

#### TITLE:

Increased periostin associates with greater airflow limitation in patients receiving inhaled corticosteroids.

### AUTHOR(S):

Kanemitsu, Yoshihiro; Matsumoto, Hisako; Izuhara, Kenji; Tohda, Yuji; Kita, Hideo; Horiguchi, Takahiko; Kuwabara, Kazunobu; ... Yokoyama, Tetsuji; Niimi, Akio; Mishima, Michiaki

### CITATION:

Kanemitsu, Yoshihiro ...[et al]. Increased periostin associates with greater airflow limitation in patients receiving inhaled corticosteroids.. The Journal of allergy and clinical immunology 2013, 132(2): 305-312.e3

### **ISSUE DATE:**

2013-08

URL:

http://hdl.handle.net/2433/179319

### RIGHT:

© 2013 American Academy of Allergy, Asthma & Immunology. Published by Mosby, Inc.; この論文は出版社版でありません。引用の際には出版社版をご確認ご利用ください。; This is not the published version. Please cite only the published version.



- 1 Increased periostin associates with greater airflow limitation in patients receiving
- 2 inhaled corticosteroids

- 4 Yoshihiro Kanemitsu MD\*<sup>1,2</sup>, Hisako Matsumoto MD, PhD\*<sup>1,2</sup>, Kenji Izuhara MD,PhD³,
- 5 Yuji Tohda MD, PhD<sup>2,4</sup>, Hideo Kita MD, PhD<sup>2,5</sup>, Takahiko Horiguchi MD, PhD<sup>2,6</sup>, Kazunobu
- 6 Kuwabara MD, PhD<sup>2,6</sup>, Keisuke Tomii MD, PhD<sup>2,7</sup>, Kojiro Otsuka MD, PhD<sup>1,2,7</sup>, Masaki
- 7 Fujimura MD, PhD<sup>2,8</sup>, Noriyuki Ohkura MD<sup>2,8</sup>, Katsuyuki Tomita MD, PhD<sup>2,4</sup>, Akihito
- 8 Yokoyama MD, PhD<sup>2,9</sup>, Hiroshi Ohnishi MD, PhD<sup>2,9</sup>, Yasutaka Nakano MD, PhD<sup>2,10</sup>, Tetsuya
- 9 Oguma MD, PhD<sup>2,10</sup>, Soichiro Hozawa MD, PhD<sup>2,11</sup>, Tadao Nagasaki MD<sup>1</sup>, Isao Ito MD,
- 10 PhD<sup>1</sup>, Tsuyoshi Oguma MD<sup>1</sup>, Hideki Inoue MD<sup>1</sup>, Tomoko Tajiri MD<sup>1</sup>, Toshiyuki Iwata MD<sup>1</sup>,
- 11 Yumi Izuhara MD<sup>1</sup>, Junya Ono BS<sup>12</sup>, Shoichiro Ohta MD, PhD<sup>13</sup>, Mayumi Tamari MD,
- PhD<sup>14</sup>, Tomomitsu Hirota DDS, PhD<sup>14</sup>, Tetsuji Yokoyama MD, PhD<sup>15</sup>, Akio Niimi MD,
- 13 PhD<sup>1,2,16</sup> and Michiaki Mishima MD, PhD<sup>1,2</sup>

- <sup>1</sup>Department of Respiratory Medicine, Graduate School of Medicine, Kyoto University,
- 16 Kyoto, Japan
- 17 <sup>2</sup>Kinki Hokuriku Airway disease Conference (KiHAC)
- <sup>3</sup>Division of Medical Biochemistry, Department of Biomolecular Sciences, Saga Medical
- 19 School, Saga, Japan
- <sup>4</sup>Department of Respiratory Medicine and Allergology, Faculty of Medicine, Kinki University,
- 21 Osaka, Japan
- <sup>5</sup>Department of Respiratory Medicine, Takatsuki Red Cross Hospital, Osaka, Japan
- <sup>6</sup>Department of Respiratory Internal Medicine, Fujita Health University Second Educational
- Hospital, Aichi, Japan
- <sup>7</sup>Department of Respiratory Medicine, Kobe City Medical Center General Hospital, Hyogo,
- 26 Japan
- <sup>8</sup>Department of Respiratory Medicine, Cellular Transplantation Biology, Kanazawa





- 28 University Graduate School of Medicine, Kanazawa, Japan
- <sup>9</sup>Department of Hematology and Respiratory Medicine, Kochi University, Kochi, Japan
- 30 <sup>10</sup>Division of Respiratory Medicine, Department of Internal Medicine, Shiga University of
- 31 Medical Science, Shiga, Japan
- 32 <sup>11</sup>Hiroshima Allergy and Respiratory Clinic, Hiroshima, Japan
- 33 <sup>12</sup>Shino-Test Corporation, Kanagawa, Japan
- 34 <sup>13</sup>Department of Laboratory Medicine, Saga Medical School, Saga, Japan
- 35 <sup>14</sup>Laboratory for Respiratory Diseases, Center for Genomic Medicine, RIKEN, Yokohama,
- 36 Kanagawa, Japan

- 37 <sup>15</sup>Department of Health Promotion, National Institute of Public Health, Wako, Saitama, Japan
- 38 <sup>16</sup>Department of Medical Oncology and Immunology, Nagoya City University School of
- 39 Medical Sciences, Aichi, Japan
- \*YK and HM contributed equally to this study.
- 42 Correspondence should be addressed to Hisako Matsumoto MD, PhD
- 43 Department of Respiratory Medicine, Graduate School of Medicine, Kyoto University
- 44 53 Kawahara-cho, Shogoin, Sakyo-ku, Kyoto 606-8507, Japan
- 45 E-mail: hmatsumo@kuhp.kyoto-u.ac.jp
- 46 Funded by KiHAC, as a project of 2009 KiHAC Respiratory Medicine Group and the
- 47 Adaptable and Seamless Technology Transfer Program through target-driven R&D, JST.

KURENAI KI



48

49

50

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

67

68

69

70

### Abstract

**Background:** Periostin, an extracellular matrix protein, contributes to subepithelial thickening in asthmatic airways, and its serum levels reflect airway eosinophilic inflammation. However, the relationship between periostin and the development of airflow limitation, a functional consequence of airway remodeling, remains unknown. **Objective:** To determine the relationship between serum periostin levels and pulmonary function decline in asthmatic patients on inhaled corticosteroid (ICS) treatment. **Methods:** 224 asthmatic patients (average age 62.3 years) treated with ICS for at least 4 years were enrolled. Annual changes in forced expiratory volume in one second (FEV<sub>1</sub>), from at least one year after the initiation of ICS treatment to the time of enrollment or later (average 16.2 measurements over 8 years per individual), were assessed. At enrollment, clinical indices, biomarkers including serum periostin, and periostin gene polymorphisms were examined. Associations between clinical indices or biomarkers and a decline in FEV<sub>1</sub> of 30 mL·yr<sup>-1</sup> or greater were analyzed. **Results:** High serum periostin levels ( $\geq 95 \text{ ng/mL}$ ) at enrollment, the highest treatment step, higher ICS daily doses, a history of admission due to asthma exacerbation, comorbid or a history of sinusitis, and ex-smoking were associated with a decline in FEV<sub>1</sub> of 30 mL·yr<sup>-1</sup> or greater. Multivariate analysis revealed that high serum periostin, the highest treatment step, and ex-smoking were independent risk factors for the decline. Polymorphisms of periostin gene were related to higher serum periostin levels (rs3829365) and a decline in FEV<sub>1</sub> of 30  $mL \cdot yr^{-1}$  or greater (rs9603226). **Conclusions:** Serum periostin appears to be a useful biomarker for the development of airflow limitation in asthmatic patients on ICS.

71

72

73

### Clinical implications (25 words)

Serum periostin levels reflect greater FEV<sub>1</sub> decline in asthmatic patients on inhaled





93

4

corticosteroid treatment. POSTN gene polymorphisms may also be helpful for identifying 74 rapid FEV<sub>1</sub> decliners. 75**Key words** 76 Asthma, inhaled corticosteroids, lung function decline, periostin, *POSTN* gene polymorphism, 77sinusitis, treatment step 78 79 80 **Abbreviations** ACT: asthma control test 81 82 ECP: eosinophil cationic protein FAS I: fasciclin I 83 FEV<sub>1</sub>: forced expiratory volume in one second 84 FVC: forced vital capacity 85 hsCRP: high sensitivity C-reactive protein 86 ICS: inhaled corticosteroids 87 88 IgE: immunoglobulin E IL: interleukin 89 ROC: receiver operating characteristic 90 SNP: single-nucleotide polymorphism 91

Total word counts for the text and the abstract are 3800 and 258 words, respectively.

TGF-β: transforming growth factor beta







- 95 Capsule summary (32 words)
- This is the first study to identify a relationship between high serum periostin and greater
- 97 annual decline in FEV<sub>1</sub>, which sheds new light on serum periositin as a useful biomarker in
- 98 asthma.



