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Pulsed field gradients (PFGs) make it possible to record two- and three-dimensional spectra using experiments with only a single scan per increment (1–10). In these experiments PFGs are employed to select the desired coherence-transfer pathway. However, each time a combination of PFGs is used to select a coherence transfer from order \(n\) to \(m\), a second desired pathway, from \(-n\) to \(m\), is eliminated, resulting in an intrinsic sensitivity loss of a factor \(\sqrt{2}\). Consequently, coherence pathway selection by means of PFGs is suitable only for applications where the signal-to-noise ratio is not a limiting factor. However, for 3D and 4D experiments applied to dilute protein samples, the intrinsic loss in signal-to-noise of \(\sqrt{2}\) for each PFG coherence selection step is highly undesirable. To circumvent this problem, a different strategy which relies on suppression of undesired coherence-transfer pathways can be used (11). In this latter approach, coherences associated with spurious magnetization transfer, which frequently result from pulse imperfections, can be eliminated, thereby reducing the number of phase-cycling steps required. A shorter phase cycle allows a reduction in measuring time for experiments that are not limited by the signal-to-noise ratio. Alternatively, it permits an increase in resolution in the indirectly detected dimensions of experiments where measuring time limits the number of increments that can be executed.

Heteronuclear 3D and 4D J correlation techniques make it possible to obtain complete \(^1\)H, \(^{13}\)C, and \(^{15}\)N resonance assignments for uniformly \(^{13}\)C/\(^{15}\)N-enriched proteins of up to several hundred amino acids (12, 13). Measurement of \(^1\)H–\(^1\)H NOEs, however, constitutes the basis for the protein structure determination process and both the precision and the accuracy of the resulting structures are directly related to the number of unambiguously determined NOEs (14, 15). For uniformly \(^{13}\)C/\(^{15}\)N-enriched proteins, the 4D \(^{13}\)C/\(^{15}\)N-separated NOESY (16) and the \(^{13}\)C/\(^{13}\)C-separated NOESY (17, 18) experiments are the methods of choice since they provide the highest possible dispersion of resonances. In the corresponding 4D spectra, \(^1\)H–\(^1\)H NOE cross peaks are separated in a third dimension by the frequency of the heteroatom attached to the first proton and in a fourth dimension by the frequency of the heteroatom attached to the second proton. Resolution in the heteronuclear dimensions of these 4D experiments typically has been limited by the fact that only eight complex points could be acquired in these dimensions, corresponding to very short acquisition times, on the order of a few milliseconds. Longer acquisition times, combined with the required extensive phase cycling, would result in unacceptably long measuring times. For the 4D \(^{13}\)C/\(^{13}\)C-separated HMQC–NOESY–HMQC experiment, the artifacts resulting from spurious magnetization-transfer pathways associated with the strong diagonal resonances in the spectrum were a major consideration in the design of the previously proposed eight-step phase cycle (17). This phase cycle is a compromise in which several noninteracting phase cycles are executed simultaneously. In addition, two trim pulses are needed to further reduce artifacts. In contrast, as will be demonstrated below, the gradient version of the 4D \(^{13}\)C/\(^{13}\)C-separated HMQC–NOESY–HMQC experiment does not lower the signal-to-noise ratio and requires only a two-step phase cycle that results in spectra with reduced artifacts and increased resolution.

The pulse scheme for the gradient version of the 4D \(^{13}\)C/\(^{13}\)C-separated HMQC–NOESY–HMQC experiment is shown in Fig. 1A. Because the experiment consists of simple building blocks that have extensively been discussed before (16–18), only the fate of some undesired coherences will be discussed, as these provide the rationale for the choice of the gradient pulses. In the following discussion, the spin operators I and S denote the coupled \(^1\)H and \(^{13}\)C spins.

The effects of the imperfection of a 180° refocusing pulse on transverse spin operators can be removed by the application of PFGs of identical strength and polarity at either side of this pulse (11). Both the set of PFGs labeled \(G_1\) and the set labeled \(G_4\) in Fig. 1A are used for this purpose. As
FIG. 1. Pulse schemes for the gradient versions of the 4D $^{13}$C/$^{13}$C-separated HMQC–NOESY–HMQC (A) and HMQC–NOESY–HSQC (B) experiments. Narrow and wide pulses denote 90° and 180° flip angles, respectively. Unless indicated otherwise, all pulses are applied along the $x$ axis.

