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Increasing the Potency and Breadth of an HIV Antibody by using Structure-Based Rational Design*

Ron Diskin¹, Johannes F. Scheid^{2,3}, Paola M. Marcovecchio¹, Anthony P. West Jr.¹, Florian Klein², Han Gao¹, Priyanthi N. P. Gnanapragasam¹, Alexander Abadir², Michael S. Seaman⁴, Michel C. Nussenzweig^{2,5}, and Pamela J. Bjorkman^{1,5}

¹Division of Biology, California Institute of Technology, 1200 E. California Blvd., Pasadena, CA 91125

²Laboratory of Molecular Immunology, The Rockefeller University, New York, NY 10065

³Charité Universitätsmedizin, D-10117 Berlin, Germany

⁴Beth Israel Deaconess Med. Ctr., Boston, MA 02215; United States

⁵Howard Hughes Medical Institute, California Institute of Technology, 1200 E. California Blvd., Pasadena, CA 91125; Howard Hughes Medical Institute, The Rockefeller University, New York, NY 10065

Abstract

Antibodies against the CD4 binding site (CD4bs) on the HIV-1 spike protein gp120 can show exceptional potency and breadth. We determined structures of NIH45-46, a more potent clonal variant of VRC01, alone and bound to gp120. Comparisons with VRC01–gp120 revealed that a four-residue insertion in CDRH3 contributed to increased interaction between NIH45-46 and the gp120 inner domain, which correlated with enhanced neutralization. We used structure-based design to create NIH45-46^{G54W}, a single substitution in CDRH2 that increases contact with the gp120 bridging sheet and improves breadth and potency, critical properties for potential clinical use, by an order of magnitude. Together with the NIH45-46–gp120 structure, these results indicate that gp120 inner domain/bridging sheet residues should be included in immunogens to elicit CD4bs antibodies.

Three decades after the emergence of HIV there is no vaccine, and AIDS remains a threat to global public health (1-5). However, some HIV-infected individuals eventually develop broadly neutralizing antibodies (bNAbs), i.e., antibodies that neutralize a large panel of HIV viruses (6-11) and can delay viral rebound in HIV patients (12). Such antibodies are relevant to vaccine development because passive transfer into macaques can prevent infection (13-15). Antibodies obtained by new cloning methods (7,16,17) target several epitopes on the viral spike (7,8,16,18-20). The broadest and most potent are highly active agonistic anti-CD4 binding site antibodies (HAADs) that mimic binding of the host receptor CD4 (21) by exposing the co-receptor binding site on gp120 (8,16,22-24). Despite isolation from different donors, HAADs are derived from two closely-related Ig V_H genes that share gp120 contact residues (16,25). HAADs are typically members of large expanded clones (16) with variable levels of neutralizing activity despite intraclonal sequence similarities (16,25,26).

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Structures of gp120 complexes with VRC01, a highly potent and broad HAAD (25), and VRC03 and VRC-PG04, two new CD4-binding site (CD4bs) antibodies sharing the VRC01 germline V_H gene, revealed convergence of gp120 recognition despite low sequence identities (48-57% in V_H; 62-65% in V_L) (26). However, sequence differences between these clonally-unrelated antibodies make it difficult to determine structural features that correlate with neutralization potency and breadth.

To determine structural correlates of high potency and breadth in HAADs, we solved structures of NIH45-46, alone and bound to the clade A/E 93TH057 gp120 core (27) (Table S1, Fig. 1A,1B). NIH45-46 is a more potent clonal variant of VRC01 that was isolated from the same donor using a YU2 trimer (16) instead of a resurfaced gp120 core (RSC3) as a bait (8). Comparisons of NIH45-46 Fab in its free versus gp120-bound states demonstrate that gp120 binding does not require major conformational changes (Fig. 1A). However, gp120 binding induced minor conformational changes in CDRL1, CDRH3, and in heavy chain framework region 3 (FWR3). As predicted by high sequence identity (85% in V_H; 96% in V_L) (Fig. S1), NIH45-46 resembles VRC01 (Fig. S2A,B). However, relative to VRC01, NIH45-46 includes a four-residue insertion within CDRH3 (Fig. 2A) that was acquired by somatic hypermutation (16).

