusions: Functional TRPV1 channels are present in the human airway epithelium and are over-expressed in the airways of refractory asthmatics. These channels may represent a novel therapeutic target for uncontrolled asthma.

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To The Editors
Journal of Allergy and Clinical Immunology

15th August 2013

Dear Editors,

Many thanks for the opportunity to respond to the reviewers comments and to submit a revised manuscript. We found each of the comments extremely helpful and based on these have undertaken a number of additional experiments which we and believe have resulted in an improved manuscript. We have provided our response to the specific comments from the Editor and each of the reviewers' comments below and indicated where changes have been made to the manuscript

SPECIFIC COMMENTS FROM THE EDITOR:

Although your submission is intriguing, there are issues regarding specificity of the cell types studied, sample size for some of the assessments, and examination of the impact of leukotrienes; these must be addressed.

RESPONSE: We are pleased that the Editor finds our manuscript intriguing. We have undertaken additional experiments to address the issue regarding cell specificity and addressed the important issue of studying more specific environmental irritants in particular the effect of acidic pH. We have also considered and responded to the comment regarding leukotrienes. We have also reduced the length of article to meet with the request to restrict length to 7 pages and reduced the figure legend word count where possible to fewer than 60 words.

COMMENTS FROM REVIEWER #1:

General Comments

I thought that the methodology was sound in this manuscript. My major questions center on following numbers of subjects done for each assay, which made the manuscript more difficult to follow.

RESPONSE: We thank the reviewer for this positive comment and we are pleased they believe our methodology to be 'sound'. We have carefully addressed each of the specific comments below to provide clarification on 'number of subjects done for each assay'.

Specific Comments

- 1) It is unclear why there are two separate cohorts of patients---the Belfast and Leicester. I was unclear how many from each group had IHC and gene expression assays. Did the same subjects have microarray, qPCR and IHC, or did only the Leicester cohort have microarray and the IHC

was done from subjects from Belfast---this was difficult to follow in the manuscript.

RESPONSE: We apologise for the lack of clarity. IHC was performed on the Belfast Cohort only and the microarray and qPCR was performed only on the freshly isolated brushings from Leicester cohort. We have now clarified this in our methods section and have indicated the number of subjects (see methods section revised manuscript (Marked), page 8, line 17-20)

- 2) How were subjects with possible COPD excluded? In table 1, the pack year range was 1-20 years. Or does it not matter that this group with severe asthma may have included not "pure asthma."

RESPONSE: Data from this cohort has been published previously. (Shikotra A et al. Increased expression of immunoreactive thymic stromal lymphopoietin in patients with severe asthma. *J Allergy Clin Immunol.* 2012 Jan;129(1):104-11 and Butler CA et al. Glucocorticoid receptor β and histone deacetylase 1 and 2 expression in the airways of severe asthma. *Thorax.* 2012 May;67(5):392-8). A great strength of the cohort is that the patients are well characterised in specialist centres with regard to diagnosis, physiology and adherence with therapy and all fulfil the definition of severe refractory asthma. The median pack year history is only 2.5 years and in an analysis of ex-smokers, we have also recently demonstrated that ex-smokers with a <10 pack year history have broadly similar demographic and inflammatory profiles to never smokers with severe asthma (Thomson NC et al. Clinical outcomes and inflammatory biomarkers in current smokers and ex-smokers with severe asthma. *J Allergy Clin Immunol.* 2013 Apr;131(4):1008-16) and so we believe that the smoking history does not affect the conclusions from this study.

- 3) Were subjects in Table 2 also included in Table 1? Table 2 also should read "mild asthma" instead of cough variant asthma to match the text. Also, why are the FEV₁ and the FEV₁/FVC lowest in the non-asthmatic coughers in Table 2?

RESPONSE: No, the subjects in table 2 were recruited from a specialist cough clinic and are not included in table 1. We have adjusted the text in table 2 as recommended by this reviewer. Although the absolute FEV₁ values are lower in non-asthmatic coughers the % predicted values are in fact higher. The FEV₁/FVC ratios are lower in this group although still within normal limits and not likely to be of clinical importance

- 4) Why does Figure 1A only have a representative for normal subjects? Is the conclusion that TRPV1 also found in normal subjects? You have data from qPCR that TRPV1 does not differ from normals and asthmatics.

RESPONSE: We undertook the experiments (represented in figure 1) to first establish that we could detect the presence of mRNA for the TRPV family of channels including TRPV1 in human bronchial epithelium. Once this was confirmed we undertook the subsequent sequence of experiments to examine expression of TRPV1 at protein level and its function and potential relevance to the asthmatic airway. We included this in the original manuscript to provide some chronology to our sequence of experiments. However we would be happy to remove this from the revised manuscript if this is felt to be preferable.

- 5) What is the significance of the difference of TRPV1 expression by IHC/microarray (i.e., increased with more severe asthma) and then no difference between groups in relationship to function.

RESPONSE: We thank the reviewer for pointing out this interesting observation. The functional findings described in this manuscript (i.e. IL-8 release in response to TRPV1 activation by capsaicin) represent just one consequence of TRPV1 activation. The lack of difference may reflect sample size and we have commented on this issue in our revised marked manuscript (page 14, line 11)

COMMENTS FROM REVIEWER #2:

The expression of TRPV1, a sensory cation channel was investigated in the airway epithelium of asthmatic patients. Quantitative assessment by immunohistochemistry showed significantly more TRPV1 expression in the epithelium of asthmatics when compared with healthy controls, with the highest expression measured in a group designated as "refractory asthmatics" by the authors. In vitro stimulation of TRPV1 by capsaicin induced IL-8 expression in epithelial cell cultures that was blocked by the antagonist capsazepine. The authors concluded that TRPV1 channels may represent a novel therapeutic target for uncontrolled asthma.

General Comments

1. While the study investigates an interesting question in regards to the function of TRPV1 on epithelial cells in the airways of asthmatic patients, the relevance of the demonstrated findings remain unclear and the conclusion is over stated. The authors aimed to investigate the effects of environmental irritants on TRPV1, but only capsaicin was used, which is not really an environmental irritant. It would support the claims if the effects of pH and temperature changes were studied in addition to capsaicin in epithelial cells.

RESPONSE: We thank the reviewer for this very helpful comment. Based on these comments we have undertaken a series of additional experiments on our primary bronchial epithelial cells to evaluate the effect of lowering pH from 7.4 to 6.4 on bronchial epithelial cell membrane currents. We observed large outwardly rectifying currents with typical TRPV1 signature at pH 6.4 which were abrogated with the TRPV1 antagonist capsazepine (see figures 1 and 2 below). This important finding now provides evidence of a physiologically relevant environmental irritant (i.e. acid pH) activating membrane bound TRPV1 channels on bronchial epithelial cells and is very supportive of our original hypothesis. In our revised manuscript we have detailed these additional experiments, described the results and commented on their significance within the discussion

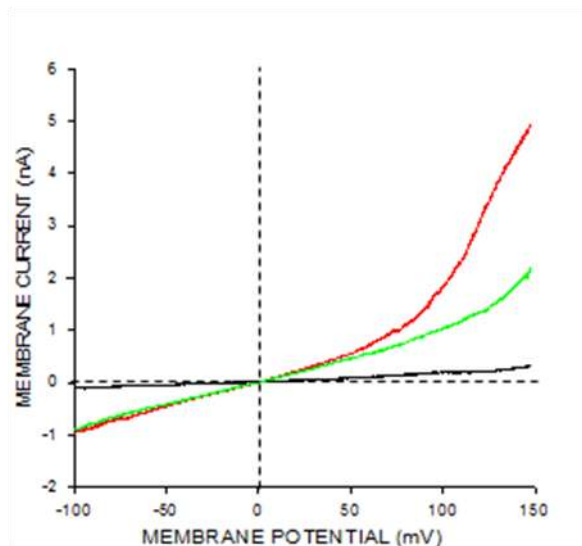


FIGURE 1

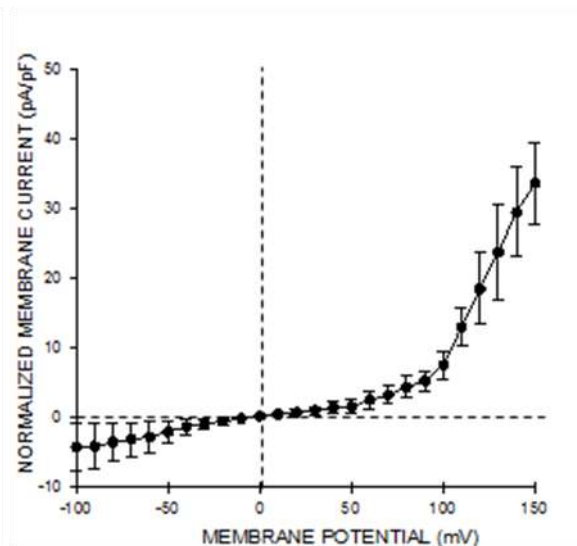


FIGURE 2

2. The rationale and significance of the epithelial cells as opposed to other structural cells of the airways such as fibroblasts and smooth muscle cells is not supported convincingly. In their introduction the authors are creating the impression that coughing and wheezing to environmental irritants maybe due to TRPV expression on the epithelial cells. These responses are in fact directly due to activation of sensory neurons that lead to bronchospasm and coughing/wheezing. This needs to be clarified.

RESPONSE: This is an important point raised by the reviewer. They are correct in pointing out that protective/irritant airway responses such as bronchospasm and cough are directly due to neuronal activation. However there is evidence that neuronal airway responses may be sensitized in disease such as asthma and we propose that activation of TRPV1 on non-neuronal cells (in this case bronchial epithelial cells) by irritants with subsequent release of sensitizing mediators represents one possible mechanism as to how this may arise. We have now revised our introduction to clarify this (see revised marked manuscript page 8, lines 1-4)

Specific Comments

3. What is the definition of "refractory asthmatics"? This is an important group of patients, and how it was defined needs to be spelled out more clearly.

RESPONSE: As stated in the manuscript '...participants with refractory asthma were recruited from the Belfast City Hospital. All had persisting symptoms despite treatment at Step 4 or 5 of the Global Initiative for Asthma (GINA) guidelines...'. We have now added the following '.....fulfilled the ATS definition of refractory asthma (reference - Proceedings of the ATS workshop on refractory asthma. Current understanding, recommendations, and unanswered questions. American Journal of Respiratory and Critical Care Medicine. 2000;162(6):2341-51)

4. Was there any statistical correlation between asthma severity and the expression of TRPV1 (either in the mRNA or the protein expression?)

RESPONSE: Although epithelial TRPV1 expression in the airway biopsies were significantly greater in asthma compared with health we found no statistical

correlation between disease severity either defined by lung function, inhaled steroid dose (BDP equivalent) or oral steroid dose.

5. Figure 1A: The mRNA bands appear at different levels, what is the explanation for that? Why is there no band at V5? What are the V1-V6 designations?

RESPONSE: We apologise for not clarifying this and have now revised the legend accompanying figure 1. The V1-V6 designations represent the sub types of the TRPV (vanilloid) family, hence slightly different band levels. TRPV1 was the focus of interest in this manuscript but others (although not TRPV5) appear to be expressed in the samples we studied.

6. Figure 1B: It appears that in lane 2 the 95kDa band is missing or is very faintly expressed. What is the explanation?

RESPONSE: We believe the differences in band intensity represent differential expression of protein levels between samples.

7. Figure 1C: In the microarray what was the control that the expression level is standardized to? It appears that all of the expression is below 1.

RESPONSE: The gene expression microarrays were conducted as described previously in *J Immunol* 2011;186:1861-1869. In brief, two color Agilent Whole Human Genome microarrays using a common reference design (utilizing Universal Human Reference RNA, Stratagene, La Jolla, CA for the reference channel) were employed in this study. Gene expression was calculated as the log₂ ratio of test and reference channel values. The reference sample facilitates array to array normalization as a technical control. As a result, the expression values (log₂ ratios) for a given gene will be arbitrarily offset based on the relative abundance of mRNA in the Human Reference RNA sample as compared to the test samples.

8. In addition to the epithelium in Figures 2 and 3 there appears to be significant TRPV1 positivity in the submucosal area. This ought to be investigated by the authors, especially in regards to the proinflammatory significance as fibroblasts and smooth muscle cells are a prominent source of IL-8 and similar proinflammatory chemokines.

RESPONSE: This is an important comment from the reviewer. We agree that the role of other non-neuronal cells (e.g. smooth muscle cells) may play a role in TRPV1 mediated inflammatory response. Indeed we have undertaken some preliminary experiments which provides evidence to support this (please see our response below to the reviewers next comment). However we are keen to confine the focus of this current manuscript to the expression and function of TRPV1 in bronchial epithelial cells. We believe our findings in relation to the bronchial epithelium represent an interesting finding. We believe our preliminary experiments in primary smooth muscle cells will form the basis for future work. However if the Editor feels this would improve the manuscript we would be provide this preliminary data.

9. Along the same line, it would be imperative to add experiments in which cultured fibroblast and smooth muscle cells are also investigated for their membrane potential and their expression of IL-8 in response to capsaicin stimulation.

RESPONSE: We have undertaken some preliminary experiments to confirm that primary human airway smooth muscle cells (ASM) are activated by capsaicin to release IL-8 and this can be almost completely inhibited by the TRPV1 antagonist capsazepine. ASM cells were treated for 24 hours with 10, 25 or 50 μM capsaicin or control medium. Supernatants were collected after 24 h and ELISA was performed to measure the release of IL-8. (see figure 3 below). As indicated in our response above we believe these findings require an extensive programme of work which is outside the focus of our current manuscript.

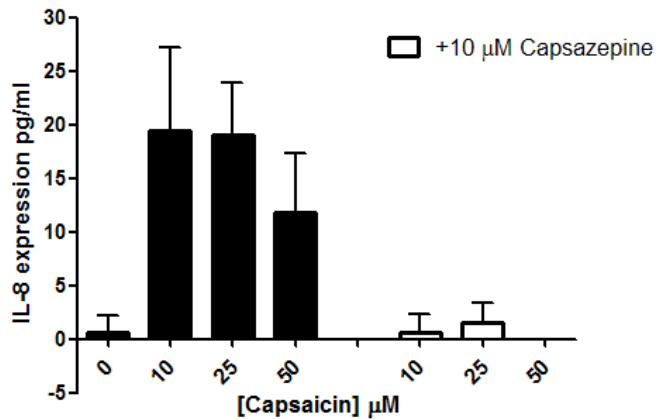


Figure3 Capsaicin induced IL-8 release from primary human airway smooth muscle cells

10. The intracytoplasmic expression of the TRPV1 receptor expression is interesting and needs further experimental support. For example, co-localization with endoplasmic reticulum markers would provide additional support. The functional significance of this TRPV1 as opposed to the membrane expressed molecule needs to be investigated.

RESPONSE: As suggested by the reviewer we have now undertaken some additional immunolabelling experiments demonstrating that TRPV1 does co-localise with ER markers in bronchial epithelial cells (see figure 4 below). We have included this in our results (page 12, line 15-19) and discussion (page 16, line 7-10) section of our revised manuscript. These channels may play a role in ER- Ca^{2+} release, but to fully establish this would require an extensive programme of work involving the development of routine methods for measuring cytoplasmic and ER Ca^{2+} in PBECs as well as patch-clamp recording of ER membranes from these cells. Whilst such studies obviously lie beyond the scope and focus of the current work, we believe that our findings do provide an important basis for such studies in the future.

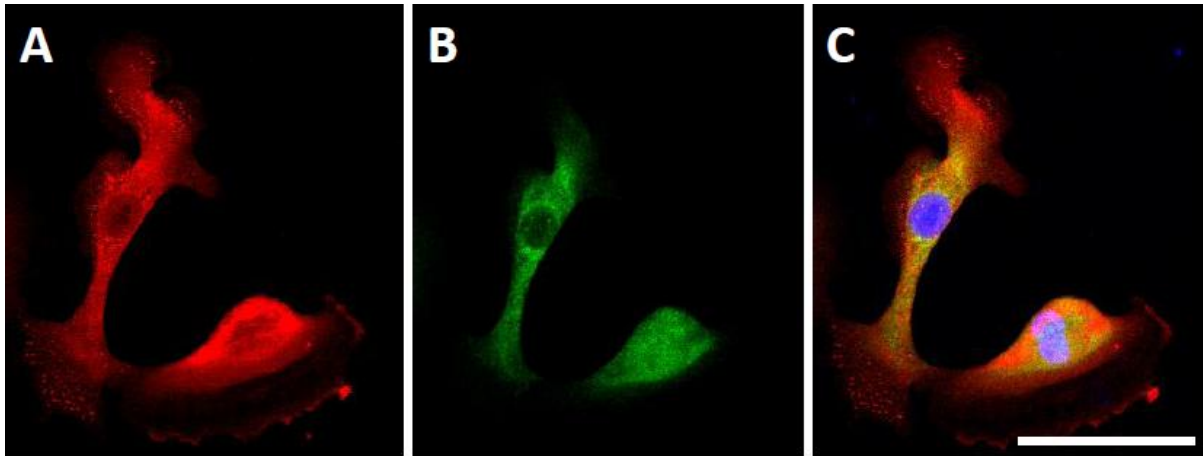


Figure 4: TRPV1 and ER colocalisation in human bronchial epithelial cells. Representative confocal microscopy images (A-C) showing immunohistochemistry staining of bronchial epithelial cells positive for TRPV1 (A) and ER (B) markers. Co-localisation was evident when the images were merged (C; orange/yellow). DAPI stained blue nuclei (C). Scale bar = 100 μ m.