### Introduction

Airway inflammation and remodeling are key features of asthma that have been demonstrated by pathological¹ and radiological findings²-³. Physiologically, patients with asthma show a greater decline in pulmonary function than subjects without asthma⁴. Studies that were mostly conducted in the era before inhaled corticosteroids (ICS) demonstrated that more severe symptoms or severe exacerbations⁵-7, long-standing asthma³, and smoking history⁴-8 were moderate to strong risk factors for greater decline in pulmonary function⁵. Blood and sputum eosinophilia³-10 and genetic predisposition¹¹¹-¹³ were also potential risk factors. Owing to early intervention with ICS, however, airway inflammation and the degree of annual decline in pulmonary function have been attenuated in a majority of asthmatic patients¹⁴-16. Meanwhile, a subset of patients still show accelerated decline in FEV₁ and develop irreversible airway obstruction despite adequate treatment¹¹-18. van Veen et al. found that exhaled nitric oxide of 20 ppb or higher is a predictor of accelerated decline in pulmonary function in patients with difficult-to-treat asthma¹¹8. However, other biomarkers for greater decline in FEV₁ despite treatment with ICS remain unknown.

The airway inflammation of asthma is classically characterized by infiltration and activation of eosinophils, mast cells, and Th2 cells with several mediators and Th2 cytokines, such as interleukin (IL)-4, IL-5, and IL-13<sup>19, 20</sup>. Periostin, a secreted, 90-kDa, extracellular matrix protein that is induced by IL-4 and IL-13, was originally isolated as an osteoblast-specific factor; it shares structural homology to the insect cell adhesion molecule fasciclin I (FAS I) and binds to fibronectin, tenascin-C, and collagen<sup>21, 22</sup>. In airway epithelial cells collected from patients with asthma, periostin is one of the up-regulated genes<sup>23</sup>, and its expression is correlated with thickness of the airway basement membrane<sup>24</sup>. Takayama et al. clearly demonstrated that periostin is deposited in the airway subepithelial layer in asthmatic patients. Moreover, serum periostin is identified as the single best predictor of airway eosinophilia in patients with severe asthma who remain symptomatic despite maximal ICS





treatment<sup>25</sup>. Therefore, we hypothesized that periostin would be a novel biomarker of Th2/eosinophil-driven airway inflammation and greater decline in pulmonary function, a functional consequence of airway remodeling in patients with asthma.

In this study, the effects of biomarkers and clinical indices on greater annual decline in pulmonary function in asthmatic patients on ICS treatment were examined, with the specific aim of determining the association between serum periostin levels and pulmonary function decline. Polymorphisms of the *POSTN* gene, which encodes periostin, were also examined on the hypothesis that *POSTN* gene polymorphisms may affect serum periostin levels.

### Methods

# For full details see Online Repository

### **Patients**

Patients with asthma were recruited from nine institutions belonging to the Kinki
Hokuriku Airway disease Conference where asthma specialists manage patients. Asthma was
diagnosed according to the American Thoracic Society criteria<sup>26</sup>. From September 2009 to
December 2011, patients were enrolled if they had received ICS treatment for 4 years or more,
undergone three or more pulmonary function tests when they were stable, and were free from
exacerbations for at least one month. The first pulmonary function test was performed at least
one year after the commencement of ICS treatment and at 25 years of age or older. Patients
who had smoked more than 10 pack-years, smoked in the past one year, or had other
pulmonary diseases were excluded.

This study was approved by the ethics committee of each participant institution and was registered in the UMIN Clinical Trials Registry (Registry ID UMIN000002414). Written informed consent was obtained from all participants.

### Measurements

At enrollment, patients underwent a work-up that included answering a self-completed questionnaire, spirometry, and blood tests. After enrollment, spirometry was repeated at least 6 months later for up to 12 months.

# **Self-completed questionnaire and clinical indices**

The self-completed questionnaire was composed of 4 major items, as presented in Table 1. The Asthma Control Test (ACT)<sup>TM</sup> was also scored. The treatment step at enrollment was determined according to the Global Initiative for Asthma 2010 guideline<sup>27</sup>.

KURENAI KI



# **Pulmonary function**

Spirometry was performed using an electrical spirometer, which was calibrated once a week, at each institution. Spirometry data were obtained only when patients were stable. To determine pulmonary function on daily medications, ICS and other controllers, including long-acting  $\beta_2$  agonists, leukotriene receptor antagonists, or slow-release theophylline, were not withdrawn before spirometry.

# **Measurement of systemic biomarkers**

Blood eosinophil and neutrophil counts, and serum levels of total immunoglobulin E (IgE), specific IgE against common inhaled allergens, eosinophil cationic protein (ECP), high sensitivity C-reactive protein (hsCRP), and periostin were determined.

Serum periostin levels were measured using an enzyme-linked immunosorbent assay at Shino-test (Kanagawa, Japan), as described previously<sup>28</sup>. Pooled serum periostin level data from 66 healthy subjects [mean (SD), 60.7 (16.7) years old, 40 males]<sup>28,29</sup> were used for comparison with those of asthmatic patients.

### Haplotype analysis, DNA extraction, and genotyping of the POSTN gene

A total of 47 single-nucleotide polymorphisms (SNPs) in the region of the *POSTN* gene and its upstream, total 39 kb, was captured in the HapMap Japanese data set. Haplotype analysis identified 4 major haplotypes and 2 minor haplotypes. Two minor haplotypes were grouped into the closest major haplotype, and 3 tag SNPs that determined the 4 haplotypes were identified (Figure 1).

Genomic DNA was isolated from blood cells using a QIAamp DNA Blood Mini Kit (Qiagen, Tokyo, Japan). SNPs were genotyped using a Taqman genotyping assay according to the manufacturer's instructions (Applied Biosystems, Tokyo, Japan) and analyzed using an Applied Biosystems 7300 Real-Time PCR System (Applied Biosystems).

KURENAI II



186

187

188

189

190

191

192

193

194

195

196

197

198

199

200

201

202

203

204

205

206

207

208

# Statistical analysis

Statistical analyses were performed using JMP version 9.0 (SAS Institute Inc., Tokyo, Japan). Annual changes in  $FEV_1$  ( $\Delta FEV_1$ ) were estimated for each subject by fitting a leastsquare regression line to all of his/her all available data points. Receiver operating characteristic (ROC) curve analysis was performed to determine a serum periostin cut-off value for asthmatic patients. The effects of serum biomarkers or other indices on  $\Delta FEV_1$  were estimated using a generalized linear mixed model with adjustment for sex, height, age at enrollment, and FEV<sub>1</sub> at the first measurement. The institutions were included as random effects in this model. On univariate analysis of  $\Delta FEV_1$ , the adjusted p value, i.e., q value, which was a measure of significance in terms of the false discovery rate, was obtained using R and QVALUE software<sup>30</sup> to determine spurious significance in multiple testing. The effects on the dichotomous data for a decline in FEV<sub>1</sub> of -30 mL·yr<sup>-1</sup> or greater<sup>31</sup> were similarly estimated using a generalized linear mixed model by IBM SPSS Advanced Statistics 19 (SPSS Inc., Tokyo, Japan). Multivariate analysis was performed using variables with p < 0.10 on univariate analysis, except for ICS daily maintenance dose because of its strong correlation with treatment step. On multivariate analysis, the periostin level was considered as a dichotomous variable (high or low) instead of a continuous variable. Correlation coefficients between serum periostin levels and clinical indices were estimated by fitting least-square regression lines to data, in which institutions were included as random effects. Unpaired t- and Chi-square tests were performed for comparisons of continuous and dichotomous variables, respectively. When data were not normally distributed, they were logtransformed. Data are presented as means (SD). P values  $\leq 0.05$  were considered significant.