A two-step phase cycle is used: $\phi_1 - \pi/2$, $\phi_2 - \pi/2$, and $\phi_3 - \pi/2$. Quadrature detection in $t_1$, $t_2$, and $t_3$ is obtained with the States–TPPI method (20), incrementing independently the phases $\phi_1$, $\phi_2$, and $\phi_3$, respectively. Sine-bell-shaped pulsed field gradients (PFG) with a strength of 7 G/cm at their maximum are applied along the $z$ axis. Gradients used have the following durations: $G_1 = 750 \mu s$, $G_2 = 2 \mu s$, $G_3 = 4 \mu s$, $G_4 = 750 \mu s$ (negative polarity). The delay $\delta$ is set to 3.5 ms and $^{13}$C decoupling during $t_1$ data acquisition is accomplished with a GARP decoupling scheme (21), using an RF field strength of 4.5 kHz.

the $G_4$ PFGs are applied before and after the $t_3$ period, evolution of the $^{13}$C spin operators during $t_3$ is not affected. Note, however, that the $\sin(\Omega t_3)/3$ fraction of the $^{13}$C magnetization, which remains as multiple-quantum coherence after the last $^{13}$C 90° pulse, is defocused by the last $G_4$ PFG. When not eliminated, this residual multiple-quantum coherence can give rise to significant decoupling modulation sidebands during the time $t_4$ (19). The effects of a nonideal 180° $^1$H pulse in the center of the $t_1$ period are eliminated in the same way as that described above for the last 180° $^1$H pulse.

Although, at the end of the $t_3$ evolution period, most $^1$H magnetization is in-phase with respect to its attached $^{13}$C, a small fraction remains antiphase because (a) for sensitivity reasons the delay $\delta$ is usually kept slightly shorter than $1/(2J_{CH})$, and (b) not all $^{13}$C spins are inverted by the composite 180° $^{13}$C pulse applied at the midpoint of $t_2$. As a consequence, the second 90° $^1$H pulse, applied at the start of the NOESY mixing period, can generate a small amount of $I_zS_z$ order. At the end of the NOESY mixing period, a fraction of this two-spin order could be converted by the remainder of the pulse sequence into spurious magnetization observable during the time $t_4$. It is therefore necessary to eliminate this type of $zz$ order during the NOE mixing period, together with all transverse magnetization components. This is accomplished by the application of the gradient $G_2$ (for the transverse terms), followed by a 90° $^{13}$C pulse and a subsequent gradient $G_3$ for eliminating the $I_zS_z$ order. Although the polarities of $G_1$ and $G_4$ relative to $G_2$ and $G_3$ can be chosen arbitrarily, $G_1$ and $G_4$ have opposite signs in order to minimize the disturbance of the deuterium lock.

The gradients used in the 4D $^{13}$C/$^{13}$C-separated HMQC–NOESY–HMQC experiment are applied either during delays that are needed for dephasing and rephasing of transverse magnetization or during the NOE mixing period. Therefore, no additional delays are required for the application of the gradient pulses. Although the gradients used are relatively weak (7 G/cm), their required duration is much shorter than the dephasing and rephasing periods, $\delta$.

In the absence of any phase cycling, the pulse scheme of Fig. 1A does not eliminate the signals from protons not attached to $^{13}$C, resulting in axial resonances in both the $F_1$ and the $F_3$ dimensions of the 4D spectrum. Removal of these axial peaks requires selection of the desired heteronuclear coherence-transfer pathways during $t_1$ and $t_3$ by means of PFGs (1–10), purging of signals from protons not attached to $^{13}$C, or additional phase cycling. We have chosen the last option, using a ±$x$ phase cycle of the first 90° $^{13}$C pulse, together with a ±$x$ phase cycle for the receiver. Because no such heteronuclear filtering is used after the NOESY mixing period, axial peaks, shifted to the edge of the spectrum in the $F_3$ dimension by the States–TPPI procedure (20), are present in the 4D spectrum. Fortunately, their intensities are weak as they result from the small fraction (≤−10%) of magnetization that is converted into multiple-quantum coherence during the time $t_3$, but that is not converted during the time $t_4$. Signals from protons not attached to $^{13}$C, such as the residual HDO signal, and signals originating from longitudinal relaxation during the NOE mixing period are effectively removed by the two-step phase cycle. However, for cases where the $^{13}$C enrichment is incomplete, or for the study of a complex between an unlabelled ligand and a labelled protein, an additional ±$x$ phase-cycling step is required for one of the 90° pulses of the last HMQC transfer, resulting in a four-step phase cycle for the 4D experiment. Alternatively, these signals may be removed by using an HSQC–instead of an HMQC-type $^1$H–$^{13}$C correlation after the NOESY mixing period (Fig. 1B), which allows for suppression of $^1$H coherence during the two INEPT stages of this experiment (17), without resorting to a four-step phase cycle.