The crystal structure of the NIH45-46-93TH057 gp120 complex verified that NIH45-46 targets the CD4bs on gp120 (Fig. 1B,2A). The primary binding surface is the outer domain, including the CD4 binding loop (Fig. S3A), loop D and loop V5, but CDRH3_{NIH45-46} reaches toward the gp120 inner domain (Fig. 1B,2A-C). Important interactions in the VRC01-93TH057 structure (25) are conserved in NIH45-46 (Fig. S2B); e.g., residues C-terminal to CDRH2 of VRC01 and NIH45-46 mimic the interaction of mainchain atoms in the C' β-strand of CD4 domain, which hydrogen bond with the CD4-binding loop of gp120 (Fig. S3). In both NIH45-46 and VRC01, hydrogen bonds between CDRH2 and gp120 are water-mediated (25,26) (except for the Gly⁵⁴_{NIH45-46}/Gly⁵⁴_{VRC01} carbonyl oxygen-Asp³⁶⁸_{gp120} backbone nitrogen H-bond (Fig. S3)), and Arg⁷¹_{VRC01}/Arg⁷¹_{NIH45-46} preserves the Arg⁵⁹_{CD4} interaction with Asp³⁶⁸_{gp120}. However, the Phe⁴³_{CD4} interaction with a hydrophobic pocket between α-helix 3_{gp120} (CD4 binding loop) and β-strand 21_{gp120} (bridging sheet) (Fig. 3A,B) is not mimicked by either antibody. Differences between VRC01 and NIH45-46 include the conformation of heavy chain residue Tyr⁷⁴, a FWR3 residue that was substituted during somatic hypermutation (16), and a tyrosine to serine substitution in CDRL1 (Fig. S4,S5, Supplementary Discussion).

The most notable difference between VRC01 and NIH45-46 is the four-residue insertion (residues 99a-99d) in CDRH3. Three inserted residues contribute to binding to gp120 (Fig. 2A-inset), consistent with deletion of the insertion resulting in ~10-fold reduced neutralization potencies (Table S2). First, the Tyr^{99d}_{NIH45-46} sidechain hydrogen bonds with the loop D Ala²⁸¹_{gp120} carbonyl oxygen (Fig. 2B), a mainchain atom, thus preventing escape through mutation. Indeed, NIH45-46-sensitive strains accommodate different sidechains at position 281_{gp120} (Table S3). The importance of Tyr^{99d}_{NIH45-46} for potency is demonstrated by alanine substitution (NIH45-46 Y99dA), which reduces the neutralization potency of NIH45-46 to values intermediate between wild-type NIH45-46 and the deletion mutant (Table S2). Second, Asp^{99c}_{NIH45-46} interacts electrostatically with Lys⁹⁷_{gp120} at the base of α-helix 1_{gp120}, and third, Arg^{99b}_{NIH45-46} hydrogen bonds with Asn⁹⁹_{gp120} (Fig. 2C). The conformation of the insertion is stabilized by two intramolecular hydrogen bonds. In one, the Tyr^{99d}_{NIH45-46} sidechain hydrogen bonds with the ε-amino group of Lys⁵²_{NIH45-46} within CDRH2 (Fig. 2B), also seen in the unbound structure of NIH45-46 (Fig. 2B-inset), thus the Tyr^{99d}_{NIH45-46} hydroxyl is poised for interacting with Ala²⁸¹_{gp120}. A second hydrogen bond between Tyr⁹⁷_{NIH45-46} and Asp^{99c}_{NIH45-46} in the gp120-bound Fab positions the negatively-charged aspartic acid for interaction with Lys⁹⁷_{gp120} (Fig. 2C).

The region of gp120 with which CDRH3_{NIH45-46} interacts was not included in the previously-defined vulnerable site of initial CD4 attachment on the gp120 outer domain (25) (Fig. 3C). Thus, gp120 residues that contact CDRH3_{NIH45-46} residues required for potent neutralization (Table S2), e.g., Lys97_{gp120}, were mutated in RSC3 (Fig. S6), the resurfaced gp120 used for isolating bNAbs (8) and as a candidate HIV immunogen (28).