COMMENTS FROM REVIEWER #3:

This paper presents strong evidence for the presence of the TRPV1 receptor in airway epithelium and suggestive evidence of the increase in its expression in asthma, particularly in severe asthma.

RESPONSE: We are delighted this reviewer believes our paper provides ‘strong evidence for the presence of the TRPV1 receptor in airway epithelium’. We have addressed each of their comments below.

My Comments

1. The subjects with severe asthma were older, had higher IgE levels, and had a greater smoking history than others studied. Do the authors have any data on non-asthmatic smokers to separate this as a potential cause of the increase identified?

RESPONSE: The age and demographic features of the severe refractory group is very typical of this group (Heaney LG et al. Refractory asthma in the UK: cross-sectional findings from a UK multicentre registry. *Thorax*. 2010 Sep;65(9):787-94). As discussed above, we have also recently published a detailed analysis on smokers, never smokers and ex-smokers with severe asthma and some of the features alluded to by the Reviewer are again typical of this more severe group (Thomson NC et al. Clinical outcomes and inflammatory biomarkers in current smokers and ex-smokers with severe asthma. *J Allergy Clin Immunol*. 2013 Apr;131(4):1008-16). It is an intriguing possibility that that ex-smoking may potentially contribute to some of the ‘steroid resistant’ mechanisms, which defines this severe population and we discuss this in our previous manuscript. However, we would argue that whatever the mechanism, the fact that our patients are precisely characterised in this cohort with regard to diagnosis, physiology and adherence with asthma therapy does not alter the significance of the findings.

2. The authors comment that TRPV1 receptors were identified in submucosal cells. To what degree could these cells have contributed to their

findings of increased numbers of receptors and to increased activity in asthma?

RESPONSE: This is an important point raised by this reviewer. It is likely that a number of other non-neuronal cells in addition to bronchial epithelial cells express functional TRPV1 receptors. In our response to one of the other reviewers (Reviewer #2) comments above, we have provided some preliminary evidence that this is the case for primary airway smooth muscle cells. However we believe this is outside of the scope of the current manuscript.

3. The authors state that TRPV1 receptors respond to LTB4. It would be of interest to know whether other leukotriene mediators share this property as it would further the link between expression of asthma and the function of these receptors.

RESPONSE: There is no current evidence that other leukotriene mediators (e.g. LTD4 or LTE4) directly activate TRPV1 in the same fashion as LTB4. However it is clear that other products of lipoxygenase (e.g. 15(s)-HETE) are potent TRPV1 agonists and as such represent additional mechanisms whereby inflammatory mediators relevant to asthma may modulate TRPV1 function.

We hope this response and revised manuscript addresses the issues raised during the review process.

With best wishes

Yours Sincerely

Lorcan McGarvey (on behalf of the authors)

1 **Increased expression of bronchial epithelial transient receptor potential vanilloid 1**
2 **channels in severe asthma**

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6 | **ABSTRACT (250 words)**

7 | **Background:** The airway epithelium is exposed to a range of physical and chemical irritants
8 | in the environment which are known to trigger asthma. Transient Receptor Potential (TRP)
9 | cation channels play a central role in sensory responses to noxious physical and chemical
10 | stimuli. Recent genetic evidence suggests an involvement of TRPV1, one member of the
11 | vanilloid subfamily of TRP channels, in the pathophysiology of asthma. The functional
12 | expression of TRPV1 on airway epithelium has yet to be elucidated.

13 | **Objective:** In this study we examined the molecular, functional and immunohistochemical
14 | expression of TRPV1 in the asthmatic and healthy airway.

15 | **Methods:** Bronchial biopsies and bronchial brushings were obtained from healthy
16 | volunteers (n=18), mild to moderate asthmatics (n=24) and refractory asthmatics (n=22). To
17 | investigate the functional role of TRPV1, cultured primary bronchial epithelial cells (PBEC)
18 | from mild asthmatics (n=4), non asthmatic coughers (n=4) and healthy subjects (n=4) were
19 | studied.

20 | **Results:** Quantitative immunohistochemistry revealed significantly more TRPV1 expression
21 | in asthmatics compared to healthy subjects and with greatest expression in the refractory
22 | asthmatics (p=0.001). PCR and Western blotting analysis confirmed gene and protein

1 expression of TRPV1 in cultured primary bronchial epithelial cells (PBEC). Patch-clamp
2 electrophysiology directly confirmed functional TRPV1 expression in all three groups. In
3 functional assays, the TRPV1 agonist capsaicin induced dose dependent interleukin-8 (IL-8)
4 release which could be blocked by the antagonist capsazepine. Reduction of external pH
5 from 7.4 to 6.4 activated a capsazepine-sensitive outwardly-rectifying membrane current.

6

7 **Conclusions:** Functional TRPV1 channels are present in the human airway epithelium and
8 are over-expressed in the airways of refractory asthmatics. These channels may represent a
9 novel therapeutic target for uncontrolled asthma.

10

1 **Clinical Implications (30 words)**

2 TRPV1 is a functional channel activated by physical and chemical irritants. We report its
3 overexpression in asthmatic airway epithelium most notably in severe disease suggesting a
4 role in uncontrolled asthma.

5 **Capsule summary (35 words)**

6 The Transient Receptor Potential vanilloid-1 (TRPV1) channel regulates responses to irritant
7 stimuli. We report TRPV1 overexpression in asthmatic airway epithelium most notably in
8 severe disease suggesting its potential as a therapeutic target for uncontrolled asthma.

9

10 **Key words:** ion channel, sensory, asthma, irritant, chemical, exacerbation, cough

11 **Abbreviations:**

12 TRP - Transient Receptor Potential

13 TRPV1 – Transient Receptor Potential Vanilloid-1

14 GINA – Global Initiative for Asthma Guidelines

15 PBEC – Primary bronchial epithelial cell

16 qPCR – Quantitative real-time PCR

17 GMA – glycol methacrylate

18

1 INTRODUCTION

2 During an exacerbation of asthma, the airways become hypersensitive and patients are
3 often troubled with bouts of cough and wheeze following exposure to relatively innocuous
4 stimuli such as aerosols, strong odours or changes in air temperature. The precise
5 mechanism for this airway hypersensitivity is not known but upregulation of receptors
6 responsible for sensing chemical and physical stimuli may be relevant. The recently
7 described Transient Receptor Potential (TRP) proteins, a family of Ca²⁺ permeable non
8 selective cation channels which sense a vast array of chemical and physical stimuli (1, 2), are
9 believed to have an important role in the regulation of airway function in both health and
10 disease (3, 4). Recent data has provided genetic evidence for the involvement of one
11 subfamily member, TRP vanilloid 1 (TRPV1) in asthma pathophysiology (5).

12 TRPV1 is activated by capsaicin (the pungent principle contained in hot chilli peppers) and
13 inhaled capsaicin causes cough (6) and bronchospasm (7) in asthmatics. The airway
14 inflammatory events which accompany an exacerbation of asthma include a rise in airway
15 temperature (8), acidosis (9) and release of mediators derived from lipoxygenase
16 metabolism of arachidonic acid (4, 10). TRPV1 is primarily recognised as a heat thermal
17 sensor, but it is also readily activated by low extracellular pH (11, 12) and lipoxygenase
18 products such as leukotriene B₄. (13). TRPV1 receptors are expressed in several cell types in
19 human airway including sensory neurones (14), smooth muscle cells (15) and epithelial cells
20 from both the upper (16) and lower airway (17). As the bronchial epithelium has an
21 important role in sensing and responding to noxious stimuli we hypothesised that TRPV1 is
22 over expressed in the airway epithelium of asthmatics, in particular those with difficult-to-

1 | ~~treat asthma, and that~~ Further we considered that the inflammatory response following
2 | ~~irritant activation of airway epithelial TRPV1 has a functional role in~~ may contribute to
3 | ~~heighted neuronal reflexes responsible for cough and bronchospasm in asthma, a~~
4 | ~~asthma may pathogenesis and airway responses to environmental irritants.~~

5 | In this study, we examined both TRPV1 expression and function in human bronchial
6 | epithelium. In addition we provide evidence for the overexpression of TRPV1 in the airways
7 | of severe asthmatics compared to mild asthmatics and healthy subjects. Some of the results
8 | of these studies have been reported in abstract form (18).

9 |

10 | **METHODS AND MATERIALS**

11 | *Full methodological details are provided in an online data repository*

12 | *Study Subjects*

13 | Participants with refractory asthma were recruited from the Belfast City Hospital. All had
14 | persisting symptoms despite treatment at Step 4 or 5 of the Global Initiative for Asthma
15 | (GINA) guidelines. Mild to moderate asthmatics had a clinical diagnosis of asthma with a
16 | current history of recurrent wheezing and documented response to asthma medication.

17 | Bronchial biopsy samples from this Belfast cohort were analysed using
18 | immunohistochemistry (IHC). For examination of *in vivo* gene expression of TRPV1 in airway
19 | epithelium an additional cohort of subjects from Leicester were recruited. Demographic
20 | details of both cohorts are shown in Table 1. To investigate the functional role of TRPV1,
21 | primary bronchial epithelial cell (PBEC) cultures were grown from bronchial brushings

1 obtained from mild asthmatics and non-asthmatic patients recruited from a specialist cough
2 clinic (19) and healthy participants recruited by advertisement. Demographic details are
3 shown in Table 2.

4 The study was approved by the appropriate Institutional Research Ethics Committees (details in
5 online data repository). Written informed consent was gained from all participants prior to their
6 involvement.

7

8 *Bronchoscopy and sample processing*

9 All asthmatic participants were clinically stable at the time of bronchoscopy, and had an
10 FEV₁ >60% predicted. Bronchial biopsies and brushings were obtained from segmental
11 airways using a standard techniques. Three biopsies from each participant (Belfast Cohort)
12 were submitted for immunohistochemistry studies. Gene expression microarray analysis
13 (and qPCR) was performed on fresh bronchial brushings (Leicester cohort). To investigate
14 the functional role of TRPV1, PBEC cultures from bronchial brushings were cultured as
15 described previously (20). These were obtained from mild asthmatic (n=4), non asthmatic
16 patients with chronic cough (n=4) and healthy volunteers (n=4).

17 Immunohistochemistry

18 Twenty eight biopsies (10 healthy volunteer, 8 mild asthmatic and 10 severe asthmatic
19 biopsies) were processed for immunohistochemistry (IHC) using local protocols as detailed
20 in the online data repository. [For TRPV1 and ER co-localisation experiments local protocols](#)
21 [were employed as detailed in the online data repository.](#)

22

1 **Gene expression analysis**

2 *Bronchial brushings and PBEC cultures*

3 Real-time PCR (qRT-PCR) analyses were performed using RNA from bronchial brushings using
4 TaqMan® Gene Expression Assays (Applied Biosystems, Foster City, CA) as previously
5 described (21). RNA was amplified (Ambion,) for Agilent (Santa Clara, CA) two color Whole
6 Human Genome (WHG) 4x44k gene expression microarray analysis.

7 PBECs were trypsinised and total RNA extracted using the RNeasy Mini Kit (Qiagen, Hilden,
8 Germany) and qRT-PCR performed using a 7300 Real Time PCR system (Applied Biosystems,
9 Foster City, CA). More information including detail on GAPDH and TRPV1 forward and
10 reverse primers is provided in the online data repository.

11 *Western Blotting*

12 Protein concentration was determined using BCA protein assay kit (Thermo Scientific,
13 Wilmington, USA) and protein expression determined using a rabbit polyclonal TRPV1
14 antibody (ab63083; Abcam, Cambridge, UK), at a dilution of 1:3000 followed by incubation
15 for 1 h with the secondary antibody, HRP-labelled anti-rabbit (1:3000; Bio-Rad Laboratories,
16 Hercules, CA, USA). More details provided in online data repository.

17 *TRPV1 agonists / antagonist experiments in PBECs*

18 Capsaicin is a selective TRPV1 agonist and in this study cultured PBECs were treated with either 10,
19 25 or 50 µM capsaicin or control medium for 24 h and the concentration of IL-8 measured by ELISA
20 (R & D Systems Europe Abingdon, UK). For the antagonist experiments, PBECs were pre-
21 treated for 20 min with 10 µM of the selective TRPV1 antagonist capsazepine.

22 *Patch clamp experiments*

1 Whole-cell currents were recorded using borosilicate heat-polished patch pipettes (2-3 M Ω
2 resistance when filled with the pipette solution) and an Axopatch 200B amplifier (Molecular
3 Devices, Union City, CA, USA) interfaced to Digidata 1322A using the pClamp 9 software
4 (Molecular Devices) as previously described by ourselves (22) and detailed in the online data
5 repository.

6 *Data Analysis*

7 Data are presented as means \pm SEM. For statistical analysis a one way ANOVA was performed
8 with Bonferroni test for multiple comparisons. Kendall's tau b statistical analysis was used to
9 examine the trend in biopsy expression by participant group. Patch clamp data were
10 analysed and plotted using Origin 8 (OriginLab, Northampton, MA, USA). A P value < 0.05
11 was considered significant.

12

13 **RESULTS**

14 **Confirmation of TRPV1 expression in cultured human bronchial epithelial cells.**

15 Gene expression for multiple members of the TRPV family including TRPV1 (but not TRPV5)
16 was identified in PBECs from healthy controls (n=3) (Figure 1A). TRPV1 protein was detected
17 at its expected molecular mass of 95 kDa by Western blotting in the same samples (Figure
18 1B).

19 **Molecular identification and localisation (immunohistochemical expression) of TRPV1 in** 20 **healthy and asthmatic epithelium**

21 *Microarray*

1 Using gene expression microarray analysis of bronchial brush samples from the Leicester
2 cohort we found significantly greater levels of TRPV1 expression (median, IQR) in severe
3 asthmatics (0.673, 0.452-0.761) compared to mild/moderate asthmatics (0.341, 0.167-0.538,
4 $p=0.008$) and healthy volunteers (0.322, 0.155-0.410, $p=0.004$) (Figure 1C).

5 *Quantitative PCR.*

6 TRPV1 mRNA was detected in all of the epithelial brush samples. There was no difference in
7 median (IQR) relative fold expression of TRPV1 mRNA calculated as Delta Delta C_T values
8 between groups: -1.262 (-3.598, 2.518), (-2.523 (-4.040, 1.362) and -1.289 (-4.746, 1.624) in
9 healthy volunteers, mild / moderate asthmatics and severe asthmatics respectively (see
10 Figure E1 in online data repository)

11 *Immunohistochemistry*

12 Immunohistochemistry revealed bronchial epithelial expression of TRPV1 in healthy
13 volunteers, mild/moderate asthmatics and severe asthmatics (Figures 2A, B and C,
14 respectively). Dual labelling immunofluorescence confirmed the expression of TRPV1 on

15 cytokeratin positive epithelial cells (Figures 3 A, B and C). We observed an intracytoplasmic
16 distribution of the TRPV1 and to further characterise this we undertook additional
17 experiments demonstrating immunohistochemistry staining of bronchial epithelial cells
18 positive for TRPV1 (Figure 4A) and ER (Figure 4B) markers. Colocalisation was evident when
19 the images were merged (Figure 4C).

20 We observed ~~There was~~ greater TRPV1 expression in asthmatic epithelium compared to
21 healthy subjects (Figure 45; $p = 0.001$).

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1 **Membrane current responses to TRPV1 agonist capsaicin**

2 Mean membrane capacitance of PBEC was 30.8 ± 4.3 pF (n=6). In unstimulated cells, only
3 small background currents could be recorded in the whole range of membrane potentials
4 tested, from -100 to +140 mV (Figure [5A6A](#), left panel). The I-V relationship of the mean
5 background current normalised by the membrane capacitance to account for some
6 variations in cell size is shown in Figure [5B6B](#). It should be noted that composition of the
7 external and pipette solutions was such that this facilitated isolation of cation currents but
8 precluded activation of any potassium or calcium-dependent conductances. Capsaicin
9 application (50 μ M) caused rapid activation of large outwardly rectifying currents (Figure
10 [5A6A](#), middle panel), which is the characteristic “biophysical” signature of TRPV1 channels.
11 This effect was completely reversible upon capsaicin wash-out (Figure [5A6A](#), right panel).
12 Corresponding I-V relationships are shown in Figure [5C6C](#). The capsaicin-induced currents
13 reversed close to 0 mV indicating their non-selective cation nature.

14 To investigate the time course of these responses, voltage ramps over the same range of
15 potentials were applied at 10 s interval. Figure [6A-7A](#) shows a typical response to 50 μ M
16 capsaicin application consisting of rapid activation and desensitisation in the continuous
17 presence of the agonist, which developed over 2-3 min. The I-V relationships which were
18 measured in this experiment in control, at the peak response and after agonist wash-out are
19 shown in Figure [6B-7B](#) (compare to I-V curves in Fig. 6C measured by voltage steps), while
20 the mean results are summarised in panel C. The increase in current amplitude in response
21 to capsaicin was significant (p=0.007).

22 **Membrane responses to reduction of external pH**

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1 Reduction of external pH from 7.4 to 6.4 activated inward and outward currents in cells from
2 healthy patients which were partially inhibited by the addition of the selective TRPV1
3 antagonist capsazepine (5 μ M; Fig. 8A). The mean capsazepine-sensitive I-V relation (Fig. 8B
4 n=4) was similar in profile to the capsaicin-dependent currents exhibiting strong outward
5 rectification and reversal close to 0 mV.