210

211

212

213

214

215

216

217

218

219

220

221

222

223

224

225

226

227

228

229

230

231

232

233



### **Results**

### **Patients' characteristics**

Initially, 233 patients were enrolled in this study, but 9 patients were excluded: 5 with a smoking history of more than 10 pack-years and 4 who did not have enough pulmonary function data available. The demographic data of the remaining 224 patients are presented in Table 2. The mean age at enrollment was 62.3 (13.7) years. Overall, 130 (58%) had onset of asthma at 40 years or older. The average number of measurements of FEV<sub>1</sub>, follow-up period, and  $\Delta FEV_1$  of 224 patients were 16.2 (13.9) times, 8.0 (4.5) years, and -7.8 (34.6) mL·yr<sup>-1</sup>, respectively. The distribution of  $\Delta FEV_1$  in this population is shown in Figure E1 in the Online Repository. Within 2 years after diagnosis, 46% of patients started ICS treatment. At enrollment, 82% of patients took controllers such as long-acting β<sub>2</sub> agonists, leukotriene receptor antagonists, or sustained release theophylline to achieve adequate asthma control. Based on a questionnaire, adherence to medication was satisfactory; 49% of the participants never and 38% seldom forgot to take ICS or other medications. Based on ACT scores, 50% was totally controlled, and 38% scored from 20 to 24, indicating that they were well controlled at enrollment. Serum periostin levels of asthmatic patients [92.8 (38.4) ng/mL] were significantly higher than those of healthy subjects [39.1 (24.5) ng/mL, p < 0.001]. The ROC curve analysis was performed to discriminate patients with asthma who were thought to have refractory Th2 inflammation despite long-term ICS treatment from healthy subjects. The highest specificity among the 4 cut-off values tested was achieved at 95 ng/mL (0.985) in the comparison study of 224 asthmatic patients and 66 healthy subjects. Therefore a cut-off value of 95 ng/mL was used to define a high serum periostin group, although it had relatively lesser sensitivity (0.379) (see Figure E2 in the Online Repository). In asthmatic patients, 85 patients (38%) had high serum periostin levels ( $\geq 95$  ng/mL). Of the 85 patients, 40 patients (47%) were on



京都大学学術情報リボジトリ KURENAI 「「 Kyoto University Research Information Repository

treatment step 4, according to the treatment step classification<sup>27</sup>, and 9 patients (11%) were on treatment step 5.

# Associations between serum periostin levels and greater annual decline in $FEV_1$ and a decline in $FEV_1$ of 30 mL·yr<sup>-1</sup> or greater

In an analysis of continuous values of  $\Delta FEV_1$ , greater decline in  $FEV_1$  was associated with higher serum periostin levels at enrollment, treatment step 5, lower ACT scores, incomplete adherence to medications, comorbid or a history of sinusitis, and comorbid diabetes mellitus (Table 3). When patients were stratified into two groups according to their serum periostin levels, high serum periostin ( $\geq$  95 ng/mL) was also associated with greater decline in  $FEV_1$  (Table 3). Of these, high serum periostin was significant after controlling for multiple testing using the false discovery rate (q = 0.03, data not shown in Table 3). Multivariate analysis revealed that greater decline of  $FEV_1$  was solely associated with high serum periostin ( $\geq$  95 ng/mL) (estimated effect -5.39, 95% confidence interval -10.0 to -0.77, p = 0.02).

Fifty-two patients (23%) showed a decline in FEV<sub>1</sub> of 30 mL·yr<sup>-1</sup> or greater [mean - 51.8 (18.4) mL·yr<sup>-1</sup>] and were considered rapid decliners<sup>31</sup>. When adjusted by confounders, higher serum periostin levels at enrollment, treatment step 5, a history of admission due to asthma exacerbation, higher ICS daily doses, comorbid or a history of sinusitis, and exsmoking were associated with a decline in FEV<sub>1</sub> of 30 mL·yr<sup>-1</sup> or greater. High serum periostin ( $\geq$  95 ng/mL) was also associated with a decline in FEV<sub>1</sub> of 30 mL·yr<sup>-1</sup> or greater (Table 4). On multivariate analysis, high serum periostin ( $\geq$  95 ng/mL), treatment step 5, and ex-smoking were independent risk factors for a decline in FEV<sub>1</sub> of 30 mL·yr<sup>-1</sup> or greater (Table 4).

Of the 224 patients, 19 patients were on treatment step 5, and 36 patients took high-dose ICS (1,000 µg or higher doses of ICS equivalent to fluticasone propionate daily). When



patients were stratified into the high periostin group, the average  $\Delta FEV_1$  of patients on treatment step 5 (n = 9) was -41.0 (49.3) mL·yr<sup>-1</sup>, and 7 of them (78%) had excess decline; the average  $\Delta FEV_1$  of patients on high-dose ICS (n=18) was -34.3 (39.4) mL·yr<sup>-1</sup>, and 11 of them (61%) had a decline in  $FEV_1$  of 30 mL·yr<sup>-1</sup> or greater.

# Serum periostin levels and clinical indices

In 224 patients, serum periostin levels were weakly associated with blood eosinophil counts (Figure 2), serum IgE (Figure 2) and ECP levels (r = 0.25, p = 0.0005), ICS-untreated period, i.e. period between onset of asthma and the initiation of ICS therapy (r = 0.16, p = 0.01), daily maintenance doses of ICS at enrollment (r = 0.13, p = 0.05), and a history of admission due to asthma exacerbation (r = 0.15, p = 0.03). Serum periostin levels were significantly higher in patients on high-dose ICS ( $\geq 1,000$  µg daily) than in the remaining patients (110.3 ng/mL vs. 89.5 ng/mL, p = 0.003). Lastly, serum periostin levels were higher in patients with sinusitis than in those without sinusitis (103.9 ng/mL vs. 88.3 ng/mL, p = 0.007). Serum periostin levels did not show any seasonal variability or association with age at onset of asthma (data not shown).

### **POSTN** gene polymorphisms

Associations between polymorphisms of the *POSTN* gene, which encodes periostin, and both serum periostin levels and pulmonary function decline were then investigated. In one patient, DNA quality was insufficient for genotyping; thus, 3 tag SNPs of the *POSTN* gene were analyzed in 223 patients. All genotyped data were in Hardy-Weinberg equilibrium. The frequencies of the 3 tag SNPs and analysis results using dominant and recessive models for serum periostin levels and a decline in FEV<sub>1</sub> of 30 mL·yr<sup>-1</sup> or greater are presented in Table 5.

Serum periostin levels were higher in patients with the GG genotype of rs3829365 than





in those with the GC/CC genotype (GG 98.7 ng/mL vs. GC/CC 86.1 ng/mL, p = 0.003). 286 287rs1028728 was not associated with serum periostin levels or with the frequency of rapid decliners, but patients with the TT genotype of rs1028728, 4 patients only, showed no 288 significant decline compared with the AA/AT genotype (AA/AT -8.6 mL·yr<sup>-1</sup> vs. TT 29.3 289  $mL \cdot yr^{-1}$ , p = 0.03). Rapid decliners were more frequently observed in patients with the minor 290 A allele of rs9603226 than in the GG genotype (GG 16% vs. AG/AA 30%, p = 0.02). A 291 marked difference in the frequency of rapid decliners was observed when patients were 292 stratified into the high periostin group [GG of rs9630226 (n = 37) 19% vs. AG/AA (n = 47) 293 45%, p = 0.01]. 294

KURENAI 紅



295

296

297

298

299

300

301

302

303

304

305

306

307

308

309

310

311

312

313

314

315

316

317

318

319

320

### **Discussion**

To the best of our knowledge, this is the first study to identify a relationship between greater decline in FEV<sub>1</sub> and higher serum periostin levels, particularly if they were 95 ng/mL or more, in asthmatic patients on ICS treatment. It was also shown that high serum periostin, together with treatment step 5 and light ex-smoking, was an independent risk factor for a decline in FEV<sub>1</sub> of 30 mL·yr<sup>-1</sup> or greater. In addition, polymorphisms of the *POSTN* gene, which encodes periostin, were associated with serum periostin levels and a decline in FEV<sub>1</sub> of 30 mL·yr<sup>-1</sup> or greater in asthmatic patients. These findings suggest that serum periostin may be a useful biomarker for the development of airflow limitation in asthmatic patients on ICS. In this study, despite long-term treatment with ICS with or without other controllers, 23% of asthmatic patients were rapid decliners who showed a decline in FEV<sub>1</sub> of 30 mL·yr<sup>-1</sup> or greater, for which treatment step 5 was an independent risk factor. Adherence to ICS treatment and the frequency of early intervention with ICS did not differ between rapid decliners and non-decliners, although long-term adherence to ICS was undetermined in the present study. In previous studies of patients who were not treated with ICS, severe exacerbation of asthma contributed to greater annual decline of pulmonary function<sup>6,7</sup>, but the exacerbation-related greater annual decline disappeared in an early intervention group with ICS treatment in the START study<sup>6</sup>, which might be interpreted to mean that asthmatic patients on ICS treatment have little risk of accelerated FEV<sub>1</sub> decline. However, since the START study originally recruited mild persistent asthmatic patients, its results cannot simply be applied to severe asthmatic patients. As observed in the present study, there would be a subset of asthmatic patients still at risk of greater annual decline of pulmonary function despite intensive treatment for asthma. Persistent eosinophilic airway inflammation is a key process in irreversible airway

obstruction<sup>10</sup>. Indeed, exhaled nitric oxide of 20 ppb or higher is a risk factor for accelerated