The insertion in CDRH3 contributes to a higher total buried surface area between the NIH45-46 heavy chain and gp120 compared with VRC01 (Table S4). The extra contacts with gp120 created by the CDRH3 insertion allow the NIH45-46 footprint on gp120 to more closely resemble the CD4 footprint on gp120 than does the VRC01 footprint (Fig. 3C, Table S4). The observation that NIH45-46 shows more extensive contacts relative to VRC01 with the inner domain and bridging sheet of gp120 (Fig. 3C), yet exhibits higher potency and breadth (16), is inconsistent with the suggestion that increased contact area with regions outside of the outer domain of gp120 correlate with decreased neutralization potency and/or breadth (25,26). Indeed, the observed CDRH3 contacts with the inner domain imply that the crystallographically-observed conformation of this region, whether pre-existing or induced, actively played a role in the affinity maturation events that resulted in the four-residue insertion with CDRH3.

Although NIH45-46 increases its contacts with the inner domain/bridging sheet area of gp120, like VRC01, it lacks a critical CD4 contact to a hydrophobic pocket at the boundary between the gp120 bridging sheet and outer domain made by burying Phe43_{CD4}. This residue alone accounts for 23% of the interatomic contacts between CD4 and gp120, serving as a “linchpin” that welds CD4 to gp120 (29). On gp120, the Phe43 binding cavity is a binding site of small-molecule CD4 mimics (30) and a desirable target for compounds to disrupt CD4–gp120 interactions (29), yet it remains unfilled in the 93TH057 complexes with VRC01 (25) and NIH45-46. In a superimposition of a CD4–gp120 structure (31) and NIH45-46–gp120 (Fig. 3B), the C α atom of heavy chain residue Gly54_{NIH45-46} is only ~1.4 Å from the Phe43_{CD4} C α , suggesting that this important interaction might be mimicked by substituting Gly54_{NIH45-46} with a large hydrophobic residue. Indeed, residue 54 of VRC03 is a tryptophan, and Trp54_{VRC03} is accommodated within gp120’s Phe43 binding cavity to mimic Phe43_{CD4}, while still maintaining its mainchain hydrogen bond with Asp368_{gp120} (PDB 3SE8) (Fig. S3). If increasing contacts with the inner domain/bridging sheet enhances antibody activity, as suggested by analysis of the NIH45-46–gp120 structure, then substituting Gly54_{NIH45-46} with a large hydrophobic residue should increase the potency and breadth of NIH45-46.

We constructed a series of NIH45-46 mutants to test the possibility that a hydrophobic sidechain at position 54 in NIH45-46 would improve activity. We first verified that substitutions at residue 54 did not interfere with antigen binding by assessing the ability of one mutant, NIH45-46^{G54W}, to bind core gp120s. Surface plasmon resonance (SPR) binding analyses demonstrated that NIH45-46^{G54W} Fab bound core gp120s with slightly higher affinities than did NIH45-46 Fab, with differences largely due to slower dissociation rates (Fig. S7). We next evaluated mutant IgGs in neutralization assays using a panel of six viruses chosen to include NIH45-46–sensitive and resistant strains (Fig. S8A). NIH45-46^{G54W} and NIH45-46^{G54F} showed increased potencies and NIH45-46^{G54W} increased breadth by neutralizing three strains that are resistant to >50 µg/mL of NIH45-46. The apparent increase in breadth is likely due to increased potency as evidenced by the extrapolated IC₅₀ for NIH45-46 against strain DU172.17 (Fig. S8B).

We further tested 82 viruses including 13 NIH45-46–resistant, 14 weakly-neutralized, and 55 sensitive strains representing all clades, of which 12 are transmitted founder viruses (Fig 4A, Tables S5,S6). This panel is more difficult for NIH45-46 to neutralize than a recently-