7 **Functional expression of TRPV1 in cultured human primary bronchial epithelial cells**

8 *Dose-dependent induction of IL-8 release from cultured human PBECs by capsaicin*

9 Dose-dependent induction of IL-8 release by capsaicin was observed within each study
10 group although there was no significant difference in fold increase in IL-8 between asthmatic
11 and non asthmatic groups which may relate to sample size (data not shown). The dose
12 dependent increase in IL-8 release when data from all three groups was therefore pooled
13 has been presented in figure ~~7A9A~~. When compared to basal a marginal 1.25-fold increase in
14 IL-8 release was observed following incubation with 10 μ M capsaicin with significant
15 increases of 1.76-fold ($p < 0.01$) and 2.15 -fold ($p < 0.001$) compared to basal observed
16 following incubation with 25 and 50 μ M doses of capsaicin, respectively.

17 To further test the specificity of the capsaicin effect, we used selective TRPV1 blocker
18 capsazepine. Capsazepine significantly inhibited capsaicin-induced IL-8 release compared to
19 25 μ M capsaicin alone ($p < 0.001$) and compared to 50 μ M capsaicin alone ($p < 0.001$). The
20 data for n=9 individual experiments is shown in Figure ~~7B9B~~.

1

2 **DISCUSSION**

3 In this study we have shown for the first time that TRPV1, which is primarily recognised as
4 the major neuronal hot receptor (23, 24), is expressed and functional in human bronchial
5 epithelium. In primary bronchial epithelial cell cultures we have shown that pharmacological
6 activation of the TRPV1 channel with capsaicin induces IL-8 release which can be attenuated
7 by pre-treatment with the selective TRPV1 antagonist capsazepine.

8 Interestingly, TRPV1 expression in the airway was increased in asthmatic subjects compared
9 to healthy controls with a further increase in subjects with refractory asthma. Thus, over-
10 expression may have a role in symptoms associated with irritant exposure in asthma,
11 particularly in patients with persisting symptoms despite treatment with high dose anti-
12 inflammatory therapy. Understanding the role of TRPV1 in refractory asthma may therefore
13 identify a novel target for anti-inflammatory therapy in more severe disease.

14 We believe that bronchial epithelial TRPV1 channels are ideally positioned to detect
15 chemical and physical irritants (e.g. inflammatory mediators, environmental pollutants, acid
16 pH and changes in temperature) in the airway lumen. To date, most of the focus has been
17 on ~~the role of~~ neuronal TRPV1 in the human airway (14, 25). ~~Direct activation of irritant~~
18 ~~neural receptors mediates reflex responses such as cough and bronchospasm, thus~~
19 ~~protecting the airway from noxious chemical and physical irritants.~~ However, TRPV1
20 expression ~~at both an mRNA and protein level~~ has been reported in a variety of non-
21 neuronal tissues ~~and cell types~~ and therefore we hypothesised that the airway responses to
22 environmental stimuli might in part be regulated by the bronchial epithelial TRPV1

1 receptors. Recent studies in immortalised ~~human~~ airways epithelial cell lines provide some
2 support for this (17, 26) although evidence ~~for this~~ in human disease is lacking. ~~In this~~
3 ~~study~~ Here we ~~have provided~~ provide *in vivo* and *ex vivo* evidence of TRPV1 expression and
4 function in human airway biopsies and cultured primary bronchial epithelial cells with
5 specific relevance to refractory asthma.

6 We observed an intracytoplasmic distribution of the receptor in bronchial epithelial cells
7 ~~which has been as~~ observed in other cell types (27, 28). ~~The intracellular localisation of~~
8 ~~TRPV1 (in particular~~ Our finding of TRPV1 staining in the endoplasmic reticulum ~~) in the~~
9 ~~airway epithelium~~ is consistent with its functional role as a Ca²⁺ store release channel
10 ~~regulating intracellular Ca²⁺ homeostasis~~. Robust membrane current responses to the TRPV1
11 agonist capsaicin (Figs. 5 and 6) suggest strongly that plasmalemmal TRPV1 is also expressed
12 and functional in PBEC, ~~although the precise location and function of bronchial epithelial cell~~
13 ~~membrane and intracellular TRPV1 requires further study~~. We also observed TRPV1 staining
14 on a number of submucosal cells although the focus of this present study is confined to the
15 bronchial epithelial expression. Previous studies from chronic coughers (14) and healthy
16 subjects (15) have reported only sparse TRPV1 expression in the airway epithelium, although
17 greater expression ~~has been reported~~ in the ~~human~~ upper airway (29). ~~We suggest~~
18 ~~d~~ Differences in tissue processing, antibodies used and clinical phenotype may explain the
19 discrepancy. ~~To overcome issues regarding specificity of antibodies we~~ In our studies to
20 evaluated the functional properties of TRPV1 on bronchial epithelium. ~~In these experiments~~
21 we saw that ~~TRPV1 on~~ PBECs responded to the TRPV1 agonist capsaicin with the release of
22 pro-inflammatory mediators ~~and that this~~ which was blocked by capsazepine, a TRPV1
23 selective antagonist. ~~We chose to measure~~ Ccapsaicin induced IL-8 release ~~from bronchial~~

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1 ~~epithelial cells as this had~~has been previously observed in both human esophageal (30) and
2 human corneal epithelial cells (31) but not ~~before~~ in ~~cultured primary bronchial epithelial~~
3 ~~cultures~~PBECs. ~~In addition,~~As IL-8 is known to induce and maintain neural sensitization ~~and~~
4 ~~activation of nociceptors~~ (32, 33). ~~Therefore,~~our data provide a mechanistic link whereby
5 ~~activation of~~ activation of bronchial epithelial TRPV1 ~~receptors~~ by environmental irritants or
6 changes in airway pH or temperature (during exacerbations) may sensitize airway sensory
7 neurones and contribute to the airway and cough hypersensitivity that characterises asthma
8 ~~through the cross-talk between epithelial and neuronal TRPV1.~~

9 Studies in immortalised respiratory cell lines suggest ~~A~~airway epithelial TRPV1 ~~has been~~
10 ~~shown to~~ regulates cell apoptosis in response to environmental irritants ~~such as particulate~~
11 ~~matter and ash~~ (34). ~~These have been confined to immortalised respiratory cell lines~~
12 ~~whereas our findings~~Here, we provide the first evidence of a functional airway epithelial
13 TRPV1 channel ~~function~~ in primary cells ~~from the airway. We also provide further evidence~~
14 ~~of TRPV1 channel function~~ in our the whole-cell patch clamp experiments on PBECs our.
15 ~~These~~ results provided ~~a~~ direct evidence ~~of for~~ the presence of functional TRPV1 in the
16 plasma membrane that can ~~thus~~ mediate membrane depolarisation and calcium influx;
17 steps which are necessary for release of pro-inflammatory mediators (35). Both kinetics and
18 voltage-dependent properties of capsaicin-induced currents in PBEC are consistent with
19 TRPV1 biophysical features, such as slow activation/deactivation and strong outward
20 rectification.

21 ~~Asthmatics commonly report symptoms triggered by a variety of chemical and physical~~
22 ~~irritants and because~~ TRP channels have the capacity to sense a vast range of stimuli, and
23 their role in airways disease is currently under intense investigation (1). In our

1 immunohistochemical studies we have shown that TRPV1 is over expressed in the bronchial
2 epithelium of asthmatics compared to healthy subjects and patients with refractory asthma
3 express more epithelial TRPV1 than those with mild/moderate asthma. This was supported
4 by our microarray analysis ~~demonstrating~~ confirming increased expression of TRPV1 genes
5 in the more severe asthmatics. Although on quantitative PCR, mRNA was not concordant
6 with protein abundance, it has become increasingly clear that at the genomic level, mRNA
7 expression does not consistently predict corresponding protein levels (36, 37). This may
8 because of complex post-transcriptional and translational modification and /or that different
9 biological mRNA and protein degradation rates may affect correlation. Post translational
10 modification of TRPV1 has been described in experimentally induced tissue injury (38) and
11 therefore may occur in the inflamed asthmatic airway.

12 While the vast majority of asthmatics respond to standard doses of current treatments (in
13 particular corticosteroids), a proportion remain symptomatic despite treatment at GINA
14 Step 4/5 and are considered to have difficult or treatment refractory asthma. Our finding
15 that bronchial epithelial TRPV1 is over expressed ~~in human bronchial epithelium and~~
16 ~~upregulated~~ in refractory (steroid-resistant) asthma suggests ~~that it its activation~~ may
17 represent a novel pro-inflammatory pathway. Clinical trials of TRP receptor antagonists ~~in~~
18 ~~man~~ have been undertaken in other abnormal hypersensitive states including neuropathic
19 pain and these drugs may prove effective in improving symptom control in asthma as well.

20 Our findings suggest that the bronchial epithelium may sense and respond to environmental
21 stimuli via TRPV1 activation. However it is not clear how bronchial epithelial cells signal to
22 afferent sensory nerves. We suggest that release of sensitizing mediators such as IL-8 may
23 be one such mechanism. Such mediators may enhance and augment neurogenic

1 inflammation and nociceptive signalling. Therefore, pharmacological modulation of this
2 receptor may be useful in the treatment of uncontrolled asthma.

3

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9 **Author contributions:** CB, SA and SMcQ performed immunostaining, analysed the data and
10 contributed to the writing of the manuscript. SS and HA performed cell culture, molecular analysis of
11 samples and subsequent ELISAs. DC and JA performed and analysed the gene expression data,
12 analysed the data and contributed to the writing of the manuscript. CB, LH and PB recruited subjects,
13 performed bronchoscopy on the asthmatic subjects. LMG and LP recruited subjects for bronchial
14 brush sampling and with TJW performed the bronchoscopies. LH supervised CB and LMG and RC
15 supervised LP. AZ and MMcG performed and analysed patch-clamp experiments, ~~and with~~ LMG, AZ
16 and ME supervised SS. LMG and LH conceived and designed the research, contributed to data
17 analysis and interpretation and drafted the original manuscript. All authors approved the final
18 manuscript.

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8 receptor mRNA and protein in dorsal root ganglion neurons following local capsaicin
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13

- 1 **Table 1.** Demographic characteristics of the Asthmatic Participants and Healthy Volunteers (Belfast
2 and Leicester cohorts combined). Comparison with the Healthy Volunteer group [†]p<0.001, [§]p<0.001,
3 ^{||}p<0.001, ^{**}p<0.001, ^{§§}p<0.001, ^{‡‡}p<0.05, ^{||||}p<0.001. Comparison with Mild to Moderate
4 Asthmatics: [†]p<0.001, [‡]p=0.007, [§]p<0.001, ^{||}p<0.001, ^{**}p<0.001, ^{††}p<0.0001, ^{§§}p<0.05, ^{***}p<0.001.

	Healthy Volunteers (n=22)	Mild to Moderate Asthmatics (n=30)	Severe Asthmatics (n=31)	p Value*
Age (y), median (IQR)	30 (22 - 45)	29 (24 - 39)	48 (38 - 54) [†]	<0.0001
Sex, M/F	10/12	14/16	22/9	0.09
Duration of asthma (y), mean ± SD	N/A	12 ±10	22±15 [‡]	0.007
Atopy, (n)	11	21	23	0.16
FEV ₁ (L/min), mean ± SD	3.67±0.78	3.32±0.77	2.48±0.67 [§]	<0.0001
FEV ₁ (%predicted), mean ± SD	102.8± 11.9	93.5±16.0	75.9± 16.9	<0.0001
FEV ₁ / FVC, mean ± SD	82.1± 9.8	75.9± 10.6	64.6± 11.1 ^{**}	<0.0001
Inhaled steroid (n)	0	19	31	
BDP equivalent (µg), median (IQR)	0	400 (0 - 800)	1600 (1200 - 1600) ^{††}	<0.0001
Maintenance systemic steroids (n) ^b	0	0	15 (median dose prednisolone 5 mg [0 – 10 mg] plus 1 subject maintenance intramuscular triamcinolone)	
Theophylline, (n)	0	1	17	<0.0001
Long-acting β ₂ -agonists, (n)	0	14	31	<0.0001
Prior smoking	1	5	13	0.004

history (n) [pack years, median range]	[4]	[1, 1-10]	[2.5, 1 – 20]	
Eosinophils (10⁹/L), median (IQR)	0.1 (0.07 - 0.21)	0.26 (0.14 - 0.45) ^{††}	0.49 (0.25 - 0.74) ^{§§}	<0.0001
IgE (kU/L), geometric mean (95% confidence interval)	24.6 (13.3 - 45.8)	159.8 (89.8 - 284.6)	238.5 (136.2 - 417.5) ^{***}	<0.0001

1 * Calculated by ANOVA with Bonferroni's Multiple Comparisons Test (parametric) or Kruskal-Wallis
2 with Dunn's Multiple Comparison Test (non-parametric) or t-test when only asthma groups
3 compared or Chi-squared for categorical variables (sex, atopy, theophylline, long-acting β 2-agonists,
4 smoking history).

5

6

1 | **Table 2.** Demographic characteristics of the ~~cough-variant asthmatic~~mild asthmatic, non-asthmatic
 2 | cough and healthy volunteers recruited for experiments on cultured primary bronchial epithelial
 3 | cells.

	Healthy volunteers (n=4)	Cough-variant <u>mild</u> asthmatics (n=4)	Non asthmatic coughers (n=4)	P value
Age, (y) Median (IQR)	35.5 (21.3-43.8)	46.0 (39.3-56.8)	58.0(45.0-72.5)	0.05
Sex, M/F	1/3	0/4	0/4	0.336
Duration of cough (m) Mean ±SD	N/A	49.5 (30.0)	57.5 (27.0)	0.950
FEV1 (L/min) Mean ±SD	3.45 (0.65)	2.52 (0.29)	2.3 (0.34)*	0.014
FEV1, % pred Mean ±SD	107.3 (10.59)	105.6 (15.23)	118.0 (21.28)	0.530
FEV1/ FVC Mean ±SD	86.6 (10.6)	82.73 (4.11)	80.2 (3.2)	0.446
Atopy (n)	0	0	0	N/A
Inhaled steroid (n)	0	4	0	N/A
Prior smoking (n)	0	0	0	N/A

4 | Calculated by ANOVA with Bonferroni's Multiple Comparisons Test (parametric) or Kruskal-Wallis
 5 | with Dunn's Multiple Comparison Test (non-parametric) or t-test when only cough groups compared
 6 | or Chi-squared for categorical variables (gender).

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2 Legends to Figures

3 **Figure 1: A. Molecular expression of TRPV channels subtypes in human primary bronchial**
4 **epithelial cells.** Conventional RT-PCR showing the expression of TRPV family subtypes
5 including TRPV1 in PBECs from healthy controls (n=3) **B. Expression of TRPV1 protein in**
6 **~~primary bronchial epithelial cells (PBECs). Protein was extracted and analysed by Western~~**
7 **~~blotting (30 µg protein per sample). Blots were probed using a TRPV1 specific primary~~**
8 **~~antibody (Abcam 63083).~~** TRPV1 specific bands were observed at the molecular weight of 95
9 KDa in PBECs (lanes 2 to 4, upper panel). TRPV1 transfected HEK cells were used as a positive
10 control (lane 1, upper panel). β-Tubulin (lower panel) was used to ascertain equal loading
11 (Abcam 6046). Results are representative of two independent experiments. **C. Gene**
12 **expression microarray analysis of bronchial brush samples from healthy, mild/moderate**
13 **and severe asthmatics.** Gene expression microarray analysis was performed on fresh
14 bronchial brush samples from the Leicester cohort comprising severe asthmatics (n=12),
15 mild/moderate asthmatics (n=16) and healthy subjects (n=8). TRPV1 was significantly
16 elevated in severe asthma versus mild moderate asthma (* p=0.008) and healthy volunteers
17 (** p=0.004).

18

19 **Figure 2 - Digital photographic images (A-D) of TRPV1 expression in JB4 sections from (A)**
20 **healthy control; ~~showing~~ only patchy or focal areas of low intensity epithelial expression; (B)**
21 **moderate asthmatic; ~~where the~~ majority of epithelial and submucosal cells express antigen;**
22 **(C) severe asthmatic; ~~where there is~~ widespread and strong epithelial and submucosal cell**

1 | expression of TRPV1. Sections incubated in the absence of antibodies showed no staining; a
2 | representative image is shown in (D). Representative immunohistochemistry shown at X 400
3 | magnification

4 |
5 | **Figure 3** – Confocal microscopy images (A-C) of double staining immunocytochemistry
6 | demonstrating localisation of TRPV1 channels in bronchial epithelium (healthy subject).
7 | Bronchial epithelial cells identified by positive red staining with the cytokeratin antibody
8 | AE1AE3 (A) were also positive for the TRPV1 (green) (B) and co-localisation was evident
9 | when the images were merged (yellow) (C). Representative immunohistochemistry shown
10 | at X 400 magnification

11 |
12 | Figure 4: TRPV1 and ER colocalisation in human bronchial epithelial cells. Representative
13 | confocal microscopy images at X 40 magnification (A-C) showing immunohistochemistry
14 | staining of bronchial epithelial cells positive for TRPV1 (A) and ER (B) markers. Colocalisation
15 | was evident when the images were merged (C; orange/yellow). DAPI stained blue nuclei (C).
16 | Scale bar = 100 µm.