However, the sensitivity of the scheme in Fig. 1B was found to be lower than that of the scheme in Fig. 1A, mainly due to the additional 180° pulses in the HSQC type of correlation, which are far from ideal near the edges of the $^1$H and $^{13}$C spectrum. Indeed, the loss in sensitivity is largest for interactions to methyl and aromatic protons. Therefore, we found the scheme in Fig. 1A preferable, using only a two-step phase cycle. For this experiment, the residual axial peaks at the edges of the $^{13}$C ($F_3$) spectral window can obscure the NOE...
correlation from a proton $H_A$ to a proton $H_B$ if its attached carbon, $C_B$, resonates close to the edge of the $F_3$ spectral window. However, its mirror image, corresponding to NOE transfer from $H_B$ to $H_A$, will be clearly observable unless $C_A$ also resonates close to the edge of the $F_3$ spectral window.

The experiment is demonstrated for a 1.7 mM solution of the protein interleukin-4 (IL-4) (~15.4 kDa), uniformly enriched (>95%) with $^{13}$C and $^{15}$N, dissolved in D$_2$O, pH 5.7. The experiment was recorded at 36°C on a Bruker AMX600, operating at a $^{1}$H resonance frequency of 600.13 MHz. A Bruker triple-resonance probe equipped with a self-shielded $z$ gradient was used, together with an in-house-developed gradient pulse-shaping unit and amplifier. Sine-bell-shaped gradient pulses had a strength of 7 G/cm at the center of the sine bell. To adjust different relative strengths of the PFGs, the durations and not the amplitudes of these pulses were varied.

Figure 2 shows two 2D cross sections taken through the 4D $^{13}$C/$^{15}$C-separated gradient-IMQ-C-NOESY-IMQ-C spectrum of IL-4, taken perpendicular to the $F_3$ ($^{13}$C) and $F_4$ ($^{1}$H) coordinates of the two $C^6$ methyl groups of Leu-113. These cross sections therefore show the frequencies of the protons (together with the chemical shifts of their attached $^{13}$C nuclei) that have an NOE interaction to these two methyl groups. Several features deserve special note. First, the resolution in the $^{13}$C ($F_3$) dimension is high for a 40 spectrum, permitting the unique identification of many NOEs in crowded parts of the spectrum, such as the methyl-
methyl region. For example, the structurally important NOE interactions between Leu113-C61H and Leu56-C62H or Leu83-C61H would be unresolvable with poorer resolution (Fig. 2A). Second, the level of artifacts is very low. Only some sinc wiggles resulting from truncation of the time-domain signal of the intense "diagonal resonance" of Leu113-C61H are observed. In fact, comparison between the 4D spectrum recorded with the previously published eight-step phase cycle (17) (data not shown) and the present data set indicates that the artifacts associated with the intense diagonal resonances are reduced nearly 10-fold. This low level of artifacts makes it possible to unambiguously identify the structurally important NOE between Leu113-C61H and Val33-C61H, despite the fact that the proton chemical shifts of these two methyl groups are nearly degenerate (Fig. 2A).

In Fig. 2B, NOEs to Leu113-C62H and Lys88-C61H are shown in a cross section taken at (F3, F4) = (25.2, 0.18) ppm. As expected, many of the NOEs observed for Leu113-C61H in Fig. 2A are also observed for Leu113-C62H. The cross section also shows intraresidue NOEs from the α, β, δ, and ε protons to the γ protons of Lys88. Interestingly, as a result of the high resolution in the F3 dimension (perpendicular to the plane shown), the F3 coordinates of the NOE cross peaks to Leu113-C61H and Lys88-C61H are sufficiently different (0.3 ppm) to unambiguously establish the NOE between Leu113-C62H and Lys88-C61H and to exclude the possibility of an NOE from Lys88-C61H to Leu87-C61H.

Our results show that straightforward incorporation of PFGs in the 4D 13C-separated HMQC–NOESY–HMQC experiment makes it possible to record high-quality 4D spectra with high resolution in all four frequency dimensions in a reasonable amount of measuring time (3.5 days). Incorporation of the PFGs does not decrease the sensitivity of the experiment but greatly reduces the level of spectral artifacts. Acquisition times in the 13C dimensions of the 4D experiment much longer than the 5.7 ms used in the present case would degrade the signal-to-noise obtainable per unit case of measuring time because the 13C signal in these dimensions rapidly decreases as the acquisition times approach 1/(2JCC) (~14 ms). Consequently, the durations of the acquisition times in the t1 and t2 13C dimensions used in the present experiment are a good compromise between optimum sensitivity and maximum spectral resolution. In principle, it is possible to record the 4D spectrum without any phase cycling by replacing both HMQC elements by HSQC elements with appropriate PFGs. However, because of the signal losses associated with the larger number of RF pulses required for an HSQC-type correlation, the intrinsic sensitivity of the HSQC version of the experiment is somewhat lower than that of the HMQC version.

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