published panel (16) (Fig. 4B). NIH45-46^{G54W} showed increases in potency and breadth compared to NIH45-46 and VRC01: geometric mean IC₅₀s (32) of 0.04 µg/mL for NIH45-46^{G54W}, 0.41 µg/mL for NIH45-46, and 0.92 µg/mL for VRC01 (calculated for 65 viruses against which VRC01 was previously evaluated (16)) (Tables S5,S6, Fig. 4C). Notably, NIH45-46^{G54W} gained *de novo* neutralization activity against six NIH45-46 resistant strains, including the only three that were sensitive to VRC01 but resistant to NIH45-46 in the panel tested in (16). Remarkably, for some strains that NIH45-46 neutralizes poorly, NIH45-46^{G54W} was significantly more potent (e.g., improvements of >700-fold for T255-34 and 2000-fold for 3718.v3.c11). The enhanced neutralization activity of NIH45-46^{G54W} implies that Trp54 forms a favorable hydrophobic interaction with Phe43 cavity of gp120 as seen in VRC03–gp120 (PDB 3SE8). NIH45-46^{G54F} showed some increased activity, but other substitutions were less favorable (Fig. S8A, Tables S5,S6), including serine (25), which is found in the NIH45-46/VRC01 germline precursor. Substituting Gly54 with tryptophan adds ~140 Å² of buried surface area on V_H when complexed with gp120, consistent with the reduced dissociation rates observed in SPR experiments (Fig. S7), and provide a critical contact to a region of gp120 that cannot change without loss of CD4 tropism. By providing a tryptophan in the Phe43 cavity of gp120, NIH45-46^{G54W} may use higher affinities and/or slower dissociation rates to overcome incompatible surface variations that render some viruses less sensitive or resistant to its effects. If so, resistance to NIH45-46^{G54W} might require restricted access to the binding site on a native trimeric spike. Inspection of sequences of strains resistant to NIH45-46^{G54W} and/or NIH45-46 suggests that variations at contact sites near V5 and in loop D may be responsible (Table S7) (25), consistent with mutagenesis studies of neutralization by VRC01 (33).

Heavy chain residue 54 is not conserved in HAADs; in addition to glycine (NIH45-46 and VRC01), residue 54 can be threonine (3BNC60, 3BNC117, 3BNC115; VRC-PG04), tyrosine (12A12), phenylalanine (12A21), or arginine (1B2530 and 1NC9) (16, 26). Tryptophan substitution did not improve all HAADs that were tested (Table S8), thus increased potency requires a precise geometry.

Passive immunization and/or gene therapy to deliver HIV antibodies is increasingly being considered an option for prevention of HIV infection. To reduce the concentrations and numbers of antibodies required for protection to realistic and affordable levels, highly potent and broadly neutralizing antibodies are the reagents of choice for passive delivery. Although it is difficult to compare the potencies and breadth of antibodies characterized using different virus panels, the natural form of NIH45-46 exhibits superior potency to VRC01 when compared against a panel of 82 Tier 2 and 3 viruses representing all known HIV clades (16). A new set of HIV antibodies, the PGT antibodies that recognize the gp120 V3 loop and associated carbohydrates, exhibited median IC₅₀s up to 10-fold lower than VRC01 (18), but are less potent and broad than NIH45-46^{G54W} (Fig. 4B, Table S9). The successful improvement of the potency and breadth of NIH45-46, already one of the most potent and broad of the currently-available bNAbs (16), suggests that NIH45-46^{G54W} should be among a small number of characterized HIV antibodies considered for clinical testing. Since escape from even the best bNAbs is possible, optimal results may be achieved using NIH45-46^{G54W} in combination with one or more other bNAbs.

Our results also have important implications for vaccine development, i.e., the findings that contacts to the gp120 inner domain by NIH45-46 and increased interactions with the bridging sheet by NIH45-46^{G54W} improve the breadth and potency of a HAAD antibody suggest that immunogens used in vaccine strategies should include native inner domain/bridging sheet residues. Although this sort of immunogen may elicit some non-neutralizing CD4bs antibodies, raising the most potent and broad CD4bs antibodies will require contact

with regions within the gp120 inner domain and bridging sheet in addition to the initial site of CD4 attachment on the gp120 outer domain.