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19 | **Figure 4-5** Quantitative immunohistochemical epithelial expression for TRPV1 in bronchial
20 | biopsies. TRPV1 expression is increased in the bronchial epithelium of severe asthmatic
21 | patients. There was a trend to increased TRPV1 expression with severity of asthma ($p=0.001$,
22 | Kendall's tau-B statistic).

1 | **Figure 56.** Whole-cell PBEC (healthy subject) membrane current responses to capsaicin
2 | application (50 μ M) measured by voltage-steps. **A.** Superimposed current traces evoked by 1
3 | s voltage steps before, in the presence of, and after capsaicin wash-out, as indicated. ~~The~~
4 | ~~dotted~~-Dotted line indicates zero-level current. **B.** Mean normalised I-V relationship of
5 | background current (n=4). **C.** I-V relationships for the current amplitude measured at the
6 | end of each voltage step in the experiment illustrated in panel A.

7 | **Figure 67.** Time course of the capsaicin membrane current response. **A.** Current
8 | amplitude was monitored by applying voltage ramps from -100 to 140 mV at 10 s interval.
9 | Circles show current amplitude at 140 mV. **B.** The I-V relationships were derived from the
10 | ramps applied before, at the peak response and 3 min after wash-out of the drug as
11 | indicated. **C.** Mean normalised I-V relationships measured before (squares) and after
12 | (circles) capsaicin application (n=3). In each cell, current amplitude at 140 mV in control was
13 | normalised as 1.0.

14 | **Figure 8.** Whole-cell PBEC (healthy subject) membrane current responses to reduced
15 | external pH measured by voltage ramps. **A.** Superimposed current traces evoked by voltage
16 | ramps from -100 to 140 mV at 10 s interval in pH 7.4 and pH 6.4 (\pm capsazepine; 5 μ M)
17 | external solutions as indicated. The dotted line indicates zero-level current. **B.** Mean \pm SEM
18 | I-V relationship (normalized for cell capacitance) of capsazepine-sensitive current (n=4).

19 | **Figure 79.** Capsaicin induces IL-8 release from human PBECs (asthmatic and non-asthmatic
20 | subjects). PBECs ~~were~~-treated ~~(-for~~24 hours) with 10, 25 or 50 μ M capsaicin or control
21 | medium. ~~Supernatants were collected after 24 h and ELISA was performed to measure the~~
22 | ~~release of IL-8.~~ Values are expressed as the mean release over basal \pm SEM. **A.** Capsaicin-

1 induced IL-8 release *** $p < 0.001$ compared to control; ~~measured using a~~ Kruskal-Wallis test
2 with Dunn's multiple comparison ~~test for~~ (n=12 individual experiments). **B.** Capsazepine
3 inhibits capsaicin induced IL-8 release from human PBECs. PBECs were pre-treated for 20
4 min with 10 μ M capsazepine. ~~Following which media was replaced then~~ with 10, 25 or 50 μ M
5 capsaicin or control medium with or without capsazepine. ~~Supernatants were collected after~~
6 ~~24 h and ELISA was performed to measure the release of IL-8.~~ Values are expressed as the
7 mean release over basal \pm SEM. Capsazepine significantly inhibited capsaicin-induced IL-8
8 release ~~* $p < 0.001$~~ compared to 25 μ M (~~* $p < 0.001$~~) ~~and capsaicin alone~~ and 50 μ M
9 (~~** $p < 0.001$~~) ~~compared to 50 μ M~~ capsaicin alone; ~~measured using~~ one way ANOVA with
10 Bonferroni's multiple comparison test ~~for~~ (n=9 individual experiments).

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ONLINE DATA REPOSITORY

Increased expression of bronchial epithelial transient receptor potential vanilloid 1 channels in severe asthma

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Study Subjects

The studies were approved by the Research Ethics Committee of both Institutions (Belfast - Office of Research and Ethics Committee of Northern Ireland reference 06/NIR02/114 and 09/NIR02/51, Leicester - Leicestershire, Northamptonshire, & Rutland Research Ethics Committee reference 04/Q2502/74). Written informed consent was gained from all participants prior to their involvement.

Bronchial biopsies:

Belfast Cohort: Participants with refractory asthma were recruited from the Belfast City Hospital where systematic evaluation protocols ensure patients have well-characterised refractory asthma. All participants had persisting symptoms despite treatment at Step 4 or 5 of the GINA guidelines Global Initiative for Asthma (GINA) (E1). Mild to moderate asthmatic and normal participants were recruited by advertisement and had a clinical diagnosis of asthma with a current history of recurrent wheezing and response to asthma medication. Healthy volunteers had no history of asthma or persistent respiratory symptoms and normal lung function. All asthmatic participants were clinically stable on their usual medication at the time of bronchoscopy, and all participants had an FEV1 of greater than 60% predicted. Demographic details are shown in Table 1 of the main paper.

Leicester cohort: For the gene expression studies, asthmatic subjects (n=36) and healthy volunteers (n=12) were recruited from respiratory clinics including the Leicester Difficult Asthma Clinic, from staff at Glenfield Hospital, and from the general population through

advertisement (Leicester cohort). Subjects with asthma gave a suggestive history and had objective evidence of variable airflow obstruction as indicated by one or more of the following: (1) methacholine airway hyperresponsiveness ($PC_{20} FEV_1 < 8 \text{ mg/mL}$), (2) $>15\%$ improvement in FEV_1 10 minutes after 200 μg inhaled salbutamol, (3) peak expiratory flow ($>20\%$ maximum within-day amplitude from twice-daily peak expiratory flow measurements over a period of 14 days). Subjects underwent spirometry; allergen skin prick tests for *Dermatophagoides pteronyssinus*, dog, cat, grass pollen and *Aspergillus fumigatus*; a methacholine inhalation test using the tidal breathing method; and sputum induction using incremental concentrations of nebulized hypertonic saline (i.e., 3%, 4%, and 5%, each for 5 min)(E2). Subjects with asthma also kept a diary card for 2 weeks prior to bronchoscopy, recording daytime and night-time symptoms, daily short-acting β_2 -agonist use and twice daily peak expiratory flow (PEF). Asthma severity was defined by British Guideline on the Management of Asthma treatment steps (mild = step 1, β_2 -agonist only; moderate = steps 2 and 3, inhaled corticosteroid $\leq 800 \text{ mg}$ beclomethasone equivalent per day \pm long-acting β_2 -agonist; severe = step 4 and 5) (E3). Of the 16 severe patients at step 4/5, 13 met the American Thoracic Society criteria for refractory asthma (E4).

In both cohorts, all participants were current non-smokers with no upper or lower respiratory tract infection in the 6 weeks prior to their bronchoscopy. All asthmatic participants were clinically stable on their usual medication at the time of bronchoscopy. Demographic details for each cohort are shown in Table 1 in the main manuscript.

Primary bronchial epithelial cell (PBEC) cultures: To investigate the functional role of TRPV1 we performed a series of experiments on primary bronchial epithelial cell (PBEC) cultures from bronchial brushings obtained from mild asthmatic (n=4) and non asthmatic patients with chronic cough (n=4) referred to the Belfast City Hospital specialist cough clinic (E5). Healthy volunteers (n=4) were recruited by local advertisement. Demographic details for this cohort are shown in Table 2 in the main manuscript.

Bronchoscopy and samples

Bronchoscopy was performed using a standard technique. In brief, after intravenous sedation and local anaesthesia with topical lignocaine, bronchial biopsies and bronchial brushings were obtained from lobar / segmental airways. One biopsy from each participant was immediately placed in RNA preservative (RNAlater, Ambion, Austin, TX) and submitted for microarray analysis and qPCR and 3 biopsies placed immediately into dry acetone containing protease inhibitors (iodoacetamide and PMSF) and fixed overnight at -20°C. Brushings were placed in RNAlater or Promocell™ medium containing penicillin/streptomycin and Primocin™ for cell culture (Promocell UK). PBECs were cultured using methods developed in our laboratory which resulted in homogenous cultures of basal epithelial cells (E6).

For immunohistochemistry (IHC), biopsy samples were processed using local protocols. Twenty eight biopsies (10 healthy volunteer, 8 mild asthmatic and 10 severe asthmatic biopsies) were available for immunohistochemistry. Infiltration and embedding with JB4 resin was performed as previously described (E7). Two µm sections were cut from each biopsy. Endogenous peroxidase was blocked with 0.3% hydrogen peroxide. All sections were pressure cooked in 0.01M Tris-EDTA pH9.0 on full steam for 3 minutes for antigen retrieval.

Sections were incubated with TRPV1 polyclonal antibody (1:200, Abcam), overnight at 4°C, then primary antibodies were detected by incubation for 30 min at RT in peroxidase-labelled EnVision anti-rabbit secondary antibody (Dako, Ely, UK) using 3,3'-diaminobenzidine (DAB; Dako, Ely, UK) as chromagen. On selected tissue sections from this series, immunofluorescence was carried out to permit high resolution analysis of TRPV1 antigen distribution. Sections were treated as for IHC and following incubation in a mix of TRPV1 and a monoclonal antibody to the epithelial specific antibody AE1AE3 (1:50, Dako). Antigen binding sites were detected with a mixture of anti-mouse Alexa 568 and anti-rabbit Alexa 488 (Invitrogen). Sections were counterstained with DAPI hardset mounting medium (Vector). All fluorescently stained slides were assessed and digital fluorescent images acquired with a Leica DFC350 FX digital camera and processed using Leica FW4000 software. Immunostained sections were evaluated by two independent persons, following which results were discussed and consensus scoring applied to any conflicting results. A 4-point scoring system was used: 0 - no detectable expression in epithelium or submucosa; 1 - patchy or focal areas of low intensity epithelial expression, not all cells expressing positivity for protein, scattered positivity in submucosa; 2 - moderate / majority of epithelial and submucosal cell expression; 3 - widespread and strong epithelial and submucosal cell expression.

TRPV1 and ER colocalisation in human bronchial epithelial cells.

Human bronchial epithelial cells were grown on coverslips and fixed in 4% paraformaldehyde (PFA) for 30 minutes at RT. Cells were briefly washed in Phosphate Buffered Saline (PBS) before being permeabilised with 0.1% Triton X-100 in PBS for 20 minutes at RT. Cells were blocked with 10% Bovine Serum Albumin (BSA) in PBS for 1 hour at

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RT and incubated with primary antibodies, mouse monoclonal anti-KDEL (1:500), ER marker and rabbit polyclonal anti-VR1 (1:200), TRPV1 marker (AB12223 and Ab63083 respectively; Abcam, Cambridge, UK) overnight at 4⁰C, followed by incubation for 1 hour at RT with secondary antibodies, Alexa Fluor goat anti-mouse 488 (1:500; Invitrogen) and Alexa Fluor goat anti-rabbit 568 (1:500; Invitrogen). Cell nuclei were stained using Vectashield mounting medium containing DAPI (Vector Labs, Peterborough, UK). Staining was visualised and images captured using Nikon EZ-C1 confocal system running C1 acquisition software (Nikon UK Ltd. Surrey, UK) at x40 magnification.

Gene expression analyses

RNA was isolated from homogenized bronchial biopsies and real-time PCR (qPCR) was performed as described previously (E8). TaqMan[®] Gene Expression Assays (Applied Biosystems, Foster City, CA) were purchased and conducted per manufacturer's instructions for TRPV1. RNA was amplified (Ambion,) for Agilent (Santa Clara, CA) two color Whole Human Genome (WHG) 4x44k gene expression microarray analysis. Universal Human Reference RNA (Stratagene, La Jolla, CA) was used for the reference channel. Probe intensities were transformed as log₂ ratios of test and reference channels calculated by the Agilent Feature Extraction software, protocol GE2-v5_95 (Agilent). Flagged outliers were not included in any subsequent analyses. All gene expression analyses and plotting were performed using the R Project software package, version 2.10.1 (refer to <http://www.R-project.org>).

Quantitative Real-Time PCR

Bronchial brushings

Real-time PCR (qRT-PCR) analyses were performed using RNA from homogenized bronchial brushings (36 subjects; 10 HV, 16 MA and 15 SA) using TaqMan® Gene Expression Assays (Applied Biosystems, Foster City, CA) as previously described. The delta-delta Ct method was used to calculate relative fold expression with GAPDH as housekeeping gene

PBEC cultures

PBECs were trypsinised and cells lysed using Qiagen RLT buffer (Qiagen, Hilden, Germany) before homogenisation using QIAshredders (Qiagen, Hilden, Germany). RNA was then extracted using the RNeasy Mini Kit (Qiagen, Hilden, Germany). RNA concentration was measured with the nanodrop spectrophotometer (Thermo Scientific, Wilmington, USA) and 1µg RNA was used to synthesise cDNA using the high capacity cDNA reverse transcription kit (Applied Biosystems, Foster City, CA). PBEC cDNA was mixed with Power Sybr Green PCR master mix (Applied Biosystems, Foster City, CA) and either GAPDH or TRPV1 forward and reverse primers (Table E1). The following PCRs were performed in a 7300 Real Time PCR system (Applied Biosystems, Foster City, CA). Melting curves showed formation of only one gene product per primer pair. Normalisations and calibrations of the data from all studies were performed with the $2^{-\Delta\Delta C_T}$ method as described in Applied Biosystems User Bulletin No. 2 (P/N 4303859) (27). Delta C_T was calculated as target gene expression (C_T) minus internal housekeeping control, GAPDH gene expression (C_T). Human Universal Reference RNA (Clontech, P/N 639654) was prepared as cDNA and utilised as plate calibrators. DeltaDelta C_T was calculated as sample Delta C_T minus plate calibrator Delta C_T . GAPDH and TRPV1 forward and reverse primers are detailed in table E1 below.

Table E1. Forward and reverse primers for TRPV1 and GAPDH

Primer name	Forward	Reverse	Size (bp)
TRPV1	GCCTGGAGCTGTTCAAGTTC	GCCTGAAACTCTGCTTGACC	452
TRPV2	CCAGCTCTCCAGTTTTTCAGG	GTTCCAGCACAGCCTTCATCA	365
TRPV3	TTCCTCATGCACAAGCTGAC	TGCGCTTCACAAAGTCATTC	490
TRPV4	CCCGTGAGAACACCAAGTTT	GAAGAGCGTAATGACCTCGC	374
TRPV5	TTGCTCAGTTGCTGTCATCC	GCTCCTCTTTGTGCGAACGTC	307
TRPV6	TCACTTCGCTTCCTGGAAGT	ACACGCTTTCCACAAGCTCT	401
GAPDH	GCTCTCTGCTCCTCTGTTC	AAATCCGTTGACTCCGACC	100

Source: Invitrogen

Western Blotting

PBECs were collected and snap frozen, homogenised and lysed in RIPA buffer supplemented with protease inhibitor cocktail (Roche Diagnostics, Mannheim, Germany). Protein concentration was determined using BCA protein assay kit (Thermo Scientific, Wilmington, USA). 30µg of total protein was separated on the 8% SDS-Polyacrylamide gel by electrophoresis and transferred to Hybond-P PVDF membrane (GE Healthcare Life Sciences, Buckinghamshire, UK). The membrane was blocked for 30 min in 3% BSA/ PBS and then probed overnight in 3% BSA/PBS with a rabbit polyclonal TRPV1 antibody (ab63083; Abcam, Cambridge, UK), at a dilution of 1:3000. Followed by incubation for 1 h with the secondary antibody, HRP-labelled anti-rabbit (1:3000; Bio-Rad Laboratories, Hercules, CA, USA). Images

were obtained using UVP bioimager (UVP, Cambridge, UK). Benchmark Pre-stained protein standards (Invitrogen, Paisley, UK) were used to determine molecular mass.

Patch clamp experiments

Whole-cell currents were recorded using borosilicate heat-polished patch pipettes (2-3 M Ω resistance when filled with the pipette solution) and an Axopatch 200B amplifier (Molecular Devices, Union City, CA, USA) interfaced to Digidata 1322A using the pClamp 9 software (Molecular Devices). Series resistance was compensated by about 70%. Holding potential was -40 mV. Steady-state current-voltage (I-V) relationships were measured by applying either voltage steps (test potentials from -100 to +140 mV with a 20 mV increment) or slow voltage ramps from -100 to +140 mV. To monitor the time course of TRPV1 activation by capsaicin, the voltage ramps were applied at 10 s interval. The I-V curves obtained by the voltage ramp and step protocols were similar confirming steady-state channel activity at each test potential when ramps were used.

[For experiments involving the addition of capsaicin the](#) ~~The~~ external solution contained (mM): NaCl 120, glucose 12, HEPES 10, pH 7.4 (adjusted with NaOH). Pipette solution contained (mM): CsCl 80, MgATP 1, creatine 5, GTP 1, D-glucose 5, HEPES 10, BAPTA 10, CaCl₂ 4.6 ([Ca²⁺]_i=100 nM), pH 7.4 (adjusted with CsOH). [For experiments involving altered pH the external solution contained \(mM\): NaCl 150, CsCl 6, MgCl₂ 1.3, CaCl₂ 2, Glucose, 5; HEPES, 10; pH7.4 or 6.4 with Tris. Pipette solution contained \(mM\): CsOH 130, D Gluconic Acid 120, MgCl₂ 1, EGTA 0.5 \(\[Ca²⁺\]_i= 30 nM\), pH 7.2 with Tris.](#)

IL-8 ELISA

IL-8 concentrations were measured from aliquots of culture medium taken 24 h following treatment with capsaicin (Enzo Life Sciences, Exeter, UK) using a commercial ELISA kit from R & D Systems Europe (Abingdon, UK).