322

323

324

325

326

327

328

329

330

331

332

333

334

335

336

337

338

339

340

341

342

343

344

345

346

FEV<sub>1</sub> decline in patients with difficult-to-treat asthma<sup>18</sup>. Studies on novel therapies for refractory eosinophilic asthma, i.e., anti-IL-5 therapy<sup>32</sup> and anti-IL-13 therapy<sup>33</sup>, revealed that these treatments may reverse airway remodeling when patients are adequately targeted, suggesting the necessity of establishing "companion diagnostics" for this population. According to the most recent study, serum periostin is the single best biomarker reflecting sputum and tissue eosinophilia among several biomarkers, including blood eosinophils and exhaled nitric oxide<sup>25</sup>. In the current study, the serum periostin level, which was associated with the blood eosinophil count, was the sole biomarker that reflected greater decline in FEV<sub>1</sub>. Periostin is secreted by airway epithelial cells<sup>23, 24</sup> and lung fibroblasts<sup>21</sup> in response to IL-4 and IL-13 and is thought to be secreted into the capillary vessels. Downstream of IL-13, which plays a pivotal role in subepithelial airway fibrosis<sup>34</sup>, airway remodeling<sup>35</sup>, and steroid insensitivity<sup>36</sup>, periostin mediates collagen synthesis<sup>24</sup> and fibrillogenesis<sup>24, 37</sup> by binding to collagen<sup>37</sup> and activates TGF- $\beta^{24}$ . In the asthmatic airway, periostin is deposited in the subepithelial layer, colocalizing with collagens I, III, and V, fibronectin, tenascin-C, and periostin itself<sup>21</sup>, which indicates involvement of periostin in airway remodeling in asthma. Collectively, periostin may be a key molecule that links eosinophilic inflammation and remodeling via IL-13 in asthmatic airways. Further roles of periostin in allergic inflammation and remodeling in the airways remain undetermined because studies using periostin-deficient mice with acute allergen exposure have yielded conflicting findings<sup>38-40</sup>; one study showed that periostin facilitates eosinophil infiltration into the lung<sup>38</sup>, whereas two other studies<sup>39, 40</sup> suggested protective roles of periostin. Meanwhile, a recent study of a chronic mouse model of atopic dermatitis demonstrated periostin's role in the chronicity of Th2 inflammation<sup>29</sup>. In the present study, patients on high-dose ICS showed higher serum periostin levels than the other patients. Although a longitudinal study is needed to determine responses of serum periostin levels to ICS treatment, we do not think that the high serum periostin levels in patients on high-dose ICS were induced by ICS treatment, because periostin expression in



the airway epithelium was decreased with ICS treatment<sup>23</sup>. Rather, the elevation of serum periostin in this population may reflect IL-13-mediated inflammation that is partly refractory to ICS, as was reported in a recent study by Jia and colleagues<sup>25</sup>. They showed that, in patients with severe asthma who were treated with high doses ICS (> 1000 µg daily), elevation of serum periostin levels was associated with persistent airway tissue eosinophilia, concluding that serum periostin is a systemic biomarker of airway eosinophilia refractory to high-dose ICS<sup>25</sup>. Providing further support, among patients with moderate to severe asthma who are inadequately controlled despite ICS treatment, patients with high serum periostin levels are likely to benefit from anti-IL-13 antibody, lebrikizumab, treatment<sup>33</sup>. The novelty of the present finding is that high serum periostin is an independent risk factor for greater decline in FEV<sub>1</sub>, providing the first evidence for the potential association between persistent Th2- or IL-13-driven inflammation refractory to ICS treatment and greater decline in FEV<sub>1</sub>, a functional consequence of airway remodeling.

Needless to say, current smokers with asthma have more accelerated FEV<sub>1</sub> decline<sup>4</sup> than those not smoking, and current smoking impairs the therapeutic response to ICS or oral corticosteroids<sup>41</sup>. Meanwhile, smoking cessation improves their FEV<sub>1</sub> levels<sup>42</sup>, and exsmokers with asthma with 10 pack-years or more show an intermediate response to short-term oral corticosteroid treatment, between current smokers and never-smokers<sup>41</sup>. In the present study, rather unexpectedly, ex-smoking with 10 pack-years or less was still an independent risk factor for a decline in FEV<sub>1</sub> of 30 mL·yr<sup>-1</sup> or greater. It should be recognized that even light ex-smoking increases the risk of airway remodeling in asthmatic patients on ICS, and its underlying mechanisms should be clarified.

Chronic sinusitis is a well-known comorbidity with severe asthma<sup>43, 44</sup>. In the present study, rapid decliners were more frequently observed in asthmatic patients with sinusitis than those without sinusitis on univariate analysis, and their periostin levels were higher than in patients without sinusitis. In the present study, polypoid lesions in the sinuses were not



evaluated by otolaryngologists at enrollment. However, considering that periostin is upregulated in nasal polyp tissue in patients with chronic rhinosinusitis<sup>45</sup>, asthmatic patients with sinusitis may have had severe upper and lower airway inflammation with persistent increases in periostin expression, which may have resulted in a decline in FEV<sub>1</sub> of 30 mL·yr<sup>-1</sup> or greater. Periostin is a potential molecule that unifies sinusitis and severe asthma.

Periostin is encoded on the *POSTN* gene, which is located on chromosome 13q13.3. rs3829365, which is located at the 5'UTR region that may contain sequences to regulate translation efficiency or mRNA stability, was associated with serum periostin levels. This finding suggests that, besides IL-13, a master regulator of periostin, genetic background partly determines periostin levels, although a replication study would be necessary to confirm this. The minor A allele of rs9603226, located 66 bp upstream of exon 21 in the C-terminal region, was associated with a decline in FEV<sub>1</sub> of 30 mL·yr<sup>-1</sup> or greater. In periostin, FAS I domains are thought to be primary binding sites to fibronectin, tenascin-C, and collagen V<sup>21</sup>, whereas the C-terminal region in its intact form may down-regulate the binding activity of periostin to these extracellular matrix proteins<sup>21</sup>. We therefore speculate that the minor A allele of rs9603226 might modify the binding activity at the C-terminal region and facilitate airway remodeling, particularly if the airway is in periostin enriched milieu. Further studies are needed to clarify if these SNPs are functional variants.

The age of patients in this study appears to be older than in other Euro-American studies<sup>6,7,14,18,20,23,25</sup>. One reason for the age distribution would be the entry criteria of this study. Another reason would be explained by population aging including population with asthma in Japan. According to a patient survey by the Japanese Ministry of Health, Labour and Welfare in 2008, patients aged 70 to 74 years were the most frequent age group of adult patients with asthma<sup>46</sup>, which is still older than the average age of patients in this study.

There are several limitations to the present study. First, since this study was observational in nature, ICS doses and numbers or types of controllers were not fixed during





the follow-up period. Controllers such as long-acting β2 agonists were not withdrawn at pulmonary function testing to evaluate function on daily medications, which may have resulted in the small average ΔFEV1, -7.8 mL·yr¹. Meanwhile, averages of 16.2 measurements of FEV1 and 8.0 years of follow-up were satisfactory for a longitudinal analysis of pulmonary function⁴7, and ΔFEV1 was normally distributed. Secondly, serum biomarkers were measured only once at enrollment, but the significant associations between *POSTN* gene polymorphisms and serum periostin levels or a decline in FEV1 of 30 mL·yr¹¹ or greater may circumvent the inherent insufficiency of single measurement of serum periostin. Thirdly, most of the clinical information, including smoking history and chronic sinusitis, was based on a self-completed questionnaire, which might be biased by recall memory. Despite these limitations, the current findings may provide directions for future research.

In conclusion, serum periostin appears to be a useful biomarker that reflects the development of airflow limitation in patients on prolonged treatment with ICS. *POSTN* gene

polymorphisms may also be helpful for identification of rapid decliners.





# Acknowledgments

The authors would like to acknowledge Dr Nobuo Ohta, Department of
Otolaryngology, Head and Neck Surgery, Yamagata University for fruitful discussion on
periostin expression in nasal tissue of chronic sinusitis. The authors would also like to thank
Ms Maki Futamata (Saga Medical School), Dr Guergana Petrova Stoyanoya, Dr Cui Shilei,
Ms Aya Inazumi, and Ms Yuko Maeda (Kyoto University) for their technical assistance.