Supplementary Material

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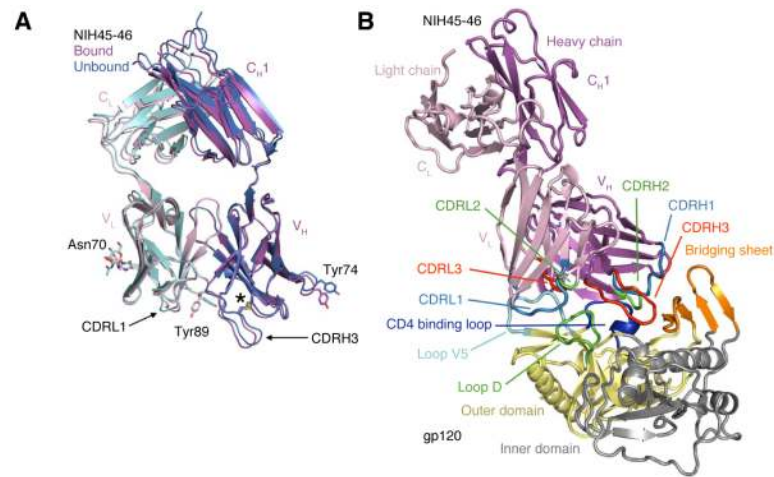
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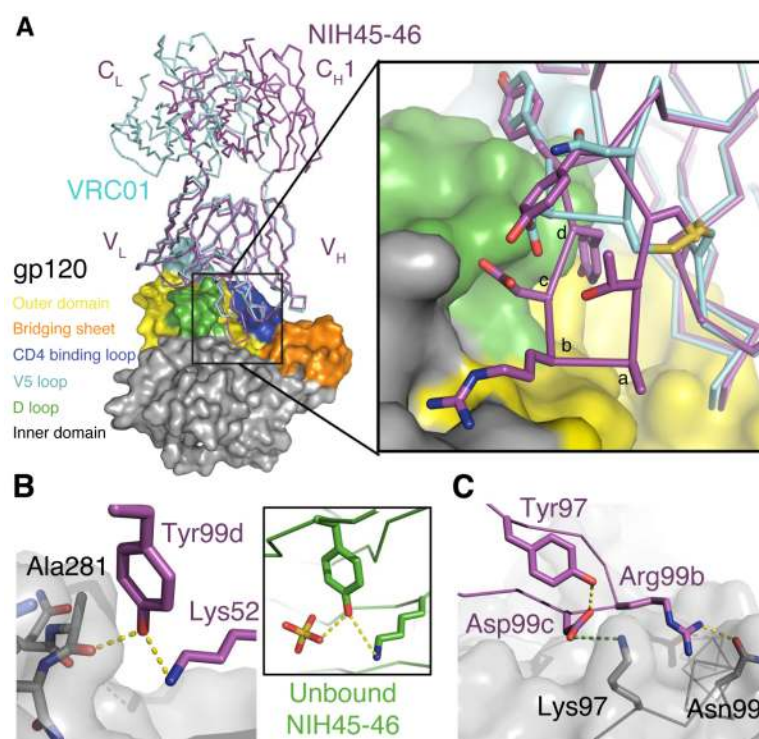
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**Fig. 1.**

Crystal structures of NIH45-46 Fab alone and bound to gp120. **(A)** Superimposition of the structures of the free (blue heavy chain and cyan light chain) and bound (magenta heavy chain and pink light chain) NIH45-46 Fab. RMSDs for free and bound V_H-V_H and V_L-V_L superimpositions (123 and 99 C α atoms, respectively) are each 0.5 Å. The location of an extra disulfide bond that joins Cys32–Cys98 in V_H is marked with an asterisk and N-linked carbohydrate attached to Asn70 of V_L is shown as sticks. Arrows point to slightly different conformations in CDRL1 and CDRH3 in bound and free NIH45-46. Highlighted sidechains, Tyr89_{CDRL3} and Tyr74_{FWRH3}, adopt notably different conformations upon binding gp120. **(B)** Structure of the NIH45-46-gp120 complex. Ribbon diagram of the NIH45-46 Fab (magenta and pink for the heavy and light chains, respectively) complexed with the inner (yellow) and outer (gray) domains of the 93TH053 gp120 core, which lacks three variable loops (V1-V2 and V3) and has N- and C-terminal truncations (25). Lines point to structural features of the gp120 core discussed in the text.