Data Analysis

Data are presented as means \pm SEM. For statistical analysis a one way ANOVA was performed with Bonferroni test for multiple comparisons. Kendall's tau b statistical analysis was used to examine the trend in biopsy expression by participant group. Patch clamp data were analysed and plotted using Origin 8 (OriginLab, Northampton, MA, USA). A P value < 0.05 was considered significant.

RESULTS

Real time PCR (qRT PCR) analysis of fresh bronchial brush samples from the Leicester cohort comprising healthy, mild/moderate and severe asthmatics. Analysis was performed on fresh bronchial brush samples from the Leicester cohort comprising severe asthmatics (n=15), mild/moderate asthmatics (n=16) and healthy subjects (n=10). TRPV1 mRNA was detected in all of the epithelial brush samples. There was no difference in median (IQR) relative fold expression of TRPV1 mRNA calculated as Delta Delta C_T values between groups: -1.262 (-3.598, 2.518), (-2.523 (-4.040, 1.362) and -1.289 (-4.746, 1.624) in healthy volunteers, mild / moderate asthmatics and severe asthmatics respectively (see figure E1 below).

Online Figure Legend

Figure E1: Real time PCR (qRT PCR) expression of TRPV1 in fresh bronchial epithelial brush samples from healthy volunteers, mild/moderate asthmatics and severe asthmatics.

Analysis of fresh bronchial brush samples from the Leicester cohort comprising healthy (n=10), mild/moderate (n=16) and severe asthmatics (n=15). Data presented as median (IQR) relative fold expression of TRPV1 mRNA calculated as delta delta C_T values.

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1 **Increased expression of bronchial epithelial transient receptor potential vanilloid 1**
2 **channels in severe asthma**

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4 **ABSTRACT (250 words)**

5 **Background:** The airway epithelium is exposed to a range of physical and chemical irritants
6 in the environment which are known to trigger asthma. Transient Receptor Potential (TRP)
7 cation channels play a central role in sensory responses to noxious physical and chemical
8 stimuli. Recent genetic evidence suggests an involvement of TRPV1, one member of the
9 vanilloid subfamily of TRP channels, in the pathophysiology of asthma. The functional
10 expression of TRPV1 on airway epithelium has yet to be elucidated.

11 **Objective:** In this study we examined the molecular, functional and immunohistochemical
12 expression of TRPV1 in the asthmatic and healthy airway.

13 **Methods:** Bronchial biopsies and bronchial brushings were obtained from healthy
14 volunteers (n=18), mild to moderate asthmatics (n=24) and refractory asthmatics (n=22). To
15 investigate the functional role of TRPV1, cultured primary bronchial epithelial cells (PBEC)
16 from mild asthmatics (n=4), non asthmatic coughers (n=4) and healthy subjects (n=4) were
17 studied.

18 **Results:** Quantitative immunohistochemistry revealed significantly more TRPV1 expression
19 in asthmatics compared to healthy subjects and with greatest expression in the refractory
20 asthmatics (p=0.001). PCR and Western blotting analysis confirmed gene and protein
21 expression of TRPV1 in cultured primary bronchial epithelial cells (PBEC). Patch-clamp
22 electrophysiology directly confirmed functional TRPV1 expression in all three groups. In

1 functional assays, the TRPV1 agonist capsaicin induced dose dependent interleukin-8 (IL-8)
2 release which could be blocked by the antagonist capsazepine. Reduction of external pH
3 from 7.4 to 6.4 activated a capsazepine-sensitive outwardly-rectifying membrane current.

4

5 **Conclusions:** Functional TRPV1 channels are present in the human airway epithelium and
6 are over-expressed in the airways of refractory asthmatics. These channels may represent a
7 novel therapeutic target for uncontrolled asthma.

8

1 **Clinical Implications (30 words)**

2 TRPV1 is a functional channel activated by physical and chemical irritants. We report its
3 overexpression in asthmatic airway epithelium most notably in severe disease suggesting a
4 role in uncontrolled asthma.

5 **Capsule summary (35 words)**

6 The Transient Receptor Potential vanilloid-1 (TRPV1) channel regulates responses to irritant
7 stimuli. We report TRPV1 overexpression in asthmatic airway epithelium most notably in
8 severe disease suggesting its potential as a therapeutic target for uncontrolled asthma.

9

10 **Key words:** ion channel, sensory, asthma, irritant, chemical, exacerbation, cough

11 **Abbreviations:**

12 TRP - Transient Receptor Potential

13 TRPV1 – Transient Receptor Potential Vanilloid-1

14 GINA – Global Initiative for Asthma Guidelines

15 PBEC – Primary bronchial epithelial cell

16 qPCR – Quantitative real-time PCR

17 GMA – glycol methacrylate

18

1 INTRODUCTION

2 During an exacerbation of asthma, the airways become hypersensitive and patients are
3 often troubled with bouts of cough and wheeze following exposure to relatively innocuous
4 stimuli such as aerosols, strong odours or changes in air temperature. The precise
5 mechanism for this airway hypersensitivity is not known but upregulation of receptors
6 responsible for sensing chemical and physical stimuli may be relevant. The recently
7 described Transient Receptor Potential (TRP) proteins, a family of Ca^{2+} permeable non
8 selective cation channels which sense a vast array of chemical and physical stimuli (1, 2), are
9 believed to have an important role in the regulation of airway function in both health and
10 disease (3, 4). Recent data has provided genetic evidence for the involvement of one
11 subfamily member, TRP vanilloid 1 (TRPV1) in asthma pathophysiology (5).

12 TRPV1 is activated by capsaicin (the pungent principle contained in hot chilli peppers) and
13 inhaled capsaicin causes cough (6) and bronchospasm (7) in asthmatics. The airway
14 inflammatory events which accompany an exacerbation of asthma include a rise in airway
15 temperature (8), acidosis (9) and release of mediators derived from lipoxygenase
16 metabolism of arachidonic acid (4, 10). TRPV1 is primarily recognised as a heat thermal
17 sensor, but it is also readily activated by low extracellular pH (11, 12) and lipoxygenase
18 products such as leukotriene B_4 . (13). TRPV1 receptors are expressed in several cell types in
19 human airway including sensory neurones (14), smooth muscle cells (15) and epithelial cells
20 from both the upper (16) and lower airway (17). As the bronchial epithelium has an
21 important role in sensing and responding to noxious stimuli we hypothesised that TRPV1 is
22 over expressed in the airway epithelium of asthmatics, in particular those with difficult-to-

1 treat asthma. Further we considered that the inflammatory response following irritant
2 activation of airway epithelial TRPV1 may contribute to heightened neuronal reflexes
3 responsible for cough and bronchospasm in asthma.may .

4 In this study, we examined both TRPV1 expression and function in human bronchial
5 epithelium. In addition we provide evidence for the overexpression of TRPV1 in the airways
6 of severe asthmatics compared to mild asthmatics and healthy subjects. Some of the results
7 of these studies have been reported in abstract form (18).

8

9 **METHODS AND MATERIALS**

10 *Full methodological details are provided in an online data repository*

11 *Study Subjects*

12 Participants with refractory asthma were recruited from the Belfast City Hospital. All had
13 persisting symptoms despite treatment at Step 4 or 5 of the Global Initiative for Asthma
14 (GINA) guidelines. Mild to moderate asthmatics had a clinical diagnosis of asthma with a
15 current history of recurrent wheezing and documented response to asthma medication.
16 Bronchial biopsy samples from this Belfast cohort were analysed using
17 immunohistochemistry (IHC). For examination of *in vivo* gene expression of TRPV1 in airway
18 epithelium an additional cohort of subjects from Leicester were recruited. Demographic
19 details of both cohorts are shown in Table 1. To investigate the functional role of TRPV1,
20 primary bronchial epithelial cell (PBEC) cultures were grown from bronchial brushings
21 obtained from mild asthmatics and non-asthmatic patients recruited from a specialist cough

1 clinic (19) and healthy participants recruited by advertisement. Demographic details are
2 shown in Table 2.

3 The study was approved by the appropriate Institutional Research Ethics Committees (details in
4 online data repository). Written informed consent was gained from all participants prior to their
5 involvement.

6

7 *Bronchoscopy and sample processing*

8 All asthmatic participants were clinically stable at the time of bronchoscopy, and had an
9 FEV₁ >60% predicted. Bronchial biopsies and brushings were obtained from segmental
10 airways using a standard techniques. Three biopsies from each participant (Belfast Cohort)
11 were submitted for immunohistochemistry studies. Gene expression microarray analysis
12 (and qPCR) was performed on fresh bronchial brushings (Leicester cohort). To investigate
13 the functional role of TRPV1, PBEC cultures from bronchial brushings were cultured as
14 described previously (20). These were obtained from mild asthmatic (n=4), non asthmatic
15 patients with chronic cough (n=4) and healthy volunteers (n=4).

16 Immunohistochemistry

17 Twenty eight biopsies (10 healthy volunteer, 8 mild asthmatic and 10 severe asthmatic
18 biopsies) were processed for immunohistochemistry (IHC) using local protocols as detailed
19 in the online data repository. For TRPV1 and ER co-localisation experiments local protocols
20 were employed as detailed in the online data repository.

21

1 ***Gene expression analysis***

2 *Bronchial brushings and PBEC cultures*

3 Real-time PCR (qRT-PCR) analyses were performed using RNA from bronchial brushings using
4 TaqMan® Gene Expression Assays (Applied Biosystems, Foster City, CA) as previously
5 described (21). RNA was amplified (Ambion,) for Agilent (Santa Clara, CA) two color Whole
6 Human Genome (WHG) 4x44k gene expression microarray analysis.

7 PBECs were trypsinised and total RNA extracted using the RNeasy Mini Kit (Qiagen, Hilden,
8 Germany) and qRT-PCR performed using a 7300 Real Time PCR system (Applied Biosystems,
9 Foster City, CA). More information including detail on GAPDH and TRPV1 forward and
10 reverse primers is provided in the online data repository.

11 *Western Blotting*

12 Protein concentration was determined using BCA protein assay kit (Thermo Scientific,
13 Wilmington, USA) and protein expression determined using a rabbit polyclonal TRPV1
14 antibody (ab63083; Abcam, Cambridge, UK), at a dilution of 1:3000 followed by incubation
15 for 1 h with the secondary antibody, HRP-labelled anti-rabbit (1:3000; Bio-Rad Laboratories,
16 Hercules, CA, USA). More details provided in online data repository.

17 *TRPV1 agonists / antagonist experiments in PBECs*

18 Capsaicin is a selective TRPV1 agonist and in this study cultured PBECs were treated with either 10,
19 25 or 50 µM capsaicin or control medium for 24 h and the concentration of IL-8 measured by ELISA
20 (R & D Systems Europe Abingdon, UK). For the antagonist experiments, PBECs were pre-
21 treated for 20 min with 10 µM of the selective TRPV1 antagonist capsazepine.

22 *Patch clamp experiments*

1 Whole-cell currents were recorded using borosilicate heat-polished patch pipettes (2-3 M Ω
2 resistance when filled with the pipette solution) and an Axopatch 200B amplifier (Molecular
3 Devices, Union City, CA, USA) interfaced to Digidata 1322A using the pClamp 9 software
4 (Molecular Devices) as previously described by ourselves (22) and detailed in the online data
5 repository.

6 *Data Analysis*

7 Data are presented as means \pm SEM. For statistical analysis a one way ANOVA was performed
8 with Bonferroni test for multiple comparisons. Kendall's tau b statistical analysis was used to
9 examine the trend in biopsy expression by participant group. Patch clamp data were
10 analysed and plotted using Origin 8 (OriginLab, Northampton, MA, USA). A P value < 0.05
11 was considered significant.

12

13 **RESULTS**

14 **Confirmation of TRPV1 expression in cultured human bronchial epithelial cells.**

15 Gene expression for multiple members of the TRPV family including TRPV1 (but not TRPV5)
16 was identified in PBECs from healthy controls (n=3) (Figure 1A). TRPV1 protein was detected
17 at its expected molecular mass of 95 kDa by Western blotting in the same samples (Figure
18 1B).

19 **Molecular identification and localisation (immunohistochemical expression) of TRPV1 in** 20 **healthy and asthmatic epithelium**

21 *Microarray*

1 Using gene expression microarray analysis of bronchial brush samples from the Leicester
2 cohort we found significantly greater levels of TRPV1 expression (median, IQR) in severe
3 asthmatics (0.673, 0.452-0.761) compared to mild/moderate asthmatics (0.341, 0.167-0.538,
4 $p=0.008$) and healthy volunteers (0.322, 0.155-0.410, $p=0.004$) (Figure 1C).

5 *Quantitative PCR.*

6 TRPV1 mRNA was detected in all of the epithelial brush samples. There was no difference in
7 median (IQR) relative fold expression of TRPV1 mRNA calculated as Delta Delta C_T values
8 between groups: -1.262 (-3.598, 2.518), (-2.523 (-4.040, 1.362) and -1.289 (-4.746, 1.624) in
9 healthy volunteers, mild / moderate asthmatics and severe asthmatics respectively (see
10 Figure E1 in online data repository)

11 *Immunohistochemistry*

12 Immunohistochemistry revealed bronchial epithelial expression of TRPV1 in healthy
13 volunteers, mild/moderate asthmatics and severe asthmatics (Figures 2A, B and C,
14 respectively). Dual labelling immunofluorescence confirmed the expression of TRPV1 on
15 cytokeratin positive epithelial cells (Figures 3 A, B and C). We observed an intracytoplasmic
16 distribution of the TRPV1 and to further characterise this we undertook additional
17 experiments demonstrating immunohistochemistry staining of bronchial epithelial cells
18 positive for TRPV1 (Figure 4A) and ER (Figure 4B) markers. Colocalisation was evident when
19 the images were merged (Figure 4C).

20 We observed greater TRPV1 expression in asthmatic epithelium compared to healthy
21 subjects (Figure 5; $p = 0.001$).

1 **Membrane current responses to TRPV1 agonist capsaicin**

2 Mean membrane capacitance of PBEC was 30.8 ± 4.3 pF (n=6). In unstimulated cells, only
3 small background currents could be recorded in the whole range of membrane potentials
4 tested, from -100 to +140 mV (Figure 6A, left panel). The I-V relationship of the mean
5 background current normalised by the membrane capacitance to account for some
6 variations in cell size is shown in Figure 6B. It should be noted that composition of the
7 external and pipette solutions was such that this facilitated isolation of cation currents but
8 precluded activation of any potassium or calcium-dependent conductances. Capsaicin
9 application (50 μ M) caused rapid activation of large outwardly rectifying currents (Figure 6A,
10 middle panel), which is the characteristic “biophysical” signature of TRPV1 channels. This
11 effect was completely reversible upon capsaicin wash-out (Figure 6A, right panel).
12 Corresponding I-V relationships are shown in Figure 6C. The capsaicin-induced currents
13 reversed close to 0 mV indicating their non-selective cation nature.

14 To investigate the time course of these responses, voltage ramps over the same range of
15 potentials were applied at 10 s interval. Figure 7A shows a typical response to 50 μ M
16 capsaicin application consisting of rapid activation and desensitisation in the continuous
17 presence of the agonist, which developed over 2-3 min. The I-V relationships which were
18 measured in this experiment in control, at the peak response and after agonist wash-out are
19 shown in Figure 7B (compare to I-V curves in Fig. 6C measured by voltage steps), while the
20 mean results are summarised in panel C. The increase in current amplitude in response to
21 capsaicin was significant (p=0.007).

22 **Membrane responses to reduction of external pH**

1 Reduction of external pH from 7.4 to 6.4 activated inward and outward currents in cells from
2 healthy patients which were partially inhibited by the addition of the selective TRPV1
3 antagonist capsazepine (5 μ M; Fig. 8A). The mean capsazepine-sensitive I-V relation (Fig. 8B
4 n=4) was similar in profile to the capsaicin-dependent currents exhibiting strong outward
5 rectification and reversal close to 0 mV.

6

7 **Functional expression of TRPV1 in cultured human primary bronchial epithelial cells**

8 *Dose-dependent induction of IL-8 release from cultured human PBECs by capsaicin*

9 Dose-dependent induction of IL-8 release by capsaicin was observed within each study
10 group although there was no significant difference in fold increase in IL-8 between asthmatic
11 and non asthmatic groups which may relate to sample size (data not shown). The dose
12 dependent increase in IL-8 release when data from all three groups was therefore pooled
13 has been presented in figure 9A. When compared to basal a marginal 1.25-fold increase in
14 IL-8 release was observed following incubation with 10 μ M capsaicin with significant
15 increases of 1.76-fold ($p < 0.01$) and 2.15 -fold ($p < 0.001$) compared to basal observed
16 following incubation with 25 and 50 μ M doses of capsaicin, respectively.

17 To further test the specificity of the capsaicin effect, we used selective TRPV1 blocker
18 capsazepine. Capsazepine significantly inhibited capsaicin-induced IL-8 release compared to
19 25 μ M capsaicin alone ($p < 0.001$) and compared to 50 μ M capsaicin alone ($p < 0.001$). The
20 data for n=9 individual experiments is shown in Figure9B.