- 420 Reference
- 1. Pascual RM, Peters SP. Airway remodeling contributes to the progressive loss of lung
- function in asthma: an overview. J Allergy Clin Immunol 2005; 116:477-86; quiz 87.
- 2. Niimi A, Matsumoto H, Takemura M, Ueda T, Chin K, Mishima M. Relationship of
- 424 airway wall thickness to airway sensitivity and airway reactivity in asthma. Am J
- 425 Respir Crit Care Med 2003; 168:983-8.
- 426 3. Ueda T, Niimi A, Matsumoto H, Takemura M, Hirai T, Yamaguchi M, et al. Role of
- small airways in asthma: investigation using high-resolution computed tomography. J
- 428 Allergy Clin Immunol 2006; 118:1019-25.
- 429 4. Lange P, Parner J, Vestbo J, Schnohr P, Jensen G. A 15-year follow-up study of
- ventilatory function in adults with asthma. N Engl J Med 1998; 339:1194-200.
- 431 5. Ulrik CS. Outcome of asthma: longitudinal changes in lung function. Eur Respir J
- 432 1999; 13:904-18.
- 6. O'Byrne PM, Pedersen S, Lamm CJ, Tan WC, Busse WW, Group SI. Severe
- exacerbations and decline in lung function in asthma. Am J Respir Crit Care Med
- 435 2009; 179:19-24.
- 436 7. Bai TR, Vonk JM, Postma DS, Boezen HM. Severe exacerbations predict excess lung
- function decline in asthma. Eur Respir J 2007; 30:452-6.
- 438 8. Ulrik CS, Lange P. Decline of lung function in adults with bronchial asthma. Am J
- 439 Respir Crit Care Med 1994; 150:629-34.
- 440 9. Ulrik CS, Backer V, Dirksen A. A 10 year follow up of 180 adults with bronchial
- asthma: factors important for the decline in lung function. Thorax 1992; 47:14-8.
- ten Brinke A, Zwinderman AH, Sterk PJ, Rabe KF, Bel EH. Factors associated with
- persistent airflow limitation in severe asthma. Am J Respir Crit Care Med 2001;
- 444 164:744-8.
- 11. Jongepier H, Boezen HM, Dijkstra A, Howard TD, Vonk JM, Koppelman GH, et al.
- Polymorphisms of the ADAM33 gene are associated with accelerated lung function
- decline in asthma. Clin Exp Allergy 2004; 34:757-60.
- 448 12. Koppelman GH, Sayers I. Evidence of a genetic contribution to lung function decline
- in asthma. J Allergy Clin Immunol 2011; 128:479-84.
- Tantisira KG, Lasky-Su J, Harada M, Murphy A, Litonjua AA, Himes BE, et al.
- Genomewide association between GLCCI1 and response to glucocorticoid therapy in
- 452 asthma. N Engl J Med 2011; 365:1173-83.
- 14. Dijkstra A, Vonk JM, Jongepier H, Koppelman GH, Schouten JP, ten Hacken NH, et
- al. Lung function decline in asthma: association with inhaled corticosteroids, smoking
- and sex. Thorax 2006; 61:105-10.
- 15. O'Byrne PM, Pedersen S, Busse WW, Tan WC, Chen YZ, Ohlsson SV, et al. Effects
- of early intervention with inhaled budesonide on lung function in newly diagnosed





- 458 asthma. Chest 2006; 129:1478-85.
- 459 16. Selroos O, Löfroos AB, Pietinalho A, Riska H. Asthma control and steroid doses 5
- years after early or delayed introduction of inhaled corticosteroids in asthma: a real-
- life study. Respir Med 2004; 98:254-62.
- 462 17. de Marco R, Marcon A, Jarvis D, Accordini S, Bugiani M, Cazzoletti L, et al. Inhaled
- steroids are associated with reduced lung function decline in subjects with asthma
- with elevated total IgE. J Allergy Clin Immunol 2007; 119:611-7.
- van Veen IH, Ten Brinke A, Sterk PJ, Sont JK, Gauw SA, Rabe KF, et al. Exhaled
- nitric oxide predicts lung function decline in difficult-to-treat asthma. Eur Respir J
- 467 2008; 32:344-9.
- Levine SJ, Wenzel SE. Narrative review: the role of Th2 immune pathway modulation
- in the treatment of severe asthma and its phenotypes. Ann Intern Med 2010; 152:232-
- 470 7.
- Woodruff PG, Modrek B, Choy DF, Jia G, Abbas AR, Ellwanger A, et al. T-helper type
- 2-driven inflammation defines major subphenotypes of asthma. Am J Respir Crit Care
- 473 Med 2009; 180:388-95.
- Takayama G, Arima K, Kanaji T, Toda S, Tanaka H, Shoji S, et al. Periostin: a novel
- component of subepithelial fibrosis of bronchial asthma downstream of IL-4 and IL-
- 476 13 signals. J Allergy Clin Immunol 2006; 118:98-104.
- 477 22. Kii I, Nishiyama T, Li M, Matsumoto K, Saito M, Amizuka N, et al. Incorporation of
- 478 tenascin-C into the extracellular matrix by periostin underlies an extracellular
- meshwork architecture. J Biol Chem 2010; 285:2028-39.
- 480 23. Woodruff PG, Boushey HA, Dolganov GM, Barker CS, Yang YH, Donnelly S, et al.
- Genome-wide profiling identifies epithelial cell genes associated with asthma and
- with treatment response to corticosteroids. Proc Natl Acad Sci U S A 2007;
- 483 104:15858-63.
- 484 24. Sidhu SS, Yuan S, Innes AL, Kerr S, Woodruff PG, Hou L, et al. Roles of epithelial
- cell-derived periostin in TGF-beta activation, collagen production, and collagen gel
- elasticity in asthma. Proc Natl Acad Sci U S A 2010; 107:14170-5.
- 487 25. Jia G, Erickson RW, Choy DF, Mosesova S, Wu LC, Solberg OD, et al. Periostin is a
- systemic biomarker of eosinophilic airway inflammation in asthmatic patients. J
- 489 Allergy Clin Immunol 2012; 130: 647-54.
- 490 26. Standards for the diagnosis and care of patients with chronic obstructive pulmonary
- disease (COPD) and asthma. This official statement of the American Thoracic Society
- was adopted by the ATS Board of Directors, November 1986. Am Rev Respir Dis
- 493 1987; 136:225-44.
- 494 27. Global Strategy for Asthma Management and Prevention, Global Initiative for Asthma
- 495 (GINA) 2010. Available from: Global Strategy for Asthma Management and





- 496 Prevention, Global Initiative for Asthma (GINA) 2010. Available from:
- 497 http://www.ginasthma.org.
- 498 28. Okamoto M, Hoshino T, Kitasato Y, Sakazaki Y, Kawayama T, Fujimoto K, et al.
- Periostin, a matrix protein, is a novel biomarker for idiopathic interstitial pneumonias.
- 500 Eur Respir J 2011; 37:1119-27.
- 501 29. Masuoka M, Shiraishi H, Ohta S, Suzuki S, Arima K, Aoki S, et al. Periostin
- promotes chronic allergic inflammation in response to Th2 cytokines. J Clin Invest
- 503 2012; 122:2590-600.
- 504 30. Storey JD, Tibshirani R. Statistical significance for genomewide studies. Proc Natl
- 505 Acad Sci U S A 2003; 100:9440-5.
- 506 31. Broekema M, Volbeda F, Timens W, Dijkstra A, Lee NA, Lee JJ, et al. Airway
- eosinophilia in remission and progression of asthma: accumulation with a fast decline
- 508 of FEV<sub>1</sub>. Respir Med 2010; 104:1254-62.
- 509 32. Haldar P, Brightling CE, Hargadon B, Gupta S, Monteiro W, Sousa A, et al.
- Mepolizumab and exacerbations of refractory eosinophilic asthma. N Engl J Med
- 511 2009; 360:973-84.
- 512 33. Corren J, Lemanske RF, Hanania NA, Korenblat PE, Parsey MV, Arron JR, et al.
- Lebrikizumab treatment in adults with asthma. N Engl J Med 2011; 365:1088-98.
- 514 34. Zhu Z, Homer RJ, Wang Z, Chen Q, Geba GP, Wang J, et al. Pulmonary expression of
- interleukin-13 causes inflammation, mucus hypersecretion, subepithelial fibrosis,
- physiologic abnormalities, and eotaxin production. J Clin Invest 1999; 103:779-88.
- 517 35. Elias JA, Zhu Z, Chupp G, Homer RJ. Airway remodeling in asthma. J Clin Invest
- 518 1999; 104:1001-6.
- 519 36. Saha SK, Berry MA, Parker D, Siddiqui S, Morgan A, May R, et al. Increased sputum
- and bronchial biopsy IL-13 expression in severe asthma. J Allergy Clin Immunol
- 521 2008; 121:685-91.
- 522 37. Norris RA, Damon B, Mironov V, Kasyanov V, Ramamurthi A, Moreno-Rodriguez R,
- et al. Periostin regulates collagen fibrillogenesis and the biomechanical properties of
- 524 connective tissues. J Cell Biochem 2007; 101:695-711.
- 525 38. Blanchard C, Mingler MK, McBride M, Putnam PE, Collins MH, Chang G, et al.
- Periostin facilitates eosinophil tissue infiltration in allergic lung and esophageal
- 527 responses. Mucosal Immunol 2008; 1:289-96.
- 528 39. Gordon ED, Sidhu SS, Wang ZE, Woodruff PG, Yuan S, Solon MC, et al. A protective
- role for periostin and TGF-β in IgE-mediated allergy and airway hyperresponsiveness.
- 530 Clin Exp Allergy 2012; 42:144-55.
- 531 40. Sehra S, Yao W, Nguyen ET, Ahyi AN, Tuana FM, Ahlfeld SK, et al. Periostin
- regulates goblet cell metaplasia in a model of allergic airway inflammation. J
- 533 Immunol 2011; 186:4959-66.