**Fig. 2.**

Interactions of the NIH45-46 insertion with gp120. **(A)** Superimposition of the gp120 portions of VRC01–93TH053 (PDB 3NGB) and NIH45-46–93TH053 structures. The Fabs are shown as magenta (NIH45-46) or cyan (VRC01) wire, and gp120 is shown as a surface with the color scheme used in Fig. 1B. The region surrounding the four-residue insertion in the NIH45-46 CDRH3 (residues 99a – 99d) is boxed. Inset: Close-up of the boxed region in which the NIH45-46 insertion residues are labeled alphabetically to correspond with residues 99a – 99d. The sidechains of relevant CDRH3 residues are shown as sticks. **(B)** Hydrogen bond network between the mainchain carbonyl oxygen of Ala281_{gp120}, Tyr99d_{NIH45-46} in CDRH3, and Lys52_{NIH45-46} in CDRH2. Yellow dots represent hydrogen bonds. The conformation of Tyr99d_{NIH45-46} is also stabilized by a hydrogen bond with Lys52_{NIH45-46} in unbound NIH45-46 (inset) in which a sulfate ion (yellow) from the crystallization solution substitutes for the interaction with Ala281_{gp120}. **(C)** Electrostatic interaction between Asp99c_{NIH45-46} and Lys97_{gp120} (green dots) is shown with Asp99c_{NIH45-46}-Tyr97_{NIH45-46} and Arg99b_{NIH45-46}-Asn99_{gp120} hydrogen bonds (yellow dots).

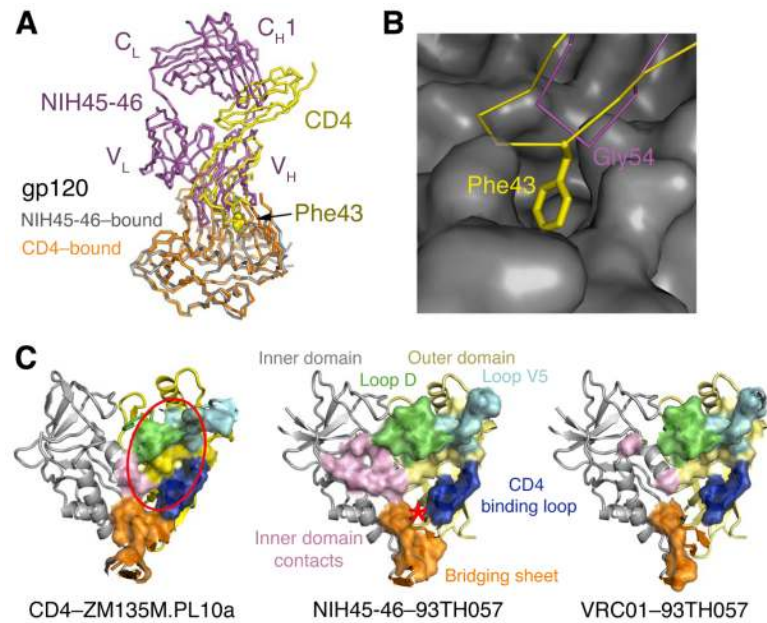
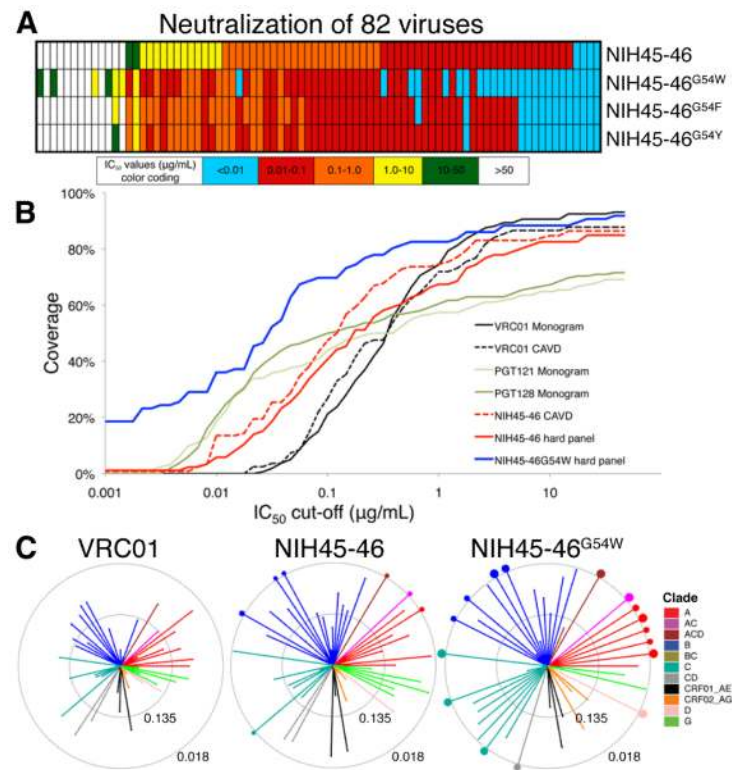