21

1

2 **DISCUSSION**

3 In this study we have shown for the first time that TRPV1, which is primarily recognised as
4 the major neuronal hot receptor (23, 24), is expressed and functional in human bronchial
5 epithelium. In primary bronchial epithelial cell cultures we have shown that pharmacological
6 activation of the TRPV1 channel with capsaicin induces IL-8 release which can be attenuated
7 by pre-treatment with the selective TRPV1 antagonist capsazepine.

8 Interestingly, TRPV1 expression in the airway was increased in asthmatic subjects compared
9 to healthy controls with a further increase in subjects with refractory asthma. Thus, over-
10 expression may have a role in symptoms associated with irritant exposure in asthma,
11 particularly in patients with persisting symptoms despite treatment with high dose anti-
12 inflammatory therapy. Understanding the role of TRPV1 in refractory asthma may therefore
13 identify a novel target for anti-inflammatory therapy in more severe disease.

14 We believe that bronchial epithelial TRPV1 channels are ideally positioned to detect
15 chemical and physical irritants (e.g. inflammatory mediators, environmental pollutants, acid
16 pH and changes in temperature) in the airway lumen. To date, most of the focus has been
17 on neuronal TRPV1 in the human airway (14, 25). However, TRPV1 expression has been
18 reported in a variety of non-neuronal tissues and therefore we hypothesised that the airway
19 responses to environmental stimuli might in part be regulated by the bronchial epithelial
20 TRPV1 receptors. Recent studies in immortalised airway epithelial cell lines provide some
21 support for this (17, 26) although evidence in human disease is lacking. Here we provide *in*

1 *vivo* and *ex vivo* evidence of TRPV1 expression and function in human airway biopsies and
2 cultured primary bronchial epithelial cells with specific relevance to refractory asthma.

3 We observed an intracytoplasmic distribution of the receptor in bronchial epithelial cells as
4 observed in other cell types (27, 28). Our finding of TRPV1 staining **in the endoplasmic**
5 **reticulum is consistent with its functional role as a Ca²⁺ store release channel regulating**
6 **intracellular Ca²⁺ homeostasis.** Robust membrane current responses to the TRPV1 agonist
7 capsaicin (Figs. 5 and 6) suggest strongly that plasmalemmal TRPV1 is also expressed and
8 functional in PBEC. We also observed TRPV1 staining on a number of submucosal cells
9 although the focus of this present study is confined to the bronchial epithelial expression.

10 Previous studies from chronic coughers (14) and healthy subjects (15) have reported only
11 sparse TRPV1 expression in the airway epithelium, although greater expression in the upper
12 airway (29). Differences in tissue processing, antibodies used and clinical phenotype may
13 explain the discrepancy. In our studies to evaluate the functional properties of TRPV1 on
14 bronchial epithelium, we saw that PBECs responded to the TRPV1 agonist capsaicin with the
15 release of pro-inflammatory mediators which was blocked by capsazepine, a TRPV1 selective
16 antagonist. Capsaicin induced IL-8 release has been previously observed in both human
17 esophageal (30) and human corneal epithelial cells (31) but not before in cultured PBECs. As
18 IL-8 is known to induce and maintain neural sensitization (32, 33) our data provide a
19 mechanistic link whereby activation of bronchial epithelial TRPV1 by environmental irritants
20 or changes in airway pH or temperature (during exacerbations) may sensitize airway sensory
21 neurones and contribute to the airway and cough hypersensitivity that characterises
22 asthma.

1 Studies in immortalised respiratory cell lines suggest airway epithelial TRPV1 regulates cell
2 apoptosis in response to environmental irritants (34). Here, we provide the first evidence of
3 a functional airway epithelial TRPV1 channel in primary cells. In the whole-cell patch clamp
4 experiments on PBECs our results provide direct evidence for the presence of functional
5 TRPV1 in the plasma membrane that can mediate membrane depolarisation and calcium
6 influx; steps which are necessary for release of pro-inflammatory mediators (35). Both
7 kinetics and voltage-dependent properties of capsaicin-induced currents in PBEC are
8 consistent with TRPV1 biophysical features, such as slow activation/deactivation and strong
9 outward rectification.

10 TRP channels have the capacity to sense a vast range of stimuli and their role in airways
11 disease is currently under intense investigation (1). In our immunohistochemical studies we
12 have shown that TRPV1 is over expressed in the bronchial epithelium of asthmatics
13 compared to healthy subjects and patients with refractory asthma express more epithelial
14 TRPV1 than those with mild/moderate asthma. This was supported by our microarray
15 analysis confirming increased expression of TRPV1 genes in the more severe asthmatics.
16 Although on quantitative PCR, mRNA was not concordant with protein abundance, it has
17 become increasingly clear that at the genomic level, mRNA expression does not consistently
18 predict corresponding protein levels (36, 37). This may be because of complex post-
19 transcriptional and translational modification and /or that different biological mRNA and
20 protein degradation rates may affect correlation. Post translational modification of TRPV1
21 has been described in experimentally induced tissue injury (38) and therefore may occur in
22 the inflamed asthmatic airway.

1 While the vast majority of asthmatics respond to standard doses of current treatments (in
2 particular corticosteroids), a proportion remain symptomatic despite treatment at GINA
3 Step 4/5 and are considered to have difficult or treatment refractory asthma. Our finding
4 that bronchial epithelial TRPV1 is over expressed in refractory (steroid-resistant) asthma
5 suggests its activation may represent a novel pro-inflammatory pathway. Clinical trials of
6 TRP receptor antagonists have been undertaken in other abnormal hypersensitive states
7 including neuropathic pain and these drugs may prove effective in improving symptom
8 control in asthma as well.

9 Our findings suggest that the bronchial epithelium may sense and respond to environmental
10 stimuli via TRPV1 activation. However it is not clear how bronchial epithelial cells signal to
11 afferent sensory nerves. We suggest that release of sensitizing mediators such as IL-8 may
12 be one such mechanism. Such mediators may enhance and augment neurogenic
13 inflammation and nociceptive signalling. Therefore, pharmacological modulation of this
14 receptor may be useful in the treatment of uncontrolled asthma.

15

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1 **Author contributions:** CB, SA and SMcQ performed immunostaining, analysed the data and
2 contributed to the writing of the manuscript. SS and HA performed cell culture, molecular analysis of
3 samples and subsequent ELISAs. DC and JA performed and analysed the gene expression data,
4 analysed the data and contributed to the writing of the manuscript. CB, LH and PB recruited subjects,
5 performed bronchoscopy on the asthmatic subjects. LMG and LP recruited subjects for bronchial
6 brush sampling and with TJW performed the bronchoscopies. LH supervised CB and LMG and RC
7 supervised LP. AZ and MMcG performed and analysed patch-clamp experiments. LMG, AZ and ME
8 supervised SS. LMG and LH conceived and designed the research, contributed to data analysis and
9 interpretation and drafted the original manuscript. All authors approved the final manuscript.

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1 **Table 1.** Demographic characteristics of the Asthmatic Participants and Healthy Volunteers (Belfast
2 and Leicester cohorts combined). Comparison with the Healthy Volunteer group [†]p<0.001, [§]p<0.001,
3 ^{||}p<0.001, ^{**}p<0.001, ^{§§}p<0.001, ^{‡‡}p<0.05, ^{||||}p<0.001. Comparison with Mild to Moderate
4 Asthmatics: [†]p<0.001, [‡]p=0.007, [§]p<0.001, ^{||}p<0.001, ^{**}p<0.001, ^{††}p<0.0001, ^{§§}p<0.05, ^{***}p<0.001.

	Healthy Volunteers (n=22)	Mild to Moderate Asthmatics (n=30)	Severe Asthmatics (n=31)	p Value*
Age (y), median (IQR)	30 (22 - 45)	29 (24 - 39)	48 (38 - 54) [†]	<0.0001
Sex, M/F	10/12	14/16	22/9	0.09
Duration of asthma (y), mean ± SD	N/A	12 ±10	22±15 [‡]	0.007
Atopy, (n)	11	21	23	0.16
FEV ₁ (L/min), mean ± SD	3.67±0.78	3.32±0.77	2.48±0.67 [§]	<0.0001
FEV ₁ (%predicted), mean ± SD	102.8± 11.9	93.5±16.0	75.9± 16.9	<0.0001
FEV ₁ / FVC, mean ± SD	82.1± 9.8	75.9± 10.6	64.6± 11.1 ^{**}	<0.0001
Inhaled steroid (n)	0	19	31	<0.0001
BDP equivalent (µg), median (IQR)	0	400 (0 - 800)	1600 (1200 - 1600) ^{††}	
Maintenance systemic steroids (n) ^b	0	0	15 (median dose prednisolone 5 mg [0 – 10 mg] plus 1 subject maintenance intramuscular triamcinolone)	
Theophylline, (n)	0	1	17	<0.0001
Long-acting β2-agonists, (n)	0	14	31	<0.0001
Prior smoking	1	5	13	0.004

history (n)				
[pack years, median range]	[4]	[1, 1-10]	[2.5, 1 – 20]	
Eosinophils (10⁹/L), median (IQR)	0.1 (0.07 - 0.21)	0.26 (0.14 - 0.45) ^{††}	0.49 (0.25 - 0.74) ^{§§}	<0.0001
IgE (kU/L), geometric mean (95% confidence interval)	24.6 (13.3 - 45.8)	159.8 (89.8 - 284.6)	238.5 (136.2 - 417.5) ^{***}	<0.0001

1 * Calculated by ANOVA with Bonferroni's Multiple Comparisons Test (parametric) or Kruskal-Wallis
2 with Dunn's Multiple Comparison Test (non-parametric) or t-test when only asthma groups
3 compared or Chi-squared for categorical variables (sex, atopy, theophylline, long-acting β2-agonists,
4 smoking history).

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1 **Table 2.** Demographic characteristics of the mild asthmatic, non-asthmatic cough and healthy
 2 volunteers recruited for experiments on cultured primary bronchial epithelial cells.

	Healthy volunteers (n=4)	mild asthmatics (n=4)	Non asthmatic coughers (n=4)	P value
Age, (y) Median (IQR)	35.5 (21.3-43.8)	46.0 (39.3-56.8)	58.0(45.0-72.5)	0.05
Sex, M/F	1/3	0/4	0/4	0.336
Duration of cough (m) Mean \pm SD	N/A	49.5 (30.0)	57.5 (27.0)	0.950
FEV1 (L/min) Mean \pm SD	3.45 (0.65)	2.52 (0.29)	2.3 (0.34)*	0.014
FEV1, % pred Mean \pm SD	107.3 (10.59)	105.6 (15.23)	118.0 (21.28)	0.530
FEV1/ FVC Mean \pm SD	86.6 (10.6)	82.73 (4.11)	80.2 (3.2)	0.446
Atopy (n)	0	0	0	N/A
Inhaled steroid (n)	0	4	0	N/A
Prior smoking (n)	0	0	0	N/A

3 Calculated by ANOVA with Bonferroni's Multiple Comparisons Test (parametric) or Kruskal-Wallis
 4 with Dunn's Multiple Comparison Test (non-parametric) or t-test when only cough groups compared
 5 or Chi-squared for categorical variables (gender).

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2 **Legends to Figures**

3 **Figure 1: A. Molecular expression of TRPV channel subtypes in human primary bronchial**
4 **epithelial cells.** Conventional RT-PCR showing the expression of TRPV family subtypes
5 including TRPV1 in PBECs from healthy controls (n=3) **B. Expression of TRPV1 protein in**
6 **PBECs.** TRPV1 specific bands were observed at the molecular weight of 95 KDa in PBECs
7 (lanes 2 to 4, upper panel). TRPV1 transfected HEK cells were used as a positive control (lane
8 1, upper panel). β -Tubulin (lower panel) was used to ascertain equal loading (Abcam 6046).
9 Results are representative of two independent experiments. **C. Gene expression microarray**
10 **analysis of bronchial brush samples from healthy, mild/moderate and severe asthmatics.**
11 Gene expression microarray analysis was performed on fresh bronchial brush samples from
12 the Leicester cohort comprising severe asthmatics (n=12), mild/moderate asthmatics (n=16)
13 and healthy subjects (n=8). TRPV1 was significantly elevated in severe asthma versus mild
14 moderate asthma (* p=0.008) and healthy volunteers (** p=0.004).

15

16 **Figure 2 -** Digital photographic images (A-D) of TRPV1 expression in JB4 sections from (A)
17 healthy control: only patchy or focal areas of low intensity epithelial expression; (B)
18 moderate asthmatic: majority of epithelial and submucosal cells express antigen; (C) severe
19 asthmatic: widespread and strong epithelial and submucosal cell expression. Sections
20 incubated in the absence of antibodies showed no staining; a representative image is shown
21 in (D). Representative immunohistochemistry shown at X 400 magnification

22

1 **Figure 3** – Confocal microscopy images (A-C) of double staining immunocytochemistry
2 demonstrating localisation of TRPV1 channels in bronchial epithelium (healthy subject).
3 Bronchial epithelial cells identified by positive red staining with the cytokeratin antibody
4 AE1/AE3 (A) were also positive for the TRPV1 (green) (B) and co-localisation was evident
5 when the images were merged (yellow) (C). Representative immunohistochemistry shown
6 at X 400 magnification

7
8 **Figure 4:** TRPV1 and ER colocalisation in human bronchial epithelial cells. Representative
9 confocal microscopy images at X 40 magnification (A-C) showing immunohistochemistry
10 staining of bronchial epithelial cells positive for TRPV1 (A) and ER (B) markers. Colocalisation
11 was evident when the images were merged (C; orange/yellow). DAPI stained blue nuclei (C).
12 Scale bar = 100 μ m.

13
14
15 **Figure 5** Quantitative immunohistochemical epithelial expression for TRPV1 in bronchial
16 biopsies. TRPV1 expression is increased in the bronchial epithelium of severe asthmatic
17 patients. There was a trend to increased TRPV1 expression with severity of asthma ($p=0.001$,
18 Kendall's tau-B statistic).

19 **Figure 6.** Whole-cell PBEC (healthy subject) membrane current responses to capsaicin
20 application (50 μ M) measured by voltage-steps. **A.** Superimposed current traces evoked by 1
21 s voltage steps before, in the presence of, and after capsaicin wash-out, as indicated. Dotted
22 line indicates zero-level current. **B.** Mean normalised I-V relationship of background current

1 (n=4). **C.** I-V relationships for the current amplitude measured at the end of each voltage
2 step in the experiment illustrated in panel A.

3 **Figure 7.** Time course of the capsaicin membrane current response. **A.** Current
4 amplitude was monitored by applying voltage ramps from -100 to 140 mV at 10 s interval.
5 Circles show current amplitude at 140 mV. **B.** The I-V relationships were derived from the
6 ramps applied before, at the peak response and 3 min after wash-out of the drug as
7 indicated. **C.** Mean normalised I-V relationships measured before (squares) and after
8 (circles) capsaicin application (n=3). In each cell, current amplitude at 140 mV in control was
9 normalised as 1.0.

10 **Figure 8.** Whole-cell PBEC (healthy subject) membrane current responses to reduced
11 external pH measured by voltage ramps. **A.** Superimposed current traces evoked by voltage
12 ramps from -100 to 140 mV at 10 s interval in pH 7.4 and pH 6.4 (\pm capsazepine; 5 μ M)
13 external solutions as indicated. The dotted line indicates zero-level current. **B.** Mean \pm SEM
14 I-V relationship (normalized for cell capacitance) of capsazepine-sensitive current (n=4).

15 **Figure 9.** Capsaicin induces IL-8 release from human PBECs (asthmatic and non-asthmatic
16 subjects). PBECs treated (24 hours) with 10, 25 or 50 μ M capsaicin or control medium.
17 Values are expressed as the mean release over basal \pm SEM. **A.** Capsaicin-induced IL-8
18 release ***p<0.001 compared to control; Kruskal-Wallis test with Dunn's multiple
19 comparison (n=12 individual experiments). **B.** Capsazepine inhibits capsaicin induced IL-8
20 release from human PBECs. PBECs were pre-treated for 20 min with 10 μ M capsazepine
21 then with 10, 25 or 50 μ M capsaicin or control medium with or without capsazepine. Values
22 are expressed as mean release over basal \pm SEM. Capsazepine significantly inhibited

1 capsaicin-induced IL-8 release compared to 25 μM (* $p < 0.001$) and and 50 μM (** $p < 0.001$)
2 capsaicin alone; one way ANOVA with Bonferroni's multiple comparison test (n=9 individual
3 experiments).