554

162:2134-8.



Thomson NC, Chaudhuri R, Livingston E. Asthma and cigarette smoking. Eur Respir 53441. J 2004; 24:822-33. 535 42. Chaudhuri R, Livingston E, McMahon AD, Lafferty J, Fraser I, Spears M, et al. 536 537 Effects of smoking cessation on lung function and airway inflammation in smokers with asthma. Am J Respir Crit Care Med 2006; 174:127-33. 538 Dixon AE, Kaminsky DA, Holbrook JT, Wise RA, Shade DM, Irvin CG. Allergic 43. 539 rhinitis and sinusitis in asthma: differential effects on symptoms and pulmonary 540 function. Chest 2006; 130:429-35. 541 Mascia K, Borish L, Patrie J, Hunt J, Phillips CD, Steinke JW. Chronic hyperplastic 542 44. eosinophilic sinusitis as a predictor of aspirin-exacerbated respiratory disease. Ann 543 Allergy Asthma Immunol 2005; 94:652-7. 544 45. Ishida A, Ohta N, Suzuki Y, Kakehata S, Okubo K, Ikeda H, et al. Expression of 545 Pendrin and Periostin in Allergic Rhinitis and Chronic Rhinosinusitis. Allergol Int 546 547 2012; 61: 589-95. Japanese Society of Allergology, Asthma Guideline Committee. Asthma Prevention 54846. and Management Guidelines 2012. Tokyo: Kyowa Kikaku; 2012 (in Japanese) 54947. Wang ML, Gunel E, Petsonk EL. Design strategies for longitudinal spirometry 550 551 studies: study duration and measurement frequency. Am J Respir Crit Care Med 2000;



### Table 1. Contents of the self-completed questionnaire

Asthma-related history

- •family history of asthma
- •age of asthma onset
- •history of pediatric asthma
- •history of admission due to asthma worsening or exacerbation
- aspirin hypersensitivity
- •asthma deterioration at the working place

# Comorbidity or a history of the following diseases

•allergic dermatitis •cardiovascular diseases including ischemic heart disease

•allergic rhinitis •gastrointestinal diseases including GERD

• seasonal rhinitis • collagen vascular diseases including rheumatoid arthritis

• allergic conjunctivitis • diabetes mellitus

•chronic sinusitis •pulmonary diseases other than asthma

other diseases including malignancy

### Lifestyle and environment

•smoking history •a highway near the home

•pet breeding •age at menopause

type of occupation

### Adherence to medication, sputum production, and exacerbations

•How often do you forget to take inhaled corticosteroids or other medications?

0: never, 1: seldom, 2: sometimes, 3: often, 4: always

• How often do you produce sputum?

0: never, 1: once in a few days, 2: every morning, 3: every morning and daytime

• How often did you receive systemic steroids due to asthma exacerbations during the recent 6 months?

0: never, 1: once, 2: twice or more

GERD: gastro-esophageal reflux disease





### Table 2. Patients' characteristics

Sex (males/ females), n	53 / 171
Age at enrollment, years	62.3 (13.7)
Age at asthma onset, years	42.0 (19.0)
Body mass index (kg/m <sup>2</sup> )	23.1 (3.5)
Smoking history (never), n	181
Atopic predisposition*, %	70
Pediatric asthma (none/ recurrent/ persistent), %	81 / 8 / 11
Disease duration, years	20.2 (14.5)
ICS-untreated period, years	9.2 (13.1)
ICS daily maintenance dose <sup>†</sup> , μg	525 (318)
Number of other controller medications, n	1.4 (1.2)
Treatment step $(2/3/4/5)^{\ddagger}$ , %	16 / 27 / 49 / 8
Sputum production (0/ 1/ 2/ 3) §, %	54 / 20 / 8 / 18
Asthma Control Test, points	22.6 (3.5)
History of admission due to asthma, n (%)	78 (35)
Allergic rhinitis, n (%)	129 (58)
Chronic sinusitis, n (%)	65 (29)
Blood neutrophils, %	60.1 (10.0)
eosinophils, %	5.2 (4.9)
Serum IgE, IU/mL	180 (0 - 16000)
periostin, ng/mL	92.8 (38.4)
high sensitivity C-reactive protein, mg/L	1341 (3147)
eosinophil cationic protein, μg/L	15.1 (29.3)
FEV <sub>1</sub> at the first measurement, L <sup>¶</sup>	2.11 (0.69)
%predicted FEV <sub>1</sub> at the first measurement, %	91.9 (19.2)
FEV <sub>1</sub> / FVC at the first measurement, %	73.9 (9.8)
FEV <sub>1</sub> at enrollment, L	2.04 (0.73)
%predicted FEV <sub>1</sub> at enrollment, %	97.4 (22.2)
FEV <sub>1</sub> / FVC at enrollment, %	72.2 (10.0)
Reversibility at enrollment, %#	3.8 (6.0)
Data at annullment and museumted analysis of amore attack Data	· · · · · · · · · · · · · · · · · · ·

Data at enrollment are presented unless otherwise stated. Data are expressed as means (SD) except for median (range) for serum IgE. \*Considered atopic when one or more specific IgE antibodies against cat or dog dander, weed, grass, or Japanese cedar pollens, moulds, or house dust mite were positive. †Equivalent to fluticasone propionate.  $^{\ddagger}$ according to the Global Initiative for Asthma 2010 guideline  $^{27}$ .  $^{\$}$ 0 = never, the details are shown in Table 1.  $^{\$}$ The first pulmonary function test was performed at least one year after the commencement of ICS treatment and at 25 years of age or older.  $^{\sharp}$ n = 206, airway reversibility to 200  $\mu$ g of inhaled salbutamol.





Table 3. Estimated effects of clinical indices and biomarkers on  $\Delta FEV_1$ 

	Estimates	95% C.I.	p value
Smoking history, ex vs. never	-8.48	-20.2, 3.27	0.16
Atopic predisposition	-1.10	-6.29, 4.09	0.68
Disease duration, years	-4.79	-18.4, 8.86	0.56
ICS-untreated period, years	0.10	-0.24, 0.45	0.65
ICS daily maintenance dose, µg	-0.01	-0.03, 0.001	0.07
Number of other controller medications, n	-0.36	-4.21, 3.49	0.86
Adherence to medication, incomplete vs. complete*	-4.56	-9.08, -0.04	0.05
Treatment step, 5 vs. 2-4 <sup>†</sup>	-7.77	-15.7, 0.13	0.05
Sputum production, never vs. others <sup>‡</sup>	0.99	-3.53, 5.51	0.67
Asthma Control Test, points	1.53	0.29, 2.77	0.02
History of admission due to asthma	-4.49	-9.45, 0.46	0.08
Aspirin hypersensitivity	-6.52	-20.0, 6.98	0.34
Asthma deterioration at the working place	-12.2	-54.4, 30.0	0.57
Allergic rhinitis	-1.21	-5.88, 3.45	0.61
Allergic dermatitis	4.51	-1.51, 10.5	0.14
Chronic sinusitis	-10.1	-19.8, -0.27	0.04
Ischemic heart disease	3.41	-16.6, 23.4	0.74
Hypertension	-3.79	-9.12, 1.53	0.16
Dyslipidemia	-3.67	-9.42, -2.06	0.21
Diabetes mellitus	-8.03	-15.4, -0.67	0.03
Gastro-esophageal reflux disease	-3.85	-9.89, 2.19	0.21
Malignancy	-3.44	-26.0, 19.1	0.76
Post-menopause	5.05	-14.2, 24.3	0.60
Pet breeding	-0.28	-12.6, 12.0	0.96
Log blood neutrophils, %	-7.40	-69.1, 54.3	0.81
eosinophils, %	-0.67	-1.60, 0.27	0.16
Log serum IgE, IU/mL	-2.85	-9.74, 4.04	0.42
periostin, ng/mL	-29.1	-56.2, -1.97	0.04
high sensitivity C-reactive protein, mg/L	-1.88	-9.85, 6.10	0.64
eosinophil cationic protein, μg/L	-4.47	-15.7, 6.81	0.44
Periostin group, high vs. low §	-6.96	-11.4, -2.51	0.002

Estimated effects were adjusted by sex, height, age at enrollment, and  $FEV_1$  at the first measurement. "Complete", when patients answered that they never forgot to take ICS or other medications; "incomplete", the remaining cases. †according to the Global Initiative for Asthma 2010 guideline<sup>27</sup>. †The details are shown in Table1. § Patients were stratified into two groups according to their serum periostin levels: high  $\geq$  95 ng/mL, low < 95 ng/mL. ICS: inhaled corticosteroids, C.I.: confidence interval



571



# Table 4. Estimated effects of clinical indices and serum periostin on a decline in $FEV_1$ of 30 mL·yr $^{-1}$ or greater

	Univariate analysis			Multivaria		
	Estimates	95% C.I.	p value	Estimates	95% C.I.	p value
Treatment step, 5 vs. 2-4*	1.63	0.51, 2.60	0.004	1.24	0.078, 2.30	0.04
History of admission due to asthma	1.09	0.37, 1.90	0.003	0.70	-0.11, 1.50	0.09
ICS daily maintenance dose, µg	0.001	0.00, 0.002	0.01	-		
Chronic sinusitis	0.82	0.11, 1.53	0.03	0.61	-0.15, 1.37	0.12
Smoking history, ex vs. never	0.87	-0.002, 1.74	0.05	0.98	0.030, 1.93	0.04
Log serum periostin, ng/mL	2.96	0.78, 5.13	0.008	-		
Periostin group, high vs. low <sup>†</sup>	1.03	0.33, 1.72	0.004	0.87	0.11, 1.63	0.03

572 Estimated effects were adjusted by sex, height, age at enrollment, and  $FEV_1$  at the first measurement.

\*according to the Global Initiative for Asthma 2010 guideline<sup>27</sup>.