Fig. 3. NIH45-46 mimicry of CD4 binding. (A) Superimposition of NIH45-46-gp120 (magenta and gray, respectively) and CD4-gp120 (yellow and orange, respectively) (31) calculated based on gp120 C α positions. Phe43_{CD4} is shown using spheres. (B) Close-up of the CDRH2 loop of NIH45-46 (magenta) and CDR2-like loop of CD4 (yellow) interacting with gp120 (gray surface). Phe43_{CD4} (labeled) inserts into a hydrophobic pocket on gp120. The closest corresponding residue on NIH45-46 (Gly54_{NIH45-46}) is labeled. (C) Comparison of contacts on gp120 made by CD4, NIH45-46, and VRC01 (left, middle and right respectively). Residues at each contact interface are highlighted on the gp120 structure (colored as in Fig. 1B) as a surface enclosing the contact residues. The location of the Phe43 cavity targeted by the tryptophan substitution in NIH45-46^{G54W} is indicated on the middle panel with a red asterisk. The approximate location of the gp120 region defined as the initial site of CD4 attachment (25, 26) is indicated with a red oval in the left panel.

**Fig. 4.**

Increased neutralization potency of NIH45-46^{G54W}. **(A)** Schematic comparing neutralization potencies for NIH45-46 and three position 54 mutants, including NIH45-46^{G54W}. IC_{50} values are color-coded as shown. Numerical values for the same 82 strains are shown in Table S5 (IC_{50} values) and Table S6 (IC_{80} values). **(B)** Graphical comparisons of neutralization coverage and potency. The y-axis shows the cumulative frequency of IC_{50} values up to the concentration shown on the x-axis. The curves for VRC01, the only antibody tested in both the CAVD panel (a panel of 118 viral strains (16)) and the Monogram panel (a panel of 162 viral strains (18)), are similar, although the CAVD panel includes a few more resistant strains (compare the solid and dotted black lines). NIH45-46 (red dotted line) was tested on the CAVD panel and is more potent than VRC01 and broader than the PGT Abs. NIH45-46^{G54W} (solid blue line) was tested with NIH45-46 (solid red line) on a panel of 82 viral strains chosen to include many NIH45-46 resistant and poorly neutralized strains (hard panel; Tables S5,S6), thus resulting in lower coverage and lower potency for NIH45-46 when evaluated using the hard panel versus using the CAVD panel (compare the solid and dotted red lines). The improved potency and breadth of NIH45-46^{G54W} versus NIH45-46 against the hard panel are apparent. Even when evaluated using a difficult to neutralize panel of viruses, NIH45-46^{G54W} is the most potent and broadly neutralizing antibody described to date. **(C)** Neutralization summary spider graphs comparing IC_{50} values for VRC01, NIH45-46, and NIH45-46^{G54W} for 65 common viruses that were tested in this work or previously (16). Each color represents a different HIV clade. Length of lines and size of circles are inversely proportional to IC_{50} . The distance between the outer and inner circle and the distance from the inner circle to the center of a spider graph each span two natural logs in IC_{50} concentration (concentrations in $\mu\text{g/mL}$ are indicated on each circle). Dots on the outer circle indicate strains with IC_{50} values less than 0.018 $\mu\text{g/mL}$ whose lines were truncated in the graph. The size of each dot is inversely proportional to the IC_{50} value.