4

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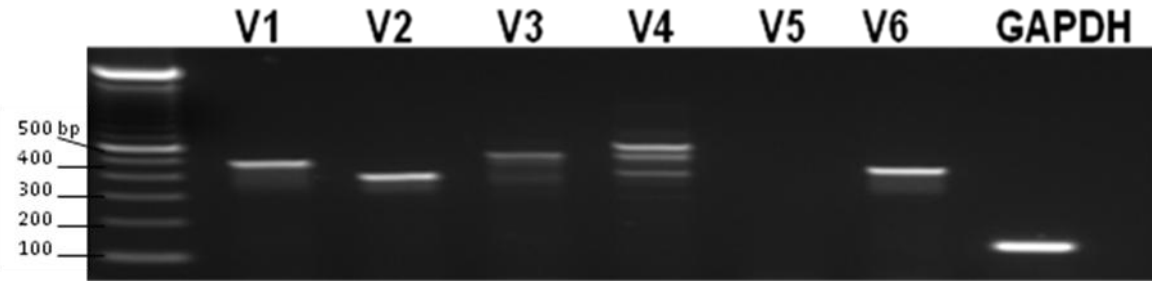
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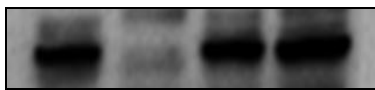
Figure 1

A



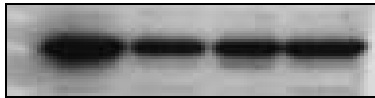
B

1 2 3 4



95 KDa

TRPV1 95 kDa



50 KDa

B-Tubulin 50 kDa

C

Gene expression microarray analysis of bronchial brush samples healthy, mild/moderate and severe asthmatics

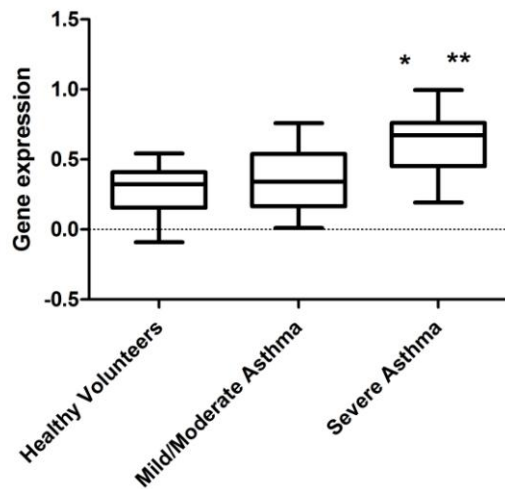


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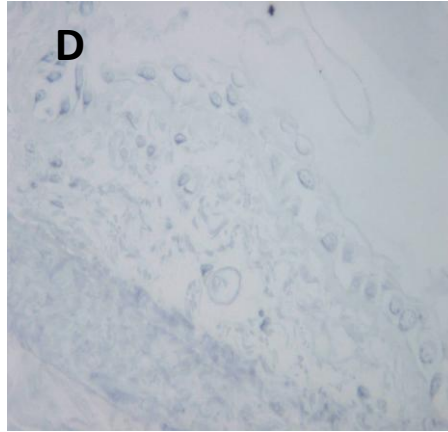
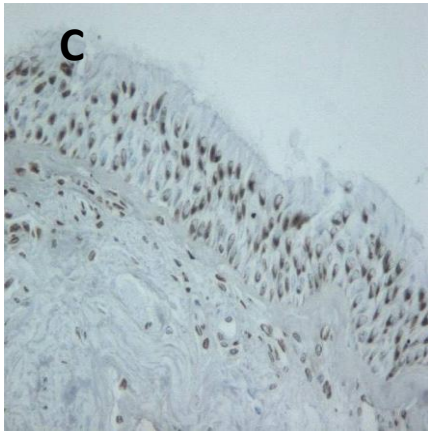
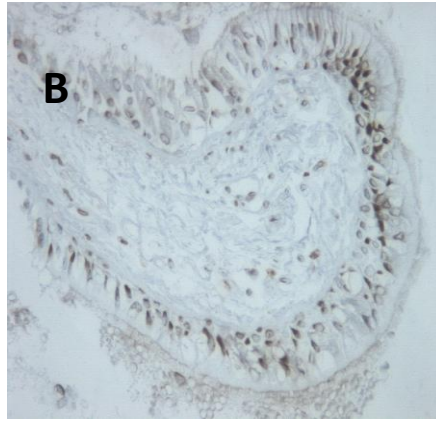
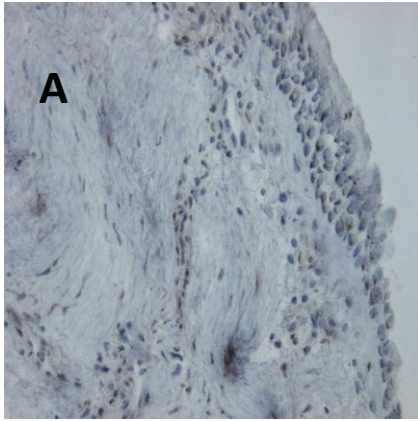


Figure 3

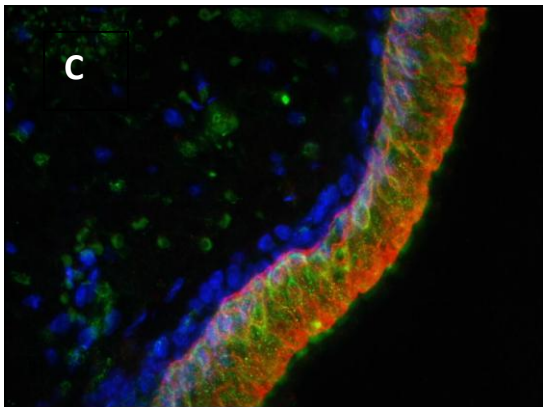
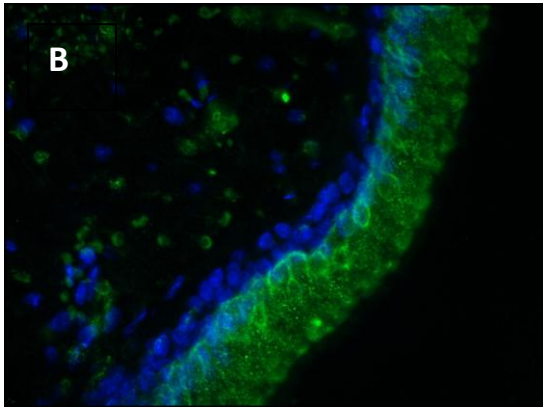
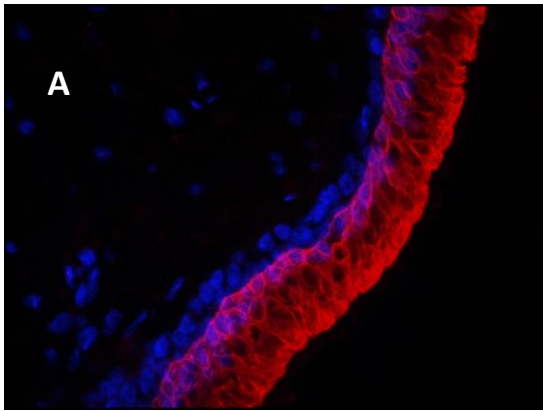


Figure 4

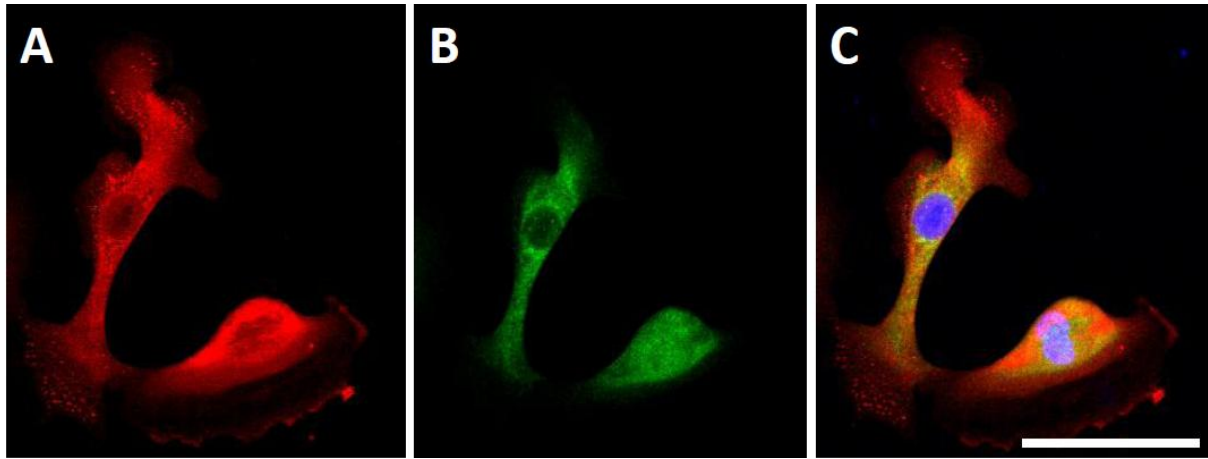


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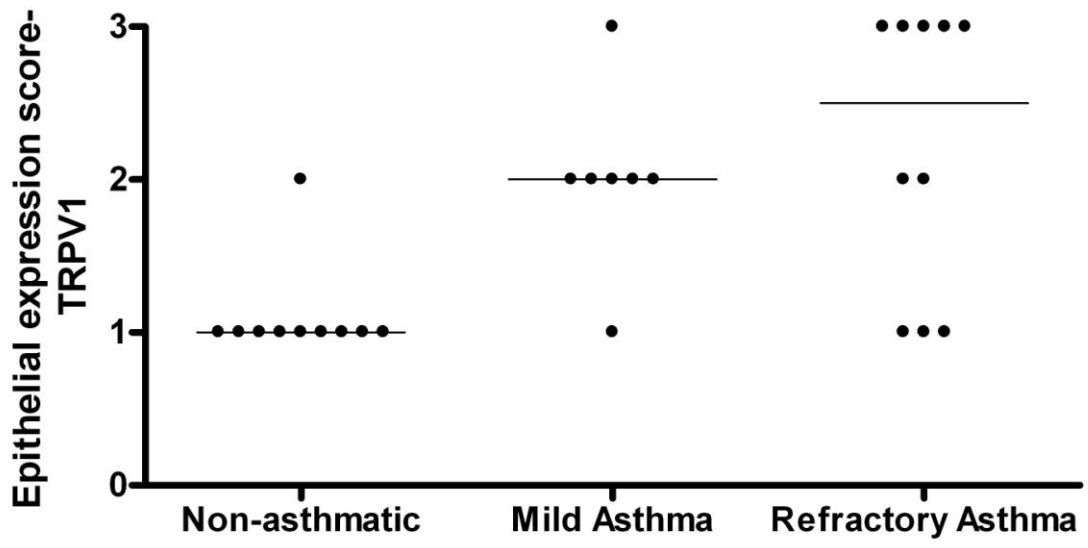


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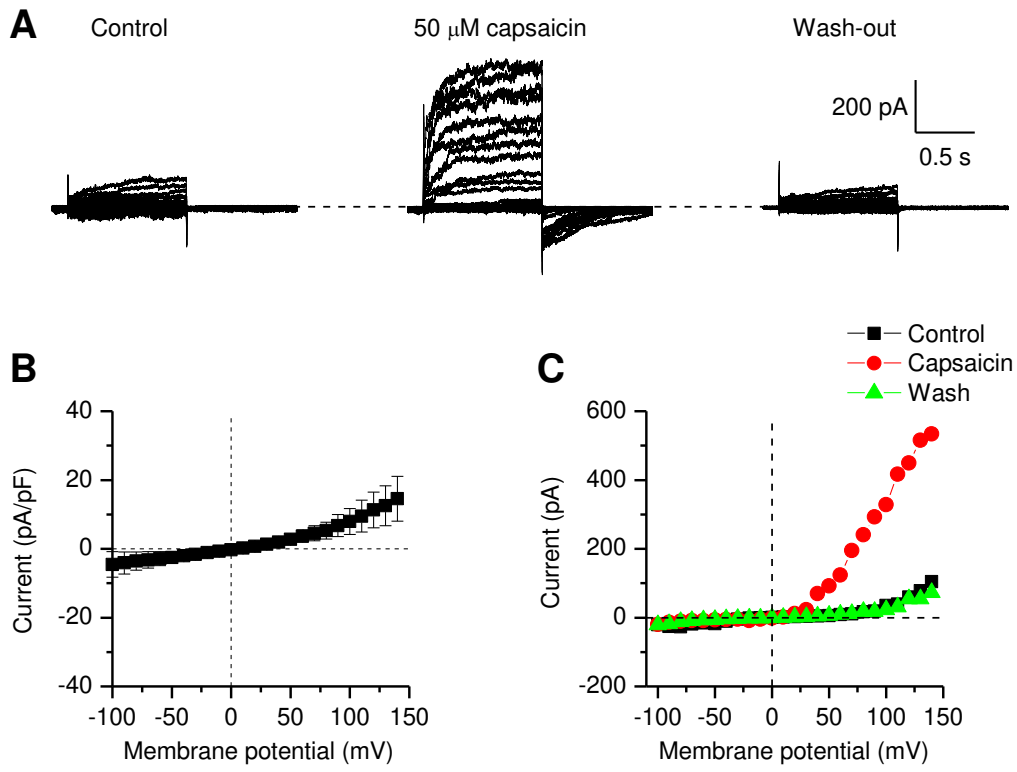


Figure 7

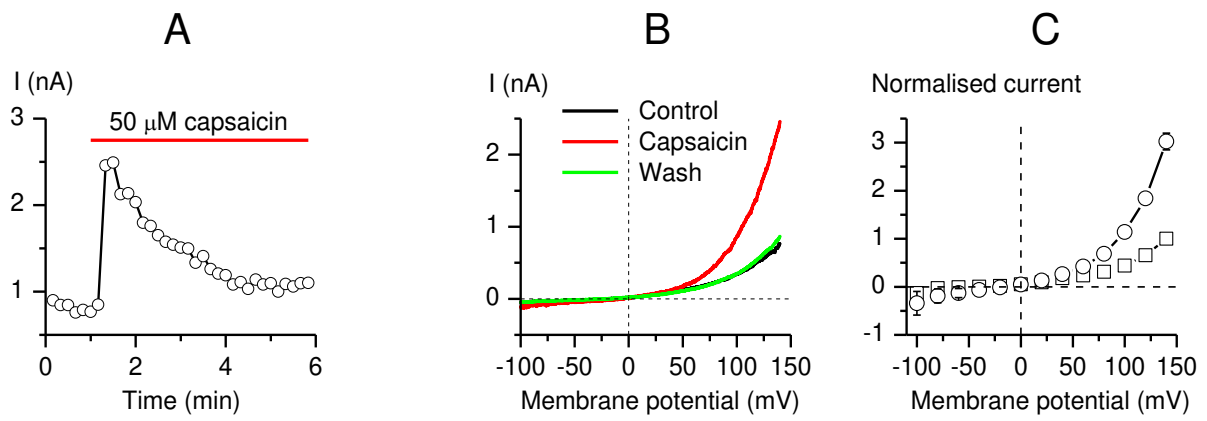


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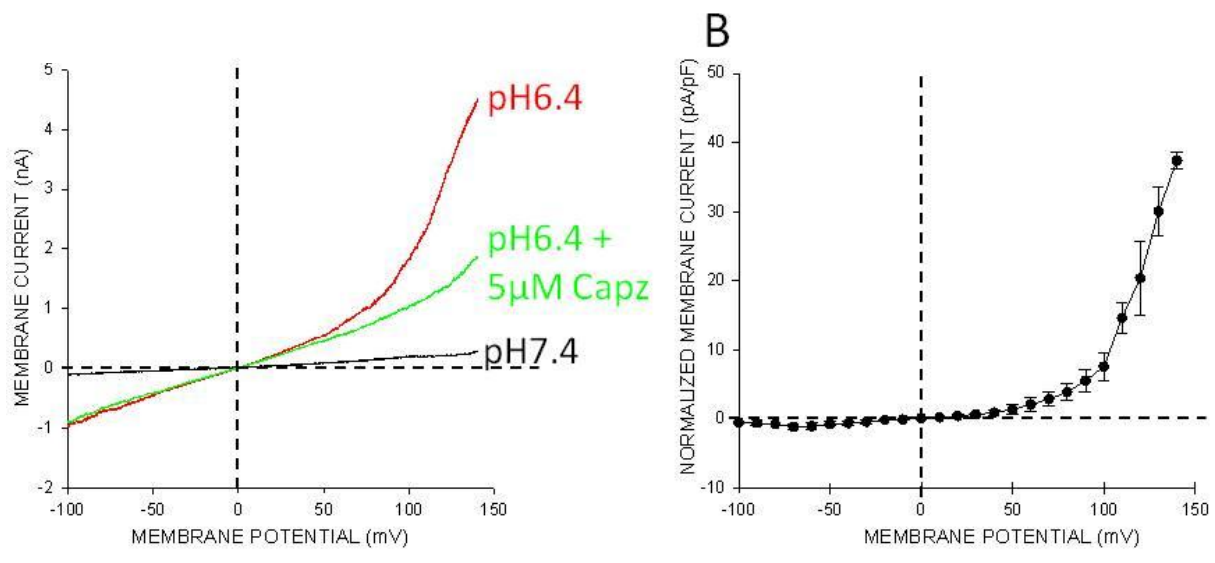
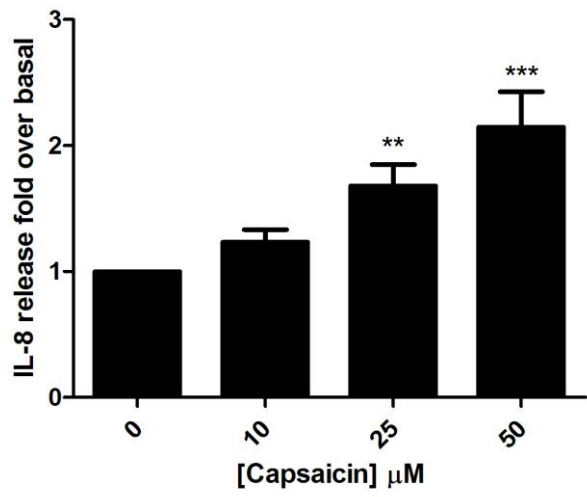
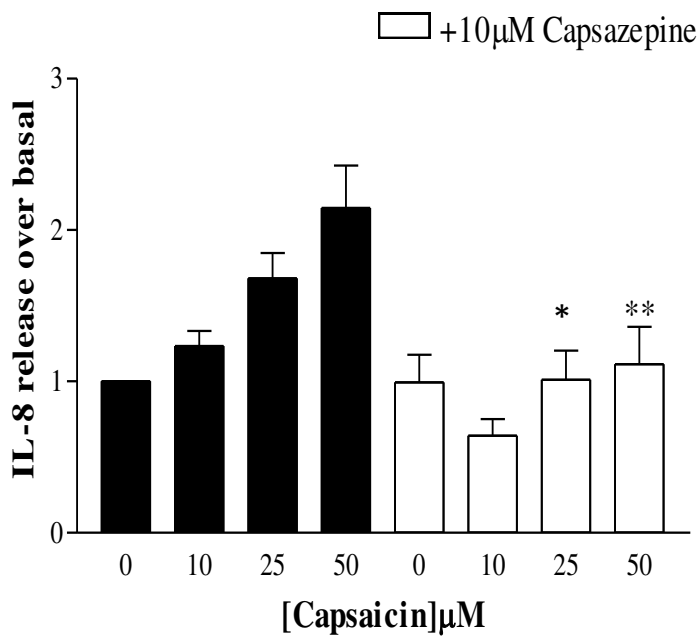


Figure 7

A



B



ONLINE DATA REPOSITORY

Increased expression of bronchial epithelial transient receptor potential vanilloid 1 channels in severe asthma

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Study Subjects

The studies were approved by the Research Ethics Committee of both Institutions (Belfast - Office of Research and Ethics Committee of Northern Ireland reference 06/NIR02/114 and 09/NIR02/51, Leicester - Leicestershire, Northamptonshire, & Rutland Research Ethics Committee reference 04/Q2502/74). Written informed consent was gained from all participants prior to their involvement.