<sup>†</sup>Patients were stratified into two groups according to their serum periostin levels: high  $\geq$  95 ng/mL, low <

575 95 ng/mL. ICS: inhaled corticosteroids, C.I.: confidence interval

576 ICS daily maintenance dose was excluded from multivariate analysis because of its strong correlation with

treatment step.





Table 5. Frequencies of 3 tag SNPs and analysis results using dominant and recessive models
 for serum periostin levels and frequency of rapid decliners\*

					Serum periostin levels		Frequency of rapid decliners	
					p value		p value	
Tag SNP	Genotype	n (%)	Allelic	n (%)	Dominant <sup>†</sup>	Recessive*	Dominant <sup>†</sup>	Recessive*
rs1028728	AA	164 (74)	A	383 (86)				
	AT	55 (25)	T	63 (14)	0.40	0.46	0.17	0.14
	TT	4 (2)						
rs3829365	GG	113 (51)	G	316 (71)				
	GC	90 (40)	C	130 (29)	0.003	0.70	0.40	0.33
	CC	20 (9)						
rs9603226	GG	107 (48)	G	311 (70)				
	AG	97 (44)	A	135 (30)	0.80	0.33	0.01	0.81
	AA	19 (9)						

<sup>\*</sup> defined as patients who showed a decline in FEV<sub>1</sub> of 30 mL·yr<sup>-1</sup> or greater

<sup>&</sup>lt;sup>†</sup> Assuming that heterozygotes have the same increased risk as minor homozygous genotypes.

<sup>&</sup>lt;sup>‡</sup>Assuming that heterozygotes have no increased risk.







585	Figure legends
586	Figure 1. Three tag SNPs that determine 4 major haplotypes of the <i>POSTN</i> gene and
587	haplotype frequencies in the Japanese population are presented.
588	*at intron 66 bp upstream of exon 21
589	
590	Figure 2. Relationships between serum periostin levels and blood eosinophil counts (left) or
591	serum IgE levels (right).
592	Presented in logarithmic scales on both the X- and Y-axes.
593	





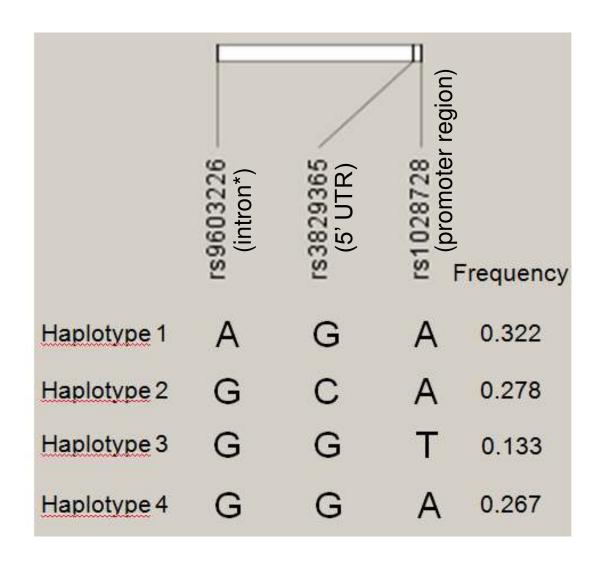
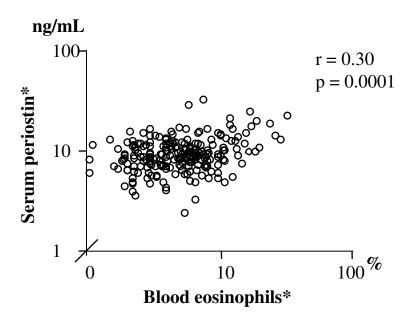


Figure 1.







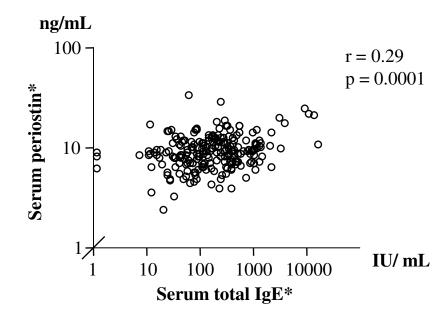


Figure 2.





- 1 Online Repository
- 2 Increased periostin associates with greater airflow limitation in patients receiving
- 3 inhaled corticosteroids
- 5 Yoshihiro Kanemitsu MD\*<sup>1,2</sup>, Hisako Matsumoto MD, PhD\*<sup>1,2</sup>, Kenji Izuhara
- 6 MD,PhD<sup>3</sup>, Yuji Tohda MD, PhD<sup>2,4</sup>, Hideo Kita MD, PhD<sup>2,5</sup>, Takahiko Horiguchi MD,
- 7 PhD<sup>2,6</sup>, Kazunobu Kuwahara MD, PhD<sup>2,6</sup>, Keisuke Tomii MD, PhD<sup>2,7</sup>, Kojiro Otsuka
- 8 MD, PhD<sup>1,2,7</sup>, Masaki Fujimura MD, PhD<sup>2,8</sup>, Noriyuki Ohkura MD<sup>2,8</sup>, Katsuyuki Tomita
- 9 MD, PhD<sup>2,4</sup>, Akihito Yokoyama MD, PhD<sup>2,9</sup>, Hiroshi Ohnishi MD, PhD<sup>2,9</sup>, Yasutaka
- Nakano MD, PhD<sup>2,10</sup>, Tetsuya Oguma MD, PhD<sup>2,10</sup>, Soichiro Hozawa MD, PhD<sup>2,11</sup>,
- Tadao Nagasaki MD<sup>1</sup>, Isao Ito MD, PhD<sup>1</sup>, Tsuyoshi Oguma MD<sup>1</sup>, Hideki Inoue MD<sup>1</sup>,
- 12 Tomoko Tajiri MD<sup>1</sup>, Toshiyuki Iwata MD<sup>1</sup>, Yumi Izuhara MD<sup>1</sup>, Junya Ono BS<sup>12</sup>,
- 13 Shoichiro Ohta MD, PhD<sup>13</sup>, Mayumi Tamari MD, PhD<sup>14</sup>, Tomomitsu Hirota DDS,
- PhD<sup>14</sup>, Tetsuji Yokoyama MD, PhD<sup>15</sup>, Akio Niimi MD, PhD<sup>1,2,16</sup> and Michiaki Mishima
- 15 MD, PhD<sup>1,2</sup>.
- 16 Affiliations
- 17 Department of Respiratory Medicine, Graduate School of Medicine, Kyoto University,
- 18 Kyoto, Japan.
- 19 <sup>2</sup> Kinki Hokuriku Airway disease Conference (KiHAC)
- 20 <sup>3</sup> Division of Medical Biochemistry, Department of Biomolecular Sciences, Saga
- 21 Medical School, Saga, Japan.
- <sup>4</sup> Department of Respiratory Medicine and Allergology, Faculty of Medicine, Kinki
- 23 University, Osaka, Japan.
- <sup>5</sup> Department of Respiratory Medicine, Takatsuki Red Cross Hospital, Osaka, Japan.
- 25 <sup>6</sup> Department of Respiratory Internal Medicine, Fujita Health University Second





- 26 Educational Hospital, Aichi, Japan.
- <sup>7</sup> Department of Respiratory Medicine, Kobe City Medical Center General Hospital,
- 28 Hyogo, Japan.
- 29 <sup>8</sup> Department of Respiratory Medicine, Cellular Transplantation Biology, Kanazawa
- 30 University Graduate School of Medicine, Kanazawa, Japan.
- 31 <sup>9</sup> Department of Hematology and Respiratory Medicine, Kochi University, Kochi,
- 32 Japan.
- 33 <sup>10</sup> Division of Respiratory Medicine, Department of Internal Medicine, Shiga University
- of Medical Science, Shiga, Japan.
- 35 <sup>11</sup> Hiroshima Allergy and Respiratory Clinic, Hiroshima, Japan.
- 36 <sup>12</sup> Shino-Test Corporation, Kanagawa, Japan.
- 37 Department of Laboratory Medicine, Saga Medical School, Saga, Japan.
- 38 14 Laboratory for Respiratory Diseases, Center for Genomic Medicine, RIKEN,
- 39 Yokohama, Kanagawa, Japan.
- 40 <sup>15</sup> Department of Health Promotion, National Institute of Public Health, Wako, Saitama,
- 41 Japan

- 42 <sup>16</sup> Department of Medical Oncology and Immunology, Nagoya City University School
- 43 of Medical Sciences, Aichi, Japan.
- \*YK and HM similarly contributed to this study.
- Correspondence should be addressed to Hisako Matsumoto M.D., Ph.D.,
- 48 Department of Respiratory Medicine, Graduate School of Medicine, Kyoto University,
- 49 53 Kawahara-cho, Shogoin, Sakyo-ku, Kyoto 606-8507, Japan.





E-mail: <u>hmatsumo@kuhp.kyoto-u.ac.jp</u>





### Methods

### **Patients**

Patients with asthma were recruited from nine institutions belonging to the Kinki Hokuriku Airway disease Conference where asthma specialists manage patients, including six university hospitals, two satellite general hospitals, and one satellite clinic. Asthma was diagnosed according to the American Thoracic Society criteria<sup>E1</sup> on the basis of a history of recurrent episodes of wheezing and chest tightness with or without cough and documented airway reversibility to a bronchodilator or hyper-responsiveness to inhaled methacholine. From September 2009 to December 2011, patients were enrolled if they had received ICS treatment for 4 years or more, undergone three or more pulmonary function tests when they were stable, and were free from exacerbations for at least one month. The first pulmonary function test was performed at least one year after the commencement of ICS treatment and at 25 years of age or older. Patients who had smoked more than 10 pack-years, smoked in the past one year, or had other pulmonary diseases were excluded.

### Self-completed questionnaire and clinical indices

The self-completed questionnaire was composed of 4 major items, as presented in Table 1.

Adherence to ICS or other medications, frequency of sputum production, and requirement for systemic corticosteroids during the last 6 months were graded as shown in Table 1. The Asthma Control Test (ACT)<sup>TM</sup> was also scored. Duration of ICS treatment and details on medication at enrollment were recorded from medical charts by patients' physicians. The treatment step at enrollment was determined according to the



Global Initiative for Asthma 2010 guideline<sup>E2</sup>.

# Measurement of systemic biomarkers

Blood eosinophil and neutrophil counts, and serum levels of total immunoglobulin E (IgE) (ImmunoCAP® total IgE, Phadia K.K., Tokyo, Japan), specific IgE against common inhaled allergens (ImmunoCAP® specific IgE), eosinophil cationic protein (ECP) (ImmunoCAP® ECP), high sensitivity C-reactive protein (hsCRP) (CardioPhase® hsCRP, Siemens Healthcare Diagnostics K.K., Tokyo, Japan), and periostin were determined.

Serum periostin levels were measured using an enzyme-linked immunosorbent assay at Shino-test (Kanagawa, Japan), as described previously<sup>E3</sup>. Briefly, two rat anti-human periostin monoclonal antibodies (SS18A and SS17B) were used. SS18A and SS17B are antibodies against the first and fourth FAS I domains, respectively. Intra- and inter-assay coefficients of variation ranged from 1.31% to 2.54% and 1.49% to 2.01%, respectively.

### Haplotype analysis, DNA extraction, and genotyping of the *POSTN* gene

A total of 47 single-nucleotide polymorphisms (SNPs) in the region of the POSTN gene and its upstream, total 39 kb, was captured in the HapMap Japanese data set with minor allele frequencies > 0.10. Pairwise tagging was performed at  $r^2 > 0.8$  using a tagger in Haploview 4.2 software. Haplotype analysis identified 4 major haplotypes and 2 minor haplotypes. Two minor haplotypes were grouped into the closest major haplotype, and 3 tag SNPs that determined the 4 haplotypes were identified (Figure 1). These 3 tag SNPs were located at promoter region (rs1028728), 5'UTR







region (rs3829365), and at intron 66 bp upstream of exon 21 (rs9603226). The frequencies of the minor alleles in the Japanese population were 0.136 (rs1028728), 0.278 (rs3829365), and 0.330 (rs9603226).

Genomic DNA was isolated from blood cells using a QIAamp DNA Blood Mini Kit (Qiagen, Tokyo, Japan). SNPs were genotyped using a Taqman genotyping assay according to the manufacturer's instructions (Applied Biosystems, Tokyo, Japan) and analyzed using an Applied Biosystems 7300 Real-Time PCR System (Applied Biosystems).







109	Refe	rences
110	E1.	Standards for the diagnosis and care of patients with chronic obstructive
111		pulmonary disease (COPD) and asthma. This official statement of the American
112		Thoracic Society was adopted by the ATS Board of Directors, November 1986.
113		Am Rev Respir Dis 1987; 136:225-44.
114		
115	E2	Global Strategy for Asthma Management and Prevention, Global Initiative for
116		Asthma (GINA) 2010. Available from: Global Strategy for Asthma Management
117		and Prevention, Global Initiative for Asthma (GINA) 2010. Available from:
118		http://www.ginasthma.org.
119		
120	E3.	Okamoto M, Hoshino T, Kitasato Y, Sakazaki Y, Kawayama T, Fujimoto K, et al.
121		Periostin, a matrix protein, is a novel biomarker for idiopathic interstitial
122		pneumonias. Eur Respir J 2011; 37:1119-27.
123		







124	Figure legends
125	
126	Figure E1. The distribution of $\Delta FEV_1$ in the study population
127	
128	Figure E2. ROC curve analysis of serum periostin levels comparing asthmatic patients
129	and healthy subjects, in which the cutoffs of 95 ng/mL, 80 ng/mL, 92 ng/mL, and 100
130	ng/mL are presented with arrows.



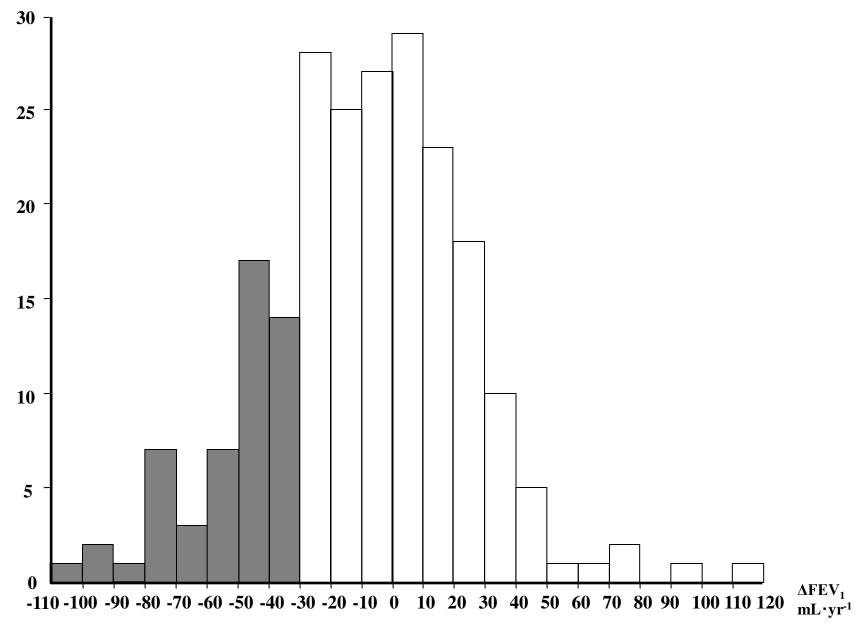


Figure E1.

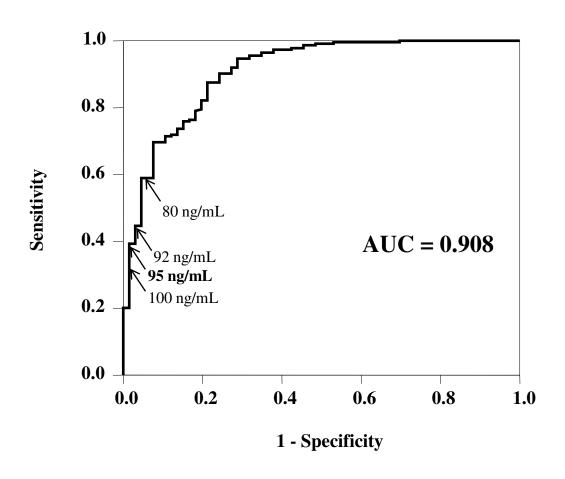


Figure E2.