Bronchial biopsies:

Belfast Cohort: Participants with refractory asthma were recruited from the Belfast City Hospital where systematic evaluation protocols ensure patients have well-characterised refractory asthma. All participants had persisting symptoms despite treatment at Step 4 or 5 of the GINA guidelines Global Initiative for Asthma (GINA) (E1). Mild to moderate asthmatic and normal participants were recruited by advertisement and had a clinical diagnosis of asthma with a current history of recurrent wheezing and response to asthma medication. Healthy volunteers had no history of asthma or persistent respiratory symptoms and normal lung function. All asthmatic participants were clinically stable on their usual medication at the time of bronchoscopy, and all participants had an FEV1 of greater than 60% predicted. Demographic details are shown in Table 1 of the main paper.

Leicester cohort: For the gene expression studies, asthmatic subjects (n=36) and healthy volunteers (n=12) were recruited from respiratory clinics including the Leicester Difficult Asthma Clinic, from staff at Glenfield Hospital, and from the general population through advertisement (Leicester cohort). Subjects with asthma gave a suggestive history and had objective evidence of variable airflow obstruction as indicated by one or more of the

following: (1) methacholine airway hyperresponsiveness ($PC_{20} FEV_1 < 8 \text{ mg/mL}$), (2) >15% improvement in FEV_1 10 minutes after 200 μg inhaled salbutamol, (3) peak expiratory flow (>20% maximum within-day amplitude from twice-daily peak expiratory flow measurements over a period of 14 days). Subjects underwent spirometry; allergen skin prick tests for *Dermatophagoides pteronyssinus*, dog, cat, grass pollen and *Aspergillus fumigatus*; a methacholine inhalation test using the tidal breathing method; and sputum induction using incremental concentrations of nebulized hypertonic saline (i.e., 3%, 4%, and 5%, each for 5 min)(E2). Subjects with asthma also kept a diary card for 2 weeks prior to bronchoscopy, recording daytime and night-time symptoms, daily short-acting β_2 -agonist use and twice daily peak expiratory flow (PEF). Asthma severity was defined by British Guideline on the Management of Asthma treatment steps (mild = step 1, β_2 -agonist only; moderate = steps 2 and 3, inhaled corticosteroid $\leq 800 \text{ mg}$ beclomethasone equivalent per day \pm long-acting β_2 -agonist; severe = step 4 and 5) (E3). Of the 16 severe patients at step 4/5, 13 met the American Thoracic Society criteria for refractory asthma (E4).

In both cohorts, all participants were current non-smokers with no upper or lower respiratory tract infection in the 6 weeks prior to their bronchoscopy. All asthmatic participants were clinically stable on their usual medication at the time of bronchoscopy. Demographic details for each cohort are shown in Table 1 in the main manuscript.

Primary bronchial epithelial cell (PBEC) cultures: To investigate the functional role of TRPV1 we performed a series of experiments on primary bronchial epithelial cell (PBEC) cultures

from bronchial brushings obtained from mild asthmatic (n=4) and non asthmatic patients with chronic cough (n=4) referred to the Belfast City Hospital specialist cough clinic (E5). Healthy volunteers (n=4) were recruited by local advertisement. Demographic details for this cohort are shown in Table 2 in the main manuscript.

Bronchoscopy and samples

Bronchoscopy was performed using a standard technique. In brief, after intravenous sedation and local anaesthesia with topical lignocaine, bronchial biopsies and bronchial brushings were obtained from lobar / segmental airways. One biopsy from each participant was immediately placed in RNA preservative (RNAlater, Ambion, Austin, TX) and submitted for microarray analysis and qPCR and 3 biopsies placed immediately into dry acetone containing protease inhibitors (iodoacetamide and PMSF) and fixed overnight at -20°C. Brushings were placed in RNAlater or Promocell™ medium containing penicillin/streptomycin and Primocin™ for cell culture (Promocell UK). PBECs were cultured using methods developed in our laboratory which resulted in homogenous cultures of basal epithelial cells (E6).

For immunohistochemistry (IHC), biopsy samples were processed using local protocols. Twenty eight biopsies (10 healthy volunteer, 8 mild asthmatic and 10 severe asthmatic biopsies) were available for immunohistochemistry. Infiltration and embedding with JB4 resin was performed as previously described (E7). Two µm sections were cut from each biopsy. Endogenous peroxidase was blocked with 0.3% hydrogen peroxide. All sections were pressure cooked in 0.01M Tris-EDTA pH9.0 on full steam for 3 minutes for antigen retrieval. Sections were incubated with TRPV1 polyclonal antibody (1:200, Abcam), overnight at 4°C, then primary antibodies were detected by incubation for 30 min at RT in peroxidase-labelled

EnVision anti-rabbit secondary antibody (Dako, Ely, UK) using 3,3'-diaminobenzidine (DAB; Dako, Ely, UK) as chromagen. On selected tissue sections from this series, immunofluorescence was carried out to permit high resolution analysis of TRPV1 antigen distribution. Sections were treated as for IHC and following incubation in a mix of TRPV1 and a monoclonal antibody to the epithelial specific antibody AE1AE3 (1:50, Dako). Antigen binding sites were detected with a mixture of anti-mouse Alexa 568 and anti-rabbit Alexa 488 (Invitrogen). Sections were counterstained with DAPI hardset mounting medium (Vector). All fluorescently stained slides were assessed and digital fluorescent images acquired with a Leica DFC350 FX digital camera and processed using Leica FW4000 software. Immunostained sections were evaluated by two independent persons, following which results were discussed and consensus scoring applied to any conflicting results. A 4-point scoring system was used: 0 - no detectable expression in epithelium or submucosa; 1 - patchy or focal areas of low intensity epithelial expression, not all cells expressing positivity for protein, scattered positivity in submucosa; 2 - moderate / majority of epithelial and submucosal cell expression; 3 - widespread and strong epithelial and submucosal cell expression.

TRPV1 and ER colocalisation in human bronchial epithelial cells.

Human bronchial epithelial cells were grown on coverslips and fixed in 4% paraformaldehyde (PFA) for 30 minutes at RT. Cells were briefly washed in Phosphate Buffered Saline (PBS) before being permeabilised with 0.1% Triton X-100 in PBS for 20 minutes at RT. Cells were blocked with 10% Bovine Serum Albumin (BSA) in PBS for 1 hour at RT and incubated with primary antibodies, mouse monoclonal anti-KDEL (1:500), ER marker and rabbit polyclonal anti-VR1 (1:200), TRPV1 marker (AB12223 and Ab63083 respectively;

Abcam, Cambridge, UK) overnight at 4⁰C, followed by incubation for 1 hour at RT with secondary antibodies, Alexa Fluor goat anti-mouse 488 (1:500; Invitrogen) and Alexa Fluor goat anti-rabbit 568 (1:500; Invitrogen). Cell nuclei were stained using Vectashield mounting medium containing DAPI (Vector Labs, Peterborough, UK). Staining was visualised and images captured using Nikon EZ-C1 confocal system running C1 acquisition software (Nikon UK Ltd. Surrey, UK) at x40 magnification.

Gene expression analyses

RNA was isolated from homogenized bronchial biopsies and real-time PCR (qPCR) was performed as described previously (E8). TaqMan[®] Gene Expression Assays (Applied Biosystems, Foster City, CA) were purchased and conducted per manufacturer's instructions for TRPV1. RNA was amplified (Ambion,) for Agilent (Santa Clara, CA) two color Whole Human Genome (WHG) 4x44k gene expression microarray analysis. Universal Human Reference RNA (Stratagene, La Jolla, CA) was used for the reference channel. Probe intensities were transformed as log₂ ratios of test and reference channels calculated by the Agilent Feature Extraction software, protocol GE2-v5_95 (Agilent). Flagged outliers were not included in any subsequent analyses. All gene expression analyses and plotting were performed using the R Project software package, version 2.10.1 (refer to <http://www.R-project.org>).

Quantitative Real-Time PCR

Bronchial brushings

Real-time PCR (qRT-PCR) analyses were performed using RNA from homogenized bronchial brushings (36 subjects; 10 HV, 16 MA and 15 SA) using TaqMan® Gene Expression Assays (Applied Biosystems, Foster City, CA) as previously described. The delta-delta Ct method was used to calculate relative fold expression with GAPDH as housekeeping gene

PBEC cultures

PBECs were trypsinised and cells lysed using Qiagen RLT buffer (Qiagen, Hilden, Germany) before homogenisation using QIAshredders (Qiagen, Hilden, Germany). RNA was then extracted using the RNeasy Mini Kit (Qiagen, Hilden, Germany). RNA concentration was measured with the nanodrop spectrophotometer (Thermo Scientific, Wilmington, USA) and 1µg RNA was used to synthesise cDNA using the high capacity cDNA reverse transcription kit (Applied Biosystems, Foster City, CA). PBEC cDNA was mixed with Power Sybr Green PCR master mix (Applied Biosystems, Foster City, CA) and either GAPDH or TRPV1 forward and reverse primers (Table E1). The following PCRs were performed in a 7300 Real Time PCR system (Applied Biosystems, Foster City, CA). Melting curves showed formation of only one gene product per primer pair. Normalisations and calibrations of the data from all studies were performed with the $2^{-\Delta\Delta C_T}$ method as described in Applied Biosystems User Bulletin No. 2 (P/N 4303859) (27). Delta C_T was calculated as target gene expression (C_T) minus internal housekeeping control, GAPDH gene expression (C_T). Human Universal Reference RNA (Clontech, P/N 639654) was prepared as cDNA and utilised as plate calibrators. DeltaDelta C_T was calculated as sample Delta C_T minus plate calibrator Delta C_T. GAPDH and TRPV1 forward and reverse primers are detailed in table E1 below.

Western Blotting

PBECs were collected and snap frozen, homogenised and lysed in RIPA buffer supplemented with protease inhibitor cocktail (Roche Diagnostics, Mannheim, Germany). Protein concentration was determined using BCA protein assay kit (Thermo Scientific, Wilmington, USA). 30µg of total protein was separated on the 8% SDS-Polyacrylamide gel by electrophoresis and transferred to Hybond-P PVDF membrane (GE Healthcare Life Sciences, Buckinghamshire, UK). The membrane was blocked for 30 min in 3% BSA/ PBS and then probed overnight in 3% BSA/PBS with a rabbit polyclonal TRPV1 antibody (ab63083; Abcam, Cambridge, UK), at a dilution of 1:3000. Followed by incubation for 1 h with the secondary antibody, HRP-labelled anti-rabbit (1:3000; Bio-Rad Laboratories, Hercules, CA, USA). Images were obtained using UVP bioimager (UVP, Cambridge, UK). Benchmark Pre-stained protein standards (Invitrogen, Paisley, UK) were used to determine molecular mass.

Patch clamp experiments

Whole-cell currents were recorded using borosilicate heat-polished patch pipettes (2-3 MΩ resistance when filled with the pipette solution) and an Axopatch 200B amplifier (Molecular Devices, Union City, CA, USA) interfaced to Digidata 1322A using the pClamp 9 software (Molecular Devices). Series resistance was compensated by about 70%. Holding potential was -40 mV. Steady-state current-voltage (I-V) relationships were measured by applying either voltage steps (test potentials from -100 to +140 mV with a 20 mV increment) or slow voltage ramps from -100 to +140 mV. To monitor the time course of TRPV1 activation by

capsaicin, the voltage ramps were applied at 10 s interval. The I-V curves obtained by the voltage ramp and step protocols were similar confirming steady-state channel activity at each test potential when ramps were used.

For experiments involving the addition of capsaicin the external solution contained (mM): NaCl 120, glucose 12, HEPES 10, pH 7.4 (adjusted with NaOH). Pipette solution contained (mM): CsCl 80, MgATP 1, creatine 5, GTP 1, D-glucose 5, HEPES 10, BAPTA 10, CaCl₂ 4.6 ([Ca²⁺]_i=100 nM), pH 7.4 (adjusted with CsOH). For experiments involving altered pH the external solution contained (mM): NaCl 150, CsCl 6, MgCl₂ 1.3, CaCl₂ 2, Glucose, 5; HEPES, 10; pH7.4 or 6.4 with Tris. Pipette solution contained (mM): CsOH 130, D Gluconic Acid 120, MgCl₂ 1, EGTA 0.5 ([Ca²⁺]_i= 30 nM), pH 7.2 with Tris.

IL-8 ELISA

IL-8 concentrations were measured from aliquots of culture medium taken 24 h following treatment with capsaicin (Enzo Life Sciences, Exeter, UK) using a commercial ELISA kit from R & D Systems Europe (Abingdon, UK).

Data Analysis

Data are presented as means ±SEM. For statistical analysis a one way ANOVA was performed with Bonferroni test for multiple comparisons. Kendall's tau b statistical analysis was used to examine the trend in biopsy expression by participant group. Patch clamp data were analysed and plotted using Origin 8 (OriginLab, Northampton, MA, USA). A P value < 0.05 was considered significant.

RESULTS

Real time PCR (qRT PCR) analysis of fresh bronchial brush samples from the Leicester cohort comprising healthy, mild/moderate and severe asthmatics. Analysis was performed on fresh bronchial brush samples from the Leicester cohort comprising severe asthmatics (n=15), mild/moderate asthmatics (n=16) and healthy subjects (n=10). TRPV1 mRNA was detected in all of the epithelial brush samples. There was no difference in median (IQR) relative fold expression of TRPV1 mRNA calculated as Delta Delta C_T values between groups: -1.262 (-3.598, 2.518), (-2.523 (-4.040, 1.362) and -1.289 (-4.746, 1.624) in healthy volunteers, mild / moderate asthmatics and severe asthmatics respectively (see figure E1 below).

Online Figure Legend

Figure E1: Real time PCR (qRT PCR) expression of TRPV1 in fresh bronchial epithelial brush samples from healthy volunteers, mild/moderate asthmatics and severe asthmatics.

Analysis of fresh bronchial brush samples from the Leicester cohort comprising healthy (n=10), mild/moderate (n=16) and severe asthmatics (n=15). Data presented as median (IQR) relative fold expression of TRPV1 mRNA calculated as delta delta C_T values.

References

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Table E1. Forward and reverse primers for TRPV1 and GAPDH

Primer name	Forward	Reverse	Size (bp)
TRPV1	GCCTGGAGCTGTTCAAGTTC	GCCTGAAACTCTGCTTGACC	452
TRPV2	CCAGCTCTCCAGTTTTTCAGG	GTTTCAGCACAGCCTTCATCA	365
TRPV3	TTCCTCATGCACAAGCTGAC	TGCGCTTCACAAAGTCATTC	490
TRPV4	CCCGTGAGAACACCAAGTTT	GAAGAGCGTAATGACCTCGC	374
TRPV5	TTGCTCAGTTGCTGTCATCC	GCTCCTCTTTGTCGAACGTC	307
TRPV6	TCACTTCGCTTCCTGGAAC	ACACGCTTTCCACAAGCTCT	401
GAPDH	GCTCTCTGCTCCTCCTGTTC	AAATCCGTTGACTCCGACC	100

Source: Invitrogen

Online Repository Figure E1. Real time PCR (qRT PCR) expression of TRPV1 in fresh bronchial epithelial brush samples from healthy volunteers, mild/moderate asthmatics and severe asthmatics.

UK EPITHELIAL BRUSHINGS

